1 Title: Interpretable deep-learning pKa prediction for small molecule drugs via

- 2 atomic sensitivity analysis
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## 22 ABSTRACT

Machine learning (ML) models play a crucial role in predicting properties essential to drug 23 24 development, such as a drug's logscale acid-dissociation constant (pKa). Despite recent architectural advances, these models often generalize poorly to novel compounds due to a 25 26 scarcity of ground-truth data. Further, these models lack interpretability, in part due to a dependence on explicit encodings of input molecules' molecular substructures. To this end, 27 atomic-resolution information is accessible in chemical structures by observing model response 28 29 to atomic perturbations of an input molecule; however, no methods exist that systematically utilize this information for model and molecular analysis. Here, we present BCL-XpKa, a 30 substructure-independent, deep neural network (DNN)-based pK<sub>a</sub> predictor that generalizes well 31 to novel small molecules. BCL-XpKa discretizes pK<sub>a</sub> prediction from a regression problem into 32 a multitask-classification problem, which accumulates data for prediction at biologically relevant 33 pH values and records the model's uncertainty in its prediction as a discrete distribution for each 34

pK<sub>a</sub> prediction. BCL-XpKa outperforms modern ML pK<sub>a</sub> predictors and accurately models the 35 36 effects of common molecular modifications on a molecule's ionizability. We then leverage BCL-37 XpKa's substructure independence to introduce atomic sensitivity analysis (ASA), which quickly decomposes a molecule's predicted pK<sub>a</sub> value into its respective atomic contributions without 38 model retraining. When paired with BCL-XpKa, ASA informs that BCL-XpKa has implicitly 39 40 learned high-resolution information about molecular substructures. We further demonstrate 41 ASA's utility in structure preparation for protein-ligand docking by identifying ionization sites in 97.8% and 83.4% of complex small molecule acids and bases. We then apply ASA with BCL-42 XpKa to understand the physicochemical liabilities and guide optimization of a recently 43 published KRAS-degrading PROTAC. 44

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#### 46 INTRODUCTION

Predicting a drug's behavior in the body is a key challenge in computational drug development. 47 For example, accurate prediction of compounds' bioavailability could support early modification 48 or termination of nonviable lead molecules, thereby saving years of time and millions of dollars 49 on research and development. The demand for fast and accurate predictions of a drug's 50 51 quantitative structure-activity and structure-property relationships (QSAR, QSPR) has 52 skyrocketed as our access to synthesizable chemical space approaches one trillion molecules<sup>1</sup>. 53 While advances in machine learning have improved prediction accuracy, the small amount of publicly available, high-quality experimental data for training often leads to overfitting and 54 prevents generalizability<sup>2-4</sup>. Further, QSPR model interpretability is often poor despite the 55 relatively more intuitive input (chemical structures) than in other fields of computational biology 56 (e.g., transcriptomics data). As such, additional explorations into architectures that can efficiently 57 train on chemical data, as well as general methods to interpret these models' outputs, are 58 59 warranted.

60 One of the most critical properties to a drug's downstream efficacy is its ionizability at physiologic pH values, which depends on the drug's logscale acid-dissociation constant (pKa 61 value)<sup>4,5</sup>. Quantum mechanical ( $\overline{QM}$ ) methods now calculate pK<sub>a</sub> with experimental accuracy 62 and are extremely valuable to late-stage drug development. However, small-molecule drug 63 development often begins with virtual high-throughput screening (vHTS) of billions of 64 compounds, and QM methods are too computationally expensive to assist meaningfully in vHTS. 65 As such, scientists have made tremendous investment in ML-based QSAR/QSPR predictors for 66 67 faster – though potentially less accurate – prediction of physicochemical properties like pKa.

These ML methods generally embed molecules using molecular fingerprints, which are twodimensional (2D)- or 3D chemical substructures centered around each atom in the molecule. Recently, groups have realized significant gains in prediction accuracy with graph neural networks (GNNs), which embed molecules as a graph in addition to standard chemical descriptors<sup>6-8</sup>. Improvements in molecular featurization strategies and network architecture have driven state-of-the-art pK<sub>a</sub> prediction accuracy to within 0.75-1.00 pK<sub>a</sub> units of experimental values.

Despite these advances, several limitations persist in ML-based pKa prediction and QSPR 75 prediction generally. First, all ML-based pK<sub>a</sub> predictors to date use regression. While regression 76 77 is the natural setting for predicting continuous values, small training set sizes restrict the 78 accuracy of regression outputs, particularly at extreme – but still physically relevant – values. Second, many of these models directly encode common molecular substructures in their feature 79 set<sup>9</sup>. This strategy may limit generalizability by preventing complete consideration of each 80 atom's local context in a molecule. For example, amide Nitrogen atoms are generally not 81 ionizable at physiologic pH values, but their acidity can be greatly increased by appending 82 neighboring electron-withdrawing groups (e.g., diacetamide). Therefore, encoding atom-specific 83 local environments may increase generalizability. 84

85 Finally, model explainability and interpretability in computational chemistry largely focus on feature-set-level analysis, encompassing step-wise<sup>10,11</sup>, feature-masking<sup>12</sup>, feature-set-86 perturbation<sup>13</sup>, feature-attribution<sup>14</sup>, and response-randomization<sup>15</sup> methods. However, feature-87 set-level analysis is often slow and unintuitive, as feature sets are often large and complex. 88 Recently, ML frameworks have been developed that utilize direct chemical representations (as 89 SMILES strings) to identify important substructures for transformer predictions<sup>16</sup>, but no group 90 to date has leveraged perturbations to the input chemical structure itself to gain insights into 91 model learning and output. Indeed, chemical structures are unique in computational biology in 92 that they can be perturbed in consistent, physical meaningful ways. With an appropriately 93 constructed feature set, measuring a model's response to these perturbations would provide 94 granular details into both model learning and molecular hotspots for prediction in real time, 95 without the need for model retraining. For example, replacing a molecule's acidic carboxylic 96 acid functional group with an inert ketone increases the  $pK_a$  from ~4 to ~20, thereby 97 demonstrating the carboxylic acid's importance to acidity. As a counterexample, increasing or 98 99 decreasing the expression of a gene in a transcriptomics-based predictor of cellular activity may not be physically meaningful, as gene expression is highly dependent on the network of 100 expressed genes in a cell/tissue. This presents an underutilized opportunity for computational 101 chemists to gain valuable insights into model learning, performance, and molecular structure. 102

To address these limitations, we present BCL-XpKa, a substructure-independent multitask 103 classifier for rapid and accurate pKa prediction built in the Biology and Chemistry Library 104 (BCL), an open-source cheminformatics platform developed and maintained by our lab. We use 105 pK<sub>a</sub> prediction to illustrate that discretizing continuous problems in chemical biology into 106 multitask classification problems can increase prediction accuracy without meaningful 107 information loss, thereby circumventing many of the problems associated with regression 108 models. We couple BCL-pKa with a novel method of atomic sensitivity analysis (ASA), which 109 provides unprecedented, atomic-level insights into which regions of a molecule are most 110 important for the model's final prediction. We demonstrate ASA's utility in probing model 111 learning, as well as its direct applicability to introducing targeted modifications in molecules that 112 reduce ionizability. Importantly, ASA is relevant to all forms of QSAR/QSPR prediction and can 113 be easily implemented to existing algorithms. 114

BCL-XpKa is a multi-layer perceptron (MLP) that embeds molecules using 2D chemical 115 descriptors features that only encode information about each atom's local environment (up to 1 116 bond away) in the molecule<sup>17</sup>. This scheme is substructure independent and enables increased 117 sensitivity to atom-level perturbations, which is particularly important for hit-to-lead and lead 118 119 optimization in late-stage drug development. Small molecule drugs often have both basic and 120 acidic regions that vary greatly in pK<sub>a</sub> values (e.g., amino acids have both an acidic carboxylic acid and a basic free amine group). To account for this, we trained two models: one to predict a 121 molecule's most acidic pKa value (BCL-XpKaAcid), and one to predict its most basic pKa value 122 (BCL-XpKaBase). This is a common practice in modern pK<sub>a</sub> prediction. We trained BCL-XpKa 123 on datasets of both predicted and experimental pKa values, and we evaluate our models on an 124 external test set of challenging acids and bases with experimental pK<sub>a</sub> values. We find that our 125 model has competitive accuracy and reduced substructure dependence than state-of-the-art pKa 126 127 predictors, including GNN-based models.

128 Overall, the work presented here has the following significant contributions to the field:

- We developed a novel, substructure-independent framework for QSPR prediction that uses local atomic environment embeddings and replaces regression with multitask classification, using pK<sub>a</sub> prediction to illustrate competitive performance with modern ML models.
- We developed a method that rapidly assesses QSPR model learning and provides atomic level insights to molecular ionizability without requiring model retraining
- We integrate these two tools in a workflow for lead optimization and apply it to optimize
   a pan-KRAS degrading Proteolysis Targeting Chimera (PROTAC).
- 137
- 138 **RESULTS**

## 139 Multitask Classification is competitive with Regression for small-molecule pK<sub>a</sub> prediction

Regression models naturally dominate machine-learning (ML) approaches to QSPR prediction. 140 However, regression models require large, high-quality datasets to train properly, and 141 understanding the model's uncertainty in its prediction is challenging. Here, we construct BCL-142 XpKa, a multilayer perceptron (MLP)-based pK<sub>a</sub> prediction tool that transforms a continuous 143 prediction problem into a multitask classification problem. Rather than predict pK<sub>a</sub> values on a 144 continuous range, BCL-XpKa predicts the probability that a molecule's pKa lies within a certain 145 range (Figure 1A-B). The expected value of this probability distribution corresponds to BCL-146 XpKa's predicted pK<sub>a</sub> for a molecule, and the variance of this distribution directly informs its 147 confidence in that prediction. 148

- 149 To train BCL-XpKa, pK<sub>a</sub> values in the training set were converted into vectors in  $\mathbb{Z}_2^n$ , where n is
- 150 the number of bins the continuous interval has been divided into, and a 1 at position i indicates
- 151 the molecule's  $pK_a$  lies in bin *i* (Figure 1B). BCL-XpKa models were trained to predict the most
- acidic and most basic pK<sub>a</sub> values of a molecule using training data from ChEMBL augmented
- 153 with negative data (i.e., nonionizable molecules).

Binning training data in this way necessarily leads to some information loss. Using an external 154 155 test set, we demonstrate that mean absolute error (MAE) increases as the number of bins 156 increases, as increasing the number of bins reduces the amount of data in each bin for training (Figure 1C). A bin size of 1 pK<sub>a</sub> unit yielded the lowest MAE on this test set and is used in the 157 BCL-XpKa production model. For both acids and bases, BCL-XpKa marginally outperforms the 158 159 best-performing regression models trained on the same data (0.79 vs 0.83 for acids, 0.86 vs 0.92 for bases, Figure 1D). Additional model details, including an evaluation of model 160 hyperparameters, can be found in Supplemental Figure 1. 161



Figure 1 BCL-XpKa model description and internal performance (A-B) Overview of the
 regression and MTC architectures for pK<sub>a</sub> prediction. BCL-XpKa utilizes the MTC architecture
 with bin size of 1 pK<sub>a</sub> unit. (C) MTC model performance by pK<sub>a</sub> bin size on acids (red) and

bases (blue). (D) Model performance comparison between best performing MTC (BCL-XpKa)and regression architectures on acids (red) and bases (blue).

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## 169 BCL-XpKa accurately captures complex trends in ionizability for druglike small molecules

We then compared BCL-XpKa's performance head-to-head against several state-of-the-art pK<sub>a</sub> predictors of varying architecture using a challenging test set external to BCL-XpKa's training data<sup>7,18-21</sup>. Despite its relatively simple architecture, BCL-XpKa achieves competitive performance to the best machine-learning (MolGpKa, graph-convolutional neural network) and rule-based pK<sub>a</sub> predictors on both acids and bases, with BCL-XpKaAcid and BCL-XpKaBase achieving mean absolute error (MAE) of 0.79 and 0.86, respectively (Figure 2A).

Beyond accurate pK<sub>a</sub> prediction, correctly predicting the effect of small perturbations to a lead 176 molecule's ionizability is of key importance in drug development<sup>22</sup>. To assess BCL-XpKa's 177 sensitivity to such changes, we identified 71 pairs of molecules in our test set that vary by a 178 slight modification, such as the replacement of an amide with an ester. BCL-XpKa correctly 179 predicts the direction of pKa change in 81.7% of these pairs (Figure 2B). To illustrate this effect, 180 BCL-XpKa correctly predicts the inductive effect of electron-withdrawing groups on acidity 181 using a series of phenol derivatives (Figure 2C). Here, fluorination at the ortho position increases 182 acidity more than fluorination at the para position (8.74 vs 9.28), and substitution with multiple 183 fluorine atoms has a greater effect than monosubstitution (7.36). While phenol was in our 184 training data, the remaining molecules were not. For bases, BCL-XpKa correctly predicts the 185 complex impact of aromaticity on nitrogen basicity in a series of piperidine derivatives relevant 186 to drug development. Introducing a neighboring phenyl group reduces predicted pK<sub>a</sub> from 10.45 187 188 (true  $pK_a$  11.2) to 4.80 (5.00). Similarly, aromatization of piperidine to pyridine decreases predicted pK<sub>a</sub> to 5.45 (5.20), and appending the same phenyl group to produce quinoline reduces 189 pK<sub>a</sub> to 4.54 (4.92) (Figure 2D). 190

191 Finally, substructure independence is critical to QSPR model generalizability to novel compounds. As described above, BCL-XpKa embeds molecules solely using the 1-bond-length 192 193 neighborhoods of each atom to limit substructure dependence. To investigate this strategy's impact, we subset BCL-XpKa's training set according to 30 ionizable functional groups. We 194 iteratively retrained BCL-XpKa leaving each substructural class out, then tested each model on 195 its withheld substructural class (Figure 2E). BCL-XpKa demonstrates robust performance on this 196 leave-class-out (LCO) test, with an average MAE of 1.1 pK<sub>a</sub> units across all LCO models. 197 Training on MACCS descriptors rather than Mol2D yielded systematically worse results and an 198 199 average MAE of 1.46 pK<sub>a</sub> units (Figure 2F).



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202 Figure 2: BCL-XpKa external performance and molecular series (A) Performance of various 203  $pK_a$  predictors on an external test set of acids (red) and bases (blue). (B) BCL-XpKa prediction vs experimental pK<sub>a</sub> value for families of related druglike molecules. Green denotes correct 204 change in predicted pK<sub>a</sub> due to chemical modification, and red denotes incorrect change in 205 predicted pK<sub>a</sub>. (C-D) Example molecular families from (B). Predicted and experimental pK<sub>a</sub> 206 values provided. (E) Schematic for LCO testing. Testi and Traini denote the subsets of the 207 training set that contain or do not contain, respectively, substructure i. Modeli was trained with 208 Traini and evaluated on Testi to give MAELCO-i. (F) LCO performance of BCL-XpKa (blue) vs an 209 equivalent model trained with a MACCS-based descriptor set (orange). 210

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# Atomic sensitivity analysis provides actionable, atomic-resolution information on model predictions

Computational chemistry currently lacks rapid, reliable tools for interpreting ML model 214 predictions<sup>23</sup>. Such atom-, substructure-, or pharmacophore-level information could accelerate 215 computer-aided drug development on multiple fronts, from assisting in model training, to 216 217 preparing and filtering molecules in virtual high-throughput screens, to guiding lead compound modification in lead optimization. To address this deficit, we developed atomic sensitivity 218 analysis (ASA, Figure 3A), and we demonstrate its utility in decomposing BCL-XpKa's 219 predictions to assess model learning, identify ionization sites in complex small molecules, and 220 guide lead optimization efforts by reducing molecular ionizability. 221

ASA compares an ML model's prediction on a parent molecule before and after some perturbation. Here, we sequentially replace heteroatoms in the parent molecule with correctly hybridized carbons, then generate probability distributions for each with BCL-XpKa. The final ASA score is a scaled version of the KL-Divergence between these parent and perturbed pKa distributions (see *Methods*). Importantly, we anticipate that, with careful feature-set selection, this replace-rescore-compare scheme will generalize well to substructure- and pharmacophore-level perturbations (Figure 3A).

We benchmarked ASA on BCL-XpKa on all test-set molecules with nontrivial ionization sites. 229 We first hypothesized that perturbing an acid's most acidic hydroxyl group or a base's most 230 basic Nitrogen atom would have the most significant impact on the predicted pKa, and therefore 231 the largest ASA scores. We tested this hypothesis by performing ASA on the Oxygen atoms in 232 233 the acid set and the Nitrogen atoms in the base set and considering only the atom with the 234 maximum ASA score in each molecule. This strategy correctly identifies 97.8% of the most 235 acidic Oxygen atoms in the acid set and 83.4% of the most basic Nitrogen atoms in the base set, 236 thereby demonstrating ASA's potential utility in high-throughput structure preparation (Figure 237 3B).

238 This benchmark revealed surprisingly consistent ASA scores for each atom in the substructures 239 that recurred throughout the test set. For example, free amines are the most basic group in 33.9% 240 of our experimentally characterized bases, and in each of these molecules the amine Nitrogen atom dominates the molecule's ASA scores (33.1 +/- 16.0). These scores were significantly 241 242 higher than average scores for Nitrogen atoms in amide (0.225 +/- 0.332), indole (0.261 +/-0.444), and nitrile groups (0.180 +/- 0.357) (p < 0.001), functional groups which are not 243 traditionally ionizable at physiologic pH and which dominated 0% of the test-set ASA scores 244 (Figure 3C). Further, molecules where the dominant ionizable group is less basic than typical 245 246 amines also demonstrated consistent ASA scores, and this ASA dominance was reliably ablated by the introduction of an amine functional group (Figure 3D-E). 247



Figure 3: Atomic sensitivity for molecular analysis (A) Schematic of the ASA protocol. Parent and Perturbed distributions refer to the localPPV distributions output by BCL-XpKa. (B) ASA accuracy at detecting the most acidic Oxygen atom and most basic Nitrogen atom in all nontrivial test-set molecules. (C) ASA scores of positive- and negative-control substructures for BCL-XpKaBase decomposition. Blue denotes the positive control, red denotes the negative controls. (D-E) Modulation of pyridine Nitrogen ASA score by addition of an amine group.

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# ASA reveals BCL-XpKa implicitly learns substructural information despite substructure free embeddings

To investigate ASA scoring consistency further, we performed ASA on Nitrogen atoms in the most frequently occurring substructures in the basic test set, beyond the controls discussed above. All subgroups examined demonstrated statistically significant loss of ASA signal when a more dominant subgroup (i.e., more relevant to the molecule's basicity) was present (Figure 4A-D), indicating that retaining the most dominant ASA atom also preserves BCL-XpKa's predicted

- $pK_a$  distribution for the molecule.
- 264 Filtering out these non-dominant molecules reveals that most substructures have consistent, substructure-specific trends in ASA scores (Figure 4E). This substructure-specificity even 265 persists when separate substructures have Nitrogen atoms with identical local atomic 266 environments. For example, indole and imidazole both have a Nitrogen bound to a Hydrogen and 267 two sp<sup>2</sup> Carbon atoms, but only imidazole is ionizable at physiologic pH values<sup>24,25</sup>. While ASA 268 correctly distinguishes that imidazole's other Nitrogen (which has a lone pair of electrons) is the 269 most basic Nitrogen in imidazole, it also scores the N-H Nitrogen significantly higher than the 270 271 identical motif in indole (Figure 3C), suggesting that this Nitrogen is critical to imidazole's 272 observed basicity.
- 273 Some of these structures have surprisingly high variance in ASA scores, particularly the pyridine 274 and aniline substructures. From manual inspection, we hypothesized a portion of this variance is attributable to the impact of neighboring electron-donating and -withdrawing groups (EDGs, 275 276 EWGs), which respectively increase and decrease basicity of neighboring Nitrogen atoms. We tested this hypothesis by scoring manually created sets of pyridine derivatives with various EDG 277 and EWG substituents, which confirmed as suspected that neighboring EDGs tend to increase 278 ASA scores, and neighboring EWGs tend to decrease ASA scores (Figure 4F). Interestingly, 279 symmetric substructures also contributed to this variance, as the symmetric substructure masks 280 the effect of the removed atom during ASA scoring (Figure 4G). 281
- Together, these ASA findings suggest that BCL-XpKa has learned impressive substructural insights that are adaptable to molecular context without directly encoding these substructures in the feature set.



Figure 4: Atomic sensitivity analysis of substructures (A-D) Violin plots of ASA scores for 286 commonly occurring substructures when these substructures are the dominant site of a 287 molecule's ionization vs when a more dominant substructure was present. (E) Violin plots of 288 ASA scores for commonly occurring substructures when these substructures were the dominant 289 site of ionization. Notably, all test-set bases containing 2-Aminopyridine and Imidazole featured 290 them as their dominant ionization site. (F) Change in pyridine Nitrogen's ASA Score by 291 neighboring EWG or EDG groups. (F) Masking effect of molecular symmetry on ASA score. 292 293 ASA = Atomic Sensitivity Analysis; EWG = electron-withdrawing group; EDG = electrondonating group. 294

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### 296 Atomic sensitivity analysis can inform lead compound optimization in drug development

Atomic sensitivity analysis also has promising utility in prospective drug design. Significant 297 interest has developed in the past two decades in targeted protein degradation via small-molecule 298 Proteolysis Targeting Chimeras (PROTACs). PROTACs consist of two small molecules joined 299 by a linker and form flexible ternary complexes with the target and an E3 Ligase, which allows 300 for target ubiquitination and subsequent degradation by the 26S proteasome (Figure 5A, PDB: 301 8QU8). PROTACs degrade targets catalytically, making them an attractive strategy for 302 challenging targets that have evaded small-molecule inhibition; however, PROTAC size and 303 complexity plagues design efforts with poor bioavailability and cell permeability<sup>26</sup>. These 304 properties are generally optimized through modification of the PROTAC linker, as the Ligase-305 and target-binding domains make specific contacts with their respective proteins. Here, we 306 demonstrate how atomic sensitivities can guide rational changes to the PROTAC linker to 307 308 minimize PROTAC ionizability.

KRAS is a challenging target in oncology because it lacks a deep, well-defined pocket for small-309 molecule inhibition<sup>27</sup>. Recently, Popow et al. created a KRAS-degrading PROTAC using the 310 VHL E3 Ligase (Figure 5A-B)<sup>28</sup>. Our model predicts this PROTAC has a  $pK_a$  of 6.51 (P-1, 311 Figure 4B), suggesting protonatability at physiologic pH values. Atomic sensitivity analysis of 312 the linker reveals one of two tertiary amines in P-1 drives this prediction (Figure 5C). The crystal 313 314 structure of the P-1 ternary complex demonstrates this amine forms a salt bridge with KRAS Q62 when protonated. While salt-bridge interactions promote strong drug binding, PROTACs 315 only need to bind their targets transiently, and a protonatable amine is a liability for permeability. 316 Based on this analysis, we evaluated several P-1 bioisosteric modifications at this amine that 317 reduce predicted  $pK_a$  (Figure 5D). 318

We evaluated each modification's performance computationally using a well-benchmarked ternary complex (TC) generation algorithm, which produces a TC ensemble and "double clusters" this ensemble by structural similarity of both the Ligase-Target interactions and PROTAC conformation<sup>29</sup>. Per the algorithm authors' benchmark, the largest structural double cluster often yields the most crystal-structure-like poses and filters out false positives from the full ensemble. Each modification produced comparable TC ensembles and double clusters to P-1 (Figure 5E).

Emerging experimental evidence suggests that a PROTAC's degradation efficiency depends on 326 its ability to place the target protein in proximity to ubiquitin (<60Å) in the complete E3 Ligase 327 complex<sup>30</sup>. As such, we also evaluated the modifications' ability to place KRAS in proximity to 328 ubiquitin by superposing each TC ensemble member into the full E3 Ligase complex. Here, we 329 find that the amide modification in P-2 provides KRAS similar ubiquitin access to P-1 and 330 superior access to all other modifications tested (Figure 5F). Furthermore, docking P-2 into the 331 VHL-KRAS pose identified in the 8QU8 crystal structure demonstrates that P-2 can recapitulate 332 the binding pose of P-1 (heavy atom RMSD <2.5Å), including the key hydrogen bond with 333 KRAS Q62 that P-1 utilizes using the tertiary amine (Figure 5G). 334

Together, these results demonstrate a computationally validated use of atomic sensitivities to guide lead-molecule optimization.



Figure 5: Atomic sensitivity for drug design (A) Crystal structure (PDB: 8QU8) of pan-KRAS 339 degrading PROTAC P-1 in ternary complex with VHL and KRAS. (B) PROTAC P-1 colored 340 according to 5A, with pK<sub>a</sub> calculated by BCL-XpKa. (C) ASA scores for P-1 Nitrogen atoms. 341 VHL- and KRAS-binders omitted for space. (D) Proposed bioisosteric P-1 linker modifications 342 with pK<sub>a</sub> values predicted by BCL-XpKa. (E) Ternary-complex ensemble size and size of largest 343 Protein-PROTAC conformational double cluster for each linker modification in 5D. (F) 344 Cumulative number of ternary complexes near Ubiquitin at increasing distances from Ubiquitin 345 in the closed conformation of the CUL2 E3 Ligase complex. Red line at 60Å denotes an 346 empirically estimated distance beyond which target ubiquitination is improbable. (G) 347 Representative images of the 8QU8 crystal structure and the Amide-based linker modification P-348 2 supporting similar PROTAC conformations that preserve the hydrogen bond to KRAS Q62. P-349 1 complex shown in brighter colors; P-2 complex shown in muted colors; P-2 linker modification 350 351 highlighted in yellow.

### 352 **DISCUSSION**

Here, we have presented BCL-XpKa, a deep-learning based pKa predictor that reframes QSPR 353 prediction as a classification problem and avoids explicit substructural embeddings while 354 355 maintaining competitiveness with contemporary machine learning pK<sub>a</sub> predictors. We found that this multitask classification approach directly informs the model's uncertainty in its prediction, 356 and that, beyond its absolute accuracy, BCL-XpKa reliably predicts the effects of common 357 molecular modifications made to a hit/lead compound in a drug development program. We also 358 359 showed that BCL-XpKa generalizes to foreign substructures better than equivalent models 360 trained on MACCS-based descriptors via leave-substructural-class-out validation.

We then used BCL-XpKa as a model system to introduce atomic sensitivity analysis (ASA), a 361 first-in-class ML interpretability method we designed to provide actionable insights into QSPR 362 model output by decomposing a molecule's QSPR prediction into its atomic contributions 363 364 through direct perturbation of the input chemical structure. When applied to BCL-XpKa, ASA 365 identifies the most ionizable atoms in both acids and bases with remarkable accuracy. ASA also 366 revealed surprisingly consistent results for how BCL-XpKa considers ionizable substructures at 367 the atomic level. These substructural ASA scores were responsive to neighboring electron 368 donating and withdrawing groups, demonstrating that BCL-XpKa learns context-dependent substructural information without explicit substructural embeddings. Finally, we showed that 369 370 pairing a QSPR model's molecule-level predictions with atomic-level contributions is a powerful tool for guiding lead optimization using a published KRAS-degrading PROTAC. Here, BCL-371 XpKa and ASA directed linker modifications that reduced PROTAC ionizability while retaining 372 critical PROTAC-KRAS contacts from the original crystal structure. 373

Several limitations exist in our current framework. First, while regression models can predict arbitrarily extreme values given enough quality data, BCL-XpKa must place all extreme values in two catch-all bins, " $pK_a < 0$ " and " $pK_a > 12$ ", given its multitask classifier architecture. While this limits BCL-XpKa's theoretical output range to -0.5 to 12.5, this is not consequential for biologically relevant pH scales and only marginally affects prediction accuracy.

Further, ASA is currently limited to atomic-level model explainability. While this provides excellent resolution for atomic properties like pK<sub>a</sub>, there are many QSPR tasks where understanding the contribution of entire substructures or pharmacophores would be valuable. Generalizing ASA to higher order molecular substructures will further expand our understanding of QSPR ML model predictions and allow ASA to be tailored to specific tasks.

BCL-XpKa and ASA have fundamental applications in computational chemistry generally, as well as early- and late-stage drug development. First, ASA is a generalizable strategy that can increase the explainability of any machine learning model that uses chemical structures as input data. As shown here, ASA scores can help scientists understand what their model has learned from their training set. This information can then guide training-set data augmentation or featureset modifications.

Further, BCL-XpKa paired with ASA is positioned well to support high-quality small-molecule
 structure preparation for virtual high-throughput screening (vHTS). vHTS involves screening

ultra-large libraries (ULLs) of small molecules (currently nearing  $10^{11}$  molecules) for their ability to bind to a protein target. vHTS has notoriously low hit rates, and improper protonation of ULL molecules can contribute to both false-positive and false-negative vHTS screens. BCL-XpKa and ASA's speed and accuracy at predicting pK<sub>a</sub> and ionization sites in multiprotic species make this tool a valuable asset for ULL structure preparation and downstream protein-ligand analysis in vHTS.

Finally, as demonstrated here, BCL-XpKa paired with ASA can identify ionizable regions in a compound for modification in hit-to-lead or lead optimization. While ionization-site identification is relatively straightforward, this model-ASA strategy generalizes to any QSPR/QSAR model. Therefore, applying ASA to predictors of ADMET/DMPK may facilitate understanding of important but less readily interpretable liabilities in a hit or lead compound.

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### 404 METHODS

### 405 *Training Datasets*

406 ChEMBL27 is an open-source database contains over 2 million molecules with various 407 physicochemical descriptors<sup>31</sup>. ACDlabs was used to calculate acidic  $pK_a$  and basic  $pK_a$  values 408 (chembl\_acid\_pka and chembl\_base\_pka, respectively) for ChEMBL molecules, and molecules 409 that were included in our test sets were excluded<sup>18</sup>. We also generated negative data (molecules 410 with no ionization site) in the BCL and set chembl\_acid\_pka = 50, chembl\_base\_pka = 0. In 411 sum, acidic  $pK_a$  models were trained on 988,643 molecules, and basic  $pK_a$  models on 812,918 412 molecules.

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## 414 *Molecule Preparation*

Molecular 3D structures were standardized using Corina for training and testing BCL-XpKa<sup>32</sup>. For external models, structure preparation followed the authors' direction, and Corina was used if no structure preparation method was mentioned. This standardization was used exclusively for downstream usability, as BCL-XpKa solely uses 2D descriptors. All PROTAC modifications introduced in Figure 5 were minimized in the Molecular Operating Environment (MOE) prior to ternary complex ensemble generation.

- 421
- 422 *Molecular Features*

The Mol2D molecular descriptor set was used to encode molecules as described elsewhere<sup>17</sup>. Briefly, for each atom in a molecule, Mol2D encodes information about that atom, the bonds made to that atom, and the atoms one bond length away from that atom.

426 To train a multitask classifier with N output labels, ChEMBL pK<sub>a</sub> values were encoded in an 427  $N \ge 1$  result label  $R \in \mathbb{Z}_2^N$ , where the first entry corresponds to pK<sub>a</sub> < 0, the last entry to pK<sub>a</sub> ≥ 428 12, and the  $i^{\text{th}}$  entry to  $(12/N) * (i - 1) \le pK_a < (12/N) * i$ . For regression models, the 429 pK<sub>a</sub> in the ChEMBL set was used directly as the result label.

430

#### 431 Model Training and Validation

Artificial neural networks were built in C++ for the Biology and Chemistry Library (BCL), an 432 open-source cheminformatics platform created and maintained by our lab. Each model was 433 trained for 250 iterations without early stopping, which our lab previously found to be 434 unnecessary when dropout is used<sup>33</sup>. An upper-bound for model performance was calculated 435 through random-split cross validation. A lower-bound for performance was calculated through 436 leave-class-out cross validation (LCO-CV), in which the training set was divided into 30 subset 437  $\{C_i\}_{i=1}^{30}$  based on ionizable groups defined in literature<sup>34</sup>. 30 models were then trained in an 438 iterative all-but-one scheme. Model internal performance was evaluated using the logarithmic 439 characteristic curve (logAUC) 440 receiver operating and AUC using the BCL model:ComputeStatistics application. 441

442

#### 443 *Model Output and Evaluation*

444 MTCs with N output labels calculate N local positive predictive values (localPPV), where the i<sup>th</sup> 445 localPPV denotes the probability that the  $pK_a$  lies in the i<sup>th</sup>  $pK_a$  interval (see *Molecular Features* 446 above). For model evaluation, we report mean absolute error (MAE) for reader familiarity. In the 447 supplement, we also provide a Brier score for each model, which is a proper scoring rule<sup>1</sup> for 448 more rigorous evaluation of classification model output.

For each molecule  $m_i \in Y$  in a test set Y with M molecules, the pK<sub>a</sub> of  $m_i$  was encoded with a binary result label  $R_i \in \mathbb{Z}_2^N$  as described above. MTC scored each molecule, providing a discrete probability distribution  $P_i$  describing the molecule's likely pK<sub>a</sub> interval membership. From these distributions, MAE was calculated as:

453 
$$MAE(P,Y) = \frac{1}{M} \sum_{i=1}^{M} |y_i - E[P_i]|$$

454 where  $E[P_i]$  is the expected value of  $P_i$ . Similarly, for several models a Brier score was 455 calculated as:

456

$$BS(P,Y) = \frac{1}{M} \sum_{i=1}^{M} \sum_{j=1}^{N} (r_{ij} - p_{ij})^{2}$$

<sup>&</sup>lt;sup>1</sup> "Proper scoring rule" is a term in statistics for a loss function that is minimized if the probability distribution output from the model is identical to the ground-truth probability distribution. When the bidirectional holds, the scoring rule is further labeled a *strictly proper* scoring rule.

458 Where  $r_{ij} \in R_i$  is the j<sup>th</sup> result label for the i<sup>th</sup> molecule in the test set (i.e., whether its pK<sub>a</sub> value 459 lies in the j<sup>th</sup> pK<sub>a</sub> interval), and  $p_{ij}$  is the localPPV that the i<sup>th</sup> molecule's pK<sub>a</sub> value lies in the j<sup>th</sup> 460 pK<sub>a</sub> interval.

Throughout, MAE is used throughout to compare MTC to Regression and MTC to MTC models.
 Percent accuracy of categorization is not included, as it is an improper and discontinuous scoring

463 metric.

464

465 *Atomic sensitivity analysis* 

466 Atom replacement schemes were coded in C++ within the BCL. A parent molecule m is scored 467 by BCL-XpKa to produce P, a discrete probability distribution of potential pK<sub>a</sub> values. 468 Heteroatom a in the parent molecule is replaced with an appropriately hybridized carbon atom, 469 and the perturbed molecule is rescored to produce P'<sub>a</sub>. The dissimilarity between these 470 distributions was calculated by their Kullback-Leibler (KL) divergence:

471 
$$D_{KL}(P'_{a}||P) = \sum_{j=1}^{N} P'_{a,j} \ln (\frac{P'_{a,j}}{P_{j}}),$$

where  $P_j$  and  $P'_{a,j}$  are localPPVs as described in *Model Output and Evaluation*. Briefly, the KL divergence of these two probability distributions is best interpreted as the relative entropy between these distributions, where  $D_{KL}(P'||P) = 0$  denotes the distributions are identical (there would be no "surprises" if a given sample came from P vs P'), and higher values denote more dissimilarity. Finally, KL divergences were empirically denoised to generate ASA scores:

477 
$$ASA(m, a) = e^{[5*D_{KL}(P'_a||P)]} - 1$$

478

### 479 PROTAC ternary complex ensemble generation

480 Ternary complexes (TCs) were constructed according to Drummond et al (2020). Briefly, 481 protein-protein interactions (PPIs) with the PROTAC binding pockets near each other, as well as 482 a set of up to 10000 PROTAC conformations, were produced in the Molecular Operating 483 Environment (MOE). PROTAC conformations were then docked into the PPIs and filtered 484 according to the authors' criteria. TCs were then clustered on both protein- and PROTAC-485 conformations to produce "double clusters." Protein-conformational clustering was done at CA-486 RMSD < 10Å. PROTAC clustering was done at heavy-atom RMSD < 2.5Å.</p>

- 487
- 488 Hardware

All models were trained with 18 Intel Xenon W-2295 CPU cores. PROTAC TC formation was
 performed using an Nvidia RTX A5000 GPU.

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