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2 **Mitigation of Cyanobacterial Harmful Algal Blooms by Electrochemical Ozonation: From**
3 **Bench-scale Studies to Field Applications**

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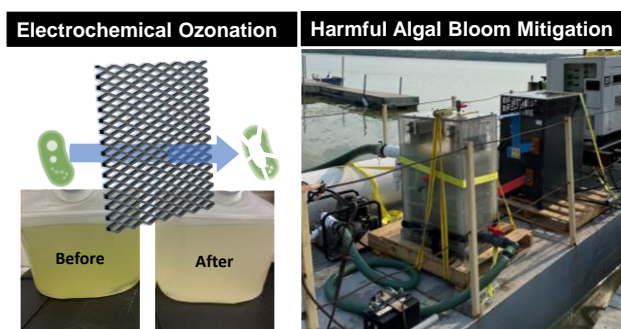
19 **ABSTRACT**

20 Cyanobacterial harmful algal blooms (HABs) are an emerging threat to ecosystems, drinking
21 water safety, and the recreational industry. As an environmental challenge intertwined with
22 climate change and excessive nutrient discharge, HAB events occur more frequently and
23 irregularly. This dilemma calls for fast-response treatment strategies. This study developed an
24 electrochemical ozonation (ECO) process, which uses a Ni-Sb-SnO₂ anode to produce locally
25 concentrated ozone (O₃) on electrodes to inactivate cyanobacteria and destroy microcystins
26 within minutes. More importantly, the proof-of-concept was evolved into a full-scale boat-mounted
27 completely mixed flow reactor for the treatment of HAB-impacted lake water at a treatment
28 capacity of 544 m³/d and energy consumption of < 1 Wh/L. Both lab-scale and full-scale
29 investigations show that the byproducts (e.g., chlorate, bromate, trihalomethanes, and haloacetic
30 acids) in the ECO-treated lake water were below the regulatory limits for drinking water. The whole
31 effluent toxicity tests suggest that ECO treatment at 10 mA/cm² posed certain chronic toxicity to
32 the model invertebrate (*Ceriodaphnia dubia*). However, the treatment at 7 mA/cm² (identified as
33 the optimum condition) did not increase toxicity to model invertebrate and fish (*Pimephales
34 promelas*) species. This study is a successful example of leveraging fundamental innovations in
35 electrocatalysis to solve real-world problems.

36

37 **Keywords:** Harmful algal bloom, Microcystin, Ozone, Electrocatalysis, Tin oxide.

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42 INTRODUCTION

43 Widespread cyanobacterial harmful algal blooms (HABs) have become an emerging threat to
44 ecosystems, recreational use of lakes, and drinking water supplies. Due to increased nutrient
45 discharge and global climate change, the occurrence of HABs is expected to be more frequent.^{1,2}
46 In the United States, the cyanobacterial HAB occurrence was 7 days per year per waterbody in
47 2017, and the frequency was projected to be 18-39 days per year per water body by 2090.¹ A
48 recent study based on a 45-year record (1970 – 2015) revealed that the frequency of HABs
49 occurring along the Chinese coast has increased by about 40% per decade.³ The surge
50 production of hepatotoxic, cytotoxic, and neurotoxic microcystins can be expected in the HAB
51 events.⁴

52 Chemical oxidation (using chlorine, permanganate, H₂O₂, etc.)⁵⁻⁷ and advanced oxidation
53 processes (UV/H₂O₂ and UV/chlorine)⁸ are promising for HAB mitigation at drinking water
54 treatment works. However, they are less practical for on-site lake remediation due to the uncertain
55 efficacy limited by hydrological conditions and concerns about the environmental impact of
56 chemicals.

57 There is a critical need for *in situ* treatment technology that can proactively curb HABs at the
58 early stage and precisely eliminate algae plumes in the impacted high-value areas (lake shores,
59 public beaches, etc.). Since the bloom is usually concentrated at the top 0–1 m of the water
60 column,^{9,10} we envision that mobile devices with pump-and-treat functionality could realize the
61 precise treatment of plume (instead of the whole water body) for maximum cost-effectiveness.
62 Electrochemical oxidation (EO)-based technologies, with the advantages of small footprint, high
63 efficiency, and chemical-free operation, could be the best fit. Previously, The EO treatment of
64 algae and microcystins equipped with anodes made of boron-doped diamond,¹¹ metal oxide
65 (RuO₂ and IrO₂),^{12,13} and graphite¹⁴ was investigated at the lab scale. We advanced the EO
66 process by developing Ti₄O₇ filter anodes with pore sizes of 24–53 μm for the treatment of HAB-
67 impacted lake water at both lab and full scales.¹⁵ Similar to other reported anode materials, the
68 Ti₄O₇ filter anodes oxidize chloride in lake water to chlorine to inactivate algae cells and oxidize
69 cyanotoxins. The flow-through operation (by pumping water out of the filter anodes immersed in
70 the plume) significantly promoted convective mass transfer and thereby achieved higher
71 treatment performance than homogenous chlorination. Though the scaled-up demonstration at a
72 capacity of 110 m³/d was successful, two limitations were identified: (1) the chlorine yield, which
73 is associated with treatment performance, depends on chloride concentrations in lake water. (2)
74 The micropores of filter anode create large pressure drops ahead of the pump. Thus the treatment

75 capacity is limited by the pump suction head, which is significantly smaller than the lift head. The
76 pumps were also vulnerable to damage by cavitation.

77 This study aims to address these engineering challenges. First, the anode configuration was
78 transformed from microporous cartridge filters to an array of mesh electrodes with rhombus
79 0.5×0.8 mm openings. Correspondingly, as will be discussed below, the treatment operation was
80 changed to pumping water through the mesh electrode array in a boat-mount reactor. Second,
81 the electrocatalysts were shifted from Ti₄O₇ to nickel-doped antimony tin oxide (**NATO**: Ni-Sb-
82 SnO₂). The incentive is that NATO has unique reactivity toward ozone (O₃) evolution by water
83 oxidation ($3\text{H}_2\text{O} \rightarrow \text{O}_3(\text{g}) + 6\text{H}^+ + 6\text{e}^-$, $E^0=1.51 \text{ V}_{\text{RHE}}$).^{16,17} Notably, NATO was deployed in
84 organic degradation and disinfection.^{18,19} However, its application in HAB mitigation was not
85 explored. Because water is the only required precursor for ozone production, the shift of reactive
86 species from free chlorine to O₃ enables the treatment performance to be independent of chloride
87 concentration. In addition to process innovation, this study provides comprehensive evaluations
88 of the environmental implications, covering process impacts on byproduct formation and effluent
89 toxicity on an invertebrate and fish.

90

91 **METHODS**

92 **Cyanobacteria Cultivation and Materials.** Two cyanobacteria strains, *Synechococcus* sp. and
93 *Microcystis aeruginosa* (UTEX Culture Collection of Algae), were incubated in a shaker incubator
94 (Innova S44i, Eppendorf) with 10-30% photosynthetic light following our previous study.¹⁵ The
95 bench-scale tests involved mesh-type (0.5 × 0.8 mm rhombus openings) anode with either NATO
96 or antimony tin oxide (ATO: Sb-SnO₂) coatings. The NATO and ATO anodes were prepared by
97 dip-coating titanium mesh into sol-gel solutions of metal-citrate complexes, followed by
98 calcination.¹⁸ The NATO anode comprises NATO out-layer coating (1 mg/cm² projected area) and
99 ATO base coating (1 mg/cm² projected area) on Ti mesh, while the ATO anode only contains the
100 ATO base coating. The dual-layer design of the NATO anode bestows the electrode with superior
101 durability and O₃ yield. Details were provided in our previous publication.¹⁸ The full-scale electrode
102 array contains 20 pieces of mesh NATO anode (50 × 50 cm; 0.5 × 0.8 mm rhombus openings)
103 sandwiched by 21 pieces of perforated stainless steel sheet cathode (50 × 50 cm; round opening
104 at Ø 3 mm) at an interspace of 0.5 cm. The full-scale mesh NATO anodes were manufactured by
105 Square One Coating Systems following the same sol-gel coating + calcination procedure
106 described above.

107 **Testing Conditions.** In bench-scale investigations, a mesh anode (NATO or ATO; 5 × 5 cm) was
108 coupled with a stainless steel cathode (5 × 5 cm) at a distance of 0.5 cm and operated at current
109 densities of 7 or 10 mA/cm². All tests were conducted in 95 mL phosphate buffer solution (**PBS**;
110 pH = 7.7, conductivity = 329 μS/cm) or lake water collected from Lake Neatahwanta, NY. The
111 synthetic water or lake water was spiked with cyanobacteria culture to reach an initial
112 concentration of 100 μg/L as chlorophyll-a (Chl-a).

113 **Analytical Approaches.** Chl-a was measured by a fluorometer, and Microcystin LR (MC-LR) was
114 quantified by liquid chromatography coupled with a quadrupole mass spectrometer (LC-MS/MS;
115 Thermo Scientific, Vanquish-TSQ ALTIS). Details were described previously.¹⁵ In the tests
116 involving *Microcystis aeruginosa* (an MC-LR toxin-producing strain), MC-LR detected in the bulk
117 solution was reported as extracellular MC-LR. Intracellular MC-LR was extracted by acetone after
118 retaining cells by filtering the water sample through a polycarbonate membrane with a pore size
119 of 0.2 μm.

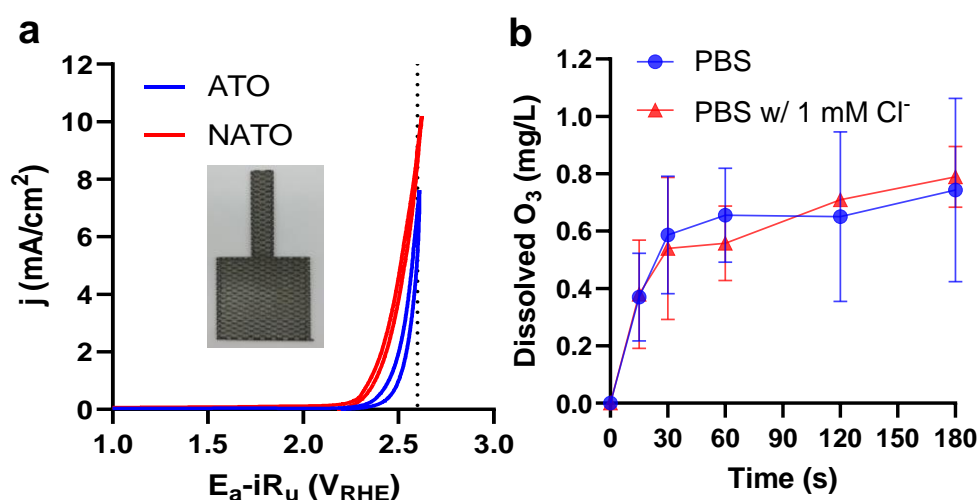
120 Free chlorine was measured by a portable Hach DR900 colorimeter (HACH, CO) using a
121 DPD (N,N-diethyl-p-phenylenediamine) reagent. Dissolved O₃ concentrations were determined
122 by the indigo method on a NanoDrop OneC spectrophotometer (Thermo Fisher Scientific).²⁰
123 When measuring electrochemical O₃ evolution in the presence of chloride, the anodically
124 generated free chlorine was masked by 1 M malonic acid.¹⁸ Trihalomethanes (THMs) and
125 haloacetic acids (HAAs) were measured by gas chromatography/mass spectrometry (GC/MS)
126 following EPA Method 624.1. Inorganic anions were analyzed by ion chromatography (Thermo
127 Dionex Integrion HPIC).²¹ The whole effluent toxicity tests on *Pimephales promelas* and
128 *Ceriodaphnia dubia* were independently conducted by AquaTOX Research, Inc., following EPA
129 Methods 2000.0 and 2002.0 respectively (EPA-821-R-02-013).

130

131 RESULTS AND DISCUSSION

132 Electrode Characterization.

133 **Figure 1a** shows the cyclic voltammetric (CV) profiles of NATO and ATO. We adopted the
134 Ni dopant level of 1 at.% (Ni/(Ni+Sb+Sn)) as optimized in previous studies.^{16,22} The Ni doping
135 shifts the CV hysteresis toward lower potentials. The rise of the current response at lower
136 potentials can be attributed to the enhanced evolution of oxygen and O₃.¹⁷ In the lake water
137 treatment, the mesh electrodes were operated at current densities between 6-10 mA/cm²
138 projected area, which leads to anodic potentials around 2.6-2.7 V_{RHE} (**Figure 1a**), surpassing the
139 thermodynamic criteria for •OH radical evolution (•OH/H₂O: 2.7 V_{NHE}).²³



140
141 **Figure 1.** (a) Cyclic voltammetry of ATO and NATO anodes measured in 100 mM NaClO₄
142 electrolyte. (b) Dissolved O₃ produced by NATO at 10 mA/cm² in PBS electrolyte with and without
143 1 mM Cl⁻. Data in Figure 1b are presented as the mean value of triplicate ± standard deviation.

144 Electrolysis using ATO could not generate O₃. In contrast, the NATO generated dissolved
145 O₃ in PBS electrolyte in the presence or absence of Cl⁻ (**Figure 1b**). The evolution rate and current
146 efficiency in PBS electrolyte at 10 mA/cm² are 0.0078 mmol/m²/s and 4.5%, respectively. Chlorine
147 evolution capability was tested in PBS electrolyte amended with 1 mM Cl⁻, a typical Cl⁻
148 concentration found in lake waters treated in this study (**Table S1**). NATO and ATO demonstrated
149 comparable chlorine evolution rates of 0.022 and 0.024 mmol/m²/s, respectively, at 10 mA/cm²
150 (**Figure S1**).

151 The electrochemical reactive surface area (ECSA) of the NATO mesh anode was
152 measured by the double-layer capacitance method.²⁴ The capacitance is measured as 16.64 mF,

153 corresponding to an ECSA of 443.6 cm² (Figure S2). The estimated ECSA is 8.9 times larger than
154 the projected surface area (50 cm²).

155 **Bench-scale Investigation**

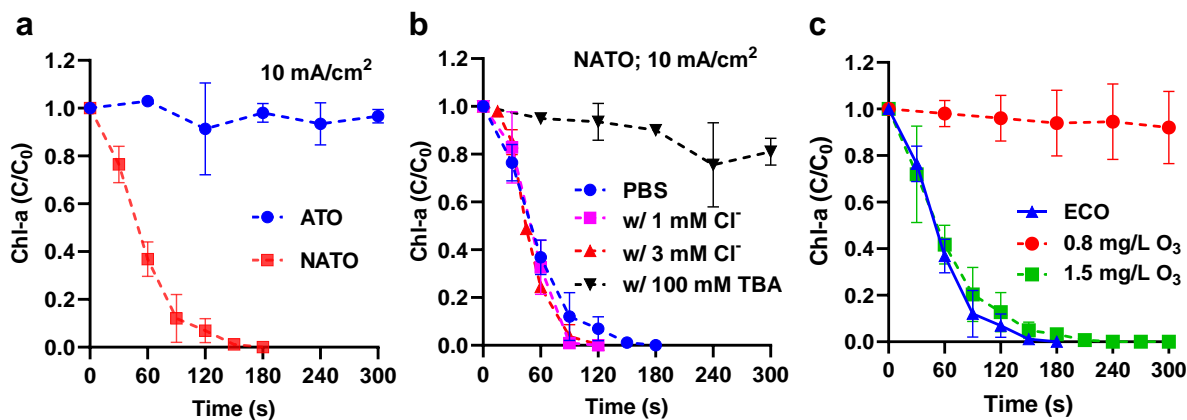
156 The treatment performance of mesh-type ATO and NATO was evaluated based on the
157 inactivation efficiency of *Synechococcus* and *Microcystis*, two strains of common cyanobacteria
158 in algal blooms.^{25,26} Because Chl-a cannot maintain its structural integrity and light-adsorbing
159 properties after cell damage, it was used as an indicator to quantify the viable cell concentration
160 as a generally accepted approach.^{27–30} *Synechococcus* does not produce toxins. It was used in
161 most bench-scale tests to study the kinetics of Chl-a degradation (i.e., cyanobacteria inactivation)
162 without the interference of MC-LR. The optimized reaction conditions were then applied to treat
163 electrolyte spiked with *Microcystis aeruginosa* culture with co-existing MC-LR.

164 The tests of inactivation of *Synechococcus* were first performed in PBS electrolyte at a
165 current density of 10 mA/cm². Note that both ATO and NATO have sufficiently high anodic
166 potentials of ~2.7 V_{RHE} to produce •OH (Figure 1a). If •OH is responsible for algae inactivation,
167 ATO and NATO should exhibit similar performance. In contrast, it was found that ATO showed
168 negligible reactivity in Chl-a removal, and NATO significantly outperformed ATO (Figure 2a).
169 These results exclude the contribution of •OH-mediated oxidation and direct oxidation to cell
170 inactivation, leaving electrochemically generated O₃ the only possible oxidant. The O₃ evolution
171 on NATO involves the combination of surface-bound •OH with O₂ to form •HO₃, then O₃.^{17,31}
172 Therefore, the addition of 100 mM tert-butyl alcohol (TBA) to scavenge •OH halted the O₃
173 production (below detection limit), and thereby retarded Chl-a removal (Figure 2b).

174 In the presence of chloride, NATO could produce free chlorine and O₃. However, the
175 addition of up to 3 mM Cl⁻ (a high record in lakes of the United States³²) did not accelerate Chl-a
176 removal (Figure 2b). Previously, we determined the pseudo-first-order rate constant of Chl-a in
177 *Synechococcus* and free chlorine as 0.027 L/(mg min).³³ With the presence of 1 mM Cl⁻, the time-
178 average free chlorine concentrations produced by NATO at 10 mA/cm² within 120 s electrolysis
179 should be ~7 mg/L (Figure S1). If free chlorine is responsible for Chl-a removal, the half-life of the
180 reaction should be 220 s instead of the 48 s shown in Figure 2b.

181 The above investigations have excluded direct oxidation, •OH-mediated oxidation, and
182 free chlorine-mediated oxidation as the dominant cyanobacteria inactivation (i.e., Chl-a removal)
183 mechanisms, leaving electrochemical ozonation (ECO) the only pathway. The efficacy of ECO
184 was compared with homogeneous ozonation (Figure 2c). The homogeneous ozonation at 1.5
185 mg/L O₃ achieved Chl-a removal kinetics similar to ECO. In contrast, ECO generated 0.48 mg/L

186 O₃ at 120 s, corresponding to a time-average [O₃] of 0.24 mg/L. ECO produced less bulk [O₃]
187 (0.24 vs. 1.5 mg/L) to realize faster Chl-a removal kinetics than homogeneous ozonation. In other
188 words, the cyanobacteria inactivation could be attributed to the locally concentrated O₃ produced
189 at the anode surface, and the surficial O₃ concentration is equivalent to ~1.5 mg/L.
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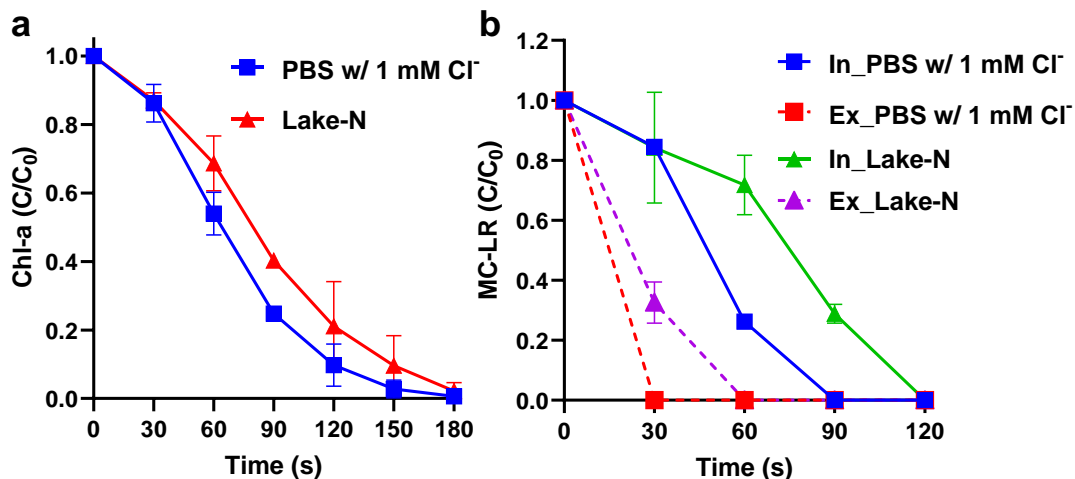


191
192 **Figure 2.** Inactivation of *Synechococcus* by electrolysis using (a) ATO and NATO anodes in PBS
193 electrolyte and (b) NATO anode in different electrolytes. (c) Comparison of Chl-a destruction by
194 ECO and homogeneous ozonation with various initial O₃ dosages (0.8 and 1.5 mg/L dissolved
195 O₃). ECO tests were conducted in PBS at 10 mA/cm². Data are presented as the mean value of
196 triplicate ± standard deviation.

197 After elucidating the reaction mechanism, the treatment tests were operated in more field-
198 like scenarios. In most of the field tests, the ECO full-scale system was operated at 7 mA/cm²
199 (see discussion below). At 7 mA/cm², ECO was effective in the removal of Chl-a in electrolytes
200 with and without Cl⁻ (Figure S3). As shown in Figure 3a, the ECO process is equally efficient for
201 the inactivation of *Microcystis aeruginosa* in PBS amended with 1 mM Cl⁻. The treatment was
202 further operated in Lake Neatahwanta water (collected before the HAB season and filtered to
203 exclude background cyanobacteria and MC-LR) spiked with *Microcystis aeruginosa*. The Chl-a
204 removal kinetics were slightly retarded by the water matrices, but the >90% removal efficiency
205 can still be achieved after 180 s electrolysis. In contrast to *Synechococcus*, the *Microcystis*
206 *aeruginosa* culture contains both intra- and extracellular MC-LR. The destruction of MC-LR
207 consists of two parallel steps: (1) the lysed cells release intracellular MC-LR, which then become
208 extracellular MC-LR in water, and (2) the destruction of existing and nascent extracellular MC-LR.
209 The degradation of co-existing MC-LR in extra- and intra-cellular forms was investigated in PBS
210 electrolyte amended with 1 mM Cl⁻ and lake water. In both media, extra-cellular MC-LR was

211 subjected to faster degradation than intra-cellular MC-LR, as the degradation of the former
212 required the destruction of cell structures (Figure 3b).

213



214

215 **Figure 3.** (a) Inactivation of *Microcystis aeruginosa* and (b) destruction of intracellular (In) and
216 extracellular (Ex) MC-LR by electrolysis using NATO anode in PBS amended with 1 mM Cl⁻ and
217 Lake Neatahwanta (Lake-N) water at 7 mA/cm². The initial Chl-a concentration is 100 µg/L. The
218 initial concentrations of intracellular and extracellular MC-LR in PBS electrolyte are 1 and 3 µg/L ,
219 respectively. Data are presented as the mean value of triplicate ± standard deviation.

220

221 The bench-scale treatability study concluded that the ECO process is efficient for the
222 removal of cyanobacteria and MC-LR at short, minute-level retention time. The remaining
223 roadblock toward field development is understanding the environmental implications of the ECO
224 technology. Given the fast Chl-a degradation kinetics, we hypothesize that significant
225 cyanobacteria inactivation could be achieved before the yield of byproducts. Concern about the
226 formation of inorganic byproducts was first addressed in the bench-scale study under field-like
227 conditions. Other environmental aspects (organic byproducts and eco-toxicity) were investigated
228 in the field application (see the following content). Chlorate (ClO₃⁻) was identified as the product
229 stemming from the electrochemical oxidation of Cl⁻. Figure S4 shows the evolution of ClO₃⁻ in PBS
230 electrolyte amended with 1 mM Cl⁻ and lake water containing ~1 mM Cl⁻ at 10 mA/cm². The results
231 indicate that the evolution of ClO₃⁻ was suppressed in the lake water compared with electrolysis
232 in PBS electrolytes, possibly due to the depletion of free chlorine by reacting with matrix
233 components. As will be discussed below, the full-scale ECO reactor was operated at a Chl-a

234 removal efficiency of ~40%, corresponding to a treatment duration of 60 s in batch mode. In this
235 scenario, the $[\text{ClO}_3^-]$ was below the detection limit (5 $\mu\text{g/L}$) and the World Health Organization
236 (WHO) guideline of 0.7 mg/L.³⁴

237 Bromate is a byproduct derived from the ozonation of bromide (Br^-).³⁵ We do not believe
238 this is a concern in the lake water treatment in this study because Br^- was not detected in the lake
239 water of two test sites (Table S1). To address the concern about the future applications in treating
240 Br^- containing water, we investigated the formation BrO_3^- in the ECO treatment of PBS electrolyte
241 containing 0.2 mg/L Br^- , a concentration reported in some surface water.³⁶ The results show that,
242 throughout the 180 s ECO treatment process at 7 mA/cm², the dominant product is HOBr and
243 BrO_3^- formation was not detected (Figure S5). This finding highlights the advantage of ECO over
244 homogeneous ozonation as the former yields less bulk O_3 to form BrO_3^- .

245 The bench-scale investigation concludes that (1) ECO effectively removes cyanobacteria
246 and cyanotoxins at short, minute-level retention times; (2) the reaction is built upon the locally
247 concentrated O_3 produced at the NATO anode. The efficacy is independent of chloride
248 concentration, which broadens the treatment scenarios in fresh waters.

249 **Field Applications**

250 A boat-mount ECO system was developed for the scaled-up treatment of HAB-impacted
251 lake water (Figure 4a). The full-scale electrode array contains 20 pieces of mesh NATO anode
252 (50 × 50 cm; 0.5 × 0.8 mm rhombus openings) sandwiched by 21 pieces of perforated stainless
253 steel sheet cathode (50 × 50 cm; round opening at \varnothing 3 mm) at an interspace of 0.5 cm (Figure
254 4b). The polycarbonate reactor has a 190 L effective volume (excluding electrode array volume).
255 Other on-board components include a custom-made DC power supply, generators, and a water
256 pump. Algae plumes were captured by an intake pipe at an adjustable depth (0-1 m) underwater.
257 An intake screen with a mesh opening size of 1 mm was installed on the intake port to prevent
258 the entry of fish and other large aquatic life (Figure S6). Lake water was transferred from the pipe
259 to the pump and pushed through the 190 L reactor at a flow rate of 378 L/min (i.e., treatment
260 capacity of 544 m³/d), corresponding to a hydraulic retention time of 30 s.

261 The electrode array has a total projected anode area of 10 m². Applying 600-1000 A total
262 current on the electrode array resulted in current densities of 6-10 mA/cm² with a cell voltage
263 ranging from 12-20 V. The electrode array was tightly packed in the reactor. In operation, water
264 passing through the electrode mesh incurred efficient mass transfer. We have conducted
265 preliminary tests in the field to show that the Chl-a concentrations in samples collected from the

266 mid-height sampling port (representing bulk concentration) and outlet were the same (Figure S7).
267 Therefore, the reactor can be considered a completely mixed-flow reactor (CMFR).

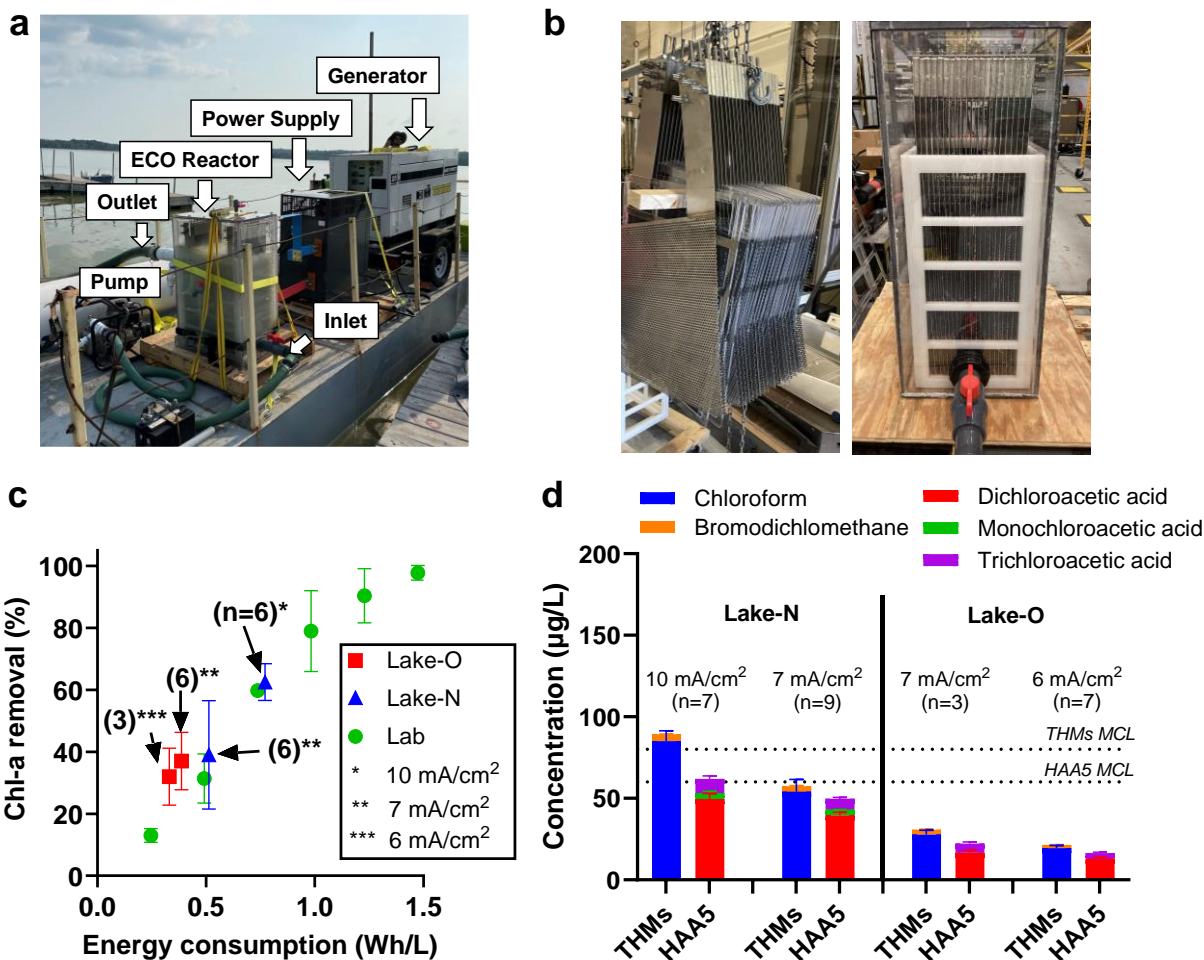
268 Based on the bench-scale batch reactions (data set of “PBS w/ 1 mM Cl⁻” in Figures 2b
269 and S3), the pseudo-first-order rate constants (k_{ECO} 's) at 7 and 10 mA/cm² are 0.026 and 0.061 s⁻¹,
270 respectively. When transforming the reaction from batch to CMFR, the Chl-a removal efficiency
271 can be estimated by

$$272 \quad \text{Removal efficiency (\%)} = (1 - C/C_0) \times 100\% = 1 - 1/(1 + \tau k_{ECO}) \quad (1)$$

273 where C_0 and C are Chl-a concentrations in water entering and exiting the reactor, respectively.
274 τ is the hydraulic retention time (30 s). Consequently, the removal efficiencies were projected as
275 44% and 64% for 7 and 10 mA/cm² operation, respectively, in CMFR mode.

276 The performance of the full-scale system was validated in the treatment of algal blooms
277 that occurred in Lake Neatahwanta (43°18'30"N 76°26'14"W) and Oneida Lake (43°10'25"N
278 75°55'50"W) in New York state. Lake Neatahwanta was treated in the midst of an algal bloom
279 with 105 µg/L Chl-a and 3 µg/L MC-LR, while tests on Oneida Lake happened in the early
280 blooming stage with Chl-a concentration of 5.4 µg/L (Table S1). Each field deployment lasted for
281 one day, with 9 to 12 pairs of influent and effluent samples taken. Tests in Oneida Lake were
282 performed at current densities of 6-7 mA/cm², while those in Lake Neatahwanta were conducted
283 at 7-10 mA/cm². As shown in Figure 4c, treatment operated at 7 and 10 mA/cm² achieved Chl-a
284 removal efficiency of 39% and 62%, respectively, approaching the predicted values based on the
285 CMFR model. More importantly, plotting removal efficiency data against specific energy
286 consumption (Wh/L) shows that the field test performances align well with the bench-scale data
287 set. The convergence of laboratory results and full-scale performance on Chl-a removal
288 demonstrates the promising linear scalability of the ECO process in cyanobacteria inactivation
289 using specific energy consumption as the key design parameter, given that the same anodes (i.e.,
290 NATO) are used.

291 Intracellular MC-LR was not found in the Lake Neatahwanta water samples. The treatment
292 at 7 mA/cm² led to an effluent extracellular MC-LR concentration (i.e., MC-LR resides in lake
293 water) of 0.41 µg/L (vs. $C_0 = 3$ µg/L), corresponding to a destruction efficiency of 86%. Note that
294 it is challenging to study the extracellular MC-LR degradation kinetics due to the rapid degradation
295 in ECO within a minute. Nonetheless, the 86% destruction of MC-LR when 40% Chl-a degradation
296 was obtained in the full-scale CMFR reactor is comparable with the results observed in the lab-
297 scale batch reactor (>95% destruction of MC-LR when ~40% Chl-a degradation was achieved,
298 as shown in Figure 3).



299
 300 **Figure 4.** (a) Layout of the boat-mount ECO system. (b) Side view of ECO reactor with mesh
 301 electrodes installed. (c) Chl-a removal efficiencies benchmarked by energy consumption. Data
 302 were collected from lab-scale investigation and field tests performed in Lake Neatahwanta (Lake-
 303 N) and Oneida Lake Oneida (Lake-O). n is the number of pairs of inlet and outlet samples taken.
 304 (d) Trifluoromethanes (THMs) and five haloacetic acids (HAA5) in the treated Lake Neatahwanta
 305 effluent when the reactor was operated at 7 mA/cm². MCL is Maximum Contaminant Level
 306 regulated by U.S. EPA.

307
 308 The current density of 7 mA/cm² (total current of 700 A and cell voltage of 14 V in treating
 309 Lake Neatahwanta) was identified as the optimum condition, balancing the significant algae
 310 removal, low byproduct formation, and insignificant toxicity (discussed below). It is important to
 311 note that the ~40% Chl-a removal at a short retention time of 30 s already incurred instant
 312 improvement in water clarity (Figure S8). Improvement in local water quality can be expected by

313 the circulative treatment of water. The energy consumption of 0.5 Wh/L is also lower than the
314 chlorine-based electrochemical HAB mitigation system (1.1 Wh/L) reported in our previous
315 study.³³

316

317 **Environmental Implications.**

318 The formation of halogenated byproducts, including trihalomethanes (THMs) and five
319 haloacetate acids (HAA5), was investigated in the treatment of Lake Neatahwanta and Oneida
320 Lake. THMs and HAA5 were not detected in the influent samples. In the effluent samples,
321 chloroform and dichloroacetic acid were the dominant THMs and HAAs, respectively. For
322 treatment operated at current densities of 6 and 7 mA/cm², the total concentrations of THMs and
323 HAA5 were below the Maximum Contaminant Level (MCL) values regulated by the U.S. EPA
324 (Figure 4d).

325 Whole effluent toxicity tests were conducted to evaluate whether the effluent from the ECO
326 process would negatively impact the aquatic life in the receiving water. Samples of influent (INF)
327 and treated effluent (EFF) were collected during the operation in Lake Neatahwanta (Table S2)
328 and Oneida Lake (Table S3) at 6-10 mA/cm². In the toxicity tests, model freshwater invertebrate
329 (*Ceriodaphnia dubia*) and fish (*Pimephales promelas*) species were exposed to INF samples, EFF
330 samples, and lab control water (CON). Ten replicates were used for *Ceriodaphnia dubia*, and four
331 replicates for *Pimephales promelas* for each INF, EFF, and CON exposure. Comparing the
332 toxicity test results of samples obtained at 10 mA/cm² (Table S2), although *Ceriodaphnia dubia*
333 survival was 100% at both 48 h (Acute) and 6-7 days (Chronic), reproduction is lower in EFF than
334 INF, suggesting the treatment induced toxicity for the invertebrate species. Conversely, no effects
335 were observed on the survival and growth of *Pimephales promelas*. The analysis of samples
336 derived from operation at 6 and 7 mA/cm² show that there were no significant survival impacts at
337 either 48 h (acute) or 6-7 days (chronic) for either the invertebrate or fish species, with survival
338 rates of 100%. Similarly, there were no significant effects on the reproduction of invertebrates or
339 growth of fish in EFF compared to INF. The whole effluent toxicity tests confirmed that the ECO
340 process operated at 6 and 7 mA/cm² did not induce acute or chronic toxicity in the treated effluent,
341 echoing our conclusion that 7 mA/cm² is the optimum current density.

342

343 **CONCLUSIONS**

344 This study developed an ECO process for the rapid inactivation of cyanobacteria and the
345 destruction of MC-LR. The process used NATO anodes to generate locally concentrated O₃ as

346 the major reactive species. Compared with homogeneous ozonation, the ECO process achieved
347 the same degree of treatment but yielded lower bulk $[O_3]$. Note that a majority of electrochemical
348 water treatment processes rely on the electrolysis of chloride to produce free chlorine as oxidants,
349 and the treatment performance depends on the chloride concentration^{37,38}. The shift of the
350 dominant oxidant from chlorine to O_3 provides a new strategy for sustaining high treatment
351 efficiency in freshwater with a limited chloride source and suppressing the formation of byproducts.

352 The significant merit of the engineering of this study is the scaled-up application of the
353 ECO process to solve real-world problems. The performance of batch mode lab-scale reactor was
354 replicated in a boat-mounted CMFR reactor using specific energy consumption as a benchmark.
355 In addition to the successful demonstration of HAB mitigation, we also evaluated whole effluent
356 toxicity to show that the ECO process operated at the optimized current density of 7 mA/cm^2 did
357 not increase the effluent toxicity. This case study sets an example for the scaled-up application
358 of electrified water treatment technology in environmental remediation.

359

360 **ASSOCIATED CONTENT**

361 **Supporting Information**

362 The Supporting Information is available free of charge at [Link].

363 The materials contain water composition, ECSA analysis, byproduct formation, and pictures of
364 reactors.

365

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