

# Rare diseases: Insights from landscape analysis of current research, spotlighting amyotrophic lateral sclerosis, Huntington's disease, and myasthenia gravis

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## Abstract

Rare diseases are a diverse group of disorders that despite each individual condition's rarity, collectively affect a significant portion of the global population. Currently approximately 10,000 rare diseases exist globally, 80% of these diseases have been identified as of genetic origins. In this review, we examine data from the CAS Content Collection to summarize scientific progress in the area of rare diseases. We examine the publication landscape in the area in effort to provide insights into current advances and developments. We then discuss the evolution of key concepts in the field, genetic associations, as well as the major technologies and development pipelines of rare disease treatments.

We focus our attention on three specific rare diseases: (i) amyotrophic lateral sclerosis, a terminal neurodegenerative disease affecting the central nervous system resulting in progressive loss of motor neurons that control voluntary muscles; (ii) Huntington's disease, another terminal neurodegenerative disease that causes progressive degeneration of nerve cells in the brain, with a wide impact on a person's functional abilities; and (iii) myasthenia gravis, a chronic autoimmune synaptopathy leading to skeletal muscle weakness. While the pathogenesis of these rare diseases is being elucidated, there is neither a cure nor preventative treatment available, only symptomatic treatment. The objective of the paper is to provide a broad overview of the evolving landscape of current knowledge on rare diseases, to outline challenges, and evaluate growth opportunities, all with an aim to further efforts in solving the remaining challenges.

**Key words:** rare disease; amyotrophic lateral sclerosis; Huntington's disease; myasthenia gravis; gene; pathogenesis; neurodegeneration; autoimmunity

## 1. Introduction

Rare diseases, also known as orphan diseases, are a diverse group of disorders that, despite each individual condition's rarity, collectively affect a significant portion of the global population. The exact definition of a rare disease varies from country to country, but generally, a disease is considered rare when it affects a limited number of people within a specific region or population. The World Health Organization (WHO) defines a rare disease as one that strikes fewer than 65 per 100,000 people.<sup>1</sup> According to the National Organization of Rare Disorders of the USA,<sup>2</sup> a rare disorder is a disease or condition that affects fewer than 200,000 Americans (i.e., less than ~60 per 100,000).<sup>3,4</sup> In the European Union, a rare disease is one that affects no more than 1 person in 2000<sup>5</sup> (i.e., <50 per 100,000). According to Global Genes, a global non-profit advocacy organization for individuals fighting rare and genetic diseases<sup>6</sup>, currently approximately 10,000 rare diseases exist globally, with new ones being continually discovered. About 80% of these diseases have been identified as of genetic origins<sup>7</sup>, and they often manifest in diverse and unpredictable ways.

Despite their individual rarity, rare diseases collectively impact millions of individuals worldwide – approximately 4-6% of the worldwide population, equivalent to from 300 to over 400 million people.<sup>1, 8, 9</sup> These conditions often present unique challenges due to their unfamiliarity, limited treatment options, and the difficulties associated with diagnosis. One of the primary challenges associated with rare diseases is their diagnosis. Due to their rarity and often complex clinical presentations, rare diseases are frequently misdiagnosed or undiagnosed altogether. This diagnostic struggle can be emotionally and financially difficult for patients and their families, leading to delays in appropriate treatment and care.

Moreover, the limited understanding of many rare diseases poses significant obstacles to the development of effective therapies. With only a handful of patients available for clinical trials, research into these conditions is often underfunded and progresses at a slower pace compared to more common diseases. Consequently, individuals living with rare diseases may have few, if any, treatment options available to them. As rare diseases each individually affect a small number of individuals, they have been 'orphaned' by the pharmaceutical industry, which has promoted the use of the term 'orphan disease' when referring to these conditions.<sup>1</sup>

Another critical issue facing the rare disease community is the lack of specialized healthcare providers and support services. Many rare diseases require multidisciplinary care from experts in various medical specialties, but access to such expertise can be limited, particularly in rural or underserved areas. Globally, less than 10% of patients with rare diseases receive disease-specific treatment.<sup>1</sup>

However, recent years have seen a growing awareness of rare diseases, leading to increased efforts to address the unmet needs of those affected. Since most rare diseases are genetic in their etiology, systematic research on them starts with efforts to identify genetic variants causative for each particular disease, with links between genetic mutations and diseases identified.<sup>10, 11</sup> Advances in genomic sequencing technologies and precision medicine hold promise for improved diagnosis and targeted treatments for many rare diseases. Additionally, initiatives such as orphan drug legislation<sup>12-14</sup> and incentives for rare disease research<sup>15-17</sup> have motivated pharmaceutical companies to invest in the development of therapies for these often-neglected conditions.

In this paper, we examine data from the CAS Content Collection<sup>18</sup>, the largest human-curated collection of published scientific information, and analyze the publication landscape of

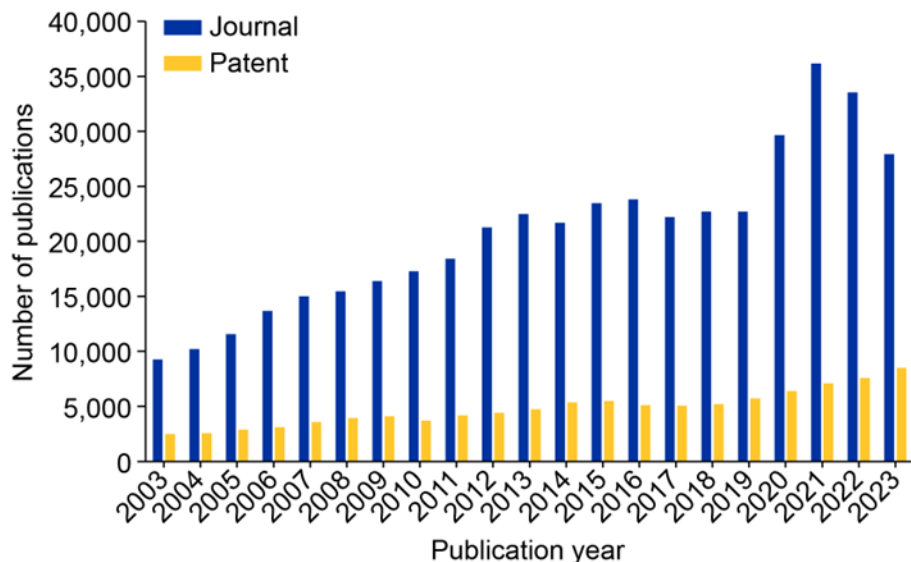
recent research in order to provide insights into the scientific progress in the area of rare diseases. We discuss the evolution of key concepts in the field as well as the major technologies and the development pipelines of rare disease treatments. We focus our attention on three specific rare diseases to perform a deeper dive into the research outlook in order to identify and understand obscure connections in these topics, namely: (i) amyotrophic lateral sclerosis (ALS), a terminal neurodegenerative disease affecting the central nervous system (CNS) resulting in progressive loss of motor neurons that control voluntary muscles; (ii) Huntington's disease (HD), another terminal neurodegenerative disease that causes progressive degeneration of nerve cells in the brain, with a wide impact on a person's functional abilities; and (iii) myasthenia gravis (MG), a chronic autoimmune synaptopathy leading to skeletal muscle weakness.

The objective of the paper is to provide a broad overview of the evolving landscape of current knowledge on rare diseases, to outline challenges, and evaluate growth opportunities, all with an aim to further efforts in solving the problems that still plague the field. The novelty and merit of the article stem from the extensive, wide-ranging coverage of the most up-to-date scientific information accumulated in the CAS Content Collection, allowing unique, unmatched breadth of landscape analysis and in-depth insights. We hope this report can serve as a useful resource for understanding the current state of knowledge and the importance of raising awareness in the field of rare disease research and development.

## **2. Landscape analysis of rare diseases research: Publication and patent trends from the CAS Content Collection**

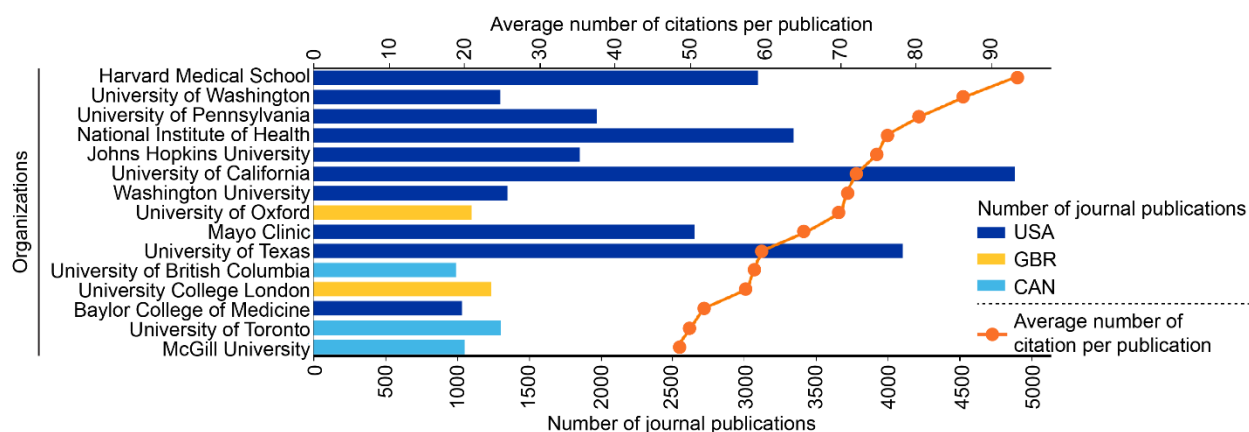
In this section we present our findings from a comprehensive analysis of more than 530,000 publications (journals and patents) in the field of rare diseases sourced from the CAS Content Collection™. Our aim for this analysis was to identify interesting trends in the field such as – leading research organizations and scientific journals, as well as identify leading rare diseases in terms of commercial exploration. In addition, we have focused on three of the most voluminous (in terms of journal and patent publications) rare diseases – ALS, HD and MG. Finally, we leveraged CAS REGISTRY, the CAS substance collection, to identify substances across different substance classes co-occurring with the three chosen rare diseases.

To fully capture the field, our subject matter experts utilized more than >650 search terms to ensure both identification of relevant publications in the field as well as capture a wide breath of pertinent information. The last two decades have seen an increasing interest in rare diseases as shown by the steady increase in journal publications, with a marked and steep increase between 2019-2021. Patent publications on the other hand, have increased consistently but at a much more moderate pace (Figure 1).



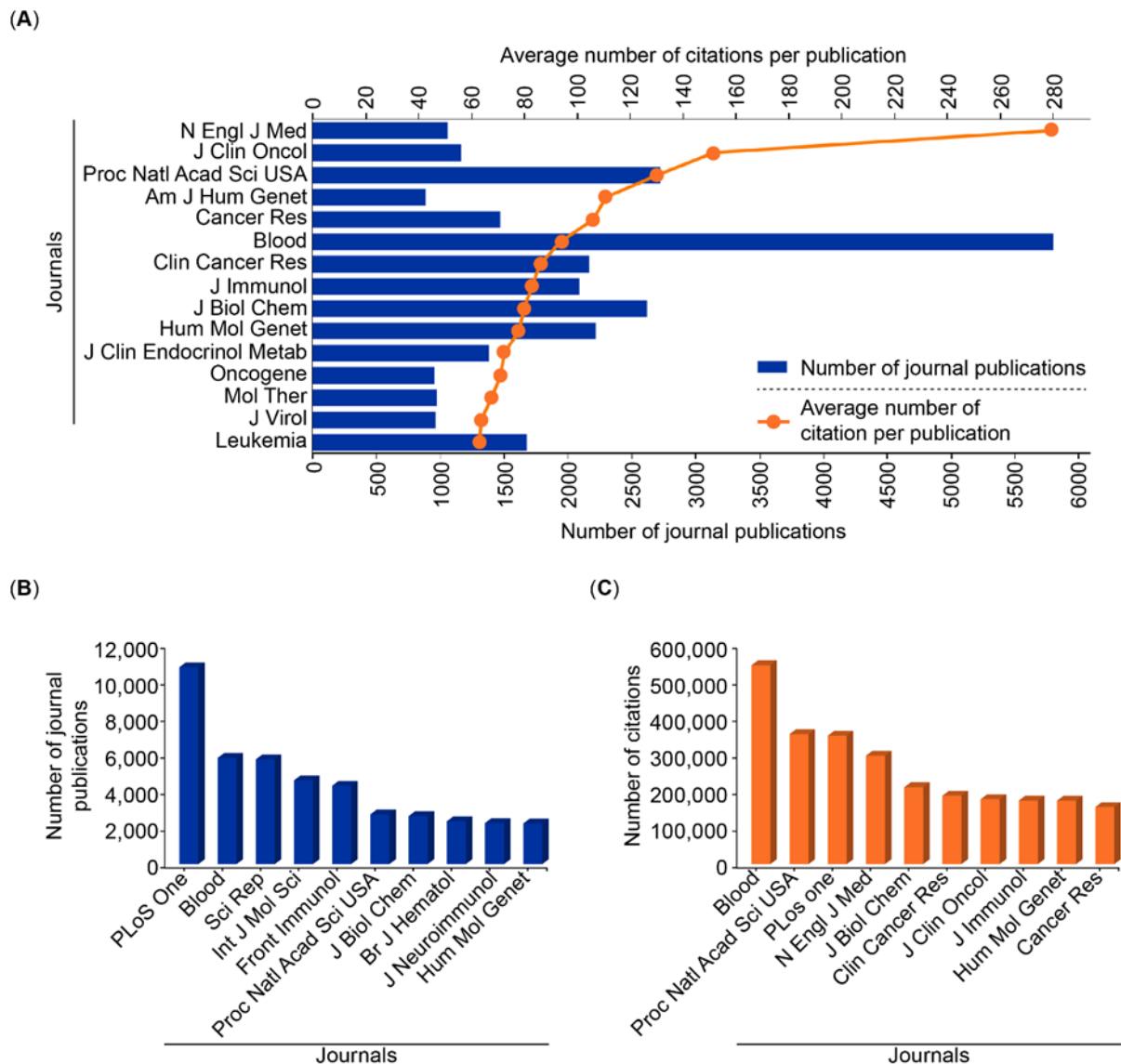
**Figure 1.** Publications in the field of rare diseases from the CAS Content Collection for the period 2003 to 2023.

To identify leading research organizations actively involved in publishing scientific content related to rare diseases, we first ranked them by the volume of journal articles followed by calculating the average number of citations per publication, a viable indicator of the scientific impact for published work. The top 100 research organizations in terms of number of journal articles were then ranked by the average number of citations per publication to give rise to the list in Figure 2. In terms of geographic distribution, ~67% of the leading 15 research institutions originated from the United States (USA). The remaining 35% consisted of research organizations from Canada (CAN) and the United Kingdom (GBR) accounting for 20% and 13%, respectively (Figure 2). Within the USA, key universities identified were University of Washington, University of Pennsylvania, Johns Hopkins University, University of California, Washington University, and University of Texas. Other important research organizations from the USA were Harvard Medical School, which ranked 1<sup>st</sup> among the leading 15 organizations, the US government run agency National Institute of Health (NIH), as well as the Mayo Clinic, and Baylor College of Medicine. Examples of recent journal articles published by researchers at Harvard Medical School include articles related to HD,<sup>19, 20</sup> MG,<sup>21</sup> ALS,<sup>19, 20</sup> as well as rare tumors and cancers such as thymomas,<sup>22</sup> multiple myeloma,<sup>21</sup> and thyroid cancer.<sup>23-25</sup> Similarly, examples of research output from other leading US-based organizations revolve around rare diseases such as multiple sclerosis,<sup>26, 27</sup> ALS,<sup>28-30</sup> systemic lupus erythematosus (SLE),<sup>31, 32</sup> sickle cell anemia,<sup>33, 34</sup> among others and rare cancers such as hematological malignancies including multiple myeloma,<sup>22</sup> acute myeloid leukemia (AML),<sup>35, 36</sup> and non-Hodgkin's lymphoma.<sup>37, 38</sup> Research organizations originating in Canada and those originating in the United Kingdom (GBR) also appear to be directing research efforts towards rare diseases such as multiple sclerosis,<sup>39-42</sup> HD,<sup>43-46</sup> SLE,<sup>47-50</sup> and ALS,<sup>51, 52</sup> among others.



**Figure 2.** Leading research organizations in the field of rare diseases based on data from the CAS Content Collection for the period 2003 to 2023. Ranking was based on total number of journal publications and average number of citations per publication, an indicator of publication impact.

Using a similar methodology as described above, we identified leading scientific journals in the area of rare diseases – first by ranking journals by the volume of publications, followed by calculating average number of citations per publication, and finally by ranking them using the calculated average number of citations per publication. The leading scientific journals identified include the *New England Journal of Medicine* (*New Engl J Med*), *Journal of Clinical Oncology* (*J Clin Oncol*), and *Proceedings of the National Academy of Sciences* (*Pro Natl Acad Sci USA*), among others (Figure 3A). The scientific journal *New England Journal of Medicine* has half as many journal articles as the *Proceedings of the National Academy of Sciences* but two times the average number of citations per publication. When looked at from the lens of sheer volume of publications or number of citations a somewhat different group of scientific journals emerge. There is a moderate degree of overlap between the two lists (number of journal publications and number of citations) with scientific journals such as *Blood*, *PLOS One*, *Proceedings of the National Academy of Sciences*, *Journal of Biological Chemistry* (*J Biol Chem*), and *Human Molecular Genetics* (*Hum Mol Genet*) featuring in both lists (Figure 3B). However, the order of appearance differs, for instance, while the open-access journal *PLOS One* leads in terms of sheer number of publications (nearly 2X fold higher than the journal *Blood*, the 2<sup>nd</sup> leading journal), it is 3<sup>rd</sup> in terms of number of citations.



**Figure 3.** Leading research journals in the field of rare diseases based on data from the CAS Content Collection for the period 2003 to 2023. (A) Ranking was based on both total number of journal publications and average number of citations per publication, an indicator of publication impact. Leading research journals when ranking was based solely on total number of (B) journal publications and (C) citations.

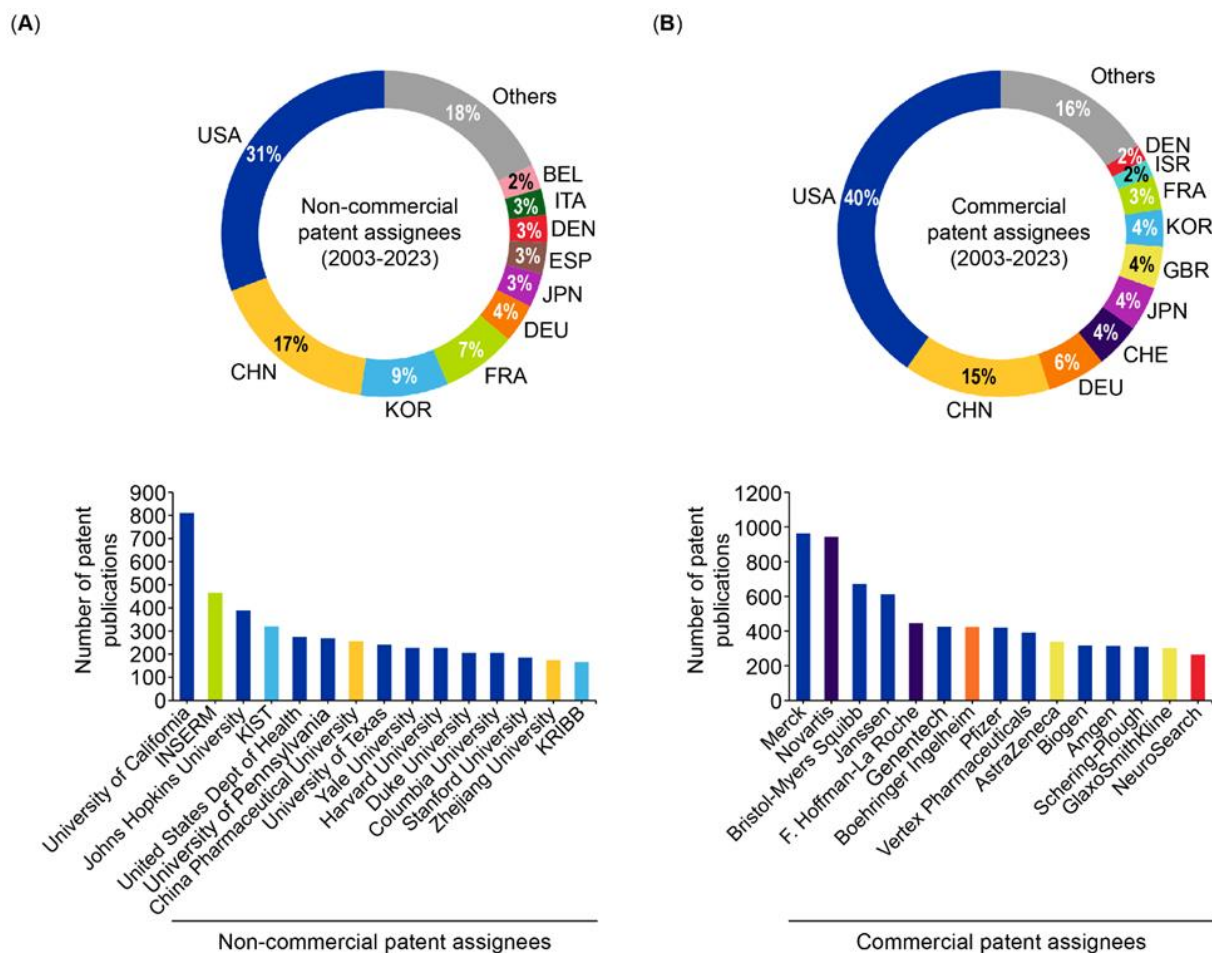
A few highly cited journal articles published in the New England Journal of Medicine revolve around rare cancers such as melanoma,<sup>53, 54</sup> multiple myeloma,<sup>55</sup> and AML<sup>53, 54</sup> as well as other rare diseases such as SLE,<sup>55</sup> multiple sclerosis,<sup>56, 57</sup> and cystic fibrosis.<sup>58</sup> The 2019 article titled “Targeting huntingtin expression in patients with Huntington's disease”<sup>59</sup> by researchers in the United Kingdom and published in the New England Journal of Medicine described results of a Phase I/IIa trial for an oligonucleotide designed by Ionis Pharmaceuticals and F. Hoffmann–La Roche to inhibit mRNA of HTT, the main gene responsible for HD and has been cited more than 400 times. Another example of a highly cited article was “Neurotoxic reactive astrocytes are induced by activated microglia” published in 2017 in the journal Nature and described the role of A1 astrocytes in neuronal cell death and their abundance in HD, ALS, and other

neurodegenerative disorders.<sup>60</sup> Similar well-cited publications for MG include a review article published in the New England Journal of Medicine.<sup>61</sup>

Geographical distribution of patent assignees in the field of rare diseases indicates a high degree of overlap in leading countries or regions with 7 out of 10 being common between commercial as well as non-commercial entities – United States (USA), China (CHN), South Korea (KOR), France (FRA), Germany (DEU), Japan (JPN), Denmark (DEN) (Figure 4). Spain (ESP), Italy (ITA), and Belgium (BEL) appear to have greater non-commercial presence while the inverse appears to be true for the United Kingdom (GBR) and Israel (ISR). Overall, the USA contributes 31% and 40% to non-commercial and commercial patents, respectively, the highest by a country/region. This is followed by China (CHN) which appears to have a more comparable commercial and non-commercial contribution.

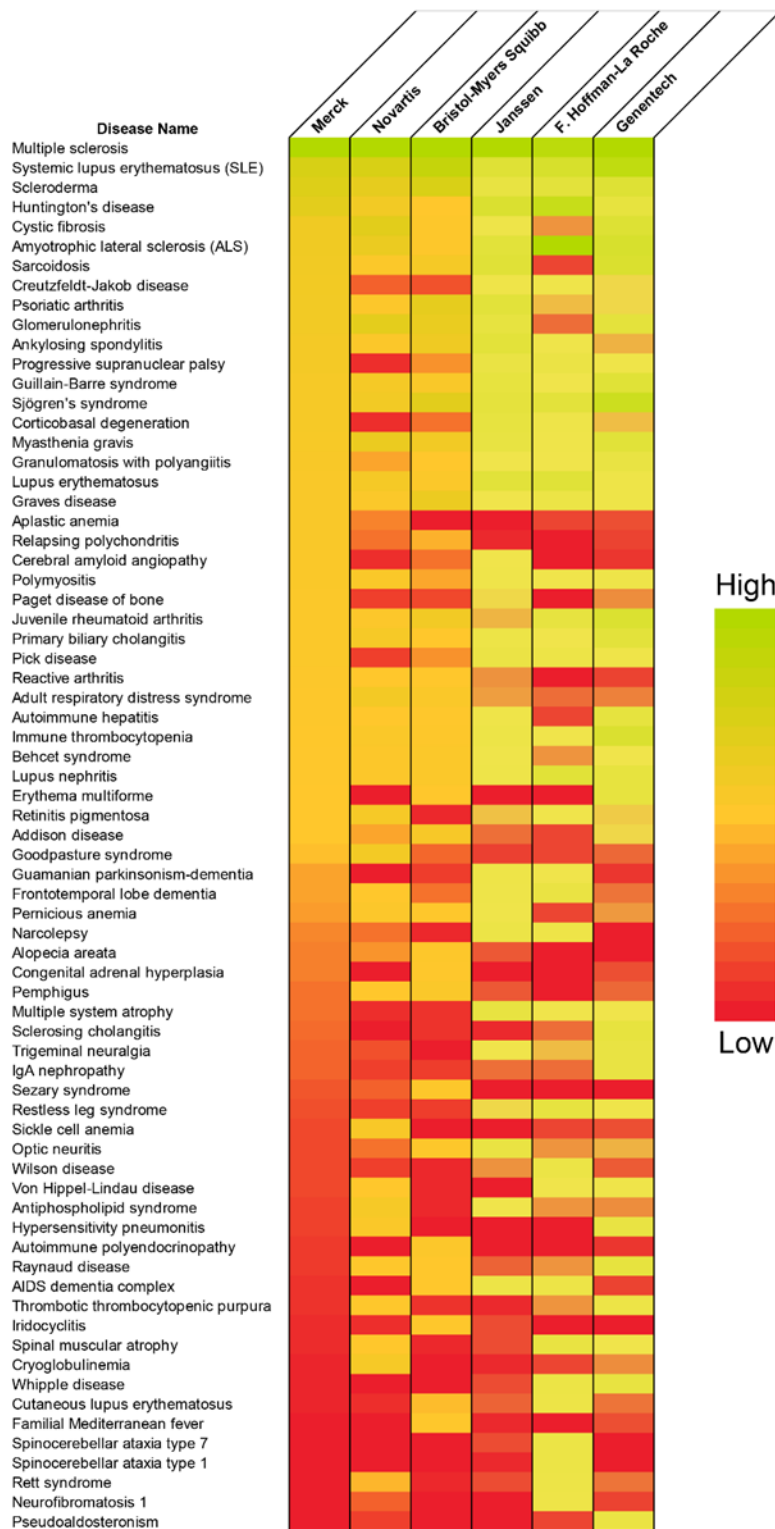
Among the non-commercial organizations, more than 65% of the leading organizations originate from the United States with the University of California leading overall (Figure 4A). The remaining consist of two research organizations each from South Korea (KOR) and China (CHN) with the L'Institut national de la santé et de la recherche médicale (INSERM) being the only French organization featuring in the top 15 non-commercial organizations. In recent years, these leading organizations appear to be involved in research across a variety of rare diseases such as SLE,<sup>31</sup> HD,<sup>31</sup> multiple sclerosis,<sup>62-64</sup> and the more obscure ones such as disabling pansclerotic morphea<sup>65</sup> and Von Hippel-Lindau disease.<sup>66</sup> Patents filed by researchers from the University of California in recent years (2015 onwards) cover a diverse group of ailments including developing small molecule inhibitors targeting various proteins and potentially useful in the treatment of ALS (WO2023086603,<sup>67</sup> WO2022104148<sup>68</sup>), using a SARS-Cov-2 pseudoviral delivery system for therapeutic transgene as potential treatment for HD (WO2022165538A1<sup>69</sup>), and development of GABA positive allosteric modulators (WO2018236955<sup>70</sup>) and peptide-based inhibitors for treatment of MG, (WO2018049053<sup>71</sup>).





**Figure 4.** Leading organizations in the field of rare diseases based on patent publication data from the CAS Content Collection for the period 2003 to 2023. **(A)** Donut and **(B)** bar charts showing geographical distribution and leading organizations, respectively. Entities were separated into non-commercial and commercial organizations. Bars colored to correspond to countries or regions shown in the donut charts. Countries or regions represented by their standard three letter codes – United States (USA), China (CHN), South Korea (KOR), France (FRA), Germany (DEU), Japan (JPN), Spain (ESP), Denmark (DEN), Italy (ITA), Belgium (BEL), Switzerland (CHE), and United Kingdom (GBR).

Next, guided by our identification of leading commercial patent assignees we analyzed in greater detail patents by the top 6 companies (Figure 4B) to understand the spread of commercial interest across different rare diseases and shown as heat maps in Figure 5 and Supporting Information Figure S1 based on CAS indexing. Rare diseases such as multiple sclerosis,<sup>72</sup> SLE,<sup>73</sup> scleroderma, and HD appears to have high commercial interest across the six companies we chose to focus on as seen by the higher number of patent publications. In contrast, examples of rare diseases with apparent low commercial interest include iridocyclitis, familial Mediterranean fever,<sup>74</sup> Rett syndrome<sup>75</sup> and pseudoaldosteronism<sup>76</sup> (Figure 5). Similar heat map for rare cancers indicates that among these, kidney cancer,<sup>77</sup> thyroid cancer,<sup>78</sup> melanoma<sup>79</sup> and multiple myeloma,<sup>80</sup> a type of hematological malignancy, appears to have number of patents filed by all six commercial companies. Examples of rare cancer that continue to be under explored commercially as exhibited by low patent publications include nasopharyngeal carcinoma,<sup>81</sup> blastic plasmacytoid dendritic cell cancer<sup>82</sup> and B-cell prolymphocytic leukemia<sup>83, 84</sup> among others (Supporting Information, Figure S1).



**Figure 5.** Heat map indicating number of patents filed by commercial organizations with respect to rare diseases. Patent assignees shown here are among the top 6 commercial patent assignees shown in Figure 4B. Data includes patent publications in the field of rare diseases from the CAS Content Collection for 2003-2023.

In our dataset we identified leading rare diseases and rare cancers based on the number of documents (journals and patents) associated with them using the robust data curation performed by CAS and are shown as a heat map in Figure 6 – ALS, HD, and MG all feature in the list. Other well-studied rare diseases include autoimmune diseases such as multiple sclerosis,<sup>72</sup> SLE,<sup>73</sup> scleroderma,<sup>85</sup> Sjögren's syndrome,<sup>86</sup> and inherited disorders such as cystic fibrosis<sup>87</sup> and sickle cell anemia.<sup>88</sup> Among the top 10 well-studied rare cancers, ~30% are hematological malignancies (or blood cancers) such as multiple myeloma,<sup>80</sup> non-Hodgkin's lymphoma,<sup>89</sup> and AML.<sup>90</sup> The list is also populated with rare cancers such as pheochromocytoma, type of neuroendocrine tumor<sup>91</sup>; cholangiocarcinoma, cancer of the bile duct,<sup>92</sup> and melanoma, a type of skin cancer.<sup>79</sup> Other examples of leading rare cancers include hepatocellular carcinoma,<sup>93</sup> a type of liver cancer, mesothelioma<sup>94</sup> which is cancer occurring in the tissue surrounding internal organs (mesothelium), and an aggressive form of brain cancer called glioblastoma.<sup>95</sup>

(A)

		Number of journal publications (2003-2023)	Number of patent publications (2003-2023)
<b>Non-cancer</b>	<b>Disease</b>		
	Multiple sclerosis	25511	22979
	Systemic lupus erythematosus (SLE)	22038	10584
	Amyotrophic lateral sclerosis (ALS)	12871	12042
	Cystic fibrosis	12213	6727
	Huntington's disease	8917	11484
	Scleroderma	6725	7567
	Sickle cell anemia	4816	2180
	Sjögren's syndrome	3157	5492
	Myasthenia gravis	2842	4716
Graft-versus-host reaction	2654	4503	

(B)

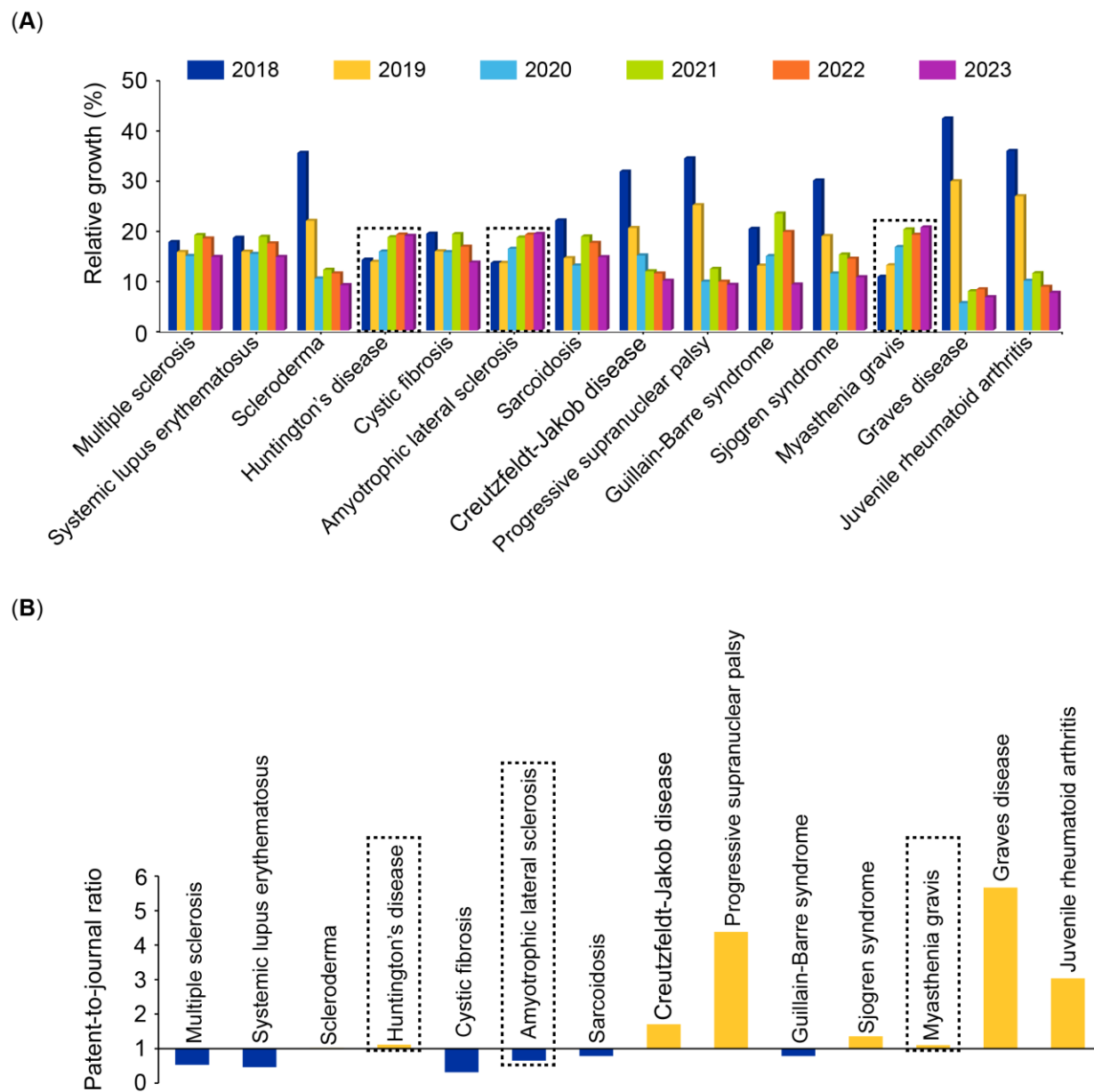
		Number of journal publications (2003-2023)	Number of patent publications (2003-2023)
<b>Cancer</b>	<b>Disease</b>		
	Multiple myeloma	23575	17303
	Non-Hodgkin's lymphoma	8510	8035
	Acute myeloid leukemia	5861	7534
	Hodgkin disease	5706	6809
	Pheochromocytoma	5667	1195
	Cholangiocarcinoma	5563	2309
	Melanoma	5224	15304
	Hepatocellular carcinoma	5110	6464
	Mesothelioma	4253	5326
Glioblastoma	3964	7564	

**Figure 6.** Heat map depicting number of journal and patent publications for leading rare diseases and rare cancers in our dataset of rare diseases. Data includes journal publications sourced/extracted from the CAS Content Collection for the period 2003-2023.

Having identified leading rare diseases and rare cancers in our dataset, we wanted to see how interest in these, and other rare diseases have played out over the last few years. To determine this, we plotted the relative increase in publications (journal and patent) for the period 2018-2022 normalized with respect to total publications for that period for each disease (Figure 7A). HD, ALS, and myasthenia gravis show a clear, steady, and consistent increase in publications over the 5-year period. This increase is perhaps most evident for myasthenia gravis with publications nearly doubling between 2018 and 2022. A few rare diseases such as scleroderma,<sup>85</sup> an autoimmune disease resulting in hardening of skin, Creutzfeld-Jakob disease,<sup>96</sup> an aggressive neurodegenerative disorder resulting in death and progressive supranuclear palsy,<sup>97</sup> neurodegenerative disorder affecting muscles and movement, show waning publications over the years. Sjögren's syndrome,<sup>86</sup> Graves' disease,<sup>98</sup> and juvenile rheumatoid arthritis<sup>99</sup> also show a downward trend in publications indicative of decrease in interest from the scientific community. Multiple sclerosis, SLE, sarcoidosis,<sup>100</sup> and Guillain-Barre syndrome,<sup>101</sup> all of which involve the body's immune system, show more or less consistent number of publications with minor fluctuations.

Calculating the patent-to-journal ratios for these diseases splits them more or less evenly – with 7 out of 14 analyzed rare diseases having a patent-to-journal ratio greater than 1, indicative of greater commercial interest. Out of the remaining 7, scleroderma has a patent-to-journal ratio of 1 while the rest six have a patent-to-journal ratio of less than 1 indicative of not as much commercial interest (Figure 7B). Out of the three rare diseases that we chose to focus on – HD and MG exhibit a patent-to-journal ratio greater than 1 by a modest extent. On the other hand, ALS has a patent-to-journal ratio of 0.7 indicating greater interest from the scientific community that is yet to translate to commercial prospects – this might also be attributable to the increased interest in ALS following the viral ice bucket phenomenon translating as increased journal publications as a result of increased funding. Among the 14 rare diseases analyzed, progressive supranuclear palsy and Graves disease lead in terms of having more than 4 times as many patents as journal publications (Figure 7B).

Similar analysis with respect to rare cancers reveals increase in publications across the board with all selected rare cancers showing clear, consistent, and rapid increase in publications (Supporting Information, Figure S2A). Of special note are Kaposi's sarcoma,<sup>102</sup> cancer affecting the lining of blood vessels and lymph nodes, glioblastoma,<sup>95</sup> cancer of the brain and/or spinal cord, and thyroid cancer<sup>78</sup> showing the greatest increase with publications more than doubling between 2019 and 2022 (Supporting Information, Figure S2A). The patent-to-journal ratios indicate seemingly high commercial interest for the selected/analyzed rare cancers with >70% exhibiting patent-to-journal ratios greater than 1 (Supporting Information, Figure S2B). The exceptions to this are multiple myeloma, esophagus cancer<sup>103</sup> and hepatocellular carcinoma,<sup>93</sup> a type of liver cancer, with low patent-to-journal ratios (Supporting Information, Figure S2B).



**Figure 7.** (A) Number of publications (journal and patent) and (B) patent-to-journal ratios for selected leading rare diseases in our dataset. Highlighted in dashed black boxes are the three rare diseases that we analyzed in detail. Data includes patent and journal publications sourced/extracted from the CAS Content Collection for the period 2018-2023 in the field of rare diseases.

### 3. Amyotrophic lateral sclerosis (ALS), Huntington's disease, myasthenia gravis

#### Amyotrophic Lateral Sclerosis

##### Overview

Amyotrophic lateral sclerosis (ALS), also known as motor neuron disease or Lou Gehrig's disease, is a rare progressive neurodegenerative disorder affecting the nerve cells in the brain and spinal cord that control voluntary muscle movement.<sup>104-109</sup> Global estimates of ALS range from 1.9 per 100,000 to 6 per 100,000.<sup>110-112</sup> ALS shot to public attention because of the viral “ice bucket challenge” in 2014.<sup>113</sup> Though the exact impact of the viral challenge remains unknown<sup>114-116</sup> with opinions divided on whether the challenge truly had a real world impact, it is undeniable that it catapulted the disease to the public's attention. Individuals well-known in their respective fields have been afflicted by this disorder, perhaps one of the most famous ones being the theoretical physicist Stephen Hawking.<sup>117</sup>

ALS leads to the gradual degeneration and death of motor neurons, which are the nerve cells responsible for transmitting signals from the brain to the muscles.<sup>118, 119</sup> As motor neurons deteriorate, voluntary muscle control and movement are progressively impaired, leading to muscle weakness, twitching (fasciculations), stiffness, and eventually paralysis. The symptoms of ALS can vary widely among individuals and may initially affect different muscle groups. Common early symptoms include weakness in the hands, arms, legs, or muscles involved in speech, swallowing, or breathing. As the disease progresses, muscle weakness and atrophy spread to other parts of the body.<sup>107</sup> ALS can also affect the muscles involved in speech, swallowing, and breathing, leading to difficulties in speaking, chewing, swallowing (dysphagia), and breathing (respiratory compromise). Bulbar symptoms often contribute to significant disability and can pose life-threatening complications in advanced stages of the disease. In some cases, ALS may be associated with cognitive and behavioral changes, including difficulties with executive function, language, decision-making, and emotional regulation. This constellation of symptoms is referred to as frontotemporal dementia (FTD) or ALS-FTD, which represents a spectrum of overlapping neurodegenerative disorders.<sup>118, 120-122</sup>

ALS currently has no cure, and the underlying mechanisms driving the disease remain incompletely understood. Most cases of ALS are sporadic, meaning they occur without a clear family history, while a smaller percentage are inherited (familial ALS) due to genetic mutations. Several genes have been implicated in familial ALS, including C9orf72, SOD1, TARDBP, FUS, and others.<sup>123-125</sup>

Diagnosis of ALS is typically based on a thorough medical history, neurological examination, electromyography (EMG), nerve conduction studies, and exclusion of other possible causes of muscle weakness and motor dysfunction. Magnetic resonance imaging (MRI) and other imaging tests may also be used to rule out other conditions.<sup>126, 127</sup>

While there is no cure for ALS, various treatments and supportive interventions can help manage symptoms, improve quality of life, and prolong survival. These may include medications to alleviate muscle cramps, spasticity, or excessive saliva production, as well as physical therapy, occupational therapy, speech therapy, and assistive devices to maintain mobility and communication abilities.<sup>128</sup> In some cases, non-invasive ventilation or feeding tubes may be

necessary to support respiratory and nutritional needs. The US Food and Drug Administration (FDA) has approved a few medications for treating symptoms related to ALS<sup>129, 130</sup> including the benzothiazole Riluzole (Rilutek, Exservan, Tiglutik)<sup>131, 132</sup> in 1995, the antioxidant Edaravone (Radicava)<sup>133</sup> in 2017, and Sodium phenylbutyrate-Taurursodiol (Relyvrio)<sup>134, 135</sup>, a combination of a histone deacetylase inhibitor and a bile acid, in 2022. Sadly, Relyvrio has been just reported to have failed Phase III trial.<sup>136</sup> The Phase 3b trial of Radicava has been also discontinued in 2023, as well as its extension study.<sup>137, 138</sup> The prognosis for ALS varies, with most individuals experiencing progressive disability over time. The rate of disease progression and life expectancy can vary widely among individuals, ranging from a few years to more than a decade from the onset of symptoms. Respiratory failure is the most common cause of death in ALS.<sup>139-141</sup>

### *Pathogenesis of amyotrophic lateral sclerosis*

The pathogenesis of ALS is complex and involves a combination of genetic, environmental, and cellular factors. While the exact cause of ALS remains incompletely understood, several key mechanisms have been implicated in the degeneration of motor neurons characteristic of the disease.<sup>122, 123, 142-145</sup>

ALS primarily affects motor neurons, which are the nerve cells responsible for transmitting signals from the brain and spinal cord to the muscles, controlling voluntary muscle movement. The progressive degeneration and death of motor neurons lead to muscle weakness, atrophy, and eventual paralysis.<sup>146, 147</sup> Glutamate is the primary excitatory neurotransmitter in the CNS, including the motor neurons. In ALS, dysregulation of glutamate signaling occurs, leading to excessive levels of glutamate in the synaptic cleft. This excitotoxicity can damage motor neurons, contributing to their degeneration and death.<sup>148-150</sup> Motor neurons are particularly vulnerable to oxidative stress, which occurs when there is an imbalance between the production of reactive oxygen species (ROS) and the body's ability to neutralize them with antioxidants. Oxidative stress can damage cellular structures, including proteins, lipids, and DNA, leading to motor neuron dysfunction and death.<sup>151, 152</sup> Abnormal protein aggregation is a hallmark feature of ALS pathology. Mutations in genes such as SOD1, TARDBP (encoding TDP-43), FUS, and C9orf72 can lead to the misfolding and aggregation of proteins within motor neurons and surrounding cells. These protein aggregates can disrupt cellular function and contribute to neurodegeneration.<sup>153-156</sup>

Mitochondria are the cellular organelles responsible for energy production and play a crucial role in maintaining neuronal function and viability. Dysfunction of mitochondria, including impaired energy metabolism and increased production of ROS, has been implicated in ALS pathogenesis, contributing to motor neuron degeneration.<sup>157-159</sup> In addition to motor neurons, non-neuronal cells such as astrocytes, microglia, and oligodendrocytes play important roles in ALS pathogenesis. Dysfunctional glial cells can contribute to neuroinflammation, excitotoxicity, and impaired neuronal support and repair mechanisms, further exacerbating motor neuron degeneration.<sup>160-163</sup> While most cases of ALS are sporadic, meaning they occur without a clear family history, approximately 5-10% of cases are familial and are associated with specific genetic mutations. Mutations in genes such as SOD1, TARDBP, FUS, and C9orf72 have been identified in familial ALS and can contribute to motor neuron dysfunction through various mechanisms.<sup>164, 165</sup> While the majority of ALS cases have no known cause, environmental factors such as exposure to toxins, heavy metals, pesticides, and traumatic brain injury have been suggested as potential risk factors for ALS. These factors may interact with genetic susceptibility to contribute to disease onset and progression.<sup>166-168</sup> Thus, ALS is likely a multifactorial disease involving

interactions between genetic susceptibility, environmental factors, and various cellular mechanisms leading to motor neuron degeneration and progressive muscle weakness and paralysis. The high heterogeneity of ALS and the failures of the efforts to find a singular cure have led some researchers to propose that it may not be a single disease, but rather a miscellany of overlapping conditions that share common characteristics, similarly to cancer.<sup>169, 170</sup> Further research is needed to fully elucidate the complex pathogenesis of ALS and identify effective therapeutic strategies for the treatment of this devastating neurodegenerative disorder.

### *Genetic background of amyotrophic lateral sclerosis*

The genetic background of ALS is multifaceted, involving both familial and sporadic forms of the disease. While the majority of ALS cases are sporadic, meaning they occur without a clear family history, approximately 5-10% of cases are familial and have a known genetic component.<sup>124, 125, 171-174</sup>

Familial ALS (fALS) accounts for a small percentage of ALS cases and is characterized by a clear family history of the disease. In these cases, ALS is inherited in an autosomal dominant manner, meaning that a mutation in a single copy of a specific gene is sufficient to cause the disease.<sup>175</sup> Several genes have been implicated in familial ALS, including:

- SOD1 (Superoxide Dismutase 1): Mutations in the SOD1 gene were the first identified genetic cause of ALS. SOD1 encodes an enzyme involved in antioxidant defense, and mutations in this gene can lead to protein misfolding and aggregation, mitochondrial dysfunction, and motor neuron degeneration.<sup>176-178</sup>

- C9orf72 (Chromosome 9 Open Reading Frame 72): Expansion of a hexanucleotide repeat sequence (GGGGCC) within the C9orf72 gene is the most common genetic cause of ALS and frontotemporal dementia (FTD). The exact mechanism by which the repeat expansion leads to neurodegeneration is not fully understood but likely involves RNA toxicity, protein aggregation, and impaired nucleocytoplasmic transport.<sup>123, 179-182</sup>

- TARDBP (TAR DNA-Binding Protein): Mutations in the TARDBP gene, which encodes the TDP-43 protein involved in RNA processing and metabolism, have been identified in familial ALS cases. Abnormal accumulation of TDP-43 protein aggregates is a pathological hallmark of ALS.<sup>183-185</sup>

- FUS (Fused in Sarcoma): Mutations in the FUS gene, which encodes a DNA/RNA-binding protein involved in RNA processing and transport, have also been implicated in familial ALS. Like TDP-43, abnormal accumulation of FUS protein aggregates is observed in ALS.<sup>186-188</sup>

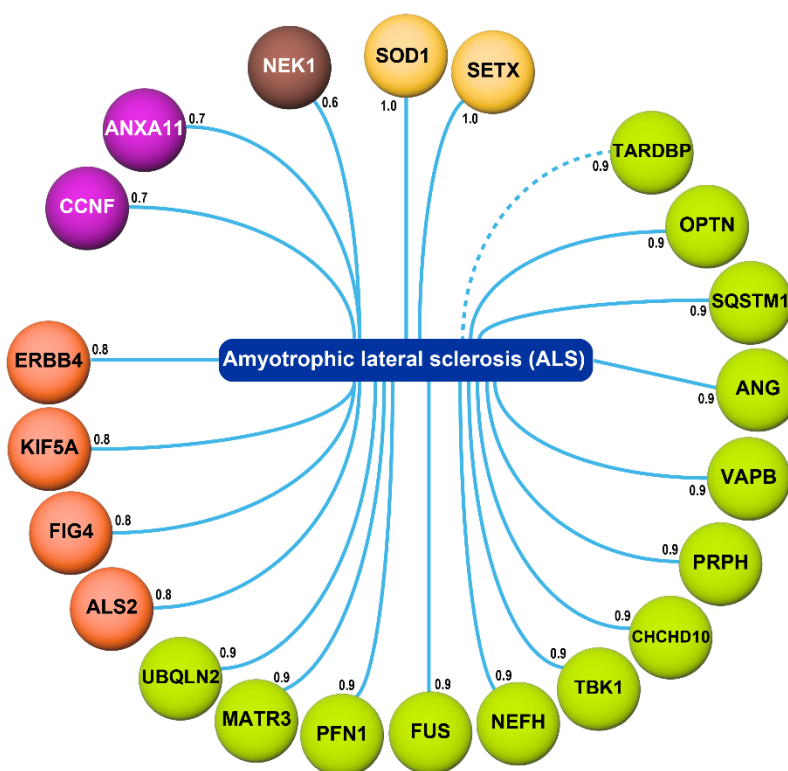
- Other Genes: Mutations in other genes, such as VCP (valosin-containing protein), OPTN (optineurin) and SQSTM1 (sequestosome 1) have been identified in rare familial ALS cases, although their contribution to disease pathogenesis is less well understood.<sup>189-192</sup>

Sporadic ALS (sALS) occurs in individuals with no family history of the disease and is thought to result from a combination of genetic susceptibility and environmental factors. While specific genetic mutations are less common in sALS compared to fALS cases, genome-wide association studies (GWAS) have identified common genetic variants associated with an increased risk of developing ALS. These variants are often found in genes involved in neuronal function, inflammation, and other biological pathways implicated in ALS pathogenesis.<sup>193, 194</sup>



Genes associated with ALS based on data from the CAS Content Collection are explored in Figure 8 to facilitate the identification of potential therapeutic targets and biological mechanisms underlying disease processes. Details about each of the identified genes can be found in Table 1. For a detailed methodology please refer to the Supporting Information.

ALS exhibits significant genetic heterogeneity, with different genetic mutations associated with distinct clinical phenotypes and disease progression. Additionally, genetic modifiers and environmental factors may influence the penetrance and expressivity of ALS-associated mutations, leading to variability in disease onset, severity, and progression among affected individuals.<sup>124, 195, 196</sup> Understanding the genetic basis of ALS is critical for elucidating disease mechanisms, identifying potential therapeutic targets, and developing personalized treatment approaches for individuals with ALS. Ongoing research efforts continue to uncover novel genetic risk factors and pathways underlying ALS pathogenesis, with the ultimate goal of improving outcomes for patients with this devastating neurodegenerative disorder.



**Figure 8.** Genes associated with amyotrophic lateral sclerosis (ALS) based on data from the CAS Content Collection. Only genes with an association score of greater than 0.6 and with at least 10 records are shown here. Color corresponds to association score – yellow (1.0), green (0.9), orange (0.8), purple (0.7), and brown (0.6). The nature of the line indicates association source with dashed lines indicating a majority of records resulting from text mining.

**Table 1.** List of genes associated with amyotrophic lateral sclerosis (ALS), as shown in Figure 8, with the association type as causal or contributing.

Gene	Association score	Protein	Expression profile, function and other diseases	Amyotrophic lateral sclerosis (ALS)
SOD1 <sup>197</sup>	1.0	Superoxide dismutase 1	An enzyme that scavenges superoxide radicals, superoxide dismutase 1 is crucial in converting ROS to molecular oxygen (O <sub>2</sub> ) and hydrogen peroxide (H <sub>2</sub> O <sub>2</sub> ). <sup>198, 199</sup> Also referred to as copper-zinc superoxide dismutase (Cu/Zn SOD1, CuZn SOD1) due to the presence of a copper and Zn ion in the enzyme. <sup>199</sup> Superoxide dismutase are expressed in the neurons of CNS and are linked to neurological disorders. <sup>200</sup>	Mutations in superoxide dismutase 1 are thought to result in protein misfolding leading to ALS. <sup>201, 202</sup> In silico studies have tried to decipher changes in structure of mutant superoxide dismutase 1 resulting in “pathogenic conformational states” that might promote aggregation. <sup>203</sup> Researchers have also found a disulfide bond (C57-C146) to be critical for the normal functionality of superoxide dismutase 1. <sup>204, 205</sup> Research continues to be conducted to understand the exact effect of a mutation on protein structure using molecular modeling and simulation studies. <sup>178</sup>
SETX <sup>206</sup>	1.0	Senataxin	An enzyme classified as a superfamily 1B helicase, senataxin plays a critical role in resolving R-loops acting as a RNA:DNA helicase during DNA damage repair. <sup>207</sup> Mutations in SETX have been found in ataxia with oculomotor apraxia (AOA2). <sup>208-211</sup>	Mutations in senataxin have been linked to ALS4 (juvenile amyotrophic lateral sclerosis-4 with onset occurring before the age of 25 <sup>212</sup> ), <sup>213, 214</sup> These mutations are thought to interfere with senataxin’s ability to regulate RNA polymerase II, control R loops, and binding to other protein partners among other things. <sup>215, 216</sup>
TARDBP <sup>217</sup>	0.9	Transactive response (TAR) DNA-binding Protein 43 kDa (TDP-43)	Protein composed of ~400 amino acid belonging to the heterogeneous nuclear ribonucleoprotein family. <sup>218</sup> TDP-43 is largely composed of 4 domains <sup>219</sup> and plays a role in RNA regulation. <sup>218</sup> Primarily a nuclear protein, TDP-43 shuttles between the nucleus and cytoplasm. <sup>220</sup> TDP-43 has been implicated in neurological disorders such as Alzheimer’s, <sup>221-224</sup> Parkinson’s <sup>225</sup> disease,	Mutations in TDP-43 have been observed in ALS. <sup>229-232</sup> These mutations can lead to formation of amyloid-like fibrils or cause TDP-43 to localize in the cytoplasm instead of the nucleus affecting RNA regulation. <sup>220, 233</sup> Recently researchers have determined that the aggregated forms of TDP-43 exhibit distinct folds depending on the type of disease (ALS <sup>234</sup> vs. type B FTLD-TDP

			as well as frontotemporal dementia (FTD). <sup>226-228</sup>	<sup>235</sup> ). A functional dimer, monomerization of TDP-43 has been shown to be important in ALS. <sup>236</sup>
OPTN <sup>237</sup>	0.9	Optineurin	A ~570 amino acid cytosolic protein that is expressed in a variety of tissues including the brain, heart, skeletal muscle, and eye among others. <sup>238</sup> Optineurin is involved in vesicular trafficking <sup>239</sup> and immune signaling. <sup>240</sup> Optineurin aggregates have been reported in Parkinson's disease <sup>241</sup> and its expression altered in a Parkinson's disease model. <sup>242</sup> Optineurin has a possible role in promoting aggregation of mutant huntingtin. <sup>243</sup> On the other hand, optineurin might have a beneficial role in Alzheimer's disease. <sup>244</sup>	Mutations in optineurin have been reported in ALS <sup>245-250</sup> with mutations affecting optineurin's ability to interact with mitochondria and affecting mitophagy. <sup>238, 251</sup> OPTN gene therapy is being explored as possible treatment option for ALS. <sup>252</sup>
SQSTM1 <sup>253</sup>	0.9	Sequestosome 1	Also referred to as p62, sequestosome 1 is a selective autophagy receptor. <sup>254</sup> Composed of multiple protein-protein interaction motifs, sequestosome 1 functions to eliminate misfolded and aggregated protein. <sup>255</sup> In addition, sequestosome 1 is important in aging and age-related diseases. <sup>254</sup> Besides ALS, SQSTM1 has been implicated in FTD <sup>256-259</sup> and Alzheimer's disease <sup>260</sup> among other neurological disorders.	Similar to the other genes listed in this table, mutations in sequestosome 1 have been identified in individuals suffering from ALS. <sup>191, 192, 261, 262</sup> For some mutations in sequestosome 1 its ability to bind to other protein binding partners is impaired leading to reduced autophagy. <sup>263</sup> Therefore, compounds enhancing autophagy may be a potential treatment strategy. <sup>264</sup>
ANG <sup>265</sup>	0.9	Angiogenin	As the name suggests, angiogenin is a small (~120 amino acid residues) protein that is crucial in angiogenesis (the formation of new blood vessels). <sup>266, 267</sup> Besides angiogenesis, other processes involving angiogenin include rRNA and mRNA transcription, tRNA metabolism <sup>268</sup> as well as playing a role in immune response. <sup>269</sup> Angiogenin belongs to the ribonuclease A superfamily.	Mutations in ANG, often loss-of-function, have been reported in ALS. <sup>270, 271</sup> Research is ongoing to determine the exact effect of reported/known ALS mutations on the structure and ribonuclease A activity of angiogenin. <sup>272, 273</sup> In a recent report, the role of angiogenin in ALS was shown to be more complex than was earlier thought. <sup>274</sup>
VAPB <sup>275</sup>	0.9	Vesicle-associated membrane protein (VAMP)-associated protein B and C (VAPB)	VAPB is a type IV membrane protein consisting of ~243 amino acids associated with the endoplasmic reticulum. <sup>276</sup> Known to interact with a wide variety of partners <sup>277</sup>	Mutations in VAPB have been detected in ALS patients. <sup>281, 282</sup> One mutation in particular appears to have been studied extensively in the context of ALS – a

			often (but not always) via interaction between its MSP domain and a FFAT motif of the binding partner. <sup>278</sup> VAPB has been implicated in a number of neurological disorders including Alzheimer's and Parkinson's disease. <sup>277</sup> In addition, the role of VAPB in cancer is being actively studied. <sup>279, 280</sup>	Pro-to-Ser (P56S) mutation. <sup>283-290</sup> The P56S mutation appears to a toxic gain-of-function mutation and number of mechanisms have been proposed including affecting autophagy, <sup>290</sup> causing endoplasmic reticulum stress, <sup>287, 289</sup> changing the morphology of endoplasmic reticulum, <sup>291</sup> and diminishing VAPB's ability to mediate unfolded protein response. <sup>283</sup> Other mutations perhaps less well studied include P56H, <sup>292</sup> T46I, <sup>293</sup> V234I, <sup>294</sup> and a deletion mutant ( $\Delta$ S160). <sup>295</sup>
PRPH <sup>296</sup>	0.9	Peripherin	A type III intermediate filament protein, which are part of the cytoskeleton (other members include desmin, vimentin and glial fibrillary acidic protein). <sup>297</sup> Unlike other type III intermediate filament proteins, peripherin is expressed primarily in the peripheral and CNS. <sup>297, 298</sup> Peripherin is thought to play a role in axonal growth. <sup>299, 300</sup> Researchers are exploring peripherin as potential biomarker for motor neuron diseases <sup>301</sup> and axonal damage. <sup>302</sup>	Upregulation of peripherin has been reported in ALS <sup>303</sup> with this upregulation proposed to have a detrimental effect as observed in mouse models. <sup>304</sup> Others have reported mutations in peripherin in ALS patients. <sup>305</sup> Peripherin splice variants prone to aggregation have also been reported. <sup>303, 304</sup>
CHCHD10 <sup>306</sup>	0.9	Coiled-coil-helix-coiled-coil-helix domain containing protein 10	A small protein (~149 amino acid residues) associated with the intermembrane space of mitochondria, <sup>307</sup> coiled-coil-helix-coiled-coil-helix domain containing protein 10 is also known as protein N27C7-4. <sup>308</sup> Like other members of the CHCHD-containing protein family, CHCH10 protein contain (CX <sub>9</sub> C) <sub>2</sub> motifs. <sup>307</sup> Appears to play a role in mitochondrial structure and function. <sup>307</sup> Other diseases associated with CHCHD10 include late onset spinal motor neuronopathy <sup>309</sup> and mitochondrial myopathy. <sup>310</sup>	Mutations in CHCHD10 have been reported in ALS patients. <sup>311-314</sup> Overexpression of a mutant CHCHD10 was associated with abnormalities in mitochondria <sup>312</sup> and could underlie pathogenesis of ALS and FTD.
TBK1 <sup>315</sup>	0.9	TANK binding kinase 1	A serine/threonine protein kinase composed of ~729 amino acid residues, TBK1 plays a role in innate immunity, autophagy and	Among the many binding partners of TBK1 are optineurin and sequestosome 1/p62. Mutations in TBK1 have been

			apoptosis among other cellular functions. <sup>316</sup> TBK1 binds to and interacts with a wide variety of protein partners utilizing different domains present in its structure. <sup>316-318</sup> Activation of TBK1 requires autophosphorylation at Ser172. <sup>317, 319</sup> Diseases that involve TBK1 include cancer <sup>316, 320, 321</sup> and autoimmune disorders <sup>322, 323</sup> with TBK1 inhibitors being explored as a viable strategy. <sup>322, 324, 325</sup>	reported in ALS <sup>326-328</sup> with mutations thought to interfere with autophagic function of TBK1 <sup>329</sup> by affecting kinase activity. <sup>330</sup> Besides ALS, TBK1 mutations are also implicated in FTD. <sup>331, 332</sup>
NEFH <sup>333</sup>	0.9	Neurofilament heavy chain	Neurofilaments are heavily expressed in the central and peripheral nervous system. <sup>334</sup> They are made up of three subunits based on molecular weight, light, medium and heavy (NEFH). <sup>335</sup> They are also involved with cytoskeletal functions such as the regulation of axon diameter and growth. <sup>336</sup> Besides ALS, the other disease associated with NEFH includes Charcot-Marie-Tooth disease. <sup>337-339</sup>	Mutations in NEFH proteins have been reported to be associated with ALS such as those within the KSP repeat region of the tail domain. <sup>340-342</sup> Another study reports variants in NEFH associated with sALS such as the NEFH Ser787Arg. <sup>343</sup>
FUS <sup>344</sup>	0.9	Fused in sarcoma (FUS) RNA binding protein	A DNA/RNA binding protein, FUS is composed of ~506 amino acids. <sup>345</sup> FUS is expressed both in the nucleus and cytoplasm <sup>345</sup> and participates in a number of important cellular processes such as DNA repair <sup>346</sup> including mitochondrial DNA repair <sup>347</sup> and mRNA transport, <sup>347</sup> among others. Some other names that FUS is known by include translocated in liposarcoma (TLS) and heterogeneous nuclear ribonucleoprotein P2 (HNRNPP2). Besides ALS, FUS has also been linked to FTD. <sup>348, 349</sup>	Mutations in the FUS gene have been linked to ALS over the last two decades. <sup>187, 350-356</sup> Some ALS-associated mutations have been found in the nuclear localization signal domain dampening the ability of FUS to bind transportin 1, a nuclear import receptor, <sup>357</sup> resulting in accumulation of mutant FUS in the cytoplasm. <sup>358</sup> An article published in 2018 describes FUS R495X ALS-associated mutation resulting in alterations in the mitochondria. <sup>359</sup>
PFN1 <sup>360</sup>	0.9	Profilin 1	Profilin 1 is a small actin binding protein composed of ~139 amino acid residues with a wide expression profile. <sup>361</sup> Profilin 1 helps in actin polymerization <sup>362</sup> by accelerating nucleotide exchange or by lowering actin concentration required to jump start polymerization. <sup>361</sup> Besides actin	Mutations in profilin have been observed in ALS <sup>370-372</sup> – aggregation of mutant profilin 1 often alongside TDP-43 is thought to be a contributing factor to ALS. Mutation in the phosphorylation site of profilin 1, important for actin polymerization, has also been

			polymerization, profilin 1 appears to be important for a number of cellular processes including cell division <sup>363</sup> and membrane trafficking. <sup>364, 365</sup> In the last decade, profilin 1's role in cancer has investigated <sup>366</sup> with dysregulation of profilin 1 observed in breast cancer, <sup>367</sup> endometrial cancer, <sup>368</sup> as well as non-small cell lung cancer, <sup>369</sup> among others.	reported. <sup>373</sup> Molecular modeling and dynamic studies indicate that some mutations in profilin1 might affect the stability and functionality of the protein. <sup>374</sup>
MATR3 <sup>375</sup>	0.9	Matrin 3	A DNA/RNA binding nuclear protein, matrin 3 is composed of ~847 amino acid residues <sup>376</sup> organized into several domains such as RNA recognition motifs, zinc finger domain, and intrinsically disordered regions. <sup>377, 378</sup> The various functions that matrin 3 is involved in includes mRNA stabilization, <sup>379</sup> alternative splicing regulation, <sup>380</sup> DNA repair, <sup>381</sup> etc. <sup>378</sup>	Matrin 3 binds to and interacts with a number of different proteins including TDP-43 and FUS, which are also implicated in ALS. <sup>378</sup> Mutations in MATR3 has been detected in ALS patients. <sup>382-386</sup>
UBQLN2 <sup>387</sup>	0.9	Ubiquilin 2	A ubiquitin-like protein, ubiquilin 2 is an important component of the ubiquitin-proteasome system helping in the degradation of misfolded proteins. <sup>388</sup> The ubiquitin-like domain and ubiquitin-associated domains of ubiquilin 2 interact with the proteasome and the polyubiquitin chains on proteins, respectively. <sup>389</sup>	Mutations in UBQLN2 linked to ALS <sup>390-392</sup> possibly by hampering protein degradation by affecting its interaction with binding partners/other proteins. <sup>393, 394</sup>
ALS2 <sup>395</sup>	0.8	Alsin	Composed of ~1657 amino acids arranged into four domains, alsin is a large protein that acts as a guanine nucleotide exchange factor for Rac1 and Rac5 <sup>396</sup> and plays a role in membrane trafficking. <sup>396, 397</sup> Alsin expression has been observed in the CNS in mice <sup>398</sup> and studies with alsin knockdown and Rac1 mutants indicate that alsin-Rac1 signaling is important for axonal growth in motor neurons. <sup>399</sup>	Mutations in alsin have been detected in ALS patients. <sup>400-403</sup> Some ALS-associated mutations result in expression of short and long form of alsin lacking crucial domains, diminishing alsin's ability to effectively function as a guanine nucleotide exchange factor. <sup>400</sup>
FIG4 <sup>404</sup>	0.8	Polyphosphoinositide phosphatase	Also known as SAC domain-containing protein 3 (Sac3) and is composed of ~907 amino acid residues. <sup>405, 406</sup> An enzyme, polyphosphoinositide phosphatase cleaves the phosphatase group from the 5-position of the inositol ring of phosphatidylinositol 3,5-	FIG4 mutations have been observed in ALS patients. <sup>412-418</sup> While the exact mechanism underlying ALS-associated disease mutations remains under investigation, evidence from FIG4's involvement in Charcot-Marie-Tooth

			bisphosphate (PI(3,5)P <sub>2</sub> ). <sup>406, 407</sup> PI(3,5)P <sub>2</sub> is a lipid signaling molecule and thought to be important for development of the nervous system. <sup>408</sup> Mutation in FIG4 and deficiency of FIG4 are associated with Charcot-Marie-Tooth disease, <sup>409</sup> an inherited genetic condition affecting peripheral nerves, <sup>410</sup> and lysosomal storage disorder. <sup>411</sup>	disease indicates disruption of intracellular trafficking due to excessive vacuoles could be a possible reason. <sup>419</sup>
KIF5A <sup>420</sup>	0.8	Kinesin family member 5A	Kinesin family member 5A belongs to the kinesin family of ATP-driven motor proteins. <sup>421</sup> Also referred to as kinesin-1, they are expressed in neurons <sup>422</sup> and are engaged in intracellular transport of protein complexes, mRNAs and others <sup>423</sup> as well as cell division. <sup>424</sup> Composed of ~1039 amino acid residues, kinesin family member 5A appears to be involved in trafficking of GABA receptors. <sup>425</sup> Under physiological conditions, kinesin family member 5A are autoinhibited <sup>426</sup> rendering them incapable of interacting with cargo and only upon binding of two partners – c-Jun N-terminal kinase–interacting protein 1 (JIP1) and fasciculation and elongation protein ζ1 (FEZ1) – is the autoinhibition lifted allowing kinesin family member 5A to carry out transport. <sup>427</sup> Besides ALS, expression levels of KIF5A appear to be altered in Alzheimer's disease <sup>428, 429</sup> and mutations in KIF5A appear to be associated with spastic paraplegia. <sup>430-432</sup>	In the last few years, there have been reports of KIF5A mutations observed in ALS patients. <sup>385, 433-438</sup> Recently, Baron et al. <sup>439</sup> elucidated the mechanism underlying an ALS-associated mutations in KIF5A – as per their explanation, the mutation rendered the autoinhibition of kinesin family member 5A ineffective resulting in increased mitochondrial transport. A research group from Tohoku university showed that an ALS-associated mutant of KIF5A had increased propensity to form aggregates. <sup>440</sup> Other ALS-associated mutations in KIF5A have also been identified. <sup>441</sup>
ERBB4 <sup>442</sup>	0.8	Erb-b2 receptor tyrosine kinase 4	A member of the epidermal growth factor receptor family, ErbB4 is a type of receptor tyrosine kinase. <sup>443</sup> Also referred to as HER4, ErbB4 is classified as a type I single transmembrane protein and is composed of ~1300 amino acid residues with a large extracellular and intracellular domain. <sup>444</sup> ErbB4 has been implicated in various types of cancer including breast, lung, and colorectal cancer, among others. <sup>445, 446</sup>	Mutations in ERBB4 have been reported in ALS patients. <sup>447-450</sup> ErbB4 binds to and interacts with a number of partners, one of which is neuregulin, small cell signaling protein molecules, <sup>451</sup> and undergo dimerization accompanied by autophosphorylation. <sup>452</sup> Mutations associated with ALS in the tyrosine kinase or C terminal domain of ErbB4

				was accompanied by reduced autophosphorylation. <sup>448</sup>
CCNF <sup>453</sup>	0.7	Cyclin F <sup>453</sup>	Cyclins are proteins that regulate the activity of cyclin-dependent protein kinases (CDK). <sup>454</sup> Described first in 1994 by Bai et al, <sup>454</sup> cyclin F is composed of ~786 amino acid residues and is structurally similar to other cyclins (especially A and B) with a wide distribution profile. Cyclin F is a part of the Skp1–Cul1–F-box (SCF) E3 ubiquitin ligase complex and is involved in ubiquitination of proteins <sup>455</sup> with ubiquitinated protein eventually being degraded by the ubiquitin-proteasome system. Cyclin F has been implicated in cancer. <sup>456-458</sup>	Similar to other proteins important for autophagy such as TDP-43 and p62/sequestosome 1, mutations in CCNF has been observed in and linked to ALS. <sup>385, 459-462</sup> Mutant CCNF results in abnormal ubiquitination, and this is thought to play a role in ALS. <sup>459, 460</sup> Another mechanism/pathway that may be important in ALS involves mutant cyclin F increasing the ATPase activity of valosin-containing protein eventually resulting in TDP-43 aggregation. <sup>463</sup>
ANXA11 <sup>464</sup>	0.7	Annexin A11	Annexin A11 is a ~504 amino acid residue long Ca <sup>2+</sup> -dependent membrane protein that is wide expressed. <sup>465</sup> Annexin A11 is thought to be important for vesicle trafficking by aiding in stabilization of Sec31A, a component of the coat protein complex II. <sup>466</sup> In 2023, the full-length structure of annexin A11 was published <sup>467</sup> and is bound to help in understanding how disease causing mutations affect the protein. Annexin A11 appears to play a role in sarcoidosis, an inflammatory disorder. <sup>465, 468</sup> Another disease with possible annexin A11 involvement is immunoglobulin G4-related disease. <sup>469, 470</sup>	As with other genes noted here, mutations in ANXA11 have been reported in the context of ALS. <sup>471-475</sup> The various mechanisms by which mutant annexin A11 results in ALS included formation of insoluble aggregates, disruption of Ca <sup>2+</sup> homeostasis, <sup>195, 476</sup> and interference with RNA transport. <sup>477</sup> Structural studies of annexin A11 including crystal structure <sup>478</sup> have been reported that suggest possible effects of mutation on the structure and functionality of annexin A11. <sup>479</sup>
NEK1 <sup>480</sup>	0.6	NIMA related kinase 1	The never in mitosis A (NIMA)-related kinase 1 is a serine/threonine kinase. <sup>480</sup> It is highly expressed in neuronal and germ cells <sup>481, 482</sup> and plays a role in several cellular processes such as cell cycle regulation, cell death, cilia formation, and DNA damage response. <sup>483, 484</sup> NEK1 mutations appear to be associated with Mohr syndrome, <sup>485</sup> short rib-polydactyly syndrome, Majewski type, <sup>486</sup> and axial spondylometaphyseal dysplasia. <sup>487</sup>	Different NEK1 mutations have been discovered and associated with ALS in various cohorts. <sup>488</sup> Two main mutations, NEK1 loss-of-function variants and missense variants such as p.Arg261His have been discovered and are believed to be associated with ALS. <sup>489-492</sup>



## Huntington's disease

### Overview

Huntington's disease (HD), also known as Huntington's chorea, is a rare hereditary neurodegenerative disorder characterized by progressive motor dysfunction, cognitive decline, and psychiatric symptoms (the term 'chorea' refers to the jerky, unpredictable movements of the muscles in the face, arms and legs that is a prevalent symptom of Huntington's disease).<sup>493-499</sup> The estimates are that the global prevalence of HD is ~2.7 per 100,000 persons.<sup>500</sup>

HD is caused by a mutation in the HTT gene, located on chromosome 4, which encodes the huntingtin protein. This mutation involves an expansion of a trinucleotide repeat sequence, known as CAG, within the gene. Individuals with HD inherit an expanded CAG repeat from one of their parents, leading to the production of a mutant huntingtin protein. The length of the CAG repeat is inversely correlated with the age of onset and severity of symptoms, with longer repeats generally associated with earlier onset and more severe disease.<sup>501-504</sup>

HD follows an autosomal dominant pattern of inheritance, meaning that a person who inherits a single copy of the mutant gene from one parent will develop the disease, regardless of whether the other parent carries the mutation. Each child of an affected individual has a 50% chance of inheriting the mutated gene.<sup>505, 506</sup> The mutant huntingtin protein disrupts normal cellular functions and leads to neuronal dysfunction and death, particularly in the basal ganglia and cerebral cortex of the brain. These regions are involved in movement control, cognition, and emotion regulation. Progressive degeneration of these brain areas results in the characteristic symptoms of HD, which typically worsen over time.<sup>501, 507, 508</sup>

The most prominent early symptoms of HD are often motor-related and may include involuntary movements called chorea, which are rapid, jerky, and random. Other motor symptoms can include dystonia (sustained muscle contractions causing twisting or repetitive movements), rigidity, bradykinesia (slowness of movement), and impaired coordination and balance.<sup>509-511</sup> HD also affects cognitive function, leading to impairments in memory, executive function, attention, and decision-making. As the disease progresses, individuals may experience difficulties with language, planning, problem-solving, and other cognitive tasks.<sup>512, 513</sup> Psychiatric symptoms are common in HD and can precede motor symptoms by several years. These can include depression, anxiety, irritability, apathy, impulsivity, and obsessive-compulsive behaviors. Psychiatric symptoms can significantly impact quality of life for individuals with HD and their caregivers.<sup>514, 515</sup> While most cases of HD typically manifest in adulthood, a small percentage of individuals develop symptoms during childhood or adolescence, known as juvenile-onset HD. Juvenile-onset HD tends to progress more rapidly and may have distinct clinical features compared to adult-onset HD.<sup>493, 509, 516</sup>

Currently, there is no cure for HD, and available treatments focus on managing symptoms and improving quality of life. Medications can help alleviate motor symptoms, psychiatric symptoms, and complications such as chorea and depression. Doctors utilize an arsenal of medications to help alleviate the symptoms of Huntington's disease including antipsychotics and antidepressants. Tetrabenazine (Xenazine)<sup>517</sup> and deutetrabenazine (Austedo)<sup>518, 519</sup> are two US FDA drugs approved in 2008 and 2017, respectively, for treating chorea in Huntington's disease. Physical therapy, speech therapy, and occupational therapy may also be beneficial in managing motor and functional impairments.<sup>503, 520, 521</sup> Genetic testing can confirm the diagnosis of HD in individuals with symptoms or a family history of the disease. Genetic counseling is recommended

for individuals considering testing to discuss the implications of the test results and the potential impact on themselves and their families.<sup>509, 522</sup> Ongoing research efforts aim to better understand the underlying mechanisms of HD and develop disease-modifying treatments. Clinical trials are underway to test potential therapies targeting the mutant huntingtin protein, neuroinflammation, and other pathways implicated in HD pathogenesis.<sup>523-525</sup> Despite the challenges posed by HD, advances in research and clinical care offer hope for improved treatments and ultimately a cure for this devastating neurodegenerative disorder.

### *Pathogenesis of Huntington's disease*

The pathogenesis of Huntington's disease (HD) involves a complex interplay of genetic, molecular, and cellular mechanisms that ultimately lead to neurodegeneration in specific regions of the brain.<sup>504, 526-528</sup>

HD is caused by a mutation in the HTT gene, located on chromosome 4. This mutation involves an abnormal expansion of a trinucleotide repeat sequence, known as CAG, within the gene. The CAG repeat encodes a polyglutamine tract in the huntingtin protein. The mutant huntingtin protein (mHTT) has an expanded polyglutamine tract, which is toxic to neurons and disrupts normal cellular functions.<sup>501, 509, 529</sup> The expanded polyglutamine tract in mHTT leads to the misfolding and aggregation of the protein. These aggregates, known as inclusion bodies, accumulate within neurons and disrupt cellular processes. Protein aggregation is a hallmark feature of HD pathology and contributes to neuronal dysfunction and death.<sup>530-532</sup>

mHTT disrupts mitochondrial function, leading to impaired energy production, increased oxidative stress, and mitochondrial fragmentation. Mitochondrial dysfunction contributes to neuronal vulnerability and exacerbates neurodegeneration in HD.<sup>533-535</sup> Dysregulation of glutamate signaling and excitotoxicity play a role in HD pathogenesis. mHTT disrupts glutamate homeostasis, leading to excessive glutamate release and activation of NMDA receptors. Prolonged activation of NMDA receptors results in calcium influx, mitochondrial dysfunction, and neuronal damage.<sup>536-539</sup>

mHTT interferes with axonal transport, the process by which cellular components are transported along axons. Impaired axonal transport disrupts the delivery of essential proteins and organelles to synapses, leading to synaptic dysfunction and neuronal degeneration.<sup>540-542</sup> Neuroinflammatory processes contribute to HD pathogenesis and exacerbate neuronal damage. Activation of microglia and astrocytes, the resident immune cells of the CNS, leads to the release of pro-inflammatory cytokines, ROS, and other toxic molecules that contribute to neuronal dysfunction and death.<sup>543-545</sup>

mHTT disrupts gene transcription and expression, leading to widespread alterations in gene expression patterns in affected neurons. Transcriptional dysregulation affects multiple cellular pathways involved in neuronal function, survival, and plasticity, contributing to neurodegeneration in HD.<sup>546-548</sup> HD pathology affects synaptic function and plasticity, leading to impaired neurotransmission and synaptic loss. Synaptic dysfunction contributes to cognitive and motor impairments in HD and is an early feature of the disease.<sup>549, 550</sup> Certain neuronal populations, particularly those in the striatum and cerebral cortex, are selectively vulnerable to the toxic effects of mHTT. The striatum, which plays a crucial role in motor control and cognition, is severely affected in HD, leading to the characteristic motor and cognitive symptoms of the disease.<sup>551-553</sup>

### *Genetic background of Huntington's disease*

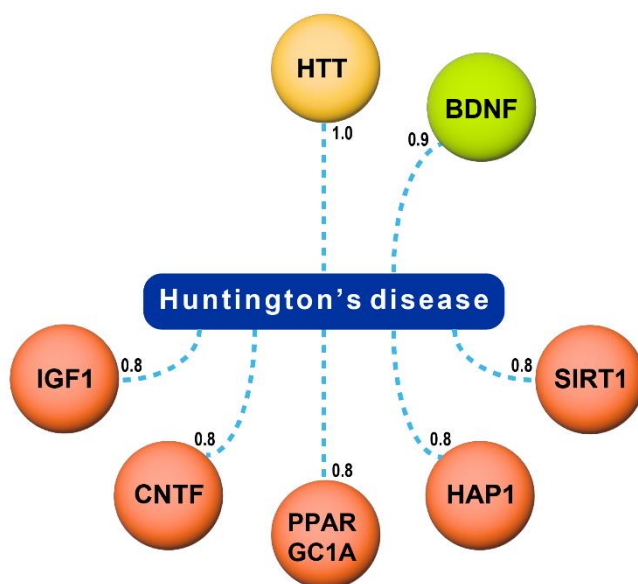
The genetic background of Huntington's disease (HD) is primarily characterized by a mutation in the HTT gene, which is located on the short arm of chromosome 4 (4p16.3).<sup>502, 506</sup> The genetic mutation responsible for HD involves an abnormal expansion of a trinucleotide repeat sequence, known as CAG, within the HTT gene. The CAG repeat encodes a polyglutamine tract in the huntingtin protein. In individuals with HD, the CAG repeat is expanded beyond a certain threshold, resulting in an increased number of glutamine residues in the huntingtin protein.<sup>526, 528, 554, 555</sup>

HD follows an autosomal dominant pattern of inheritance, meaning that a person who inherits a single copy of the mutant HTT gene from one parent will develop the disease, regardless of whether the other parent carries the mutation. Each child of an affected individual has a 50% chance of inheriting the mutated gene.<sup>502, 509</sup> A notable feature of HD inheritance is anticipation, where the age of onset of symptoms tends to decrease and the severity of symptoms tends to increase in successive generations. This phenomenon is thought to be due to further expansion of the CAG repeat during transmission from one generation to the next.<sup>556, 557</sup> While the presence of an expanded CAG repeat in the HTT gene is necessary for the development of HD, the age of onset and severity of symptoms can vary widely among individuals with the mutation. Some individuals may carry the mutation but never develop symptoms (referred to as incomplete penetrance), while others may have an earlier onset and more severe disease.<sup>558-560</sup>

In addition to the CAG repeat expansion in the HTT gene, other genetic factors may influence the age of onset and progression of HD. Genetic modifiers, such as variations in other genes or non-genetic factors, may interact with the mutant HTT gene to modify disease onset, severity, and progression. Research into genetic modifiers may provide insights into the variability of HD phenotypes and identify potential targets for therapeutic intervention.<sup>561-564</sup>

While the role of HTT in HD is well-established<sup>565-568</sup>, we wished to collate a comprehensive list of other genes that might play a role in the development of HD. Results from our are visualized in Figure 9. Besides HTT, other genes that might play a crucial role in the etiology of HD include BDNF, NPY, SIRT1, HAP1, PPARGC1A, JPH3, GRIN2B, CNTF, and IGF1. Details about each of the identified genes can be found in Table 2.

Genetic testing for the CAG repeat expansion in the HTT gene can confirm the diagnosis of HD in individuals with symptoms or a family history of the disease. Predictive genetic testing can also be offered to individuals who are at risk of inheriting the mutation but do not yet have symptoms. Genetic counseling is recommended for individuals considering testing to discuss the implications of the test results and the potential impact on themselves and their families.<sup>569-571</sup> Understanding the genetic basis of HD is essential for accurate diagnosis, genetic counseling, and the development of targeted treatments for this devastating neurodegenerative disorder.



**Figure 9.** Genes associated with Huntington's disease (HD) based on data from the CAS Content Collection. Only genes with an association score of greater than 0.8 and with at least 10 records are shown here. Color corresponds to association score – yellow (1.0), green (0.9) and orange (0.8). The nature of the line indicates association source with dashed lines indicating a majority of records resulting from text mining.

**Table 2.** List of genes associated with Huntington's disease (HD), as shown in Figure 9, with the association type as causal or contributing.

Gene	Association score	Protein	Expression profile, function and other diseases	Huntington's disease (HD)
BDNF <sup>572</sup>	0.9	Brain derived neurotrophic factor	Brain derived neurotrophic factor are small proteins expressed widely through various parts of the brain <sup>573, 574</sup> and important for synaptic plasticity. <sup>574</sup> Implicated in psychiatric illness. <sup>575</sup>	Reduction in levels of BDNF are observed in HD. <sup>576, 577</sup> Targeting BDNFs is also a potential therapeutic strategy in HD. <sup>578, 579</sup> Increasing BDNF levels are indirectly potentially beneficial in treating HD. <sup>580, 581</sup> There are conflicting reports about the utility of BDNF as a potential biomarker for HD. <sup>582-584</sup>
SIRT1 <sup>585</sup>	0.8	Sirtuin 1	Sirtuin is a type of deacetylase that is dependent on nicotinamide adenosine dinucleotide (NAD) <sup>586</sup> with a wide expression profile (brain as well as other organs such as kidney, liver, pancreas, spleen). <sup>587</sup> Sirtuin 1 appears to have a role in obesity <sup>588</sup> , metabolic disorders including diabetes, <sup>589, 590</sup> as well as immune response. <sup>591</sup>	Sirtuin 1 appear to have a neuroprotective role in HD. <sup>592, 593</sup> Modulation of sirtuin 1 activity is being explored as viable therapeutic strategy <sup>593-596</sup> with arguments being made for sirtuin 1 activation <sup>597</sup> as well as inhibition. <sup>598</sup> Selisistat (CAS RN: 49843-98-3) was being developed as a small molecule inhibitor of sirtuin 1 for the treatment of HD. <sup>599, 600</sup>
HAP1 <sup>601</sup>	0.8	Huntingtin associated protein 1	Huntingtin associated protein 1 is expressed in various areas of the CNS including amygdala and hypothalamus among others. <sup>602</sup> HAP1 found to be important for trafficking of cellular components. <sup>603, 604</sup> HAP1 is also implicated in various neurodegenerative diseases such as Alzheimer's, Parkinson's as well as rare diseases such as ALS. <sup>605</sup>	HAP1 protein has been shown to interact with mutant huntingtin protein. <sup>605</sup> The HAP1-mHTT association impairs normal/physiological cellular trafficking carried out by HAP1 in conjunction with other proteins. <sup>606, 607</sup> There are contradictory studies indicating HAP1 has a bearing on age of onset. <sup>608, 609</sup>
PPARGC1A <sup>610</sup>	0.8	Peroxisome proliferator-activated receptor (PPAR)-γ coactivator 1 α (PGC-1α)	PGC-1α acts as a coactivator to PPARα and plays a role in mitochondrial energy metabolism. <sup>611</sup> PGC-1α has been studied in the context of physical fitness especially in athletes. <sup>612-616</sup> Recently, PGC-1α has been linked to Parkinson's disease. <sup>617, 618</sup>	Impaired levels of PGC-1α are observed in HD <sup>619</sup> with overexpression of PGC-1α allowing rescue in HD mice. <sup>620</sup> Mutant huntingtin protein is shown to inhibit PGC-1α expression. <sup>621</sup> Studies suggest that PGC-1α may have an impact on age of onset of HD. <sup>622, 623</sup>

CNTF <sup>624</sup>	0.8	Ciliary neurotrophic factor	Ciliary neurotrophic factor is a cytokine belonging to the IL-6 family. <sup>625</sup> Expressed in Schwann cells as well as T cells. <sup>626</sup> The are also important for the survival of oligodendrocytes <sup>627</sup> and have been implicated in CNS diseases involving demyelination <sup>628-630</sup> as well as motor neuron disease. <sup>631</sup>	CNTF appears to have a neuroprotective role in HD. <sup>632</sup> Administration of CNTF has been explored as a potential treatment for HD with varying results. <sup>633-636</sup>
IGF1 <sup>637</sup>	0.8	Insulin-like growth factor 1	Also referred to as somatomedin C or somatomedin 1, <sup>638</sup> IGF1 is a small protein of 70 amino acid residues <sup>639</sup> secreted primarily by the liver. <sup>640</sup> Acts as a mediator of growth hormone and is critical in somatic growth. <sup>640</sup> Elevated IGF1 levels has been linked to rare disorders such as gigantism and acromegaly. <sup>641</sup> IGF1 has also been studied in the context of aging. <sup>642, 643</sup>	IGF1 has been associated with declining social cognition in HD <sup>644</sup> with IGF1 levels as potential indicators. <sup>645</sup> Targeting IGF1 has also been explored as a therapeutic strategy in HD with both IGF1 inhibitors <sup>646</sup> as well as administration of IGF1 itself having been studied. <sup>647</sup>

## Myasthenia gravis

### Overview

Myasthenia gravis (MG) is a rare chronic neuromuscular disorder characterized by weakness and rapid fatigue of voluntary muscles.<sup>648-650</sup> With the earliest accounts dating back to the late 1600s,<sup>651</sup> MG is described as an autoimmune disorder affecting neuromuscular junctions.<sup>652</sup> Thought to arise as a result of the body generating antibodies against the acetylcholine receptor (AChR) or muscle specific kinase,<sup>652, 653</sup> resulting in the immune system mistakenly attacking receptors on muscle cells, particularly at the neuromuscular junction where nerve impulses stimulate muscle contractions,<sup>654-656</sup> MG leads to muscle weakness and a host of other symptoms.<sup>657, 658</sup> During the Covid-19 pandemic, reports have emerged of onset of MG after SARS-Cov-2 infection.<sup>659, 660</sup> Global prevalence rates range from 150 to 200 cases per 1,000,000 people. The prevalence of MG in the United States is estimated at 14 to 20 cases per every 100,000 people or between 36,000 and 60,000 cases. In Europe, an estimated 56,000 to 123,000 individuals live with MG.<sup>661, 662</sup>

The hallmark symptom of MG is muscle weakness, which typically worsens with activity and improves with rest. This weakness can affect various muscles, including those controlling eye movements, facial expressions, chewing, swallowing, and breathing. Fatigue is also a common feature, with muscles becoming progressively weaker during periods of activity. Weakness in the muscles that control eye movements often leads to double vision or drooping of the eyelids (ptosis). Difficulty swallowing (dysphagia) can occur due to weakness in the muscles involved in chewing and swallowing. In severe cases, weakness of the muscles involved in breathing can lead to respiratory difficulties, which can be life-threatening. Symptoms of MG can vary widely among individuals and may fluctuate over time, making diagnosis challenging.<sup>663-667</sup>

Factors such as stress, illness, fatigue, or certain medications can exacerbate symptoms in people with MG. Diagnosis typically involves a thorough medical history, physical examination, blood tests to check for specific antibodies associated with MG, and specialized tests such as electromyography and nerve conduction studies. While there is no cure for MG, various treatments can help manage symptoms and improve quality of life. These may include medications such as acetylcholinesterase (AChE) inhibitors, immunosuppressants, and corticosteroids.<sup>668-670</sup> Some individuals may also benefit from procedures such as plasmapheresis or intravenous immunoglobulin therapy. In severe cases, surgical removal of the thymus gland (thymectomy) may be recommended.<sup>671</sup> With appropriate treatment, many people with MG can lead fulfilling lives. However, the course of the disease can be unpredictable, and long-term management often requires close monitoring and adjustments to treatment by developing a personalized treatment plan and receive ongoing support and care.<sup>672-678</sup>

### *Pathogenesis of myasthenia gravis*

The pathogenesis of MG involves an autoimmune response targeting components of the neuromuscular junction (NMJ), where nerve impulses trigger muscle contractions.<sup>662, 679</sup> MG is primarily driven by an autoimmune response, where the body's immune system mistakenly identifies components of the NMJ as foreign and attacks them. The primary target of this autoimmune response is the AChRs on the muscle cell membrane. These receptors normally bind acetylcholine, a neurotransmitter released by motor neurons, to initiate muscle contractions.<sup>652, 680, 681</sup>

In MG, the immune system produces autoantibodies called anti-AChR antibodies. These antibodies bind to the AChR on the muscle cell membrane, leading to several effects: (i) some antibodies block the binding sites on the AChR, preventing acetylcholine from binding and initiating muscle contractions; (ii) other antibodies may cross-link adjacent AChR molecules, leading to internalization and degradation of the AChR complex, reducing the number of functional receptors on the muscle cell membrane; (iii) the binding of antibodies to AChR can also activate the complement system, a part of the immune system involved in inflammation and cell destruction. This further contributes to damage and dysfunction at the NMJ.<sup>653, 682-684</sup>

The presence of anti-AChR antibodies and complement activation disrupts neuromuscular transmission, leading to: (i) reduced signal transmission, since with fewer functional AChR available, the binding of acetylcholine released by motor neurons is impaired, resulting in weakened muscle contractions; (ii) endplate destruction, since chronic immune-mediated damage to the NMJ can lead to structural changes, including destruction of the postsynaptic membrane and alterations in the distribution of AChR.<sup>685-687</sup>

The thymus gland plays a role in the development of MG in some individuals.<sup>688</sup> It is commonly associated with thymic abnormalities, such as thymic hyperplasia or thymoma (a tumor of the thymus). The thymus is thought to contribute to the production of autoantibodies and the maturation of autoreactive T cells involved in the autoimmune response seen in MG. In general, the pathogenesis of MG involves a complex interplay between autoantibodies, complement activation, and immune-mediated damage at the neuromuscular junction, leading to impaired neuromuscular transmission and muscle weakness characteristic of the disease.<sup>688-690</sup>

### *Genetic background of myasthenia gravis*

Myasthenia gravis has a complex genetic background, but it is not typically considered a purely genetic disorder like some other conditions. Instead, MG is primarily regarded as an autoimmune disease with genetic predispositions. While the exact cause of MG is unknown, there is evidence suggesting a genetic predisposition to the disease. Certain genetic variations or polymorphisms have been associated with an increased risk of developing MG. These variations are often related to genes involved in immune system function, such as genes encoding human leukocyte antigens (HLAs), specifically the HLA-B8 and HLA-DR3 alleles.<sup>691-697</sup>

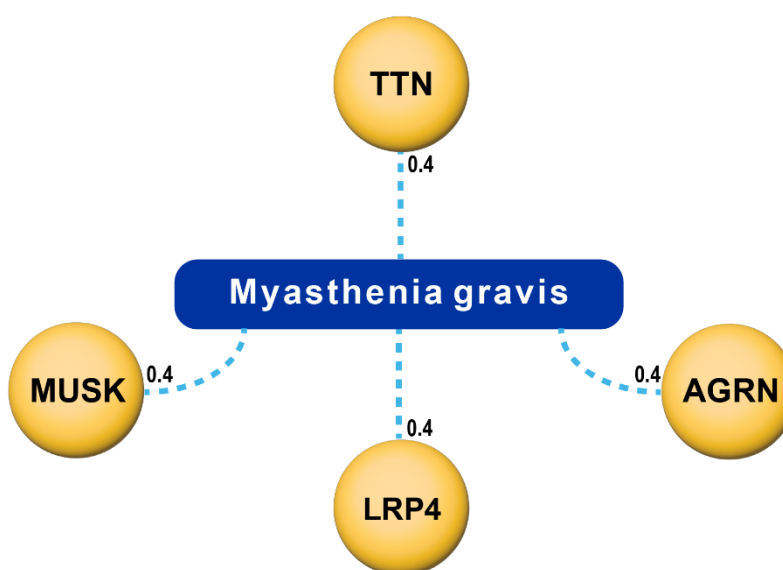
MG can sometimes run in families, indicating a potential genetic component.<sup>698</sup> Family studies have shown that first-degree relatives of individuals with MG have a higher risk of developing the condition compared to the general population. However, the inheritance pattern is usually not straightforward, suggesting the involvement of multiple genetic and environmental factors.<sup>698, 699</sup>

Certain HLA alleles, particularly those within the major histocompatibility complex (MHC) region, have been consistently linked to an increased risk of MG.<sup>700</sup> For example, the HLA-DR3 allele has been associated with MG, especially in individuals with early-onset disease and thymic abnormalities.<sup>696</sup> However, HLA associations alone are not sufficient to explain the development of MG, indicating the involvement of other genetic and environmental factors. In addition to HLA alleles, studies have identified other genetic factors that may contribute to the risk of MG. These include genes involved in immune regulation, such as those encoding cytokines, chemokines, and components of the complement system. Variations in these genes may affect immune function and predispose individuals to autoimmune diseases like MG.<sup>691, 701, 702</sup>



Using data from the CAS Content Collection, we have put together genes known to have an association with MG (Figure 10). These genes include AGRN, LRP4, MUSK, and TTN and are explored in more detail in Table 3.

It is important to note that the development of MG likely involves complex interactions between genetic susceptibility factors and environmental triggers.<sup>696</sup> Environmental factors such as infections, medications, and hormonal changes may play a role in triggering the autoimmune response in genetically susceptible individuals. While genetic factors contribute to the susceptibility to MG, the disease's development is likely multifactorial, involving a combination of genetic predisposition, environmental triggers, and immune dysregulation. Further research is needed to elucidate the specific genetic mechanisms underlying MG and their interactions with environmental factors.<sup>682, 703, 704</sup>



**Figure 10.** Genes associated with myasthenia gravis based on data from the CAS Content Collection. Only genes with an association score<sup>705</sup> greater than 0.4 and with at least 10 records are shown here. Color corresponds to association score with yellow corresponding to association scores of 0.4. The nature of the line indicates association source with dashed lines indicating a majority of records resulting from text mining.

**Table 3.** List of genes associated with myasthenia gravis (MG), as shown in Figure 10, with the association type as causal or contributing.

Gene	Association score	Protein	Expression profile, function and other diseases	Myasthenia gravis (MG)
AGRN <sup>706</sup>	0.4	Agrin	Agrin is a glycosylated protein involved in activation of MuSK and the development of NMJs. <sup>707</sup> Two isoforms of agrin exist – neural agrin and muscle agrin. <sup>707</sup> Abnormalities in agrin have been observed in Alzheimer's <sup>708, 709</sup> and Parkinson's <sup>710</sup> diseases and agrin targeted therapy has also been suggested for treatment of cardiac injury. <sup>711</sup> Agrin's role in immune system has also been explored. <sup>712</sup>	Autoantibodies against agrin have been detected in individuals suffering from MG. <sup>707, 713-715</sup> Mice immunized with agrin generated agrin antibodies and exhibited reduced muscle weakness along with morphological changes in NMJs. <sup>716</sup>
LRP4 <sup>717</sup>	0.4	Low-density lipoprotein (LDL) receptor related protein 4	LDL receptor related protein 4 is a receptor with a single transmembrane domain that binds to neural agrin and induces activation of MuSK leading to development of NMJs. <sup>707</sup> LRP4 has been implicated in neurological disorders. <sup>718</sup>	Similar to agrin, autoantibodies against LRP4 have been detected in individuals with MG <sup>715, 719-721</sup> – especially true of individuals suffering from double-seronegative myasthenia gravis.
MUSK <sup>722</sup>	0.4	Muscle associated receptor tyrosine kinase (MuSK)	The agrin-LRP4 complex binds to and activates MuSK, a receptor tyrosine kinase, which in turn interacts with other proteins (including docking-protein 7 and tumorous imaginal disc 1) to eventually cause clustering of acetylcholine receptors in the NMJs. <sup>723, 724</sup> Similar to agrin, MuSK is also a single transmembrane domain protein. <sup>725</sup> Involvement of MUSK has been studied in the context of muscle disorders including congenital myasthenic syndrome <sup>726, 727</sup> as well as fetal akinesia deformation sequence (FADS) <sup>728, 729</sup> a rare lethal disorder affecting infants. <sup>730, 731</sup>	MuSK-MG is a subtype of MG characterized by an acute onset <sup>725, 732</sup> and seemingly appear to be associated with worse outcomes than AChR antibody MG. <sup>733</sup> MuSK-MG tends to show greater prevalence among females <sup>734</sup> with recent studies published by researchers in China showing similar trends. <sup>735, 736</sup> Autoantibodies against MuSK prevent binding of LRP4 to MuSK disrupting NMJs. <sup>737</sup>
TTN <sup>738</sup>	0.4	Titin	Titin is one of the largest proteins in the human body and is composed of 27,000-34,000 amino acid residues. <sup>739</sup> Along with actin and myosin, titin is integral for muscle contraction. <sup>740</sup> Diseases arising due to mutations in the TTN gene are referred to as titanopathy and characterized by muscle weakness. <sup>741-743</sup>	Autoantibodies against titin have been reported in MG patients with thymoma. <sup>744-746</sup>

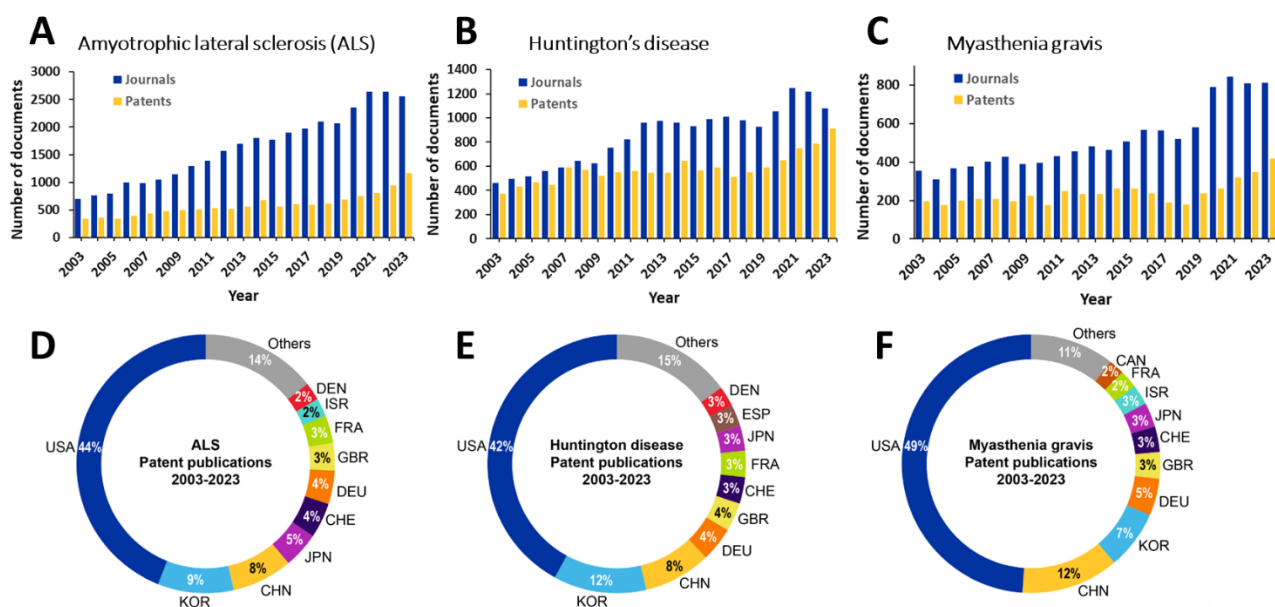
## Landscape analysis of data on ALS, HD, and MG in CAS Content Collection

We focused our attention to the three rare diseases, ALS, HD, and MG, to perform a deeper dive into the publication landscape as reflected in the CAS Content Collection, in order to identify and understand indistinct connections and correlations in these areas.

### Documents yearly growth and geographic distribution

A look at journal and patent publication trends specific to the three diseases in focus indicates the following:

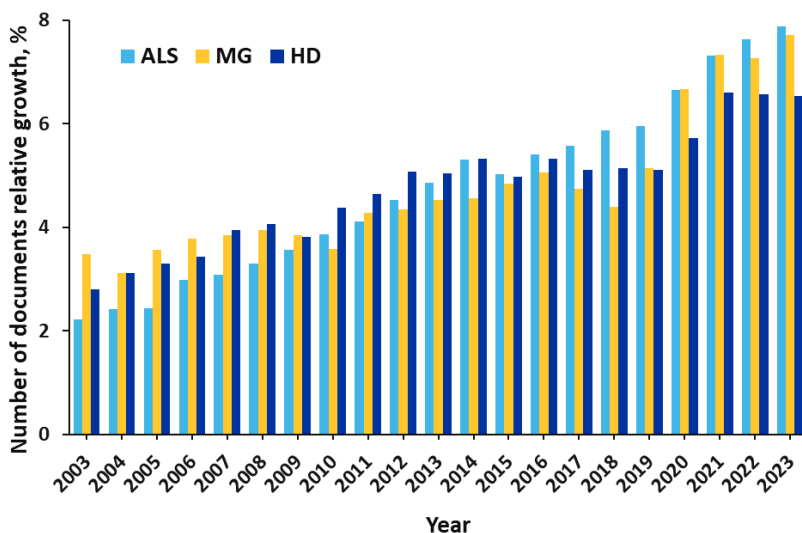
- Publications related to ALS show a steady incline over the last two decades with journal publications increasing by over 20% and the patent publications – by the impressive ~70% over 2019 to 2022 (Figure 11A).
- Growth in journal publications related to HD exhibit an upward trend increasing consistently till 2012, followed by a few years of pleateauing and growth in the period 2019-2021. Patent publications grow consistently after 2018 (Figure 11B).
- For MG, journal publications exhibit slow but generally consistent growth. Patent publications on the other hand appear to grow consistently after 2018, after years of pleateauing (Figure 11C).
- For all three diseases journal publications outnumber the patent publications.
- Comparison of geographical distribution of patent assignees in the three diseases areas indicate a high degree of overlap, with countries such as the United States (USA), China (CHN), South Korea (KOR), Japan (JPN), Germany (DEU), Switzerland (CHE), the United Kingdom (GBR), and France (FRA) being common amongst the three. Denmark (DEN) shows up in the top 10 countries or regions for patent assignees in ALS and HD while Israel (ISR) features in the top 10 for ALS and MG (Figure 11D-F).



**Figure 11.** Publications for specific rare diseases: (A) amyotrophic lateral sclerosis (ALS), (B) Huntington's disease (HD) and (C) myasthenia gravis (MG). Donut graphs showing geographical distribution of patent assignees (both non-commercial and commercial organizations) for each of three diseases: (D) ALS, (E) HD and (F) MG. Countries are indicated by their standard three letter

codes – United States (USA), South Korea (KOR), China (CHN), Japan (JPN), Switzerland (CHE), Germany (DEU), United Kingdom (GBR), France (FRA), Israel (ISR), Denmark (DEN), Spain (ESP), Canada (CAN). Data includes journal and patent publications related to each individual rare disease sourced from the CAS Content Collection for 2003-2023.

The relative growth in the number of documents related to the three rare diseases in the CAS Content Collection in the last two decades (2003-2023) is shown in Figure 12. While MG exhibited the fastest growth in the years 2003-2006, ALS took the lead since 2014, with nearly 8% relative growth in 2023. This increase coincides with and therefore might be attributed to the viral “ice bucket” challenge that started in 2014.



**Figure 12.** Relative growth in the number of documents related to ALS, MG, and HD in 2003-2023.

Using our access to robust CAS indexing data, we further explored co-occurrences of the three rare diseases – ALS, HD, and MG, with a host of concepts such as other non-rare and rare diseases (Figure 13A), therapy types and drugs (Figure 13B), as well as proteins and cells (Figure 13C).

## Disease comorbidities

### *Amyotrophic lateral sclerosis (ALS)*

Hypertension and dyslipidemia are the most commonly reported comorbidities.<sup>747</sup> These results are of particular interest considering the debate related to the protective role of hypertension and other cardiovascular disorders for the prognosis and survival of ALS.<sup>748, 749</sup> The occurrence of autoimmune diseases in ALS patients is frequently reported, but little is known about the related clinical phenotype.<sup>750</sup> Association of ALS and cancer (overall cancer, as well as certain specific cancers) has been examined, and the results have been ambiguous and inconsistent regarding the risk of cancer in general, and of specific cancers.<sup>751-754</sup> A study examined co-occurrences of ALS and multiple sclerosis, both associated with upper motor neuron degeneration, checking the possibility for common biological pathways. The study concluded

though that rather than a shared biology, the co-occurrences are random, although a common risk factor cannot be excluded.<sup>755</sup>

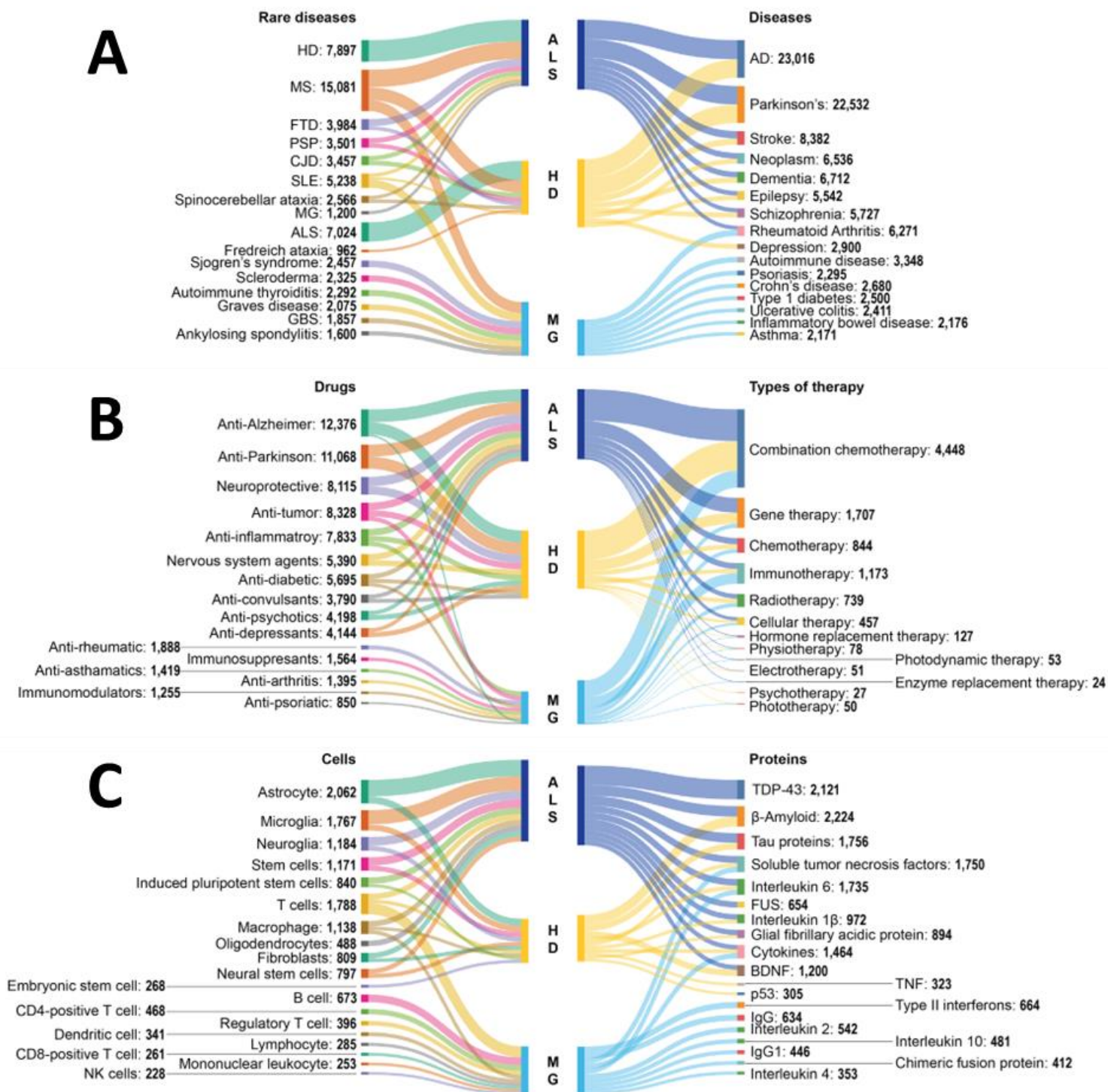
### *Huntington's disease*

Examination of comorbidities associated with HD showed depression as the most common, affecting nearly 43% of patients, with females more frequently affected than males.<sup>756</sup> Other reported comorbidities of HD include dementia (~ 38% of patients), urinary incontinence (over 32.% of patients), extrapyramidal and movement disorders (over 30% of patients), dysphagia (nearly 30% of patients), and disorders of lipoprotein metabolism (over 28% of patients).<sup>756</sup> Another study found that the prevalence of comorbidities, especially in the musculoskeletal, cardiovascular, and psychiatric diseases, was higher in patients with HD than in a control group of healthy individuals.<sup>757</sup> The observed psychiatric comorbidities comprise obsessive-compulsive disorder, depression, insomnia, bipolar affective disorders, dementia, and neurosis.<sup>757</sup> A recent study reported higher incidence rate of multiple comorbidities, such as obsessive-compulsive disorder, psychosis, communication disorders, depression, anxiety, dementia, and others, in individuals with adult-onset HD than in controls; in patients with juvenile-onset HD, the incidence rates of epilepsy, and acute respiratory symptoms have been found higher.<sup>758</sup> Reports exist of the co-occurrence of HD with ALS.<sup>759</sup> The number of reported cases of co-occurrence are low, which obstruct systematic observational studies or clinical trials. A single case of multiple sclerosis with comorbid HD has been reported.<sup>760</sup>

### *Myasthenia gravis*

Patients with MG may be associated with autoimmune as well as non-autoimmune comorbidities.<sup>761, 762</sup> Autoimmune comorbidities such as autoimmune thyroiditis, followed by systemic lupus erythematosus, and rheumatoid arthritis have been reported as the most frequent comorbidities of MS.<sup>763</sup> Co-occurrence of thymoma MG and late-onset MG with cardiomyositis and subclinical cardiac dysfunction have been reported, however, these conditions have not been considered a significant risk. Lymphomas and some other cancers have been documented with a slightly higher frequency, autoimmune MG does not appear to be a separate cancer risk factor.<sup>763</sup> MG has been reported in 0.2% of the diagnosed cases of autoimmune thyroid disease. Pernicious anemia, psoriasis, systemic vasculitis, and other disorders have also been reported.<sup>761-764</sup> The prevalence of autoimmune comorbidity is different for MG subtypes: individuals with early-onset MG are more likely to develop an additional autoimmune disease than those with late-onset MG.<sup>763</sup> Another group of diseases co-occurring frequently with MG are cardiovascular diseases. Arterial hypertension is noted to prevail in patients with MG.<sup>764</sup> With respect to cancers – lymphoma, breast cancer, and lung cancer have been found more common in the group of MG patients.<sup>762, 764</sup> Mental health disorders such as depression and anxiety are frequent among MG patients. Concurrent MG and ALS have also been registered in many cases.<sup>762, 764</sup>

The results of examination of co-occurrences of ALS, HD, and MG terms with other rare and non-rare diseases in the CAS Content Collection are illustrated in Figure 13A. ALS and HD most frequently co-occurs with Alzheimer's, and Parkinson's diseases, and multiple sclerosis, as well as between themselves. MG most frequently co-occurs with multiple sclerosis, rheumatoid arthritis, and systemic lupus erythematosus (Figure 13A).



**Figure 13.** Co-occurrences of amyotrophic lateral sclerosis (ALS), Huntington's disease (HD) and myasthenia gravis (MG) with various concepts such as (A) other rare and non-rare diseases, (B) types of therapy and drugs used to treat symptoms and (C) cells and proteins. Data includes both patent and journal publications sourced from the CAS Content Collection for the period 2003-2023 in the field of rare diseases. Abbreviations: HD – Huntington's disease, MS – multiple sclerosis, FTD – frontotemporal dementia, PSP – progressive supranuclear palsy, CJD – Creutzfeldt-Jakob disease, SLE – systemic lupus erythematosus,

MG – myasthenia gravis, ALS – amyotrophic lateral sclerosis, GBS – Guillain Barre syndrome and AD - Alzheimer's disease.

## Types of therapy

Combination therapies are the most common type of therapy for the three rare diseases, according to the co-occurrence analysis on data from the CAS Content Collection (Figure 13B). Current research indicates that drugs acting at a single target may be insufficient for the treatment of multifactorial neurodegenerative diseases such as HD and ALS, typified by the coexistence of multiple etiopathologies including oxidative stress, protein misfolding and aggregation, mitochondrial dysfunction, inflammation, and metal accumulation at the sites of neurodegeneration. Clearly, combination drug therapy of neurodegenerative diseases with multifunctional remedies exhibiting diverse biological properties is supposed to have distinct advantage.<sup>765</sup>

ALS and HD frequently co-occur with gene therapy, while for MG, immunotherapy is the second most frequently occurring after combination therapy (Figure 13B). Indeed, with respect to ALS, over 50 genes have been identified as either cause or modifier in ALS and ALS/frontotemporal dementia spectrum disease. Substantial effort has been made to discover pathways underlying the pathogenesis of these gene mutations. Accordingly, targeting etiologic genes to suppress their toxic impacts have been investigated widely, with the major strategies including: removal or inhibition of abnormally transcribed RNA using miRNA or antisense oligonucleotides (ASOs); degradation of abnormal mRNA using RNA interference (RNAi); decrease or inhibition of mutant proteins by using, e.g., antibodies against misfolded proteins; and/or DNA genome editing with methods such as CRISPR or CRISPR/Cas.<sup>766</sup> The favorable results of these studies have resulted in application of some of these strategies in clinical trials for ALS, especially for C9orf72 and SOD1.<sup>766-769</sup> Regarding HD, gene therapies are being explored using genetic material to ramp up expression of genes whose functions are declined or are damaged over the course of the disease, to boost the brain and body's natural resilience against disease progression.<sup>770</sup> As a monogenic disease, HD is a good target for gene therapy approaches, including the use of programmable endonucleases. A protocol for HTT gene knock-out using a modified Cas9 protein (nickase, Cas9n) has been recently tested with promising results.<sup>771</sup>

Regarding MG, immunotherapeutic biologics are emerging as important therapeutic tools. The monoclonal antibody eculizumab has been approved by the US FDA for refractory MG on the basis of a Phase III trial.<sup>772</sup> Another monoclonal antibody, rituximab, is in advanced stage of clinical trials. A selection of newer anti-CD20 antibodies such as ocrelizumab (CAS RN: 637334-45-3), ofatumumab (CAS RN: 679818-59-8), obinutuzumab (CAS RN: 949142-50-1), ublituximab (CAS RN: 1174014-05-1) or inebilizumab (CAS RN: 1299440-37-1) are also being tested.<sup>772</sup> Enhanced availability of new biologics provides targeted immunotherapies and the chance to develop more specific therapies for MG.

## Drugs

A co-occurrence search in the CAS Content Collection showed the highest frequency of co-occurrence of ALS and HD with anti-Alzheimer and anti-Parkinson drugs (Figure 13B).

For example, ropinirole, a drug used to treat Parkinson's disease, has showed potential in delaying the progression of ALS.<sup>773</sup> The multifunctional brain permeable iron chelator M30 was shown to possess neuroprotective activities against various neurodegenerative diseases, such as Alzheimer's and Parkinson's disease, and ALS.<sup>774,775</sup> Recently, allopurinol (CAS RN: 315-30-0) and carvedilol (CAS RN: 72956-09-3), medications used to manage gout and high blood pressure, respectively, were reported to significantly reduce the risk of developing ALS, Alzheimer's, or Parkinson's disease.<sup>776,777</sup>

MG frequently co-occurs with immunosuppressant and anti-rheumatic drugs (Figure 13B). Immunosuppressant medications act to lessen the immune system's response in order to avoid the immune attacks on NMJs, thereby limiting muscle fatigue. Common immunosuppressive medications used in treating MG include prednisone (CAS RN: 53-03-2), azathioprine (CAS RN: 446-86-6), cyclophosphamide (CAS RN: 50-18-0), methotrexate (CAS RN: 59-05-2), tacrolimus (CAS RN: 104987-11-3), and mycophenolate mofetil (CAS RN: 128794-94-5).<sup>778-782</sup> Patients with rheumatoid arthritis exhibit high prevalence of MG compared to the general population.<sup>783</sup> The rheumatoid arthritis drug abatacept has been reported to prevent MG in clinical trial.<sup>782</sup> Another drug used in the treatment of rheumatoid arthritis, rituximab (CAS RN: 174722-31-7), has been also shown able to reduce the risk of deterioration in MG.<sup>784</sup>

Since there is no cure for these rare diseases, drug repurposing studies have been intensely searching to identify existing drugs that could be repositioned to be used as viable treatment options. As the pharmaceutical development process is both time-consuming and costly, drug repurposing provides a chance to accelerate it by exploring the beneficial effects of agents approved for other disorders. These drugs have established safety profiles, pharmacokinetic description, formulations, dosages, and manufacturing procedures. Recently, *in silico* pharmacology has been widely applied and various computer applications including machine learning and artificial intelligence approaches have been explored in identifying potential drugs for repurposing to various diseases.

There have been number of studies on the potential of old drugs for the treatment of neurodegenerative diseases, including HD and ALS.<sup>785</sup> Tetrabenazine (CAS RN: 58-46-8) has been developed as antipsychotic drug but has been later repurposed for diseases involving abnormal, involuntary hyperkinetic movements, such as HD.<sup>786</sup> Another antipsychotic, tiapride (CAS RN: 51012-32-9), has been also tested for the treatment of HD.<sup>787</sup> Olanzapine (CAS RN: 132539-06-1)<sup>786,788</sup>, risperidone (CAS RN: 106266-06-2)<sup>789</sup>, quetiapine (CAS RN: 111974-69-7)<sup>790</sup> are also antipsychotic drugs widely prescribed for the treatment of the motor and behavioral symptoms of HD.<sup>785</sup> Other examples of drugs under clinical trial to be repurposed for the treatment ALS include the anti-cancer masitinib (CAS RN: 790299-79-5), the anti-inflammatory ibudilast (CAS RN: 50847-11-5), the antiretroviral triumeq (CAS RN: 1319715-09-7), the anticonvulsant retigabine (CAS RN: 150812-12-7), and the antiestrogen tamoxifen (CAS RN: 10540-29-1).<sup>785</sup>

## Proteins

A search for co-occurrence with proteins showed that ALS exhibits the highest co-occurrence with TDP-43 (Figure 13C). TDP-43 (TAR DNA-binding protein 43) is a key pathological hallmark associated with ALS and related motor neuron diseases.<sup>791</sup> Loss of TDP-43 from the nucleus and abnormal accumulation of TDP-43 aggregates in the cytoplasm of affected neurons is a prominent pathological feature observed in the majority (>97%) of cases, particularly in



sporadic ALS. <sup>218, 792-794</sup> TDP-43 pathology correlates with disease severity and progression, suggesting a central role in ALS pathogenesis. <sup>224</sup>

While HD etiology has been traditionally associated with abnormalities in the huntingtin protein, recent research has explored the involvement of  $\beta$ -amyloid and tau ( $\tau$ ) protein, typically associated with Alzheimer's disease, in HD pathology, as reflected by the frequent co-occurrence of these proteins with HD in the CAS Content Collection (Figure 13C). Studies have shown the presence of  $\beta$ -amyloid deposits as well as elevated levels of phosphorylated tau in the brains of HD patients, particularly in regions affected by neurodegeneration. <sup>795-798</sup>

## Cells

Astrocytes and microglial cells are the most frequently co-occurring cell types with ALS and HD in the rare diseases-related documents of the CAS Content Collection, while MG exhibits high co-occurrence with T-cells and B-cells (Figure 13C). While ALS is traditionally considered a motor neuron disease and HD – a neuronal disorder caused by mutant huntingtin protein (mHTT), emerging evidence suggests that non-neuronal cells, particularly astrocytes, microglia, and other types of neuroglia, play crucial roles in ALS and HD pathogenesis. <sup>162, 799-803</sup>

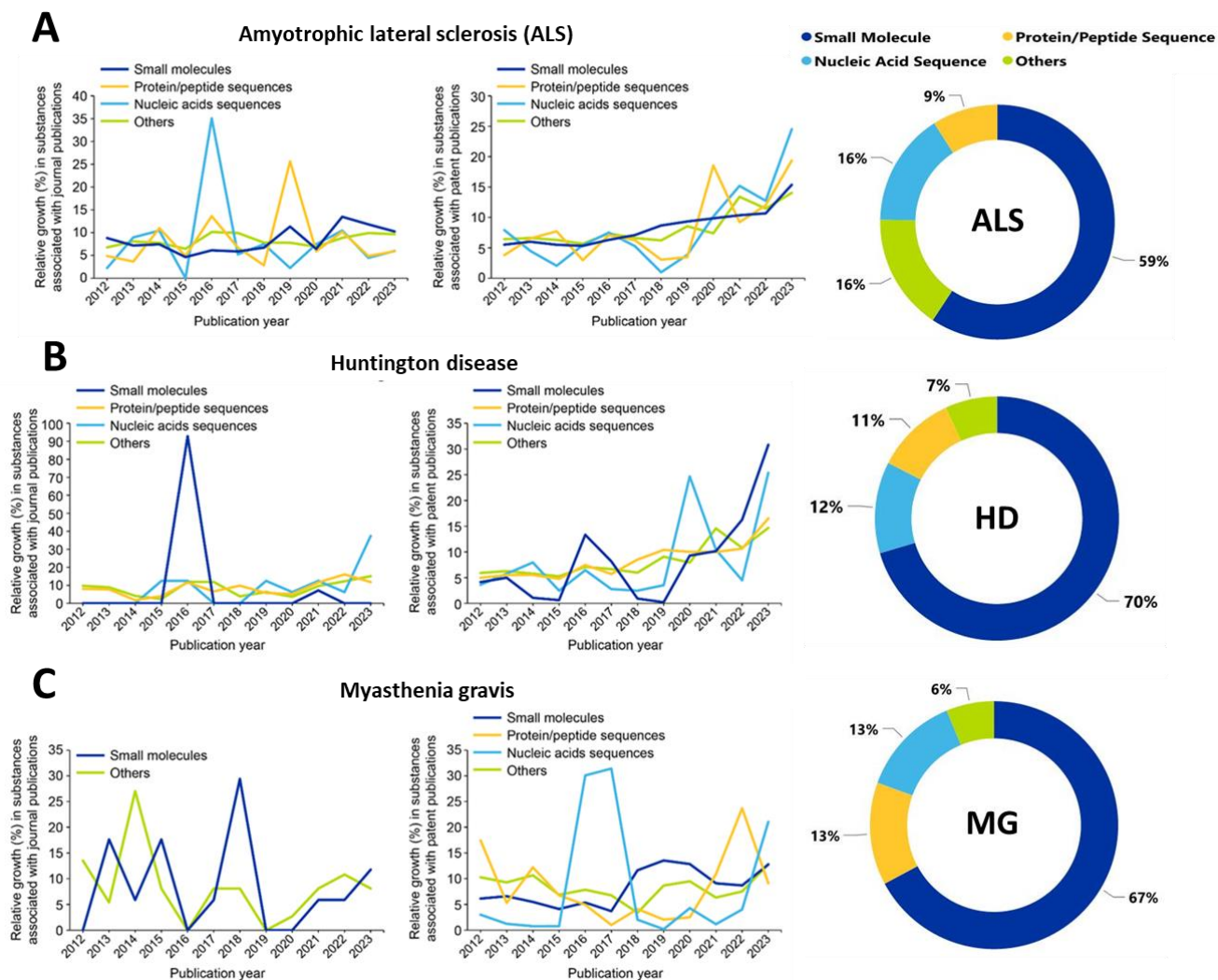
Astrocytes, the most abundant glial cells in the central nervous system, are implicated in ALS and HD through various mechanisms. <sup>799, 804, 805</sup> Reactive astrocytes can secrete toxic molecules, such as pro-inflammatory cytokines and ROS, contributing to neuronal damage and death. Dysfunctional astrocytes also fail to provide adequate support to neurons, impairing their survival and function. Furthermore, mutations in genes like SOD1 and C9orf72, associated with familial ALS cases, have been shown to induce astrocyte dysfunction, exacerbating disease progression. <sup>806-808</sup> Reactive astrocytes in HD exhibit altered morphology and dysregulated function, including impaired glutamate uptake and disrupted calcium signaling. <sup>809-811</sup> Dysfunction of astrocytes contributes to excitotoxicity, oxidative stress, and neuronal dysfunction in HD. Additionally, astrocytes expressing mHTT can release toxic factors, exacerbating neuronal damage and disease progression. <sup>812-814</sup>

Microglia, the resident immune cells of the central nervous system, play dual roles in ALS and HD pathology. <sup>799, 802, 815-817</sup> While initially recruited to sites of neuronal injury to clear cellular debris and promote tissue repair, microglia can become chronically activated and contribute to neuroinflammation and neurotoxicity. Dysregulated microglial responses, characterized by the release of pro-inflammatory cytokines and neurotoxic factors, have been implicated in motor neuron degeneration. Modulating microglial activation and promoting their neuroprotective functions represent potential therapeutic strategies for ALS and HD.

Other neuroglial cells, such as oligodendrocytes and NG2 glia, may also contribute to ALS and HD pathology. <sup>801, 818-820</sup> Oligodendrocyte dysfunction can disrupt myelination and impair neuronal signaling, contributing to cognitive and motor impairments. NG2 glia, also known as oligodendrocyte progenitor cells, respond to injury and participate in remyelination processes. Dysregulation of these neuroglial cell types may exacerbate neuronal dysfunction and degeneration in ALS and HD.

## Overview of substance data

We leveraged our access to the CAS Registry, consisting of data for >250 million substances, and examined substances belonging to diverse classes such as small molecules, protein/peptide sequences, nucleic acid sequences, and others, explored in the documents related to ALS, HD, and MG in the CAS Content Collection (Figure 14). For all three diseases small molecules constituted the largest fraction of explored substances (Figure 14, right column). While for journal publications (Figure 14, left column) the growth in the number of explored substances does not exhibit any particular trend, for patents there is clear upward trend in the number of substances as seen by their relative growths (Figure 14, middle column). All three classes of substances (small molecules, protein/peptide sequences, and nucleic acid sequences) show an increase in patent publications indicative of commercial interest in developing these substances as therapeutics. In particular, the nucleic acid sequence and small molecule sub-class of substances exhibit a marked increase around 2018-2019 for ALS and HD, respectively. Some specific representative substances from the protein/peptide classes are displayed in Figure S3 in the Supporting Information and include immunosuppressive agents such as cyclosporin (CAS RN: 59865-13-3), chemotherapeutic agents such as actinomycin D (CAS RN: 50-76-0) and peptide hormone amylin (CAS RN: 106602-62-4). Other protein/peptide molecules co-occurring frequently with ALS, HD and MG include antibodies such as the CD-52 directed antibody alemtuzumab (CAS RN: 216503-57-0), VEGF-A directed antibody bevacizumab (CAS RN: 216974-75-3), and  $\alpha$ 4 integrin directed antibody natalizumab (CAS RN: 189261-10-7), among others.

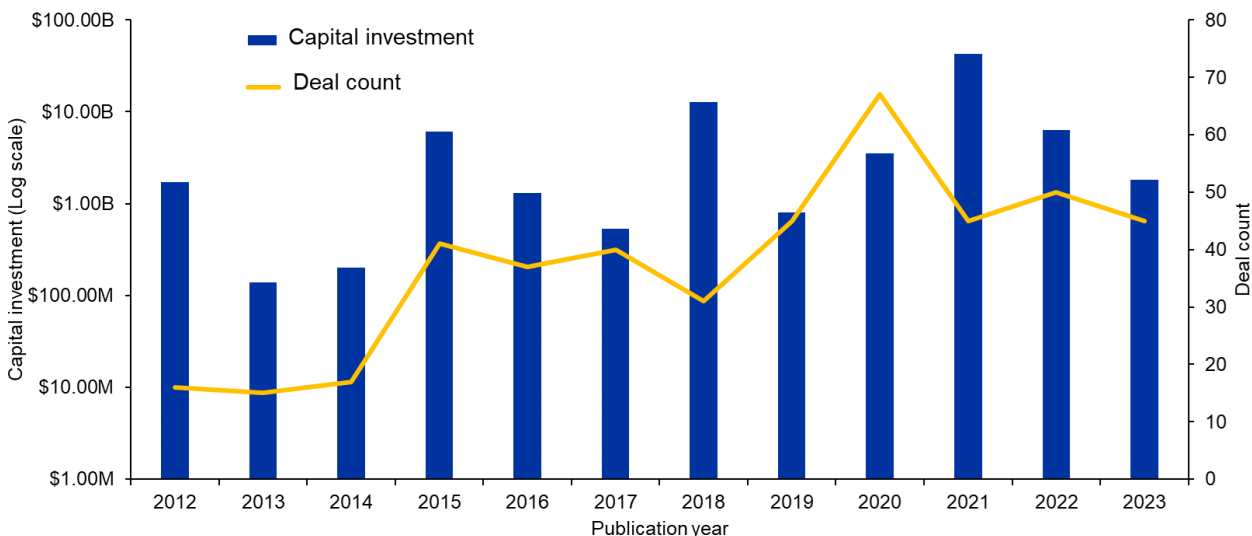


**Figure 14.** Substance data from the CAS Registry associated with (A) amyotrophic lateral sclerosis (ALS), (B) Huntington's disease (HD) and (C) myasthenia gravis (MG). Data includes substances associated with both patent and journal publications sourced from the CAS Registry and the CAS Content Collection for the period 2012-2023.

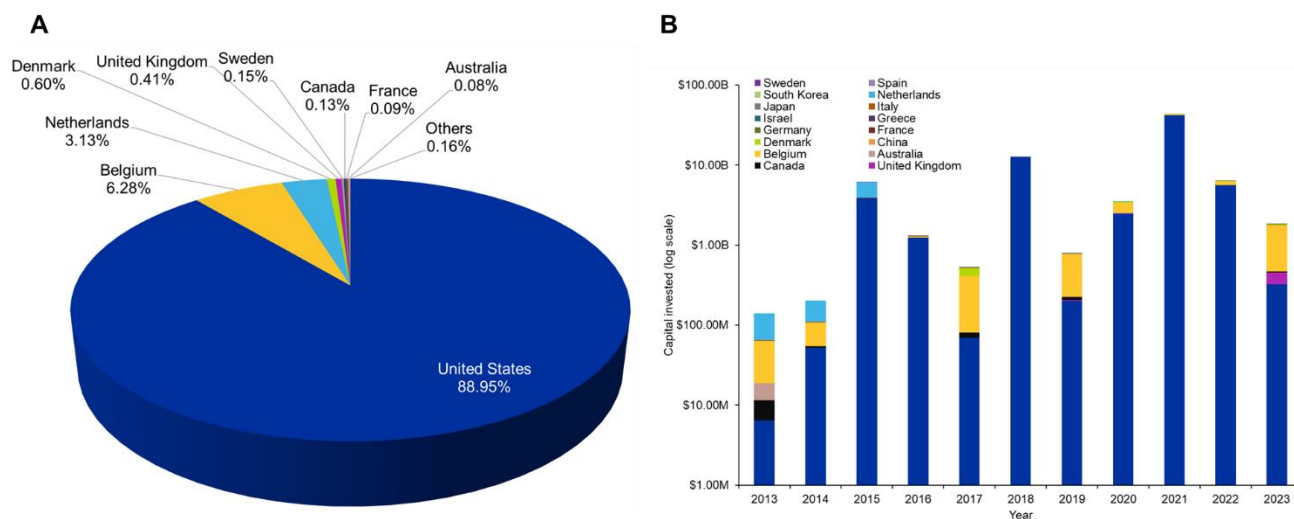
## 4. Capital investment

Capital investment data from Pitchbook – an online platform for investment data, reveals a more or less consistent level of capital that has been invested in the fields of ALS, HD, and MG over the past 10 years (Figure 15) while the number of deals shows a moderate increase during the same period. Interestingly, there is a mild decline in the amount of money invested in this field over the last 3 years (2021-2023) which could be indicative of a slight decrease in commercial interest but the exact reason for this remains unknown. In terms of the geographical distribution of capital investment in the field of rare diseases, the US led with respect to capital investment from 2013 to 2023, followed by Belgium, the Netherlands, and the UK (Figure 16A). The investment in the US is ~14 times that of the next top contributor-Belgium and ~27 times that of

the Netherlands. Growth in the capital investment made from 2013-2023 for the few leading countries/regions (Figure 16B) shows a moderate overall increase till 2021, post which it shows a minor dip till 2023. Of note, the capital investment by Belgium and the UK has increased in the last 2 years (2021-2023).



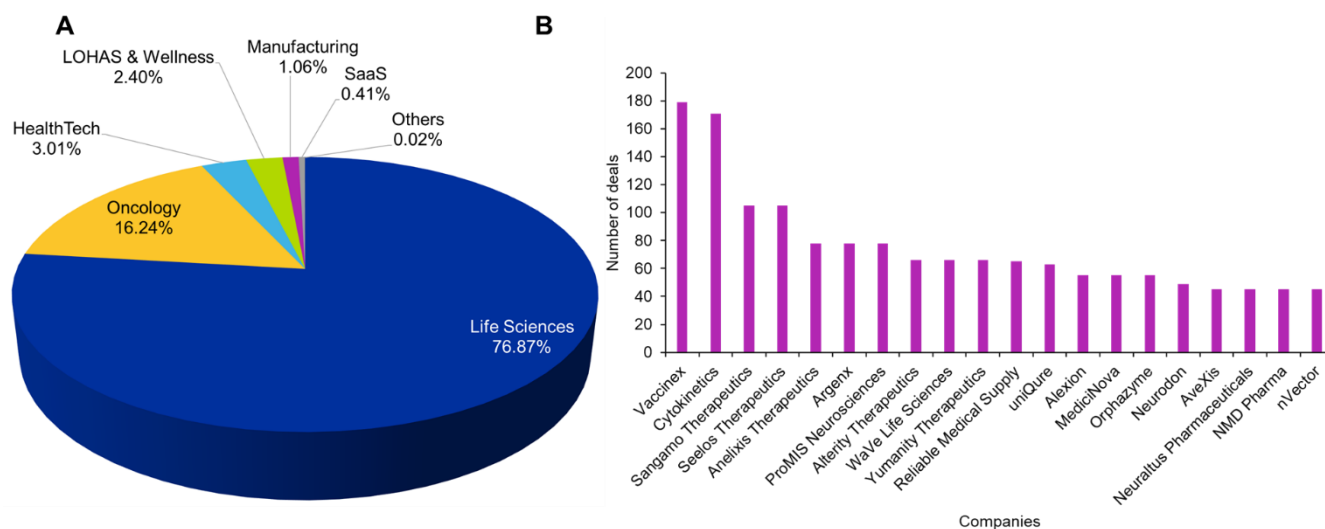
**Figure 15.** Commercial interest in rare diseases (data sourced from PitchBook). Capital invested and deals related to rare diseases (ALS, HD, and MG) for the past decade (2012–2023).



**Figure 16. (A)** Geographical distribution of the number of companies engaged in the field of rare diseases (ALS, HD, and MG). **(B)** Growth in the number of deals in the field of rare diseases by different countries/regions from 2013 to 2022.

A closer look at the data suggests that life sciences-based industries are the top investors followed by oncology-based and health technology industries (Figure 17). Vaccinex has the maximum number of deals in this field. Vaccinex lead drug candidate, pepinemab (CAS RN: 2097151-87-4), blocks semaphorin 4D (SEMA4D), a key driver of neuroinflammation. This drug

has the potential as a disease-modifying treatment for HD, Alzheimer's, and other neurodegenerative diseases and is currently in Phase II clinical trials.<sup>821, 822</sup> Similarly, Cytokinetics is investing in diseases linked to neuromuscular junction such as ALS.<sup>823</sup> Sangamo Therapeutics is working towards the preclinical development of a zinc finger transcriptional repressor targeting the SCN9A gene as a novel therapy for peripheral neuropathic pain which could be effective against ALS.<sup>824</sup> These investment trends indicate the steady interest of companies in the field of rare diseases such as ALS, HD, and MG.



**Figure 17. (A)** Sector-wise distribution of various industry types investing in the field of rare diseases (ALS, HD, and MG) **(B)** Growth in the number of deals in the field of rare diseases by different companies from 2013 to 2022.

## 5. Therapeutic Development Pipelines

### Commercial preclinical development

Nearly 250 substances are being researched and developed preclinically for the treatment of ALS, HD, and MG (Table 4). The vast majority (74%) of these substances are for the treatment of ALS but therapies for the treatment of HD (18%) and MG (8%) are also in the development pipeline (Table 4). A wide range of therapies are being investigated. Small molecule drugs dominate in the total number of drug candidates followed by gene, antibody, RNA, antisense oligonucleotide (ASO), and stem cell therapies, amongst others (Figure 18). Small molecule drugs make up about 50% of the drug candidates for ALS and MG, in contrast to only 10% for HD where gene therapy, RNA interference agents, and other biologic treatments dominate (Figure 18).

**Table 4.** Rare disease therapeutic drug candidates in commercial preclinical development (Source: <https://clinicalintelligence.citeline.com/>)

Therapy type	Drug	Mechanism	Company, location
<b>Amyotrophic lateral sclerosis</b>			
Antibody therapy	VTX-001	Protein degrader, targeting oxidized phosphatidylcholines	VectoryTx, Netherlands
	VTX-002	Protein degrader; TDP-43 inhibitor	VectoryTx, Netherlands
Antisense oligonucleotide therapy	ALS therapy, AcuraStem	PIKFYVE inhibitor	AcuraStem, USA
	ALS therapy, AcuraStem	SYF2 pre-mRNA splicing factor antagonist	AcuraStem, USA
	ALS therapy, AcuraStem	Unc-13 homolog A agonist	AcuraStem, USA
	ALS therapy, Genetic Leap-1	Targeting FLCN	Genetic Leap, USA
	ALS therapy, Maze Therapeutics	UNC13A inhibitor, Protein synthesis inhibitor	Maze Therapeutics, USA
	ALS therapy, Regulus	MicroRNA inhibitor	Regulus Therapeutics, USA
	AMX-114	Calpain 2 inhibitor	Amylyx Pharmaceuticals, USA
	AS-202	Phosphatidylinositol 3-phosphate 5-kinase inhibitor; TDP-43 inhibitor	AcuraStem, USA; Takeda, Japan
	BMD-001	MicroRNA inhibitor	Biorchestra, South Korea
	QRL-202	TDP-43 inhibitor	QurAlis, USA
	QRL-204	Undisclosed	QurAlis, USA
CAR T-cell therapy	ALS therapy, PoITREG	T cell stimulant	PoITREG, Poland
Cognition enhancer, neuroprotectant	ADV-368	Sigma 1 receptor agonist, sigma 2 receptor antagonist	Advantx Pharmaceuticals, USA
	AGS-499	Telomerase stimulant	Neuromagen Pharma, Israel
	Alpha-cyclodextrin derivatives	Arachidonic acid inhibitor, down-regulates the activity of the phosphatidylinositol cycle	ASDERA, USA

ALS therapy, Myrobalan Therapeutics	Colony stimulating factor 1 receptor antagonist	Myrobalan Therapeutics, USA
ALS therapy, Myrobalan Therapeutics	G protein-coupled receptor 17 antagonist	Myrobalan Therapeutics, USA
ALS therapy, Nevrargenics	RAR modulators	Nevrargenics, UK
ALS therapy, WaveBreak	TDP-43 inhibitor	WaveBreak, USA
ALTA-808	TDP-43 inhibitor	Alteron Therapeutics, USA
ALS therapy, Sharp Therapeutics	Undisclosed	Sharp Therapeutics, USA
ALS therapy, Yumanity Therapeutics	Undisclosed	Kineta, USA
ALS therapy, Yumanity Therapeutics	Undisclosed	Merck & Co., USA
ALS therapy, Yumanity Therapeutics	Undisclosed frontotemporal lobar dementia therapy	Kineta, USA
Anhydrous enol-oxaloacetate, Metvital	Undisclosed	MetVital, USA
Antroquinonol	Ras inhibitor	Golden Biotechnology, Taiwan
AS-201	Phosphatidylinositol 3-phosphate 5-kinase inhibitor	AcuraStem, USA
BLX-0279	Undisclosed	Biolexis Therapeutics, USA
BrAD-R13	TrkB tyrosine kinase stimulant	Shanghai Braegen Pharmaceutical, China
BSC-3301	Receptor-interacting serine/threonine kinase 1 inhibitor	BiSiChem, South Korea
FB101	Abl receptor tyrosine kinase inhibitor	1st Biotherapeutics, South Korea
FB418	Abl receptor tyrosine kinase inhibitor	1st Biotherapeutics, South Korea
GP-119	Undisclosed	GliaPharm, Switzerland

	INF-11	Undisclosed	Beijing Joekai Biotechnology, China
	IRX-4204	Retinoid X receptor agonist	Io Therapeutics, USA
	Mitometin	Carnitine palmitoyltransferase 1 inhibitor	2N Pharma, Denmark
	NB-001	Parkin E3 ubiquitin ligase stimulant	NysnoBio, USA
	NMRA-CK1d	Casein kinase 1 inhibitor	Neumora Therapeutics, USA
	NVG-300	Undisclosed	NervGen Pharma, Canada
	NX-210c	Beta 1 integrin agonist	Axoltis Pharma, France
	Resveratrol	Apoptosis stimulant; Reducing agent	Jupiter Neurosciences, USA
	Rifampicin	DNA directed RNA polymerase inhibitor	Medilabo RFP, Japan
	Progranulin	Undisclosed	Alpha Cognition, Canada
	Pur XX-01	Beta-N-acetylglucosaminidase inhibitor	PurMinds NeuroPharma, Canada
	SEL-148,742	Undisclosed	Sharp Therapeutics, USA
	SLS-009	Ubiquitin ligase E3 stimulant; Protein degrader	Seelos Therapeutics, USA
	SMB-001	Undisclosed	Symbinas Pharmaceuticals, USA
	SRT-055 series	Mitogen-activated protein kinase kinase 5 inhibitor	Seal Rock Therapeutics, USA
Exosome therapy	AB-126	Reduces neuroinflammation, promotes neuroprotection, and stimulates neuroregeneration	Aruna Bio, USA
Fusion protein therapy	PF-1802	Undisclosed	Immunoforge, South Korea
Fusion protein therapy; Cytokine therapy	COYA-302	CD80 antagonist; CD86 antagonist; Immune checkpoint modulator; Interleukin 2 receptor agonist; T cell inhibitor; T cell stimulant	Coya Therapeutics, USA; Dr. Reddy's Laboratories, India
Gene therapy	ABO-201	Genome editing, C9orf72 inhibitor	Arbor Biotechnologies, USA
	ABO-202	Genome editing, stathmin 2 agonist	Arbor Biotechnologies, USA; 4D Molecular Therapeutics, USA
	ABO-204	Genome editing, superoxide dismutase-1 inhibitor	Arbor Biotechnologies, USA



ALS therapy, AviadoBio	Gene supplementation, PGRN	AviadoBio, UK
ALS therapy, Hui-Gen Therapeutics	Genome (DNA) editing	HuidaGene Therapeutics, China
ALS therapy, Korro Bio	Genome editing, TDP-43 stimulant	Korro Bio, USA
ALS therapy, Maze Therapeutics	Ataxin 2 inhibitor, Protein synthesis inhibitor	Maze Therapeutics, USA
ALS therapy, NeuShen Therapeutics	SOD1 silencing	NeuShen Therapeutics, China
ALS therapy, NeuShen Therapeutics	Undisclosed	NeuShen Therapeutics, China
ALS therapy, Prime Medicine	Genome editing	Prime Medicine, USA
ALS therapy, Sangamo Therapeutics	C9orf72 inhibitor, ZFP-TF gene therapy	Alexion, UK; Sangamo Therapeutics, USA
ALS therapy, Scribe Therapeutics	CRISPR-Cas9 Genome editing	Biogen, USA; Scribe Therapeutics, USA
AMA-007	Undisclosed	Amarna Therapeutics, Netherlands
ANEW-202	Undisclosed	Anew Medical, USA
ANL-303	Undisclosed	ANLBIO, South Korea
CTx-1000	TDP-43 inhibitor	Celosia Therapeutics, Australia
CTx-2000	TDP-43 inhibitor	Celosia Therapeutics, Australia
CTx-TFEBx	Undisclosed	Coave Therapeutics, France
ET-101	Caveolin stimulant	Eikonoklastes Therapeutics, Japan
NM-301	Undisclosed	Helixmith, South Korea; Neuromyon, South Korea
NM-302	Undisclosed	Helixmith, South Korea
NXL-003	Undisclosed	NeuExcell Therapeutics, USA
PBAL-05	Undisclosed	Passage Bio, USA
SNUG-01	Unidentified pharmacological activity	Sineugene, China
SOL-257	TDP-43 inhibitor	SOLA Biosciences, USA

	SOL-258	Undisclosed	SOLA Biosciences, USA
	VY-SOD102	Superoxide dismutase-1 inhibitor	Voyager Therapeutics, USA
Monoclonal antibody	BT-3814	TDP-43 inhibitor	BioArctic Neuroscience, Sweden
	LP-005	C5 and C3 inhibitor	LongBio Pharma, China
	ND-3014	TDP-43 inhibitor	BioArctic Neuroscience, Sweden
	PAS-003	Undisclosed	Pasithea Therapeutics, USA
	PMN-267	TDP-43 inhibitor	ProMIS Neurosciences, Canada
	Anti-TDP-43 antibody, Prothena	TDP-43 inhibitor	Bristol-Myers Squibb, USA; Prothena, Ireland
Monoclonal antibody, humanized	ALS therapy, Mabyon	Undisclosed	Mabyon, Switzerland; SciNeuro Pharmaceuticals, USA
	ARGX-119	Muscle-specific receptor kinase stimulant	Argenx, Netherlands
	EME-023	Undisclosed	Epsilon Molecular Engineering, Japan
	Foralumab	CD3 antagonist	Tiziana Life Sciences, UK
	Hu-GNK-301	Undisclosed	GeNeuro, Switzerland
	NC-B8	IgG4 inhibitor	ENCEFA, France
	NI-308	Immunostimulant	Neurimmune, Switzerland
Monoclonal antibody, murine	ALS therapy, ProMIS Neurosciences	Protein kinase C inhibitor, RACK1-targeting antibodies	ProMIS Neurosciences, Canada
	ALS therapy, ProMIS Neurosciences	SOD1-targeting antibodies, Superoxide dismutase-1 inhibitor	ProMIS Neurosciences, Canada
Neuroprotectant	AKV-9	Protein aggregation inhibitor	Akava Therapeutics, USA
	ALS therapy, Amylyx Pharmaceuticals	Bax and Bak protein inhibitors	Amylyx Pharmaceuticals, USA
	ALS therapy, Dewpoint Therapeutics	TDP-43 inhibitor	Dewpoint Therapeutics, USA
	ALS therapy, Genetic Leap-2	RNA binder targeting FLCN	Genetic Leap, USA

ALS therapy, ImmunoBrain Checkpoint	Immune checkpoint modulator	ImmunoBrain Checkpoint, Israel
ALS therapy, Miramoon Pharma	Ryanodine receptor antagonist; Oxygen scavenger	Miramoon Pharma, Spain
ALS therapy, Nine Square Therapeutics	Undisclosed	Nine Square Therapeutics, USA
ALS therapy, NRG Therapeutics	mPTP inhibitor	NRG Therapeutics, UK
ALS therapy, Origami Therapeutics	Undisclosed	Origami Therapeutics, USA
ALS therapy, Path Biotech	Undisclosed	Path Biotech, USA
ALS therapy, Pikralida	Undisclosed	Pikralida, Poland
ALS therapy, PrecisionLife	Undisclosed	PrecisionLife, UK
ALS therapy, reMYND	Undisclosed	reMYND, Belgium
ALS therapy, Treventis	TDP-43 inhibitor	Treventis, Canada
ALS therapy, Zhittya Genesis Medicine	Fibroblast growth factor 1 agonist	Zhittya Genesis Medicine, USA
Amisodin	Superoxide dismutase-1 inhibitor	Pioneer of Rare Genopathies, South Korea
ALS therapy, Verge Genomics-1	Undisclosed	Eli Lilly, USA
ALS therapy, Verge Genomics-1	Undisclosed	Verge Genomics, USA
ATC-104	Protein degrader; Sequestosome 1 stimulant; TDP-43 inhibitor	Autotac Bio, South Korea
ATH-1105	Undisclosed	Athira Pharma, USA
BEN-34712	Retinoic acid alpha receptor agonist; Retinoic acid beta receptor agonist	BenevolentAI, UK
BIOIO-1001	Sirtuin 3 stimulant	BIOIO, USA
BL-002, Bloom Science	Microbiome modulator, live microorganisms	Bloom Science, USA
BREN-02	Undisclosed	BrainEver, France
BT-003	Undisclosed	BAKX Therapeutics, USA
CB03-154	Potassium channel agonist	Shanghai Zhimeng Biopharma, China

Donepezil hydrochloride + nebivolol hydrochloride	Acetylcholinesterase inhibitor; Beta 1 adrenoceptor antagonist	Dr. Noah Biotech, South Korea
EVT-8683	Protein degrader; Translation initiation factor 2B stimulant	Bristol-Myers Squibb, USA
EVT-8683	Protein degrader; Translation initiation factor 2B stimulant	Evotec, Germany
FB-1002	Unidentified pharmacological activity	4B Technologies, China
FP-802	Transient receptor potential cation channel M antagonist; NMDA receptor antagonist	FundaMental Pharma, Germany
Glycolic acid + D-lactic acid	Undisclosed	Neurevo, Germany
HLX-94	Undisclosed	Shanghai Fosun Pharmaceutical, China
HS-03	Undisclosed	Foshan Rexiu Biotechnology, China
ILB	Undisclosed	TikoMed, Sweden
iN3011-K23	Undisclosed	iN Therapeutics, South Korea
IPG 008	Hydrolase inhibitor	Nanjing Immunophage Biotech, China
KFRX-03	Tyrosine kinase inhibitor (TKI)	KeifeRx, USA
LAUR-301	Glial cell derived neurotrophic growth factor agonist	Lauren Sciences, USA
M-102	Undisclosed	Acclipse Therapeutics, USA
mecobalamin	Vitamin B12 agonist	Eisai, Japan
MP-101	Undisclosed	Mitochon Pharmaceuticals, USA
MT-2	Undisclosed	MimeTech, Italy
Nibrozetone (RRx-001)	NLRP3 inhibitor and Nrf2 upregulator	EpicentRx, USA
OC-514	Undisclosed	Oncocross, South Korea
ORY-4001	Histone deacetylase 6 inhibitor	Oryzon, Spain
otaplimastat	Undisclosed	Shin Poong Pharmaceutical, South Korea
P-202320	Undisclosed	EDDC, Singapore
Pimicotinib	Colony stimulating factor 1 receptor antagonist; Immuno-oncology therapy	Sperogenix Therapeutics, China

	Riluzole	Dopamine receptor agonist; Glutamate antagonist; Voltage-gated sodium channel antagonist	Brain Trust Bio, USA
	RT-1968	Undisclosed	Raya Therapeutic, Canada
	RT-1972	Undisclosed	Raya Therapeutic, Canada
	RT-1978	Undisclosed	Raya Therapeutic, Canada
	RT-1999	Undisclosed	Raya Therapeutic, Canada
	RT-2010	Undisclosed	Raya Therapeutic, Canada
	RVL-027	Undisclosed	Unravel Biosciences, USA
	SAM-001	Transient receptor potential cation channel member 1 agonist	Samsara Therapeutics, UK
	SAM-19272	GTPase stimulant	Samsara Therapeutics, UK
	SNP-210	TDP-43 inhibitor	SciNeuro Pharmaceuticals, USA
	S-oxprenolol	Beta adrenoreceptor antagonist	Actimed Therapeutics, UK
	TDP-43 inhibitors, Aquinnah	TDP-43 inhibitor	Aquinnah Pharmaceuticals, USA; Roche, Switzerland
	WD-920	Undisclosed	Zhejiang Wenda Medical Technology, China
	Y-8	Undisclosed	Neurodawn Pharmaceutical, China
RNA interference	ALN-SOD1	Gene expression inhibitor, Superoxide dismutase-1 inhibitor	Alnylam, USA; Regeneron, USA
	ALS therapy, AviadoBio	Gene expression inhibitor, ATXN2	AviadoBio, UK
	ALS therapy, AviadoBio	Gene expression inhibitor, c9orf72	AviadoBio, UK
	ALS therapy, AviadoBio	Gene expression inhibitor, SOD1	AviadoBio, UK
	AMT-161	C9orf72 inhibitor; Gene expression inhibitor	uniQure, Netherlands
Stem cell therapy	ALS therapy, Neuroplast	Undisclosed	Neuroplast, Netherlands
	ALS therapy, Takeda	Undisclosed	Takeda, Japan
	BMS-xxxx	Not applicable	Bristol-Myers Squibb, USA; Evotec, Germany
	Cognistem	Undisclosed	Amniotics, Sweden
	CUR-201	Undisclosed	Curamys, South Korea

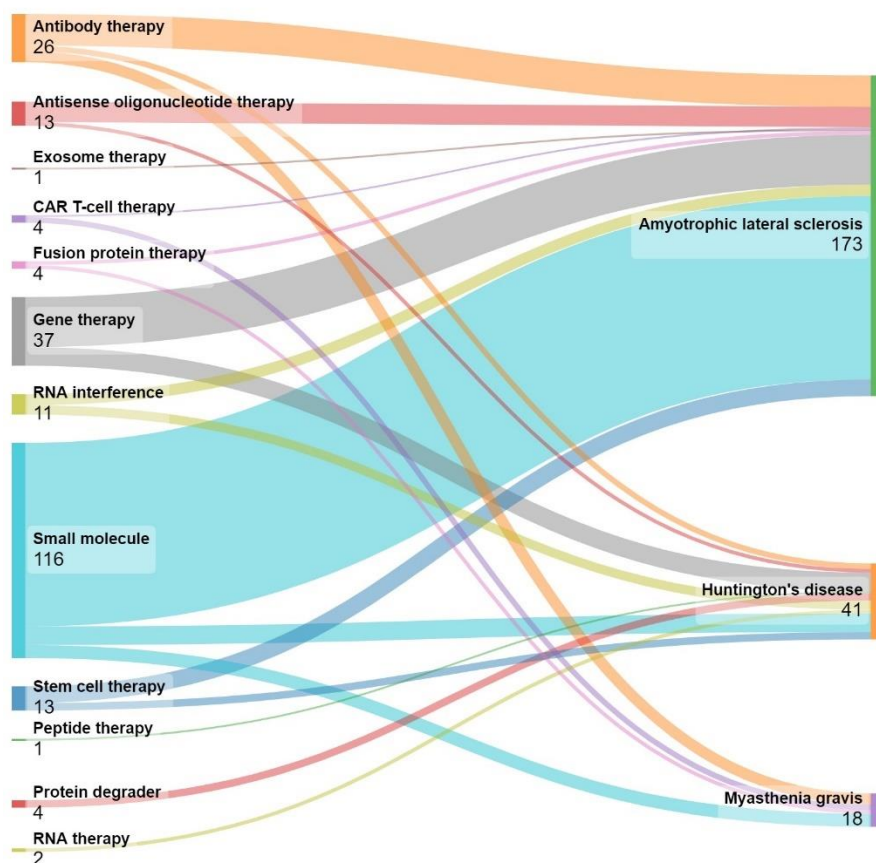
	CUR-202	Undisclosed	Curamys, South Korea
	CUR-204	Undisclosed	Curamys, South Korea
	hNPC-02	Not applicable	Hopstem Biotechnology, China
	NCP-01	Undisclosed	Hemostemix, Canada
<b>Huntington's disease</b>			
Antibody therapy	VTX-003	mHTT protein inhibitor, selectively binds mutant HTT (and not normal HTT) to clear mutant HTT	VectorY Therapeutics, Netherlands
Anti inflammatory	IC 100-05	Inflammasome ASC Inhibitor	ZyVersa Therapeutics, USA
Antisense oligonucleotide therapy	Huntington's disease therapy, Ionis	Undisclosed	Ionis, USA; Roche, Switzerland
Cognition enhancer	ASK-005	Arachidonic acid inhibitor	ASDERA, USA
Cognition enhancer, neuroprotectant	Huntington's disease therapy, MitoRx	Reverses mitochondrial dysfunction	MitoRx Therapeutics, UK
	NP-001	Oxidizing agent	Neuraltus, USA; Neuvivo, USA
Gene therapy	ET-101	Caveolin stimulant	Eikonoklastes Therapeutics, USA
	Huntington's disease therapy, Hudiagene	DNA editing, CRISPR	HuidaGene Therapeutics, China
	Huntington's disease therapy, Life edit	Genome editing targeting both the T and C alleles of an exonic SNP in the HTT gene utilizing AAV vector	Life Edit Therapeutics, USA
	Huntington's disease therapy, Passage	Deliver functional gene utilizing AAV vector	Passage Bio, USA
	Huntington's disease therapy, Prime	Genome editing	Prime Medicine, USA
	NXL-002	Regenerates neurons	NeuExcell Therapeutics, USA; Spark Therapeutics, USA
	SOL175	Reduces abnormally folded protein	SOLA Biosciences, USA
	INT41	mHTT protein inhibitor, selectively binds to mHTT protein	Vybion, USA
	SOL176	Reduces abnormally folded protein	SOLA Biosciences, USA
	TAK-686	Zinc finger nucleases can target regions of DNA to modify them or stop RNA from being made	Sangamo Therapeutics, USA; Takeda, Japan

	ReS18-H	Restores function and improve survival of medium spiny neurons leading to reactivation of corticostriatal transmission	reMYND, Belgium
Monoclonal antibody, humanized	ATLX-1095	HTT inhibitor	Alchemab Therapeutics, UK
Monoclonal antibody	Huntington's disease therapy ProMIS	Targeting protein RACK-1, Protein kinase C inhibitor	ProMIS Neurosciences, USA
mRNA therapy	Anima Huntingtin translation inhibitor	Gene expression inhibitor	Anima Biotech, USA; Takeda, Japan
Neuroprotectant	AJ-201	Transcription factor Nrf2 stimulant	Avenue Therapeutics, USA
	Huntington's disease therapy, BPGbio	Undisclosed	BPGbio, USA
	Huntington's disease therapy LoQus23	DNA damage repair	LoQus23 Therapeutics, USA
	M102	Activates Nrf2 and HSF1	Acclipse Therapeutics, USA
	TQS-168	PPARG coactivator 1 alpha agonist	Tranquis Therapeutics, USA
Peptide therapy	TT-P34	Activates pathways that can bypass mHTT to reactivate CREB	Teitur Trophics, Denmark
	NT-0100	Gene expression inhibitor	NeuBase Therapeutics, USA
PROTAC	Huntington's disease therapy, Arvinas	HTT inhibitor, E3 ubiquitin ligase stimulant, protein degrader	Arvinas, USA
Protein degrader	ORI-113	HTT protein degrader	Origami Therapeutics, USA
Protein conformation correctors	ORI-503	HTT proteins conformation corrector	Origami Therapeutics, USA
Protein degrader	SLS009	Protein targeted autophagy	Seelos Therapeutics, USA
RNA interference	ALN-HTT	Gene expression inhibitor	Alnylam, USA
	ALN-HTT02	Gene expression inhibitor	Alnylam, USA
	Huntington's disease therapy Atalanta	Gene expression inhibitor	Atalanta Therapeutics, USA; Biogen, USA
	Huntington's disease therapy, novartis	Utilized a small hairpin RNA or short hairpin RNA for gene expression inhibition	Novartis, Switzerland; Voyager Therapeutics, USA
	OCCT-HTT siRNA	Gene expression inhibitor, HTT inhibitor	Ophidion, USA

RNA therapy	SMDG-HD11	Undisclosed	S. M. Discovery Group, UK
Stem cell therapy	Debamestrocel	Glial cell derived neurotrophic growth factor agonist	BrainStorm Cell Therapeutics, USA
	HB AdMSC	Undisclosed	Hope Biosciences, USA
	Huntington's disease therapy, trailhead	Mesenchymal Stem Cells	Trailhead Biosystems, USA
	SC-379	Glial progenitor cells	Sana Biotechnology, USA
Undisclosed	Huntington's disease therapy Aitia	Undisclosed	UCB, Belgium; Aitia, USA
<b>Myasthenia gravis</b>			
CAR T-cell therapy	CABA-201	T cell stimulant	Cabaletta Bio, USA
	Equecabtagene autoleucl	Immuno-oncology therapy; T cell stimulant	Nanjing IASO Biotechnology, China
	KYV-101	T cell stimulant	Kyverna Therapeutics, USA
Fusion protein therapy	Myasthenia gravis therapy, Rongchang Pharmaceuticals	Lymphocyte stimulant	Rongchang Pharmaceuticals, China
	TOL-2	Undisclosed	Toleranzia, Sweden
Monoclonal antibody	EA-5	Complement inhibitor	Lan-yi Therapeutics, China
	LP-005	C5a inhibitor	LongBio Pharma, China
	MV-2C2	Survivin inhibitor	MimiVax, USA
Monoclonal antibody, humanized	CAN-106	Complement factor C5 inhibitor	CANbridge Life Sciences, China
	MIL-62	CD20 antagonist; Immuno-oncology therapy	Beijing Mabworks Biotech, China
	Pozelimab	C5a inhibitor	Regeneron, USA
Musculoskeletal therapy	BHV-1310	Protein degrader	Biohaven, British Virgin Islands
	Myasthenia gravis therapy, BioCryst Pharmaceuticals	Complement factor C5 inhibitor	BioCryst Pharmaceuticals, USA
	CNP-MYG	B-cell inhibitor; CD8 antagonist	Cour Pharmaceuticals, USA



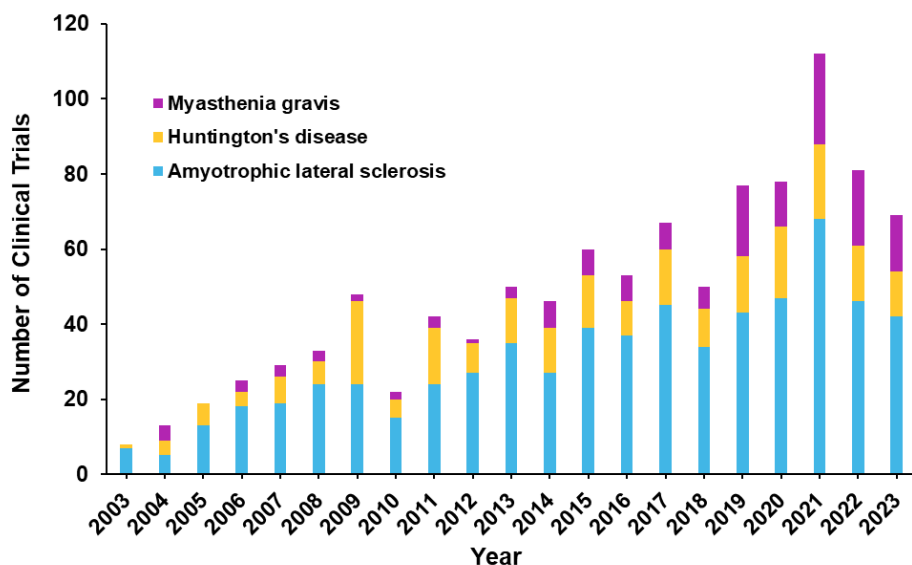
CV-MG-02	Immunostimulant	CuraVac, USA
Myasthenia gravis vaccine, ImCyse	Immunosuppressant	ImCyse, Belgium
Myasthenia gravis therapy, NovelMed Therapeutics	Complement inhibitor	NovelMed Therapeutics, USA
prednisone, Sarcomed AB	Glucocorticoid agonist	Sarcomed AB
ZP-10068	Complement C3 inhibitor	Alexion, UK; Zealand Pharma, Denmark



**Figure 18.** Preclinical drug therapy candidates and their respective rare disease indication currently in the development pipeline

## Clinical Trials

Clinical trials researching the treatment of rare diseases ALS, HD, and MG are explored in this section to gain an overall view of the past and current state of clinical development. Over 1000 clinical trials have been registered on the US National Institutes of Health (NIH) clinical trial website over the last 10 years for these rare diseases. In terms of sheer number, ALS has the highest number of registered clinical trials, followed by HD and MG.<sup>825</sup> Figure 19 shows an oscillating curve for clinical trials of these rare disease, between the years 2013 to 2023. ALS clinical trials gradually increase over the years, while HD's stayed more consistent. One note of interest is that the number of clinical trials for MG stay consistent through 2018 and then triple into 2019, maintaining consistency moving forward. One contributing factor to this surge is the US FDA approval of Solaris (eculizumab) in 2017, the first for MG, increasing industry-sponsored clinical research to monitor and develop new therapy options for patients with this rare disease.



**Figure 19.** Number of clinical trials by rare disease indications for years 2003-2023

Further analysis of these rare disease clinical trials reveals that nearly half of all trials for the different indications are not phased (Figure 20A). The phase that contains the next largest group of trials is Phase II studies for ALS and HD and Phase III studies for MG. Nearly half of all clinical trials for ALS, HD, and MG have been completed (Figure 20B). The status with the next largest group of trials is the recruiting status, which is encouraging as new clinical trials are created and carried out to research the treatment of these rare diseases, offering hope to patients worldwide.

### A

Rare disease indication	Early Phase I	Phase I	Phase I/II	Phase II	Phase II/III	Phase III	Phase IV	NA
Amyotrophic lateral sclerosis	1%	10%	7%	19%	4%	7%	1%	51%
Huntington's disease	2%	14%	6%	22%	0%	1%	1%	53%
Myasthenia gravis	2%	3%	2%	16%	4%	26%	2%	45%

### B

Rare disease indication	Not yet recruiting	Recruiting	Active	Completed	Withdrawn/Terminated/Suspended
Amyotrophic Lateral Sclerosis		5%	21%	6%	55%
Huntington's disease		4%	19%	6%	61%
Myasthenia gravis		9%	38%	7%	39%

**Figure 20.** Percentage of rare disease clinical trials in various: (A) phases; (B) statuses.

Finally, representative clinical trials examining rare disease therapeutics are highlighted in Table 5 categorized by rare disease indication and therapy type. These are examined in further detail below to showcase a variety of promising therapeutic strategies, interventions, and targeted conditions in clinical development along with their status, phase, and any published results.

Therapy types in clinical development highlighted below for the treatment of ALS include cell and gene therapies along with small molecule drugs. An autologous T-cell therapy, RAPA-501, was developed by RAPA Therapeutics to address the lack of compounds for treatment of

neuroinflammation in ALS and is currently recruiting for a Phase II/III clinical trial (NCT04220190). This trial is an expansion cohort that was added to access RAPA-501 cells efficiency in standard-risk ALS patients. In this ongoing clinical trial, RAPA-501 cells were found to be safe, have multiple anti-inflammatory effects, and showed early signs of preserving pulmonary function.<sup>826</sup> RAPA-501 cells express both the TREG and Th2 transcription factors forkhead box P3 (FOXP3) and GATA-binding protein 3 (GATA3), are enriched for expression of the T-cell homing molecule CD103 and the ATP ectonucleotidase molecules CD39 and CD73.<sup>827</sup> They also suppress effector T-cell inflammatory molecules and CNS microglial cell inflammatory molecules.<sup>827</sup> RAPA-501 cells are also available through an expanded access clinical trial (NCT06169176) to patients living with high-risk ALS and not eligible for other ALS clinical trials. These trials will research therapy feasibility, safety, and efficacy including biomarker measurements for neuroinflammation.

Gene therapy agent, AMT-162, developed by UniQure Biopharma, is not yet actively recruiting but will soon start a clinical trial, NCT06100276, to research the use of AMT-162 in patients with rapidly progressive ALS and SOD1 mutations. AMT-162, a one-time treatment, is comprised of a recombinant AAVrh10 vector that expresses a miRNA targeting the SOD1 gene. This clinical trial will research the safety and efficacy of AMT-162, evaluating if it will silence the expression of mutant SOD1 and improve the course of ALS.<sup>828</sup>

Small molecule drug FB1006, fully discovered and developed using artificial intelligence (AI), is being advanced as a new potential treatment for ALS. 4B Technologies with collaborative efforts completed the development process of FB1006, from target identification and compound screening to patient enrollment, in less than 2 years.<sup>829</sup> Phase IV clinical trial NCT05923905 recently completed enrollment of 64 patients which will evaluate the efficacy of safety of FB1006 in the treatment of ALS patients. The trial is being conducted at the Third Hospital of Peking University and is expected to complete double-blind dosing in August 2024, followed by 1-year of clinical observation in February 2025.<sup>829</sup>

The last two small molecules we highlight for the treatment of ALS are both part of the HEALEY ALS Platform trial (NCT04297683). This trial is a perpetual multi-center, multi-regimen clinical trial evaluating the safety and efficacy of ALS treatments. There is a single master protocol dictating the conduct of the trials with each regimen sharing placebo patients. The first drug ibudilast is being investigated by MediciNova. A Phase I/II clinical trial NCT02714036 revealed ibudilast to be safe and showed no drug related severe adverse reactions. However, the tolerability was limited due to gastrointestinal side effects, fatigue, and insomnia.<sup>830</sup> Data from another completed Phase II study NCT02238626 showed ibudilast, when in combination with approved therapy Rilutek showed a marked increase in number of patients who saw no functional decline in six months compared to Rilutek alone and it also helped increase patient lifespan.<sup>831</sup> Ibudilast will also be investigated in a Phase II/III clinical trial (NCT04057898) currently recruiting and enrolling up to 230 participants across the USA and Canada to evaluate the efficacy, safety, and tolerability of ibudilast for 12 months followed by a 6-month open-label extension phase. Ibudilast is a glial attenuator that suppresses pro-inflammatory cytokines IL-1 $\beta$ , TNF- $\alpha$ , and IL-6, and may upregulate the anti-inflammatory cytokine IL-10.<sup>832</sup> It has additionally been shown to be a toll-like receptor 4 antagonist that may contribute to its attenuation of neuroinflammation.<sup>832</sup> Ibudilast received both fast track<sup>833</sup> and orphan drug designations<sup>834</sup> in 2016 from the US FDA for treating ALS.

The second small molecule drug pridopidine (CAS RN: 346688-38-8), also a regimen in the HEALEY ALS Platform trial (NCT04297683), was investigated in a completed Phase II/III

clinical trial (NCT04615923). While there was no significant improvements in the primary outcome measures of disease progression and mortality rate, there were positive results reported for secondary outcome measures such as improved respiratory and speech measurements. Improvement in disease progression and neurofilament light levels (biomarker for neuronal injury) for rapidly declining ALS participants who were early in the disease were also seen. Pridopidine is also available through an expanded access clinical trial (NCT06069934) for up to 200 patients with ALS who are ineligible for other clinical trials. Pridopidine is a highly selective sigma-1 agonist. Sigma-1 receptor is highly expressed in the brain and CNS and activation by pridopidine stimulates multiple cellular pathways, including autophagy, which are essential to neuronal function and survival, and may lead to neuroprotective effects.<sup>835</sup> Prilenia Therapeutics was granted orphan drug designation for pridopidine in 2021 from the US FDA for treating ALS.<sup>836</sup>

Highlighted therapies in clinical development for the treatment of HD include antisense oligonucleotides, cell-based, and monoclonal antibody therapies along with computer based cognitive stimulation and small molecule agents. One such antisense oligonucleotide (ASO) therapeutic is Wave Life Sciences' gene silencing therapeutic, WVE-003. Interim results for Phase I/Phase II clinical trial NCT05032196, reveals that a single dose of WVE-003 (30 or 60 mg) led to a mean 35% reduction in mHTT in the cerebrospinal fluid compared to a placebo.<sup>837</sup> More upcoming trial findings are expected by June 2024. Phase I and Phase II/Phase III clinical trials (NCT02728115 and NCT04219241) assessing the safety and efficacy of Cellavita's NestaCell, a stem cell therapy derived from immature human dental pulp, are currently active for the treatment of HD. Another Phase III clinical trial (NCT06097780) researching NestaCell is not yet recruiting but has an estimated start date of June 2024 and will also investigate the efficacy and safety of NestaCell. A previous Phase I clinical trial revealed no serious adverse events and improved HD motor symptoms with the use of NestaCell in the treatment of HD.<sup>838</sup>

Another treatment of HD utilizing a computer based cognitive rehabilitation program is being researched by the Santa Cre Hospital in Spain. Researchers are currently recruiting for their study (NCT05769972) to examine the use of this method in patients with HD with expectations that the program will have a greater beneficial effect on the cognitive status of HD patients compared to control modalities such as music therapy. Monoclonal antibody therapeutic Pepinemab, a semaphorin 4D blocking antibody developed by Vaccinex, was researched in a Phase II clinical trial (NCT02481674) to determine safety, tolerability, pharmacokinetics, and efficacy. While this trial did not meet its primary endpoints, it did have a favorable safety profile, showed a reduction in brain atrophy, and improvement in decline in brain metabolic activity that is typically seen in HD progression.<sup>821</sup>

Lastly for the treatment of HD, Sage Therapeutics' small molecule drug SAGE-718 is currently recruiting for their Phase II/Phase III clinical trials. These trials (NCT05107128, NCT05358821, and NCT05655520) will investigate the safety, tolerability, and efficacy of these drugs for the treatment of HD. SAGE-718, a NMDA receptor positive allosteric modulator has completed initial single and multiple ascending dose clinical studies, where it demonstrated efficacy in disease-relevant populations.<sup>839</sup> In addition, SAGE-718 was granted both FDA Fast track designation<sup>840</sup> in 2022 and FDA Orphan Drug Designation in 2023.<sup>841</sup>

Therapy types in clinical development highlighted for the treatment of MG include antigen, cell, and fusion protein therapies along small molecule drugs. COUR Pharmaceutical is developing CNP-106, an antigen specific therapeutic designed to prevent immune mediated neuromuscular destruction and aims to reprogram the immune system to address the immunological root cause

of myasthenia gravis.<sup>842</sup> The not yet recruiting Phase I/II clinical trial (NCT06106672) will enroll up to 54 adult patients and assess the treatment's safety, tolerability, pharmacological properties, and efficacy. Descartes-08, a mRNA CAR T-cell therapy expressing a chimeric antigen receptor directed to B-cell maturation antigen is currently recruiting for an ongoing Phase II clinical trial (NCT04146051). Results from the Phase IIa portion of the study revealed that Descartes-08 is well tolerated and participants saw meaningful improvement in MG disease scorings.<sup>843</sup> Descartes-08 was granted Orphan Drug Designation by the US FDA for the treatment of MG in 2024.<sup>844</sup>

RemeGene is currently recruiting for its Phase III clinical trial (NCT05737160) investigating fusion protein Telitacicept for the treatment of MG. Telitacicept is constructed with the extracellular domain of the transmembrane activator and calcium modulator and cyclophilin ligand interactor receptor and the fragment crystallizable domain of immunoglobulin G.<sup>845</sup> Telitacicept targets two cell-signaling molecules critical for B-lymphocyte development: B-cell lymphocyte stimulator and a proliferation inducing ligand, which allows it to effectively reduce B-cell mediated autoimmune responses. Results from a previous Phase II study showed that Telitacicept improved MG symptoms and had a good safety profile.<sup>845</sup> Lastly, we look at a small molecule factor D inhibitor developed by Alexion Pharmaceuticals. ALXN2050 is currently being investigated in an active Phase II clinical trial (NCT05218096) for its ability to improve the disease symptoms and the daily life of people with MG along with its safety. Seventy patients are enrolled with a study completion date of late 2025, results will be forthcoming soon.

**Table 5.** Highlighted rare disease therapeutic clinical trials

Therapy type	Intervention	CAS RN	Sponsor, location	Status	Phase	NCT Number
<b>Amyotrophic lateral sclerosis</b>						
Cell therapy	RAPA-501	N/A	Rapa Therapeutics, USA	Recruiting	Phase II/ III	NCT04220190
Gene therapy	AMT-162	N/A	UniQure Biopharma, Netherlands	Not yet recruiting	Phase I/ II	NCT06100276
Small molecule	FB1006	N/A	4B Technologies, China	Recruiting	Phase IV	NCT05923905
Small molecule	Ibudilast (MN-166)	50847-11-5	MediciNova, USA	Completed	Phase I/ II	NCT02714036
					Phase II	NCT02238626
				Recruiting	Phase II/ III	NCT04057898
Small molecule	Pridopidine	346688-38-8	Prilenia, USA	Recruiting	Phase II/ III	NCT04297683
				Completed	Phase II/ III	NCT04615923
				Available	Expanded access	NCT05281484
<b>Huntington's disease</b>						
Antisense oligo-nucleotide	WVE-003	3029749-53-6	Wave Life Sciences, USA	Recruiting	Phase I/Phase II	NCT05032196
				Active, not recruiting	Phase II/Phase III	NCT04219241
				Not yet recruiting	Phase III	NCT06097780
Computer based cognitive stimulation	Virtual reality computer simulation	N/A	Santa Creu Hospital, Spain	Active, not recruiting	NA	NCT05769972
Monoclonal antibody	Pepinemab	2097151-87-4	Vaccinex, USA	Completed	Phase II	NCT02481674
Small molecule	SAGE-718	1629853-48-0	Sage Therapeutics, USA	Recruiting	Phase II	NCT05107128
					Phase II	NCT05358821
					Phase III	NCT05655520
Stem cell therapy	NestaCell	N/A	Azidus, Brazil	Active, not recruiting	Phase I	NCT02728115
				Active, not recruiting	Phase II/Phase III	NCT04219241
				Not yet recruiting	Phase III	NCT06097780
<b>Myasthenia gravis</b>						
Antigen therapy	CNP-106	N/A	COUR Pharmacetucial, USA	Not yet recruiting	Phase I/III	NCT06106672
Cell therapy	Descartes-08	2784598-58-7	Cartesian Therapeutics, USA	Recruiting	Phase II	NCT04146051
Fusion protein	Telitacicept	2136630-26-5	RemeGen, China	Recruiting	Phase III	NCT05737160
Small molecule	Vemircopan (ALXN2050)	2086178-00-7	Alexion, UK	Active, not recruiting	Phase II	NCT05218096

## FDA approved therapeutic agents

While there is no current cure for ALS, HD, or MG, there are treatments to slow disease progression and treat symptoms. Table 6 examines the US FDA approved treatments for these rare diseases along with their CAS RN, therapy types, mechanism of action, and company information. Small molecule drugs dominate the approved drugs for both ALS and HD, with biologic therapies such as monoclonal antibodies, antibody fragments, and peptide therapy making up the approved treatments for MG.

There are 13 unique drugs currently approved by the US FDA for the treatment of these rare diseases. Three of these compounds have multiple approved formulations, as well. One example, the drug riluzole has three different formulations approved. The first being an oral tablet, there is also an oral film, and a thickened suspension available for patients dealing with muscle tone and swallowing issues. One of the approved drugs for the treatment of ALS, Relyvrio, has been recently discontinued. Amylyx Pharmaceuticals has started the process with the US FDA of discontinuing authorizations for Relyvrio and removing it from the market.<sup>846</sup> While Relyvrio is generally safe and well tolerated, it unfortunately failed to meet primary and secondary endpoints in a Phase III clinical trial (NCT05021536).<sup>846</sup>



**Table 6.** US FDA approved drugs for the treatment of specified rare diseases (Source: The CAS Content Collection)

Drug	Therapy type	CAS RN	Rare disease indication	Mechanism/ notes	Company, location
Exservan (riluzole)	Small molecule	1744-22-5	Amyotrophic lateral sclerosis	Glutamate signaling blocker/ oral film formulation	Mitsubishi Tanabe Pharma America, USA
Nuedexta (dextromethorphan hydrobromide and quinidien sulfate)	Small molecule	2445595-41-3	Amyotrophic lateral sclerosis	Sigma-1 receptor agonist, NMDA receptor antagonist	Otsukac America Pharmaceutical, USA
Qalsody (tofersen)	Gene therapy	2088232-70-4	Amyotrophic lateral sclerosis	Targets SOD1 mRNA to reduce SOD1 protein production	Biogen, USA
Radicava (edaravone)	Small molecule	89-25-8	Amyotrophic lateral sclerosis	Free radical scavenger	Mitsubishi Tanabe Pharma America, USA
Relyvrio (sodium phenylbutyrate and taurursodiol)	Small molecule	2436469-04-2	Amyotrophic lateral sclerosis	Small molecule chaperone and Bax inhibitor/ withdrawn 2024	Amylyx, USA
Rilutek (riluzole)	Small molecule	1744-22-5	Amyotrophic lateral sclerosis	Glutamate signaling blocker/ oral tablet formulation	Sanofi, USA
Tiglutik (riluzole)	Small molecule	1744-22-5	Amyotrophic lateral sclerosis	Glutamate signaling blocker/ oral thickened suspension	ITF Pharma, USA
Austedo (deutetrabenazine)	Small molecule	1392826-25-3	Huntington's disease	VMAT2 inhibitor	Teva Pharmaceutical, Israel
Austedo XR (deutetrabenazine)	Small molecule	1392826-25-3	Huntington's disease	VMAT2 inhibitor/ extended release formulation	Teva Pharmaceutical, Israel
Ingrezza (valbenazine)	Small molecule	1025504-45-3	Huntington's disease	VMAT2 inhibitor	Neurocrine Biosciences, USA
Xenazine (tetrabenazine)	Small molecule	58-46-8	Huntington's disease	VMAT2 inhibitor	Lundbeck Pharmaceuticals, Denmark
Rystiggo (rozanolixizumab-noli)	Monoclonal antibody	1584645-37-3	Myasthenia gravis	Targets FcRn to prevent IgG recycling	UCB, USA
Soliris (eculizumab)	Monoclonal antibody	219685-50-4	Myasthenia gravis	Complement factor C5 inhibitor	Alexion, UK

Ultomiris (ravulizumab-cwvz)	Monoclonal antibody	1803171-55-2	Myasthenia gravis	Complement factor C5 inhibitor	Alexion, UK
Vyvgart (efgartigimod alfa-fcab) intravenous injection	Antibody fragment	1821402-21-4	Myasthenia gravis	Fc receptor blocker	Argenx, Netherlands
Vyvgart Hytrulo (efgartigimod alfa and hyaluronidase-qvfc) subcutaneous injection	Antibody fragment	1821402-21-4	Myasthenia gravis	Fc receptor blocker	Argenx, Netherlands
Zilbrysq (zilucoplan)	Peptide therapy	1841136-73-9	Myasthenia gravis	Complement factor C5 inhibitor	UCB, USA

## 6. Notable patents on rare diseases

**Table 7** shows notable patents in the field of rare diseases such as ALS, HD and MG published in recent years (2020 to 2023). Patents were selected based on relevance, novelty, applicability, and field of study. Most of these involve therapeutic strategies, disease markers and recent advancements in these disease areas.

For instance, a recently published patent CN117050134 describes the synthesis and characterization of a novel oleanamide derivative for activating KEAP/NRF2/ARE signaling pathway which is a signature of oxidative stress. This treatment activates NRF2 transcription factor that can help in preventing and treating various neurological disorders including ALS. In context of HD, WO2023099648 by AstraZeneca AB, Sweden, describes pyrazolo- and triazolo-azinone compounds that inhibits receptor-interacting protein kinase 1 (RIPK1) and can be used in the treatment of neurological disorders including HD. In another example, patent application WO2023236967A1 by RemeGen Co., Ltd., China, describes the development of the drug, a dosage regimen, an administration interval, and a mode for treating MG using - Telitacicept (TACI-Fc fusion protein). It is shown that this formulation exhibits good clinical efficacy and safety in the treatment of MG patients. These patents highlight the constant research endeavours in the field of rare diseases.

**Table 7.** List of notable patents pertaining to the three rare diseases – ALS, HD, and MG – identified from the CAS Content Collection.

Patent Number	Year	Patent assignee, location	Description
<b>Amyotrophic lateral sclerosis</b>			
US20200002723	2020	Deutsches Krebsforschungszentrum, Germany	It describes nucleotide sequences called MSBI (Multiple Sclerosis Brain Isolate) as well as probes, primers, and antibodies against polypeptides encoded by MSBI sequences. These could serve as early markers for the future development of cancer and diseases of the CNS (multiple sclerosis, prion-linked diseases, ALS, transmissible spongiform encephalitis, Parkinson's disease, and Alzheimer's disease).
WO2020010049	2020	The General Hospital Corporation, AZ Therapies, Inc., USA	It describes a composition comprising micronized cromolyn sodium, $\alpha$ -lactose, and salt of fatty acid (preferably magnesium stearate) used to treat certain neurological diseases including Alzheimer's disease, ALS, and Parkinson's disease.
CN117050134	2023	Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, China	It describes a novel oleanamide derivative for activating a KEAP/NRF2/ARE signaling pathway which can be used for treating and preventing various neurological disorders including ALS.

EP4255406A1	2020	Massey Ventures Ltd	It relates to (2S)-2-Aminopentanethioic S-acid or a pharmaceutically acceptable salt as a medication for the treatment of ALS.
WO2022138707	2022	Eisai R&D Management Co., Ltd., Japan	It describes the development of pharmaceutical composition comprising anti-EphA4 antibodies capable of binding to and promoting the cleavage of EphA4, which is used for treating ALS.
<b>Huntington's disease</b>			
WO2023099648	2023	AstraZeneca AB, Sweden	It describes pyrazolo- and triazolo-azinone compounds that inhibit receptor-interacting protein kinase 1 ("RIPK1") and can be used in the treatment of neurological disorders including HD.
US20240076310A1	2023	Sage Therapeutics Inc, USA	Neuroactive steroids (or their combinations), that target GABA receptor complex (GRC) can be used for the treatment of neurodegenerative disorders including HD.
WO2022235329A1	2022	University of South Carolina, USA	It comprises hydrophilic nanogels based on polyethylene glycol (PEG) copolymers. The nanogels can encapsulate an antibody for delivery to the brain and can include ligands for blood brain barrier (BBB) receptors on the surface. These nanogels can the treatment of neurological disorders including HD.
WO2022132894A1	2022	Rush University Medical Center, USA	It describes a pharmaceutical composition comprising glycerol tribenzoate and glycerol phenylbutyrate which can be used for treating HD.
WO2020068913A1	2020	Chase Therapeutics Corporation, United States	It describes a combination 5HT <sub>3</sub> antagonist and/or a NK-1 antagonist, in combination with 6-propylamino-4,5,6,7-tetrahydro-1,3-benzothiazole-2-amine and with fluoxetine, zonisamide, or a statin to treat protein misfolding neurodegenerative diseases.
<b>Myasthenia gravis</b>			
WO2020106724A1	2020	Alexion Pharmaceuticals, Inc., United States	It describes a formulation that specifically binds complement component 5 (C5) and can be used for treating MG in the pediatric population.
WO2020014072A1	2020	GT Biopharma, Inc., United States	It describes the use of an NK1-antagonist (e.g. aprepitant), in combination with neostigmine, to facilitate the treatment of a patient suffering from MG.
WO2023236967A1	2023	RemeGen Co., Ltd., China	This describes the development of the drug, a dosage regimen, an administration interval, and a mode for treating MG using TACI-Fc fusion protein. It is shown that this formulation exhibits good clinical efficacy and safety in the treatment of MG patients.
WO2020086506A1	2020	Ra Pharmaceuticals, Inc., United States	The present disclosure relates methods of treating MG with zilucoplan (complement inhibitor), including devices and kits available for administering zilucoplan.
CN112048565A	2020	Shijiazhuang People's Hospital, China	The invention discloses a microbial marker (comprising <i>Megamonas hypermegale</i> and/or <i>Fusobacterium mortiferum</i> ) for the diagnosis of MG which offers the advantages of good specificity and high sensitivity.

## 7. Outlook and Perspectives

In the vast landscape of medical conditions, rare diseases occupy a unique and often overlooked niche. Defined by their low prevalence, these disorders collectively affect millions worldwide<sup>8, 9</sup>, presenting a multitude of challenges to patients, healthcare professionals, and researchers. Moreover, rare diseases offer a window into the diversity of human health and the complexity of biological systems. Each condition represents a unique manifestation of genetic, environmental, or infectious factors, often with distinct clinical presentations and treatment challenges. From rare genetic disorders like cystic fibrosis and Huntington's disease to autoimmune conditions like lupus, myasthenia gravis, and rare cancers, the spectrum of rare diseases encompasses a broad array of pathologies.

Significant roadblocks remain on the path to progress in rare disease research and care. The small size of patient populations presents challenges for conducting robust clinical trials, leading to limited evidence-based treatment options. Furthermore, the fragmented nature of rare disease research and healthcare delivery can impede collaboration and knowledge sharing. Fragmented approaches to research and care may limit opportunities for interdisciplinary collaboration and hinder the translation of scientific discoveries into clinical practice. Commercial incentives for developing treatments for rare diseases have been relatively low accounting for low interest from pharmaceutical industry.

Despite the challenges, there is reason for optimism in the rare disease landscape. Advances in genomics, molecular biology, and precision medicine hold promise for improved diagnosis and targeted therapies. Technologies such as next-generation sequencing have revolutionized our ability to identify genetic mutations underlying rare diseases. Whole exome sequencing (WES)<sup>847-849</sup> and whole genome sequencing (WGS)<sup>850, 851</sup> have become indispensable tools for unraveling the genetic basis of rare diseases, facilitating personalized medicine approaches and targeted therapies. Research into rare diseases has uncovered a plethora of novel disease mechanisms, shedding light on fundamental biological processes and pathways underlying human health and disease. Insights gained from studying rare diseases have broad implications for understanding more prevalent disorders and have led to the identification of druggable targets and therapeutic strategies. The general public is also becoming increasingly aware of the existence of these rare diseases, driving the overall interest up.

Despite the inherent challenges in developing treatments for rare diseases, there have been significant advancements in therapeutic innovations. From small molecule drugs to gene and cell-based therapies, researchers are exploring diverse modalities to address the unmet medical needs of individuals with rare diseases. Collaborative initiatives, regulatory incentives, and patient-centered trial designs are accelerating the translation of scientific discoveries into clinically meaningful interventions. AI is also being utilized from drug target discovery to the clinical pipeline, and beyond, to speed pharmaceutical progress and development. The sharing of data and resources through collaborative platforms and consortia has emerged as a cornerstone of rare disease research. Initiatives such as the Global Alliance for Genomics and Health (GA4GH)<sup>852</sup> and the Undiagnosed Diseases Network (UDN)<sup>853</sup> facilitate data exchange, harmonize standards, and foster interdisciplinary collaborations, thereby maximizing the impact of research efforts and empowering patients and families with rare diseases.

Looking ahead, continued investment in rare disease research, infrastructure, and policy initiatives is critical for overcoming existing challenges and maximizing the potential of scientific advancements to improve the lives of individuals with rare diseases. Multidisciplinary collaborations and inclusive research practices are needed to effectively address the complex and evolving landscape of rare diseases.

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## Notes

The authors declare no competing financial interest.

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## Supporting Information

Supporting Information. Methodology for collating genes associated with rare diseases utilizing the CAS Content Collection

Supporting Information Figure S1. Heat map indicating number of patents filed by commercial organizations with respect to rare cancers. Patent assignees shown here are among the top 6 commercial patent assignees shown in Figure 4B. Data includes patent publications in the field of rare diseases from the CAS Content Collection for 2003-2023

Supporting Information Figure S2. (A) Number of publications (journal and patent) and (B) patent-to-journal ratios for leading rare cancers in our dataset. Data includes both patent and journal publications sourced/extracted from the CAS Content Collection for the period 2018-2023 in the field of rare diseases.

Supporting Information Figure S3. Leading substances from the CAS Registry associated with amyotrophic lateral sclerosis (ALS), Huntington's disease (HD) and myasthenia gravis (MG) for the protein/peptide sequences subclass of substances. Data includes substances associated with both patent and journal publications sourced from the CAS Registry and the CAS Content Collection for the period 2012-2023.

## References

1. Abozaid, G. M., Kerr, K., McKnight, A., and Al-Omar, H. A. (2022) Criteria to define rare diseases and orphan drugs: a systematic review protocol. *BMJ Open* 12, e062126.
2. National Organization for Rare Disorders. <https://rarediseases.org/> (accessed Mar 21, 2024).
3. Rare Disease Database. <https://rarediseases.org/rare-diseases/> (accessed Mar 21, 2024).
4. Genetic and Rare Diseases Information Center. <https://rarediseases.info.nih.gov/> (accessed Mar 21, 2024).
5. Rare diseases. [https://research-and-innovation.ec.europa.eu/research-area/health/rare-diseases\\_en](https://research-and-innovation.ec.europa.eu/research-area/health/rare-diseases_en) (accessed Mar 21, 2024).
6. Global Genes. <https://globalgenes.org/> (accessed Mar 21, 2024).
7. Fu, M. P., Merrill, S. M., Sharma, M., Gibson, W. T., Turvey, S. E., and Kobor, M. S. (2023) Rare diseases of epigenetic origin: Challenges and opportunities. *Front Genet* 14, 1113086.
8. Nguengang Wakap, S., Lambert, D. M., Olry, A., Rodwell, C., Gueydan, C., Lanneau, V., Murphy, D., Le Cam, Y., and Rath, A. (2020) Estimating cumulative point prevalence of rare diseases: analysis of the Orphanet database. *European Journal of Human Genetics* 28, 165-173.
9. Joszt, L. Not So Rare: 300 Million People Worldwide Affected by Rare Diseases. <https://www.ajmc.com/view/not-so-rare-300-million-people-worldwide-affected-by-rare-diseases> (accessed Mar 27, 2024).
10. Stolk, P., Willemsen, M. J., and Leufkens, H. G. (2006) Rare essentials: drugs for rare diseases as essential medicines. *Bull World Health Organ* 84, 745-751.
11. Melnikova, I. (2012) Rare diseases and orphan drugs. *Nature reviews. Drug discovery* 11, 267-268.
12. Roberts, A., and Wadhwa, R. Orphan Drug Approval Laws. <https://www.ncbi.nlm.nih.gov/books/NBK572052/> (accessed Mar 26, 2024).
13. Herder, M. (2017) What Is the Purpose of the Orphan Drug Act? *PLoS Med* 14, e1002191.
14. Fermaglich, L. J., and Miller, K. L. (2023) A comprehensive study of the rare diseases and conditions targeted by orphan drug designations and approvals over the forty years of the Orphan Drug Act. *Orphanet Journal of Rare Diseases* 18, 163.
15. Orphan drugs incentives and guidelines in 2023. <https://within3.com/blog/orphan-drug-incentives-guidelines> (accessed Mar 26, 2024).
16. Seoane-Vazquez, E., Rodriguez-Monguio, R., Szeinbach, S. L., and Visaria, J. (2008) Incentives for orphan drug research and development in the United States. *Orphanet J Rare Dis* 3, 33.
17. Aartsma-Rus, A., Dooms, M., and Le Cam, Y. (2021) Orphan Medicine Incentives: How to Address the Unmet Needs of Rare Disease Patients by Optimizing the European Orphan Medicinal Product Landscape Guiding Principles and Policy Proposals by the European Expert Group for Orphan Drug Incentives (OD Expert Group). *Front Pharmacol* 12, 744532.
18. CAS Content Collection. <https://www.cas.org/about/cas-content> (accessed Mar 31, 2024).
19. Kovalenko, M., Erdin, S., Andrew, M. A., St Claire, J., Shaughnessey, M., Hubert, L., Neto, J. L., Stortchevoi, A., Fass, D. M., Mouro Pinto, R., et al. (2020) Histone deacetylase knockouts modify transcription, CAG instability and nuclear pathology in Huntington disease mice. *Elife* 9.
20. Wilton, D. K., Mastro, K., Heller, M. D., Gergits, F. W., Willing, C. R., Fahey, J. B., Frouin, A., Daggett, A., Gu, X., Kim, Y. A., et al. (2023) Microglia and complement mediate early corticostriatal synapse loss and cognitive dysfunction in Huntington's disease. *Nat Med* 29, 2866-2884.
21. Narayanaswami, P., Sanders, D. B., Thomas, L., Thibault, D., Blevins, J., Desai, R., Krueger, A., Bibeau, K., Liu, B., Guptill, J. T., et al. (2024) Comparative effectiveness of azathioprine and mycophenolate mofetil for myasthenia gravis (PROMISE-MG): a prospective cohort study. *Lancet Neurol* 23, 267-276.

22. Suster, D. I., Craig Mackinnon, A., DiStasio, M., Basu, M. K., Pihan, G., and Suster, S. (2022) Atypical thymomas with squamoid and spindle cell features: clinicopathologic, immunohistochemical and molecular genetic study of 120 cases with long-term follow-up. *Mod Pathol* 35, 875-894.
23. Xu, D., Jin, T., Zhu, H., Chen, H., Ofengeim, D., Zou, C., Mifflin, L., Pan, L., Amin, P., Li, W., et al. (2018) TBK1 Suppresses RIPK1-Driven Apoptosis and Inflammation during Development and in Aging. *Cell* 174, 1477-1491 e1419.
24. Paganoni, S., Macklin, E. A., Hendrix, S., Berry, J. D., Elliott, M. A., Maiser, S., Karam, C., Caress, J. B., Owegi, M. A., Quick, A., et al. (2020) Trial of Sodium Phenylbutyrate-Taurursodiol for Amyotrophic Lateral Sclerosis. *The New England journal of medicine* 383, 919-930.
25. Raheja, R., Regev, K., Healy, B. C., Mazzola, M. A., Beynon, V., Von Glehn, F., Paul, A., Diaz-Cruz, C., Gholipour, T., Glanz, B. I., et al. (2018) Correlating serum micrnas and clinical parameters in amyotrophic lateral sclerosis. *Muscle Nerve* 58, 261-269.
26. International Multiple Sclerosis Genetics, C., and Multiple, M. S. C. (2023) Locus for severity implicates CNS resilience in progression of multiple sclerosis. *Nature* 619, 323-331.
27. Fitzgerald, K. C., Smith, M. D., Kim, S., Sotirchos, E. S., Kornberg, M. D., Douglas, M., Nourbakhsh, B., Graves, J., Rattan, R., Poisson, L., et al. (2021) Multi-omic evaluation of metabolic alterations in multiple sclerosis identifies shifts in aromatic amino acid metabolism. *Cell Rep Med* 2, 100424.
28. Baughn, M. W., Melamed, Z., Lopez-Erauskin, J., Beccari, M. S., Ling, K., Zuberi, A., Presa, M., Gonzalo-Gil, E., Maimon, R., Vazquez-Sanchez, S., et al. (2023) Mechanism of STMN2 cryptic splice-polyadenylation and its correction for TDP-43 proteinopathies. *Science (New York, N.Y.)* 379, 1140-1149.
29. Bloom, A. J., Mao, X., Strickland, A., Sasaki, Y., Milbrandt, J., and DiAntonio, A. (2022) Constitutively active SARM1 variants that induce neuropathy are enriched in ALS patients. *Mol Neurodegener* 17, 1.
30. Baxi, E. G., Thompson, T., Li, J., Kaye, J. A., Lim, R. G., Wu, J., Ramamoorthy, D., Lima, L., Vaibhav, V., Matlock, A., et al. (2022) Answer ALS, a large-scale resource for sporadic and familial ALS combining clinical and multi-omics data from induced pluripotent cell lines. *Nat Neurosci* 25, 226-237.
31. Perez, R. K., Gordon, M. G., Subramaniam, M., Kim, M. C., Hartoularos, G. C., Targ, S., Sun, Y., Ogorodnikov, A., Bueno, R., Lu, A., et al. (2022) Single-cell RNA-seq reveals cell type-specific molecular and genetic associations to lupus. *Science (New York, N.Y.)* 376, eabf1970.
32. Hasni, S. A., Gupta, S., Davis, M., Poncio, E., Temesgen-Oyelakin, Y., Carlucci, P. M., Wang, X., Naqi, M., Playford, M. P., Goel, R. R., et al. (2021) Phase 1 double-blind randomized safety trial of the Janus kinase inhibitor tofacitinib in systemic lupus erythematosus. *Nat Commun* 12, 3391.
33. Li, C., Georgakopoulou, A., Newby, G. A., Chen, P. J., Everette, K. A., Paschoudi, K., Vlachaki, E., Gil, S., Anderson, A. K., Koob, T., et al. (2023) In vivo HSC prime editing rescues sickle cell disease in a mouse model. *Blood* 141, 2085-2099.
34. Jones, R. J., and DeBaun, M. R. (2021) Leukemia after gene therapy for sickle cell disease: insertional mutagenesis, busulfan, both, or neither. *Blood* 138, 942-947.
35. Richardson, P. G., Oriol, A., Larocca, A., Blade, J., Cavo, M., Rodriguez-Otero, P., Leleu, X., Nadeem, O., Hiemenz, J. W., Hassoun, H., et al. (2021) Melflufen and Dexamethasone in Heavily Pretreated Relapsed and Refractory Multiple Myeloma. *J Clin Oncol* 39, 757-767.
36. Hideshima, T., and Anderson, K. C. (2021) Signaling Pathway Mediating Myeloma Cell Growth and Survival. *Cancers (Basel)* 13.
37. Breen, W. G., Hathcock, M. A., Young, J. R., Kowalchuk, R. O., Bansal, R., Khurana, A., Bennani, N. N., Paludo, J., Villasboas Bisneto, J. C., Wang, Y., et al. (2022) Metabolic characteristics and prognostic differentiation of aggressive lymphoma using one-month post-CAR-T FDG PET/CT. *J Hematol Oncol* 15, 36.



38. Wagner-Johnston, N. D., Sharman, J., Furman, R. R., Salles, G., Brown, J. R., Robak, T., Gu, L., Xing, G., Chan, R. J., Rajakumaraswamy, N., et al. (2021) Idelalisib immune-related toxicity is associated with improved treatment response. *Leuk Lymphoma* 62, 2915-2920.
39. Jhelum, P., Santos-Nogueira, E., Teo, W., Haumont, A., Lenoel, I., Stys, P. K., and David, S. (2020) Ferroptosis Mediates Cuprizone-Induced Loss of Oligodendrocytes and Demyelination. *J Neurosci* 40, 9327-9341.
40. Rojas, O. L., Probstel, A. K., Porfilio, E. A., Wang, A. A., Charabati, M., Sun, T., Lee, D. S. W., Galicia, G., Ramaglia, V., Ward, L. A., et al. (2019) Recirculating Intestinal IgA-Producing Cells Regulate Neuroinflammation via IL-10. *Cell* 176, 610-624 e618.
41. Cortese, R., Tur, C., Prados, F., Schneider, T., Kanber, B., Moccia, M., Wheeler-Kingshott, C. A. G., Thompson, A. J., Barkhof, F., and Ciccarelli, O. (2021) Ongoing microstructural changes in the cervical cord underpin disability progression in early primary progressive multiple sclerosis. *Mult Scler* 27, 28-38.
42. Desai, R. A., Davies, A. L., Del Rossi, N., Tachrount, M., Dyson, A., Gustavson, B., Kaynezhad, P., Mackenzie, L., van der Putten, M. A., McElroy, D., et al. (2020) Nimodipine Reduces Dysfunction and Demyelination in Models of Multiple Sclerosis. *Ann Neurol* 88, 123-136.
43. Caron, N. S., Banos, R., Yanick, C., Aly, A. E., Byrne, L. M., Smith, E. D., Xie, Y., Smith, S. E. P., Potluri, N., Findlay Black, H., et al. (2021) Mutant Huntingtin Is Cleared from the Brain via Active Mechanisms in Huntington Disease. *J Neurosci* 41, 780-796.
44. O'Day, D. H. (2022) Calmodulin Binding Domains in Critical Risk Proteins Involved in Neurodegeneration. *Curr Issues Mol Biol* 44, 5802-5814.
45. O'Regan, G. C., Farag, S. H., Casey, C. S., Wood-Kaczmar, A., Pocock, J. M., Tabrizi, S. J., and Andre, R. (2021) Human Huntington's disease pluripotent stem cell-derived microglia develop normally but are abnormally hyper-reactive and release elevated levels of reactive oxygen species. *J Neuroinflammation* 18, 94.
46. Scahill, R. I., Zeun, P., Osborne-Crowley, K., Johnson, E. B., Gregory, S., Parker, C., Lowe, J., Nair, A., O'Callaghan, C., Langley, C., et al. (2020) Biological and clinical characteristics of gene carriers far from predicted onset in the Huntington's disease Young Adult Study (HD-YAS): a cross-sectional analysis. *Lancet Neurol* 19, 502-512.
47. Mai, L., Asaduzzaman, A., Noamani, B., Fortin, P. R., Gladman, D. D., Touma, Z., Urowitz, M. B., and Wither, J. (2021) The baseline interferon signature predicts disease severity over the subsequent 5 years in systemic lupus erythematosus. *Arthritis Res Ther* 23, 29.
48. Munoz-Grajales, C., Prokopec, S. D., Johnson, S. R., Touma, Z., Ahmad, Z., Bonilla, D., Hiraki, L., Bookman, A., Boutros, P. C., Chruscinski, A., et al. (2022) Serological abnormalities that predict progression to systemic autoimmune rheumatic diseases in antinuclear antibody-positive individuals. *Rheumatology (Oxford)* 61, 1092-1105.
49. Bradford, H. F., Haljasmagi, L., Menon, M., McDonnell, T. C. R., Sarekannu, K., Vanker, M., Peterson, P., Wincup, C., Abida, R., Gonzalez, R. F., et al. (2023) Inactive disease in patients with lupus is linked to autoantibodies to type I interferons that normalize blood IFNalpha and B cell subsets. *Cell Rep Med* 4, 100894.
50. Isenberg, D., Furie, R., Jones, N. S., Guibord, P., Galanter, J., Lee, C., McGregor, A., Toth, B., Rae, J., Hwang, O., et al. (2021) Efficacy, Safety, and Pharmacodynamic Effects of the Bruton's Tyrosine Kinase Inhibitor Fenebrutinib (GDC-0853) in Systemic Lupus Erythematosus: Results of a Phase II, Randomized, Double-Blind, Placebo-Controlled Trial. *Arthritis Rheumatol* 73, 1835-1846.
51. Paulson, V. A., Shivdasani, P., Angell, T. E., Cibas, E. S., Krane, J. F., Lindeman, N. I., Alexander, E. K., and Barletta, J. A. (2017) Noninvasive Follicular Thyroid Neoplasm with Papillary-Like Nuclear Features Accounts for More Than Half of "Carcinomas" Harboring RAS Mutations. *Thyroid* 27, 506-511.

52. Chu, Y. H., Wirth, L. J., Farahani, A. A., Nose, V., Faquin, W. C., Dias-Santagata, D., and Sadow, P. M. (2020) Clinicopathologic features of kinase fusion-related thyroid carcinomas: an integrative analysis with molecular characterization. *Mod Pathol* 33, 2458-2472.
53. Snyder, A., Makarov, V., Merghoub, T., Yuan, J., Zaretsky, J. M., Desrichard, A., Walsh, L. A., Postow, M. A., Wong, P., Ho, T. S., et al. (2014) Genetic basis for clinical response to CTLA-4 blockade in melanoma. *The New England journal of medicine* 371, 2189-2199.
54. Curtin, J. A., Fridlyand, J., Kageshita, T., Patel, H. N., Busam, K. J., Kutzner, H., Cho, K. H., Aiba, S., Brocker, E. B., LeBoit, P. E., et al. (2005) Distinct sets of genetic alterations in melanoma. *The New England journal of medicine* 353, 2135-2147.
55. Richardson, P. G., Sonneveld, P., Schuster, M. W., Irwin, D., Stadtmauer, E. A., Facon, T., Harousseau, J. L., Ben-Yehuda, D., Lonial, S., Goldschmidt, H., et al. (2005) Bortezomib or high-dose dexamethasone for relapsed multiple myeloma. *The New England journal of medicine* 352, 2487-2498.
56. Hauser, S. L., Waubant, E., Arnold, D. L., Vollmer, T., Antel, J., Fox, R. J., Bar-Or, A., Panzara, M., Sarkar, N., Agarwal, S., et al. (2008) B-cell depletion with rituximab in relapsing-remitting multiple sclerosis. *The New England journal of medicine* 358, 676-688.
57. Kappos, L., Radue, E. W., O'Connor, P., Polman, C., Hohlfeld, R., Calabresi, P., Selmaj, K., Agoropoulou, C., Leyk, M., Zhang-Auberson, L., et al. (2010) A placebo-controlled trial of oral fingolimod in relapsing multiple sclerosis. *The New England journal of medicine* 362, 387-401.
58. Patel, J. P., Gonen, M., Figueroa, M. E., Fernandez, H., Sun, Z., Ravevskis, J., Van Vlierberghe, P., Dolgalev, I., Thomas, S., Aminova, O., et al. (2012) Prognostic relevance of integrated genetic profiling in acute myeloid leukemia. *The New England journal of medicine* 366, 1079-1089.
59. Tabrizi, S. J., Leavitt, B. R., Landwehrmeyer, G. B., Wild, E. J., Saft, C., Barker, R. A., Blair, N. F., Craufurd, D., Priller, J., Rickards, H., et al. (2019) Targeting Huntingtin Expression in Patients with Huntington's Disease. *The New England journal of medicine* 380, 2307-2316.
60. Liddelow, S. A., Guttenplan, K. A., Clarke, L. E., Bennett, F. C., Bohlen, C. J., Schirmer, L., Bennett, M. L., Munch, A. E., Chung, W. S., Peterson, T. C., et al. (2017) Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* 541, 481-487.
61. Gilhus, N. E. (2016) Myasthenia Gravis. *The New England journal of medicine* 375, 2570-2581.
62. Guglielmetti, C., Levi, J., Huynh, T. L., Tiret, B., Blecha, J., Tang, R., VanBrocklin, H., and Chaumeil, M. M. (2022) Longitudinal Imaging of T Cells and Inflammatory Demyelination in a Preclinical Model of Multiple Sclerosis Using (18)F-FARAG PET and MRI. *Journal of nuclear medicine : official publication, Society of Nuclear Medicine* 63, 140-146.
63. Hauser, S. L., Zielman, R., Das Gupta, A., Xi, J., Stoneman, D., Karlsson, G., Robertson, D., Cohen, J. A., and Kappos, L. (2023) Efficacy and safety of four-year ofatumumab treatment in relapsing multiple sclerosis: The ALITHIOS open-label extension. *Mult Scler* 29, 1452-1464.
64. Rojo, D., Dal Cengio, L., Badner, A., Kim, S., Sakai, N., Greene, J., Dierckx, T., Mehl, L. C., Eisinger, E., Ransom, J., et al. (2023) BMAL1 loss in oligodendroglia contributes to abnormal myelination and sleep. *Neuron* 111, 3604-3618 e3611.
65. Baghdassarian, H., Blackstone, S. A., Clay, O. S., Philips, R., Matthiasardottir, B., Nehrebecky, M., Hua, V. K., McVicar, R., Liu, Y., Tucker, S. M., et al. (2023) Variant STAT4 and Response to Ruxolitinib in an Autoinflammatory Syndrome. *The New England journal of medicine* 388, 2241-2252.
66. Hasanov, E., and Jonasch, E. (2021) MK-6482 as a potential treatment for von Hippel-Lindau disease-associated clear cell renal cell carcinoma. *Expert Opin Investig Drugs* 30, 495-504.
67. Compositions and methods for preserving and/or restoring neural function. WO2023086603, 2023.
68. Ire1alpha inhibitors and uses thereof. WO2022104148, 2022.
69. Viral delivery of therapeutics to the central nervous system. WO2022165538A1, 2022.
70. Enhancing gaba's ability to modulate immune responses. WO2018236955, 2018.

71. Peptides and uses thereof for diagnosing and treating myasthenia gravis. WO2018049053, 2018.
72. Multiple sclerosis. [https://www.mayoclinic.org/diseases-conditions/multiple-sclerosis/symptoms-causes/syc-20350269#:~:text=Multiple%20sclerosis%20\(MS\)%20is%20a,the%20rest%20of%20your%20body.](https://www.mayoclinic.org/diseases-conditions/multiple-sclerosis/symptoms-causes/syc-20350269#:~:text=Multiple%20sclerosis%20(MS)%20is%20a,the%20rest%20of%20your%20body.) (accessed 11th March).
73. Systemic Lupus Erythematosus (SLE). <https://www.cdc.gov/lupus/facts/detailed.html> (accessed 11th March).
74. Familial Mediterranean fever. <https://www.mayoclinic.org/diseases-conditions/familial-mediterranean-fever/symptoms-causes/syc-20372470> (accessed 11th March).
75. Rett syndrome. <https://www.mayoclinic.org/diseases-conditions/rett-syndrome/symptoms-causes/syc-20377227> (accessed 11th March).
76. Rizzolo, K., Beck, N. M., and Ambruso, S. L. (2022) Syndromes of Pseudo-Hyperaldosteronism. Clin J Am Soc Nephrol 17, 581-584.
77. Kidney cancer. <https://www.mayoclinic.org/diseases-conditions/kidney-cancer/symptoms-causes/syc-20352664> (accessed 11th March).
78. Thyroid cancer. <https://www.mayoclinic.org/diseases-conditions/thyroid-cancer/symptoms-causes/syc-20354161> (accessed 11th March).
79. Melanoma. <https://www.mayoclinic.org/diseases-conditions/melanoma/symptoms-causes/syc-20374884#:~:text=deeper%20skin%20layers.-,Melanoma%20is%20a%20kind%20of%20skin%20cancer%20that%20starts%20in,often%20exposed%20to%20the%20sun.> (accessed 11th March).
80. Multiple myeloma. <https://www.mayoclinic.org/diseases-conditions/multiple-myeloma/symptoms-causes/syc-20353378#:~:text=Multiple%20myeloma%20is%20a%20cancer,build%20up%20in%20bone%20marrow.> (accessed 11th March).
81. Nasopharyngeal carcinoma. <https://www.mayoclinic.org/diseases-conditions/nasopharyngeal-carcinoma/symptoms-causes/syc-20375529> (accessed 11th March).
82. Cuglievan, B., Connors, J., He, J., Khazal, S., Yedururi, S., Dai, J., Garces, S., Quesada, A. E., Roth, M., Garcia, M., et al. (2023) Blastic plasmacytoid dendritic cell neoplasm: a comprehensive review in pediatrics, adolescents, and young adults (AYA) and an update of novel therapies. Leukemia 37, 1767-1778.
83. Freedman, A. S. A., J. C.; Dearden, C. B cell prolymphocytic leukemia. [https://www.uptodate.com/contents/b-cell-prolymphocytic-leukemia#:~:text=B%20cell%20prolymphocytic%20leukemia%20\(B,are%20mature%20activated%20B%20cells.](https://www.uptodate.com/contents/b-cell-prolymphocytic-leukemia#:~:text=B%20cell%20prolymphocytic%20leukemia%20(B,are%20mature%20activated%20B%20cells.) (accessed 11th March).
84. Bindra, B. S., Kaur, H., Portillo, S., Emiloju, O., and Garcia de de Jesus, K. (2019) B-cell Prolymphocytic Leukemia: Case Report and Challenges on a Diagnostic and Therapeutic Forefront. Cureus 11, e5629.
85. Scleroderma. <https://www.mayoclinic.org/diseases-conditions/scleroderma/symptoms-causes/syc-20351952> (accessed 11th March).
86. Sjogren's syndrome. <https://www.mayoclinic.org/diseases-conditions/sjogrens-syndrome/symptoms-causes/syc-20353216> (accessed 11th March).
87. Cystic fibrosis. <https://www.mayoclinic.org/diseases-conditions/cystic-fibrosis/symptoms-causes/syc-20353700#:~:text=Cystic%20fibrosis%20is%20a%20disorder,mucus%2C%20sweat%20and%20digestive%20juices.> (accessed 11th March).
88. Sickle cell anemia. <https://www.mayoclinic.org/diseases-conditions/sickle-cell-anemia/symptoms-causes/syc-20355876> (accessed 11th March).

89. Non-Hodgkin Lymphoma Treatment (PDQ®)–Patient Version. <https://www.cancer.gov/types/lymphoma/patient/adult-nhl-treatment-pdq#:~:text=Non%2DHodgkin%20lymphoma%20is%20a,risk%20of%20non%2DHodgkin%20lymphoma>. (accessed 11th March).
90. Acute myelogenous leukemia. <https://www.mayoclinic.org/diseases-conditions/acute-myelogenous-leukemia/symptoms-causes/syc-20369109> (accessed 11th March).
91. Pheochromocytoma. <https://www.cancer.gov/pediatric-adult-rare-tumor/rare-tumors/rare-endocrine-tumor/pheochromocytoma#:~:text=Pheochromocytoma%20is%20a%20type%20of,on%20top%20of%20the%20kidneys>. (accessed 11th March).
92. Cholangiocarcinoma (bile duct cancer). <https://www.mayoclinic.org/diseases-conditions/cholangiocarcinoma/symptoms-causes/syc-20352408> (accessed 11th March).
93. Liver cancer. <https://www.mayoclinic.org/diseases-conditions/liver-cancer/symptoms-causes/syc-20353659> (accessed 11th March).
94. Mesothelioma. <https://www.mayoclinic.org/diseases-conditions/mesothelioma/symptoms-causes/syc-20375022> (accessed 11th March).
95. Glioma. <https://www.mayoclinic.org/diseases-conditions/glioblastoma/cdc-20350148> (accessed 11th March).
96. Creutzfeldt-Jakob disease. <https://www.mayoclinic.org/diseases-conditions/creutzfeldt-jakob-disease/symptoms-causes/syc-20371226> (accessed 11th March).
97. Progressive supranuclear palsy. <https://www.mayoclinic.org/diseases-conditions/progressive-supranuclear-palsy/symptoms-causes/syc-20355659> (accessed 11th March).
98. Graves' disease. <https://www.mayoclinic.org/diseases-conditions/graves-disease/symptoms-causes/syc-20356240> (accessed 11th March).
99. Juvenile idiopathic arthritis. <https://www.mayoclinic.org/diseases-conditions/juvenile-idiopathic-arthritis/symptoms-causes/syc-20374082> (accessed 11th March).
100. Sarcoidosis. <https://www.mayoclinic.org/diseases-conditions/sarcoidosis/symptoms-causes/syc-20350358> (accessed 11th March).
101. Guillain-Barre syndrome. <https://www.mayoclinic.org/diseases-conditions/guillain-barre-syndrome/symptoms-causes/syc-20362793> (accessed 11th March).
102. Soft tissue sarcoma. <https://www.mayoclinic.org/diseases-conditions/kaposi-sarcoma/cdc-20387726> (accessed 11th March).
103. Esophageal cancer. <https://www.mayoclinic.org/diseases-conditions/esophageal-cancer/symptoms-causes/syc-20356084#:~:text=Esophageal%20cancer%20occurs%20when%20cells,other%20parts%20of%20the%20body>. (accessed 11th March).
104. Kiernan, M. C., Vucic, S., Cheah, B. C., Turner, M. R., Eisen, A., Hardiman, O., Burrell, J. R., and Zoing, M. C. (2011) Amyotrophic lateral sclerosis. *Lancet* 377, 942-955.
105. Siddique, N., and Siddique, T., Amyotrophic Lateral Sclerosis Overview. In *Gene Reviews*, Adam, M. P.; Feldman, J.; Mirzaa, G. M.; Pagon, R. A.; Wallace, S. E.; Bean, L. J. H.; Gripp, K. W.; Amemiya, A., Eds. University of Washington, Seattle

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106. Wijesekera, L. C., and Leigh, P. N. (2009) Amyotrophic lateral sclerosis. *Orphanet J Rare Dis* 4, 3.
107. Amyotrophic lateral sclerosis (ALS). <https://www.mayoclinic.org/diseases-conditions/amyotrophic-lateral-sclerosis/symptoms-causes/syc->

[20354022#:~:text=ALS%20often%20begins%20with%20muscle,cure%20for%20this%20fatal%20disease.](#) (accessed Feb 26, 2024).

108. Goutman, S. A., Hardiman, O., Al-Chalabi, A., Chió, A., Savelieff, M. G., Kiernan, M. C., and Feldman, E. L. (2022) Recent advances in the diagnosis and prognosis of amyotrophic lateral sclerosis. *The Lancet Neurology* 21, 480-493.
109. Goutman, S. A., Hardiman, O., Al-Chalabi, A., Chió, A., Savelieff, M. G., Kiernan, M. C., and Feldman, E. L. (2022) Emerging insights into the complex genetics and pathophysiology of amyotrophic lateral sclerosis. *The Lancet Neurology* 21, 465-479.
110. Barbieri, E. Epidemiology of ALS: Incidence, Prevalence, and Suspected Clusters. <https://www.targetals.org/2022/11/22/epidemiology-of-als-incidence-prevalence-and-clusters/#:~:text=How%20common%20is%20ALS%20worldwide,100%2C000%20to%206%20per%20100%2C000.> (accessed Feb 26, 2024).
111. Wolfson, C., Gauvin, D. E., Ishola, F., and Oskoui, M. (2023) Global Prevalence and Incidence of Amyotrophic Lateral Sclerosis. *Neurology* 101, e613-e623.
112. Xu, L., Liu, T., Liu, L., Yao, X., Chen, L., Fan, D., Zhan, S., and Wang, S. (2020) Global variation in prevalence and incidence of amyotrophic lateral sclerosis: a systematic review and meta-analysis. *J Neurol* 267, 944-953.
113. ALS Ice Bucket Challenge. <https://www.als.org/IBC> (accessed Mar 18, 2024).
114. Piper, K. The Ice Bucket Challenge and the promise — and the pitfalls — of viral charity. <https://www.vox.com/future-perfect/2019/7/20/20699732/ice-bucket-challenge-viral-charity-als> (accessed Mar 18, 2024).
115. Ice Bucket Challenge dramatically accelerated the fight against ALS. <https://www.als.org/stories-news/ice-bucket-challenge-dramatically-accelerated-fight-against-als> (accessed Mar 18, 2024).
116. Onque, R. The 'Ice Bucket Challenge' funded a new ALS drug, but experts have varying opinions about its approval. <https://www.cnn.com/2022/10/08/ice-bucket-challenge-new-als-drug-approved-with-funds-raised.html> (accessed Mar 18, 2024).
117. Who Gets ALS? <https://www.als.org/understanding-als/who-gets-als> (accessed Mar 18, 2024).
118. Keon, M., Musrie, B., Dinger, M., Brennan, S. E., Santos, J., and Saksena, N. K. (2021) Destination Amyotrophic Lateral Sclerosis. *Front Neurol* 12, 596006.
119. Rojas, P., Ramírez, A. I., Fernández-Albarral, J. A., López-Cuenca, I., Salobar-García, E., Cadena, M., Elvira-Hurtado, L., Salazar, J. J., de Hoz, R., and Ramírez, J. M. (2020) Amyotrophic Lateral Sclerosis: A Neurodegenerative Motor Neuron Disease With Ocular Involvement. *Front Neurosci* 14, 566858.
120. Ling, S. C., Polymenidou, M., and Cleveland, D. W. (2013) Converging mechanisms in ALS and FTD: disrupted RNA and protein homeostasis. *Neuron* 79, 416-438.
121. Lattante, S., Ciura, S., Rouleau, G. A., and Kabashi, E. (2015) Defining the genetic connection linking amyotrophic lateral sclerosis (ALS) with frontotemporal dementia (FTD). *Trends Genet* 31, 263-273.
122. Fecto, F., and Siddique, T. (2011) Making connections: pathology and genetics link amyotrophic lateral sclerosis with frontotemporal lobe dementia. *J Mol Neurosci* 45, 663-675.
123. Chió, A., Borghero, G., Restagno, G., Mora, G., Drepper, C., Traynor, B. J., Sendtner, M., Brunetti, M., Ossola, I., Calvo, A., et al. (2012) Clinical characteristics of patients with familial amyotrophic lateral sclerosis carrying the pathogenic GGGGCC hexanucleotide repeat expansion of C9ORF72. *Brain* 135, 784-793.
124. Mejzini, R., Flynn, L. L., Pitout, I. L., Fletcher, S., Wilton, S. D., and Akkari, P. A. (2019) ALS Genetics, Mechanisms, and Therapeutics: Where Are We Now? *Front Neurosci* 13, 1310.
125. ALS Genes and Mutations. <https://www.als.org/research/als-research-topics/genetics> (accessed Feb 26, 2024).

126. Paganoni, S., Macklin, E. A., Lee, A., Murphy, A., Chang, J., Zipf, A., Cudkowicz, M., and Atassi, N. (2014) Diagnostic timelines and delays in diagnosing amyotrophic lateral sclerosis (ALS). *Amyotroph Lateral Scler Frontotemporal Degener* 15, 453-456.
127. Hardiman, O., van den Berg, L. H., and Kiernan, M. C. (2011) Clinical diagnosis and management of amyotrophic lateral sclerosis. *Nat Rev Neurol* 7, 639-649.
128. Majmudar, S., Wu, J., and Paganoni, S. (2014) Rehabilitation in amyotrophic lateral sclerosis: why it matters. *Muscle Nerve* 50, 4-13.
129. FDA-Approved Drugs for Treating ALS. <https://www.als.org/navigating-als/living-with-als/fda-approved-drugs> (accessed Mar 18, 2024).
130. Kaylor, A. Exploring FDA-Approved Drugs and Potential Treatment Options for ALS. [https://pharmanewsintel.com/features/exploring-fda-approved-drugs-and-potential-treatment-options-for-als#:~:text=Covis%20Pharma's%20Rilutek%20\(riluzole\)%20was,has%20been%20available%20since%20003](https://pharmanewsintel.com/features/exploring-fda-approved-drugs-and-potential-treatment-options-for-als#:~:text=Covis%20Pharma's%20Rilutek%20(riluzole)%20was,has%20been%20available%20since%20003). (accessed Mar 18, 2024).
131. RILUTEK (riluzole) tablets, for oral use [https://www.accessdata.fda.gov/drugsatfda\\_docs/label/2016/020599s017lbl.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/label/2016/020599s017lbl.pdf) (accessed Mar 18, 2024).
132. Riluzole. <https://medlineplus.gov/druginfo/meds/a696013.html#:~:text=Riluzole%20is%20used%20to%20treat,th at%20affect%20nerves%20and%20muscles>. (accessed Mar 26, 2024).
133. FDA approves drug to treat ALS. <https://www.fda.gov/news-events/press-announcements/fda-approves-drug-treat-als> (accessed Mar 18, 2024).
134. RELYVRIO (sodium phenylbutyrate and taurursodiol), for oral suspension. [https://www.accessdata.fda.gov/drugsatfda\\_docs/label/2022/216660s000lbl.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/label/2022/216660s000lbl.pdf) (accessed Mar 18, 2024).
135. Sodium Phenylbutyrate and Taurursodiol. <https://medlineplus.gov/druginfo/meds/a623014.html> (accessed Mar 26, 2024).
136. McKenzie, H. Future of Amylyx's ALS Drug in Question After Phase III Failure. <https://www.biospace.com/article/future-of-amylyx-s-als-drug-in-question-after-phase-iii-failure/> (accessed Mar 26, 2024).
137. Bryson, S. MTPA to stop Phase 3 trial of approved Radicava ORS for ALS. <https://alsnewstoday.com/news/mtpa-to-stop-phase-3-trial-of-approved-radicava-ors-for-als/> (accessed Mar 26, 2024).
138. Ferrer reports top-line results from Phase III ADORE study in ALS. <https://www.ferrer.com/en/results-study-ADORE-ALS> (accessed Mar 26, 2024).
139. Bali, T., and Miller, T. M. (2013) Management of amyotrophic lateral sclerosis. *Mo Med* 110, 417-421.
140. Phukan, J., and Hardiman, O. (2009) The management of amyotrophic lateral sclerosis. *J Neurol* 256, 176-186.
141. Brooks, B. R. (2009) Managing amyotrophic lateral sclerosis: Slowing disease progression and improving patient quality of life. *Annals of Neurology* 65, S17-S23.
142. Ferraiuolo, L., Kirby, J., Grierson, A. J., Sendtner, M., and Shaw, P. J. (2011) Molecular pathways of motor neuron injury in amyotrophic lateral sclerosis. *Nat Rev Neurol* 7, 616-630.
143. van den Bos, M. A. J., Geevasinga, N., Higashihara, M., Menon, P., and Vucic, S. (2019) Pathophysiology and Diagnosis of ALS: Insights from Advances in Neurophysiological Techniques. *International journal of molecular sciences* 20.
144. Warner, T. T., Chapter 10 - Motor Neuron Diseases. In *Practical Guide to Neurogenetics*, Warner, T. T.; Hammans, S. R., Eds. W.B. Saunders: Philadelphia, 2009; pp 150-174.

145. Siddique, T., Deng, H. X., and Ajroud-Driss, S., Chapter 132 - Motor Neuron Disease. In *Emery and Rimoin's Principles and Practice of Medical Genetics (Sixth Edition)*, Rimoin, D.; Pyeritz, R.; Korf, B., Eds. Academic Press: Oxford, 2013; pp 1-22.
146. Ragagnin, A. M. G., Shadfar, S., Vidal, M., Jamali, M. S., and Atkin, J. D. (2019) Motor Neuron Susceptibility in ALS/FTD. *Front Neurosci* 13, 532.
147. Boillée, S., Vande Velde, C., and Cleveland, Don W. (2006) ALS: A Disease of Motor Neurons and Their Nonneuronal Neighbors. *Neuron* 52, 39-59.
148. Foran, E., and Trotti, D. (2009) Glutamate transporters and the excitotoxic path to motor neuron degeneration in amyotrophic lateral sclerosis. *Antioxid Redox Signal* 11, 1587-1602.
149. Jiang, L. L., Zhu, B., Zhao, Y., Li, X., Liu, T., Pina-Crespo, J., Zhou, L., Xu, W., Rodriguez, M. J., Yu, H., et al. (2019) Membralin deficiency dysregulates astrocytic glutamate homeostasis leading to ALS-like impairment. *J Clin Invest* 129, 3103-3120.
150. Xie, M., Pallegar, P. N., Parusel, S., Nguyen, A. T., and Wu, L.-J. (2023) Regulation of cortical hyperexcitability in amyotrophic lateral sclerosis: focusing on glial mechanisms. *Molecular Neurodegeneration* 18, 75.
151. Singh, A., Kukreti, R., Saso, L., and Kukreti, S. (2019) Oxidative Stress: A Key Modulator in Neurodegenerative Diseases. *Molecules* 24.
152. Liu, Z., Zhou, T., Ziegler, A. C., Dimitrion, P., and Zuo, L. (2017) Oxidative Stress in Neurodegenerative Diseases: From Molecular Mechanisms to Clinical Applications. *Oxid Med Cell Longev* 2017, 2525967.
153. Duranti, E., and Villa, C. (2022) Molecular Investigations of Protein Aggregation in the Pathogenesis of Amyotrophic Lateral Sclerosis. *International journal of molecular sciences* 24.
154. Blokhuis, A. M., Groen, E. J., Koppers, M., van den Berg, L. H., and Pasterkamp, R. J. (2013) Protein aggregation in amyotrophic lateral sclerosis. *Acta Neuropathol* 125, 777-794.
155. Malik, R., and Wiedau, M. (2020) Therapeutic Approaches Targeting Protein Aggregation in Amyotrophic Lateral Sclerosis. *Frontiers in Molecular Neuroscience* 13.
156. Cicardi, M. E., Marrone, L., Azzouz, M., and Trotti, D. (2021) Proteostatic imbalance and protein spreading in amyotrophic lateral sclerosis. *The EMBO Journal* 40, e106389.
157. Zhao, J., Wang, X., Huo, Z., Chen, Y., Liu, J., Zhao, Z., Meng, F., Su, Q., Bao, W., Zhang, L., et al. (2022) The Impact of Mitochondrial Dysfunction in Amyotrophic Lateral Sclerosis. *Cells* 11.
158. Muyderman, H., and Chen, T. (2014) Mitochondrial dysfunction in amyotrophic lateral sclerosis - a valid pharmacological target? *Br J Pharmacol* 171, 2191-2205.
159. Smith, E. F., Shaw, P. J., and De Vos, K. J. (2019) The role of mitochondria in amyotrophic lateral sclerosis. *Neuroscience Letters* 710, 132933.
160. Provenzano, F., Torazza, C., Bonifacino, T., Bonanno, G., and Milanese, M. (2023) The Key Role of Astrocytes in Amyotrophic Lateral Sclerosis and Their Commitment to Glutamate Excitotoxicity. *International journal of molecular sciences* 24, 15430.
161. Lee, J., Hyeon, S. J., Im, H., Ryu, H., Kim, Y., and Ryu, H. (2016) Astrocytes and Microglia as Non-cell Autonomous Players in the Pathogenesis of ALS. *Exp Neurol* 25, 233-240.
162. You, J., Youssef, M. M. M., Santos, J. R., Lee, J., and Park, J. (2023) Microglia and Astrocytes in Amyotrophic Lateral Sclerosis: Disease-Associated States, Pathological Roles, and Therapeutic Potential. *Biology (Basel)* 12.
163. Van Harten, A. C. M., Phatnani, H., and Przedborski, S. (2021) Non-cell-autonomous pathogenic mechanisms in amyotrophic lateral sclerosis. *Trends in Neurosciences* 44, 658-668.
164. Hartzfeld, D. FYI: Familial Amyotrophic Lateral Sclerosis (FALS) and Genetic Testing. <https://www.als.org/navigating-als/resources/familial-amyotrophic-lateral-sclerosis-fals-and-genetic#:~:text=About%2010%25%20of%20cases%20are,is%20most%20often%20autosomal%20dominant.> (accessed Feb 26, 2024).

165. Siddique, T., and Ajroud-Driss, S. (2011) Familial amyotrophic lateral sclerosis, a historical perspective. *Acta Myol* 30, 117-120.
166. Oskarsson, B., Horton, D. K., and Mitsumoto, H. (2015) Potential Environmental Factors in Amyotrophic Lateral Sclerosis. *Neurol Clin* 33, 877-888.
167. Bozzoni, V., Pansarasa, O., Diamanti, L., Nosari, G., Cereda, C., and Ceroni, M. (2016) Amyotrophic lateral sclerosis and environmental factors. *Funct Neurol* 31, 7-19.
168. Andrew, A. S., Bradley, W. G., Peipert, D., Butt, T., Amoako, K., Piro, E. P., Tandan, R., Novak, J., Quick, A., Pugar, K. D., et al. (2021) Risk factors for amyotrophic lateral sclerosis: A regional United States case-control study. *Muscle & Nerve* 63, 52-59.
169. Rosenfeld, J., and Strong, M. J. (2015) Challenges in the Understanding and Treatment of Amyotrophic Lateral Sclerosis/Motor Neuron Disease. *Neurotherapeutics* 12, 317-325.
170. Zipprich, B. ALS Is Not a Singular Disease. Stop Treating It Like One. <https://www.biospace.com/article/opinion-als-is-not-a-singular-disease-stop-treating-it-like-one/> (accessed Mar 26, 2024).
171. Andersen, P. M., and Al-Chalabi, A. (2011) Clinical genetics of amyotrophic lateral sclerosis: what do we really know? *Nat Rev Neurol* 7, 603-615.
172. Marangi, G., and Traynor, B. J. (2015) Genetic causes of amyotrophic lateral sclerosis: new genetic analysis methodologies entailing new opportunities and challenges. *Brain Res* 1607, 75-93.
173. Renton, A. E., Chiò, A., and Traynor, B. J. (2014) State of play in amyotrophic lateral sclerosis genetics. *Nat Neurosci* 17, 17-23.
174. Ghasemi, M., and Brown, R. H., Jr. (2018) Genetics of Amyotrophic Lateral Sclerosis. *Cold Spring Harb Perspect Med* 8.
175. Vance, C., Rogelj, B., Hortobágyi, T., De Vos, K. J., Nishimura, A. L., Sreedharan, J., Hu, X., Smith, B., Ruddy, D., Wright, P., et al. (2009) Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6. *Science (New York, N.Y.)* 323, 1208-1211.
176. Andersen, P. M. (2006) Amyotrophic lateral sclerosis associated with mutations in the CuZn superoxide dismutase gene. *Curr Neurol Neurosci Rep* 6, 37-46.
177. Amyotrophic Lateral Sclerosis (ALS): Causes/Inheritance. <https://www.mda.org/disease/amyotrophic-lateral-sclerosis/causes-inheritance#:~:text=SOD1%20was%20the%20first%20gene,that%20can%20cause%20familial%20ALS.> (accessed Feb 26, 2024).
178. Berdyński, M., Miszta, P., Safranow, K., Andersen, P. M., Morita, M., Filipek, S., Żekanowski, C., and Kuźma-Kozakiewicz, M. (2022) SOD1 mutations associated with amyotrophic lateral sclerosis analysis of variant severity. *Scientific reports* 12, 103.
179. Farg, M. A., Sundaramoorthy, V., Sultana, J. M., Yang, S., Atkinson, R. A., Levina, V., Halloran, M. A., Gleeson, P. A., Blair, I. P., Soo, K. Y., et al. (2014) C9ORF72, implicated in amyotrophic lateral sclerosis and frontotemporal dementia, regulates endosomal trafficking. *Hum Mol Genet* 23, 3579-3595.
180. DeJesus-Hernandez, M., Mackenzie, I. R., Boeve, B. F., Boxer, A. L., Baker, M., Rutherford, N. J., Nicholson, A. M., Finch, N. A., Flynn, H., Adamson, J., et al. (2011) Expanded GGGGCC hexanucleotide repeat in noncoding region of C9ORF72 causes chromosome 9p-linked FTD and ALS. *Neuron* 72, 245-256.
181. Majounie, E., Renton, A. E., Mok, K., Dopper, E. G., Waite, A., Rollinson, S., Chiò, A., Restagno, G., Nicolaou, N., Simon-Sanchez, J., et al. (2012) Frequency of the C9orf72 hexanucleotide repeat expansion in patients with amyotrophic lateral sclerosis and frontotemporal dementia: a cross-sectional study. *Lancet Neurol* 11, 323-330.
182. Smith, B. N., Newhouse, S., Shatunov, A., Vance, C., Topp, S., Johnson, L., Miller, J., Lee, Y., Troakes, C., Scott, K. M., et al. (2013) The C9ORF72 expansion mutation is a common cause of ALS+/-FTD in Europe and has a single founder. *Eur J Hum Genet* 21, 102-108.



183. Gendron, T. F., Rademakers, R., and Petrucelli, L. (2013) TARDBP mutation analysis in TDP-43 proteinopathies and deciphering the toxicity of mutant TDP-43. *J Alzheimers Dis* 33 Suppl 1, S35-45.
184. Van Deerlin, V. M., Leverenz, J. B., Bekris, L. M., Bird, T. D., Yuan, W., Elman, L. B., Clay, D., Wood, E. M., Chen-Plotkin, A. S., Martinez-Lage, M., et al. (2008) TARDBP mutations in amyotrophic lateral sclerosis with TDP-43 neuropathology: a genetic and histopathological analysis. *Lancet Neurol* 7, 409-416.
185. Chen, S., Zhou, R.-L., Zhang, W., Che, C.-H., Feng, S.-Y., Huang, H.-P., Liu, C.-Y., and Zou, Z.-Y. (2021) Novel TARDBP missense mutation caused familial amyotrophic lateral sclerosis with frontotemporal dementia and parkinsonism. *Neurobiology of Aging* 107, 168-173.
186. Kamelgarn, M., Chen, J., Kuang, L., Jin, H., Kasarskis, E. J., and Zhu, H. (2018) ALS mutations of FUS suppress protein translation and disrupt the regulation of nonsense-mediated decay. *Proceedings of the National Academy of Sciences* 115, E11904-E11913.
187. Ticozzi, N., Silani, V., LeClerc, A. L., Keagle, P., Gellera, C., Ratti, A., Taroni, F., Kwiatkowski, T. J., Jr., McKenna-Yasek, D. M., Sapp, P. C., et al. (2009) Analysis of FUS gene mutation in familial amyotrophic lateral sclerosis within an Italian cohort. *Neurology* 73, 1180-1185.
188. An, H., Skelt, L., Notaro, A., Highley, J. R., Fox, A. H., La Bella, V., Buchman, V. L., and Shelkownikova, T. A. (2019) ALS-linked FUS mutations confer loss and gain of function in the nucleus by promoting excessive formation of dysfunctional paraspeckles. *Acta Neuropathologica Communications* 7, 7.
189. Scarian, E., Fiamingo, G., Diamanti, L., Palmieri, I., Gagliardi, S., and Pansarasa, O. (2022) The Role of VCP Mutations in the Spectrum of Amyotrophic Lateral Sclerosis-Frontotemporal Dementia. *Front Neurol* 13, 841394.
190. Pensato, V., Magri, S., Bella, E. D., Tannorella, P., Bersano, E., Sorarù, G., Gatti, M., Ticozzi, N., Taroni, F., Lauria, G., et al. (2020) Sorting Rare ALS Genetic Variants by Targeted Re-Sequencing Panel in Italian Patients: OPTN, VCP, and SQSTM1 Variants Account for 3% of Rare Genetic Forms. *Journal of clinical medicine* 9.
191. Fecto, F., Yan, J., Vemula, S. P., Liu, E., Yang, Y., Chen, W., Zheng, J. G., Shi, Y., Siddique, N., Arrat, H., et al. (2011) SQSTM1 Mutations in Familial and Sporadic Amyotrophic Lateral Sclerosis. *Archives of Neurology* 68, 1440-1446.
192. Kwok, C. T., Morris, A., and de Belleruche, J. S. (2014) Sequestosome-1 (SQSTM1) sequence variants in ALS cases in the UK: prevalence and coexistence of SQSTM1 mutations in ALS kindred with PDB. *European Journal of Human Genetics* 22, 492-496.
193. Wexler, M. Sporadic ALS. <https://alsnewstoday.com/sporadic-als/> (accessed Feb 26, 2024).
194. Logan, R., Dubel-Haag, J., Scholnicov, N., and Miller, S. J. (2022) Novel Genetic Signatures Associated With Sporadic Amyotrophic Lateral Sclerosis. *Front Genet* 13, 851496.
195. Wang, H., Guan, L., and Deng, M. (2023) Recent progress of the genetics of amyotrophic lateral sclerosis and challenges of gene therapy. *Front Neurosci* 17, 1170996.
196. Sabatelli, M., Conte, A., and Zollino, M. (2013) Clinical and genetic heterogeneity of amyotrophic lateral sclerosis. *Clinical Genetics* 83, 408-416.
197. SOD1 superoxide dismutase 1 [ *Homo sapiens* (human) ]. <https://www.ncbi.nlm.nih.gov/gene/6647> (accessed Apr 6, 2024).
198. Tainer, J. A., Getzoff, E. D., Richardson, J. S., and Richardson, D. C. (1983) Structure and mechanism of copper, zinc superoxide dismutase. *Nature* 306, 284-287.
199. Perry, J. J. P., Shin, D. S., Getzoff, E. D., and Tainer, J. A. (2010) The structural biochemistry of the superoxide dismutases. *Biochimica et Biophysica Acta (BBA) - Proteins and Proteomics* 1804, 245-262.
200. Chidambaram, S. B., Anand, N., Varma, S. R., Ramamurthy, S., Vichitra, C., Sharma, A., Mahalakshmi, A. M., and Essa, M. M. (2024) Superoxide dismutase and neurological disorders. *IBRO Neuroscience Reports* 16, 373-394.

201. Bruijn, L. I., Houseweart, M. K., Kato, S., Anderson, K. L., Anderson, S. D., Ohama, E., Reaume, A. G., Scott, R. W., and Cleveland, D. W. (1998) Aggregation and Motor Neuron Toxicity of an ALS-Linked SOD1 Mutant Independent from Wild-Type SOD1. *Science (New York, N.Y.)* 281, 1851-1854.
202. Sau, D., De Biasi, S., Vitellaro-Zuccarello, L., Riso, P., Guarnieri, S., Porrini, M., Simeoni, S., Crippa, V., Onesto, E., Palazzolo, I., et al. (2007) Mutation of SOD1 in ALS: a gain of a loss of function. *Human Molecular Genetics* 16, 1604-1618.
203. Alemasov, N. A., Ivanisenko, N. V., Ramachandran, S., and Ivanisenko, V. A. (2018) Molecular mechanisms underlying the impact of mutations in SOD1 on its conformational properties associated with amyotrophic lateral sclerosis as revealed with molecular modelling. *BMC Structural Biology* 18, 1.
204. Keskin, I., Forsgren, E., Lehmann, M., Andersen, P. M., Brännström, T., Lange, D. J., Synofzik, M., Nordström, U., Zetterström, P., Marklund, S. L., et al. (2019) The molecular pathogenesis of superoxide dismutase 1-linked ALS is promoted by low oxygen tension. *Acta Neuropathologica* 138, 85-101.
205. Baek, Y., Woo, T.-G., Ahn, J., Lee, D., Kwon, Y., Park, B.-J., and Ha, N.-C. (2022) Structural analysis of the overoxidized Cu/Zn-superoxide dismutase in ROS-induced ALS filament formation. *Communications Biology* 5, 1085.
206. SETX senataxin [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/23064> (accessed Apr 6, 2024).
207. Hasanova, Z., Klapstova, V., Porrúa, O., Stefl, R., and Sebesta, M. (2023) Human senataxin is a bona fide R-loop resolving enzyme and transcription termination factor. *Nucleic Acids Res* 51, 2818-2837.
208. Moreira, M.-C., Klur, S., Watanabe, M., Németh, A. H., Ber, I. L., Moniz, J.-C., Tranchant, C., Aubourg, P., Tazir, M., Schöls, L., et al. (2004) Senataxin, the ortholog of a yeast RNA helicase, is mutant in ataxia-ocular apraxia 2. *Nature Genetics* 36, 225-227.
209. Asaka, T., Yokoji, H., Ito, J., Yamaguchi, K., and Matsushima, A. (2006) Autosomal recessive ataxia with peripheral neuropathy and elevated AFP: Novel mutations in *SETX*. *Neurology* 66, 1580-1581.
210. Datta, N., and Hohler, A. (2013) A new SETX mutation producing AOA2 in two siblings. *International Journal of Neuroscience* 123, 670-673.
211. Nanetti, L., Cavalieri, S., Pensato, V., Erbetta, A., Pareyson, D., Panzeri, M., Zorzi, G., Antozzi, C., Moroni, I., Gellera, C., et al. (2013) SETX mutations are a frequent genetic cause of juvenile and adult onset cerebellar ataxia with neuropathy and elevated serum alpha-fetoprotein. *Orphanet Journal of Rare Diseases* 8, 123.
212. AMYOTROPHIC LATERAL SCLEROSIS 4, JUVENILE; ALS4. <https://www.omim.org/entry/602433> (accessed Apr 6, 2024).
213. Chen, Y.-Z., Bennett, C. L., Huynh, H. M., Blair, I. P., Puls, I., Irobi, J., Dierick, I., Abel, A., Kennerson, M. L., Rabin, B. A., et al. (2004) DNA/RNA Helicase Gene Mutations in a Form of Juvenile Amyotrophic Lateral Sclerosis (ALS4). *The American Journal of Human Genetics* 74, 1128-1135.
214. Bennett, C. L., Chen, Y., Vignali, M., Lo, R. S., Mason, A. G., Unal, A., Huq Saifee, N. P., Fields, S., and La Spada, A. R. (2013) Protein Interaction Analysis of Senataxin and the ALS4 L389S Mutant Yields Insights into Senataxin Post-Translational Modification and Uncovers Mutant-Specific Binding with a Brain Cytoplasmic RNA-Encoded Peptide. *PloS one* 8, e78837.
215. Tsui, A., Kouznetsova, V. L., Kesari, S., Fiala, M., and Tsigelny, I. F. (2023) Role of Senataxin in Amyotrophic Lateral Sclerosis. *Journal of Molecular Neuroscience* 73, 996-1009.
216. Giannini, M., and Porrúa, O. (2024) Senataxin: A key actor in RNA metabolism, genome integrity and neurodegeneration. *Biochimie* 217, 10-19.
217. TARDBP TAR DNA binding protein [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/23435> (accessed Apr 6, 2024).
218. Jo, M., Lee, S., Jeon, Y.-M., Kim, S., Kwon, Y., and Kim, H.-J. (2020) The role of TDP-43 propagation in neurodegenerative diseases: integrating insights from clinical and experimental studies. *Experimental & Molecular Medicine* 52, 1652-1662.

219. François-Moutal, L., Perez-Miller, S., Scott, D. D., Miranda, V. G., Mollasalehi, N., and Khanna, M. (2019) Structural Insights Into TDP-43 and Effects of Post-translational Modifications. *Frontiers in Molecular Neuroscience* 12.
220. Boer, E. M. J. d., Orié, V. K., Williams, T., Baker, M. R., Oliveira, H. M. D., Polvikoski, T., Silsby, M., Menon, P., Bos, M. v. d., Halliday, G. M., et al. (2021) TDP-43 proteinopathies: a new wave of neurodegenerative diseases. *Journal of Neurology, Neurosurgery & Psychiatry* 92, 86-95.
221. Zhang, N., Gu, D., Meng, M., and Gordon, M. L. (2020) TDP-43 Is Elevated in Plasma Neuronal-Derived Exosomes of Patients With Alzheimer's Disease. *Frontiers in Aging Neuroscience* 12.
222. Shih, Y.-H., Tu, L.-H., Chang, T.-Y., Ganesan, K., Chang, W.-W., Chang, P.-S., Fang, Y.-S., Lin, Y.-T., Jin, L.-W., and Chen, Y.-R. (2020) TDP-43 interacts with amyloid- $\beta$ , inhibits fibrillization, and worsens pathology in a model of Alzheimer's disease. *Nature Communications* 11, 5950.
223. Huang, W., Zhou, Y., Tu, L., Ba, Z., Huang, J., Huang, N., and Luo, Y. (2020) TDP-43: From Alzheimer's Disease to Limbic-Predominant Age-Related TDP-43 Encephalopathy. *Frontiers in Molecular Neuroscience* 13.
224. Meneses, A., Koga, S., O'Leary, J., Dickson, D. W., Bu, G., and Zhao, N. (2021) TDP-43 Pathology in Alzheimer's Disease. *Molecular Neurodegeneration* 16, 84.
225. Tiloca, C., Goldwurm, S., Calcagno, N., Verde, F., Peverelli, S., Calini, D., Zecchinelli, A. L., Sangalli, D., Ratti, A., Pezzoli, G., et al. (2022) TARDBP mutations in a cohort of Italian patients with Parkinson's disease and atypical parkinsonisms. *Frontiers in Aging Neuroscience* 14.
226. Neumann, M., Sampathu, D. M., Kwong, L. K., Truax, A. C., Micsenyi, M. C., Chou, T. T., Bruce, J., Schuck, T., Grossman, M., Clark, C. M., et al. (2006) Ubiquitinated TDP-43 in Frontotemporal Lobar Degeneration and Amyotrophic Lateral Sclerosis. *Science (New York, N.Y.)* 314, 130-133.
227. Hu, W. T., and Grossman, M. (2009) TDP-43 and frontotemporal dementia. *Current Neurology and Neuroscience Reports* 9, 353-358.
228. Katisko, K., Huber, N., Kokkola, T., Hartikainen, P., Krüger, J., Heikkinen, A.-L., Paananen, V., Leinonen, V., Korhonen, V. E., Helisalmi, S., et al. (2022) Serum total TDP-43 levels are decreased in frontotemporal dementia patients with C9orf72 repeat expansion or concomitant motoneuron disease phenotype. *Alzheimer's Research & Therapy* 14, 151.
229. Van Deerlin, V. M., Leverenz, J. B., Bekris, L. M., Bird, T. D., Yuan, W., Elman, L. B., Clay, D., Wood, E. M., Chen-Plotkin, A. S., Martinez-Lage, M., et al. (2008) TARDBP mutations in amyotrophic lateral sclerosis with TDP-43 neuropathology: a genetic and histopathological analysis. *The Lancet Neurology* 7, 409-416.
230. Pesiridis, G. S., Lee, V. M.-Y., and Trojanowski, J. Q. (2009) Mutations in TDP-43 link glycine-rich domain functions to amyotrophic lateral sclerosis. *Human Molecular Genetics* 18, R156-R162.
231. Sreedharan, J., Blair, I. P., Tripathi, V. B., Hu, X., Vance, C., Rogelj, B., Ackerley, S., Durnall, J. C., Williams, K. L., Buratti, E., et al. (2008) TDP-43 Mutations in Familial and Sporadic Amyotrophic Lateral Sclerosis. *Science (New York, N.Y.)* 319, 1668-1672.
232. Prasad, A., Bharathi, V., Sivalingam, V., Girdhar, A., and Patel, B. K. (2019) Molecular Mechanisms of TDP-43 Misfolding and Pathology in Amyotrophic Lateral Sclerosis. *Frontiers in Molecular Neuroscience* 12.
233. Nilaver, B. I., and Urbanski, H. F. (2023) Mechanisms underlying TDP-43 pathology and neurodegeneration: An updated Mini-Review. *Frontiers in Aging Neuroscience* 15.
234. Arseni, D., Hasegawa, M., Murzin, A. G., Kametani, F., Arai, M., Yoshida, M., and Ryskeldi-Falcon, B. (2022) Structure of pathological TDP-43 filaments from ALS with FTLD. *Nature* 601, 139-143.
235. Arseni, D., Chen, R., Murzin, A. G., Peak-Chew, S. Y., Garringer, H. J., Newell, K. L., Kametani, F., Robinson, A. C., Vidal, R., Ghetti, B., et al. (2023) TDP-43 forms amyloid filaments with a distinct fold in type A FTLD-TDP. *Nature* 620, 898-903.

236. Oiwa, K., Watanabe, S., Onodera, K., Iguchi, Y., Kinoshita, Y., Komine, O., Sobue, A., Okada, Y., Katsuno, M., and Yamanaka, K. (2023) Monomerization of TDP-43 is a key determinant for inducing TDP-43 pathology in amyotrophic lateral sclerosis. *Science Advances* 9, eadf6895.
237. OPTN optineurin [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/10133> (accessed Apr 6, 2024).
238. Zhao, S., Chen, R., Gao, Y., Lu, Y., Bai, X., and Zhang, J. (2023) Fundamental roles of the Optineurin gene in the molecular pathology of Amyotrophic Lateral Sclerosis. *Frontiers in Neuroscience* 17.
239. Ryan, T. A., and Tumbarello, D. A. (2018) Optineurin: A Coordinator of Membrane-Associated Cargo Trafficking and Autophagy. *Frontiers in Immunology* 9.
240. O'Loughlin, T., Kruppa, A. J., Ribeiro, A. L. R., Edgar, J. R., Ghannam, A., Smith, A. M., and Buss, F. (2020) OPTN recruitment to a Golgi-proximal compartment regulates immune signalling and cytokine secretion. *Journal of Cell Science* 133.
241. Osawa, T., Mizuno, Y., Fujita, Y., Takatama, M., Nakazato, Y., and Okamoto, K. (2011) Optineurin in neurodegenerative diseases. *Neuropathology* 31, 569-574.
242. Wise, J. P., Jr., and Cannon, J. (2016) From the Cover: Alterations in Optineurin Expression and Localization in Pre-clinical Parkinson's Disease Models. *Toxicological Sciences* 153, 372-381.
243. Moharir, S. C., Raghawan, A. K., and Swarup, G. (2020) Optineurin promotes aggregation of mutant huntingtin and mutant ataxin-3, and reduces cytotoxicity of aggregates. *bioRxiv*, 2020.2008.2013.249201.
244. Xu, Y., Liu, Y., Chen, X., Xu, Q., Liu, L., Liu, H., Guo, R., and Qin, Y. (2022) OPTN attenuates the neurotoxicity of abnormal Tau protein by restoring autophagy. *Translational Psychiatry* 12, 230.
245. Maruyama, H., Morino, H., Ito, H., Izumi, Y., Kato, H., Watanabe, Y., Kinoshita, Y., Kamada, M., Nodera, H., Suzuki, H., et al. (2010) Mutations of optineurin in amyotrophic lateral sclerosis. *Nature* 465, 223-226.
246. Bo, R. D., Tiloca, C., Pensato, V., Corrado, L., Ratti, A., Ticozzi, N., Corti, S., Castellotti, B., Mazzini, L., Sorarù, G., et al. (2011) Novel optineurin mutations in patients with familial and sporadic amyotrophic lateral sclerosis. *Journal of Neurology, Neurosurgery & Psychiatry* 82, 1239-1243.
247. Weishaupt, J. H., Waibel, S., Birve, A., Volk, A. E., Mayer, B., Meyer, T., Ludolph, A. C., and Andersen, P. M. (2013) A novel optineurin truncating mutation and three glaucoma-associated missense variants in patients with familial amyotrophic lateral sclerosis in Germany. *Neurobiology of Aging* 34, 1516.e1519-1516.e1515.
248. Feng, S.-M., Che, C.-H., Feng, S.-Y., Liu, C.-Y., Li, L.-Y., Li, Y.-X., Huang, H.-P., and Zou, Z.-Y. (2019) Novel mutation in optineurin causing aggressive ALS+/-frontotemporal dementia. *Annals of Clinical and Translational Neurology* 6, 2377-2383.
249. Stezin, A., Chaithra, S. P., Holla, V. V., Kamble, N., Yadav, R., and Pal, P. K. (2019) A novel <em>OPTN</em> variant causing PSP-CBS-like phenotype in familial amyotrophic lateral sclerosis. *Parkinsonism & Related Disorders* 69, 147-149.
250. Liu, Z., Li, H., Hong, C., Yue, T., Chen, C., Wang, Z., You, Q., Li, C., Xie, H., and Hu, R. (2018) ALS-Associated E478G Mutation in Human OPTN (Optineurin) Promotes Inflammation and Induces Neuronal Cell Death. *Frontiers in Immunology* 9.
251. Wong, Y. C., and Holzbaur, E. L. F. (2014) Optineurin is an autophagy receptor for damaged mitochondria in parkin-mediated mitophagy that is disrupted by an ALS-linked mutation. *Proceedings of the National Academy of Sciences* 111, E4439-E4448.
252. Wen, D., Ji, Y., Li, Y., Duan, W., Wang, Y., Li, Z., Tao, M., and Liu, Y. (2024) OPTN gene therapy increases autophagy and protects mitochondria in SOD1-G93A-expressing transgenic mice and cells. *The FEBS journal* 291, 795-813.

253. SQSTM1 sequestosome 1 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/8878> (accessed Apr 6, 2024).
254. Kumar, A. V., Mills, J., and Lapierre, L. R. (2022) Selective Autophagy Receptor p62/SQSTM1, a Pivotal Player in Stress and Aging. *Frontiers in Cell and Developmental Biology* 10.
255. Lin, X., Li, S., Zhao, Y., Ma, X., Zhang, K., He, X., and Wang, Z. (2013) Interaction Domains of p62: A Bridge Between p62 and Selective Autophagy. *DNA and Cell Biology* 32, 220-227.
256. Rubino, E., Rainero, I., Chiò, A., Rogaeva, E., Galimberti, D., Fenoglio, P., Grinberg, Y., Isaia, G., Calvo, A., Gentile, S., et al. (2012) SQSTM1 mutations in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Neurology* 79, 1556-1562.
257. Le Ber, I., Camuzat, A., Guerreiro, R., Bouya-Ahmed, K., Bras, J., Nicolas, G., Gabelle, A., Didic, M., De Septenville, A., Millicamps, S., et al. (2013) SQSTM1 Mutations in French Patients With Frontotemporal Dementia or Frontotemporal Dementia With Amyotrophic Lateral Sclerosis. *JAMA Neurology* 70, 1403-1410.
258. Sun, L., Rong, Z., Li, W., Zheng, H., Xiao, S., and Li, X. (2018) Identification of a Novel Hemizygous SQSTM1 Nonsense Mutation in Atypical Behavioral Variant Frontotemporal Dementia. *Frontiers in Aging Neuroscience* 10.
259. Benotmane, H. (2021) Dual pathogenic mutations in SQSTM1 and C9orf72 as a cause of frontotemporal dementia. *Alzheimer's & Dementia* 17, e052078.
260. Dong, W., Cui, M.-C., Hu, W.-Z., Zeng, Q., Wang, Y.-L., Zhang, W., and Huang, Y. (2022) Genetic and Molecular Evaluation of SQSTM1/p62 on the Neuropathologies of Alzheimer's Disease. *Frontiers in Aging Neuroscience* 14.
261. Hirano, M., Nakamura, Y., Saigoh, K., Sakamoto, H., Ueno, S., Isono, C., Miyamoto, K., Akamatsu, M., Mitsui, Y., and Kusunoki, S. (2013) Mutations in the gene encoding p62 in Japanese patients with amyotrophic lateral sclerosis. *Neurology* 80, 458-463.
262. Chen, Y., Zheng, Z.-Z., Chen, X., Huang, R., Yang, Y., Yuan, L., Pan, L., Hadano, S., and Shang, H.-F. (2014) SQSTM1 mutations in Han Chinese populations with sporadic amyotrophic lateral sclerosis. *Neurobiology of Aging* 35, 726.e727-726.e729.
263. Brennan, A., Layfield, R., Long, J., Williams, H. E. L., Oldham, N. J., Scott, D., and Searle, M. S. (2022) An ALS-associated variant of the autophagy receptor SQSTM1/p62 reprograms binding selectivity toward the autophagy-related hATG8 proteins. *Journal of Biological Chemistry* 298.
264. Ma, S., Attarwala, I. Y., and Xie, X.-Q. (2019) SQSTM1/p62: A Potential Target for Neurodegenerative Disease. *ACS Chemical Neuroscience* 10, 2094-2114.
265. ANG angiogenin [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/283> (accessed Apr 6, 2024).
266. Fett, J. W., Strydom, D. J., Lobb, R. R., Alderman, E. M., Bethune, J. L., Riordan, J. F., and Vallee, B. L. (1985) Isolation and characterization of angiogenin, an angiogenic protein from human carcinoma cells. *Biochemistry* 24, 5480-5486.
267. Kishimoto, K., Liu, S., Tsuji, T., Olson, K. A., and Hu, G.-f. (2005) Endogenous angiogenin in endothelial cells is a general requirement for cell proliferation and angiogenesis. *Oncogene* 24, 445-456.
268. Lyons, S. M., Fay, M. M., Akiyama, Y., Anderson, P. J., and Ivanov, P. (2017) RNA biology of angiogenin: Current state and perspectives. *RNA biology* 14, 171-178.
269. Sheng, J., and Xu, Z. (2016) Three decades of research on angiogenin: a review and perspective. *Acta Biochimica et Biophysica Sinica* 48, 399-410.
270. Wu, D., Yu, W., Kishikawa, H., Folkerth, R. D., Iafrate, A. J., Shen, Y., Xin, W., Sims, K., and Hu, G.-f. (2007) Angiogenin loss-of-function mutations in amyotrophic lateral sclerosis. *Annals of Neurology* 62, 609-617.

271. Greenway, M. J., Andersen, P. M., Russ, C., Ennis, S., Cashman, S., Donaghy, C., Patterson, V., Swingler, R., Kieran, D., Prehn, J., et al. (2006) ANG mutations segregate with familial and 'sporadic' amyotrophic lateral sclerosis. *Nature Genetics* 38, 411-413.
272. Thiyagarajan, N., Ferguson, R., Subramanian, V., and Acharya, K. R. (2012) Structural and molecular insights into the mechanism of action of human angiogenin-ALS variants in neurons. *Nature Communications* 3, 1121.
273. Bradshaw, W. J., Rehman, S., Pham, T. T. K., Thiyagarajan, N., Lee, R. L., Subramanian, V., and Acharya, K. R. (2017) Structural insights into human angiogenin variants implicated in Parkinson's disease and Amyotrophic Lateral Sclerosis. *Scientific reports* 7, 41996.
274. Aluri, K. C., Salisbury, J. P., Prehn, J. H. M., and Agar, J. N. (2020) Loss of angiogenin function is related to earlier ALS onset and a paradoxical increase in ALS duration. *Scientific reports* 10, 3715.
275. VAPB VAMP associated protein B and C [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/9217> (accessed Apr 6, 2024).
276. Skehel, P. A., Fabian-Fine, R., and Kandel, E. R. (2000) Mouse VAP33 is associated with the endoplasmic reticulum and microtubules. *Proceedings of the National Academy of Sciences* 97, 1101-1106.
277. Kors, S., Costello, J. L., and Schrader, M. (2022) VAP Proteins – From Organelle Tethers to Pathogenic Host Interactors and Their Role in Neuronal Disease. *Frontiers in Cell and Developmental Biology* 10.
278. Loewen, C. J. R., Roy, A., and Levine, T. P. (2003) A conserved ER targeting motif in three families of lipid binding proteins and in Opi1p binds VAP. *The EMBO Journal* 22, 2025-2035.
279. Rao, M., Song, W., Jiang, A., Shyr, Y., Lev, S., Greenstein, D., Brantley-Sieders, D., and Chen, J. (2012) VAMP-Associated Protein B (VAPB) Promotes Breast Tumor Growth by Modulation of Akt Activity. *PLoS one* 7, e46281.
280. Faria Assoni, A., Giove Mitsugi, T., Wardenaar, R., Oliveira Ferreira, R., Farias Jandrey, E. H., Machado Novaes, G., Fonseca de Oliveira Granha, I., Bakker, P., Kaid, C., Zatz, M., et al. (2023) Neurodegeneration-associated protein VAPB regulates proliferation in medulloblastoma. *Scientific reports* 13, 19481.
281. Nishimura, A. L., Al-Chalabi, A., and Zatz, M. (2005) A common founder for amyotrophic lateral sclerosis type 8 (ALS8) in the Brazilian population. *Human Genetics* 118, 499-500.
282. Chadi, G., Maximino, J. R., Jorge, F. M. d. H., Borba, F. C. d., Gilio, J. M., Callegaro, D., Lopes, C. G., Santos, S. N. D., and Rebelo, G. N. S. (2017) Genetic analysis of patients with familial and sporadic amyotrophic lateral sclerosis in a Brazilian Research Center. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration* 18, 249-255.
283. Kanekura, K., Nishimoto, I., Aiso, S., and Matsuoka, M. (2006) Characterization of Amyotrophic Lateral Sclerosis-linked P56S Mutation of Vesicle-associated Membrane Protein-associated Protein B (VAPB/ALS8)\*. *Journal of Biological Chemistry* 281, 30223-30233.
284. Suzuki, H., and Matsuoka, M. (2011) Amyotrophic lateral sclerosis-linked mutant VAPB enhances TDP-43-induced motor neuronal toxicity. *Journal of Neurochemistry* 119, 1099-1107.
285. Funke, A., Esser, M., Krüttgen, A., Weis, J., Mitne-Neto, M., Lazar, M., Nishimura, A., Sperfeld, A., Trillenber, P., Senderek, J., et al. (2010) The p.P56S mutation in the VAPB gene is not due to a single founder: the first European case. *Clinical Genetics* 77, 302-303.
286. Aliaga, L., Lai, C., Yu, J., Chub, N., Shim, H., Sun, L., Xie, C., Yang, W.-J., Lin, X., O'Donovan, M. J., et al. (2013) Amyotrophic lateral sclerosis-related VAPB P56S mutation differentially affects the function and survival of corticospinal and spinal motor neurons. *Human Molecular Genetics* 22, 4293-4305.
287. Moustaqim-Barrette, A., Lin, Y. Q., Pradhan, S., Neely, G. G., Bellen, H. J., and Tsuda, H. (2013) The amyotrophic lateral sclerosis 8 protein, VAP, is required for ER protein quality control. *Human Molecular Genetics* 23, 1975-1989.

288. Di, L., Chen, H., Da, Y., Wang, S., and Shen, X.-M. (2016) Atypical familial amyotrophic lateral sclerosis with initial symptoms of pain or tremor in a Chinese family harboring VAPB-P56S mutation. *Journal of Neurology* 263, 263-268.
289. Guber, R. D., Schindler, A. B., Budron, M. S., Chen, K.-I., Li, Y., Fischbeck, K. H., and Grunseich, C. (2018) Nucleocytoplasmic transport defect in a North American patient with ALS8. *Annals of Clinical and Translational Neurology* 5, 369-375.
290. Tripathi, P., Guo, H., Dreser, A., Yamoah, A., Sechi, A., Jesse, C. M., Katona, I., Doukas, P., Nikolin, S., Ernst, S., et al. (2021) Pathomechanisms of ALS8: altered autophagy and defective RNA binding protein (RBP) homeostasis due to the VAPB P56S mutation. *Cell Death & Disease* 12, 466.
291. Fasana, E., Fossati, M., Ruggiano, A., Brambillasca, S., Hoogenraad, C. C., Navone, F., Francolini, M., and Borgese, N. (2010) A VAPB mutant linked to amyotrophic lateral sclerosis generates a novel form of organized smooth endoplasmic reticulum. *The FASEB Journal* 24, 1419-1430.
292. Sun, Y.-m., Dong, Y., Wang, J., Lu, J.-h., Chen, Y., and Wu, J.-j. (2017) A novel mutation of VAPB in one Chinese familial amyotrophic lateral sclerosis pedigree and its clinical characteristics. *Journal of Neurology* 264, 2387-2393.
293. Chen, H.-J., Anagnostou, G., Chai, A., Withers, J., Morris, A., Adhikaree, J., Pennetta, G., and de Bellerocche, J. S. (2010) Characterization of the Properties of a Novel Mutation in VAPB in Familial Amyotrophic Lateral Sclerosis\*. *Journal of Biological Chemistry* 285, 40266-40281.
294. van Blitterswijk, M., van Es, M. A., Koppers, M., van Rheezen, W., Medic, J., Schelhaas, H. J., van der Kooij, A. J., de Visser, M., Veldink, J. H., and van den Berg, L. H. (2012) VAPB and C9orf72 mutations in 1 familial amyotrophic lateral sclerosis patient. *Neurobiology of Aging* 33, 2950.e2951-2950.e2954.
295. Landers, J. E., Leclerc, A. L., Shi, L., Virkud, A., Cho, T., Maxwell, M. M., Henry, A. F., Polak, M., Glass, J. D., Kwiatkowski, T. J., et al. (2008) New VAPB deletion variant and exclusion of VAPB mutations in familial ALS. *Neurology* 70, 1179-1185.
296. PRPH peripherin [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/5630> (accessed Apr 6, 2024).
297. Hol, E. M., and Capetanaki, Y. (2017) Type III Intermediate Filaments Desmin, Glial Fibrillary Acidic Protein (GFAP), Vimentin, and Peripherin. *Cold Spring Harbor Perspectives in Biology* 9.
298. Parysek, L., and Goldman, R. (1988) Distribution of a novel 57 kDa intermediate filament (IF) protein in the nervous system. *The Journal of Neuroscience* 8, 555-563.
299. Troy, C. M., Muma, N. A., Greene, L. A., Price, D. L., and Shelanski, M. L. (1990) Regulation of peripherin and neurofilament expression in regenerating rat motor neurons. *Brain Research* 529, 232-238.
300. Helfand, B. T., Mendez, M. G., Pugh, J., Delsert, C., and Goldman, R. D. (2003) A Role for Intermediate Filaments in Determining and Maintaining the Shape of Nerve Cells. *Molecular Biology of the Cell* 14, 5069-5081.
301. Sabbatini, D., Raggi, F., Ruggero, S., Seguso, M., Mandrioli, J., Cagnin, A., Briani, C., Toffanin, E., Gizzi, M., Fortuna, A., et al. (2021) Evaluation of peripherin in biofluids of patients with motor neuron diseases. *Annals of Clinical and Translational Neurology* 8, 1750-1754.
302. Keddie, S., Smyth, D., Keh, R. Y. S., Chou, M. K. L., Grant, D., Surana, S., Heslegrave, A., Zetterberg, H., Wieske, L., Michael, M., et al. (2023) Peripherin is a biomarker of axonal damage in peripheral nervous system disease. *Brain* 146, 4562-4573.
303. Xiao, S., Tjostheim, S., Sanelli, T., McLean, J. R., Horne, P., Fan, Y., Ravits, J., Strong, M. J., and Robertson, J. (2008) An Aggregate-Inducing Peripherin Isoform Generated through Intron Retention Is Upregulated in Amyotrophic Lateral Sclerosis and Associated with Disease Pathology. *The Journal of Neuroscience* 28, 1833-1840.

304. Robertson, J., Doroudchi, M. M., Nguyen, M. D., Durham, H. D., Strong, M. J., Shaw, G., Julien, J.-P., and Mushynski, W. E. (2003) A neurotoxic peripherin splice variant in a mouse model of ALS. *Journal of Cell Biology* 160, 939-949.
305. Corrado, L., Carlomagno, Y., Falasco, L., Mellone, S., Godi, M., Cova, E., Cereda, C., Testa, L., Mazzini, L., and D'Alfonso, S. (2011) A novel peripherin gene (PRPH) mutation identified in one sporadic amyotrophic lateral sclerosis patient. *Neurobiology of Aging* 32, 552.e551-552.e556.
306. CHCHD10 coiled-coil-helix-coiled-coil-helix domain containing 10 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/400916> (accessed Apr 6, 2024).
307. Modjtahedi, N., Tokatlidis, K., Dessen, P., and Kroemer, G. (2016) Mitochondrial Proteins Containing Coiled-Coil-Helix-Coiled-Coil-Helix (CHCH) Domains in Health and Disease. *Trends in Biochemical Sciences* 41, 245-260.
308. Q8WYQ3 · CHC10\_HUMAN. <https://www.uniprot.org/uniprotkb/Q8WYQ3/entry> (accessed Apr 9, 2024).
309. Penttilä, S., Jokela, M., Bouquin, H., Saukkonen, A. M., Toivanen, J., and Udd, B. (2015) Late onset spinal motor neuronopathy is caused by mutation in CHCHD10. *Annals of Neurology* 77, 163-172.
310. Ajroud-Driss, S., Fecto, F., Ajroud, K., Lalani, I., Calvo, S. E., Mootha, V. K., Deng, H.-X., Siddique, N., Tahmouh, A. J., Heiman-Patterson, T. D., et al. (2015) Mutation in the novel nuclear-encoded mitochondrial protein CHCHD10 in a family with autosomal dominant mitochondrial myopathy. *neurogenetics* 16, 1-9.
311. Chausse, A., Le Ber, I., Ait-El-Mkadem, S., Camuzat, A., de Septenville, A., Bannwarth, S., Genin, E. C., Serre, V., Augé, G., Brice, A., et al. (2014) Screening of CHCHD10 in a French cohort confirms the involvement of this gene in frontotemporal dementia with amyotrophic lateral sclerosis patients. *Neurobiology of Aging* 35, 2884.e2881-2884.e2884.
312. Bannwarth, S., Ait-El-Mkadem, S., Chausse, A., Genin, E. C., Lacas-Gervais, S., Fragaki, K., Berg-Alonso, L., Kageyama, Y., Serre, V., Moore, D. G., et al. (2014) A mitochondrial origin for frontotemporal dementia and amyotrophic lateral sclerosis through CHCHD10 involvement. *Brain* 137, 2329-2345.
313. Chiò, A., Mora, G., Sabatelli, M., Caponnetto, C., Traynor, B. J., Johnson, J. O., Nalls, M. A., Calvo, A., Moglia, C., Borghero, G., et al. (2015) CHCH10 mutations in an Italian cohort of familial and sporadic amyotrophic lateral sclerosis patients. *Neurobiology of Aging* 36, 1767.e1763-1767.e1766.
314. Jiao, B., Xiao, T., Hou, L., Gu, X., Zhou, Y., Zhou, L., Tang, B., Xu, J., and Shen, L. (2015) High prevalence of CHCHD10 mutation in patients with frontotemporal dementia from China. *Brain* 139, e21-e21.
315. TBK1 TANK binding kinase 1 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/29110> (accessed Apr 6, 2024).
316. Runde, A. P., Mack, R., S.J, P. B., and Zhang, J. (2022) The role of TBK1 in cancer pathogenesis and anticancer immunity. *Journal of Experimental & Clinical Cancer Research* 41, 135.
317. Larabi, A., Devos, Juliette M., Ng, S.-L., Nanao, Max H., Round, A., Maniatis, T., and Panne, D. (2013) Crystal Structure and Mechanism of Activation of TANK-Binding Kinase 1. *Cell Reports* 3, 734-746.
318. Zhang, C., Shang, G., Gui, X., Zhang, X., Bai, X.-c., and Chen, Z. J. (2019) Structural basis of STING binding with and phosphorylation by TBK1. *Nature* 567, 394-398.
319. Ma, X., Helgason, E., Phung, Q. T., Quan, C. L., Iyer, R. S., Lee, M. W., Bowman, K. K., Starovasnik, M. A., and Dueber, E. C. (2012) Molecular basis of Tank-binding kinase 1 activation by transautophosphorylation. *Proceedings of the National Academy of Sciences* 109, 9378-9383.
320. Revach, O.-Y., Liu, S., and Jenkins, R. W. (2020) Targeting TANK-binding kinase 1 (TBK1) in cancer. *Expert Opinion on Therapeutic Targets* 24, 1065-1078.



321. Sun, Y., Revach, O.-y., Anderson, S., Kessler, E. A., Wolfe, C. H., Jenney, A., Mills, C. E., Robitschek, E. J., Davis, T. G. R., Kim, S., et al. (2023) Targeting TBK1 to overcome resistance to cancer immunotherapy. *Nature* 615, 158-167.
322. Hasan, M., and Yan, N. (2016) Therapeutic potential of targeting TBK1 in autoimmune diseases and interferonopathies. *Pharmacological Research* 111, 336-342.
323. Bodewes, I. L. A., Huijser, E., van Helden-Meeuwssen, C. G., Tas, L., Huizinga, R., Dalm, V. A. S. H., van Hagen, P. M., Groot, N., Kamphuis, S., van Daele, P. L. A., et al. (2018) TBK1: A key regulator and potential treatment target for interferon positive Sjögren's syndrome, systemic lupus erythematosus and systemic sclerosis. *Journal of Autoimmunity* 91, 97-102.
324. Ding, C., Song, Z., Shen, A., Chen, T., and Zhang, A. (2020) Small molecules targeting the innate immune cGAS–STING–TBK1 signaling pathway. *Acta Pharmaceutica Sinica B* 10, 2272-2298.
325. Thomson, D. W., and Bergamini, G. (2021) Recent progress in small molecule TBK1 inhibitors: a patent review (2015– 2020). *Expert Opinion on Therapeutic Patents* 31, 785-794.
326. Cirulli, E. T., Lasseigne, B. N., Petrovski, S., Sapp, P. C., Dion, P. A., Leblond, C. S., Couthouis, J., Lu, Y.-F., Wang, Q., Krueger, B. J., et al. (2015) Exome sequencing in amyotrophic lateral sclerosis identifies risk genes and pathways. *Science (New York, N.Y.)* 347, 1436-1441.
327. Tsai, P.-C., Liu, Y.-C., Lin, K.-P., Liu, Y.-T., Liao, Y.-C., Hsiao, C.-T., Soong, B.-W., Yip, P.-K., and Lee, Y.-C. (2016) Mutational analysis of TBK1 in Taiwanese patients with amyotrophic lateral sclerosis. *Neurobiology of Aging* 40, 191.e111-191.e116.
328. Piaceri, I., Bessi, V., Matà, S., Polito, C., Tedde, A., Berti, V., Bagnoli, S., Braccia, A., Del Mastio, M., Pignone, A. M., et al. (2018) Association of the New Variant Tyr424Asp at TBK1 Gene with Amyotrophic Lateral Sclerosis and Cognitive Decline. *Journal of Alzheimer's Disease* 61, 41-46.
329. Oakes, J. A., Davies, M. C., and Collins, M. O. (2017) TBK1: a new player in ALS linking autophagy and neuroinflammation. *Molecular Brain* 10, 5.
330. Ye, J., Cheung, J., Gerbino, V., Ahlsén, G., Zimanyi, C., Hirsh, D., and Maniatis, T. (2019) Effects of ALS-associated TANK binding kinase 1 mutations on protein–protein interactions and kinase activity. *Proceedings of the National Academy of Sciences* 116, 24517-24526.
331. Le Ber, I., De Septenville, A., Millecamps, S., Camuzat, A., Caroppo, P., Couratier, P., Blanc, F., Lacomblez, L., Sellal, F., Fleury, M.-C., et al. (2015) TBK1 mutation frequencies in French frontotemporal dementia and amyotrophic lateral sclerosis cohorts. *Neurobiology of Aging* 36, 3116.e3115-3116.e3118.
332. Gijssels, I., Van Mossevelde, S., van der Zee, J., Sieben, A., Philtjens, S., Heeman, B., Engelborghs, S., Vandenbulcke, M., De Baets, G., Bäumer, V., et al. (2015) Loss of *TBK1* is a frequent cause of frontotemporal dementia in a Belgian cohort. *Neurology* 85, 2116-2125.
333. NEFH neurofilament heavy chain [ *Homo sapiens* (human) ]. <https://www.ncbi.nlm.nih.gov/gene/4744> (accessed Apr 6, 2024).
334. Yuan, A., Rao, M. V., Veeranna, and Nixon, R. A. (2017) Neurofilaments and Neurofilament Proteins in Health and Disease. *Cold Spring Harbor Perspectives in Biology* 9.
335. Lee, M., Xu, Z., Wong, P., and Cleveland, D. (1993) Neurofilaments are obligate heteropolymers in vivo. *Journal of Cell Biology* 122, 1337-1350.
336. Hoffman, P. N., Cleveland, D. W., Griffin, J. W., Landes, P. W., Cowan, N. J., and Price, D. L. (1987) Neurofilament gene expression: a major determinant of axonal caliber. *Proceedings of the National Academy of Sciences* 84, 3472-3476.
337. Jacquier, A., Delorme, C., Belotti, E., Juntas-Morales, R., Solé, G., Dubourg, O., Giroux, M., Maurage, C.-A., Castellani, V., Rebelo, A., et al. (2017) Cryptic amyloidogenic elements in mutant NEFH causing Charcot-Marie-Tooth 2 trigger aggregates formation and neuronal death. *Acta Neuropathologica Communications* 5, 55.

338. Ikenberg, E., Reilich, P., Abicht, A., Heller, C., Schoser, B., and Walter, M. C. (2019) Charcot-Marie-Tooth disease type 2CC due to a frameshift mutation of the neurofilament heavy polypeptide gene in an Austrian family. *Neuromuscular Disorders* 29, 392-397.
339. Pipis, M., Cortese, A., Polke, J. M., Poh, R., Vandrovцова, J., Laura, M., Skorupinska, M., Jacquier, A., Juntas-Morales, R., Latour, P., et al. (2022) Charcot-Marie-Tooth disease type 2CC due to *NEFH* variants causes a progressive, non-length-dependent, motor-predominant phenotype. *Journal of Neurology, Neurosurgery & Psychiatry* 93, 48-56.
340. Figlewicz, D. A., Krizus, A., Martinoli, M. G., Meininger, V., Dib, M., Rouleau, G. A., and Julien, J.-P. (1994) Variants of the heavy neurofilament subunit are associated with the development of amyotrophic lateral sclerosis. *Human Molecular Genetics* 3, 1757-1761.
341. Tomkins, J., Usher, P., Slade, J. Y., Ince, P. G., Curtis, A., Bushby, K., and Shaw, P. J. (1998) Novel insertion in the KSP region of the neurofilament heavy gene in amyotrophic lateral sclerosis (ALS). *NeuroReport* 9, 3967-3970.
342. Al-Chalabi, A., Andersen, P. M., Nilsson, P., Chioza, B., Andersson, J. L., Russ, C., Shaw, C. E., Powell, J. F., and Leigh, P. N. (1999) Deletions of the heavy neurofilament subunit tail in amyotrophic lateral sclerosis. *Hum Mol Genet* 8, 157-164.
343. Lin, F., Lin, W., Zhu, C., Lin, J., Zhu, J., Li, X.-Y., Wang, Z., Wang, C., and Huang, H. (2021) Sequencing of neurofilament genes identified *NEFH* Ser787Arg as a novel risk variant of sporadic amyotrophic lateral sclerosis in Chinese subjects. *BMC Medical Genomics* 14, 222.
344. FUS FUS RNA binding protein [ *Homo sapiens* (human) ]. <https://www.ncbi.nlm.nih.gov/gene/2521> (accessed Apr 6, 2024).
345. Dormann, D., and Haass, C. (2013) Fused in sarcoma (FUS): An oncogene goes awry in neurodegeneration. *Molecular and Cellular Neuroscience* 56, 475-486.
346. Jia, W., Kim, S. H., Scalf, M. A., Tonzi, P., Millikin, R. J., Guns, W. M., Liu, L., Mastrocola, A. S., Smith, L. M., Huang, T. T., et al. (2021) Fused in sarcoma regulates DNA replication timing and kinetics. *Journal of Biological Chemistry* 297.
347. Kodavati, M., Wang, H., Guo, W., Mitra, J., Hegde, P. M., Provasek, V., Rao, V. H. M., Vedula, I., Zhang, A., Mitra, S., et al. (2024) FUS unveiled in mitochondrial DNA repair and targeted ligase-1 expression rescues repair-defects in FUS-linked motor neuron disease. *Nature Communications* 15, 2156.
348. Deng, H., Gao, K., and Jankovic, J. (2014) The role of FUS gene variants in neurodegenerative diseases. *Nature Reviews Neurology* 10, 337-348.
349. Neumann, M., Rademakers, R., Roeber, S., Baker, M., Kretzschmar, H. A., and Mackenzie, I. R. A. (2009) A new subtype of frontotemporal lobar degeneration with FUS pathology. *Brain* 132, 2922-2931.
350. Kwiatkowski, T. J., Bosco, D. A., LeClerc, A. L., Tamrazian, E., Vanderburg, C. R., Russ, C., Davis, A., Gilchrist, J., Kasarskis, E. J., Munsat, T., et al. (2009) Mutations in the *FUS/TLS* Gene on Chromosome 16 Cause Familial Amyotrophic Lateral Sclerosis. *Science (New York, N.Y.)* 323, 1205-1208.
351. Vance, C., Rogelj, B., Hortobágyi, T., De Vos, K. J., Nishimura, A. L., Sreedharan, J., Hu, X., Smith, B., Ruddy, D., Wright, P., et al. (2009) Mutations in FUS, an RNA Processing Protein, Cause Familial Amyotrophic Lateral Sclerosis Type 6. *Science (New York, N.Y.)* 323, 1208-1211.
352. Chiò, A., Restagno, G., Brunetti, M., Ossola, I., Calvo, A., Mora, G., Sabatelli, M., Monsurrò, M. R., Battistini, S., Mandrioli, J., et al. (2009) Two Italian kindreds with familial amyotrophic lateral sclerosis due to FUS mutation. *Neurobiology of Aging* 30, 1272-1275.
353. Millicamps, S., Salachas, F., Cazeneuve, C., Gordon, P., Bricka, B., Camuzat, A., Guillot-Noël, L., Russaouen, O., Bruneteau, G., Pradat, P.-F., et al. (2010) *SOD1*, *ANG*, *VAPB*, *TARDBP*, and *FUS* mutations in familial amyotrophic lateral sclerosis: genotype–phenotype correlations. *Journal of Medical Genetics* 47, 554-560.

354. Hou, L., Jiao, B., Xiao, T., Zhou, L., Zhou, Z., Du, J., Yan, X., Wang, J., Tang, B., and Shen, L. (2016) Screening of SOD1, FUS and TARDBP genes in patients with amyotrophic lateral sclerosis in central-southern China. *Scientific reports* 6, 32478.
355. Liu, Z.-J., Lin, H.-X., Liu, G.-L., Tao, Q.-Q., Ni, W., Xiao, B.-G., and Wu, Z.-Y. (2017) The investigation of genetic and clinical features in Chinese patients with juvenile amyotrophic lateral sclerosis. *Clinical Genetics* 92, 267-273.
356. Dodd, K. C., Power, R., Ealing, J., and Hamdalla, H. (2019) FUS-ALS presenting with myoclonic jerks in a 17-year-old man. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration* 20, 278-280.
357. Mboukou, A., Rajendra, V., Kleinova, R., Tisné, C., Jantsch, M. F., and Barraud, P. (2021) Transportin-1: A Nuclear Import Receptor with Moonlighting Functions. *Frontiers in Molecular Biosciences* 8.
358. Dormann, D., Rodde, R., Edbauer, D., Bentmann, E., Fischer, I., Hruscha, A., Than, M. E., Mackenzie, I. R. A., Capell, A., Schmid, B., et al. (2010) ALS-associated fused in sarcoma (FUS) mutations disrupt Transportin-mediated nuclear import. *The EMBO Journal* 29, 2841-2857.
359. Nakaya, T., and Maragkakis, M. (2018) Amyotrophic Lateral Sclerosis associated FUS mutation shortens mitochondria and induces neurotoxicity. *Scientific reports* 8, 15575.
360. PFN1 profilin 1 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/5216> (accessed Apr 6, 2024).
361. Alkam, D., Feldman, E. Z., Singh, A., and Kiaei, M. (2017) Profilin1 biology and its mutation, actin(g) in disease. *Cellular and Molecular Life Sciences* 74, 967-981.
362. Karlsson, R., and Lindberg, U., Profilin, an Essential Control Element for Actin Polymerization. In *Actin-Monomer-Binding Proteins*, Lappalainen, P., Ed. Springer New York: New York, NY, 2007; pp 29-44.
363. Witke, W., Sutherland, J. D., Sharpe, A., Arai, M., and Kwiatkowski, D. J. (2001) Profilin I is essential for cell survival and cell division in early mouse development. *Proceedings of the National Academy of Sciences* 98, 3832-3836.
364. Witke, W. (2004) The role of profilin complexes in cell motility and other cellular processes. *Trends in Cell Biology* 14, 461-469.
365. Jockusch, B. M., Murk, K., and Rothkegel, M., The profile of profilins. In *Reviews of Physiology, Biochemistry and Pharmacology*, Amara, S. G.; Bamberg, E.; Fleischmann, B.; Gudermann, T.; Hebert, S. C.; Jahn, R.; Lederer, W. J.; Lill, R.; Miyajima, A.; Offermanns, S.; Zechner, R., Eds. Springer Berlin Heidelberg: Berlin, Heidelberg, 2007; pp 131-149.
366. Wang, Y., Wang, Y., Wan, R., Hu, C., and Lu, Y. (2021) Profilin 1 Protein and Its Implications for Cancers. *Oncology (Williston Park)* 35, 402-409.
367. Zou, L., Jaramillo, M., Whaley, D., Wells, A., Panchapakesa, V., Das, T., and Roy, P. (2007) Profilin-1 is a negative regulator of mammary carcinoma aggressiveness. *British Journal of Cancer* 97, 1361-1371.
368. George, L., Winship, A., Sorby, K., Dimitriadis, E., and Menkhorst, E. (2020) Profilin-1 is dysregulated in endometrioid (type I) endometrial cancer promoting cell proliferation and inhibiting pro-inflammatory cytokine production. *Biochemical and Biophysical Research Communications* 531, 459-464.
369. Wang, Y., Lu, Y., Wan, R., Wang, Y., Zhang, C., Li, M., Deng, P., Cao, L., and Hu, C. (2022) Profilin 1 Induces Tumor Metastasis by Promoting Microvesicle Secretion Through the ROCK 1/p-MLC Pathway in Non-Small Cell Lung Cancer. *Frontiers in Pharmacology* 13.
370. Wu, C.-H., Fallini, C., Ticozzi, N., Keagle, P. J., Sapp, P. C., Piotrowska, K., Lowe, P., Koppers, M., McKenna-Yasek, D., Baron, D. M., et al. (2012) Mutations in the profilin 1 gene cause familial amyotrophic lateral sclerosis. *Nature* 488, 499-503.
371. Tanaka, Y., Nonaka, T., Suzuki, G., Kametani, F., and Hasegawa, M. (2016) Gain-of-function profilin 1 mutations linked to familial amyotrophic lateral sclerosis cause seed-dependent intracellular TDP-43 aggregation. *Human Molecular Genetics* 25, 1420-1433.

372. Chi, J., Chen, J., Li, Y., Huang, Z., Wang, L., and Zhang, Y. (2020) A Familial Phenotypic and Genetic Study of Mutations in PFN1 Associated with Amyotrophic Lateral Sclerosis. *Neuroscience Bulletin* 36, 174-178.
373. Ingre, C., Landers, J. E., Rizik, N., Volk, A. E., Akimoto, C., Birve, A., Hübers, A., Keagle, P. J., Piotrowska, K., Press, R., et al. (2013) A novel phosphorylation site mutation in profilin 1 revealed in a large screen of US, Nordic, and German amyotrophic lateral sclerosis/frontotemporal dementia cohorts. *Neurobiology of Aging* 34, 1708.e1701-1708.e1706.
374. Pereira, G. R. C., Tellini, G. H. A. S., and De Mesquita, J. F. (2019) In silico analysis of PFN1 related to amyotrophic lateral sclerosis. *PloS one* 14, e0215723.
375. MATR3 matrin 3 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/9782> (accessed Apr 6, 2024).
376. P43243 · MATR3\_HUMAN. <https://www.uniprot.org/uniprotkb/P43243/entry> (accessed Apr 6, 2024).
377. Malik, A. M., and Barmada, S. J. (2021) Matrin 3 in neuromuscular disease: physiology and pathophysiology. *JCI Insight* 6.
378. Salem, A., Wilson, C. J., Rutledge, B. S., Dilliot, A., Farhan, S., Choy, W.-Y., and Duennwald, M. L. (2022) Matrin3: Disorder and ALS Pathogenesis. *Frontiers in Molecular Biosciences* 8.
379. Salton, M., Elkon, R., Borodina, T., Davydov, A., Yaspo, M.-L., Halperin, E., and Shiloh, Y. (2011) Matrin 3 Binds and Stabilizes mRNA. *PloS one* 6, e23882.
380. Coelho, M. B., Attig, J., Bellora, N., König, J., Hallegger, M., Kayikci, M., Eyra, E., Ule, J., and Smith, C. W. (2015) Nuclear matrix protein Matrin3 regulates alternative splicing and forms overlapping regulatory networks with PTB. *The EMBO Journal* 34, 653-668.
381. Salton, M., Lerenthal, Y., Wang, S.-Y., Chen, D. J., and Shiloh, Y. (2010) Involvement of Matrin 3 and SFPQ/NONO in the DNA damage response. *Cell Cycle* 9, 1568-1576.
382. Johnson, J. O., Pioro, E. P., Boehringer, A., Chia, R., Feit, H., Renton, A. E., Pliner, H. A., Abramzon, Y., Marangi, G., Winborn, B. J., et al. (2014) Mutations in the Matrin 3 gene cause familial amyotrophic lateral sclerosis. *Nature Neuroscience* 17, 664-666.
383. Lin, K.-P., Tsai, P.-C., Liao, Y.-C., Chen, W.-T., Tsai, C.-P., Soong, B.-W., and Lee, Y.-C. (2015) Mutational analysis of MATR3 in Taiwanese patients with amyotrophic lateral sclerosis. *Neurobiology of Aging* 36, 2005.e2001-2005.e2004.
384. Origone, P., Verdiani, S., Bandettini Di Poggio, M., Zuccarino, R., Vignolo, M., Caponnetto, C., and Mandich, P. (2015) A novel Arg147Trp MATR3 missense mutation in a slowly progressive ALS Italian patient. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration* 16, 530-531.
385. Tripolszki, K., Gampawar, P., Schmidt, H., Nagy, Z. F., Nagy, D., Klivényi, P., Engelhardt, J. I., and Széll, M. (2019) Comprehensive Genetic Analysis of a Hungarian Amyotrophic Lateral Sclerosis Cohort. *Frontiers in Genetics* 10.
386. Narain, P., Padhi, A. K., Dave, U., Mishra, D., Bhatia, R., Vivekanandan, P., and Gomes, J. (2019) Identification and characterization of novel and rare susceptible variants in Indian amyotrophic lateral sclerosis patients. *neurogenetics* 20, 197-208.
387. UBQLN2 ubiquilin 2 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/29978> (accessed Apr 6, 2024).
388. Zheng, T., Yang, Y., and Castañeda, C. A. (2020) Structure, dynamics and functions of UBQLNs: at the crossroads of protein quality control machinery. *Biochemical Journal* 477, 3471-3497.
389. Zhang, K. Y., Yang, S., Warraich, S. T., and Blair, I. P. (2014) Ubiquilin 2: A component of the ubiquitin–proteasome system with an emerging role in neurodegeneration. *The International Journal of Biochemistry & Cell Biology* 50, 123-126.

390. Deng, H.-X., Chen, W., Hong, S.-T., Boycott, K. M., Gorrie, G. H., Siddique, N., Yang, Y., Fecto, F., Shi, Y., Zhai, H., et al. (2011) Mutations in UBQLN2 cause dominant X-linked juvenile and adult-onset ALS and ALS/dementia. *Nature* 477, 211-215.
391. Synofzik, M., Maetzler, W., Grehl, T., Prudlo, J., vom Hagen, J. M., Haack, T., Rebassoo, P., Munz, M., Schöls, L., and Biskup, S. (2012) Screening in ALS and FTD patients reveals 3 novel UBQLN2 mutations outside the PXX domain and a pure FTD phenotype. *Neurobiology of Aging* 33, 2949.e2913-2949.e2917.
392. Williams, K. L., Warraich, S. T., Yang, S., Solski, J. A., Fernando, R., Rouleau, G. A., Nicholson, G. A., and Blair, I. P. (2012) UBQLN2/ubiquilin 2 mutation and pathology in familial amyotrophic lateral sclerosis. *Neurobiology of Aging* 33, 2527.e2523-2527.e2510.
393. Xia, Y., Yan, L. H., Huang, B., Liu, M., Liu, X., and Huang, C. (2014) Pathogenic mutation of UBQLN2 impairs its interaction with UBXD8 and disrupts endoplasmic reticulum-associated protein degradation. *Journal of Neurochemistry* 129, 99-106.
394. Renaud, L., Picher-Martel, V., Codron, P., and Julien, J.-P. (2019) Key role of UBQLN2 in pathogenesis of amyotrophic lateral sclerosis and frontotemporal dementia. *Acta Neuropathologica Communications* 7, 103.
395. ALS2 alsin Rho guanine nucleotide exchange factor ALS2 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/57679> (accessed Apr 6, 2024).
396. Topp, J. D., Gray, N. W., Gerard, R. D., and Horazdovsky, B. F. (2004) Alsln Is a Rab5 and Rac1 Guanine Nucleotide Exchange Factor \*. *Journal of Biological Chemistry* 279, 24612-24623.
397. Hadano, S., Otomo, A., Suzuki-Utsunomiya, K., Kunita, R., Yanagisawa, Y., Showguchi-Miyata, J., Mizumura, H., and Ikeda, J.-E. (2004) ALS2CL, the novel protein highly homologous to the carboxy-terminal half of ALS2, binds to Rab5 and modulates endosome dynamics. *FEBS Letters* 575, 64-70.
398. Devon, R. S., Schwab, C., Topp, J. D., Orban, P. C., Yang, Y.-z., Pape, T. D., Helm, J. R., Davidson, T.-L., Rogers, D. A., Gros-Louis, F., et al. (2005) Cross-species characterization of the ALS2 gene and analysis of its pattern of expression in development and adulthood. *Neurobiology of Disease* 18, 243-257.
399. Jacquier, A., Buhler, E., Schäfer, M. K. E., Bohl, D., Blanchard, S., Beclin, C., and Haase, G. (2006) Alsln/Rac1 signaling controls survival and growth of spinal motoneurons. *Annals of Neurology* 60, 105-117.
400. Yang, Y., Hentati, A., Deng, H.-X., Dabbagh, O., Sasaki, T., Hirano, M., Hung, W.-Y., Ouahchi, K., Yan, J., Azim, A. C., et al. (2001) The gene encoding alsin, a protein with three guanine-nucleotide exchange factor domains, is mutated in a form of recessive amyotrophic lateral sclerosis. *Nature Genetics* 29, 160-165.
401. Kress, J. A., Kühnlein, P., Winter, P., Ludolph, A. C., Kassubek, J., Müller, U., and Sperfeld, A.-D. (2005) Novel mutation in the ALS2 gene in juvenile amyotrophic lateral sclerosis. *Annals of Neurology* 58, 800-803.
402. Luigetti, M., Lattante, S., Conte, A., Romano, A., Zollino, M., Marangi, G., and Sabatelli, M. (2013) A novel compound heterozygous ALS2 mutation in two Italian siblings with juvenile amyotrophic lateral sclerosis. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration* 14, 470-472.
403. Farrugia Wismayer, M., Farrugia Wismayer, A., Borg, R., Bonavia, K., Abela, A., Chircop, C., Aquilina, J., Soler, D., Pace, A., Vella, M., et al. (2023) Genetic landscape of ALS in Malta based on a quinquennial analysis. *Neurobiology of Aging* 123, 200-207.
404. FIG4 FIG4 phosphoinositide 5-phosphatase [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/9896> (accessed Apr 6, 2024).
405. Chow, C. Y., Zhang, Y., Dowling, J. J., Jin, N., Adamska, M., Shiga, K., Szigeti, K., Shy, M. E., Li, J., Zhang, X., et al. (2007) Mutation of FIG4 causes neurodegeneration in the pale tremor mouse and patients with CMT4J. *Nature* 448, 68-72.

406. Sbrissa, D., Ikononov, O. C., Fu, Z., Ijuin, T., Gruenberg, J., Takenawa, T., and Shisheva, A. (2007) Core Protein Machinery for Mammalian Phosphatidylinositol 3,5-Bisphosphate Synthesis and Turnover That Regulates the Progression of Endosomal Transport: NOVEL SAC PHOSPHATASE JOINS THE ArPIKfyve-PIKfyve COMPLEX \*. *Journal of Biological Chemistry* 282, 23878-23891.
407. Takasuga, S., and Sasaki, T. (2013) Phosphatidylinositol-3,5-bisphosphate: metabolism and physiological functions. *The Journal of Biochemistry* 154, 211-218.
408. Jin, N., Lang, Michael J., and Weisman, Lois S. (2016) Phosphatidylinositol 3,5-bisphosphate: regulation of cellular events in space and time. *Biochemical Society Transactions* 44, 177-184.
409. Lenk, G. M., Ferguson, C. J., Chow, C. Y., Jin, N., Jones, J. M., Grant, A. E., Zolov, S. N., Winters, J. J., Giger, R. J., Dowling, J. J., et al. (2011) Pathogenic Mechanism of the FIG4 Mutation Responsible for Charcot-Marie-Tooth Disease CMT4J. *PLOS Genetics* 7, e1002104.
410. Charcot-Marie-Tooth Disease. <https://www.ninds.nih.gov/health-information/disorders/charcot-marie-tooth-disease> (accessed Apr 9, 2024).
411. Martyn, C., and Li, J. (2013) Fig4 deficiency: A newly emerged lysosomal storage disorder? *Progress in Neurobiology* 101-102, 35-45.
412. Chow, C. Y., Landers, J. E., Bergren, S. K., Sapp, P. C., Grant, A. E., Jones, J. M., Everett, L., Lenk, G. M., McKenna-Yasek, D. M., Weisman, L. S., et al. (2009) Deleterious Variants of *FIG4*, a Phosphoinositide Phosphatase, in Patients with ALS. *The American Journal of Human Genetics* 84, 85-88.
413. Cady, J., Allred, P., Bali, T., Pestronk, A., Goate, A., Miller, T. M., Mitra, R. D., Ravits, J., Harms, M. B., and Baloh, R. H. (2015) Amyotrophic lateral sclerosis onset is influenced by the burden of rare variants in known amyotrophic lateral sclerosis genes. *Annals of Neurology* 77, 100-113.
414. Nakamura, R., Sone, J., Atsuta, N., Tohnai, G., Watanabe, H., Yokoi, D., Nakatochi, M., Watanabe, H., Ito, M., Senda, J., et al. (2016) Next-generation sequencing of 28 ALS-related genes in a Japanese ALS cohort. *Neurobiology of Aging* 39, 219.e211-219.e218.
415. Krüger, S., Battke, F., Sprecher, A., Munz, M., Synofzik, M., Schöls, L., Gasser, T., Grehl, T., Prudlo, J., and Biskup, S. (2016) Rare Variants in Neurodegeneration Associated Genes Revealed by Targeted Panel Sequencing in a German ALS Cohort. *Frontiers in Molecular Neuroscience* 9.
416. Osmanovic, A., Rangnau, I., Kosfeld, A., Abdulla, S., Janssen, C., Auber, B., Raab, P., Preller, M., Petri, S., and Weber, R. G. (2017) FIG4 variants in central European patients with amyotrophic lateral sclerosis: a whole-exome and targeted sequencing study. *European Journal of Human Genetics* 25, 324-331.
417. Zhang, H., Cai, W., Chen, S., Liang, J., Wang, Z., Ren, Y., Liu, W., Zhang, X., Sun, Z., and Huang, X. (2018) Screening for possible oligogenic pathogenesis in Chinese sporadic ALS patients. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration* 19, 419-425.
418. Liu, C.-Y., Lin, J.-L., Feng, S.-Y., Che, C.-H., Huang, H.-P., and Zou, Z.-Y. (2022) Novel Variants in the FIG4 Gene Associated With Chinese Sporadic Amyotrophic Lateral Sclerosis With Slow Progression. *J Clin Neurol* 18, 41-47.
419. Zhang, X., Chow, C. Y., Sahenk, Z., Shy, M. E., Meisler, M. H., and Li, J. (2008) Mutation of FIG4 causes a rapidly progressive, asymmetric neuronal degeneration. *Brain* 131, 1990-2001.
420. KIF5A kinesin family member 5A [ *Homo sapiens* (human) ]. <https://www.ncbi.nlm.nih.gov/gene/3798> (accessed Apr 6, 2024).
421. Miki, H., Setou, M., Kaneshiro, K., and Hirokawa, N. (2001) All kinesin superfamily protein, KIF, genes in mouse and human. *Proceedings of the National Academy of Sciences* 98, 7004-7011.
422. Wang, N., and Xu, J. (2015) Functions of Kinesin Superfamily Proteins in Neuroreceptor Trafficking. *BioMed Research International* 2015, 639301.
423. Hirokawa, N., Noda, Y., Tanaka, Y., and Niwa, S. (2009) Kinesin superfamily motor proteins and intracellular transport. *Nature Reviews Molecular Cell Biology* 10, 682-696.

424. Nakagawa, T., Tanaka, Y., Matsuoka, E., Kondo, S., Okada, Y., Noda, Y., Kanai, Y., and Hirokawa, N. (1997) Identification and classification of 16 new kinesin superfamily (KIF) proteins in mouse genome. *Proceedings of the National Academy of Sciences* 94, 9654-9659.
425. Nakajima, K., Yin, X., Takei, Y., Seog, D.-H., Homma, N., and Hirokawa, N. (2012) Molecular Motor KIF5A Is Essential for GABA<sub>A</sub> Receptor Transport, and KIF5A Deletion Causes Epilepsy. *Neuron* 76, 945-961.
426. Kaan, H. Y. K., Hackney, D. D., and Kozielski, F. (2011) The Structure of the Kinesin-1 Motor-Tail Complex Reveals the Mechanism of Autoinhibition. *Science (New York, N.Y.)* 333, 883-885.
427. Blasius, T. L., Cai, D., Jih, G. T., Toret, C. P., and Verhey, K. J. (2007) Two binding partners cooperate to activate the molecular motor Kinesin-1. *Journal of Cell Biology* 176, 11-17.
428. Hares, K., Miners, J. S., Cook, A. J., Rice, C., Scolding, N., Love, S., and Wilkins, A. (2017) Overexpression of Kinesin Superfamily Motor Proteins in Alzheimer's Disease. *Journal of Alzheimer's Disease* 60, 1511-1524.
429. Wang, Q., Tian, J., Chen, H., Du, H., and Guo, L. (2019) Amyloid beta-mediated KIF5A deficiency disrupts anterograde axonal mitochondrial movement. *Neurobiology of Disease* 127, 410-418.
430. Wang, L., and Brown, A. (2010) A hereditary spastic paraplegia mutation in kinesin-1A/KIF5A disrupts neurofilament transport. *Molecular Neurodegeneration* 5, 52.
431. Cuchanski, M., and Baldwin, K. (2018) Novel Mutation in KIF5A Causing Hereditary Spastic Paraplegia with Axonal Sensorimotor Neuropathy (P2.444). *Neurology* 90, P2.444.
432. Qiu, Y., Zhong, S., Cong, L., Xin, L., Gao, X., Zhang, J., and Hong, D. (2018) A novel KIF5A gene variant causes spastic paraplegia and cerebellar ataxia. *Annals of Clinical and Translational Neurology* 5, 1415-1420.
433. Simone, M., Trabacca, A., Panzeri, E., Losito, L., Citterio, A., and Bassi, M. T. (2018) KIF5A and ALS2 Variants in a Family With Hereditary Spastic Paraplegia and Amyotrophic Lateral Sclerosis. *Frontiers in Neurology* 9.
434. Filosto, M., Piccinelli, S. C., Palmieri, I., Necchini, N., Valente, M., Zanella, I., Biasiotto, G., Lorenzo, D. D., Cereda, C., and Padovani, A. (2019) A Novel Mutation in the Stalk Domain of KIF5A Causes a Slowly Progressive Atypical Motor Syndrome. *Journal of clinical medicine* 8, 17.
435. Faruq, M., Kumar, D., Wadhwa, S., Shamim, U., Mathur, A., Parveen, S., Garg, A., and Srivastava, A. K. (2019) Intrafamilial variable spastic paraplegia/ataxia/ALS phenotype linked to a novel KIF5A mutation. *Clin Genet* 96, 271-273.
436. Zhang, K., Liu, Q., Shen, D., Tai, H., Liu, S., Wang, Z., Shi, J., Fu, H., Wu, S., Ding, Q., et al. (2019) Mutation analysis of KIF5A in Chinese amyotrophic lateral sclerosis patients. *Neurobiology of Aging* 73, 229.e221-229.e224.
437. Naruse, H., Ishiura, H., Mitsui, J., Takahashi, Y., Matsukawa, T., Sakuishi, K., Nakamagoe, K., Miyake, Z., Tamaoka, A., Goto, J., et al. (2021) Splice-site mutations in KIF5A in the Japanese case series of amyotrophic lateral sclerosis. *neurogenetics* 22, 11-17.
438. Nakamura, R., Tohnai, G., Atsuta, N., Nakatochi, M., Hayashi, N., Watanabe, H., Yokoi, D., Watanabe, H., Katsuno, M., Izumi, Y., et al. (2021) Genetic and functional analysis of KIF5A variants in Japanese patients with sporadic amyotrophic lateral sclerosis. *Neurobiology of Aging* 97, 147.e111-147.e117.
439. Baron, D. M., Fenton, A. R., Saez-Atienzar, S., Giampetruzzi, A., Sreeram, A., Shankaracharya, Keagle, P. J., Doocy, V. R., Smith, N. J., Danielson, E. W., et al. (2022) ALS-associated KIF5A mutations abolish autoinhibition resulting in a toxic gain of function. *Cell Reports* 39.
440. Nakano, J., Chiba, K., and Niwa, S. (2022) An ALS-associated KIF5A mutant forms oligomers and aggregates and induces neuronal toxicity. *Genes to Cells* 27, 421-435.

441. Nicolas, A., Kenna, K. P., Renton, A. E., Ticozzi, N., Faghri, F., Chia, R., Dominov, J. A., Kenna, B. J., Nalls, M. A., Keagle, P., et al. (2018) Genome-wide Analyses Identify KIF5A as a Novel ALS Gene. *Neuron* 97, 1267-1288.
442. ERBB4 erb-b2 receptor tyrosine kinase 4 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/2066> (accessed Apr 6, 2024).
443. Wieduwilt, M. J., and Moasser, M. M. (2008) The epidermal growth factor receptor family: Biology driving targeted therapeutics. *Cellular and Molecular Life Sciences* 65, 1566-1584.
444. Bouyain, S., Longo, P. A., Li, S., Ferguson, K. M., and Leahy, D. J. (2005) The extracellular region of ErbB4 adopts a tethered conformation in the absence of ligand. *Proceedings of the National Academy of Sciences* 102, 15024-15029.
445. El-Gamal, M. I., Mewafi, N. H., Abdelmottaleb, N. E., Emar, M. A., Tarazi, H., Sbenati, R. M., Madkour, M. M., Zarai, S.-O., Shahin, A. I., and Anbar, H. S. (2021) A Review of HER4 (ErbB4) Kinase, Its Impact on Cancer, and Its Inhibitors. *Molecules* 26, 7376.
446. Lucas, L. M., Dwivedi, V., Senfeld, J. I., Cullum, R. L., Mill, C. P., Piazza, J. T., Bryant, I. N., Cook, L. J., Miller, S. T., IV, J. H. L., et al. (2022) The Yin and Yang of ERBB4: Tumor Suppressor and Oncoprotein. *Pharmacological Reviews* 74, 18-47.
447. Borg, R., Farrugia Wismayer, M., Bonavia, K., Farrugia Wismayer, A., Vella, M., van Vugt, J. J. F. A., Kenna, B. J., Kenna, K. P., Vassallo, N., Veldink, J. H., et al. (2021) Genetic analysis of ALS cases in the isolated island population of Malta. *European Journal of Human Genetics* 29, 604-614.
448. Takahashi, Y., Fukuda, Y., Yoshimura, J., Toyoda, A., Kurppa, K., Moritoyo, H., Belzil, Veronique V., Dion, Patrick A., Higasa, K., Doi, K., et al. (2013) *ERBB4* Mutations that Disrupt the Neuregulin-ErbB4 Pathway Cause Amyotrophic Lateral Sclerosis Type 19. *The American Journal of Human Genetics* 93, 900-905.
449. Wang, F., Liu, X., He, J., Zhang, N., Chen, L., Tang, L., and Fan, D. (2022) Analysis of ERBB4 Variants in Amyotrophic Lateral Sclerosis Within a Chinese Cohort. *Frontiers in Neurology* 13.
450. Zhang, N., Chen, K.-L., Huang, Y.-Y., Chen, S.-F., Dong, Q., Tan, L., and Yu, J.-T. (2023) A new ERBB4 variant in amyotrophic lateral sclerosis type 19: Case report and review of the literature. *Clinical Neurology and Neurosurgery* 227, 107636.
451. Falls, D. L., - Neuregulins: Functions, forms, and signaling strategies. In *The EGF Receptor Family*, Carpenter, G., Ed. Academic Press: Burlington, 2003; pp 15-31.
452. Lemmon, M. A. (2009) Ligand-induced ErbB receptor dimerization. *Experimental Cell Research* 315, 638-648.
453. CCNF cyclin F [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/899> (accessed Apr 6, 2024).
454. Bai, C., Richman, R., and Elledge, S. J. (1994) Human cyclin F. *The EMBO Journal* 13, 6087-6098.
455. Galper, J., Rayner, S. L., Hogan, A. L., Fifita, J. A., Lee, A., Chung, R. S., Blair, I. P., and Yang, S. (2017) Cyclin F: A component of an E3 ubiquitin ligase complex with roles in neurodegeneration and cancer. *The International Journal of Biochemistry & Cell Biology* 89, 216-220.
456. Fu, J., Qiu, H., Cai, M., Pan, Y., Cao, Y., Liu, L., Yun, J., and Zhang, C. Z. (2013) Low cyclin F expression in hepatocellular carcinoma associates with poor differentiation and unfavorable prognosis. *Cancer Science* 104, 508-515.
457. Krajewski, A., Gagat, M., Żuryń, A., Hałas-Wiśniewska, M., Grzanka, D., and Grzanka, A. (2020) Cyclin F is involved in response to cisplatin treatment in melanoma cell lines. *Oncol Rep* 43, 765-772.
458. Zelong, Y., Han, Y., Ting, G., Yifei, W., Kun, H., Haoran, H., and Yong, C. (2021) Increased expression of Cyclin F in liver cancer predicts poor prognosis: A study based on TCGA database. *Medicine* 100.



459. Williams, K. L., Topp, S., Yang, S., Smith, B., Fifita, J. A., Warraich, S. T., Zhang, K. Y., Farrarwell, N., Vance, C., Hu, X., et al. (2016) CCNF mutations in amyotrophic lateral sclerosis and frontotemporal dementia. *Nature Communications* 7, 11253.
460. Lee, A., Rayner, S. L., Gwee, S. S. L., De Luca, A., Shahheydari, H., Sundaramoorthy, V., Ragagnin, A., Morsch, M., Radford, R., Galper, J., et al. (2018) Pathogenic mutation in the ALS/FTD gene, CCNF, causes elevated Lys48-linked ubiquitylation and defective autophagy. *Cellular and Molecular Life Sciences* 75, 335-354.
461. Tian, D., Li, J., Tang, L., Zhang, N., and Fan, D. (2018) Screening for CCNF Mutations in a Chinese Amyotrophic Lateral Sclerosis Cohort. *Frontiers in Aging Neuroscience* 10.
462. Tsai, P.-C., Liao, Y.-C., Chen, P.-L., Guo, Y.-C., Chen, Y.-H., Jih, K.-Y., Lin, K.-P., Soong, B.-W., Tsai, C.-P., and Lee, Y.-C. (2018) Investigating CCNF mutations in a Taiwanese cohort with amyotrophic lateral sclerosis. *Neurobiology of Aging* 62, 243.e241-243.e246.
463. Yu, Y., Nakagawa, T., Morohoshi, A., Nakagawa, M., Ishida, N., Suzuki, N., Aoki, M., and Nakayama, K. (2019) Pathogenic mutations in the ALS gene CCNF cause cytoplasmic mislocalization of Cyclin F and elevated VCP ATPase activity. *Human Molecular Genetics* 28, 3486-3497.
464. ANXA11 annexin A11 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/311> (accessed Apr 6, 2024).
465. Mirsaedi, M., Gidfar, S., Vu, A., and Schraufnagel, D. (2016) Annexins family: insights into their functions and potential role in pathogenesis of sarcoidosis. *Journal of Translational Medicine* 14, 89.
466. Shibata, H., Kanadome, T., Sugiura, H., Yokoyama, T., Yamamuro, M., Moss, S. E., and Maki, M. (2015) A New Role for Annexin A11 in the Early Secretory Pathway via Stabilizing Sec31A Protein at the Endoplasmic Reticulum Exit Sites (ERES) <sup>\*</sup>. *Journal of Biological Chemistry* 290, 4981-4993.
467. Dudas, E. F., Tully, M. D., Foldes, T., Kelly, G., Tartaglia, G. G., and Pastore, A. (2024) The structural properties of full-length annexin A11. *Frontiers in Molecular Biosciences* 11.
468. Sarcoidosis. <https://www.mayoclinic.org/diseases-conditions/sarcoidosis/symptoms-causes/syc-20350358> (accessed Apr 6, 2024).
469. Hubers, L. M., Vos, H., Schuurman, A. R., Erken, R., Elferink, R. P. O., Burgering, B., Graaf, S. F. J. v. d., and Beuers, U. (2018) Annexin A11 is targeted by IgG4 and IgG1 autoantibodies in IgG4-related disease. *Gut* 67, 728-735.
470. Herta, T., Kersten, R., Chang, J.-C., Hubers, L., Go, S., Tolenaars, D., Paulusma, C. C., Nathanson, M. H., Elferink, R. O., van de Graaf, S. F. J., et al. (2022) Role of the IgG4-related cholangitis autoantigen annexin A11 in cholangiocyte protection. *Journal of Hepatology* 76, 319-331.
471. Smith, B. N., Topp, S. D., Fallini, C., Shibata, H., Chen, H.-J., Troakes, C., King, A., Ticozzi, N., Kenna, K. P., Soragia-Gkazi, A., et al. (2017) Mutations in the vesicular trafficking protein annexin A11 are associated with amyotrophic lateral sclerosis. *Science Translational Medicine* 9, eaad9157.
472. Zhang, K., Liu, Q., Liu, K., Shen, D., Tai, H., Shu, S., Ding, Q., Fu, H., Liu, S., Wang, Z., et al. (2018) ANXA11 mutations prevail in Chinese ALS patients with and without cognitive dementia. *Neurology Genetics* 4, e237.
473. Sainouchi, M., Hatano, Y., Tada, M., Ishihara, T., Ando, S., Kato, T., Tokunaga, J., Ito, G., Miyahara, H., Toyoshima, Y., et al. (2021) A novel splicing variant of ANXA11 in a patient with amyotrophic lateral sclerosis: histologic and biochemical features. *Acta Neuropathologica Communications* 9, 106.
474. Teyssou, E., Muratet, F., Amador, M.-D.-M., Ferrien, M., Lautrette, G., Machat, S., Boillée, S., Larmonier, T., Saker, S., Leguern, E., et al. (2021) Genetic screening of ANXA11 revealed novel mutations linked to amyotrophic lateral sclerosis. *Neurobiology of Aging* 99, 102.e111-102.e120.
475. Wang, Y., Duan, X., Zhou, X., Wang, R., Zhang, X., Cao, Z., Wang, X., Zhou, Z., Sun, Y., and Peng, D. (2022) ANXA11 mutations are associated with amyotrophic lateral sclerosis–frontotemporal dementia. *Frontiers in Neurology* 13.

476. Nahm, M., Lim, S. M., Kim, Y.-E., Park, J., Noh, M.-Y., Lee, S., Roh, J. E., Hwang, S.-M., Park, C.-K., Kim, Y. H., et al. (2020) *ANXA11* mutations in ALS cause dysregulation of calcium homeostasis and stress granule dynamics. *Science Translational Medicine* 12, eaax3993.
477. Liao, Y.-C., Fernandopulle, M. S., Wang, G., Choi, H., Hao, L., Drerup, C. M., Patel, R., Qamar, S., Nixon-Abell, J., Shen, Y., et al. (2019) RNA Granules Hitchhike on Lysosomes for Long-Distance Transport, Using Annexin A11 as a Molecular Tether. *Cell* 179, 147-164.e120.
478. Lillebostad, P. A. G., Raasakka, A., Hjellbrekke, S. J., Patil, S., Røstbø, T., Hollås, H., Sakya, S. A., Szigetvari, P. D., Vedeler, A., and Kursula, P. (2020) Structure of the ALS Mutation Target Annexin A11 Reveals a Stabilising N-Terminal Segment. *Biomolecules* 10, 660.
479. Shihora, A., Elias, R. D., Hammond, J. A., Ghirlando, R., and Deshmukh, L. (2023) ALS Variants of Annexin A11's Proline-Rich Domain Impair Its S100A6-Mediated Fibril Dissolution. *ACS Chemical Neuroscience* 14, 2583-2589.
480. NEK1 NIMA related kinase 1 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/4750> (accessed Apr 6, 2024).
481. Letwin, K., Mizzen, L., Motro, B., Ben-David, Y., Bernstein, A., and Pawson, T. (1992) A mammalian dual specificity protein kinase, Nek1, is related to the NIMA cell cycle regulator and highly expressed in meiotic germ cells. *Embo j* 11, 3521-3531.
482. Arama, E., Yanai, A., Kilfin, G., Bernstein, A., and Motro, B. (1998) Murine NIMA-related kinases are expressed in patterns suggesting distinct functions in gametogenesis and a role in the nervous system. *Oncogene* 16, 1813-1823.
483. Peres de Oliveira, A., Kazuo Issayama, L., Betim Pavan, I. C., Riback Silva, F., Diniz Melo-Hanchuk, T., Moreira Simabuco, F., and Kobarg, J. (2020) Checking NEKs: Overcoming a Bottleneck in Human Diseases. *Molecules* 25, 1778.
484. Chen, Y., Gaczynska, M., Osmulski, P., Polci, R., and Riley, D. J. (2010) Phosphorylation by Nek1 regulates opening and closing of voltage dependent anion channel 1. *Biochemical and Biophysical Research Communications* 394, 798-803.
485. Monroe, G. R., Kappen, I. F. P. M., Stokman, M. F., Terhal, P. A., van den Boogaard, M.-J. H., Savelberg, S. M. C., van der Veken, L. T., van Es, R. J. J., Lens, S. M., Hengeveld, R. C., et al. (2016) Compound heterozygous NEK1 variants in two siblings with oral-facial-digital syndrome type II (Mohr syndrome). *European Journal of Human Genetics* 24, 1752-1760.
486. Thiel, C., Kessler, K., Giessler, A., Dimmler, A., Shalev, S. A., von der Haar, S., Zenker, M., Zahnleiter, D., Stöss, H., Beinder, E., et al. (2011) *NEK1* Mutations Cause Short-Rib Polydactyly Syndrome Type Majewski. *The American Journal of Human Genetics* 88, 106-114.
487. Wang, Z., Horemuzova, E., Iida, A., Guo, L., Liu, Y., Matsumoto, N., Nishimura, G., Nordgren, A., Miyake, N., Tham, E., et al. (2017) Axial spondylometaphyseal dysplasia is also caused by NEK1 mutations. *Journal of Human Genetics* 62, 503-506.
488. Lattante, S., Doronzio, P. N., Conte, A., Marangi, G., Martello, F., Bisogni, G., Meleo, E., Colavito, D., Del Giudice, E., Patanella, A. K., et al. (2021) Novel variants and cellular studies on patients' primary fibroblasts support a role for NEK1 missense variants in ALS pathogenesis. *Hum Mol Genet* 30, 65-71.
489. Riva, N., Pozzi, L., Russo, T., Pipitone, G. B., Schito, P., Domi, T., Agosta, F., Quattrini, A., Carrera, P., and Filippi, M. (2022) NEK1 Variants in a Cohort of Italian Patients With Amyotrophic Lateral Sclerosis. *Frontiers in Neuroscience* 16.
490. Yao, L., He, X., Cui, B., Zhao, F., and Zhou, C. (2021) NEK1 mutations and the risk of amyotrophic lateral sclerosis (ALS): a meta-analysis. *Neurol Sci* 42, 1277-1285.
491. Kenna, K. P., van Doormaal, P. T. C., Dekker, A. M., Ticozzi, N., Kenna, B. J., Diekstra, F. P., van Rheenen, W., van Eijk, K. R., Jones, A. R., Keagle, P., et al. (2016) NEK1 variants confer susceptibility to amyotrophic lateral sclerosis. *Nature Genetics* 48, 1037-1042.

492. Nguyen, H. P., Van Mossevelde, S., Dillen, L., De Bleecker, J. L., Moisse, M., Van Damme, P., Van Broeckhoven, C., and van der Zee, J. (2018) NEK1 genetic variability in a Belgian cohort of ALS and ALS-FTD patients. *Neurobiol Aging* 61, 255.e251-255.e257.
493. Parsons, M. P., and Raymond, L. A., Chapter 20 - Huntington Disease. In *Neurobiology of Brain Disorders*, Zigmond, M. J.; Rowland, L. P.; Coyle, J. T., Eds. Academic Press: San Diego, 2015; pp 303-320.
494. Wagner, L. A., Menalled, L., Goumeniouk, A. D., Brunner, D., and Leavitt, B. R., CHAPTER 6 - Huntington Disease. In *Animal and Translational Models for CNS Drug Discovery*, McArthur, R. A.; Borsini, F., Eds. Academic Press: San Diego, 2008; pp 207-266.
495. Ghosh, R., and Tabrizi, S. J., Chapter 17 - Huntington disease. In *Handbook of Clinical Neurology*, Geschwind, D. H.; Paulson, H. L.; Klein, C., Eds. Elsevier: 2018; Vol. 147, pp 255-278.
496. Kent, A., Huntington's Disease. In *International Encyclopedia of Public Health*, Heggenhougen, H. K., Ed. Academic Press: Oxford, 2008; pp 495-501.
497. McColgan, P., and Tabrizi, S. J. (2018) Huntington's disease: a clinical review. *European Journal of Neurology* 25, 24-34.
498. Huntington's disease. <https://www.mayoclinic.org/diseases-conditions/huntingtons-disease/diagnosis-treatment/drc-20356122> (accessed Feb 12, 2024).
499. Stoker, T. B., Mason, S. L., Greenland, J. C., Holden, S. T., Santini, H., and Barker, R. A. (2022) Huntington's disease: diagnosis and management. *Practical Neurology* 22, 32-41.
500. Huntington disease (HD) epidemiology. <https://www.rarediseaseadvisor.com/disease-info-pages/huntington-disease-epidemiology/#:~:text=They%20estimated%20that%20the%20global,around%202.7%20per%20100%2C000%20persons.&text=In%202022%2C%20researchers%20published%20an,between%202010%20and%20February%202022.> (accessed Feb 26, 2024).
501. Sari, Y. (2011) Huntington's Disease: From Mutant Huntingtin Protein to Neurotrophic Factor Therapy. *Int J Biomed Sci* 7, 89-100.
502. Bates, G. P. (2005) History of genetic disease: the molecular genetics of Huntington disease - a history. *Nat Rev Genet* 6, 766-773.
503. Imarisio, S., Carmichael, J., Korolchuk, V., Chen, C. W., Saiki, S., Rose, C., Krishna, G., Davies, J. E., Ttofi, E., Underwood, B. R., et al. (2008) Huntington's disease: from pathology and genetics to potential therapies. *Biochem J* 412, 191-209.
504. Sturrock, A., and Leavitt, B. R. (2010) The clinical and genetic features of Huntington disease. *J Geriatr Psychiatry Neurol* 23, 243-259.
505. Groves, M., Vonsattel, J.-P., Mazzoni, P., and Marder, K. (2003) Huntington's Disease. *Science of Aging Knowledge Environment* 2003, dn3-dn3.
506. Mahalingam, S., and Levy, L. M. (2014) Genetics of Huntington Disease. *American Journal of Neuroradiology* 35, 1070-1072.
507. Jurcau, A. (2022) Molecular Pathophysiological Mechanisms in Huntington's Disease. *Biomedicines* 10.
508. Barron, J. C., Hurley, E. P., and Parsons, M. P. (2021) Huntingtin and the Synapse. *Frontiers in Cellular Neuroscience* 15.
509. Huntington's Disease. <https://www.ninds.nih.gov/health-information/disorders/huntingtons-disease#:~:text=Early%20signs%20of%20HD%20can,increases%20the%20chances%20of%20falling.> (accessed Feb 26, 2024).
510. Roos, R. A. C. (2010) Huntington's disease: a clinical review. *Orphanet Journal of Rare Diseases* 5, 40.
511. Tampi, R. R., and Weber, M. Early Warnings: Neuropsychiatric Manifestations of Huntington Disease. <https://www.psychiatrytimes.com/view/early-warnings-neuropsychiatric-manifestations-huntington-disease> (accessed Feb 26, 2024).

512. Paulsen, J. S. (2011) Cognitive impairment in Huntington disease: diagnosis and treatment. *Curr Neurol Neurosci Rep* 11, 474-483.
513. Friedman, N. P., and Robbins, T. W. (2022) The role of prefrontal cortex in cognitive control and executive function. *Neuropsychopharmacology* 47, 72-89.
514. McAllister, B., Gusella, J. F., Landwehrmeyer, G. B., Lee, J.-M., MacDonald, M. E., Orth, M., Rosser, A. E., Williams, N. M., Holmans, P., Jones, L., et al. (2021) Timing and Impact of Psychiatric, Cognitive, and Motor Abnormalities in Huntington Disease. *Neurology* 96, e2395-e2406.
515. Peixoto, C., Rego, D., Bicho, M., Coelho, J., and Medeiros, H., *Psychiatric symptoms in huntington's disease*. *Eur Psychiatry*. 2021 Aug 13;64(Suppl 1):S254-5. doi: 10.1192/j.eurpsy.2021.682. eCollection 2021 Apr.
516. Douglas, I., Evans, S., Rawlins, M. D., Smeeth, L., Tabrizi, S. J., and Wexler, N. S. (2013) Juvenile Huntington's disease: a population-based study using the General Practice Research Database. *BMJ Open* 3.
517. Yero, T., and Rey, J. A. (2008) Tetrabenazine (Xenazine), An FDA-Approved Treatment Option For Huntington's Disease-Related Chorea. P t 33, 690-694.
518. AUSTEDO™ (deutetrabenazine) tablets, for oral use. [https://www.accessdata.fda.gov/drugsatfda\\_docs/label/2017/208082s000lbl.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/label/2017/208082s000lbl.pdf) (accessed Mar 18, 2024).
519. TEVA announces FDA approval of AUSTEDO® XR (Deutetrabenazine) extended-release tablets, a new once-daily formulation of AUSTEDO® (Deutetrabenazine) tablets. <https://hdsa.org/news/eva-announces-fda-approval-of-austedo-xr-deutetrabenazine-extended-release-tablets-a-new-once-daily-formulation-of-austedo-deutetrabenazine-tablets/> (accessed Mar 18, 2024).
520. Dhingra, H., and Gaidhane, S. A. (2023) Huntington's Disease: Understanding Its Novel Drugs and Treatments. *Cureus* 15, e47526.
521. Kim, A., Lalonde, K., Truesdell, A., Gomes Welter, P., Brocardo, P. S., Rosenstock, T. R., and Gil-Mohapel, J. (2021) New Avenues for the Treatment of Huntington's Disease. *International journal of molecular sciences* 22.
522. Muthane, U. (2011) Predictive genetic testing in Huntington's disease. *Ann Indian Acad Neurol* 14, S29-30.
523. Andrew, K. M., and Fox, L. M. (2023) Supporting Huntington's Disease Families Through the Ups and Downs of Clinical Trials. *J Huntingtons Dis* 12, 71-76.
524. Current HSG Clinical Trials & Studies. <https://huntingtonstudygroup.org/current-clinical-trials/> (accessed Fer 26, 2024).
525. Rocha, N. P., Colpo, G. D., Teixeira, A. L., and Stimming, E. F. Clinical Trials for Huntington Disease. <https://practicalneurology.com/articles/2020-june/clinical-trials-for-huntington-disease> (accessed Feb 26, 2024).
526. Jones, L., and Hughes, A. (2011) Pathogenic mechanisms in Huntington's disease. *Int Rev Neurobiol* 98, 373-418.
527. Irfan, Z., Khanam, S., Karmakar, V., Firdous, S. M., El Khier, B., Khan, I., Rehman, M. U., and Khan, A. (2022) Pathogenesis of Huntington's Disease: An Emphasis on Molecular Pathways and Prevention by Natural Remedies. *Brain Sci* 12.
528. Jimenez-Sanchez, M., Licitra, F., Underwood, B. R., and Rubinsztein, D. C. (2017) Huntington's Disease: Mechanisms of Pathogenesis and Therapeutic Strategies. *Cold Spring Harb Perspect Med* 7.
529. The Huntington Gene. <https://health.ucdavis.edu/huntingtons/genetic-change.html> (accessed Feb 26, 2024).
530. Daldin, M., Fodale, V., Cariulo, C., Azzollini, L., Verani, M., Martufi, P., Spiezia, M. C., Deguire, S. M., Cherubini, M., Macdonald, D., et al. (2017) Polyglutamine expansion affects huntingtin conformation in multiple Huntington's disease models. *Scientific reports* 7, 5070.

531. Finkbeiner, S. (2011) Huntington's Disease. Cold Spring Harb Perspect Biol 3.
532. Kuang, X., Nunn, K., Jiang, J., Castellano, P., Hardikar, U., Horgan, A., Kong, J., Tan, Z., and Dai, W. (2021) Structural insight into transmissible mutant huntingtin species by correlative light and electron microscopy and cryo-electron tomography. *Biochemical and Biophysical Research Communications* 560, 99-104.
533. Johri, A., and Beal, M. F. (2012) Mitochondrial dysfunction in neurodegenerative diseases. *The Journal of pharmacology and experimental therapeutics* 342, 619-630.
534. Kamitsuka, P. J., Ghanem, M. M., Ziar, R., McDonald, S. E., Thomas, M. G., and Kwakye, G. F. (2023) Defective Mitochondrial Dynamics and Protein Degradation Pathways Underlie Cadmium-Induced Neurotoxicity and Cell Death in Huntington's Disease Striatal Cells. *International journal of molecular sciences* 24.
535. Dai, Y., Wang, H., Lian, A., Li, J., Zhao, G., Hu, S., and Li, B. (2023) A comprehensive perspective of Huntington's disease and mitochondrial dysfunction. *Mitochondrion* 70, 8-19.
536. Miladinovic, T., Nashed, M. G., and Singh, G. (2015) Overview of Glutamatergic Dysregulation in Central Pathologies. *Biomolecules* 5, 3112-3141.
537. Henningsen, J. B., Soylu-Kucharz, R., Björkqvist, M., and Petersén, Å. (2021) Effects of excitotoxicity in the hypothalamus in transgenic mouse models of Huntington disease. *Heliyon* 7, e07808.
538. Anglada-Huguet, M., Laura, V.-S., Nuria, C.-L., Jordi, A., and Xavier, X., Pathogenesis of Huntington's Disease: How to Fight Excitotoxicity and Transcriptional Dysregulation. In *Huntington's Disease*, Nagehan Ersoy, T., Ed. IntechOpen: Rijeka, 2017; p Ch. 3.
539. Lewerenz, J., and Maher, P. (2015) Chronic Glutamate Toxicity in Neurodegenerative Diseases—What is the Evidence? *Frontiers in Neuroscience* 9.
540. Trushina, E., Dyer, R. B., Badger, J. D., 2nd, Ure, D., Eide, L., Tran, D. D., Vrieze, B. T., Legendre-Guillemain, V., McPherson, P. S., Mandavilli, B. S., et al. (2004) Mutant huntingtin impairs axonal trafficking in mammalian neurons in vivo and in vitro. *Mol Cell Biol* 24, 8195-8209.
541. Berth, S. H., and Lloyd, T. E. (2023) Disruption of axonal transport in neurodegeneration. *J Clin Invest* 133.
542. Migazzi, A., Scaramuzzino, C., Anderson, E. N., Tripathy, D., Hernández, I. H., Grant, R. A., Rocuzzo, M., Tosatto, L., Virlogeux, A., Zuccato, C., et al. (2021) Huntingtin-mediated axonal transport requires arginine methylation by PRMT6. *Cell Reports* 35, 108980.
543. Jia, Q., Li, S., Li, X. J., and Yin, P. (2022) Neuroinflammation in Huntington's disease: From animal models to clinical therapeutics. *Front Immunol* 13, 1088124.
544. Rocha, N. P., Ribeiro, F. M., Furr-Stimming, E., and Teixeira, A. L. (2016) Neuroimmunology of Huntington's Disease: Revisiting Evidence from Human Studies. *Mediators Inflamm* 2016, 8653132.
545. Zhang, W., Xiao, D., Mao, Q., and Xia, H. (2023) Role of neuroinflammation in neurodegeneration development. *Signal Transduction and Targeted Therapy* 8, 267.
546. Kumar, A., Vaish, M., and Ratan, R. R. (2014) Transcriptional dysregulation in Huntington's disease: a failure of adaptive transcriptional homeostasis. *Drug Discov Today* 19, 956-962.
547. Pradhan, S., Gao, R., Bush, K., Zhang, N., Wairkar, Y. P., and Sarkar, P. S. (2022) Polyglutamine Expansion in Huntingtin and Mechanism of DNA Damage Repair Defects in Huntington's Disease. *Front Cell Neurosci* 16, 837576.
548. Hervás-Corpión, I., Guiretti, D., Alcaraz-Iborra, M., Olivares, R., Campos-Caro, A., Barco, Á., and Valor, L. M. (2018) Early alteration of epigenetic-related transcription in Huntington's disease mouse models. *Scientific reports* 8, 9925.
549. Smith-Dijak, A. I., Sepers, M. D., and Raymond, L. A. (2019) Alterations in synaptic function and plasticity in Huntington disease. *J Neurochem* 150, 346-365.

550. Cepeda, C., and Levine, M. S. (2022) Synaptic Dysfunction in Huntington's Disease: Lessons from Genetic Animal Models. *Neuroscientist* 28, 20-40.
551. Morigaki, R., and Goto, S. (2017) Striatal Vulnerability in Huntington's Disease: Neuroprotection Versus Neurotoxicity. *Brain Sci* 7.
552. Han, I., You, Y., Kordower, J. H., Brady, S. T., and Morfini, G. A. (2010) Differential vulnerability of neurons in Huntington's disease: the role of cell type-specific features. *J Neurochem* 113, 1073-1091.
553. Mätlik, K., Baffuto, M., Kus, L., Deshmukh, A. L., Davis, D. A., Paul, M. R., Carroll, T. S., Caron, M.-C., Masson, J.-Y., Pearson, C. E., et al. (2024) Cell-type-specific CAG repeat expansions and toxicity of mutant Huntingtin in human striatum and cerebellum. *Nature Genetics*.
554. Ramakrishnan, S., and Gupta, V. Trinucleotide Repeat Disorders. <https://www.ncbi.nlm.nih.gov/books/NBK559254/> (accessed Feb 14, 2024).
555. Brown, V., and Warren, S. T., Trinucleotide Repeats: Dynamic DNA and Human Disease. In *Brenner's Encyclopedia of Genetics (Second Edition)*, Maloy, S.; Hughes, K., Eds. Academic Press: San Diego, 2001; pp 181-185.
556. Reiner, A., Dragatsis, I., and Dietrich, P. (2011) Genetics and neuropathology of Huntington's disease. *Int Rev Neurobiol* 98, 325-372.
557. Ridley, R. M., Frith, C. D., Crow, T. J., and Conneally, P. M. (1988) Anticipation in Huntington's disease is inherited through the male line but may originate in the female. *Journal of Medical Genetics* 25, 589-595.
558. Kay, C., Collins, J. A., Miedzybrodzka, Z., Madore, S. J., Gordon, E. S., Gerry, N., Davidson, M., Slama, R. A., and Hayden, M. R. (2016) Huntington disease reduced penetrance alleles occur at high frequency in the general population. *Neurology* 87, 282-288.
559. McNeil, S. M., Novelletto, A., Srinidhi, J., Barnes, G., Kornbluth, I., Altherr, M. R., Wasmuth, J. J., Gusella, J. F., MacDonald, M. E., and Myers, R. H. (1997) Reduced Penetrance of the Huntington's Disease Mutation. *Human Molecular Genetics* 6, 775-779.
560. Jankovic, J., and Squitieri, F. (2017) Letter re: Huntington disease reduced penetrance alleles occur at high frequency in the general population. *Neurology* 88, 334-334.
561. Kaltenbach, L. S., Romero, E., Becklin, R. R., Chettier, R., Bell, R., Phansalkar, A., Strand, A., Torcassi, C., Savage, J., Hurlburt, A., et al. (2007) Huntingtin interacting proteins are genetic modifiers of neurodegeneration. *PLoS Genet* 3, e82.
562. (2015) Identification of Genetic Factors that Modify Clinical Onset of Huntington's Disease. *Cell* 162, 516-526.
563. Gusella, J. F., and MacDonald, M. E. (2009) Huntington's disease: the case for genetic modifiers. *Genome Medicine* 1, 80.
564. Arning, L., and Epplen, J. T. (2012) Genetic modifiers of Huntington's disease: beyond CAG. *Future Neurology* 7, 93-109.
565. Huntington's Disease: The Discovery of the Huntingtin Gene. <https://www.nature.com/scitable/topicpage/huntington-s-disease-the-discovery-of-the-851/#:~:text=It%20turns%20out%20that%20the,an%20individual%20will%20have%20HD>. (accessed Apr 10, 2024).
566. HTT gene. <https://medlineplus.gov/genetics/gene/htt/> (accessed Apr 10, 2024).
567. Huntington's disease. <https://medlineplus.gov/genetics/condition/huntingtons-disease/> (accessed Apr 10, 2024).
568. MacDonald, M. E., Ambrose, C. M., Duyao, M. P., Myers, R. H., Lin, C., Srinidhi, L., Barnes, G., Taylor, S. A., James, M., and Groot, N. (1993) A novel gene containing a trinucleotide repeat that is expanded and unstable on Huntington's disease chromosomes. *Cell* 72, 971-983.

569. Warby, S. C., Montpetit, A., Hayden, A. R., Carroll, J. B., Butland, S. L., Visscher, H., Collins, J. A., Semaka, A., Hudson, T. J., and Hayden, M. R. (2009) CAG expansion in the Huntington disease gene is associated with a specific and targetable predisposing haplogroup. *Am J Hum Genet* 84, 351-366.
570. Genetic Testing Protocol for Huntington's Disease. <https://hdsa.org/wp-content/uploads/2015/02/HDSA-Gen-Testing-Protocol-for-HD.pdf> (accessed Feb 26, 2024).
571. Huntington's disease: Genetic Testing and Counseling. <https://huntingtonsdiseasenews.com/genetic-testing-and-counseling/> (accessed Feb 26, 2024).
572. BDNF brain derived neurotrophic factor [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/627> (accessed Apr 4, 2024).
573. Hofer, M., Pagliusi, S. R., Hohn, A., Leibrock, J., and Barde, Y. A. (1990) Regional distribution of brain-derived neurotrophic factor mRNA in the adult mouse brain. *The EMBO Journal* 9, 2459-2464-2464.
574. Miranda, M., Morici, J. F., Zanoni, M. B., and Bekinschtein, P. (2019) Brain-Derived Neurotrophic Factor: A Key Molecule for Memory in the Healthy and the Pathological Brain. *Frontiers in Cellular Neuroscience* 13.
575. Cattaneo, A., Cattane, N., Begni, V., Pariante, C. M., and Riva, M. A. (2016) The human BDNF gene: peripheral gene expression and protein levels as biomarkers for psychiatric disorders. *Translational Psychiatry* 6, e958-e958.
576. Zuccato, C., Liber, D., Ramos, C., Tarditi, A., Rigamonti, D., Tartari, M., Valenza, M., and Cattaneo, E. (2005) Progressive loss of BDNF in a mouse model of Huntington's disease and rescue by BDNF delivery. *Pharmacological Research* 52, 133-139.
577. Zuccato, C., and Cattaneo, E. (2007) Role of brain-derived neurotrophic factor in Huntington's disease. *Progress in Neurobiology* 81, 294-330.
578. Giralt, A., Carretón, O., Lao-Peregrin, C., Martín, E. D., and Alberch, J. (2011) Conditional BDNF release under pathological conditions improves Huntington's disease pathology by delaying neuronal dysfunction. *Molecular Neurodegeneration* 6, 71.
579. Couly, S., Paucard, A., Bonneaud, N., Maurice, T., Benigno, L., Jourdan, C., Cohen-Solal, C., Vignes, M., and Maschat, F. (2018) Improvement of BDNF signalling by P42 peptide in Huntington's disease. *Human Molecular Genetics* 27, 3012-3028.
580. Borrell-Pagès, M., Canals, J. M., Cordelières, F. P., Parker, J. A., Pineda, J. R., Grange, G., Bryson, E. A., Guillemier, M., Hirsch, E., Hantraye, P., et al. (2006) Cystamine and cysteamine increase brain levels of BDNF in Huntington disease via HSI1b and transglutaminase. *The Journal of Clinical Investigation* 116, 1410-1424.
581. Conforti, P., Zuccato, C., Gaudenzi, G., Ieraci, A., Camnasio, S., Buckley, N. J., Mutti, C., Cotelli, F., Contini, A., and Cattaneo, E. (2013) Binding of the repressor complex REST-mSIN3b by small molecules restores neuronal gene transcription in Huntington's disease models. *Journal of Neurochemistry* 127, 22-35.
582. Gutierrez, A., Corey-Bloom, J., Thomas, E. A., and Desplats, P. (2020) Evaluation of Biochemical and Epigenetic Measures of Peripheral Brain-Derived Neurotrophic Factor (BDNF) as a Biomarker in Huntington's Disease Patients. *Frontiers in Molecular Neuroscience* 12.
583. Giampà, C., Montagna, E., Dato, C., Melone, M. A. B., Bernardi, G., and Fusco, F. R. (2013) Systemic Delivery of Recombinant Brain Derived Neurotrophic Factor (BDNF) in the R6/2 Mouse Model of Huntington's Disease. *PloS one* 8, e64037.
584. Ou, Z.-Y. A., Byrne, L. M., Rodrigues, F. B., Tortelli, R., Johnson, E. B., Foiani, M. S., Arridge, M., De Vita, E., Scahill, R. I., Heslegrave, A., et al. (2021) Brain-derived neurotrophic factor in cerebrospinal fluid and plasma is not a biomarker for Huntington's disease. *Scientific reports* 11, 3481.
585. SIRT1 sirtuin 1 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/23411> (accessed Apr 4, 2024).

586. Rahman, S., and Islam, R. (2011) Mammalian Sirt1: insights on its biological functions. *Cell Communication and Signaling* 9, 11.
587. Elibol, B., and Kilic, U. (2018) High Levels of SIRT1 Expression as a Protective Mechanism Against Disease-Related Conditions. *Frontiers in Endocrinology* 9.
588. Clark, S. J., Falchi, M., Olsson, B., Jacobson, P., Cauchi, S., Balkau, B., Marre, M., Lantieri, O., Andersson, J. C., Jernås, M., et al. (2012) Association of Sirtuin 1 (SIRT1) Gene SNPs and Transcript Expression Levels With Severe Obesity. *Obesity* 20, 178-185.
589. de Kreutzenberg, S. V., Ceolotto, G., Papparella, I., Bortoluzzi, A., Semplicini, A., Man, C. D., Cobelli, C., Fadini, G. P., and Avogaro, A. (2010) Downregulation of the Longevity-Associated Protein Sirtuin 1 in Insulin Resistance and Metabolic Syndrome: Potential Biochemical Mechanisms. *Diabetes* 59, 1006-1015.
590. Wang, W., Sun, W., Cheng, Y., Xu, Z., and Cai, L. (2019) Role of sirtuin-1 in diabetic nephropathy. *Journal of Molecular Medicine* 97, 291-309.
591. Kong, S., McBurney, M. W., and Fang, D. (2012) Sirtuin 1 in immune regulation and autoimmunity. *Immunology & Cell Biology* 90, 6-13.
592. Jiang, M., Wang, J., Fu, J., Du, L., Jeong, H., West, T., Xiang, L., Peng, Q., Hou, Z., Cai, H., et al. (2012) Neuroprotective role of Sirt1 in mammalian models of Huntington's disease through activation of multiple Sirt1 targets. *Nature Medicine* 18, 153-158.
593. Duan, W. (2013) Targeting Sirtuin-1 in Huntington's Disease: Rationale and Current Status. *CNS Drugs* 27, 345-352.
594. Neo, S. H., and Tang, B. L., Chapter Four - Sirtuins as Modifiers of Huntington's Disease (HD) Pathology. In *Prog Mol Biol Transl Sci*, Zheng, W., Ed. Academic Press: 2018; Vol. 154, pp 105-145.
595. Donmez, G., and Outeiro, T. F. (2013) SIRT1 and SIRT2: emerging targets in neurodegeneration. *EMBO Molecular Medicine* 5, 344-352.
596. Naia, L., and Rego, A. C. (2015) Sirtuins: double players in Huntington's disease. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease* 1852, 2183-2194.
597. Jiang, M., Zheng, J., Peng, Q., Hou, Z., Zhang, J., Mori, S., Ellis, J. L., Vlasuk, G. P., Fries, H., Suri, V., et al. (2014) Sirtuin 1 activator SRT2104 protects Huntington's disease mice. *Annals of Clinical and Translational Neurology* 1, 1047-1052.
598. Pallos, J., Bodai, L., Lukacsovich, T., Purcell, J. M., Steffan, J. S., Thompson, L. M., and Marsh, J. L. (2008) Inhibition of specific HDACs and sirtuins suppresses pathogenesis in a Drosophila model of Huntington's disease. *Human Molecular Genetics* 17, 3767-3775.
599. Westerberg, G., Chiesa, J. A., Andersen, C. A., Diamanti, D., Magnoni, L., Pollio, G., Darpo, B., and Zhou, M. (2015) Safety, pharmacokinetics, pharmacogenomics and QT concentration-effect modelling of the SirT1 inhibitor selisistat in healthy volunteers. *British Journal of Clinical Pharmacology* 79, 477-491.
600. Süßmuth, S. D., Haider, S., Landwehrmeyer, G. B., Farmer, R., Frost, C., Tripepi, G., Andersen, C. A., Di Bacco, M., Lamanna, C., Diodato, E., et al. (2015) An exploratory double-blind, randomized clinical trial with selisistat, a SirT1 inhibitor, in patients with Huntington's disease. *British Journal of Clinical Pharmacology* 79, 465-476.
601. HAP1 huntingtin associated protein 1 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/9001> (accessed Apr 4, 2024).
602. Page, K. J., Potter, L., Aronni, S., Everitt, B. J., and Dunnett, S. B. (1998) The expression of Huntingtin-associated protein (HAP1) mRNA in developing, adult and ageing rat CNS: implications for Huntington's disease neuropathology. *European Journal of Neuroscience* 10, 1835-1845.
603. Mackenzie, K. D., Lim, Y., Duffield, M. D., Chataway, T., Zhou, X.-F., and Keating, D. J. (2017) Huntingtin-associated protein-1 (HAP1) regulates endocytosis and interacts with multiple trafficking-related proteins. *Cellular Signalling* 35, 176-187.



604. Wu, L. L.-y., and Zhou, X.-F. (2009) Huntingtin associated protein 1 and its functions. *Cell Adhesion & Migration* 3, 71-76.
605. Chen, X., He, E., Su, C., Zeng, Y., and Xu, J. (2023) Huntingtin-associated protein 1-associated intracellular trafficking in neurodegenerative diseases. *Frontiers in Aging Neuroscience* 15.
606. Wu, L. L.-y., Fan, Y., Li, S., Li, X.-J., and Zhou, X.-F. (2010) Huntingtin-associated Protein-1 Interacts with Pro-brain-derived Neurotrophic Factor and Mediates Its Transport and Release <sup>\*</sup>. *Journal of Biological Chemistry* 285, 5614-5623.
607. Rong, J., McGuire, J. R., Fang, Z.-H., Sheng, G., Shin, J.-Y., Li, S.-H., and Li, X.-J. (2006) Regulation of Intracellular Trafficking of Huntingtin-Associated Protein-1 Is Critical for TrkA Protein Levels and Neurite Outgrowth. *The Journal of Neuroscience* 26, 6019-6030.
608. Metzger, S., Rong, J., Nguyen, H.-P., Cape, A., Tomiuk, J., Soehn, A. S., Propping, P., Freudenberg-Hua, Y., Freudenberg, J., Tong, L., et al. (2008) Huntingtin-associated protein-1 is a modifier of the age-at-onset of Huntington's disease. *Human Molecular Genetics* 17, 1137-1146.
609. Karadima, G., Dimovasili, C., Koutsis, G., Vassilopoulos, D., and Panas, M. (2012) Age at onset in Huntington's disease: Replication study on the association of HAP1. *Parkinsonism & Related Disorders* 18, 1027-1028.
610. PPARGC1A PPARG coactivator 1 alpha [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/10891> (accessed Apr 4, 2024).
611. Liang, H., and Ward, W. F. (2006) PGC-1 $\alpha$ : a key regulator of energy metabolism. *Advances in Physiology Education* 30, 145-151.
612. Stefan, N., Thamer, C., Staiger, H., Machicao, F., Machann, J. r., Schick, F., Venter, C., Niess, A., Laakso, M., Fritsche, A., et al. (2007) Genetic Variations in PPARD and PPARGC1A Determine Mitochondrial Function and Change in Aerobic Physical Fitness and Insulin Sensitivity during Lifestyle Intervention. *The Journal of Clinical Endocrinology & Metabolism* 92, 1827-1833.
613. Eynon, N., Meckel, Y., Sagiv, M., Yamin, C., Amir, R., Sagiv, M., Goldhammer, E., Duarte, J. A., and Oliveira, J. (2010) Do PPARGC1A and PPAR $\alpha$  polymorphisms influence sprint or endurance phenotypes? *Scandinavian Journal of Medicine & Science in Sports* 20, e145-e150.
614. Tural, E., Kara, N., Agaoglu, S. A., Elbistan, M., Tasmektepligil, M. Y., and Imamoglu, O. (2014) PPAR- $\alpha$  and PPARGC1A gene variants have strong effects on aerobic performance of Turkish elite endurance athletes. *Molecular Biology Reports* 41, 5799-5804.
615. Tharabenjasin, P., Pabalan, N., and Jarjanazi, H. (2019) Association of PPARGC1A Gly428Ser (rs8192678) polymorphism with potential for athletic ability and sports performance: A meta-analysis. *PloS one* 14, e0200967.
616. Hall, E. C. R., Lockey, S. J., Heffernan, S. M., Herbert, A. J., Stebbings, G. K., Day, S. H., Collins, M., Pitsiladis, Y. P., Erskine, R. M., and Williams, A. G. (2023) The PPARGC1A Gly482Ser polymorphism is associated with elite long-distance running performance. *Journal of Sports Sciences* 41, 56-62.
617. Soyal, S. M., Zara, G., Ferger, B., Felder, T. K., Kwik, M., Nofziger, C., Dossena, S., Schwienbacher, C., Hicks, A. A., Pramstaller, P. P., et al. (2019) The PPARGC1A locus and CNS-specific PGC-1 $\alpha$  isoforms are associated with Parkinson's Disease. *Neurobiology of Disease* 121, 34-46.
618. Li, L.-z., Zhao, Y.-w., Pan, H.-x., Xiang, Y.-q., Wang, Y.-g., Xu, Q., Yan, X.-x., Tan, J.-q., Li, J.-c., Tang, B.-s., et al. (2022) Association of rare PPARGC1A variants with Parkinson's disease risk. *Journal of Human Genetics* 67, 687-690.
619. Weydt, P., Pineda, V. V., Torrence, A. E., Libby, R. T., Satterfield, T. F., Lazarowski, Eduardo R., Gilbert, M. L., Morton, G. J., Bammler, T. K., Strand, A. D., et al. (2006) Thermoregulatory and metabolic defects in Huntington's disease transgenic mice implicate PGC-1 $\beta$ ; in Huntington's disease neurodegeneration. *Cell Metabolism* 4, 349-362.
620. La Spada, A. R. (2012) PPARGC1A/PGC-1 $\alpha$ , TFEB and enhanced proteostasis in Huntington disease. *Autophagy* 8, 1845-1847.

621. Cui, L., Jeong, H., Borovecki, F., Parkhurst, C. N., Tanese, N., and Krainc, D. (2006) Transcriptional Repression of PGC-1 $\beta$ ; by Mutant Huntingtin Leads to Mitochondrial Dysfunction and Neurodegeneration. *Cell* 127, 59-69.
622. Weydt, P., Soyak, S. M., Gellera, C., DiDonato, S., Weidinger, C., Oberkofler, H., Landwehrmeyer, G. B., and Patsch, W. (2009) The gene coding for PGC-1 $\alpha$  modifies age at onset in Huntington's Disease. *Molecular Neurodegeneration* 4, 3.
623. Soyak, S. M., Felder, T. K., Auer, S., Hahne, P., Oberkofler, H., Witting, A., Paulmichl, M., Landwehrmeyer, G. B., Weydt, P., Patsch, W., et al. (2012) A greatly extended PPARGC1A genomic locus encodes several new brain-specific isoforms and influences Huntington disease age of onset $\dagger$ . *Human Molecular Genetics* 21, 3461-3473.
624. CNTF ciliary neurotrophic factor [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/1270> (accessed Apr 4, 2024).
625. Ramaswamy, S., and Kordower, J. H., 11 - GENE AND CELLULAR TRANSPLANTATION THERAPIES FOR HUNTINGTON'S DISEASE. In *CNS Regeneration (Second Edition)*, Kordower, J. H.; Tuszynski, M. H., Eds. Academic Press: San Diego, 2008; pp 267-294.
626. CNTF Signaling Pathways. <https://www.rndsystems.com/pathways/cntf-signaling-pathways> (accessed Apr 4, 2024).
627. Barres, B. A., Burne, J. F., Holtmann, B., Thoenen, H., Sendtner, M., and Raff, M. C. (1996) Ciliary Neurotrophic Factor Enhances the Rate of Oligodendrocyte Generation. *Molecular and Cellular Neuroscience* 8, 146-156.
628. Linker, R. A., Mäurer, M., Gaupp, S., Martini, R., Holtmann, B., Giess, R., Rieckmann, P., Lassmann, H., Toyka, K. V., Sendtner, M., et al. (2002) CNTF is a major protective factor in demyelinating CNS disease: A neurotrophic cytokine as modulator in neuroinflammation. *Nature Medicine* 8, 620-624.
629. Giess, R., Mäurer, M., Linker, R., Gold, R., Warmuth-Metz, M., Toyka, K. V., Sendtner, M., and Rieckmann, P. (2002) Association of a Null Mutation in the CNTF Gene With Early Onset of Multiple Sclerosis. *Archives of Neurology* 59, 407-409.
630. Kuhlmann, T., Remington, L., Cognet, I., Bourbonniere, L., Zehntner, S., Guilhot, F., Herman, A., Guay-Giroux, A., Antel, J. P., Owens, T., et al. (2006) Continued Administration of Ciliary Neurotrophic Factor Protects Mice from Inflammatory Pathology in Experimental Autoimmune Encephalomyelitis. *The American Journal of Pathology* 169, 584-598.
631. Masu, Y., Wolf, E., Holtmann, B., Sendtner, M., Brem, G., and Thoenen, H. (1993) Disruption of the CNTF gene results in motor neuron degeneration. *Nature* 365, 27-32.
632. Alberch, J., Pérez-Navarro, E., and Canals, J. M., Neurotrophic factors in Huntington's disease. In *Progress in Brain Research*, Elsevier: 2004; Vol. 146, pp 197-229.
633. Mittoux, V., Ouary, S., Monville, C., Lisovoski, F., Poyot, T., Condé, F., Escartin, C., Robichon, R., Brouillet, E., Peschanski, M., et al. (2002) Corticostriatopallidal Neuroprotection by Adenovirus-Mediated Ciliary Neurotrophic Factor Gene Transfer in a Rat Model of Progressive Striatal Degeneration. *The Journal of Neuroscience* 22, 4478-4486.
634. Bloch, J., Bachoud-Lévi, A. C., Déglon, N., Lefaucheur, J. P., Winkel, L., Palfi, S., Nguyen, J. P., Bourdet, C., Remy, P., Brugières, P., et al. (2004) Neuroprotective Gene Therapy for Huntington's Disease, Using Polymer-Encapsulated Cells Engineered to Secrete Human Ciliary Neurotrophic Factor: Results of a Phase I Study. *Human Gene Therapy* 15, 968-975.
635. Escartin, C., Hantraye, P., and Déglon, N., 22 - Transplants of CNTF-producing Cells for the Treatment of Huntington's Disease. In *Cellular Transplantation*, Halberstadt, C.; Emerich, D., Eds. Academic Press: Burlington, 2007; pp 385-398.
636. Denovan-Wright, E. M., Attis, M., Rodriguez-Lebron, E., and Mandel, R. J. (2008) Sustained striatal ciliary neurotrophic factor expression negatively affects behavior and gene expression in normal and R6/1 mice. *Journal of Neuroscience Research* 86, 1748-1757.

637. IGF1 insulin like growth factor 1 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/3479> (accessed Apr 4, 2024).
638. Pang, A. L.-Y., and Chan, W.-Y., Chapter 22 - Molecular Basis of Diseases of the Endocrine System. In *Essential Concepts in Molecular Pathology*, Coleman, W. B.; Tsongalis, G. J., Eds. Academic Press: San Diego, 2010; pp 289-307.
639. Li, J., Choi, E., Yu, H., and Bai, X.-c. (2019) Structural basis of the activation of type 1 insulin-like growth factor receptor. *Nature Communications* 10, 4567.
640. Delafontaine, P., Song, Y.-H., and Li, Y. (2004) Expression, Regulation, and Function of IGF-1, IGF-1R, and IGF-1 Binding Proteins in Blood Vessels. *Arteriosclerosis, Thrombosis, and Vascular Biology* 24, 435-444.
641. Gigantism and Acromegaly. <https://www.ncbi.nlm.nih.gov/books/NBK538261/> (accessed Apr 4, 2024).
642. Junnila, R. K., List, E. O., Berryman, D. E., Murrey, J. W., and Kopchick, J. J. (2013) The GH/IGF-1 axis in ageing and longevity. *Nature Reviews Endocrinology* 9, 366-376.
643. Wrigley, S., Arafa, D., and Tropea, D. (2017) Insulin-Like Growth Factor 1: At the Crossroads of Brain Development and Aging. *Frontiers in Cellular Neuroscience* 11.
644. Gil-Polo, C., Martinez-Horta, S.-I., Sampedro Santalo, F., Martín-Palencia, M., Gundín-Menéndez, S., Alvarez-Baños, P., Maza-Pereg, L., Calvo, S., Collazo, C., Alonso-García, E., et al. (2023) Association Between Insulin-Like Growth Factor-1 and Social Cognition in Huntington's Disease. *Movement Disorders Clinical Practice* 10, 279-284.
645. Salem, L., Saleh, N., Désaméricq, G., Youssov, K., Dolbeau, G., Cleret, L., Bourhis, M.-L., Azulay, J.-P., Krystkowiak, P., Verny, C., et al. (2016) Insulin-Like Growth Factor-1 but Not Insulin Predicts Cognitive Decline in Huntington's Disease. *PloS one* 11, e0162890.
646. Moll, L., Ben-Gedalya, T., Reuveni, H., and Cohen, E. (2016) The inhibition of IGF-1 signaling promotes proteostasis by enhancing protein aggregation and deposition. *The FASEB Journal* 30, 1656-1669.
647. Lopes, C., Ribeiro, M., Duarte, A. I., Humbert, S., Saudou, F., Pereira de Almeida, L., Hayden, M., and Rego, A. C. (2014) IGF-1 Intranasal Administration Rescues Huntington's Disease Phenotypes in YAC128 Mice. *Molecular Neurobiology* 49, 1126-1142.
648. Myasthenia gravis. <https://www.mayoclinic.org/diseases-conditions/myasthenia-gravis/symptoms-causes/syc-20352036> (accessed Mar 18, 2024).
649. Farrugia, M. E., and Goodfellow, J. A. (2020) A Practical Approach to Managing Patients With Myasthenia Gravis—Opinions and a Review of the Literature. *Frontiers in Neurology* 11.
650. Jayam Trough, A., Dabi, A., Solieman, N., Kurukumbi, M., and Kalyanam, J. (2012) Myasthenia Gravis: A Review. *Autoimmune Diseases* 2012, 874680.
651. Hughes, T. (2005) The early history of myasthenia gravis. *Neuromuscular Disorders* 15, 878-886.
652. Berrih-Aknin, S., and Le Panse, R. (2014) Myasthenia gravis: a comprehensive review of immune dysregulation and etiological mechanisms. *J Autoimmun* 52, 90-100.
653. Lazaridis, K., and Tzartos, S. J. (2020) Autoantibody Specificities in Myasthenia Gravis; Implications for Improved Diagnostics and Therapeutics. *Frontiers in Immunology* 11.
654. Ciafaloni, E., Myasthenia Gravis. In *Pediatric Clinical Advisor (Second Edition)*, Garfunkel, L. C.; Kaczorowski, J. M.; Christy, C., Eds. Mosby: Philadelphia, 2007; pp 387-388.
655. Bonovich, D. C., Chapter 64 - Myasthenia Gravis. In *Critical Care Secrets (Fourth Edition)*, Parsons, P. E.; Wiener-Kronish, J. P., Eds. Mosby: Philadelphia, 2007; pp 414-418.
656. McComas, A. J., Chapter 18 - MYASTHENIA GRAVIS. In *Neuromuscular Function and Disorders*, McComas, A. J., Ed. Butterworth-Heinemann: 1977; pp 193-208.
657. Berrih-Aknin, S., Frenkian-Cuvelier, M., and Eymard, B. (2014) Diagnostic and clinical classification of autoimmune myasthenia gravis. *Journal of Autoimmunity* 48-49, 143-148.

658. Suzuki, S., Utsugisawa, K., and Suzuki, N. (2013) Overlooked non-motor symptoms in myasthenia gravis. *Journal of Neurology, Neurosurgery & Psychiatry* 84, 989-994.
659. Restivo, D. A., Centonze, D., Alesina, A., and Marchese-Ragona, R. (2020) Myasthenia Gravis Associated With SARS-CoV-2 Infection. *Annals of Internal Medicine* 173, 1027-1028.
660. Tugasworo, D., Kurnianto, A., Retnaningsih, Andhitara, Y., Ardhini, R., and Budiman, J. (2022) The relationship between myasthenia gravis and COVID-19: a systematic review. *The Egyptian Journal of Neurology, Psychiatry and Neurosurgery* 58, 83.
661. Myasthenia gravis epidemiology. <https://www.rarediseaseadvisor.com/disease-info-pages/myasthenia-gravis-epidemiology/#:~:text=Global%20prevalence%20rates%20range%20from%20150%20to%20200%20cases%20per%201%20C000%20C000%20people.&text=The%20prevalence%20of%20MG%20in,between%2036%20C000%20and%2060%20C000%20cases.&text=In%20Europe%2C%20an%20estimated%2056%20C000%20to%20123%20C000%20individuals%20live%20with%20MG>. (accessed Feb 26 2024).
662. Dresser, L., Wlodarski, R., Reznia, K., and Soliven, B. (2021) Myasthenia Gravis: Epidemiology, Pathophysiology and Clinical Manifestations. *Journal of clinical medicine* 10.
663. Koneczny, I., Martinez, P. M., and De Baets, M., Myasthenia Gravis. In *Encyclopedia of Immunobiology*, Ratcliffe, M. J. H., Ed. Academic Press: Oxford, 2016; pp 168-179.
664. Verschuuren, J., Strijbos, E., and Vincent, A., Chapter 24 - Neuromuscular junction disorders. In *Handbook of Clinical Neurology*, Pittock, S. J.; Vincent, A., Eds. Elsevier: 2016; Vol. 133, pp 447-466.
665. Szilagyi, E., Sundivakkam, P., Nunez, T., Premenand, K., Kenyon, N., and Bartholomew, A., Chapter 37 - Clinical Aspects of Regenerative Medicine: Immune System. In *Translational Regenerative Medicine*, Atala, A.; Allickson, J. G., Eds. Academic Press: Boston, 2015; pp 507-526.
666. Pal, J., Rozsa, C., Komoly, S., and Illes, Z. (2011) Clinical and biological heterogeneity of autoimmune myasthenia gravis. *J Neuroimmunol* 231, 43-54.
667. Nguyen, A. Myasthenia Gravis: Causes, Risk Factors, Diagnosis, Treatment and Current/Future Research. <https://app.scientist.com/blog/2023/06/08/myasthenia-gravis-causes-risk-factors-diagnosis-treatment-and-current-future-research> (accessed Feb 26, 2024).
668. Shah, A. K. Myasthenia Gravis Medication. [https://emedicine.medscape.com/article/1171206-medication?form=fpf&scode=msp&st=fpf&socialSite=google&icd=login\\_success\\_gg\\_match\\_fpf](https://emedicine.medscape.com/article/1171206-medication?form=fpf&scode=msp&st=fpf&socialSite=google&icd=login_success_gg_match_fpf) (accessed Feb 26, 2024).
669. Farmakidis, C., Pasnoor, M., Dimachkie, M. M., and Barohn, R. J. (2018) Treatment of Myasthenia Gravis. *Neurol Clin* 36, 311-337.
670. Alhaidar, M. K., Abumurad, S., Soliven, B., and Reznia, K. (2022) Current Treatment of Myasthenia Gravis. *Journal of clinical medicine* 11.
671. Aydin, Y., Ulas, A. B., Mutlu, V., Colak, A., and Eroglu, A. (2017) Thymectomy in Myasthenia Gravis. *Eurasian J Med* 49, 48-52.
672. Mantegazza, R., and Cavalcante, P. (2019) Diagnosis and treatment of myasthenia gravis. *Curr Opin Rheumatol* 31, 623-633.
673. Bird, S. J. Overview of the treatment of myasthenia gravis. <https://www.uptodate.com/contents/overview-of-the-treatment-of-myasthenia-gravis> (accessed Feb 26, 2024).
674. Hehir, M. K., 2nd, and Li, Y. (2022) Diagnosis and Management of Myasthenia Gravis. *Continuum (Minneapolis)* 28, 1615-1642.
675. Angelini, C. (2011) Diagnosis and management of autoimmune myasthenia gravis. *Clin Drug Investig* 31, 1-14.
676. Carr, A. S., Cardwell, C. R., McCarron, P. O., and McConville, J. (2010) A systematic review of population based epidemiological studies in Myasthenia Gravis. *BMC Neurol* 10, 46.

677. Mao, Z. F., Mo, X. A., Qin, C., Lai, Y. R., and Olde Hartman, T. C. (2010) Course and prognosis of myasthenia gravis: a systematic review. *Eur J Neurol* 17, 913-921.
678. Sieb, J. P. (2014) Myasthenia gravis: an update for the clinician. *Clin Exp Immunol* 175, 408-418.
679. Phillips, W. D., and Vincent, A. (2016) Pathogenesis of myasthenia gravis: update on disease types, models, and mechanisms. *F1000Res* 5.
680. Meriggioli, M. N., and Sanders, D. B. (2009) Autoimmune myasthenia gravis: emerging clinical and biological heterogeneity. *Lancet Neurol* 8, 475-490.
681. Ha, J. C., and Richman, D. P. (2015) Myasthenia gravis and related disorders: Pathology and molecular pathogenesis. *Biochim Biophys Acta* 1852, 651-657.
682. Gilhus, N. E., Tzartos, S., Evoli, A., Palace, J., Burns, T. M., and Verschuuren, J. J. G. M. (2019) Myasthenia gravis. *Nature Reviews Disease Primers* 5, 30.
683. Fichtner, M. L., Jiang, R., Bourke, A., Nowak, R. J., and O'Connor, K. C. (2020) Autoimmune Pathology in Myasthenia Gravis Disease Subtypes Is Governed by Divergent Mechanisms of Immunopathology. *Frontiers in Immunology* 11.
684. Wang, Z., and Yan, Y. (2017) Immunopathogenesis in Myasthenia Gravis and Neuromyelitis Optica. *Front Immunol* 8, 1785.
685. Serra, A., Ruff, R. L., and Leigh, R. J. (2012) Neuromuscular transmission failure in myasthenia gravis: decrement of safety factor and susceptibility of extraocular muscles. *Ann N Y Acad Sci* 1275, 129-135.
686. Ruff, R. L., and Lennon, V. A. (2008) How myasthenia gravis alters the safety factor for neuromuscular transmission. *J Neuroimmunol* 201-202, 13-20.
687. Howard Jr., J. F. (2018) Myasthenia gravis: the role of complement at the neuromuscular junction. *Annals of the New York Academy of Sciences* 1412, 113-128.
688. Tireli, H., Yuksel, G., Okay, T., and Tutkavul, K. (2020) Role of thymus on prognosis of myasthenia gravis in Turkish population. *North Clin Istanbul* 7, 452-459.
689. Sanders, D. B., and Massey, J. M., Chapter 7 Clinical features of myasthenia gravis. In *Handbook of Clinical Neurology*, Elsevier: 2008; Vol. 91, pp 229-252.
690. Abbas, M. Decoding the Thymus Gland: Unveiling Its Role in Immune Function and Myasthenia Gravis. <https://bnnbreaking.com/breaking-news/health/decoding-the-thymus-gland-unveiling-its-role-in-immune-function-and-myasthenia-gravis> (accessed Feb 26, 2024).
691. Zhong, H., Zhao, C., and Luo, S. (2019) HLA in myasthenia gravis: From superficial correlation to underlying mechanism. *Autoimmun Rev* 18, 102349.
692. Muñiz-Castrillo, S., Vogrig, A., and Honnorat, J. (2020) Associations between HLA and autoimmune neurological diseases with autoantibodies. *Autoimmunity Highlights* 11, 2.
693. Carlsson, B., Wallin, J., Pirskanen, R., Matell, G., and Smith, C. I. (1990) Different HLA DR-DQ associations in subgroups of idiopathic myasthenia gravis. *Immunogenetics* 31, 285-290.
694. Niks, E. H., Kuks, J. B., Roep, B. O., Haasnoot, G. W., Verduijn, W., Ballieux, B. E., De Baets, M. H., Vincent, A., and Verschuuren, J. J. (2006) Strong association of MuSK antibody-positive myasthenia gravis and HLA-DR14-DQ5. *Neurology* 66, 1772-1774.
695. Avidan, N., Le Panse, R., Berrih-Aknin, S., and Miller, A. (2014) Genetic basis of myasthenia gravis - a comprehensive review. *J Autoimmun* 52, 146-153.
696. Zagoriti, Z., Kambouris, M. E., Patrinos, G. P., Tzartos, S. J., and Poulas, K. (2013) Recent advances in genetic predisposition of myasthenia gravis. *Biomed Res Int* 2013, 404053.
697. Bao, S., Rajotte-Caron, J., Hammoudi, D., and Lin, J. (2016) Implications of Epigenetics in Myasthenia Gravis. *J Autoimmune Disord* 2.
698. Liu, F. C., Kuo, C. F., See, L. C., Tsai, H. I., and Yu, H. P. (2017) Familial aggregation of myasthenia gravis in affected families: a population-based study. *Clin Epidemiol* 9, 527-535.

699. Kerzin-Storarr, L., Metcalfe, R. A., Dyer, P. A., Kowalska, G., Ferguson, I., and Harris, R. (1988) Genetic factors in myasthenia gravis: a family study. *Neurology* 38, 38-42.
700. Al Naqbi, H., Mawart, A., Alshamsi, J., Al Safar, H., and Tay, G. K. (2021) Major histocompatibility complex (MHC) associations with diseases in ethnic groups of the Arabian Peninsula. *Immunogenetics* 73, 131-152.
701. Tiftikcioglu, B. I., Uludag, I. F., Zorlu, Y., Pirim, İ., Sener, U., Tokucoglu, F., and Korucuk, M. (2017) Human Leucocyte Antigen B50 Is Associated with Conversion to Generalized Myasthenia Gravis in Patients with Pure Ocular Onset. *Med Princ Pract* 26, 71-77.
702. Creary, L. E., Gangavarapu, S., Caillier, S. J., Cavalcante, P., Frangiamore, R., Lie, B. A., Bengtsson, M., Harbo, H. F., Brauner, S., Hollenbach, J. A., et al. (2021) Next-Generation Sequencing Identifies Extended HLA Class I and II Haplotypes Associated With Early-Onset and Late-Onset Myasthenia Gravis in Italian, Norwegian, and Swedish Populations. *Frontiers in Immunology* 12.
703. Chien, C. Y., Chang, C. W., Liao, M. F., Chu, C. C., Ro, L. S., Wu, Y. R., Chang, K. H., Chen, C. M., and Kuo, H. C. (2023) Myasthenia gravis and independent risk factors for recurrent infection: a retrospective cohort study. *BMC Neurol* 23, 255.
704. Westerberg, E. Environmental Factors of Importance in Myasthenia Gravis. Emphasis on Physical Activity. Uppsala University, Digital Comprehensive Summaries of Uppsala Dissertations from the Faculty of Medicine 1439, 2018.
705. Piñero, J., Queralt-Rosinach, N., Bravo, À., Deu-Pons, J., Bauer-Mehren, A., Baron, M., Sanz, F., and Furlong, L. I. (2015) DisGeNET: a discovery platform for the dynamical exploration of human diseases and their genes. Database 2015.
706. AGRN agrin [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/375790> (accessed Apr 4, 2024).
707. Yan, M., Xing, G.-L., Xiong, W.-C., and Mei, L. (2018) Agrin and LRP4 antibodies as new biomarkers of myasthenia gravis. *Annals of the New York Academy of Sciences* 1413, 126-135.
708. Donahue, J. E., Berzin, T. M., Rafii, M. S., Glass, D. J., Yancopoulos, G. D., Fallon, J. R., and Stopa, E. G. (1999) Agrin in Alzheimer's disease: Altered solubility and abnormal distribution within microvasculature and brain parenchyma. *Proceedings of the National Academy of Sciences* 96, 6468-6472.
709. Cotman, S. L., Halfter, W., and Cole, G. J. (2000) Agrin Binds to  $\beta$ -Amyloid (A $\beta$ ), Accelerates A $\beta$  Fibril Formation, and Is Localized to A $\beta$  Deposits in Alzheimer's Disease Brain. *Molecular and Cellular Neuroscience* 15, 183-198.
710. Liu, I.-H., Uversky, V. N., Munishkina, L. A., Fink, A. L., Halfter, W., and Cole, G. J. (2005) Agrin binds  $\alpha$ -synuclein and modulates  $\alpha$ -synuclein fibrillation. *Glycobiology* 15, 1320-1331.
711. Skeffington, K. L., Jones, F. P., Suleiman, M. S., Caputo, M., Brancaccio, A., and Bigotti, M. G. (2022) Determination of Agrin and Related Proteins Levels as a Function of Age in Human Hearts. *Frontiers in Cardiovascular Medicine* 9.
712. Jury, E. C., and Kabouridis, P. S. (2010) New role for Agrin in T cells and its potential importance in immune system regulation. *Arthritis Research & Therapy* 12, 205.
713. Gasperi, C., Melms, A., Schoser, B., Zhang, Y., Meltoranta, J., Risson, V., Schaeffer, L., Schalke, B., and Kröger, S. (2014) Anti-agrin autoantibodies in myasthenia gravis. *Neurology* 82, 1976-1983.
714. Zhang, B., Shen, C., Bealmeier, B., Ragheb, S., Xiong, W.-C., Lewis, R. A., Lisak, R. P., and Mei, L. (2014) Autoantibodies to Agrin in Myasthenia Gravis Patients. *PLoS one* 9, e91816.
715. Rivner, M. H., Quarles, B. M., Pan, J.-X., Yu, Z., Howard Jr, J. F., Corse, A., Dimachkie, M. M., Jackson, C., Vu, T., Small, G., et al. (2020) Clinical features of LRP4/agrin-antibody-positive myasthenia gravis: A multicenter study. *Muscle & Nerve* 62, 333-343.
716. Yan, M., Liu, Z., Fei, E., Chen, W., Lai, X., Luo, B., Chen, P., Jing, H., Pan, J.-x., Rivner, M. H., et al. (2018) Induction of Anti-agrin Antibodies Causes Myasthenia Gravis in Mice. *Neuroscience* 373, 113-121.

717. LRP4 LDL receptor related protein 4 [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/4038> (accessed Apr 4, 2024).
718. Chen, B.-H., Lin, Z.-Y., Zeng, X.-X., Jiang, Y.-H., and Geng, F. (2024) LRP4-related signalling pathways and their regulatory role in neurological diseases. *Brain Research* 1825, 148705.
719. Higuchi, O., Hamuro, J., Motomura, M., and Yamanashi, Y. (2011) Autoantibodies to low-density lipoprotein receptor-related protein 4 in myasthenia gravis. *Annals of Neurology* 69, 418-422.
720. Pevzner, A., Schoser, B., Peters, K., Cosma, N.-C., Karakatsani, A., Schalke, B., Melms, A., and Kröger, S. (2012) Anti-LRP4 autoantibodies in AChR- and MuSK-antibody-negative myasthenia gravis. *Journal of Neurology* 259, 427-435.
721. Zhang, B., Tzartos, J. S., Belimezi, M., Ragheb, S., Bealmeas, B., Lewis, R. A., Xiong, W.-C., Lisak, R. P., Tzartos, S. J., and Mei, L. (2012) Autoantibodies to Lipoprotein-Related Protein 4 in Patients With Double-Seronegative Myasthenia Gravis. *Archives of Neurology* 69, 445-451.
722. MUSK muscle associated receptor tyrosine kinase [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/4593> (accessed Apr 4, 2024).
723. Zong, Y., Zhang, B., Gu, S., Lee, K., Zhou, J., Yao, G., Figueiredo, D., Perry, K., Mei, L., and Jin, R. (2012) Structural basis of agrin-LRP4-MuSK signaling. *Genes & Development* 26, 247-258.
724. Zong, Y., and Jin, R. (2013) Structural mechanisms of the agrin-LRP4-MuSK signaling pathway in neuromuscular junction differentiation. *Cellular and Molecular Life Sciences* 70, 3077-3088.
725. Borges, L. S., and Richman, D. P. (2020) Muscle-Specific Kinase Myasthenia Gravis. *Frontiers in Immunology* 11.
726. Maselli, R. A., Arredondo, J., Cagney, Ó., Ng, J. J., Anderson, J. A., Williams, C., Gerke, B. J., Soliven, B., and Wollmann, R. L. (2010) Mutations in MUSK causing congenital myasthenic syndrome impair MuSK-Dok-7 interaction. *Human Molecular Genetics* 19, 2370-2379.
727. McLean, A., and Wilson, I. (2023) Congenital myasthenic syndrome from a <em>MUSK</em> gene mutation. *Practical Neurology*, pn-2023-003945.
728. Tan-Sindhunata, M. B., Mathijssen, I. B., Smit, M., Baas, F., de Vries, J. I., van der Voorn, J. P., Kluijft, I., Hagen, M. A., Blom, E. W., Sistermans, E., et al. (2015) Identification of a Dutch founder mutation in MUSK causing fetal akinesia deformation sequence. *European Journal of Human Genetics* 23, 1151-1157.
729. Wilbe, M., Ekvall, S., Eurenus, K., Ericson, K., Casar-Borota, O., Klar, J., Dahl, N., Ameer, A., Annerén, G., and Bondeson, M.-L. (2015) MuSK: a new target for lethal fetal akinesia deformation sequence (FADS). *Journal of Medical Genetics* 52, 195-202.
730. Fetal akinesia deformation sequence. <https://www.orpha.net/en/disease/detail/994#:~:text=The%20fetal%20akinesia%2Fhypokinesia%20sequence,sequence%20is%20decreased%20foetal%20activity>. (accessed Apr 4, 2024).
731. FETAL AKINESIA DEFORMATION SEQUENCE 1; FADS1. <https://www.omim.org/entry/208150> (accessed Apr 4, 2024).
732. Rodolico, C., Bonanno, C., Toscano, A., and Vita, G. (2020) MuSK-Associated Myasthenia Gravis: Clinical Features and Management. *Frontiers in Neurology* 11.
733. König, N., Stetefeld, H. R., Dohmen, C., Mergenthaler, P., Kohler, S., Schönenberger, S., Bösel, J., Lee, D.-H., Gerner, S. T., Huttner, H. B., et al. (2021) MuSK-antibodies are associated with worse outcome in myasthenic crisis requiring mechanical ventilation. *Journal of Neurology* 268, 4824-4833.
734. Lavrnic, D., Losen, M., Vujic, A., Baets, M. D., Hajdukovic, L. J., Stojanovic, V., Trikić, R., Djukic, P., and Apostolski, S. (2005) The features of myasthenia gravis with autoantibodies to MuSK. *Journal of Neurology, Neurosurgery & Psychiatry* 76, 1099-1102.
735. Huang, Q., Li, F., and Zhao, S. (2022) Spotlight on MuSK positive myasthenia gravis: clinical characteristics, treatment and outcomes. *BMC Neurology* 22, 73.

736. Zhou, Y., Chen, J., Li, Z., Tan, S., Yan, C., Luo, S., Zhou, L., Song, J., Huan, X., Wang, Y., et al. (2022) Clinical Features of Myasthenia Gravis With Antibodies to MuSK Based on Age at Onset: A Multicenter Retrospective Study in China. *Frontiers in Neurology* 13.
737. Huijbers, M. G., Zhang, W., Klooster, R., Niks, E. H., Friese, M. B., Straasheijm, K. R., Thijssen, P. E., Vrolijk, H., Plomp, J. J., Vogels, P., et al. (2013) MuSK IgG4 autoantibodies cause myasthenia gravis by inhibiting binding between MuSK and Lrp4. *Proceedings of the National Academy of Sciences* 110, 20783-20788.
738. TTN titin [ Homo sapiens (human) ]. <https://www.ncbi.nlm.nih.gov/gene/7273> (accessed Apr 4, 2024).
739. Molecule of the Month: Titin. <https://pdb101.rcsb.org/motm/185#:~:text=Titin%20is%20the%20largest%20protein,organizes%20the%20thin%20actin%20filaments>. (accessed Apr 4, 2024).
740. Herzog, W. (2018) The multiple roles of titin in muscle contraction and force production. *Biophysical Reviews* 10, 1187-1199.
741. Pfeffer, G., Joseph, J. T., Innes, A. M., Frizzell, J. B., Wilson, I. J., Brownell, A. K. W., and Chinnery, P. F. (2014) Titinopathy in a Canadian Family Sharing the British Founder Haplotype. *Canadian Journal of Neurological Sciences / Journal Canadien des Sciences Neurologiques* 41, 90-94.
742. Oates, E. C., Jones, K. J., Donkervoort, S., Charlton, A., Brammah, S., Smith III, J. E., Ware, J. S., Yau, K. S., Swanson, L. C., Whiffin, N., et al. (2018) Congenital Titinopathy: Comprehensive characterization and pathogenic insights. *Annals of Neurology* 83, 1105-1124.
743. Huang, K., Duan, H.-Q., Li, Q.-X., Luo, Y.-B., Bi, F.-F., and Yang, H. (2021) Clinicopathological features of titinopathy from a Chinese neuromuscular center. *Neuropathology* 41, 349-356.
744. Yamamoto, A. M., Gajdos, P., Eymard, B., Tranchant, C., Warter, J.-M., Gomez, L., Bourquin, C., Bach, J.-F., and Garchon, H.-J. (2001) Anti-Titin Antibodies in Myasthenia Gravis: Tight Association With Thymoma and Heterogeneity of Nonthymoma Patients. *Archives of Neurology* 58, 885-890.
745. Chen, X. J., Qiao, J., Xiao, B. G., and Lu, C. Z. (2004) The significance of titin antibodies in myasthenia gravis. *Journal of Neurology* 251, 1006-1011.
746. Kim, K. H., Kim, S. W., Cho, J., Chung, H. Y., and Shin, H. Y. (2022) Anti-titin antibody is associated with more frequent hospitalization to manage thymoma-associated myasthenia gravis. *Frontiers in Neurology* 13.
747. Segura, T., Medrano, I. H., Collazo, S., Maté, C., Sguera, C., Del Rio-Bermudez, C., Casero, H., Salcedo, I., García-García, J., Alcahut-Rodríguez, C., et al. (2023) Symptoms timeline and outcomes in amyotrophic lateral sclerosis using artificial intelligence. *Scientific reports* 13, 702.
748. Körner, S., Kollwe, K., Ilsemann, J., Müller-Heine, A., Dengler, R., Krampfl, K., and Petri, S. (2013) Prevalence and prognostic impact of comorbidities in amyotrophic lateral sclerosis. *European journal of neurology* 20, 647-654.
749. Diekmann, K., Kuzma-Kozakiewicz, M., Piotrkiewicz, M., Gromicho, M., Grosskreutz, J., Andersen, P. M., de Carvalho, M., Uysal, H., Osmanovic, A., and Schreiber-Katz, O. (2020) Impact of comorbidities and co-medication on disease onset and progression in a large German ALS patient group. *Journal of Neurology* 267, 2130-2141.
750. Li, J.-Y., Sun, X.-H., Shen, D.-c., Yang, X.-Z., Liu, M.-S., and Cui, L.-Y. (2022) Clinical characteristics and prognosis of amyotrophic lateral sclerosis with autoimmune diseases. *PLoS one* 17, e0266529.
751. Fang, F., Al-Chalabi, A., Ronnevi, L. O., Turner, M. R., Wirdefeldt, K., Kamel, F., and Ye, W. (2013) Amyotrophic lateral sclerosis and cancer: a register-based study in Sweden. *Amyotroph Lateral Scler Frontotemporal Degener* 14, 362-368.
752. Freedman, D. M., Curtis, R. E., Daugherty, S. E., Goedert, J. J., Kuncl, R. W., and Tucker, M. A. (2013) The association between cancer and amyotrophic lateral sclerosis. *Cancer Causes Control* 24, 55-60.



753. Gibson, S. B., Abbott, D., Farnham, J. M., Thai, K. K., McLean, H., Figueroa, K. P., Bromberg, M. B., Pulst, S. M., and Cannon-Albright, L. (2016) Population-based risks for cancer in patients with ALS. *Neurology* 87, 289-294.
754. Riancho, J., Delgado-Alvarado, M., Andreu, M. D., Paz-Fajardo, L., Arozamena, S., Gil-Bea, F. J., and López de Munáin, A. (2021) Amyotrophic lateral sclerosis (ALS), cancer, autoimmunity and metabolic disorders: An unsolved tantalizing challenge. *British Journal of Pharmacology* 178, 1269-1278.
755. Rahmlow, M., Sorenson, E., Crum, B., Lucchinetti, C., Kantarci, O., and Carter, J. (2012) The Co-Occurrence of MS and ALS: Chance or Shared Biology? (P02.147). *Neurology* 78, P02.147-P102.147.
756. Ohlmeier, C., Saum, K.-U., Galetzka, W., Beier, D., and Gothe, H. (2019) Epidemiology and health care utilization of patients suffering from Huntington's disease in Germany: real world evidence based on German claims data. *BMC Neurology* 19, 318.
757. Zielonka, D., Witkowski, G., Puch, E. A., Lesniczak, M., Mazur-Michalek, I., Isalan, M., and Mielcarek, M. (2020) Prevalence of Non-psychiatric Comorbidities in Pre-symptomatic and Symptomatic Huntington's Disease Gene Carriers in Poland. *Frontiers in Medicine* 7.
758. Furby, H., Moore, S., Nordstroem, A. L., Houghton, R., Lambrelli, D., Graham, S., Svenningsson, P., and Petersén, Å. (2023) Comorbidities and clinical outcomes in adult- and juvenile-onset Huntington's disease: a study of linked Swedish National Registries (2002-2019). *J Neurol* 270, 864-876.
759. Fung, W. L. A., Mah, H., and Gibbons, C. (2018) F08 Co-occurrence of amyotrophic lateral sclerosis and huntington's disease – a systematic review. *Journal of Neurology, Neurosurgery & Psychiatry* 89, A43-A43.
760. Maloni, H. W., and Wallin, M. T. (2015) Presentation of multiple sclerosis with comorbid Huntington's disease. *Clinical Neurology and Neurosurgery* 136, 86-88.
761. Misra, U. K., Kalita, J., Singh, V. K., and Kumar, S. (2020) A study of comorbidities in myasthenia gravis. *Acta Neurol Belg* 120, 59-64.
762. Myasthenia gravis - Comorbidities. <https://www.rarediseaseadvisor.com/disease-info-pages/myasthenia-gravis-comorbidities/#:~:text=Thyroid%20disease%2C%20systemic%20lupus%20erythematosus,not%20pose%20a%20significant%20risk>. (accessed Mar 15, 2024).
763. Gilhus, N. E., Nacu, A., Andersen, J. B., and Owe, J. F. (2015) Myasthenia gravis and risks for comorbidity. *Eur J Neurol* 22, 17-23.
764. Diaz, B., Flores-Gavilán, P., García-Ramos, G., and Lorenzana-Mendoza, N. (2015) Myasthenia gravis and its comorbidities. *J. Neurol. Neurophysiol.* 6, 1-5.
765. Weinreb, O., Mandel, S., Bar-Am, O., Yogev-Falach, M., Avramovich-Tirosh, Y., Amit, T., and Youdim, M. B. H. (2009) Multifunctional Neuroprotective Derivatives of Rasagiline as Anti-Alzheimer's Disease Drugs. *Neurotherapeutics* 6, 163-174.
766. Fang, T., Je, G., Pacut, P., Keyhanian, K., Gao, J., and Ghasemi, M. (2022) Gene Therapy in Amyotrophic Lateral Sclerosis. *Cells* 11.
767. Amado, D. A., and Davidson, B. L. (2021) Gene therapy for ALS: A review. *Molecular Therapy* 29, 3345-3358.
768. Giovannelli, I., Higginbottom, A., Kirby, J., Azzouz, M., and Shaw, P. J. (2023) Prospects for gene replacement therapies in amyotrophic lateral sclerosis. *Nature Reviews Neurology* 19, 39-52.
769. Cappella, M., Ciotti, C., Cohen-Tannoudji, M., and Biferi, M. G. (2019) Gene Therapy for ALS—A Perspective. *International journal of molecular sciences* 20, 4388.
770. Huntington's Disease: Gene Therapy Approaches. <https://patienteducation.asgct.org/disease-treatments/huntingtons-disease>.
771. Dabrowska, M., and Olejniczak, M. (2020) Gene Therapy for Huntington's Disease Using Targeted Endonucleases. *Methods in molecular biology (Clifton, N.J.)* 2056, 269-284.

772. Dalakas, M. C. (2019) Immunotherapy in myasthenia gravis in the era of biologics. *Nature Reviews Neurology* 15, 113-124.
773. Morimoto, S., Takahashi, S., Ito, D., Daté, Y., Okada, K., Kato, C., Nakamura, S., Ozawa, F., Chyi, C. M., Nishiyama, A., et al. (2023) Phase 1/2a clinical trial in ALS with ropinirole, a drug candidate identified by iPSC drug discovery. *Cell Stem Cell* 30, 766-780.e769.
774. Youdim, M. B. (2013) Multi target neuroprotective and neurorestorative anti-Parkinson and anti-Alzheimer drugs ladostigil and m30 derived from rasagiline. *Exp Neurobiol* 22, 1-10.
775. Golko-Perez, S., Amit, T., Bar-Am, O., Youdim, M. B., and Weinreb, O. (2017) A Novel Iron Chelator-Radical Scavenger Ameliorates Motor Dysfunction and Improves Life Span and Mitochondrial Biogenesis in SOD1(G93A) ALS Mice. *Neurotox Res* 31, 230-244.
776. Study: Common Gout Medication Reduces Risk of Alzheimer's, Parkinson's, ALS. <https://www.barrowneuro.org/about/news-and-articles/press-releases/study-common-gout-medication-reduces-risk-of-alzheimers-parkinsons-als/> (accessed Mar 19, 2024).
777. Wexler, M. Gout medication decreases risk of ALS and other diseases: Study. <https://alsnewstoday.com/news/gout-medication-decreases-risk-als-other-diseases-study/> (accessed Mar 19, 2024).
778. Myasthenia Gravis Treatment. <https://www.umms.org/ummc/health-services/neurology/services/myasthenia-gravis/treatment#:~:text=Immunosuppressant%20medications%20work%20to%20lower,%2C%20Azathioprine%2C%20and%20Mycophenolate%20Mofetil.> (accessed Mar 20, 2024).
779. Lascano, A. M., and Lalive, P. H. (2021) Update in immunosuppressive therapy of myasthenia gravis. *Autoimmunity Reviews* 20, 102712.
780. Sanders, D. B., and Evoli, A. (2010) Immunosuppressive therapies in myasthenia gravis. *Autoimmunity* 43, 428-435.
781. Sathasivam, S. (2008) Steroids and immunosuppressant drugs in myasthenia gravis. *Nat Clin Pract Neurol* 4, 317-327.
782. What Medications Help Treat Myasthenia Gravis? <https://www.healthline.com/health/myasthenia-gravis/medication-for-myasthenia-gravis> (accessed Mar 20, 2024).
783. Bixio, R., Bertelle, D., Pistillo, F., Pedrollo, E., Carletto, A., Rossini, M., and Viapiana, O. (2022) Rheumatoid arthritis and myasthenia gravis: a case-based review of the therapeutic options. *Clin Rheumatol* 41, 1247-1254.
784. Piehl, F., Eriksson-Dufva, A., Budzianowska, A., Feresiadou, A., Hansson, W., Hietala, M. A., Håkansson, I., Johansson, R., Jons, D., Kmezic, I., et al. (2022) Efficacy and Safety of Rituximab for New-Onset Generalized Myasthenia Gravis: The RINOMAX Randomized Clinical Trial. *JAMA Neurology* 79, 1105-1112.
785. Durães, F., Pinto, M., and Sousa, E. (2018) Old Drugs as New Treatments for Neurodegenerative Diseases. *Pharmaceuticals (Basel)* 11.
786. Paleacu, D. (2007) Tetrabenazine in the treatment of Huntington's disease. *Neuropsychiatr Dis Treat* 3, 545-551.
787. Roos, R., Buruma, O., Bruyn, G., Kemp, B., and Van der Velde, E. (1982) Tiapride in the treatment of Huntington's chorea. *Acta Neurologica Scandinavica* 65, 45-50.
788. Coppen, E. M., and Roos, R. A. (2017) Current pharmacological approaches to reduce chorea in Huntington's disease. *Drugs* 77, 29-46.
789. Duff, K., Beglinger, L. J., O'Rourke, M. E., Nopoulos, P., Paulson, H. L., and Paulsen, J. S. (2008) Risperidone and the treatment of psychiatric, motor, and cognitive symptoms in Huntington's disease. *Annals of Clinical Psychiatry* 20, 1-3.

790. Alpay, M., and Koroshetz, W. J. (2006) Quetiapine in the treatment of behavioral disturbances in patients with Huntington's disease. *Psychosomatics* 47, 70-72.
791. Gambino, C. M., Ciaccio, A. M., Lo Sasso, B., Giglio, R. V., Vidali, M., Agnello, L., and Ciaccio, M. (2023) The Role of TAR DNA Binding Protein 43 (TDP-43) as a CandiDate Biomarker of Amyotrophic Lateral Sclerosis: A Systematic Review and Meta-Analysis. *Diagnostics (Basel)* 13.
792. Mackenzie, I. R., and Rademakers, R. (2008) The role of transactive response DNA-binding protein-43 in amyotrophic lateral sclerosis and frontotemporal dementia. *Curr Opin Neurol* 21, 693-700.
793. Bright, F., Chan, G., van Hummel, A., Ittner, L. M., and Ke, Y. D. (2021) TDP-43 and Inflammation: Implications for Amyotrophic Lateral Sclerosis and Frontotemporal Dementia. *International journal of molecular sciences* 22.
794. Seddighi, S., Qi, Y. A., Brown, A.-L., Wilkins, O. G., Bereda, C., Belair, C., Zhang, Y.-J., Prudencio, M., Keuss, M. J., Khandeshi, A., et al. (2024) Mis-spliced transcripts generate de novo proteins in TDP-43-related ALS/FTD. *Science Translational Medicine* 16, eadg7162.
795. Mees, I., Nisbet, R. M., Hannan, A. J., and Renoir, T. (2023) Implications of Tau Dysregulation in Huntington's Disease and Potential for New Therapeutics. *J Huntingtons Dis* 12, 1-13.
796. Salem, S., and Cicchetti, F. (2023) Untangling the Role of Tau in Huntington's Disease Pathology. *J Huntingtons Dis* 12, 15-29.
797. Abyadeh, M., Gupta, V., Paulo, J. A., Mahmoudabad, A. G., Shadfar, S., Mirshahvaladi, S., Gupta, V., Nguyen, C. T. O., Finkelstein, D. I., You, Y., et al. (2024) Amyloid-beta and tau protein beyond Alzheimer's disease. *Neural Regeneration Research* 19.
798. McGowan, D. P., van Roon-Mom, W., Holloway, H., Bates, G. P., Mangiarini, L., Cooper, G. J., Faull, R. L., and Snell, R. G. (2000) Amyloid-like inclusions in Huntington's disease. *Neuroscience* 100, 677-680.
799. Palpagama, T. H., Waldvogel, H. J., Faull, R. L. M., and Kwakowsky, A. (2019) The Role of Microglia and Astrocytes in Huntington's Disease. *Front Mol Neurosci* 12, 258.
800. Franklin, H., Clarke, B. E., and Patani, R. (2021) Astrocytes and microglia in neurodegenerative diseases: Lessons from human in vitro models. *Prog Neurobiol* 200, 101973.
801. Wilton, D. K., and Stevens, B. (2020) The contribution of glial cells to Huntington's disease pathogenesis. *Neurobiology of Disease* 143, 104963.
802. Yang, H.-M., Yang, S., Huang, S.-S., Tang, B.-S., and Guo, J.-F. (2017) Microglial Activation in the Pathogenesis of Huntington's Disease. *Frontiers in Aging Neuroscience* 9.
803. Rostalski, H., Leskelä, S., Huber, N., Katisko, K., Cajanus, A., Solje, E., Marttinen, M., Natunen, T., Remes, A. M., Hiltunen, M., et al. (2019) Astrocytes and Microglia as Potential Contributors to the Pathogenesis of C9orf72 Repeat Expansion-Associated FTL and ALS. *Frontiers in Neuroscience* 13.
804. Miller, S. J. (2018) Astrocyte Heterogeneity in the Adult Central Nervous System. *Front Cell Neurosci* 12, 401.
805. Quan, L., Uyeda, A., and Muramatsu, R. (2022) Central nervous system regeneration: the roles of glial cells in the potential molecular mechanism underlying remyelination. *Inflammation and Regeneration* 42, 7.
806. Birger, A., Ben-Dor, I., Ottolenghi, M., Turetsky, T., Gil, Y., Sweetat, S., Perez, L., Belzer, V., Casden, N., Steiner, D., et al. (2019) Human iPSC-derived astrocytes from ALS patients with mutated C9ORF72 show increased oxidative stress and neurotoxicity. *EBioMedicine* 50, 274-289.
807. Valori, C. F., Sulmona, C., Brambilla, L., and Rossi, D. (2023) Astrocytes: Dissecting Their Diverse Roles in Amyotrophic Lateral Sclerosis and Frontotemporal Dementia. *Cells* 12.
808. Brash-Arias, D., Aranda-Abreu, G. E., Rojas-Durán, F., Hernández-Aguilar, M. E., Toledo-Cárdenas, M. R., Pérez-Estudillo, C. A., Ortega, A., and Chi-Castañeda, L. D. (2023) The role of astrocytes with genetic mutations linked to amyotrophic lateral sclerosis. *Neurology Perspectives* 3, 100117.

809. Khakh, B. S., Beaumont, V., Cachepe, R., Munoz-Sanjuan, I., Goldman, S. A., and Grantyn, R. (2017) Unravelling and Exploiting Astrocyte Dysfunction in Huntington's Disease. *Trends Neurosci* 40, 422-437.
810. Hsiao, H.-Y., Chen, Y.-C., Chen, H.-M., Tu, P.-H., and Chern, Y. (2013) A critical role of astrocyte-mediated nuclear factor- $\kappa$ B-dependent inflammation in Huntington's disease. *Human Molecular Genetics* 22, 1826-1842.
811. Faideau, M., Kim, J., Cormier, K., Gilmore, R., Welch, M., Auregan, G., Dufour, N., Guillemier, M., Brouillet, E., Hantraye, P., et al. (2010) In vivo expression of polyglutamine-expanded huntingtin by mouse striatal astrocytes impairs glutamate transport: a correlation with Huntington's disease subjects. *Human Molecular Genetics* 19, 3053-3067.
812. Oksanen, M., Lehtonen, S., Jaronen, M., Goldsteins, G., Hämäläinen, R. H., and Koistinaho, J. (2019) Astrocyte alterations in neurodegenerative pathologies and their modeling in human induced pluripotent stem cell platforms. *Cell Mol Life Sci* 76, 2739-2760.
813. Purushotham, S. S., and Buskila, Y. (2023) Astrocytic modulation of neuronal signalling. *Frontiers in Network Physiology* 3.
814. Ben Haim, L., Carrillo-de Sauvage, M.-A., Ceyzériat, K., and Escartin, C. (2015) Elusive roles for reactive astrocytes in neurodegenerative diseases. *Frontiers in Cellular Neuroscience* 9.
815. Geloso, M. C., Corvino, V., Marchese, E., Serrano, A., Michetti, F., and D'Ambrosi, N. (2017) The Dual Role of Microglia in ALS: Mechanisms and Therapeutic Approaches. *Front Aging Neurosci* 9, 242.
816. Clarke, B. E., and Patani, R. (2020) The microglial component of amyotrophic lateral sclerosis. *Brain* 143, 3526-3539.
817. Gao, C., Jiang, J., Tan, Y., and Chen, S. (2023) Microglia in neurodegenerative diseases: mechanism and potential therapeutic targets. *Signal Transduction and Targeted Therapy* 8, 359.
818. Filipi, T., Hermanova, Z., Tureckova, J., Vanatko, O., and Anderova, A. M. (2020) Glial Cells-The Strategic Targets in Amyotrophic Lateral Sclerosis Treatment. *Journal of clinical medicine* 9.
819. Rothstein, J. D., and Diamond, B. *The role of NG2 glial cells in ALS pathogenesis*; Johns Hopkins University: Oct 2013, 2013.
820. Benraiss, A., Wang, S., Herrlinger, S., Li, X., Chandler-Militello, D., Mauceri, J., Burm, H. B., Toner, M., Osipovitch, M., Jim Xu, Q., et al. (2016) Human glia can both induce and rescue aspects of disease phenotype in Huntington disease. *Nature Communications* 7, 11758.
821. Feigin, A., Evans, E. E., Fisher, T. L., Leonard, J. E., Smith, E. S., Reader, A., Mishra, V., Manber, R., Walters, K. A., Kowarski, L., et al. (2022) Pepinemab antibody blockade of SEMA4D in early Huntington's disease: a randomized, placebo-controlled, phase 2 trial. *Nature Medicine* 28, 2183-2193.
822. Vaccinex Pipeline. <https://www.vaccinex.com/pipeline/> (accessed Apr 4, 2024).
823. Cytokinetics Pipeline. <https://cytokinetics.com/medicines-research/pipeline/> (accessed Apr 4, 2024).
824. Sangamo: Neurology Conference Presentations Hub. <https://www.sangamo.com/neurology-conference-presentations-hub/> (accessed Apr 4, 2024).
825. Clinical Trials. <https://www.clinicaltrials.gov/> (accessed Apr 4, 2024).
826. BERRY, J. D. Intermediate-Size Expanded Access Trial of Autologous Hybrid TREG/Th2 Cell Therapy (RAPA-501) of Amyotrophic Lateral Sclerosis. <https://reporter.nih.gov/search/mCZMWkS8p06HsrP6zbfaTg/project-details/10834469> (accessed Apr 4, 2024).
827. RAPA-501 Therapy for ALS. <https://clinicaltrials.gov/study/NCT04220190> (accessed Apr 4, 2024).
828. Safety, Tolerability, and Efficacy Study of Intrathecally Administered Gene Therapy AMT-162 in Amyotrophic Lateral Sclerosis (ALS) Patients With SOD1 Mutations. <https://clinicaltrials.gov/study/NCT06100276> (accessed Apr 4, 2024).

829. FB1006: AI-discovered drug advances to clinical trials for ALS treatment. <https://www.news-medical.net/news/20240227/FB1006-AI-discovered-drug-advances-to-clinical-trials-for-ALS-treatment.aspx> (accessed Apr 4, 2024).
830. Babu, S., Hightower, B. G., Chan, J., Zürcher, N. R., Kivisäkk, P., Tseng, C.-E. J., Sanders, D. L., Robichaud, A., Banno, H., Evora, A., et al. (2021) Ibutilast (MN-166) in amyotrophic lateral sclerosis- an open label, safety and pharmacodynamic trial. *NeuroImage: Clinical* 30, 102672.
831. Brooks, B., Bravver, E. K., Sanjak, M., Bockenek, W., Lindblom, S. S., Lary, C., Ranzinger, L., Newell-Sturdivant, A., Langford, V., Holsten, S., et al. (2018) Ibutilast - Phosphodiesterase Type 4 Inhibitor - Bi-Modal Therapy with Riluzole in Early [ Not Requiring Non-Invasive Ventilation ( NIV ) ] Cohort ( EC ) and Advanced [Requiring NIV ] ( ANC ) Amyotrophic Lateral Sclerosis ( ALS ) Patients - Single-Center Adaptive Design Six-Month Double-Blind ( DB ) - Placebo-Controlled Phase 1b/2a Epoch Followed by Six-Month Open Label Extension ( OLE ) Epoch, Washout ( WO ) and Post-Washout Epoch ( PWO ) – Final Report and Future Directions (P6.465). *Neurology* 90, P6.465.
832. MN-166 Ibutilast (Progressive Multiple Sclerosis, Amyotrophic Lateral Sclerosis, and Drug Dependence). [https://medicinova.com/clinical-development/core/mn-166/?doing\\_wp\\_cron=1712224628.1748700141906738281250](https://medicinova.com/clinical-development/core/mn-166/?doing_wp_cron=1712224628.1748700141906738281250) (accessed Apr 4, 2024).
833. FDA Grants Fast Track Designation for MediciNova's MN-166 (ibutilast) for Progressive Multiple Sclerosis. <https://investors.medicinova.com/news-releases/news-release-details/fda-grants-fast-track-designation-medicinovas-mn-166-ibutilast-1> (accessed Apr 4, 2024).
834. Ibutilast. <https://www.accessdata.fda.gov/scripts/opdlisting/oopd/detailedIndex.cfm?cfgridkey=445814> (accessed Apr 4, 2024).
835. Prilenia Announces Topline Results for Pridopidine in Phase 2 ALS Study. [https://s202.q4cdn.com/210612175/files/doc\\_news/Prilenia-Announces-Topline-Results-for-Pridopidine-in-Phase-2-ALS-Study-2023.pdf](https://s202.q4cdn.com/210612175/files/doc_news/Prilenia-Announces-Topline-Results-for-Pridopidine-in-Phase-2-ALS-Study-2023.pdf) (accessed Apr 4, 2024).
836. About ALS. <https://www.prilenia.com/what-is-als/> (accessed Apr 4, 2024).
837. Wave Life Sciences Announces Positive Update from Phase 1b/2a SELECT-HD Trial with Initial Results Indicating Allele-Selective Target Engagement with WVE-003 in Huntington's Disease. <https://www.globenewswire.com/news-release/2022/09/20/2519092/0/en/Wave-Life-Sciences-Announces-Positive-Update-from-Phase-1b-2a-SELECT-HD-Trial-with-Initial-Results-Indicating-Allele-Selective-Target-Engagement-with-WVE-003-in-Huntington-s-Diseas.html> (accessed Apr 16, 2024).
838. Efficacy and Safety of NestaCell® in Huntington's Disease (STAR). <https://classic.clinicaltrials.gov/ct2/show/NCT06097780> (accessed Apr 4, 2024).
839. Hill, M. D., Blanco, M.-J., Salituro, F. G., Bai, Z., Beckley, J. T., Ackley, M. A., Dai, J., Doherty, J. J., Harrison, B. L., Hoffmann, E. C., et al. (2022) SAGE-718: A First-in-Class N-Methyl-d-Aspartate Receptor Positive Allosteric Modulator for the Potential Treatment of Cognitive Impairment. *Journal of Medicinal Chemistry* 65, 9063-9075.
840. SAGE THERAPEUTICS PROVIDES IMPORTANT UPDATE ON THE CLINICAL DEVELOPMENT PROGRAM FOR SAGE'S INVESTIGATIONAL DRUG, SAGE-718. <https://hdsa.org/news/sage-therapeutics-provides-important-update-on-the-clinical-development-program-for-sages-investigational-drug-sage-718/> (accessed Apr 4, 2024).
841. Sage Therapeutics Announces U.S. Food and Drug Administration Granted SAGE-718 Orphan Drug Designation for the Treatment of Huntington's Disease. <https://investor.sagerx.com/news-releases/news-release-details/sage-therapeutics-announces-us-food-and-drug-administration> (accessed Apr 4, 2024).
842. CNP-106 Myasthenia Gravis. <https://courpharma.com/pipeline-2/#cnp-106-myasthenia-gravis> (accessed Apr 4, 2024).

843. Chahin, N., Sahagian, G., Feinberg, M. H., Stewart, C. A., Jewell, C. M., Kurtoglu, M., Miljković, M. D., Vu, T., Mozaffar, T., and James F. Howard, J. (2024) Twelve-Month Follow-Up of Patients With Generalized Myasthenia Gravis Receiving BCMA-Directed mRNA Cell Therapy. medRxiv, 2024.2001.2003.24300770.
844. CARTESIAN THERAPEUTICS HIGHLIGHTS PROGRESS AND 2024 STRATEGIC PRIORITIES ACROSS INNOVATIVE PIPELINE OF MRNA CELL THERAPIES FOR AUTOIMMUNITY. <https://www.cartesiantherapeutics.com/2024/01/08/cartesian-therapeutics-highlights-progress-and-2024-strategic-priorities-across-innovative-pipeline-of-mrna-cell-therapies-for-autoimmunity/> (accessed Apr 4, 2024).
845. RemeGen Releases Phase II Clinical Study Data for Treatment of Myasthenia Gravis in Chinese Patients. <https://www.prnewswire.com/news-releases/remegen-releases-phase-ii-clinical-study-data-for-treatment-of-myasthenia-gravis-in-chinese-patients-301664085.html> (accessed Apr 4, 2024).
846. Amylyx Pharmaceuticals Announces Formal Intention to Remove RELYVRIO®/ALBRIOZA™ from the Market; Provides Updates on Access to Therapy, Pipeline, Corporate Restructuring, and Strategy. <https://www.amylyx.com/news/amylyx-pharmaceuticals-announces-formal-intention-to-remove-relyvrrior/albriozatm-from-the-market-provides-updates-on-access-to-therapy-pipeline-corporate-restructuring-and-strategy> (accessed Apr 4, 2024).
847. Rabbani, B., Tekin, M., and Mahdieh, N. (2014) The promise of whole-exome sequencing in medical genetics. *Journal of Human Genetics* 59, 5-15.
848. Warr, A., Robert, C., Hume, D., Archibald, A., Deeb, N., and Watson, M. (2015) Exome Sequencing: Current and Future Perspectives. *G3 (Bethesda)* 5, 1543-1550.
849. Whole Exome Sequencing and Analysis. [https://nisc.nih.gov/docs/FAQ\\_whole\\_exome.pdf](https://nisc.nih.gov/docs/FAQ_whole_exome.pdf) (accessed Mar 27, 2024).
850. Ng, P. C., and Kirkness, E. F. (2010) Whole genome sequencing. *Methods in molecular biology (Clifton, N.J.)* 628, 215-226.
851. Yin, R., Kwoh, C. K., and Zheng, J., Whole Genome Sequencing Analysis. In *Encyclopedia of Bioinformatics and Computational Biology*, Ranganathan, S.; Gribskov, M.; Nakai, K.; Schönbach, C., Eds. Academic Press: Oxford, 2019; pp 176-183.
852. Global Alliance for Genomics and Health (GA4GH). <https://www.ga4gh.org/> (accessed Mar 27, 2024).
853. Undiagnosed Diseases Network (UDN). <https://undiagnosed.hms.harvard.edu/> (accessed Mar 27, 2024).

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