

# 1 Title

2 Quantifying Concentration Distributions in Redox Flow Batteries with Neutron Radiography

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28

## 29 **Abstract**

30 The continued advancement of electrochemical technologies requires an increasingly detailed  
31 understanding of the microscopic processes that control their performance, inspiring the development  
32 of new multi-modal diagnostic techniques. Here, we introduce a neutron imaging approach to enable  
33 the quantification of spatial and temporal variations in species concentrations within an operating redox  
34 flow cell. Specifically, we leverage the high attenuation of redox-active organic materials (high  
35 hydrogen content) and supporting electrolytes (boron-containing) in solution and perform subtractive  
36 neutron imaging of active species and supporting electrolyte. To resolve the concentration profiles  
37 across the electrodes, we employ an in-plane imaging configuration and correlate the concentration  
38 profiles to cell performance with polarization measurements under different operating conditions.  
39 Finally, we use time-of-flight neutron imaging to deconvolute concentrations of active species and  
40 supporting electrolyte during operation. Using this approach, we evaluate the influence of cell polarity,  
41 voltage bias and flow rate on the concentration distribution within the flow cell and correlate these with  
42 the macroscopic performance, thus obtaining an unprecedented level of insight into reactive mass  
43 transport. Ultimately, this diagnostic technique can be applied to a range of (electro)chemical  
44 technologies and may accelerate the development of new materials and reactor designs.

45

46 **Keywords:** neutron imaging, redox flow batteries, *operando* diagnostics, concentration mapping, mass  
47 transfer, electrochemical energy storage.

48

## 49 Introduction

50 Contemporary energy storage technologies do not fulfil the stringent performance and cost requirements  
51 of the current and future electrical grid <sup>1-3</sup>. Developing cost-effective and high-performance storage  
52 platforms is paramount to integrate intermittent renewable energy technologies into the energy network  
53 <sup>4</sup>. Among the existing battery technologies, redox flow batteries (RFBs) have emerged as a promising  
54 candidate for large-scale energy storage. Intrinsic to their design, RFBs offer opportunities for economic  
55 scaling as the electrolyte reservoirs and the reactor size can be independently scaled, decoupling the  
56 power rating and energy capacity <sup>5</sup>. Furthermore, flow batteries are easy to manufacture, can utilize  
57 various electrolyte chemistries, are easily recyclable and safer <sup>5,6</sup>. Common RFB architectures consist  
58 of an electrochemical stack with alternating electrodes, separators and bipolar plates where the electrical  
59 power is generated, and external storage tanks containing the electrolyte solutions where the energy is  
60 stored. The electrolyte is pumped through the reactor and is composed of dissolved redox active  
61 materials that can be reversibly converted between different oxidation states. The redox reactions are  
62 sustained at the surface of the porous electrodes through which the electrolyte continuously flows, and  
63 an ionic current is carried through the separator to maintain the electroneutrality in each half-cell <sup>7</sup>.  
64 However, electrochemical losses (i.e., ohmic, charge transfer and mass transfer overpotentials) impact  
65 the overall efficiency and cost of the battery, which challenges their economic competitiveness <sup>8</sup>. Thus,  
66 materials research for RFBs is focused on advancing the component properties to improve the overall  
67 performance and durability of the system <sup>9-11</sup>.

68 The stack performance and lifetime are governed by multiple properties of the individual cell  
69 components and their complex interplay, which gives rise to phenomena such as reactive mass transport.  
70 The physicochemical properties of the electrolyte and the main components (porous electrodes and  
71 separator), together with the reactor design (flow field geometry, transport lengths) determine the  
72 available surface area for reactions and control the transport of mass and charge. The electrode  
73 microstructure dictates the trade-off between the pressure drop and the accessible surface area <sup>12</sup>,  
74 whereas the separator type, chemical composition and nanostructure <sup>13</sup> impact the RFB performance by  
75 regulating the transport of species between the positive and negative compartments. Conventionally,  
76 the cell performance is assessed using electrochemical diagnostic tools (e.g., polarization curves,  
77 charge-discharge capacity, electrochemical impedance spectroscopy) together with *ex-situ*  
78 characterization methods (e.g., scanning electron microscopy with energy dispersive X-ray analysis)  
79 where novel materials are macroscopically evaluated by empirical comparison (internal surface area,  
80 porosity, tortuosity) with the current state-of-the-art <sup>14</sup>. Although this is a valid approach to screen  
81 promising material candidates, valuable information is lost due to the complexity of identifying local  
82 performance-limiting factors related to electrode utilization and mass transport within the reactor (i.e.,  
83 flow fields, porous electrodes and separator). From the perspective of an electrochemical diagnostic  
84 technique, the cell is a homogenous system, a so-called “black box”, with inputs and outputs in the form  
85 of current or voltage. Electrochemical cells, however, are anisotropic systems with an inhomogeneous  
86 distribution of reaction rate, current and species concentration within the reactor volume. This gives  
87 rise to a myriad of limiting phenomena in the cell that cannot be observed with conventional  
88 electrochemical techniques, including gas evolution, improper electrode wetting, local depletion in  
89 species concentration, membrane crossover and precipitation. Thus, to properly correlate the material  
90 properties to the device performance, the development of locally resolved characterization methods is  
91 essential <sup>15-17</sup>.

92 The most straightforward route to access information at a microscopic level is employing  
93 *operando* imaging of electrochemical systems in tandem with complementary electrochemical  
94 diagnostics, which has been instrumental in the rapid development of polymer electrolyte fuel cells <sup>18,19</sup>

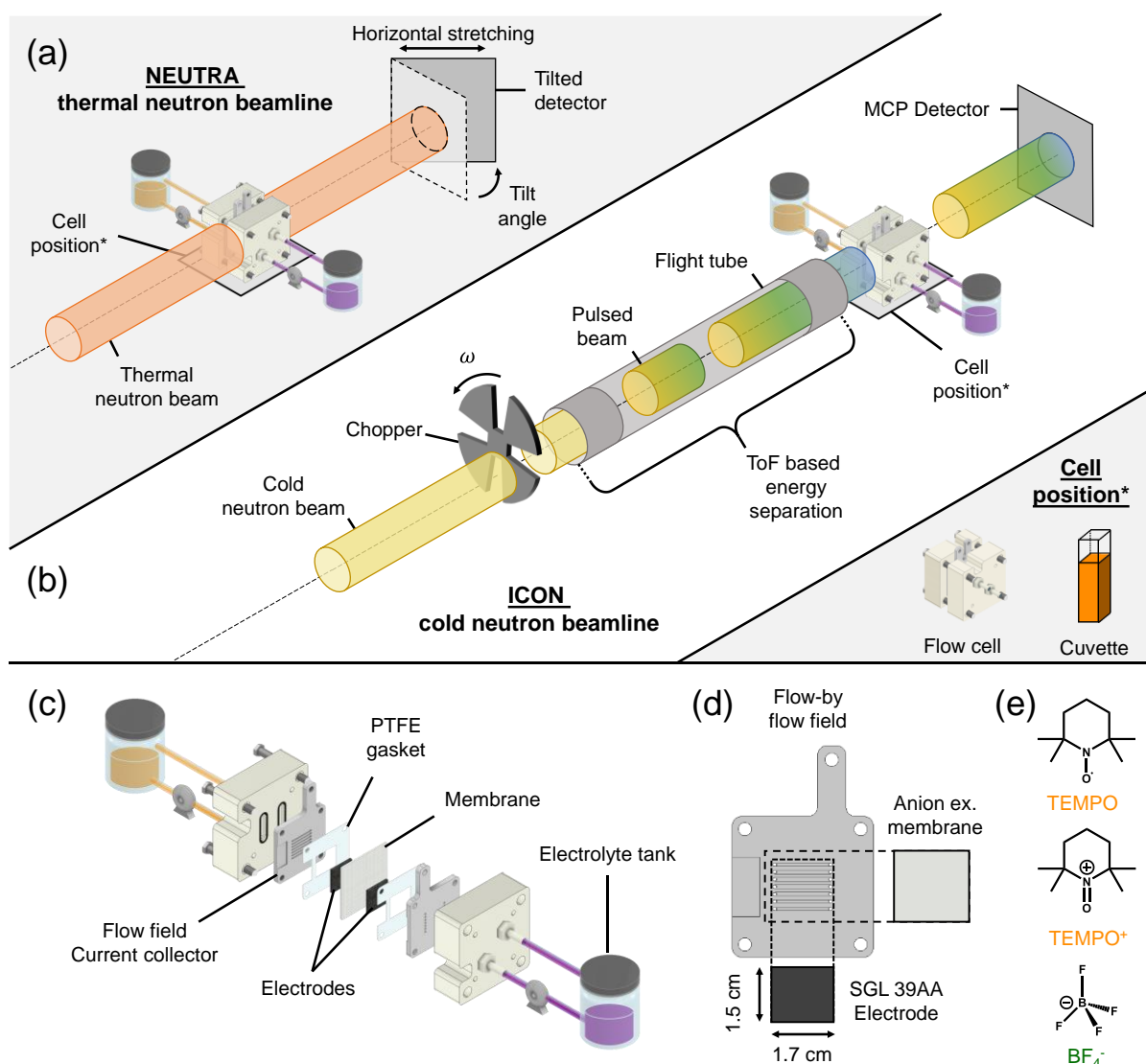
95 and lithium-ion batteries <sup>20,21</sup>. Over the last years, several groups have developed imaging and  
96 spectroscopic approaches that enable *operando* characterization of RFBs. In the following, we describe  
97 a set of representative examples, and not an exhaustive list of advances, on *operando* characterization  
98 of redox flow cells. Wong *et al.* applied fluorescence microscopy and particle velocimetry to a quinone-  
99 based flow battery to quantitatively image active species and velocity distributions near the electrode-  
100 flow field interface and highlight inhomogeneous flow profiles within the porous electrodes <sup>22</sup>. This  
101 technique holds promise to image flow velocity profiles and to resolve the electrochemical activity at  
102 the electrode-electrolyte interface. However, fluorescent species are necessary for operation, the  
103 electrochemical cell must be modified with a transparent window and the technique is restricted to a  
104 limited penetration depth through porous carbon or metallic electrodes. Moreover, Jervis *et al.*, Tariq  
105 *et al.*, Eifert *et al.* and Köble *et al.* employed X-ray tomographic microscopy to visualize gas pockets  
106 within the liquid electrolyte <sup>23–26</sup>. The authors successfully separated the carbon electrode phase from  
107 the liquid electrolyte and the gas pockets present in the porous electrodes, showing incomplete wetting  
108 of carbon fibre electrodes. X-ray imaging can yield 3D reconstructions of electrode microstructures and  
109 can uncover wetting dynamics in porous media. However, despite being considered non-destructive,  
110 high X-ray doses are generally needed to obtain high temporal and spatial resolutions, while low doses  
111 are necessary to stay under the X-ray-induced degradation threshold of the materials under  
112 investigation, causing a trade-off between resolution and exposure time <sup>27–29</sup>. Zhao *et al.* coupled *in-situ*  
113 nuclear magnetic resonance and electron paramagnetic resonance to track reaction mechanisms  
114 occurring within the electrolyte <sup>30,31</sup>. It is an effective technique to monitor the state-of-charge of the  
115 electrolyte within the tanks and to track chemical transformations. However, nuclear magnetic  
116 resonance-based techniques suffer from challenging cell-design modifications to probe the  
117 concentration within the reactor volume. While these techniques have provided important insights into  
118 flow distributions, electrode wetting, electrolyte state-of-charge and degradation, a technique that  
119 enables *operando*, reactor-level and non-invasive imaging of the electrochemical cell is necessary to  
120 understand how the microscopic properties of materials impact the cell performance.

121 Neutron radiography is an alternative technique to fulfil these requirements and has been  
122 applied to electrochemical devices for more than two decades <sup>32</sup>. Neutron imaging is a non-invasive  
123 technique that is suitable to study systems during extended operation. Neutrons are electrically neutral  
124 particles and therefore offer a high penetration depth even through high atomic weight elements, as  
125 neutrons only interact with the nuclei and not with the electron cloud, resulting in no or minimal  
126 modification of the electrochemical cell design for imaging <sup>33</sup>. One of the first studies was reported by  
127 Mosdale *et al.* who calculated the water content within the membrane of a proton exchange membrane  
128 fuel cell *in operando* <sup>34</sup>. Due to the through-plane configuration of their setup, the authors designed a  
129 cell with neutron-transparent components, enabling selective investigation of the membrane hydration.  
130 Following this work, neutron radiography was mainly applied to develop the understanding of proton  
131 exchange membrane fuel cells <sup>18,32,35–38</sup>. The interest of using neutron imaging to study fuel cells resides  
132 in the high attenuation of hydrogen atoms in liquid water and low attenuation of gases (O<sub>2</sub>, air, H<sub>2</sub>),  
133 alongside the high neutron penetration through the materials constituting the cell. Therefore, the use of  
134 neutron imaging improved the fundamental understanding of water management in fuel cells under  
135 various operating conditions <sup>39–42</sup>. Additionally, neutron imaging studies cover other electrochemical  
136 technologies such as lithium-ion batteries <sup>20,43–46</sup>, electrolyzers <sup>47,48</sup> and alkaline batteries <sup>46,49</sup>. More  
137 recently, Clement *et al.* used neutron radiography to study the gas evolution and correlated it to the  
138 properties of the electrode material during the charging process of a vanadium RFB <sup>50</sup>. We hypothesize  
139 that neutron imaging can offer more than contrast between liquid, gas and solid phases, enabling  
140 visualization of the dissolved species within the electrode pore space <sup>51,52</sup>. Unlike X-rays, neutrons have  
141 no marked trend in the distribution of mass attenuation coefficients across the periodic table, so contrast

142 between elements does not follow a predictable trajectory. However, several elements (e.g., H, Li, B),  
143 which are central to redox flow batteries, feature high attenuation of hydrogen atoms and make neutrons  
144 highly suited to study organic redox molecules<sup>53,54</sup>, which are of interest for various electrochemical  
145 technologies such as RFBs and CO<sub>2</sub>-electrolyzers. These features make it possible to perform neutron  
146 radiography of non-aqueous electrochemical systems and to obtain contrast between the redox active  
147 molecules and the supporting salts, opening an avenue to study *in situ* or *operando* motion of species  
148 in performance-defining materials such as separators and porous electrodes.

149 Here we explore, for the first time, the use of neutron imaging to reveal reactive transport  
150 phenomena and concentration distributions in non-aqueous redox flow batteries (NAqRFBs) as a model  
151 system to demonstrate this approach. NAqRFBs are an attractive option for energy storage because of  
152 their larger electrochemical stability windows and their use of organic molecules made from earth-  
153 abundant elements. In this work, neutron radiography is used to extract concentration profiles of redox-  
154 active species or supporting salts within the reactor of an *operando* NAqRFB. Resolving the  
155 concentration profiles locally in the reactor area (i.e., the performance-defining region of the RFB  
156 including the flow fields, electrodes and separator) is a powerful complementary approach to the  
157 contemporary macroscopic characterization techniques and can be used to elucidate the coupled mass  
158 transport phenomena (convection, diffusion, migration, reaction). In this work, we perform in-plane  
159 transmission neutron imaging of NAqRFBs in two beamlines offering distinct capabilities in terms of  
160 resolution and species identification. Compared to conventional through-plane imaging, in-plane  
161 imaging can reveal the concentration gradients emanating from the flow field channels towards the  
162 separator, provided appropriate spatial resolution is achievable. The NEUTRA beamline operates with  
163 thermal neutrons (white-beam) allowing cumulative concentration profiles of the active materials and  
164 the supporting salts with a high spatial resolution over the region of interest. The ICON beamline  
165 operates with cold neutrons and utilizes the time-of-flight neutron imaging (ToF-NI) as a spectral  
166 technique to image the reactor via energy-resolved neutron radiography, but the use of this technique  
167 results in a drop in temporal resolution caused by switching to a pulsed source. However, ToF-NI  
168 enables the deconvolution of concentration profiles of several species in the electrolyte (i.e., active  
169 species and supporting ions). Thus, by adding the time dimension we can resolve the movement of  
170 species between half cells under a voltage bias, giving insight into the coupled transport phenomena  
171 within the reactor area.

172 In the first part of this work (Figure 1a) the experiments performed at the NEUTRA beamline  
173 are described during RFB polarization to extract concentration profiles of 2,2,6,6-tetramethylpiperidin-  
174 1-yl)oxyl (TEMPO) in its neutral and oxidized form, dissolved in solution with a low neutron  
175 attenuating supporting salt, potassium hexafluorophosphate (KPF<sub>6</sub>) or with a highly attenuating  
176 counter-ion, tetrafluoroborate (BF<sub>4</sub><sup>-</sup>). To this end, calibration curves are obtained for all species used in  
177 this work, after which the cells are imaged during operation and concentration profiles of dissolved  
178 species in the reactor volume are extracted. In the second part of this work (Figure 1b), polarization  
179 experiments are performed using TEMPO with BF<sub>4</sub><sup>-</sup> as counter-ion at the ICON beamline. First, the  
180 neutron attenuation of the electrolyte solutions is calibrated at different neutron energies prior to  
181 *operando* imaging. By utilizing the principle of energy-dependency of their neutron cross-sections, we  
182 quantify the concentration change of active species and supporting salt separately during operation and  
183 reveal the dominant transport mechanisms within the electrodes and between half cells under voltage  
184 stimuli. This approach pushes the limits of neutron imaging by probing concentration profiles and  
185 species movement evolving in an operating flow cell and we hope it will serve as a guide for researchers  
186 intending to perform species-sensitive *operando* neutron imaging.



187

188 Figure 1: Schematic representations of (a-b) the neutron imaging set-ups, and (c-e) the flow battery cell design  
 189 and components, utilizing non-aqueous electrolytes. (a) Neutron imaging using the NEUTRA beamline, where a  
 190 cumulative concentration profile of TEMPO, TEMPO<sup>+</sup> and BF<sub>4</sub><sup>-</sup> species is obtained. (b) Neutron imaging using  
 191 the ICON beamline, where the moderation of the neutron velocities enables to select the energy of the neutrons  
 192 by means of a mechanical chopper disk, a flight tube and a microchannel plate detector, where deconvoluted  
 193 concentration profiles of active species (TEMPO and TEMPO<sup>+</sup>) and supporting electrolyte anions (BF<sub>4</sub><sup>-</sup>) are  
 194 obtained. The cell positions in (a-b) were used for both flow cell and cuvette imaging. (c) Exploded view of the  
 195 flow cell components and the connected electrolyte tanks. (d) Representation of the cell components (parallel flow  
 196 field, anion exchange membrane and SGL 39AA porous electrode) with their dimensions. (e) Chemical structures  
 197 of the attenuating active species TEMPO and TEMPO<sup>+</sup>, and the attenuating supporting species BF<sub>4</sub><sup>-</sup>.

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## 199 Results and discussion

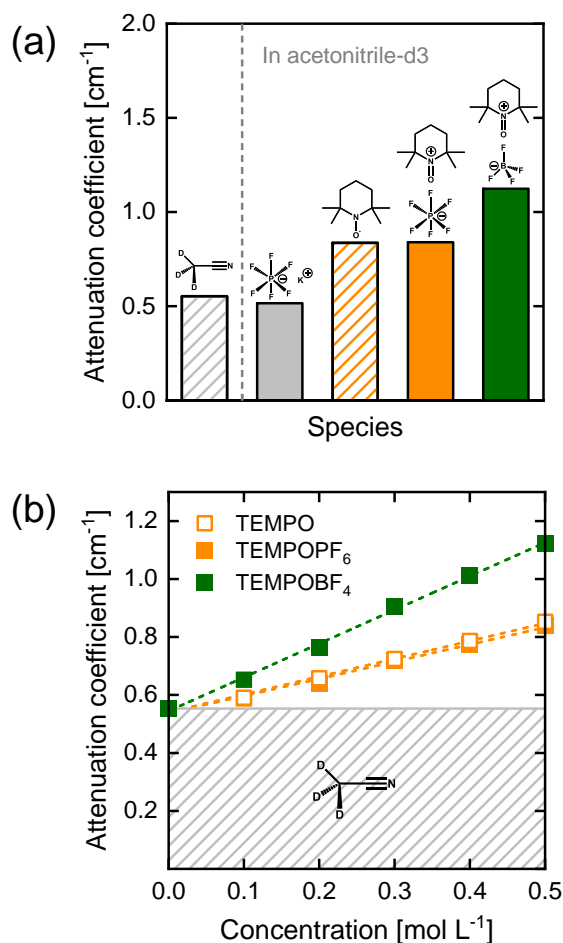
200 First, we discuss the results of the white-beam imaging obtained at the NEUTRA beamline, followed  
201 by the ToF-NI performed at the ICON beamline. Each section describes the *ex-situ* calibrations used to  
202 correlate the concentrations of species in the electrolyte with neutron attenuations, and the  
203 characterization of concentration profiles in the *operando* flow cells under various voltage biases and  
204 flow configurations. In the NEUTRA section, two sets of experiments are performed, one with a low  
205 attenuating supporting salt ( $\text{KPF}_6$ ) and one with a highly attenuating counter-ion ( $\text{BF}_4^-$ ), to differentiate  
206 between the redox active species and supporting ions.

207

### 208 White beam neutron imaging (NEUTRA)

#### 209 *Attenuation of electrolyte species*

210 Achieving contrast between the electrolyte constituents (solvent, redox-active species and supporting  
211 ions) is critical to identifying species and quantifying their dynamics within the electrochemical cell.  
212 White beam neutron imaging does technically not allow selectivity towards a target component, but it  
213 is possible to obtain insights in concentration distributions of individual electrolyte species by careful  
214 selection of the redox active species and supporting salt, coupled with subtractive imaging. We  
215 capitalize on the flexibility in the choice of solvent, supporting electrolytes and redox-active molecule  
216 for NAqRFBs, and measure attenuation coefficients for a set of electrolyte types and components using  
217 cuvettes (Figure 2a). The attenuation difference between the neat deuterated solvent ( $\text{CD}_3\text{CN}$ ) and 0.2  
218 M supporting salt solution ( $\text{KPF}_6$  in  $\text{CD}_3\text{CN}$ ) is sufficiently small to be neglected, confirming the  
219 negligible attenuation of  $\text{KPF}_6$  at this neutron energy and concentration. On the other hand, the addition  
220 of 0.5 M TEMPO in this electrolyte solution results in a larger attenuation coefficient as it has four  
221 methyl groups rich in hydrogen atoms attached to a piperidine ring (molecular formula  $\text{C}_9\text{H}_{18}\text{NO}$ ). The  
222 large number of hydrogen atoms results in a stark contrast between the supporting salt ( $\text{KPF}_6$ ) and the  
223 active species (TEMPO/TEMPO<sup>+</sup>). For the concentration range investigated in this study (0 - 0.5 M),  
224 TEMPO and TEMPO<sup>+</sup>PF<sub>6</sub><sup>-</sup> dissolved in  $\text{CD}_3\text{CN}$  show similar neutron attenuations (Figures 2a and 2b).  
225 The similar cross sections of TEMPO and TEMPO<sup>+</sup> are expected given their identical chemical  
226 composition (only one electron difference) resulting in almost identical interaction with neutrons, and  
227 further confirms the low attenuation of PF<sub>6</sub><sup>-</sup> ions. Finally, when the counter-ion (PF<sub>6</sub><sup>-</sup>) of TEMPO<sup>+</sup> is  
228 replaced with BF<sub>4</sub><sup>-</sup>, the attenuation at the same concentration is nearly doubled (Figure 2a), which  
229 indicates that TEMPO species and BF<sub>4</sub><sup>-</sup> ions have similar microscopic cross-sections. Although the  
230 counter-ion contains no hydrogen atoms, BF<sub>4</sub><sup>-</sup> contains boron which features a large neutron absorption  
231 cross-section for thermal neutrons<sup>55</sup>. Figure 2b shows a linear correlation between neutron attenuation  
232 vs. concentration for the different species employed in this study, which confirms the validity of the  
233 chosen operating range (0 – 0.5 M) where the Lambert-Beer law (equation (1)) holds. The microscopic  
234 cross-sections obtained here are then applied to obtain local concentrations in the electrochemical  
235 reactor volume during operation. The electrolyte compositions used in this work provide selectivity  
236 towards both the redox molecules and supporting salt by utilizing salts with low and high attenuation  
237 with the neutron beam. On the other hand, minimizing the neutron attenuation of the redox active  
238 molecules through a reduced hydrogen content or deuterium labelling is another strategy to obtain  
239 contrast in multicomponent electrolytes. Combining both approaches would therefore be a powerful  
240 approach for future work to tune the neutron white beam imaging selectivity by transforming a complex  
241 multicomponent system into a single component system.



242

243 Figure 2: Determination of the attenuation coefficient for the chemicals used in this study obtained at the  
 244 NEUTRA beamline. (a) The attenuation coefficients of the different species in CD<sub>3</sub>CN: the solvent only,  
 245 supporting electrolyte (0.2 M) and species TEMPO, TEMPO<sup>+</sup>PF<sub>6</sub><sup>-</sup> and TEMPO<sup>+</sup>BF<sub>4</sub><sup>-</sup> (all 0.5 M). (b) The linear  
 246 dependence of the macroscopic neutron cross-section of the TEMPO, TEMPO<sup>+</sup>PF<sub>6</sub><sup>-</sup> and TEMPO<sup>+</sup>BF<sub>4</sub><sup>-</sup> species to  
 247 the concentration (0.1, 0.2, 0.3, 0.4 and 0.5 M), where the shaded area represents the attenuation of the solvent.

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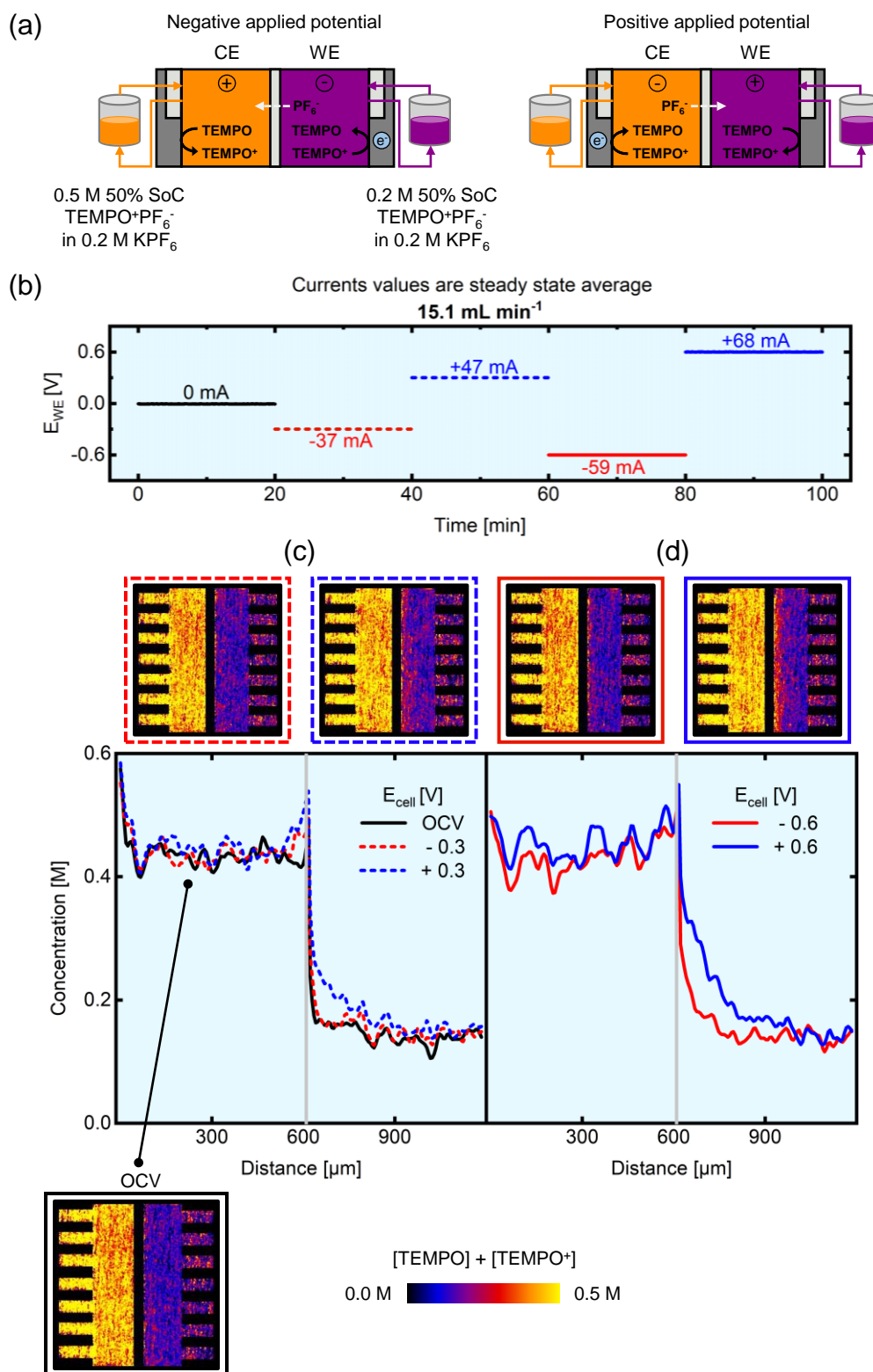
### 249 *Transport of the active species*

250 We performed neutron imaging on an operating redox flow cell to visualize concentration profiles of  
 251 TEMPO/TEMPO<sup>+</sup> (Figure 3). The cell is connected to tanks with 50% SoC TEMPO/TEMPO<sup>+</sup> at 0.5 M  
 252 concentration on the CE side and 0.2 M on the WE side, both with 0.2 M KPF<sub>6</sub> to provide ionic  
 253 conductivity and minimize supporting salt impact on neutron attenuation (Figure 3a). Because the WE  
 254 and CE compartments are separated by an anion exchange membrane, the transport of cations such as  
 255 TEMPO<sup>+</sup> and K<sup>+</sup> is significantly hindered, whereas the anions and neutral molecules such as PF<sub>6</sub><sup>-</sup> and  
 256 TEMPO can more easily pass through. Using this cell architecture and due to the negligible neutron  
 257 attenuation of KPF<sub>6</sub>, we can track the movement of TEMPO between the electrodes. The cell is  
 258 discharged (negative potential applied at the WE) and charged (positive potential applied at the WE)  
 259 alternately, such that the state-of-charge after each complete cycle does not significantly deviate from  
 260 the initial condition and two voltage magnitudes were applied to understand their impact on the  
 261 potential-driven transport processes (e.g., migration). The electrochemical sequence goes through OCV,  
 262 -0.3, +0.3, -0.6 and +0.6 V steps, each for 20 min at the highest tested inlet flow rate of 15.1 mL min<sup>-1</sup>  
 263 (Figure 3b). We also studied the impact of flow rate by performing the same electrochemical sequence  
 264 (without the OCV step) at 5.6 mL min<sup>-1</sup> (Figure S1). The current-time and voltage-time curves of the



265 entire experiment can be found in Figure S2 and a video of the experiment can be found in the  
266 Supplementary Materials. *Operando* imaging of the cell during the electrochemical protocol results in  
267 transmission images where the attenuation at each location represents the integral of neutron-matter  
268 interactions along the neutron path (Figure 1a). These images are then averaged for the duration of a  
269 voltage step (20 min) and result in concentration maps for a given condition at steady-state (Figure 3c-  
270 d). The colour scale represents the cumulative concentration of TEMPO and TEMPO<sup>+</sup> and ranges from  
271 0 - 0.5 M, resulting in a 2D map of the species concentration in the reactor area. The membrane area is  
272 omitted as the quantification of concentrations is not reliable in this region due to the high hydrogen  
273 content of the polymer membrane (perfluorinated with a polyketone reinforcement) and the reduced  
274 membrane thickness (130 μm). Finally, we calculate the concentration profiles across the thickness of  
275 the electrodes and compute these between the flow field-electrode interfaces of both half cells. Using  
276 this approach, one-dimensional concentration profiles, parallel with the electrical field, are obtained.

277 The experiment begins with an OCV step where no current is drawn from the cell. The brighter  
278 colour of the CE side in the OCV radiograph represents a higher concentration compared to the WE  
279 side, as expected by the concentrations of the electrolyte fed (0.5 M and 0.2 M TEMPO/TEMPO<sup>+</sup>). An  
280 advantage of neutron radiography is that electrolyte wetting of the porous electrodes can be visualized  
281 because of the low attenuation of gasses, which will appear as dark spots (i.e., lower concentration) in  
282 the radiographs<sup>56</sup>. During the OCV period, the concentration on both sides does not show dark regions  
283 (Figure 3c-d), suggesting full wetting, at least to the spatial resolution of the measurement. Moreover,  
284 over the course of the OCV period, the concentration profile remained fairly constant which can be  
285 attributed to the low diffusion rate of TEMPO/TEMPO<sup>+</sup> through the dense anion exchange membrane.  
286 Overall, the concentration profiles under cell polarization do not strongly deviate from the initial OCV  
287 state, except at positive potentials near the membrane area on the WE, which can be explained by the  
288 reactor configuration (i.e., anion exchange membranes, flow-by flows fields) and the low ionic  
289 conductivity of the nonaqueous electrolyte, resulting in low current densities and charge consumption  
290 (<15%, see Figure S3). In this experiment, the concentration gradient is from left to right due to the  
291 higher cumulative TEMPO/TEMPO<sup>+</sup> concentration on the CE side. Under negative potentials (-0.3 V  
292 and -0.6 V), TEMPO is converted to TEMPO<sup>+</sup> in the CE side, resulting in a build-up of TEMPO<sup>+</sup>, while  
293 the opposite reaction is taking place in the WE, resulting in TEMPO<sup>+</sup> depletion. To compensate for the  
294 charge, PF<sub>6</sub><sup>-</sup> crosses through the membrane towards the CE side, which is not visible in the images due  
295 to its low attenuation. Although favourable to sustain the electrochemical reaction, we do not expect  
296 TEMPO to cross to the CE side on the timescale of this experiment as this would be against its  
297 concentration gradient, thus the images and profiles for negative potentials are nearly identical to the  
298 OCV conditions (Figure 3c).



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Figure 3: *Operando* imaging of the active species transport in the NEUTRA beamline with the low attenuating KPF<sub>6</sub> supporting salt. (a) Schematic representation of the non-aqueous cell designs during charge and discharge mode, where the counter electrode (CE) corresponds to 0.5 M TEMPO/TEMPO<sup>+</sup>PF<sub>6</sub><sup>-</sup> at 50% state-of-charge in 0.2 M KPF<sub>6</sub> and the working electrode (WE) to 0.2 M TEMPO/TEMPO<sup>+</sup>PF<sub>6</sub><sup>-</sup> at 50% state-of-charge in 0.2 M KPF<sub>6</sub>. (b) Electrochemical sequence over time showing the applied potential steps and measured averaged current output at an inlet flow rate of 15.1 mL min<sup>-1</sup>. (c-d) Cumulative active species (TEMPO/TEMPO<sup>+</sup>) concentration profiles over the electrode thickness at an inlet flow rate of 15.1 mL min<sup>-1</sup>. The averaged snapshots of the cell after image processing and the concentration profiles are shown for various applied potential steps: (c) OCV, -0.3 V and +0.3 V and (d) -0.6 V and +0.6 V.

309 On the contrary, when positive potentials are applied (+0.3 V), the TEMPO concentration in  
310 the CE side increases, amplifying its existing concentration gradient towards the WE. This results in a  
311 stronger diffusive flux of TEMPO towards the WE side at positive potentials, supported by the bright  
312 concentration front in the corresponding radiographs, together with the steep concentration profiles near  
313 the membrane. Increasing the potential to +0.6 V amplifies this trend as more TEMPO<sup>+</sup> is converted to  
314 TEMPO on the CE side, exacerbating the concentration gradient of TEMPO and extending the  
315 concentration front deeper within the WE (Figure 3d). Decreasing the flow rate to 5.6 mL min<sup>-1</sup> further  
316 intensifies the concentration front in the WE (Figure S1) for both the positive and negative potentials.  
317 The concentration front reveals mass transfer limitations, determined by the membrane properties,  
318 applied potential, electrolyte velocity, species concentration and electrolyte and electrode properties<sup>8</sup>.  
319 To visualize such limiting phenomena, we utilized a flow-by flow field design that induces limited  
320 convection within the porous electrode. However, the intensification of the concentration fronts  
321 suggests that this flow field does have convective transport contributions in the electrode. Nevertheless,  
322 we anticipate that a convection-enhanced flow field (such as interdigitated or flow-through) would  
323 further increase species replenishment and reduce concentration gradients<sup>57</sup>. In this first set of  
324 experiments, the low neutron cross-section of the KPF<sub>6</sub> salt was utilized to maximize the contrast of  
325 TEMPO and TEMPO<sup>+</sup> compared to other electrolyte components. To visualize the motion of anions,  
326 we then employ a strongly attenuating counter-ion (BF<sub>4</sub><sup>-</sup> instead of PF<sub>6</sub><sup>-</sup>) without any additional  
327 supporting salt to amplify the contrast between all species in the electrolyte.

328

### 329 ***Transport of the counter-ion***

330 Supporting ions are essential in RFBs to provide ionic conductivity<sup>58,59</sup>. Here we leverage BF<sub>4</sub><sup>-</sup> as  
331 counterion due to its high neutron attenuation (see *Attenuation of electrolyte species*)<sup>60</sup>. To quantify the  
332 influence of migration on the charged species transport (i.e., stoichiometric operation), we do not add a  
333 supporting salt in the electrolyte (Figure 4a), which negatively impacts the obtained current density  
334 (Figure S4) but enables visualization of the counterion. Furthermore, the use of an anion-exchange  
335 membrane significantly restricts TEMPO<sup>+</sup> movement between compartments, which leaves BF<sub>4</sub><sup>-</sup> as the  
336 main charge carrier. As we perform subtractive neutron imaging, the isolation of [TEMPO], [TEMPO<sup>+</sup>]  
337 and [BF<sub>4</sub><sup>-</sup>] is not possible with white beam neutron imaging, resulting in cumulative concentration maps  
338 (see the combined colour scale in Figure 4). Although the concentration information is cumulative, by  
339 tuning experimental parameters (type of ion-exchange membrane and tank concentrations) we  
340 hypothesize that the observed changes can be attributed to the motion of certain species, which is  
341 predominantly BF<sub>4</sub><sup>-</sup> in this configuration. This resonates with the subtle changes in the concentration  
342 profiles in Figure 3 as a function of the applied potential, hinting that the neutron-transparent PF<sub>6</sub><sup>-</sup> is the  
343 main charge carrier in the system. A novel approach to isolate the concentration of species in solution  
344 with neutron imaging is discussed in the *energy selective imaging* section.

345 In the first hour of the experiment, the system is kept at OCV conditions to track diffusional  
346 crossover through the membrane (Figure 4b). Although we track a change in OCV over time, indicating  
347 crossover and concentration equilibration, the small concentration variations in this short time period  
348 are not quantitatively captured by the radiographs (OCV radiograph in Figure 4c). Although BF<sub>4</sub><sup>-</sup> has a  
349 strong concentration gradient towards the CE side, the OCV profile does not show significant deviation  
350 from the initial concentrations, which is attributed to the Donnan exclusion of TEMPO<sup>+</sup> coupled with  
351 the barrier properties of the dense anion exchange membrane<sup>61</sup>. Furthermore, the concentrations in the  
352 reactor volume are homogeneous and no local fluctuations are observed. We conclude that the  
353 timeframe of the OCV period is shorter than the time needed for diffusional crossover of species for  
354 this configuration.

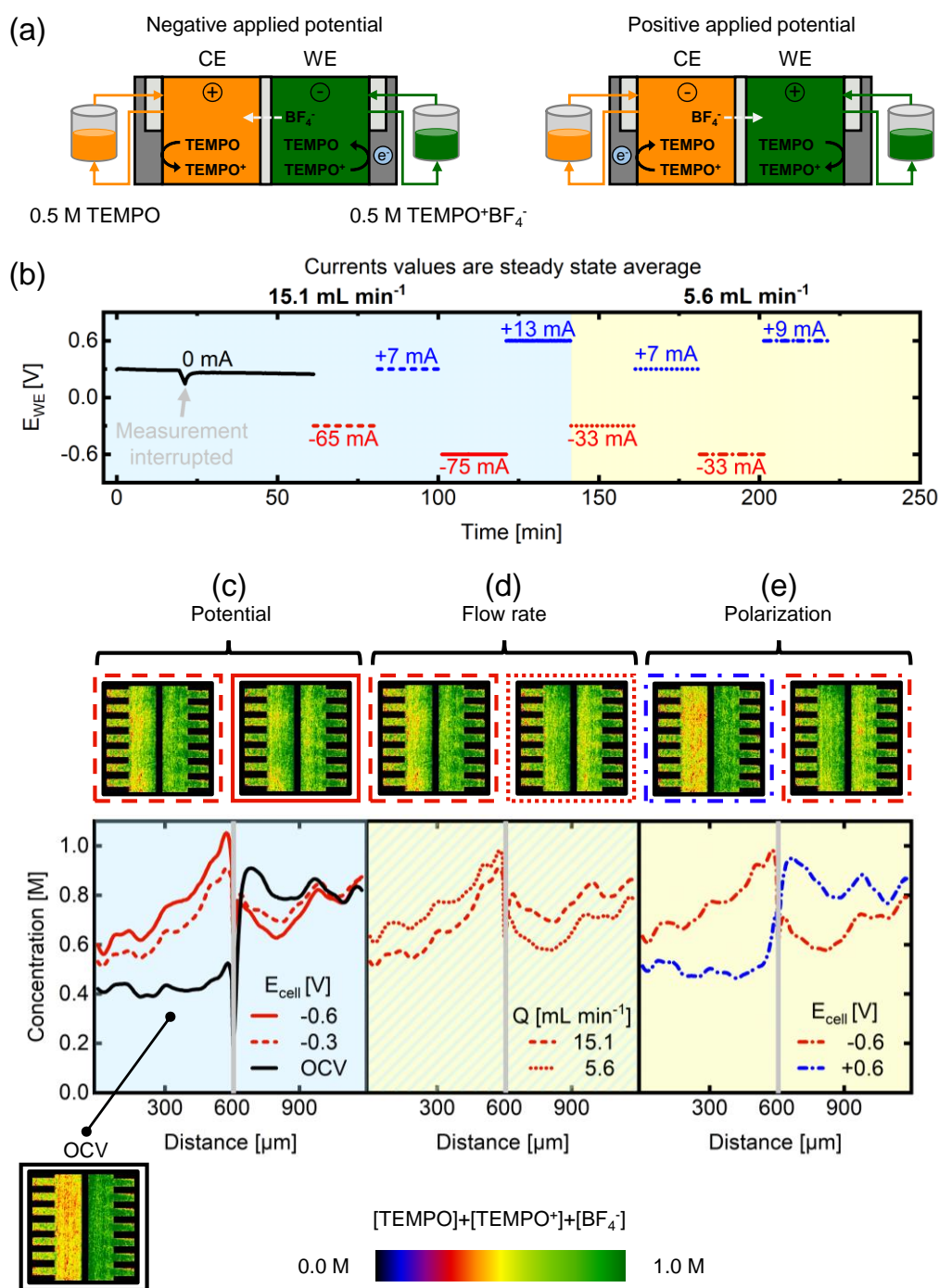
355 After the OCV period, alternating potential steps are applied to the electrochemical cell and the  
356 current response is recorded in time (Figure S4). At negative potentials, TEMPO<sup>+</sup> is converted to  
357 TEMPO in the WE, and BF<sub>4</sub><sup>-</sup> migrates to the CE compartment (Figure 4a). We can observe a local  
358 accumulation of attenuating species in the vicinity of the membrane on the CE side, while the opposite  
359 trend is observed in the WE compartment (Figure 4c), attributed to the migration of the BF<sub>4</sub><sup>-</sup> anion from  
360 the WE to the CE to maintain electroneutrality. This effect is more pronounced at a higher potential (-  
361 0.6 V vs. 0.3 V, Figure 4c), which illustrates the influence of migration. Starker concentration gradients  
362 are obtained at lower flow rates (5.6 mL min<sup>-1</sup>, Figure 4d), as the convective mass transfer is lower. At  
363 the WE, we find higher concentrations in the areas near the flow field inlets in comparison with the area  
364 under the ribs, showing an advantage of two-dimensional concentration maps obtained using neutron  
365 radiography. We hypothesize that velocity distributions within the porous electrodes, induced by the  
366 flow-by flow field design and the relatively thick porous electrode stack, explain these variations in  
367 concentration thorough the electrode volume. This phenomenon is more visible at the highest flow rate  
368 as the convective forces pushing the electrolyte in the porous electrode are larger (Figure 4d). In  
369 summary, at negative potentials, the profiles show an increasing species concentration in the CE  
370 occurring synchronously with a decrease in the WE (Figure 4c).

371 When a positive potential is applied to the WE, the reverse reactions take place, and the  
372 resulting radiographs and the concentration profiles (Figure 4e) are mirrored compared to negative  
373 applied potentials. Because of the reactor architecture used in the NEUTRA experiments (stacked paper  
374 electrodes and a flow-by flow field) in combination with the low ionic conductivity of the electrolyte,  
375 there is only a small change in the cell capacity at negative potentials, consuming only ~8% of the total  
376 capacity (Figure S5), resulting in relatively low current densities. Therefore, at positive potentials, only  
377 a small amount of TEMPO is present in the WE compartment to be converted back to TEMPO<sup>+</sup>. As a  
378 result, large overpotentials are generated throughout the cell due to the low concentration of reactants  
379 to sustain the current. This explains the asymmetry in the current magnitudes when the polarity of the  
380 cell is reversed (i.e., +7 mA vs. -65 mA at 15.1 mL min<sup>-1</sup> and +/-0.3 V, Figure 4b), and the even lower  
381 capacity recovery (~1-2%) resulting in underutilized capacity over the duration of the experiment  
382 (Figure S5).

383 When comparing the experiments with counter-ion BF<sub>4</sub><sup>-</sup> and supporting salt KPF<sub>6</sub>, we can  
384 correlate the macroscopic performance with the concentration distributions through the reactor. For the  
385 KPF<sub>6</sub> experiments, all active species (i.e., TEMPO, TEMPO<sup>+</sup>, PF<sub>6</sub><sup>-</sup>) are present in both compartments.  
386 Therefore, the macroscopic performance, i.e., the current output, is symmetric when operating at  
387 negative and positive applied potentials (Figure 3b) as the to-be-reacted species are present without the  
388 requirement of species crossover under the evaluated conditions (as the capacity change is limited to  
389 ~20%). The symmetric current output results in concentration profiles returning to the OCV profile  
390 when positive potentials are applied. Whereas for the BF<sub>4</sub><sup>-</sup> experiment, the charged species are only  
391 present in one compartment (WE) initially and are required to cross the membrane to support the  
392 reactions, which is limited by the anion exchange membrane, resulting in asymmetric current  
393 magnitudes upon changing cell polarities (Figure 4b). The asymmetric current can be correlated to the  
394 concentration profiles as for positive applied potentials, the concentration does not fully return to the  
395 OCV profiles (Figure S6).

396 Using white beam neutron imaging, we have obtained cumulative concentration maps which  
397 include active species and supporting electrolytes. Using this approach, we have coupled macroscopic  
398 electrochemical cell performance with microscopic concentration distributions, revealing mass transfer  
399 modes under different cell potentials, flow rates and cell polarities. However, we are not able to isolate  
400 concentrations of active species and supporting ions with this incident beam. Acknowledging these

401 limitations, we then utilize time-of-flight imaging to obtain quantitative insights into reactive transport  
 402 phenomena of both the active species and counter-ion, under similar experimental conditions (Figures  
 403 4 and 6).



404

405 Figure 4: *Operando* imaging of the active species transport in the NEUTRA beamline with the neutron attenuating  
 406 BF<sub>4</sub><sup>-</sup> supporting ion. (a) Schematic representation of the non-aqueous cell designs during charge and discharge  
 407 mode, where the counter electrode (CE) corresponds to 0.5 M TEMPO and the working electrode (WE) to 0.5 M  
 408 TEMPO<sup>+</sup>BF<sub>4</sub><sup>-</sup>. (b) Electrochemical sequence over time showing the applied potential steps and measured averaged  
 409 current output at two inlet flow rates of 15.1 mL min<sup>-1</sup> and 5.6 mL min<sup>-1</sup>. (c-e) Cumulative active species  
 410 (TEMPO/TEMPO<sup>+</sup>) and BF<sub>4</sub><sup>-</sup> supporting ion concentration profiles over the reactor area. The averaged snapshots  
 411 over the whole period of each individual potential step of the cell after image processing and the concentration  
 412 profiles are shown for various applied potential steps and show the influence of various operation parameters: (c)  
 413 applied potential magnitude (OCV, -0.3 V and -0.6 V at 15.1 mL min<sup>-1</sup>), (d) flow rate (-0.3 V at 15.1 mL min<sup>-1</sup>  
 414 and 5.6 mL min<sup>-1</sup>) and (e) Polarization sign (-0.6 V and +0.6 V at 5.6 mL min<sup>-1</sup>).

## 415 **Energy-resolved neutron imaging (ICON)**

416 In pursuit of deconvoluting the concentrations of different species in the electrolyte, we investigate the  
417 use of energy-resolved neutron radiography at the ICON beamline. This beamline utilizes a colder  
418 neutron spectrum by secondary moderation of the neutron beam, and slower neutrons undergo inelastic  
419 scattering events with a higher probability than thermal neutrons, allowing more variations in species  
420 cross-sections to be observed. It is also possible to perform spectral neutron imaging with a time-of-  
421 flight based technique at ICON, which is currently not possible at the NEUTRA beamline due to space  
422 limitations. Since the time-of-flight of neutrons in the flight tube is inversely proportional to the square  
423 root of their energy, the ToF-NI technique can add a fourth dimension to conventional radiography.  
424 This can provide an additional mode of contrast as neutron attenuation is a function of its energy. We  
425 anticipate that if the neutron attenuation of active species and the supporting ions have distinct energy  
426 dependency profiles, we can separate the contribution of each species from the final radiograph.

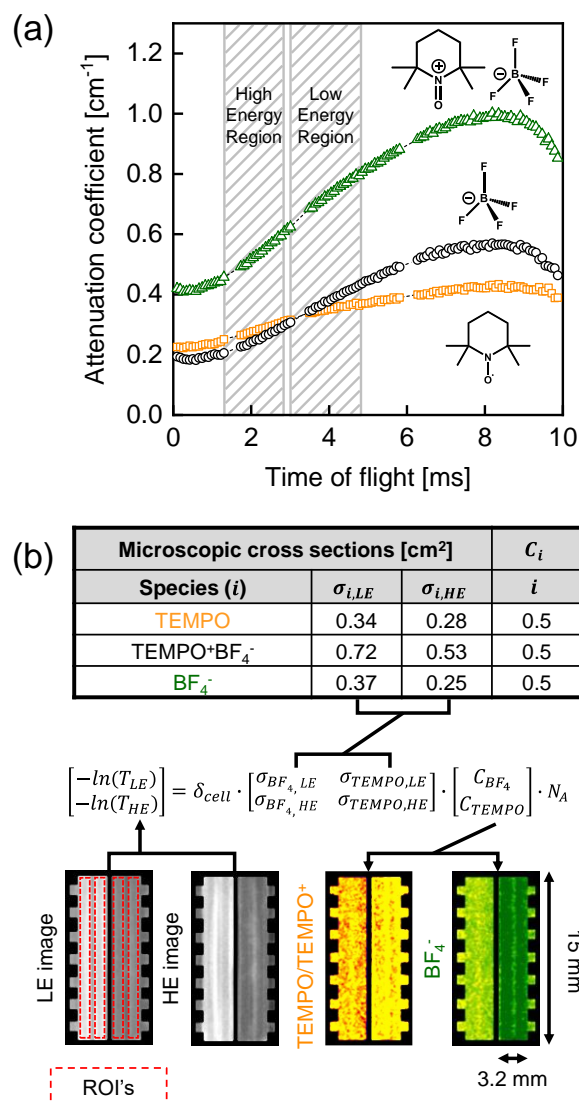
427

### 428 *Correlating attenuation with neutron energy*

429 The difference in relative neutron attenuation of materials enables tuning of the contrast between  
430 different species. To this end, we first performed calibration experiments with cuvettes, filled with  
431 0.5 M solutions of TEMPO and TEMPO<sup>+</sup>BF<sub>4</sub><sup>-</sup> in CD<sub>3</sub>CN. Figure 5a shows attenuation coefficients as a  
432 function of the time of flight, where the BF<sub>4</sub><sup>-</sup> attenuation coefficient is determined by subtracting the  
433 coefficient of TEMPO from TEMPO<sup>+</sup>BF<sub>4</sub><sup>-</sup>. Here, an increasing time-of-flight indicates a decreasing  
434 neutron energy. TEMPO<sup>+</sup>BF<sub>4</sub><sup>-</sup> reaches nearly twice the cross-section of TEMPO at higher energies,  
435 corroborating the previous observations made at the NEUTRA beamline (Figure 2) that TEMPO and  
436 BF<sub>4</sub><sup>-</sup> have similar microscopic cross-sections. The linearity of the concentration with neutron  
437 attenuation was already demonstrated in the NEUTRA beamline, thus we selected only one  
438 concentration (0.5 M) corresponding to the starting concentration in the flow cell experiments.

439 Using the matrix operation shown in equation (4) and in Figure 5b, the respective contributions  
440 of TEMPO and BF<sub>4</sub><sup>-</sup> from the total neutron attenuation can be separated. For this purpose, we need to  
441 define two regions within the spectrum, the HE and LE regions. The difference in attenuation  
442 coefficients between TEMPO and BF<sub>4</sub><sup>-</sup> varies as a function of neutron energy, this means that the slope  
443 of the graph in Figure 5a should be different between species, or in mathematical terms, the determinant  
444 of the microscopic cross-section matrix should not be zero. The neutron cross-sections of the species of  
445 interest are reported for HE and LE regions in Figure 5b. The values reported here correspond to the  
446 microscopic cross-section averaged over LE and HE ranges, described in the *Neutron Radiography*  
447 section. Although maximum contrast is achieved around 8 ms ToF, the LE region was moved towards  
448 higher energies to prevent the excessive neutron edge effects/scattering at interfaces between gaskets  
449 observed at lower energies. Finally, the matrix operation is applied pixel-wise to the greyscale  
450 transmission image to calculate the contribution of species, and a colour map is applied to designate the  
451 concentrations (Figure 5b). Achieving contrast between TEMPO and TEMPO<sup>+</sup> is still not possible, but  
452 because the movement of TEMPO<sup>+</sup> between compartments is mostly blocked by the anion exchange  
453 membrane, we can track the movement of BF<sub>4</sub><sup>-</sup> and TEMPO species separately during battery operation.

454



455

456 Figure 5: Energy selective imaging at the ICON beamline. (a) Energy dependency of the attenuation coefficient  
 457 of TEMPO, TEMPO+BF<sub>4</sub><sup>-</sup> and BF<sub>4</sub><sup>-</sup> obtained from the cuvette experiments (all 0.5 M), where the time-of-flight is  
 458 a function of the neutron energy. (b) Schematic representation of the main components in the image processing  
 459 sequence for the ICON beamline experiments including the table of microscopic cross-sections, the low and high  
 460 energy transmission images (grayscale) and deconvoluted active species and supporting ion images of the flow  
 461 cell, showing the flow fields, electrodes and membrane together with their dimensions. The transmission images  
 462 are processed using the given equation to extract the concentration maps (coloured images) of TEMPO/TEMPO+  
 463 and BF<sub>4</sub><sup>-</sup>.

464

#### 465 *Deconvoluting concentrations in a flow cell*

466 To demonstrate the potential of energy-selective and *operando* neutron imaging, a flow cell with  
 467 asymmetric concentrations (0.5 M TEMPO on the CE and 0.5 M TEMPO+BF<sub>4</sub><sup>-</sup> on the WE side, Figure  
 468 6a) was imaged. The electrolyte compositions are identical to the previous experiment that utilized BF<sub>4</sub><sup>-</sup>  
 469 as the counter-ion but we increased the electrode thickness to accommodate for the lower spatial  
 470 resolution. Previous experiments carried out on the NEUTRA beamline were set up with a tilted detector  
 471 to increase the spatial resolution of the images<sup>62</sup>, resulting in a pixel size of ~6 μm applied to study a  
 472 630 μm thick electrode. In the ICON experiments, the ToF detection system resulted in a larger pixel  
 473 size (~55 μm). Thus, to compensate for the discrepancies in spatial resolution, a thicker felt electrode  
 474 (3200 μm) was employed. The cell features stacked gaskets (incompressible PTFE and compressible

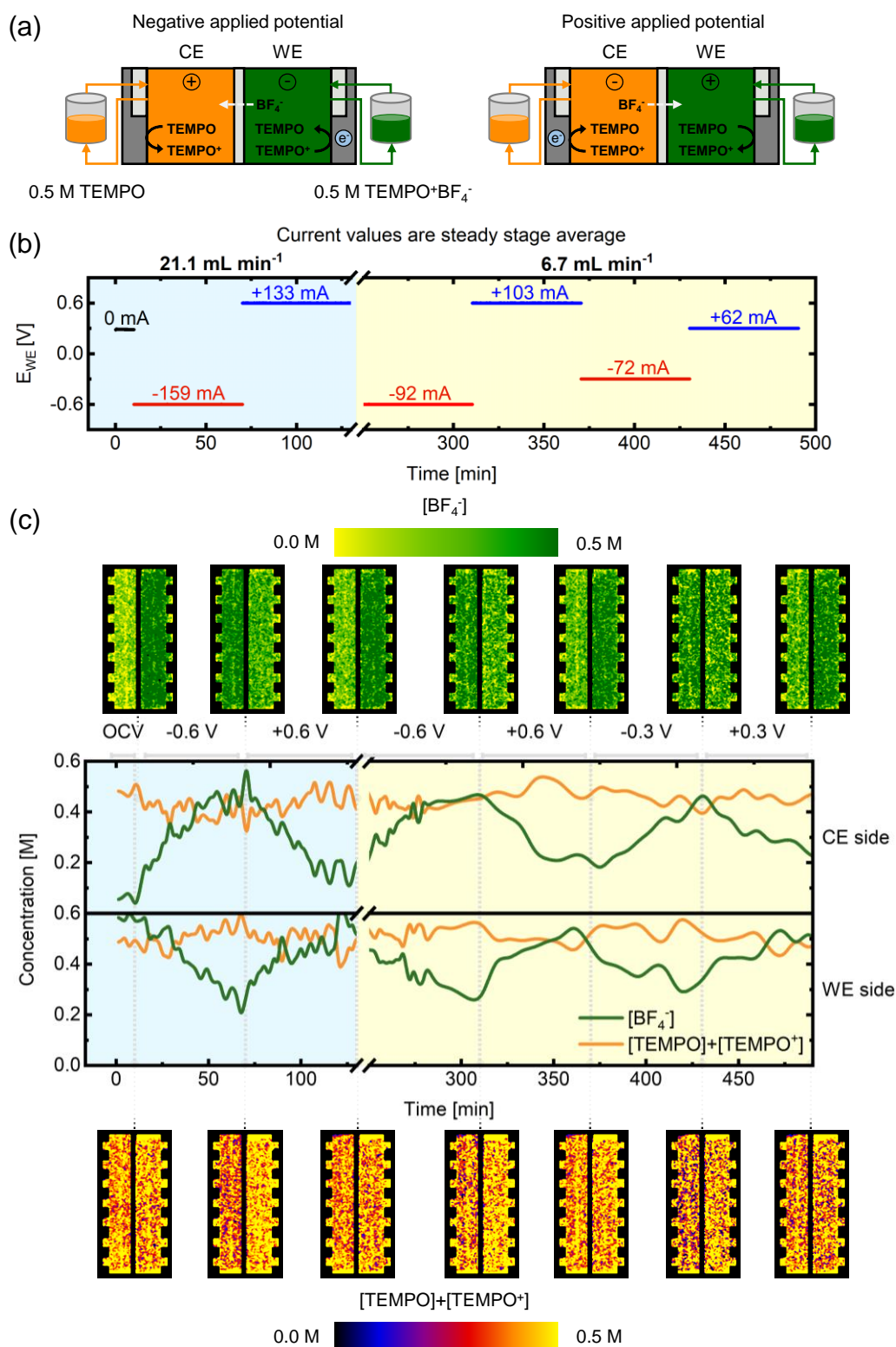
475 ePTFE) to enclose the thick felt electrode, where the interface around the ePTFE gaskets shows up in  
476 the LE image as low transmission regions (dark vertical lines in the grayscale image in Figure 5b) due  
477 to higher neutron edge effect/scattering. Nevertheless, the central regions of the incompressible gaskets  
478 (~1 mm) were large enough to define four regions of interest (Figure 5b) where the concentrations can  
479 be determined, and the reported concentrations are averaged over this volume for both compartments.  
480 To track the movement of species during the sequence, we opted for plotting the averaged  
481 concentrations over time (Figure 6c).

482 The experiment starts with an OCV period of 10 min, after which the cell was polarized at an  
483 electrolyte flow rate of 21.1 mL min<sup>-1</sup> followed by a reduced flow rate of 6.7 mL min<sup>-1</sup> (Figure 6b and  
484 Figure S7). From the deconvoluted concentration maps (Figure 6c), we confirm that BF<sub>4</sub><sup>-</sup> is the main  
485 charge carrier and that the membrane blocks the transport of TEMPO/TEMPO<sup>+</sup>, as their concentration  
486 remains relatively stable in both compartments throughout the entire electrochemical sequence. When  
487 a negative potential is applied, TEMPO<sup>+</sup> reduces to TEMPO in the WE compartment while the reverse  
488 reaction occurs in the CE (Figure 6c). Simultaneously, the concentration maps and averaged  
489 concentrations show that BF<sub>4</sub><sup>-</sup> moves through the membrane towards the CE to balance the positive  
490 charge of the generated TEMPO<sup>+</sup> species. Applying a positive potential to the WE reverses the direction  
491 of the migration flux of the BF<sub>4</sub><sup>-</sup> ions, and concentrations close to the initial state of the battery (i.e.,  
492 OCV) can be recovered. These results corroborate the observations from the experiments in NEUTRA  
493 as only minor concentration fluctuations were observed with PF<sub>6</sub><sup>-</sup> as supporting salt when an electric  
494 field is applied, whereas stark changes are detected when the supporting ion was changed to BF<sub>4</sub><sup>-</sup>.  
495 Moreover, the concentrations of the active species within the reactor area show larger variations for the  
496 highest flow rate (Figure 6c), induced by faster species conversion (i.e., higher current densities, Figure  
497 6b), and greater convective transport in the porous electrode. This brings the concentrations to extreme  
498 values due to the fast depletion of reactants in the electrolyte and promotes larger ionic currents. The  
499 difference in ionic current from the electrochemical data is correlated to the slope of the BF<sub>4</sub><sup>-</sup>  
500 concentration variations as a function of time.

501 From the capacity curves (Figure S8), we observe that after the first potential step (-0.6 V at  
502 21.1 mL min<sup>-1</sup>), 60% of the total capacity is consumed. In the next step, after an applied potential of  
503 +0.6 V, only 45% of the total capacity is recovered (due to the lower current at set time), resulting in  
504 15% underutilized capacity after a full polarization cycle because of the starting tank solutions (no BF<sub>4</sub><sup>-</sup>  
505 in the CE) as explained in the *transport of the counter-ion* section. At the lower flow rate (6.7 mL min<sup>-1</sup>)  
506 the capacity consumed at negative applied potentials is almost fully recovered at positive applied  
507 potentials, resulting in near symmetric current magnitudes. Interestingly, we find comparatively higher  
508 currents and capacity utilization with this reactor configuration in comparison with the reactor  
509 architecture used in the NEUTRA beamline (Figure 4), which can be correlated to the significant BF<sub>4</sub><sup>-</sup>  
510 concentration fluctuations. We attribute these differences to the use of a different electrode material (a  
511 thick felt vs. a stack of thin carbon papers) and lower compressive forces. The higher porosity, apparent  
512 permeability and internal surface area of the felt electrode can explain the higher current densities  
513 observed in this reactor configuration.

514 Here, we demonstrate the potential of the ToF-NI spectral technique to isolate and visualize  
515 concentration distributions of active and supporting species in redox flow cells. Compared to the use of  
516 conventional neutron radiography, the ToF method requires larger acquisition times and provides lower  
517 spatial resolution but enables detection of neutron energies necessary to deconvolute species  
518 concentrations.





519

520 Figure 6: *Operando* imaging of the active species transport in the ICON beamline with the neutron attenuating  
 521 BF<sub>4</sub><sup>-</sup> supporting ion. (a) Schematic representation of the non-aqueous cell designs during charge and discharge  
 522 mode, where the counter electrode (CE) corresponds to 0.5 M TEMPO and the working electrode (WE) to 0.5 M  
 523 TEMPO+BF<sub>4</sub><sup>-</sup>. (b) Electrochemical sequence over time showing the applied potential steps and measured averaged  
 524 current output at two inlet flow rates of 21.1 mL min<sup>-1</sup> and 6.7 mL min<sup>-1</sup>. (c) Deconvoluted active species  
 525 (TEMPO/TEMPO<sup>+</sup>) and BF<sub>4</sub><sup>-</sup> supporting ion concentration profiles. The averaged snapshots of the cell after image  
 526 processing and the concentration profiles over time are shown for various applied potential steps and flow rates:  
 527 OCV, -0.6 V and +0.6 V at 21.1 mL min<sup>-1</sup> and -0.6 V, -0.6 V, -0.3 V and +0.3 V at 6.7 mL min<sup>-1</sup>, from left to  
 528 right, where the OCV images are averaged over 5 images and the applied potentials averaged over 4 images.

17

## 529 **Practical application of the neutron imaging method**

530 This work demonstrates for the first time the use of neutron radiography to image concentrations of  
531 redox active species and supporting salts in *operando* electrochemical flow cells. By combining  
532 macroscopic electrochemical response with microscopic concentration distributions, neutron  
533 radiography can provide valuable insights into species motion within the reactor area, and this can be  
534 used to quantify mass transport mechanisms (migration, diffusion, convection) and phenomena  
535 affecting the performance of the battery in operation (e.g., electrolyte depletion, precipitation, physical  
536 failure in RFB stacks). These insights can directly be used to compare and select optimal cell  
537 components and to aid computational efforts. We anticipate that the use of molecular engineering to  
538 design redox molecular probes with controlled molecular structure, diffusivity and redox potential, can  
539 enable deconvolution of different oxidation states and degradation products. Although we focus on  
540 nonaqueous redox flow cells as a case study, we anticipate numerous applications of neutron imaging  
541 for quantifying concentration distributions in electrochemical cells and beyond. First, the resulting  
542 concentration maps can be used as experimental data to validate computational models that describe  
543 reactive mass transport. Here we used an electrolyte composed of a solvent, supporting electrolyte, and  
544 two redox active species - hence a complex, multicomponent system close to practical devices; but  
545 model experiments can be performed (e.g. with one or two analytes) to systematically deconvolute mass  
546 transport modes (e.g. diffusion, convection and migration) and their associated transport rates. Deeper  
547 fundamental understanding of reactive mass transport in electrochemical reactors and through  
548 membranes will assist designing advanced electrochemical cells. Second, the methodology enables  
549 identification of local maldistributions in concentration, which can assist in designing better flow field  
550 geometries and electrodes, as well membrane crossover, Donnan exclusion and salt precipitation within  
551 the electrochemical cell, which are deleterious to performance and lifetime in several flow battery  
552 chemistries (e.g. non-aqueous, all-vanadium and all-iron). Third, we anticipate that the method - and  
553 adaptations on the detection physics - will be instrumental in advancing hybrid redox flow batteries  
554 (e.g. all-iron, zinc-bromine), where there are phase change reactions (e.g. plating and stripping or  
555 hydrogen evolution) fundamentally limiting the performance of the system. Fourth, we anticipate that  
556 the technique can be applied to technical systems such as electrochemical stacks, where traditional  
557 neutron imaging was instrumental in advance fuel cell stacks through visualization of water  
558 distributions. Fifth, beyond redox flow batteries, the method can be applied to other (electro)chemical  
559 reactors where concentration profiles determine performance such as electrochemical separations, flow  
560 chemistry, and chemical reactor design. To further assist the design of neutron experiments to study  
561 electrochemical systems, Tables S1 and S2 summarize the neutron attenuation coefficient of commonly  
562 used materials for reactor manufacturing and redox species/supporting salts, respectively.

563 Finally, the use of molecularly engineered redox molecules acting as imaging probes might enable  
564 simultaneous visualization of concentration of multiple components (>2) when combined with energy-  
565 selective neutron imaging. Although the technique is still in its early stages, it displays considerable  
566 potential. We anticipate that ongoing advancements in neutron detectors and choppers will enable more  
567 sophisticated analyses of complex multicomponent systems using ToF-NI, offering enhanced spatial,  
568 temporal and energy resolution.

569

570

## 571 **Methods**

### 572 **Chemicals**

573 2,2,6,6-Tetramethylpiperidin-1-yl)oxyl (TEMPO, Sigma Aldrich, 98 %), nitrosonium tetrafluoroborate  
574 (NOBF<sub>4</sub>, Thermo Scientific, 98%), nitrosonium hexafluorophosphate (NOPF<sub>6</sub>, Thermo Scientific,  
575 95%), acetonitrile-d<sub>3</sub> (CD<sub>3</sub>CN, Zeochem AG, 99.8%D), acetonitrile (CH<sub>3</sub>CN, Sigma Aldrich, ≥ 99.9%)  
576 and potassium hexafluorophosphate (KPF<sub>6</sub>, Thermo Scientific, 99%) were used without further  
577 purification.

578

### 579 **Electrolyte preparation**

580 TEMPO was converted to its cation form 2,2,6,6-Tetramethyl-1-piperidinyloxy-oxo (TEMPO<sup>+</sup>) via  
581 chemical oxidation with nitrosonium salts in a nitrogen-filled glove box (MBraun, LABstar,  
582 O<sub>2</sub> < 1 ppm, H<sub>2</sub>O < 1 ppm). TEMPO (12.52 g) was dissolved in acetonitrile, where 1.1 molar  
583 equivalents of nitrosonium hexafluorophosphate (NOPF<sub>6</sub>, 15.42 g) or nitrosonium tetrafluoroborate  
584 (NOBF<sub>4</sub>, 10.29 g) dissolved in CH<sub>3</sub>CN were slowly added during 2 hours to prevent NO<sub>x</sub> build-up<sup>12</sup>.  
585 Then, a rotary evaporator (40 °C, gradual decrease from atmospheric pressure to vacuum) was used to  
586 remove the solvent and the TEMPO<sup>+</sup>PF<sub>6</sub><sup>-</sup> or TEMPO<sup>+</sup>BF<sub>4</sub><sup>-</sup> salts were recovered. The electrolytes were  
587 prepared by weighting the solid fractions (TEMPO, TEMPO<sup>+</sup>PF<sub>6</sub><sup>-</sup> and KPF<sub>6</sub> or TEMPO and  
588 TEMPO<sup>+</sup>BF<sub>4</sub><sup>-</sup>) prior to adding CD<sub>3</sub>CN until full dissolution. Finally, the volume was adjusted in a  
589 graduated flask to reach 20 mL of total volume. For the experiments using KPF<sub>6</sub> as supporting salt, a  
590 50% state-of-charge (SoC) solution was prepared for each side with two different total concentrations  
591 of TEMPO species (0.5 M at the working electrode (WE) side and 0.2 M at the counter electrode (CE)  
592 side) in 0.1 M KPF<sub>6</sub>. For the experiments using BF<sub>4</sub><sup>-</sup> ions, 0.5 M of TEMPO was used at the CE against  
593 0.5 M of TEMPO<sup>+</sup>BF<sub>4</sub><sup>-</sup> at the WE without additional added supporting salt to achieve high BF<sub>4</sub><sup>-</sup>  
594 concentrations without compromising the imaging process. For all imaging experiments, CD<sub>3</sub>CN was  
595 used as the solvent to reduce its contribution to the overall transmission as deuterium has a 10-fold  
596 lower total cross-section than hydrogen (at a neutron velocity of 2200 m s<sup>-1</sup>)<sup>60</sup>.

597

### 598 **Calibration experiments**

599 To quantify the concentration of the species within the electrochemical cells, the attenuation coefficients  
600 were determined with cuvette calibration measurements to correlate neutron transmission with solution  
601 concentration. The attenuation of the beam by the sample was calculated using the Lambert-Beer law  
602<sup>52</sup>, described as

$$T = e^{-\sigma_i n_i \delta} \quad (1)$$

603 where  $T$  is the transmitted intensity after correction for the attenuation of an empty cuvette [-],  $\sigma_i$  the  
604 conventional microscopic cross-section [m<sup>2</sup>],  $n_i$  the number density of species  $i$  [m<sup>-3</sup>], which is related  
605 to the concentration of species  $i$  by  $C_i = n_i/N_A$  [mol m<sup>-3</sup>], and  $\delta$  is the thickness of the cuvette [m].

606 For the NEUTRA and ICON beamline measurements, cuvettes (1 cm optical path) were filled  
607 with different electrolyte solutions to calibrate the attenuation of the neutron beam. For the NEUTRA  
608 beamline experiments, the reference cuvettes were filled with CD<sub>3</sub>CN (solvent) or 0.2 M KPF<sub>6</sub> in  
609 CD<sub>3</sub>CN (supporting electrolyte). To verify the linearity between concentration and neutron attenuation,  
610 solutions of TEMPO, TEMPO<sup>+</sup>PF<sub>6</sub><sup>-</sup> and TEMPO<sup>+</sup>BF<sub>4</sub><sup>-</sup> in CD<sub>3</sub>CN were measured at molarities of 0.1,

611 0.2, 0.3, 0.4 and 0.5 M. For the ICON beamline measurements, a single concentration point method  
612 was used as the linearity of the neutron attenuation as a function of the electrolyte concentration was  
613 obtained at the NEUTRA beamline. The reference cuvette was filled with CD<sub>3</sub>CN (solvent), and the  
614 single concentration points were measured for the solutions of interest of 0.5 M TEMPO and 0.5 M  
615 TEMPO<sup>+</sup>BF<sub>4</sub><sup>-</sup> in CD<sub>3</sub>CN.

616

## 617 **Flow cell parts**

618 Neutron imaging was performed using a laboratory-scale redox flow cell with minimal modifications  
619 for imaging. The flow diffusers were machined from polypropylene (McMaster-Carr) and the graphite  
620 parallel flow-by flow fields, also functioning as current collectors, featuring seven 1.6 cm long flow  
621 channels (0.5 mm depth and 1 mm width) were milled from 3.18 mm thick resin-impregnated graphite  
622 plates (G347B graphite, MWI, Inc.)<sup>12</sup>. All flow cells employed a Fumasep FAB-PK-130 (Fuel Cell  
623 Store, dry thickness 130 μm) anion exchange membrane. The electrodes had a geometric area of  
624 2.55 cm<sup>2</sup> enclosed within incompressible polytetrafluorethylene gaskets (ERIKS) and/or compressible  
625 polytetrafluorethylene (ePTFE, Gore®, 520 μm nominal) gaskets to improve the sealing of the flow  
626 cell (Figure 1c). For the experiments with the NEUTRA beamline, the flow cells contained three  
627 Sigracet 39AA electrodes (Fuel Cell Store, 280 μm nominal thickness, 89% porosity) per anode and  
628 cathode sides (six electrodes in total), sandwiching the membrane. The electrodes were compressed at  
629 25% compression by selecting an incompressible gasket thickness of 630 μm. For the experiments at  
630 the ICON beamline, one AvCarb G100 felt electrode (Fuel Cell Store, 3200 μm nominal thickness, 95%  
631 porosity) per side was used. Each side was sealed with two incompressible 1 mm gaskets and three  
632 compressible gaskets to reach a stack thickness of 3200 μm (measured from the neutron images). The  
633 reactor design was slightly modified, without impacting the transport phenomena within the cell, by  
634 grooving the current collectors and the gaskets around the active area (Figure 1c) to minimize the  
635 attenuation from cell parts with the neutron beam and thereby enhancing the sensitivity. After assembly,  
636 the cells were tightened with a torque-controlled wrench to 2 N m and the cell was mounted 1 – 3 mm  
637 in front of the neutron detector on a robotized platform. Peristaltic pumps (Cole-Parmer) were used to  
638 pump the electrolyte to the cells with rubber tubes (Masterflex LS-14 tubing) connected to two separate  
639 20 mL electrolyte tanks. No inert atmosphere was used in the beamlines, the tanks were sealed with a  
640 rubber septum, limiting the oxygen availability in the system even though the presence of oxygen was  
641 not detrimental to the electrolytes employed<sup>63</sup>. Two flow rates were employed, 15.1 mL min<sup>-1</sup> and  
642 5.6 mL min<sup>-1</sup> for the NEURA experiments and 21.1 mL min<sup>-1</sup> and 6.7 mL min<sup>-1</sup> for the ICON  
643 experiments, corresponding to superficial velocities within the flow field channels of 7.19 cm s<sup>-1</sup> and  
644 2.67 cm s<sup>-1</sup>, and 10.0 cm s<sup>-1</sup> and 3.19 cm s<sup>-1</sup>, respectively, as calculated using equation (2).

$$u = \frac{Q}{N_{ch} W_{ch} D_{ch}} \quad (2)$$

645 Where  $u$  is the fluid velocity [m s<sup>-1</sup>],  $Q$  is the electrolyte flow rate [m<sup>3</sup> s<sup>-1</sup>],  $N_{ch}$  is the number of inlet  
646 channels in the flow field configuration (7 channels) [-],  $W_{ch}$  is the width of the channel (1 x 10<sup>-3</sup> m)  
647 [m] and  $D_{ch}$  is the depth of the channel (5 x 10<sup>-4</sup> m) [m].

648

## 649 **Electrochemical protocols**

650 Electrochemical measurements were conducted with a Biologic VSP-3e potentiostat. For the  
651 experiments in the NEUTRA beamline, the open circuit voltage (OCV) of the cells was measured for

652 1 hour (for the experiments with the KPF<sub>6</sub> supporting salt) or 2 hours (for the experiments with the BF<sub>4</sub><sup>-</sup>  
653 supporting ion) at 15.1 mL min<sup>-1</sup> after the cell was filled with the electrolyte to monitor the diffusion of  
654 species between the WE and CE. Thereafter, the cell was successively held at -0.3 V, +0.3 V, -0.6 V  
655 and +0.6 V for 20 min at each potential step. Then the flow rate was decreased to 5.6 mL min<sup>-1</sup> and the  
656 same potential protocol was applied. The entire electrochemical protocol was ca. 220 min and neutron  
657 radiographs were collected during the entire duration of the experiment. All measurements were  
658 performed in the same cell by filling and emptying the cell with the different solutions with a rinsing  
659 step in between with 0.5 M TEMPO dissolved in CD<sub>3</sub>CN.

660 For the experiments at the ICON beamline, the OCV of the cells was measured for 10 min at  
661 21.1 mL min<sup>-1</sup> after which the cell was successively held at -0.6 V and +0.6 V for 1 hour at each  
662 potential step. Then the flow rate was decreased to 1.7 mL min<sup>-1</sup> and the same potential protocol was  
663 applied. Then the flow rate was increased to 6.7 mL min<sup>-1</sup> and the cell was successively held at -0.6 V,  
664 +0.6 V, -0.3 V and +0.3 V for 1 hour at each potential step. Finally, the flowrate was switched to 96  
665 mL min<sup>-1</sup> and the cell was held at -0.6 V and +0.6 V for 1 hour at each potential step. The entire  
666 electrochemical protocol took around 720 min to complete. For this study, only two flowrates  
667 (21.1 mL min<sup>-1</sup> and 6.7 mL min<sup>-1</sup>) were analysed in detail. The electrolyte was not refreshed during the  
668 entire experiment.

669

## 670 Neutron radiography

671 Neutron radiography experiments were performed at the NEUTRA thermal neutron and ICON cold  
672 neutron imaging beamlines at the Spallation Neutron Source (SINQ) facility of the Paul Scherrer  
673 Institute, Switzerland. In the SINQ facility, the neutrons were ejected from a lead spallation target that  
674 was hit with a proton beam at 590 MeV energy with 1.5 mA proton current. The ejected neutrons were  
675 moderated by heavy water and reached thermal velocities (with a mean energy of 25 meV)<sup>64</sup>. For the  
676 NEUTRA beamline, attenuated neutrons were captured by a scintillator screen (10 μm thick,  
677 Gd<sub>2</sub>O<sub>2</sub>S:Tb) and converted to visible light, which is subsequently captured by the charge-coupled device  
678 camera detector at an exposure time of 30 s. A tilted detector setup was used at the NEUTRA beamline  
679 that enables stretching in the horizontal transverse direction (with respect to the beam trajectory), see  
680 Figure 1a, meaning that the membrane-electrode assembly can be imaged with higher spatial resolution  
681<sup>65</sup>. The resulting pixel size in the direction across the membrane was 6 μm and the effective resolution,  
682 taking into account the blurring intrinsic to the detector and due to the beam divergence, was  
683 approximately 20 μm.

684 For the ICON beamline, neutrons were further moderated with a liquid deuterium (D<sub>2</sub>) tank  
685 held at 25 K, decreasing the velocity of neutrons to the cold spectrum (mean energy of 8.53 meV)<sup>66</sup>.  
686 Similarly, the neutron beam passes through a series of collimators, beam limiters and shutters but before  
687 interacting with the sample the neutron beam passes through a mechanical chopper allowing ToF-NI.  
688 The chopper creates a pulsed neutron beam whereas the travelled length of the neutrons through the  
689 flight tubes allows dispersion of the pulse based on the velocity of the constituent neutrons. The chopper  
690 rotated at a speed of 22 Hz with 4 regularly spaced openings, resulting in a pulse repetition frequency  
691 of 88 Hz. The angle of each opening was 18°, resulting in a 20% duty cycle. The path length between  
692 the chopper and the detector was 5.5 m. Finally, the neutrons were detected (exposure time 120 s per  
693 acquisition) at a microchannel plate detector having a fixed pixel size of 55 μm, an effective resolution  
694 of approximately 150 μm and a field of view of 28 x 28 mm<sup>2</sup> (512 x 512 pixel<sup>2</sup> images). In this way,  
695 the ToF-NI spectral technique in the ICON beamline allows spectral imaging of the sample, adding a  
696 new mode of contrast to conventional neutron radiography<sup>52,67</sup>. Per single ToF cycle of 11.36 ms (88

697 Hz of chopper disk rotation), 109 raw transmission images were taken. In total, 10560 cycles were  
698 completed over a total acquisition time of 2 min from high to low neutron energies, and the resulting  
699 images were binned over 109 images. Each of the recorded images represents the sum of the  
700 corresponding 10560 images acquired at one point of the cycle. Because of the relatively broad width  
701 of the neutron pulse (2.73 ms), each frame represents a blend of several neutron energies. This is not  
702 detrimental to the distinction of hydrogen and boron atoms, as the variation of neutron attenuation as a  
703 function of energy does not exhibit any stark feature. Frames were averaged from frames 16 - 30 to  
704 construct high energy (HE) images and from frames 31 – 50 to construct low energy (LE) images. The  
705 HE neutron energy ranges from 77.3 – 17.0 meV with a mean at 28.3 meV, and the LE neutron energy  
706 ranges from 16.0 – 5.7 meV with a mean at 8.5 meV. Since the image acquisition time is short compared  
707 to the experimental time (360 min), it can be assumed that HE and LE images are taken at the same cell  
708 conditions. It was not possible to utilize the lower energy neutron spectra (frames 50 - 109) due to  
709 significant beam scattering and diffraction caused by the compression gaskets at their interface.

710

## 711 **Image processing**

712 Image processing by mathematical calculations was performed to extract the sample information as  
713 described in the image processing pipeline (Figure 7). Neutron radiography was performed in a  
714 subtractive manner, meaning that the transmission data from the samples were corrected for undesirable  
715 contributions, such as scattering and absorption of the cell components, detector background effects  
716 and beam instabilities. The image processing sequence for NEUTRA is shown in detail in Figure S9 in  
717 the Supporting Information and a description of the individual steps is given below:

- 718 1 Dark current correction: Dark current images were taken with closed beam shutters and  
719 optical shutters of the camera to correct for the electronic bias within the camera circuitry.
- 720 2 White spot filtering: White spots, resulting from gamma rays or other type of radiation hitting  
721 the camera chip, were filtered from the images by an outlier filtering approach, where pixels  
722 that deviate largely from their surroundings are replaced by a median value of these  
723 surroundings.
- 724 3 Gaussian filtering: This filtering step reduces the statistical noise that is caused by high  
725 frequency photons hitting the camera.
- 726 4 Open beam correction: Open beam images were taken without the electrochemical cell in the  
727 beamline to account for the spatial variation in beam intensity by dividing the images by the  
728 averaged open beam image.
- 729 5 Registration: The registration step accounts for the physical movement of the cell during the  
730 experiments due to thermal dilation or relaxation of the cell body. All images were registered  
731 to a “reference” (the electrochemical cell filled with a 0.2 M KPF6 in CD<sub>3</sub>CN solution) by  
732 applying a correlation algorithm on certain cell regions to find the optimum geometrical  
733 transformation.
- 734 6 Intensity correction: This step corrects for beam intensity variations throughout the  
735 experiment duration by defining a non-changing area in the cell that corresponds to an area  
736 with no known changes (e.g., a part in the cell body). Gain and offset were applied to the  
737 images to match the intensity within the non-changing area region to that of the reference  
738 image.
- 739 7 Scattered background correction: To account for the scattered background resulting from  
740 neutrons that are scattered by the cell and the detector, a scattered background image was  
741 obtained by interpolating the intensity values between so-called “black bodies”. These “black  
742 bodies” are boron rods located in a grid fashion on a steel plate and are placed in front of the

743 sample. The black body images were processed with all the processing steps described above,  
 744 but the averaging on the images was performed before the registration and intensity  
 745 correction steps to reduce the amount of noise. The final scattered background image was  
 746 subtracted from the images to account for the scattered background.

747 8 Referencing: In this step, the images were divided pixel-wise by the reference image that  
 748 underwent all the processing steps above except registration and intensity correction, as these  
 749 steps were based on the position and average intensity of the non-changing area in the  
 750 reference image.

751 9 Positioning: Images were rotated so that the flow channels were vertically aligned.

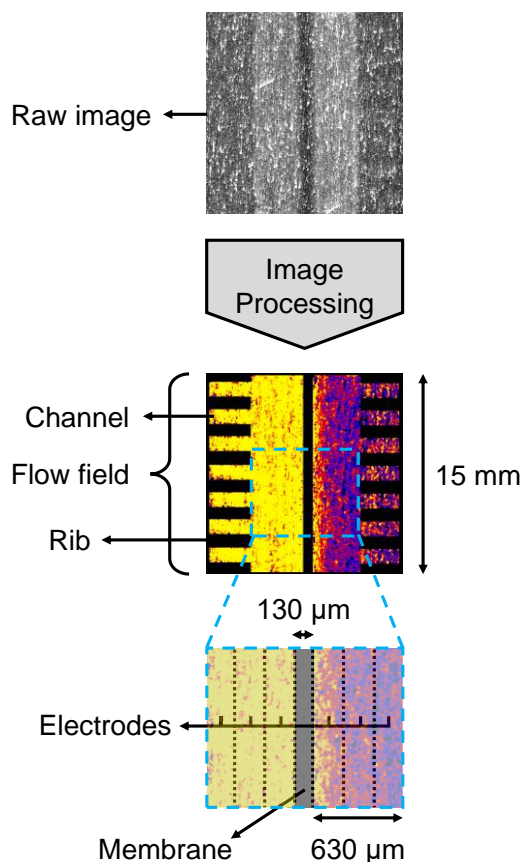
752 10 Cell mask: A cell mask was used to remove all the other parts of the image to obtain an image  
 753 with only the region of interest, i.e., the membrane, electrode and flow channels.

754 11 Concentration calculation: The final transmission values obtained per-pixel were correlated  
 755 to the species concentration via the Lambert-Beer law (equation (1)) as described in the  
 756 Calibration experiments section, where the microscopic cross-sections were obtained from  
 757 the cuvette measurements and with  $\delta_{cell}$  the electrolyte thickness within the cell [m] which  
 758 is a function of the cell geometry, electrode porosity and compression according to:

$$\delta_{cell} = L_e \varepsilon_e \quad (3)$$

759 where  $L_e$  is the electrode width ( $1.7 \times 10^{-2}$  m) [m] and  $\varepsilon_e$  the electrode porosity at the applied  
 760 compression (which is 85.3% for three stacked SGL 39AA paper electrodes) [-].

761



762

763 Figure 7: Schematic representation of the image processing for the NEUTRA beamline experiments including the  
 764 raw and final image. The final image shows the flow fields, electrodes and membrane together with their  
 765 dimensions.

766 Due to the nature of the microchannel plate detector and the ToF-NI method used at the ICON  
767 beamline, the image processing sequence is slightly different than for NEUTRA. Overlap correction  
768 was performed on all images before the image processing steps due to the characteristics of the  
769 microchannel plate detector<sup>68</sup>. The image processing sequence to obtain transmission images is listed  
770 below:

- 771 1 Outlier removal: Dead pixels (or zero pixels) were removed from the images by averaging  
772 pixels around them and setting the new value to it.
- 773 2 Scrubbing correction: To avoid bias due to the detector efficiency over time, open beam images  
774 were used to correct for any change of contrast not related to the experiment but to the detector.  
775 The function interpolates between open beam images and sets a weight to correct them.
- 776 3 Scattered background correction: Although microchannel plate detectors have less contribution  
777 of scattered background due to the transmission of neutrons parallel to the beam-axis, we still  
778 performed scattered background correction to improve accuracy<sup>69</sup>. For ICON, the black bodies  
779 were strips of Boral (2.5 mm width) in a steel plate.
- 780 4 Registration
- 781 5 Binning: Selection and merging of the HE and LE images based on the energy-dependent  
782 calibration curves of the neutron attenuation of the different species of interest and accounting  
783 for the edge scattering effects.
- 784 6 Intensity correction
- 785 7 Referencing
- 786 8 Positioning
- 787 9 Cell mask
- 788 10 Concentration calculation: The concentration of each species was obtained by solving a system  
789 of equations for the transmission of each region of interest (see Figure 5b), where the LE and  
790 HE microscopic cross-sections of the species can be correlated to the concentration distribution  
791 of the *operando* images via the following operation

$$\begin{bmatrix} -\ln(T_{LE}) \\ -\ln(T_{HE}) \end{bmatrix} = \delta_{cell} \cdot \begin{bmatrix} \sigma_{BF_4,LE} & \sigma_{TEMPO,LE} \\ \sigma_{BF_4,HE} & \sigma_{TEMPO,HE} \end{bmatrix} \cdot \begin{bmatrix} C_{BF_4} \\ C_{TEMPO} \end{bmatrix} \cdot N_A \quad (4)$$

792 where  $T_{LE}$  and  $T_{HE}$  are the low energy and high energy transmission values per-pixel of the  
793 *operando* experiment images [-],  $\sigma_{i,LE}$  and  $\sigma_{i,HE}$  are the low energy and high energy  
794 microscopic cross-sections [ $m^2$ ], and  $C_i$  is the pixel-wise concentrations of the species  $i$  [ $mol\ m^{-3}$ ],  
795 which are TEMPO or  $BF_4^-$ . The electrolyte thickness,  $\delta_{cell}$ , is defined according to equation  
796 (3), which for the felt electrode used in the ICON experiments is around 1.62 cm.

797



## 798 **Data Availability**

799 The data presented in this study can be provided by the corresponding author upon reasonable request.  
800 Source data files are provided with this paper. The data in the figures are provided as Source Data Files  
801 and can be obtained in the ChemRxiv. Supplementary data can be found in the Supporting Information  
802 and two videos are available online.

803

## 804 **Code Availability**

805 The code used in this study can be provided by the corresponding author upon reasonable request.

806

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981 R.R.J., M.v.d.H. and E.B.B. contributed equally to this work. R.R.J. contributed to the  
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## 997 **Competing Interests**

998 The authors declare no conflict of interest.

