1,2,4-Triazole-containing More than an Amide **Bioisostere:** Discovery of Pyrazolo[1,5-α]pyrimidine Host CSNK2 Inhibitors for Combatting β-Coronavirus Replication

Han Wee Ong^{1,2#*}, Xuan Yang^{1,2#}, Jeffery L. Smith², Rebekah J. Dickmander^{1,3,4,5}, Jason W. Brown⁶, Tammy M. Havener², Sharon A. Taft-Benz^{1,7}, Stefanie D. Howell², Marcia K. Sanders^{1,7}, Jacob L. Capener², Rafael M. Couñago^{2,8}, Edcon Chang⁶, Andreas Krämer⁹, Nathaniel J. Moorman^{1,3,4}, Mark T. Heise^{1,7}, Alison D. Axtman^{1,2}, David H. Drewry^{1,2,4}, Timothy M. Willson^{1,2}

¹Rapidly Emerging Antiviral Drug Development Initiative (READDI), Chapel Hill, North Carolina 27599, USA;

²Structural Genomics Consortium (SGC) and Division of Chemical Biology and Medicinal Chemistry, Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA;

³Department of Microbiology & Immunology, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA;

⁴Lineberger Comprehensive Cancer Center, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA;

⁵Department of Chemistry, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA; ⁶Takeda Development Center Americas, Inc., San Diego, California 92121, USA;

⁷Department of Genetics, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA;

⁸Centro de Química Medicinal (CQMED), Centro de Biologia Molecular e Engenharia Genética (CBMEG), University of Campinas, Campinas, São Paulo, 13083-886, Brazil;

⁹SGC, Institute of Pharmaceutical Chemistry, Goethe University Frankfurt am Main, Max-von-Laue-Str. 9, 60438, Frankfurt am Main, Germany.

[#]co-first authors

*corresponding author

ABSTRACT

The pyrazolo[1,5-a]pyrimidine scaffold is a promising scaffold to develop potent and selective CSNK2 inhibitors with antiviral activity against β -coronaviruses. Herein, we describe the discovery of a 1,2,4-triazole group to substitute a key amide group for CSNK2 binding present in many potent pyrazolo[1,5-a]pyrimidine inhibitors. Crystallographic evidence demonstrates that the 1,2,4-triazole replaces the amide in forming key hydrogen bonds with Lys68 and a water molecule buried in the ATP-binding pocket. This isosteric replacement improves potency and metabolic stability at a cost of solubility. Optimization for potency, solubility and metabolic stability led to the discovery of the potent and selective CSNK2 inhibitor 53. Despite excellent in vitro metabolic stability, rapid decline in plasma concentration of 53 in vivo was observed and may be attributed to lung accumulation, although in vivo pharmacological effect was not observed. Further optimization of this novel chemotype may validate CSNK2 as an antiviral target in vivo.

potency metabolic stability solubility metabolic stability SGC-CK2-1 (1) 53 ✓ Selective CSNK2 inhibitor CSNK2 chemical probe Lung accumulation in vivo

GRAPHICAL ABSTRACT / TOC GRAPHIC

https://doi.org/10.26434/chemrxiv-2024-h2x2b ORCID: https://orcid.org/0000-0003-3232-2373 Content not peer-reviewed by ChemRxiv. License: CC BY-NC-ND 4.0

INTRODUCTION

The COVID-19 pandemic, caused by the β -coronavirus SARS-CoV-2, has resulted in 775 million infections and has claimed 7 million lives to date¹. The rapid emergence and spread of COVID-19 highlight the need for effective treatments against this disease. Other members of the β -coronavirus family have caused previous pandemics or are primed for emergence to cause future pandemics^{2,3}. While most antiviral drugs currently target viral factors, host-directed therapy is a promising alternative avenue to develop antiviral drugs. The main advantages of host-directed therapy for antivirals are the reduced propensity of the virus to develop resistance, and, importantly, the broad-spectrum ability of such therapeutics to inhibit viral replication of different viruses in the same family^{4–6}. Developing compounds for host-directed therapy thus has the potential to accelerate drug discovery against β -coronaviruses that emerge in the future, and safeguard humanity against the perils of a future pandemic.

Kinases are amongst the host proteins hijacked by viruses for viral replication, as phosphorylation events are important for viral replication across various virus genera^{4,7}. Casein kinase 2 (CSNK2) is a host kinase which plays a key role in replication of β -coronaviruses, including SARS-CoV-2⁸. Both CSNK2A1 and CSNK2A2 are highly upregulated upon SARS-CoV-2 infection⁹. Both CSNK2A subunits interact with the N protein of SARS-CoV-2, SARS-CoV and MERS-CoV^{10,11}, and the N protein is responsible for this upregulation of CSNK2 activity⁹. While kinase inhibitors have traditionally been developed for oncology indications, there is an increasing recognition of the role of kinases in other diseases, and a corresponding increase in interest for developing kinase inhibitors for other indications, including infectious diseases^{12–14}. We and others have previously demonstrated that CSNK2 inhibitors possess antiviral activity against multiple β coronaviruses including SARS-CoV-2, emphasizing the broad-spectrum antiviral activity of CSNK2 inhibitors^{9,15,16}. CSNK2 knockdown also demonstrates antiviral effect against the β -coronavirus MHV¹⁵. Efforts have been put into elucidation of the mechanism of CSNK2 inhibition on viral replication. The CSNK2 substrates with increased phosphorylation post-infection includes cytoskeletal proteins, and it is proposed that CSNK2 mediates remodeling of the extracellular matrix for viral egress⁹. An alternative mechanism of action is the modulation of the stress granule antiviral response¹⁰. We have also previously shown that CSNK2 inhibition disrupts viral entry via the clathrin-mediated endocytosis (CME) pathway¹⁵.

Many small molecule CSNK2 inhibitors have been developed and reported in the literature^{17,18}. Among these, the pyrazolo[1,5-*a*]pyrimidine series of inhibitors, as exemplified by the CSNK2 chemical probe SGC-CK2-1 (**1**, Figure 1), has demonstrated good cellular potency and possess excellent selectivity¹⁹. We have previously reported that both **1** and **2**, a related inhibitor also of the pyrazolo[1,5-*a*]pyrimidine series, inhibit replication of SARS-CoV-2, MHV, and two bat coronaviruses, SHC014-CoV and WIV1-CoV, in vitro¹⁵. However, antiviral activity was only investigated in infected cells in vitro. To further support CSNK2 as a host target for antiviral therapy, our project aims to validate CSNK2 as an antiviral target in vivo, in a mouse model of COVID-19 infection²⁰, through the development of an in vivo tool molecule. To do so, we sought to better characterize and understand structure-activity relationship (SAR) around the pyrazolo[1,5-*a*]pyrimidine scaffold to guide lead optimization.

Based on the crystal structure of analogues of the pyrazolo[1,5-*a*]pyrimidine series bound to CSNK2 (PDB ID: 3U4U²¹, 4GUB²², 5H8B²³, 5H8E²³, 5H8G²³, 6Z83¹⁹, 6Z84¹⁹), we noted that the meta-position amide substituent of the aniline ring played a critical role in CSNK2 binding. As exemplified by the co-crystal structure of **1** with CSNK2A1 (PDB ID: 6Z83) (Figure 2, left)¹⁹, the amide binds in a s-cis conformation, with the NH of the amide forming a hydrogen bond with the Asp residue of the DWG motif (Asp175 in CSNK2A1), and the carbonyl forming critical hydrogen bonding interactions with the catalytic lysine (Lys68 in CSNK2A1) and a water molecule buried in the ATP-binding pocket. Despite the key interactions the acetamide forms with CSNK2, only limited SAR exploration at this position had been reported in the literature, involving alkylation of the NH of the amide^{21,22}, and replacement with a sulfone or primary alcohol group²¹. Herein, we report the successful identification of a 1,2,4-triazole as a bioisosteric replacement for the amide. Co-crystal structures with CSNK2A1 support the binding hypothesis. We identify a lead compound **53** with a balance of potency, solubility, metabolic stability, and low cytotoxicity. Despite the low hepatic clearance measured in vitro, **53** demonstrated a rapid decrease in plasma concentration in vivo, attributed to distribution and accumulation in lungs, although this was insufficient to achieve pharmacological effect. Nevertheless, this successful isosteric replacement afforded an alternative chemotype for investigation of in vitro CSNK2 activity.



Figure 1. Exemplar CSNK2 inhibitors of the previously reported pyrazolo[1,5-*a*]pyrimidine series

RESULTS AND DISCUSSION

Identification of the 1,2,4-triazole as an amide isostere.

To characterize the SAR in the region of the meta-position amide substituent, we evaluated other substituents with a hydrogen bond acceptor three bond lengths away from the aniline ring, with an objective of recapitulating the same hydrogen bonding interactions the amide makes (Table 1). We first screened our compounds for CSNK2A1 and CSNK2A2 inhibition using the NanoBRET assay^{15,19}. For compounds of the pyrazolo[1,5-*a*]pyrimidine series reported previously^{15,19,24} and here, we did not observe significant selectivity between the two highly homologous CSNK2A subunits. Hence, we elected to focus on CSNK2A2 in the current screening campaign, since this subunit routinely gave a larger dynamic range in the NanoBRET assay. Data for CSNK2A1 inhibition, where collected, is available in Table S1. Additionally, we screened our compounds for inhibition of mouse hepatitis virus (MHV), a β -coronavirus in the same genus as SARS-CoV-2 used as a model for virulence of SARS-CoV-2²⁵, due to biosafety and technical advantages.

As compared to the amide (**3**), a 5-to-8-fold drop in CSNK2A2 and MHV potency was observed with the sulfonamide **4**. Replacement with the sulfone analogue **5** restored potency against both CSNK2A2 and MHV, indicating that the hydrogen bond formed by the NH of the amide was dispensable for CSNK2A2 inhibition. The sulfoximines **6** and **7** were found to be an order of magnitude less potent against CSNK2A2 than the sulfone **5** and correspondingly showed an almost complete loss of activity against MHV, demonstrating that replacement of either oxygen atom of the sulfone with an NH was not tolerated. Replacement of the sulfone with the racemic sulfoxide **8** largely maintained potency, and the phosphine oxide **9** led to a 10-fold drop in potency. Replacement of the amide with alcohol groups (**10**, **11**) proved unsuccessful, with a >25-fold drop in CSNK2A2 and MHV potency. A carboxyl group at this position (**12**) was also found not to be tolerated by CSNK2A2, and was likewise inactive against MHV. Our attempt to introduce a nitrile as a hydrogen bond acceptor (**13**) also led to a drastic loss in CSNK2 and MHV potency. As the acetamide was observed to form two hydrogen bonds, one with the catalytic lysine and another with the water molecule, we rationalized that the single hydrogen bond accepting capability of the nitrile was insufficient to recapitulate the same interactions.

Five-membered ring heterocycles are established bioisosteres of the amide bond²⁶. We hypothesized that CSNK2A2 would similarly accept five-membered ring heterocycles directly attached to the phenyl ring with hydrogen bond acceptors three bond lengths away. To our delight, replacement of the acetamide with a 1,2,4-triazol-4-yl group (14) successfully improved CSNK2A2 activity 4-fold. In concert with the increased CSNK2A2 potency, the potency of 14 in the MHV assay was also improved 4-fold. While the isomeric 1,2,3-triazol-5-yl group (15) had a <3-fold decrease in MHV and CSNK2A2 activity as compared to 14, the 1,2,3-triazol-1-yl group (16) was two orders of magnitude less potent against MHV and CSNK2A2. The sensitivity of CSNK2A2 inhibition to the positions of the nitrogen atoms on the ring suggests that both nitrogen atoms on the 1- and 2-position of the triazole in 14 form crucial interactions with the kinase.

Introducing a methyl group at the 3-position of the 1,2,4-triazole (17) resulted in decreased activity of CSNK2A2 and MHV, similar to the effects observed upon introducing a methyl group at the equivalent positions in the imidazole 18 and

triazole **19**. The corresponding imidazolinone **20** was similarly inactive. Together, these analogues demonstrate the steric limitations of the binding pocket around this group. Although, an alternative explanation may be that substituents adjacent to the biaryl bond may reduce the planarity of this biaryl system, preventing efficient interactions with the kinase. The introduction of a methylene linker between the phenyl ring and the triazole (**21**) also led to a drastic drop in both CSNK2A2 and MHV activity. This suggests that there are tight steric requirements within the kinase's ATP-binding pocket, and underscores the necessity of positioning the nitrogen atoms on the 1- and 2-position of the triazole of **14** appropriately for potent binding.

We next explored if the 1,2,4-triazole could be replaced with other heterocycles. The oxadiazole 22, thiazole 23, thiadiazole 24, and tetrazoles 25, 26, and 27 were only weakly active against CSNK2A2 and inactive against MHV. Expanding the ring size to a six-membered ring was shown to be similarly unfavorable as exemplified by the pyridine (28) and pyrazine (29) analogues. These results suggest that the 1,2,4-triazole is indeed a privileged amide bioisostere at this position, in terms of fitting the steric requirement and the optimal positioning of the hydrogen bond acceptors.

Table 1. SAR at the meta-position of the aniline.



Compound	R	MHV pIC50 ^a	CSNK2A2 NanoBRET pIC50 ^b	Compound	R	MHV pIC ₅₀ ^a	CSNK2A2 NanoBRET pIC50 ^b
3	K _N HO	6.8	7.8	17		6.2	7.4
4	−s=0 NH NH	6.1	6.9	18	K _N N	<5	7.5
5	, S=0 ℃	6.8	7.3	19		<5	<5
6°	∫S=NH ℃	5.0	6.4	20	C → NH	<5	<5
7°	S=NH V O	<5	6.1	21		<5	5.4
8	,	6.4	7.4	22	N-N	5.0	6.2
9	∕_P _{≥0}	5.1	6.2	23	S N	5.6	6.1
10	∕∕_он	5.1	6.2	24	S N N	<5	7.3
11	<u>, </u>	5.1	6.4	25		5.6	<5
12	Д ОН О	<5	<5	26	K _N -N _N	<5	8.3

13	N	<5	6.1
14	K N N N	7.4	8.4
15	Hz,z,z	7.1	8.0
16	∕ _N ·N _N	5.5	6.3

27		<5	6.7
28	N.	<5	5.3
29	N N	<5	<5

^aMean of three independent experiments performed in triplicate. ^bMean of two independent experiments. ^cCompounds were separated by chiral chromatography but stereochemistry not determined.

To better understand the binding mode of the triazole and the importance of the nitrogen atoms, we obtained a co-crystal structure of CSNK2A1 bound to **14** (Figure 2, right). **14** binds to CSNK2A1 in a very similar manner as compared to the CSNK2 chemical probe SGC-CK2-1 (1)¹⁹. Like **1**, the pyrazolo[1,5-*a*]pyrimidine core of **14** binds at the hinge region of the kinase, with the N1 atom and the 7-position exocyclic NH forming key hydrogen bonding interactions with the backbone of Val116. The nitrile group at the 3-position of the pyrazolo[1,5-*a*]pyrimidine core forms a hydrogen bond with a water molecule buried in the ATP binding pocket. The co-crystal structure with **1** shows the carbonyl group of the amide forming two hydrogen bonding interactions with Lys68 and the buried water molecule. These same two hydrogen bonding interactions were captured by the triazole of **14** using both the 1- and 2-position N atoms separately. Thus, the crystallographic evidence supports our conclusions from the SAR regarding the importance of both the 1- and 2-position N atoms on the triazole, and its suitability as a bioisosteric replacement for the critical amide substituent^{21–23}.



Figure 2. Co-crystal structure of **1** (left, PDB ID: 6Z83)¹⁹ and **14** (right, PDB ID: 8P07) with CSNK2A1. Part of the kinase P-loop is hidden for clarity. Water molecules are represented as red spheres. Hydrogen bonds are denoted as yellow dashed lines.

We next decided to investigate how this bioisosteric replacement modified other important properties. Given that metabolic stability is a primary concern for the pyrazolo[1,5-*a*]pyrimidines²⁴, we investigated the microsomal and whole hepatocyte stability of compounds **3** and **14** (Table 2). Compared to **3**, compound **14** exhibited improved metabolic stability in mouse and human liver microsomes and hepatocytes, providing another advantage of this isosteric replacement, although we noted that the metabolic clearance rates were still high. However, further profiling of compound **14** revealed poor aqueous solubility of 0.47 μ g/mL. Despite this, given the excellent potency and improved metabolic stability of the triazole, we decided to maintain this group at the meta-position of the aniline and aimed to enhance aqueous solubility by incorporating further modifications elsewhere on the molecule.

Ta	ble	e 2.	Com	parison	of	metab	olic	stability	/ and	solubility	of 3	3 and	14	

	Mouse liver				
	microsomal stability	Human liver			
	(% remaining after	microsomal CL _{int}	Mouse hepatocyte	Human hepatocyte	Kinetic Solubility
Compound	30 min) ^a	(mL/min/kg) ^b	CLint (mL/min/kg)c	CLint (mL/min/kg)c	$(\mu g/mL)^d$
3	22	25	520	58	2.8
14	74	7.6	160	16	0.47

^aMetabolism in MLM measured at 30 min by LC-MS. ^bMetabolism in HLM quantified by LC-MS over five time points over 1 h, scaled by scaling factors of mass of liver per body weight and microsomal concentration in liver. ^cMetabolism in hepatocytes quantified by LC-MS over 2 h, scaled by scaling factors of mass of liver per body weight and hepatocyte concentration in liver. ^dKinetic solubility is determined from 10 mM DMSO stock solutions in PBS buffered at pH 7.4.

Modifications to improve solubility and metabolic stability.

Based on the abovementioned crystal structures of the pyrazolo[1,5-*a*]pyrimidine inhibitors bound to CSNK2, we observed that the 7-position cyclopropylamino substituent points towards the solvent, offering an opportunity to add solubilizing groups at this position (Table 3). Replacement of the cyclopropyl ring with a slightly larger cyclobutyl ring (**30**) was found to be tolerated by CSNK2A2 and MHV, although this increase in lipophilicity led to a further decrease in solubility of the compound. To our surprise, however, we observed two orders of magnitude decrease in potency against CSNK2A2 and a loss of MHV activity with the oxetane ring (**31**), a result that contrasts with previous reports of the tolerability of CSNK2 towards the oxetane substituent^{21,22}. The 3,3-difluorocyclobutyl substituent (**32**) was similarly disfavored by CSNK2A2 and was inactive against MHV. We next introduced cyclic ethers as solubilizing groups at this position (**33** and **34**), but both compounds were inactive against CSNK2A2 and MHV. A (1-methyl-1H-pyrazol-4-yl)methyl substituent (**35**) and a N-acetylpiperidin-4-yl substituent (**36**) also demonstrated no activity against CSNK2A2 and MHV. Together, these results demonstrated that the steric requirement around this region of the CSNK2 active site is rather strict. We also demonstrated that the compound with a hydroxyethyl group (**37**) had weak activity against CSNK2A2 and was inactive against MHV. We infer from these results that the introduction of polar heteroatoms in this region could be detrimental for inhibition of CSNK2A2.

To inform selection of suitable substituents at this position, we surveyed literature for examples of groups successfully incorporated at this position. It has been demonstrated previously that imidazole or pyrazole substituents were tolerated by CSNK2A1 at this position, and pendant solubilizing groups may be successfully attached here without loss of CSNK2A1 potency²². We initially investigated the imidazole substituents. Despite an improvement in solubility, in contrast with the literature reports, the replacement of the cyclopropyl ring with an N-methyl imidazole (**38**) resulted in a 17-fold drop in CSNK2A2 potency and a 19-fold drop in MHV potency. Introduction of basic amines on the imidazole (**39**, **40**, **41**, **42**) led to a further drop in CSNK2A2 potency and a loss of MHV activity. We then turned to the pyrazole substituents. Similarly to the compounds with an imidazole, the compounds containing a pyrazole with pendant amines (**43**, **44**, **45**, **46**) improved solubility compared to **14**. However, we still observed a 5- to 1000-fold drop in CSNK2A2 potency for these compounds, which was accompanied by a \geq 25-fold drop in MHV potency. A similar trend was observed for CSNK2A1 potency as well (Table S1). Interestingly, comparing match pairs of imidazoles and pyrazoles (**39** with **43**, **40** with **44**, **41** with **45**) we observed that the pyrazoles were preferred by CSNK2, demonstrating that the exact positioning of the N atoms on this ring leads to significantly different inhibitory effects. The inconsistency of the SAR with the findings by Dowling *et al.*²² are puzzling, but nevertheless consistent with our previous findings that the cyclopropylamino group is the most optimal 7-position substituent for both CSNK2 and MHV inhibition¹⁵.

Selected analogues (30–35, 40–46) were also tested for metabolic stability in mouse liver microsomes. All analogues demonstrated moderate stability with 60–90% remaining after 30 min of incubation. Because these modifications at the 7-position of the pyrazolo[1,5-*a*]pyrimidine core did not significantly affect metabolic stability, this suggested that the major site of metabolism was unlikely to be at the cyclopropylamine group, in agreement with the metabolite identification study conducted previously for another analogue of this series²⁴.

Table 3. SAR at the 7-position of the pyrazolo[1,5-*a*]pyrimidine core.



Compound	R	MHV pIC50 ^a	CSNK2A2 NanoBRET pIC50 ^b	Kinetic solubility	Mouse liver microsomal stability (% remaining after 30 min) ^d
14		7.4	8.4	0.47	74
30	T,	7.1	8.1	0.18	74
31	27	<5	6.6	n.d.	85
32	F	<5	6.4	n.d.	75
33	\sim	5.3	<5	n.d.	80
34	\sim	<5	<5	n.d.	81
35		<5	<5	n.d.	77
36	o	<5	<5	n.d.	n.d.
37	HO	<5	<5	n.d.	n.d.
38		6.1	7.1	3.1	n.d.
39	HN	<5	5.8	8.4	n.d.
40		<5	6.3	30	63
41		<5	6.7	37	67
42		<5	5.3	4.2	81
43	HNNN	<5	7.1	2.1	61
44	N N N	5.3	7.5	21	66
45		5.8	7.6	35	71
46		6.0	7.1	4.0	86

^aMean of three independent experiments performed in triplicate. ^bMean of two independent experiments. ^cKinetic solubility measurements were carried out in phosphate buffered saline solution (PBS) at pH 7.4 from DMSO stock solutions. ^dMetabolism in MLM measured at 30 min by LC-MS. n.d. = not determined

Having ruled out the feasibility of improving solubility and metabolic stability by attaching solubilizing groups at the 7-position of the pyrazolo [1,5-a] pyrimidine while maintaining cellular potency, we next turned to optimization of the substituents on the aniline ring of the molecule (Table 4). We first explored the effect of introducing fluorine atoms on the aniline ring. Interestingly, the introduction of a fluorine atom at the ortho-position of the aniline ring (47) improved MHV potency while maintaining CSNK2A2 potency, increased stability in mouse liver microsomes, and led to a slight improvement in solubility when compared to 14. However, 47 demonstrated moderate cytotoxicity in A549-ACE2 cells (59% viability at 1 µM, Table 5) and we chose not to proceed further with this compound. Introduction of a fluorine atom at the para-position of the aniline ring (48) maintained CSNK2A2 potency, decreased MHV potency, and improved metabolic stability, although it did not improve solubility compared to 14, demonstrating that a fluorine atom was not favored at this position. Replacement of an aromatic CH with a nitrogen has been recognized as an effective strategy to improve solubility of compounds in multi-parameter optimization programs²⁷. Replacement of the phenyl ring with a pyridine ring with the aromatic N atom at the "ortho-position" (49) or "para-position" (50) both slightly decreased CSNK2A2 and MHV potency. The solubility was indeed slightly improved with 49 but, surprisingly, decreased with 50. The microsomal stability of 49 and 50 was improved compared to 14. Altogether, these four analogues demonstrate that a reduction in electron-rich nature of the aniline ring was a fruitful strategy in reducing metabolic clearance in liver microsomes, presumably by reducing the propensity for cytochrome-P450-mediated oxidation into an arene oxide or a quinone-imine ring^{28–30}.

We note that the successful attachment of solubilizing groups containing basic amines at the para-position of the aniline ring has been reported when the meta-position of the aniline ring possesses an acetamide or propionamide group^{15,19,23}. We hypothesized that this strategy could be similarly employed with our triazole compounds.

As hypothesized, introduction of an *N*-(2-aminoethyl)-*N*-methyl group at this position (**51**) improved solubility drastically. However, an unexpected 58-fold drop in CSNK2A2 potency and a 9-fold drop in MHV potency was observed. We further investigated other solubilizing groups. With a *N*-(2-aminoethyl)-*N*-ethyl group at the para-position (**53**), a modest two-fold improvement in MHV potency was observed when compared to **51**, while the CSNK2A2 potency remained in the micromolar range. Methylation at other positions in this para-position group (**55**, **56**, **57**) led to similar MHV and CSNK2A2 potency. Notably, all these analogues maintained excellent solubility. Adding an ethyl group to the aliphatic amine (**58**) decreased MHV potency drastically, although potency was recovered by the addition of a second ethyl group (**59**) and maintained good solubility. Sub-micromolar potency against CSNK2A2 and MHV was obtained with an alcohol (**60**) or methyl ether (**62**) in place of the amino group, suggesting that the basic amine may be replaced with neutral substituents. While **60** demonstrated excellent solubility, the solubility of **62** was poor. To investigate the steric requirements in this region, we replaced the pendant amine of **51** with a pyrrolidine (**63**) or morpholine (**64**). While this resulted in an improvement in CSNK2A2 potency, a decrease in potency against MHV was observed. The solubility of **64** remained favorable.

We then investigated alterative solubilizing groups with cyclic amines at this position. A basic amine was maintained with a distance of three atoms between the phenyl ring and the amine, in accordance to literature findings about the optimal linker length²³. Both enantiomers with the 3-aminopyrrolidin-1-yl group (**65**, **66**) maintained sub-micromolar potency against CSNK2A2, but the (*S*)-enantiomer **66** was 8-fold more potent than the (*R*)-enantiomer **65** against MHV. Expansion of the ring size by one methylene unit (**68**) did not significantly improve CSNK2A2 or MHV potency. Methylation of the amine (**69**) resulted in comparable CSNK2A2 potencies while decreasing the MHV potency 3-fold. The (*R*)-enantiomer (**70**) was less potent against CSNK2A2, with similar MHV potency. The solubility of these compounds remained excellent, as expected from the presence of a basic amine.

We next introduced a morpholine ring at this position (72). While we found that this compound possesses good CSNK2A2 and MHV potency as compared to the other analogues with basic amines, the modest increase in solubility as compared to 14 did not justify the 7.5-fold drop in CSNK2A2 potency and 3-fold drop in MHV potency. Changing the morpholine ring to an *N*-methylpiperazine ring (73) led to a 5-fold drop in potency against CSNK2A2 and an almost 50-fold drop in potency against MHV, while the solubility surprisingly decreased further.

Selected analogues with para-position solubilizing groups (51, 53, 55, 60, 62, 64, 66–69, 71, 72) were selected for metabolic stability screening. Except for 64, all analogues exhibited excellent stability in mouse liver microsomes, with 84–100% remaining after 30 min of incubation. This result suggested that the decreased lipophilicity resulting from the introduction of solubilizing groups was beneficial for decreasing hepatic metabolism.

Our results showed that despite improvements in solubility and microsomal stability, introducing solubilizing groups at the para-position decreases both CSNK2A2 and MHV potency. Having established earlier with compound **47** that an orthoposition fluorine atom could improve the MHV potency, we hypothesized that the introduction of this ortho-position fluorine atom could compensate for the lower potency with the solubilizing groups. We thus synthesized five compounds **52**, **54**, **61**, **67**, and **71**, which are the match-pair analogues of **51**, **53**, **60**, **66**, and **70**, respectively. However, for all the match pairs except **71** and **70**, what we observed instead was a general trend where MHV potency decreased at least 6-fold with the addition of this ortho-position fluorine atom.

Table 4. SAR at the para-position of the aniline.



Compound	v	D.	v	D.	MHV	CSNK2A2	Kinetic solubility	Mouse liver microsomal stability
Compound	л	K 1	I	K 2	pIC ₅₀ ^a	NanoBRET pIC50 ^b	(µg/mL) ^c	(% remaining after 30 min) ^d
14	С	Н	С	Н	7.4	8.4	0.47	74
47	С	F	С	Н	8.2	8.5	1.5	100
48	С	Н	С	F	6.5	8.3	0.30	93
49	Ν		С	Н	6.8	8.0	1.2	88
50	С	Η	Ν		7.2	8.0	0.27	99
51	С	Н	С	H ₂ N N	6.4	6.6	45	100
52	С	F	С		5.6	6.0	41	n.d.
53	C	Н	С	H ₂ N N	6.7	6.5	56	92
54	С	F	С	H_2N N N	5.8	6.3	39	n.d.
55	С	Н	С	$H_2 N^{(1)} N_{1}$	6.3	6.6	62	84
56	С	Н	C	H ₂ N N	6.7	6.5	50	n.d.
57	С	Н	С	T Z Z	6.0	6.9	16	n.d.
58	С	Н	С		5.1	6.2	n.d.	n.d.
59	С	Н	С		6.1	7.0	60	n.d.
60	С	Н	С	HO	6.7	6.9	38	95
61	С	F	С	HO	5.6	7.2	2.6	n.d.

62	С	Н	С	NH	6.5	6.9	0.80	86
63	С	Н	С	N N	5.0	7.3	n.d.	n.d.
64	С	Н	С		6.2	7.1	25	0.0
65	С	Н	С	$H_2N^{(R)}$	5.6	6.6	44	n.d.
66	С	Н	С	$H_2N^{(S)}$	6.4	6.8	49	99
67	С	F	С	H ₂ N ^(S)	5.7	6.6	37	97
68	С	Н	С	H ₂ N ¹ , (5)	6.2	6.9	47	98
69	С	Н	C		5.7	6.9	56	97
70	С	Н	С		5.5	6.1	63	n.d.
71	С	F	С		5.8	6.8	60	94
72	С	Н	С		6.9	7.5	1.1	93
73	С	Н	С		5.3	6.8	0.20	n.d.

^aMean of three independent experiments performed in triplicate. ^bMean of two independent experiments. ^cKinetic solubility measurements were carried out in phosphate buffered saline solution (PBS) at pH 7.4 from DMSO stock solutions. ^dMetabolism in MLM measured at 30 min by LC-MS. n.d. = not determined.



Figure 3. Co-crystal structure of 50 (left, PDB ID: 8P06) and 53 (right, PDB ID: 9EZG) with CSNK2A1. Part of the kinase P-loop is hidden for clarity. Water molecules are represented as red spheres. Hydrogen bonds are denoted as yellow dashed lines.

We have additionally obtained co-crystal structures of **50** and **53** (Figure 3) with CSNK2A1. Both compounds bind to CSNK2A1 in an almost identical manner as **14**. For both compounds, we observed hydrogen bonds between the N1 of the pyrazolo[1,5-*a*]pyrimidine and the 7-position exocyclic NH with the backbone of Val116, and with both the 1- and 2-position nitrogen atoms of the triazole with the buried water molecule and Lys68 respectively. For **53**, the ethyl group points towards the P-loop while the 2-aminoethyl group oriented towards the C-lobe, forming a hydrogen bond with Asn161 and consistent with a literature crystal structure of the 2-aminoethyl group at this position (PDB ID: 5H8E)²³.

Unlike previously reported for compounds with a meta-position acetamide and propionamide^{15,19,23}, the addition of a para-position substituent to compounds with a meta-position triazole generally lead to a loss of CSNK2A2 and MHV activity. While one possible explanation could be decreased cellular permeability of the more polar compounds, this might also be attributed to disruption of the high degree of coplanarity between the triazole ring and the phenyl ring. A high degree of coplanarity was observed for both **14** and **50**, with dihedral angles of $9.6/9.9^{\circ}$ and $3.5/4.2^{\circ}$ respectively. As a reference, the measured dihedral angle across the C–N bond between the phenyl ring and the N(H)–C(=O) bond in the crystal structure of CSNK2A1 with **1** was 55.6°. A para-position substituent on the aniline ring would be expected to increase the dihedral angle, deviating from the optimal angle for binding. Because the triazole uses both 1- and 2-position N atoms to form separate hydrogen bonds with Lys68 and the water molecule, and hydrogen bonds are directional interactions sensitive to changes in bond angle, an increase in dihedral angle would be expected to diminish the binding energy of the two hydrogen bonds and preclude efficient binding to CSNK2. We noted that the dihedral angle of **53** in its co-crystal structure with CSNK2A1 was $8.7/15.2^{\circ}$, which was slightly higher than that for **14** and **50**. This rationale was also supported by the decrease in potency following introduction of a methyl group at the 3-position of the 1,2,4-triazole (**17**), at the equivalent positions in the imidazole (**18**) and triazole (**19**), and the exocyclic oxygen atom of the imidazolinone (**20**).

We selected compounds that demonstrated MHV IC₅₀ $\leq 1 \mu$ M for evaluation of cytotoxicity in A549-ACE2 cells at 1.0 μ M and 0.1 μ M inhibitor concentrations using a CellTiter-Glo assay (Table 5). All compounds except **47** and **56** demonstrated \geq 85% viability at 1 μ M and negligible cytotoxicity at 0.1 μ M.

Despite excellent potency against CSNK2 and MHV, low cytotoxicity and favorable metabolic stability of **14** and **50**, their low solubility resulted in significant challenges during chemical synthesis and purification upon reaction scale-up, as well as during formulation development for in vivo studies. As a compromise between potency and aqueous solubility, we selected **53**, which had sub-micromolar potency against both CSNK2 and MHV, and also possessed excellent solubility and exemplary microsomal stability, for further evaluation.

Compound	A549-ACE2 % viability at 1 μM ^a	A549-ACE2 % viability at 0.1 μM ^a		
14	85	100		
15	85	100		
30	93	102		
47	59	86		
48	99	99		
49	96	106		
50	98	99		
51	87	99		
53	98 ^b	101 ^b		
55	91	95		
56	76	93		
59	100	98		
60	97	95		
62	99	99		
64	101	96		
66	85	101		
68	102	106		
72	105	105		

Table 1. Cytotoxicity of compounds with potent

antiviral activity.

^aMean of quadruplicate experiments. ^bn = 9. n.d. = not determined

Table 2. Selectivity of 53 in the NanoBRET K192 Panel.

Kinase	NanoBRET K192 % occupancy at 10 μM ^a	NanoBRET pIC50
CSNK2A2	97	6.5 ^b
CSNK2A1	97	n.d.
CLK1	74	5.3 ^b
CLK2	73	5.5°
DAPK2	66	5.6°
HIPK4	63	4.6 ^b
CLK4	59	5.6°
PHKG1	59	5.2°
DYRK1A	56	n.d. ^d
CDK7	50	4.8°

^aPerformed in singlicate. ^bMean of two independent experiments. ^cDetermined from one experiment with two dilution curves. ^dPoor assay window observed but no occupancy observed up to 3 μ M. n.d. = not determined.

Potency and selectivity characterization of compound 53.

To ascertain the potency of **53** in orthogonal assays, we first evaluated **53** against CSNK2A1 and CSNK2A2 in the Eurofins KinaseProfiler assay, an in vitro radiometric enzyme assay performed with [ATP] at the K_m of the kinases. **53** was found to be a potent inhibitor of both CSNK2A1 and CSNK2A2, with IC₅₀ values of 1.7 nM and 0.66 nM respectively (Figure S1). The difference between the potencies as measured by the cellular NanoBRET assay and the in vitro KinaseProfiler assay are likely due to cellular permeability of the inhibitor or due to the higher cellular

concentrations of ATP than used in the KinaseProfiler assay. We next assayed **53** in the NanoBRET assay using a modified procedure, where digitonin was added to permeabilize the cells to eliminate the factor of membrane permeability in the assay³¹. Under these conditions, a dose-dependent inhibition of CSNK2A2 was also observed, with an IC₅₀ of 80 nM (Figure S2), representing a 4-fold increase in potency as compared to the regular conditions for the NanoBRET. Additionally, when A549-ACE2 cells were treated with 1 or 5 μ M of **53**, we measured a reduction in phosphorylation levels of the CSNK2 substrate EIF2S2³² after 24 hours (Figure S3), indicating that **53** inhibits CSNK2 downstream signaling. Altogether, these results confirmed that **53** is a potent CSNK2 inhibitor. Clearly, despite minor permeability limitations, **53** was still sufficiently permeable to achieve efficacy against MHV, and the NanoBRET assay was a good predictor of the cellular activity in the antiviral assay.

Our next step was to evaluate the kinome-wide selectivity of **53**. We measured the % occupancy of **53** at 10 μ M against 192 human kinases using the NanoBRET K192 assay panel, a selectivity panel that determines target engagement in the cellular context³³ (Figure 4). Out of 192 kinases, only two kinases, CSNK2A1 and CSNK2A2, were engaged by **53** with 97% occupancy (Table 6). Eight other kinases (CLK1, CLK2, DAPK2, HIPK4, CLK4, PHKG1, DYRK1A, and CDK7) were identified with % occupancy values between 50–75%, while all other kinases possess <50% occupancy at 10 μ M (Table

S3). A follow-up dose-response experiment in the NanoBRET assay was conducted for the off-target kinases, which verified that CLK1, CLK2, DAPK2, HIPK4, CLK4, PHKG1, and CDK7 were inhibited only with micromolar IC₅₀s (Table 6). With at least approximately one order of magnitude of potency difference between inhibition of CSNK2 and inhibition of identified off-target kinases, we concluded that **53** will be a selective inhibitor of CSNK2 in cells.



Figure 4. Selectivity profile of **53** determined at 10 μ M using the NanoBRET K192 panel³³. Only CSNK2A1 and CSNK2A2 had 97% occupancy (red). Kinases with 50-75% occupancy are shown in yellow. Kinases with <50% occupancy are shown in green. Kinases not in the NanoBRET K192 panel are shown in grey. Image generated using CORAL³⁴. No kinases had occupancy >75% except CSNK2A1 and CSNK2A2. Detailed % occupancy values for each kinase are described in Table S3.



Figure 2. (A) Structure of 53. (B) Dose-dependent effect of 53 against SARS-CoV-2 in A549-ACE2 cells and cell viability of A549-ACE2 cells as measured by the LDH assay.

Having established that **53** is a potent and selective CSNK2A2 inhibitor with good antiviral activity against MHV, we next sought to evaluate it for activity against SARS-CoV-2 in vitro (Figure 5). A dose-dependent inhibition of SARS-CoV-2 replication was observed with an IC₅₀ of 390 nM. No decrease in cell viability by the LDH assay was observed at concentrations below 10 μ M, demonstrating that this assay result was not confounded by host cell cytotoxicity. The cytotoxicity results in the LDH assay were also consistent with the CellTiter-Glo assay results.

In vitro ADME and in vivo PK of compound 53.

Encouraged by these results, we further characterized **53** in in vitro ADME studies (Table 7) and in vivo mice pharmacokinetic experiments (Table 8). As discussed previously, **53** possesses excellent solubility of 56 μ g/mL. However, **53** had poor permeability in MDCK cells transfected with MDR1, although it was not susceptible to rapid efflux by MDR1 (efflux ratio of 2.3). Compound **53** was also not highly plasma protein bound in human plasma, with an unbound fraction of 15%. **53** possessed good metabolic stability in mouse and human liver microsomes and whole hepatocytes, a promising finding considering that inhibitors of the pyrazolo[1,5-*a*]pyrimidine scaffold are known to demonstrate high mouse hepatocyte clearance due to phase I and phase II metabolism, commensurate with rapid metabolism in vivo²⁴.

In vivo, 53 demonstrates good bioavailability by i.p. dosing at 10 mg/kg or 30 mg/kg (Table 8). However, it showed no bioavailability when dosed at 10 mg/kg p.o. (data not shown), suggesting that intestinal absorption was a key pharmacokinetic challenge moving forward. This is possibly related to its limited cellular permeability. Nevertheless, the i.p. route of administration is a well-recognized and acceptable means of dosing for in vivo proof-of-concept studies³⁵, and we opted for this route of administration moving forward. Given the excellent in vitro metabolic stability, we were surprised by the short plasma half-life and high apparent intrinsic clearance of 53 when mice were dosed at 3 mg/kg i.v., 10 mg/kg i.p. or 30 mg/kg i.p. The moderate volume of distribution of 3.1 L/kg indicated that this compound distributes evenly throughout blood and tissues. This result was unsurprising, considering the basic character of 53 might predispose it to bind to negatively-charged phospholipids³⁶. We hypothesized that the high apparent intrinsic clearance was due to a rapid drop in plasma concentrations measured during the distribution phase, as opposed to the elimination phase, of the compound. The distribution of **53** into tissues might account for the high apparent clearance. Since the lung concentration is of particular interest for the effective treatment of SARS-CoV-2, we decided to investigate the pharmacokinetic parameters of the lung compartment of mice dosed with 10 mg/kg i.p. of 53 (Figure 6). Gratifyingly, we observed that from the 0.5 to 4 h time point, the lung/plasma ratio of 53 increased from 0.6 to 48, before slowly declining to a ratio of 18 by 24 h, supporting our hypothesis that 53 does partition into the lung tissues. The rapid initial increase of the lung/plasma ratio over the first four hours also supported our hypothesis that the low plasma half-life of the compound measured initially was due to drug distribution rather than metabolism. We measured the plasma half-life of 53 using the data points from 0.5-4 h, and found it to be comparable to the value determined previously (0.63 h). In contrast, the half-life of 53 from 4-24 h was much longer, at 8 h in the plasma compartment and 5 h in the lung compartment. These results suggest that the low rate of in vitro metabolism did in fact translate to a reduction in the in vivo metabolic clearance in mice. While the plasma concentration of 53 had dropped below the in vitro SARS-CoV-2 IC₅₀ by two hours, importantly, the lung concentration of 53 remained

above this level for at least 12 hours from a single dose, only dropping below this level by 24 hours. From these results, we hypothesized that b.i.d. dosing of 10 mg/kg i.p. might be able to maintain an efficacious concentration of this compound at its target tissue.

Table 7. In vitro ADME characterization of compound 53.

				Mouse liver			
				microsomal	Human liver	Mouse	Human
Kinetic	MDCK-MDR1		Human Plasma	stability (%	microsomal	hepatocyte	hepatocyte
Solubility ^a	Papp A-B/B-A		Protein	remaining after	CLint	CLint	CLint
(µg/mL)	(10 ⁻⁶ cm/s) ^b	Efflux Ratio ^b	Binding f_u (%) ^c	30 min) ^d	(mL/min/kg)e	(mL/min/kg)f	(mL/min/kg)f
56	0.16/0.36	2.3	15	92	<6.8	<32.0	<6.9

^aKinetic solubility is determined from 10 mM DMSO stock solutions in PBS buffered at pH 7.4. ^bPermeability assay in MDCK cells transfected with human MDR1, in apical-to-basolateral (A-B) and basolateral-to-apical (B-A) directions. ^cPlasma protein binding measurements performed in duplicate. ^dMetabolism in MLM measured at 30 min by LC-MS. ^cMetabolism in HLM quantified by LC-MS over five time points over 1 h, scaled by scaling factors of mass of liver per body weight and microsomal concentration in liver. ^fMetabolism in hepatocytes quantified by LC-MS over 2 h, scaled by scaling factors of mass of liver per body weight and hepatocyte concentration in liver.

Table 8. In vivo (5 h) mice pharmacokinetic data for compound 53.

Dose	Route of			CLint			AUClast	
(mg/kg)	Administration	Compartment	t1/2 (h)	(mL/min/kg)	t _{max} (h)	C _{max} (nM)	(h×nM)	Vd _{ss} (L/kg)
3	i.v.	Plasma	0.91	86	n.d.	n.d.	1300	3.1
10	i.p.	Plasma	0.63	n.d.	0.5	6300	6400	n.d.
30	i.p.	Plasma	0.63	n.d.	0.5	27000	31000	n.d.

75

n.d. = not determined.

A. 100000



B.	Compartment	t _{1/2} (0.5-4 h) (h)	t _{1/2} (4-24 h) (h)	t _{1/2} (0.5-24 h) (h)	C _{max} (nM)	AUC _{last} (h×nM)
	Plasma	0.45	7.8	n.d.	19000	15000
	Lung	n.d.	5.1	4.3	11000	39000

Figure 6. (A) Concentration over time of **53** in CD-1 mice, dosed i.p. at 10 mg/kg at t = 0 h. Plasma concentrations (light blue line) and lung concentrations (dark blue line) of the compound were measured at 0.5, 1, 2, 4, 8, 12, and 24 h, and the lung/plasma ratio is calculated (black dashed line). Data points are shown as mean \pm s.d. (n=3). The SARS-CoV-2 IC₅₀ of **53** determined in A549-ACE2 cells is plotted as a yellow dotted line. (B) Pharmacokinetic parameters.

In vivo efficacy.

We investigated **53** in a prophylactic treatment model for SARS-CoV-2 MA10 infection in mice²⁰. Briefly, **53** was dosed i.p. at 10 mg/kg every 12 h, with the first dose performed 12 h before viral inoculation. The mice were euthanized 24 h after viral inoculation (36 h after the first dose of **53**). The SARS-CoV-2 viral titer measured in mouse lung was similar in mice treated with **53** and mice given a vehicle treatment (Figure 7A). As a biomarker for CSNK2 inhibition, we measured phosphorylation of CSNK2 substrates EIF2S2³² and AKT³⁷ in mouse lung 36 h after treatment with **53** dosed at 10 mg/kg i.p. b.i.d. Compared to vehicle controls, no statistically-significant reduction in phosphorylation levels of Ser2 of EIF2S2 and Ser129 of AKT were observed (Figure 7B–G). The disconnect between the pharmacokinetics and efficacy of **53** is unexpected. We measured the protein binding levels of **53** in mouse lung homogenate, and found it to be 96.8% bound. As such, the free drug concentration of **53** is likely to be much lower than expected based on the measured total drug concentration, and may not be sufficient to achieve in vivo efficacy. For antivirals, it has been recommended that the in vivo free drug concentration to remain above the EC₉₀ levels determined in cellular assays for therapeutic effect^{38,39}. We acknowledge that the measured lung concentrations represented the organ-wide average, which may not reflect the true concentration in the specific microenvironment of the target, given the heterogeneity in cell types and subcellular compartments present. It was also possible that the concentration of **53** at the target microenvironment of CSNK2 was insufficient to achieve pharmacological effect.



Figure 7. (A) Viral titer measured in mouse lungs 24 hours post-inoculation when treated with vehicle or with compound 53 (10 mg/kg i.p. b.i.d.) (n=5). (B) Western blot for total EIF2S2 expression in mouse lung samples after treatment with vehicle or 53 (10 mg/kg i.p. b.i.d.) (n=3). (C) Western blot for EIF2S2 (phospho-Ser2) in mouse lung samples after treatment with vehicle or 53 (10 mg/kg i.p. b.i.d.) (n=3). (D) EIF2S2 (phospho-Ser2) levels normalized to total EIF2S2 levels in mouse lung. (E) Western blot for total AKT expression in mouse lung samples after treatment with vehicle or 53 (10 mg/kg i.p. b.i.d.) (n=3). (F) Western blot for AKT (phospho-Ser129) in mouse lung samples after treatment with vehicle or 53 (10 mg/kg i.p. b.i.d.) (n=3). (G) AKT (phospho-Ser129) levels normalized to total AKT levels in mouse lung.

CONCLUSION

In conclusion, we have successfully identified the 1,2,4-triazole as an appropriate isostere of the amide group. This amide group was previously found to be critical for binding to CSNK2 through its involvement in the hydrogen bonding network with Lys68 and a water molecule buried in the ATP-binding pocket. The 1,2,4-triazole successfully captures these hydrogen bonds using two different nitrogen atoms. This bioisosteric replacement improved on-target potency against CSNK2 and antiviral potency against MHV, and led to favorable metabolic stability profiles. Through a multiparameter optimization campaign, we have characterized SAR at the 7-position of the pyrazolo[1,5-*a*]pyrimidine and at the para-position of the aniline for this new chemotype. Compound **53** maintained a balance of potency, solubility, metabolic stability profile in vitro, a rapid decline in plasma levels of **53** was observed in vivo, which may be attributed to the distribution and accumulation into organs such as the lung. Pharmacological efficacy was not achieved in vivo, possibly attributed to insufficient free drug concentrations. Nevertheless, the SAR described herein further contributes to the development of improved CSNK2 inhibitors for use in vivo. **53** is a potent and selective CSNK2 inhibitor with well-characterized physiochemical and pharmacokinetic properties both in vitro and in vivo. We thus envisage that **53** may be used to investigate CSNK2 activity as an inhibitor of a new chemotype.

CHEMISTRY



^aReagents and conditions: (a) cyclopropylamine, EtOH, 25°C, 2 h; (b) Pd(OAc)₂, BINAP, Cs₂CO₃ or 'BuOLi or 'BuOK, dioxane, μ W, 130°C, 0.5 h or 100°C, 2 h; (c) Brettphos Pd G3, Cs₂CO₃, dioxane, μ W, 130°C, 0.5 h; (d) TFA, DCM, 25°C, 2 h.

Scheme 2. Synthesis of aniline intermediates 79-81, 83, 87, 89, 92, 93, 95, 96, 99, 101.^a



https://doi.org/10.26434/chemrxiv-2024-h2x2b ORCID: https://orcid.org/0000-0003-3232-2373 Content not peer-reviewed by ChemRxiv. License: CC BY-NC-ND 4.0

Scheme 1. Synthesis of analogues varying the meta-position of the aniline 3-29.^a

^aReagents and conditions: (a) NaSMe, MeOH, 0°C-25°C, 3 h; (b) mCPBA, DCM, 0°C, 1 h; (c) Rh(OAc)₂, MgO, iodobenzene diacetate, tert-butyl carbamate, DCM, 25°C, 16 h; (d) Fe, NH₄Cl, EtOH, H₂O, 65-90°C, 1-3 h; (e) Me₂P(=O)H, NaHMDS, THF, -30°C, 1 h then 25°C, 12 h; (f) TFA, DCM, 25°C, 2-10 h; (g) H₂, Pd/C, MeOH, 25°C, 2-12 h; (h) CbzCl, DIPEA, DCM, 25°C, 16 h; (i) MeMgBr, THF, 25°C, 16 h; (j) TMSN₃, CuI, DMF, MeOH, 100°C, 12 h; (k) acetohydrazide, DBU, EtOH, 85°C, 10 h; (l) NaNO₂, AcOH, H₂O, 0°C-100°C, 5 h; (m) SnCl₂.H₂O, HCl, EtOH, 70°C, 10 h; (n) NH₂CH₂CH(OMe)₂, DCM, 0°C-25°C, 3 h; (o) 4H-1,2,4-triazole, K₂CO₃, MeCN, 100°C, 10 h; (p) ArBr, Pd(dppf)Cl₂, Cs₂CO₃, dioxane, H₂O, 100°C, 5 h; (q) 'BuONO, HBF₄, EtOH, 0°C-25°C, 1 h; (r) TMSCHN₂, (CF₃CO₂)Ag, Et₃N, THF, -78°C, 1 h; (s) CsF, MeOH, rt, 0.5 h.

Synthesis of analogues varying the meta-position of the aniline (Scheme 1) was achieved by displacing the 7-chloro substituent on 5,7-dichloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (74) with cyclopropylamine to obtain the common intermediate 75. Buchwald-Hartwig coupling between 75 and anilines 76-101 furnished the desired analogues 3-29, removing a Boc or 'Bu protecting group where necessary. Aniline coupling partners were purchased where commercially available, and synthesized individually otherwise (Scheme 2). Anilines 79 and 80 were synthesized by a $S_N 2$ reaction between benzyl bromide 104 and NaSMe, followed by oxidation using mCPBA to obtain the sulfoxide 106, subsequent Rh(II)-catalyzed oxidation to obtain the Boc-protected sulfaneylidene 107, and finally nitro reduction. An analogous S_N2 reaction between benzyl bromide 108 and dimethylphosphine oxide, followed by Boc deprotection afforded aniline 81. Aniline 83 was synthesized by the attack of the ester group of 112 by two equivalents of MeMgBr, with the necessary Cbz protection and deprotection steps for the aniline. The 1H-1,2,3-triazol-5-yl group of aniline 87 was prepared through a copper-catalyzed azide-alkyne "click" reaction between the alkyne of 114 and TMSN₃, followed by nitro reduction of the nitro group to afford 87. A base-promoted condensation between the isothiocynate group of 116 and acetohydrazide afforded 117, which was desulfurized by $NaNO_2$ in acetic acid to form 118, and underwent nitro reduction to yield aniline 89. The reaction between isocyanate 119 and 2,2-dimethoxyethan-1-amine afforded the urea 120, which cyclized to the imidazol-2-one 121, and the nitro group was then reduced to the aniline 92. Synthesis of aniline 93 proceeded through a similar $S_N 2$ reaction between 1,2,4-triazole and benzyl bromide 104, with a subsequent nitro reduction. The boronate ester of 123 underwent Suzuki coupling reactions with the appropriate aryl bromide to furnish anilines 95, 96, and 101. Finally, the aniline 99 was synthesized by first converting the amino group of 124 to a diazonium salt 125, which reacted with TMSCHN₂ in a silver(I)-catalyzed [3+2] cycloaddition to form the tetrazole 126, before reduction of the nitro group to afford 99.

Scheme 3. Synthesis of analogues varying the 7-position substituent of the pyrazolo[1,5-a]pyrimidine 30-37.^a



^aReagents and conditions: (a) RNH₂, EtOH, rt, 2 h; (b) **86**, Pd(OAc)₂, BINAP, Cs₂CO₃ or ^tBuOLi, dioxane, μ W, 130°C, 0.5 h; (c) **86**, Brettphos Pd G3, Cs₂CO₃, dioxane, μ W, 130°C, 0.5 h; (d) TFA, DCM, 35°C, 2 h.



Scheme 4. Synthesis of analogues varying the 7-position substituent of the pyrazolo[1,5-a]pyrimidine 38-46.^a

^aReagents and conditions: (a) H₂, Pd/C, MeOH or THF, 20-40°C, 2-12 h; (b) **74**, EtOH, 25°C, 2-5 h; (c) **86**, Pd(OAc)₂, BINAP, Cs₂CO₃, dioxane, μ W, 130°C, 0.5-6 h; (d) 2-bromoethanol, K₂CO₃, MeCN, 60°C, 10 h; (e) BocNHCH₂CH₂Br, K₂CO₃, DMF, 90°C, 3 h; (f) SOCl₂, DCM, DMF, 25°C, 2 h; (g) MeNH₂, K₂CO₃, NaI, μ W, 80°C, 4 h; (h) Boc₂O, K₂CO₃, THF, 25°C, 3 h; (i) MeI, NaH, THF, 0-25°C, 10 h; (j) TFA, DCM, 25-35°C, 2-6 h; (k) RCH₂CH₂Cl, K₂CO₃, MeCN or DMF, 60-120°C, 4-12 h; (l) RCH₂CH₂Cl, NaH, DMF, 15°C, 3.5 h; (m) **86**, Brettphos Pd G3, Cs₂CO₃, dioxane, μ W, 130°C, 4 h.

The synthesis of analogues varying the 7-position of the pyrazolo[1,5-*a*]pyrimidine (Scheme 3) proceeded through an S_NAr reaction between 74 and commercially available amines to furnish intermediates 127-134, which reacted with 86 in Buchwald-Hartwig coupling reactions to yield analogues 30-36 and 135, which was converted to 37 by deprotection of the TBDMS group. Synthesis of analogues with an amino-imidazole or amino-pyrazole at the 7-position of the pyrazolo[1,5-*a*]pyrimidine (Scheme 4) began by functionalizing 4-nitro-1H-imidazole (139) or 3-nitro-1H-pyrazole (140) with the appropriate alkyl halide using an S_N2 reaction, followed by functional group interconversion where necessary. The nitro groups of functionalized imidazoles and pyrazoles 136, 144, 146, 153-158 were reduced to an amino group via hydrogenation (137, 147-148, 159-164), which next reacted readily with 74 in an S_NAr reaction to form intermediates 138, 149-150, 165-170. Buchwald-Hartwig coupling with 86 furnished the analogues 38-46, after Boc deprotection where necessary.



^aReagents and conditions: (a) 1,2-diformylhydrazine, Me₃SiCl, Et₃N, pyridine, 100°C, 12 h; (b) for **179**, **180** and **182**, H₂, Pd/C, MeOH, 25°C, 10 h; (c) for **181**, NH₃, H₂O, 100°C, 72 h; (d) **75**, Brettphos Pd G3, 'BuOLi, dioxane, μ W, 130°C, 0.5 h; (e) **75**, Pd(OAc)₂, BINAP, Cs₂CO₃ or 'BuOLi, dioxane, μ W, 130°C, 0.5 h.

The synthesis of analogues **47-50** (Scheme 5) started with a condensation between the aniline of **171-174** and 1,2diformylhydrazine to furnish the 1,2,4-triazole ring of **175-178**. The nitro group of **175**, **176**, and **178** were reduced via hydrogenation to afford anilines **179**, **180** and **182** respectively, while **177** underwent an S_NAr reaction with ammonia to yield **181**. A final Buchwald-Hartwig coupling step with **75** completes the synthesis of **47-50**.

Scheme 6. Synthesis of non-fluorinated analogues varying the para-position of the aniline 51, 53, 55-60, 62-66, 68-70, 72-73.^a



^aReagents and conditions: (a) R-H, K₂CO₃, MeCN, 100°C, 10-12 h; (b) H₂, Pd/C, MeOH or THF, 25-35°C, 2-12 h; (c) **75**, Pd(OAc)₂, BINAP, Cs₂CO₃, dioxane, μ W, 130°C, 0.5 h; (d) **75**, Brettphos Pd G3, Cs₂CO₃ or 'BuOLi, dioxane, μ W, 130°C, 0.5 h; (e) TFA, DCM, 25-35°C, 1-2 h; (f) for **60**, TBAF, THF, 25°C, 10 h.

Non-fluorinated analogues varying the para-position of the aniline were prepared starting with 176 (Scheme 6). S_NAr reactions with the appropriate amine nucleophile followed by nitro reduction yielded anilines 201-218. Subsequent Buchwald-Hartwig coupling with 75, and Boc or TBDMS deprotection where necessary, completed analogues 51, 53, 55-60, 62-66, 68-70, 72-73.



Scheme 7. Synthesis of fluorinated analogues varying the para-position of the aniline 52, 54, 61, 67, 71.ª

^aReagents and conditions: (a) R-H, K₂CO₃, MeCN, 60-100°C, 10-12 h; (b) for **234**, 2-(methylamino)ethan-1-ol, K₂CO₃, 60°C, 2 h, then TBDPSCl, imidazole, DMF, 25°C, 12 h; (c) CbzCl, K₂CO₃, THF, 25°C, 2-12 h; (d) Fe, NH₄Cl, EtOH, H₂O, 80-100°C, 2-12 h; (e) 1,2-diformylhydrazine, Me₃SiCl, Et₃N, pyridine, 100°C, 12 h; (f) H₂, Pd/C, MeOH, 25°C, 2-5 h; (g) **75**, Pd(OAc)₂, BINAP, Cs₂CO₃, dioxane, μ W, 130°C, 0.5-6 h; (h) TFA, DCM, 25°C, 1-3 h; (i) for **61**, TBAF, THF, 25°C, 2 h.

Fluorinated analogues varying the para-position of the aniline were synthesized starting with 231 (Scheme 7). S_NAr reactions with the appropriate amine nucleophile furnished 232-236 (234 included TBDPS protection of the alcohol group). The anilines were Cbz-protected, before the nitro groups were reduced to an aniline and condensed with 1,2-diformylhydrazine to afford the 1,2,4-triazole-containing compounds 247-251. The Cbz group was then removed via hydrogenation, and coupled with 75 in Buchwald-Hartwig coupling reactions to yield analogues 52, 54, 61, 67, and 71 after Boc or TBDPS deprotection.

EXPERIMENTAL SECTION

NanoBRET Assay.

Assays were run with a modified version of the previously published protocols^{15,19,24}. HEK293 cells were cultured at 37°C in 5% CO2 in Dulbecco's modified Eagle medium (DMEM; Gibco) supplemented with 10% fetal bovine serum (VWR/Avantor). A transfection complex of DNA at 10 µg/mL was created, consisting of 9 µg/mL carrier DNA (Promega) and 1 µg/mL CSNK2A-NLuc fusion DNA in Opti-MEM without serum (Gibco). FuGENE HD (Promega) was added at 30 µL/mL to form a lipid:DNA complex. The solution was then mixed and incubated at room temperature for 20 min. The transfection complex was mixed with a 20× volume of HEK293 cells in DMEM/FBS to arrive at a final concentration of 200,000 cells/mL, and 100 μ L/well was added to a 96-well plate that was incubated overnight at 37°C and 5% CO₂. The following day, the media were removed via aspiration and replaced with 85 µL of Opti-MEM without phenol red. A total of 5 µL per well of 20× NanoBRET Tracer K10 (Promega) at 10 µM for CSNK2A1 or 5 µM for CSNK2A2 in Tracer Dilution Buffer (Promega N291B) was added to all wells, except the "no tracer" control wells. Test compounds (10 mM in DMSO) were diluted 100× in Opti-MEM media to prepare stock solutions and evaluated at 11 concentrations. A total of 10 µL per well of the 10-fold test compound stock solutions (final assay concentration of 0.1% DMSO) were added. For "no compound" and "no tracer" control wells, DMSO in Opti-MEM was added for a final concentration of 1.1% across all wells; 96-well plates containing cells with NanoBRET Tracer K10 and test compounds (100 µL total volume per well) were equilibrated (37°C/5% CO₂) for 2 h. The plates were cooled to room temperature for 15 min. The NanoBRET NanoGlo substrate (Promega) at a ratio of 1:166 to Opti-MEM media in combination with an extracellular NLuc Inhibitor (Promega) diluted at 1:500 (10 µL of 30 mM stock per 5 mL of the Opti-MEM plus substrate) was combined to create a 3× stock solution. A total of 50 µL of the 3× substrate/extracellular NL inhibitor was added to each well. The plates were read within 30 min on a GloMax Discover luminometer (Promega) equipped with a 450 nm BP filter (donor) and 600 nm LP filter (acceptor) using 0.3 s of integration time. Raw milliBRET (mBRET) values were obtained by dividing the acceptor emission values (600 nm) by the donor emission values (450 nm) and multiplying by 1000. Averaged control values were used to represent complete inhibition (no tracer control: Opti-MEM + DMSO only) and no inhibition (tracer only control: no compound, Opti-MEM + DMSO + Tracer K10 only) and were plotted alongside the raw mBRET values. The data was first normalized and then fit using the Sigmoidal 4PL binding curve in Prism Software to determine IC₅₀ values.

NanoBRET Assay in Digitonin-Permeabilized Cells.

HEK293 cells were cultured at 37°C in 5% CO₂ in Dulbecco's modified Eagle medium (DMEM; Gibco) supplemented with 10% fetal bovine serum (VWR/Avantor). A transfection complex of DNA at 10 µg/mL was created, consisting of 9 µg/mL carrier DNA (Promega) and 1 µg/mL CSNK2A2-NLuc fusion DNA in Opti-MEM without serum (Gibco). FuGENE HD (Promega) was added at 30 µL/mL to form a lipid:DNA complex. The solution was then vortexed and incubated at room temperature for 20 min. The transfection complex was mixed with a 20× volume of HEK293 cells in DMEM/FBS to arrive at a final concentration of 200,000 cells/mL, and 100 µL was added to each well in a 96-well plate that was incubated overnight at 37°C and 5% CO₂. The following day, the media was removed via aspiration and replaced with 75 µL of Opti-MEM without phenol red. A total of 5 µL per well of 20× NanoBRET Tracer K10 (Promega) at 5 µM in Tracer Dilution Buffer (Promega N291B) was added to all wells except the "no tracer" control wells. Test compounds (10 mM in DMSO) were diluted 100× in Opti-MEM media to prepare stock solutions and evaluated at 11 concentrations. A total of 10 μ L per well of the 10-fold test compound stock solutions (final assay concentration of 0.1% DMSO) was added. For "no compound" and "no tracer" control wells, DMSO in Opti-MEM was added for a final concentration of 1.1% across all wells. A $10\times$ digitonin solution was prepared with Opti-MEM from a 400× stock solution (Promega). 10 µL of the 10× digitonin solution was then added to each well of the 96-well plate (50 µg/mL). The plate, now containing cells with NanoBRET Tracer K10, test compounds, and digitonin (100 μ L total volume per well), was then incubated at room temperature for a period no longer than 25 minutes. The NanoBRET NanoGlo substrate (Promega), at a ratio of 1:166 with Opti-MEM media, in combination with an extracellular NLuc Inhibitor (Promega) diluted at 1:500 (10 µL of 30 mM stock per 5 mL of the Opti-MEM plus substrate) was combined to create a 3× stock solution. A total of 50 µL of the 3× substrate/extracellular NLuc

inhibitor was added to each well. The plates were read within 30 min of substrate addition on a GloMax Discover luminometer (Promega) equipped with a 450 nm BP filter (donor) and 600 nm LP filter (acceptor) using 0.3 s of integration time. Raw milliBRET (mBRET) values were obtained by dividing the acceptor emission values (600 nm) by the donor emission values (450 nm) and multiplying by 1000. Averaged control values were used to represent complete inhibition (no tracer control: Opti-MEM + DMSO only) and no inhibition (tracer only control: no compound, Opti-MEM + DMSO + Tracer K10 only). The data was first normalized and then fit using the Sigmoidal 4PL binding curve in Prism Software to determine IC_{50} values.

In-cell Selectivity Profiling Using NanoBRET K192 Assay.

The K192 selectivity assay was run according to the Draft Promega technical manual, NanoBRETTM Target Engagement K192 Kinase Selectivity System. Reagents were supplied by Promega (Promega NP 4101). For the assay, DNA from the prepared kinase vector panel plates A&B were mixed with Fugene in 96 well plates (Corning 3917) and incubated at room temperature for 30 minutes. Control vectors used are the NanoLuc® Low control vector is pNL1.1.CMV [Nluc/CMV] Vector (Cat.# N1091) and the transfection control vector is NanoLuc®-HIPK2 Fusion Vector (Cat.# NV3221).

HEK293 cells were grown to 75-95% confluency in DMEM (Gibco 11995-065) supplemented with FBS (Avantor 97068-085) at 37°C in 5% RH. On the first day of the assay, cells were harvested and resuspended in Opti-MEM (Gibco 11058-021) supplemented with 1% FBS (Avantor 97068-085) at 2.5×10^5 cells per mL. 60 µL of cell suspension was mixed with 10 µL of prepared DNA (10X concentration) and 30 µL of Fugene (30 µL/mL in Opti-MEM) as outlined by Promega and incubated overnight at 37°C in a 5% CO₂ incubator.

On day two of the assay, 5 µL of 20X K10 tracer was prepared and added at concentrations recommended by Promega. Then 10 µL of compound **53** at 100 µM in Opti-MEM (diluted from a 10 mM solution in DMSO) was added to the test wells while an equivalent volume of Opti-MEM was added to the high-control wells. Plates were kept at 37°C in a 5% CO₂ incubator for two hours. After two hours, plates were allowed to equilibrate to room temperature for 15 minutes. A solution of 3X Complete Substrate plus Inhibitor Solution was freshly prepared, consisting of a 1:166 dilution of NanoBRETTM Nano-Glo® Substrate plus a 1:500 dilution of Extracellular NanoLuc® Inhibitor in Opti-MEM® medium without serum or phenol red. 50 µL of the 3X Complete Substrate plus Inhibitor Solution was added to each assay well, including control wells. After 2-3 minutes, the plate was shaken at 300 RPM for 10 seconds and the donor emission wavelength (450 nm) and acceptor emission wavelength (610 nm) were measured using the Glomax® Discover System.

As a quality check, the donor signal-to-background ratio was calculated for each individual kinase by dividing the mean donor signal for each kinase by the mean donor signal for the signal-to-background control wells. Fractional occupancy for the test drug for each kinase was determined using the following formula:

Occupancy (%) = $[1 - (Sample - Bottom) / (Top - Bottom)] \times 100$

Where:

Sample = Mean BRET value across all Sample (tracer + compound) wells for an individual kinase.

Top = Mean BRET value across all Top (tracer + vehicle) control wells for an individual kinase.

Bottom = Mean BRET value of NanoLuc® control wells (calculated either on a plate-by-plate basis or across the entire experiment).

MHV Assay.

DBT cells were cultured at 37°C in Dulbecco's modified Eagle medium (DMEM; Sigma-Aldrich) supplemented with 10% fetal bovine serum (Gibco) and penicillin and streptomycin (Sigma-Aldrich). DBT cells were plated in 96-well plates to be 80% confluent at the start of the assay. Test compounds or positive control EIDD-1931 were diluted to 15 μ M in DMEM. Serial 4-fold dilutions were made in DMEM, providing a concentration range of 15–0.22 μ M. Media were aspirated from the DBT cells, and 100 μ L of the diluted test compounds was added to the cells for 1 h at 37°C. After 1 h, MHV-nLuc5 was added at an MOI of 0.1 in 50 μ L of DMEM so that the final concentration of the first dilution of the compound was 10 μ M (T = 0). After 10 h, the media were aspirated, and the cells were washed with PBS and lysed with passive lysis buffer (Promega) for 20 min at room temperature. Relative light units (RLUs) were measured by using a luminometer

(Promega; GloMax). Triplicate data was analyzed in Graphpad Prism to generate IC_{50} values. A dose-response of EIDD-1931 was used as a positive control for the assay; each plate also contained a set of wells treated with EIDD-1931 at the IC_{50} for the assay (1.2 μ M).

Kinetic Solubility.

Phosphate buffered saline (50 mL, PBS, Fisher, pH 7.4) was added to HPLC grade H₂O (450 mL) for a total dilution factor of 1:10 and final PBS concentration of 1×. The test compound (6 μ L) as a 10 mM DMSO stock solution was combined with the aqueous PBS solution (294 μ L) for 50-fold dilution in a Millipore solubility filter plate with a 0.45 μ M polycarbonate filter membrane using a Hamilton Starlet liquid handler. The final DMSO concentration was 2.0%, and the maximum theoretical compound concentration was 200 μ M. The filter plate was heat-sealed for the duration of a 24 h incubation period. The sample was placed on a rotary shaker (200 rpm) for 24 h at ambient temperature (21.6–22.8°C) and then vacuum-filtered. All filtrates were injected into a chemiluminescent nitrogen detector for quantification. The equimolar nitrogen response of the detector was calibrated using standards that span the dynamic range of the instrument from 0.08 to 4500 μ g/mL nitrogen. The filtrates were quantified with respect to this calibration curve. The calculated solubility values were corrected for background nitrogen present in DMSO and the media used to prepare the samples.

SARS-CoV-2 Assay.

Human lung epithelial A549-ACE2 cells were cultured in DMEM containing 10% heat-inactivated FBS, nonessential amino acids, and pen strep. A549-ACE2 cells were seeded at 20,000 cells per well in a 96-well solid black plate 1 day prior to infection. To assay drug effect, cells were pretreated with drug for 1 h and then infected with SARS-CoV-2, with drug maintained during the infection. Then, 2 h after infection, the supernatant was removed, monolayers were rinsed with PBS, and media containing drug was added to each well. At 48 h post infection start, Nano-glo was added to each well as per the manufacturer's protocol (Promega), and RLUs were measured using a Promega GloMax.

LDH Cytotoxicity Assay.

DBT cells were plated to be 80% confluent at the start of the assay. Compounds were diluted as done for the MHV assay and incubated with cells at 37 °C for 1 h. After 1 h, 50 μ L of DMEM was added to the cells (T = 0); 45 min before harvest, lysis buffer was added to positive wells. LDH activity in cell-free supernatants was measured at 10 h after infection using the Sigma Tox7 kit as per the manufacturer's directions. A549-ACE2 cells were seeded at 20,000 cells per well 1 day prior to infection in 96-well plates. Cells were pretreated for 1 h and then mock-infected. Then, 2 h post-mock infection, the media was removed, the monolayer was rinsed one time with PBS, and media containing drug was added to each well. Typically, 48 h after mock infection, plates were centrifuged, and an aliquot of the cell culture supernatant was removed. For LDH assays using Sigma Tox7 kit, the clarified supernatant was transferred to a clean plate and assayed following the manufacturer's protocol.

Cell-Titer-Glo Cytotoxicity Assay.

A549-ACE2 cells were maintained in low-glucose DMEM (Gibco) supplemented with 10% FBS, 1% NEAA, 1% Lglutamine. No antibiotics were used. Cells were plated at 2000 cells/well in 384-well plate (Costar) and incubated overnight (37°C, 5% CO₂) before adding compound. Compounds were added in quadruplicate and incubated for 48 h. DMSO percentage was constant across all concentrations of compound. Cell viability was measured using CellTiter-Glo2 (Promega) and luminescence signal was read on a GloMax plate reader (Promega). Dose response analysis was performed using GraphPad Prism.

MDCK-MDR1 Permeability Assay.

 $50 \ \mu\text{L}$ and $25 \ \text{mL}$ of cell culture medium were added to each well of the Transwell insert and reservoir, respectively. The HTS transwell plates were incubated at 37 °C, 5% CO₂ for 1 hour before cell seeding. MDCK-MDR1 cells were diluted to 1.56×10^6 cells/mL with culture medium. $50 \ \mu\text{L}$ of cell suspension were dispensed into the filter well of the 96-well HTS Transwell plate. Cells were cultivated for 3-8 days in a cell culture incubator at 37 °C, 5% CO₂, 95% relative humidity. Cell culture medium was replaced every other day, beginning no later than 24 hours after initial plating. Cell monolayer integrity was verified before the assay. Media was removed from the reservoir and each Transwell insert and replaced with prewarmed fresh culture medium. Transepithelial electrical resistance (TEER) across the monolayer was measured using Millicell Epithelial Volt-Ohm measuring system (Millipore, USA) and the plate was returned to the incubator after measurement. TEER was calculated by the following equation: TEER measurement (ohms) × Area of membrane (cm²) = TEER value (ohm•cm²). A well-qualified MDCK-MDR1 monolayer was defined by a TEER value greater than 42 ohm•cm². Prior to the assay, the MDCK-MDR1 plate was removed from the incubator and washed twice with pre-warmed HBSS (10 mM HEPES, pH 7.4), and then incubated at 37°C for 30 minutes.

10 mM stock solutions of compound 53 and positive controls metoprolol, prazosin, and imatinib were prepared in DMSO. The stock solutions of test compounds were diluted in DMSO to 0.2 mM and then diluted with HBSS (10 mM HEPES, pH 7.4) to 1 μ M working solutions. The final concentration of DMSO in the incubation system was 0.5%. To determine the rate of drug transport in the apical to basolateral direction, 75 μ L of 1 μ M working solution of test compound was added to the Transwell insert (apical compartment) and the wells in the receiver plate (basolateral compartment) were filled with 235 µL of HBSS (10 mM HEPES, pH 7.4). To determine the rate of drug transport in the basolateral to apical direction, 235 µL of 1 µM working solution of test compound was to the receiver plate wells (basolateral compartment) and then the Transwell inserts (apical compartment) were filled with 75 µL of HBSS (10 mM HEPES, pH 7.4). The assay was performed in duplicate. Time 0 samples were prepared by transferring 50 µL of 1 µM working solution to wells of the 96deepwell plate, followed by the addition of 200 µL cold methanol containing appropriate internal standards (100 nM alprazolam, 200 nM labetalol, 200nM caffeine and 200 nM diclofenac). The plates were incubated at 37 °C for 2 hours. At the end of the incubation, 50 μ L samples from donor sides (apical compartment for Ap \rightarrow Bl flux, and basolateral compartment for $Bl \rightarrow Ap$) and receiver sides (basolateral compartment for $Ap \rightarrow Bl$ flux, and apical compartment for $Bl \rightarrow Ap$) were transferred to wells of a new 96-well plate, followed by the addition of 4 volumes of cold methanol containing appropriate internal standards (100 nM alprazolam, 200 nM labetalol, 200 nM caffeine and 200 nM diclofenac). Samples were vortexed for 5 minutes and then centrifuged at 3.220 g for 40 minutes. An aliquot of 100 μ L of the supernatant was mixed with 100 μ L of ultra-pure water. The samples were analyzed by LC-MS/MS.

To determine the Lucifer yellow leakage after 2 hour transport period, a stock solution of Lucifer yellow was prepared in water and diluted with HBSS (10 mM HEPES, pH 7.4) to reach the final concentration of 100 μ M. 100 μ L of the Lucifer yellow solution was added to each Transwell insert (apical compartment), followed by filling the wells in the receiver plate (basolateral compartment) with 300 μ L of HBSS (10 mM HEPES, pH 7.4). The plates were incubated at 37 °C for 30 mins. 80 μ L samples were removed directly from the apical and basolateral wells (using the basolateral access holes) and transferred to wells of new 96 wells plates. The Lucifer yellow fluorescence (to monitor monolayer integrity) signal was measured in a fluorescence plate reader at 485 nM excitation and 530 nM emission. Lucifer yellow leakage was <1% for all compounds, indicating a well-qualified MDCK-MDR1 monolayer.

The apparent permeability coefficient (P_{app}), in units of cm/s, was calculated using the following equation:

 $P_{app} = (V_A \times [drug]_{acceptor}) / (Area \times Time \times [drug]_{initial,donor})$

Where V_A is the volume (in mL) in the acceptor well, Area is the surface area of the membrane (0.143 cm² for Transwell-96 Well Permeable Supports), and Time is the total transport time in seconds.

The efflux ratio was determined using the following equation:

 $Efflux \ ratio = P_{app(B-A)} / P_{app(A-B)}$

Plasma Protein Binding Assay.

Frozen human plasma (BioIVT, MSE483763, stored at -80°C) was thawed in a 37°C water bath. Working solutions of compound **53** and positive control compound ketoconazole were prepared in DMSO at a concentration of 200 μ M, then spiked into plasma to achieve a final compound concentration of 1 μ M. The final concentration of DMSO was 0.5%.

For plasma protein binding analysis, dialysis membranes were soaked in ultra-pure water for 60 minutes to separate strips, then in 20% ethanol for 20 minutes, finally in dialysis buffer for 20 minutes. The dialysis set up was assembled according to the manufacturer's protocol. Each cell containing 150 μ L of plasma sample was dialyzed against equal volume of dialysis buffer (PBS). The dialysis plate was sealed and incubated in an incubator at 37°C with 5% CO₂ at 100 rpm for 6 hours. At the end of incubation, 50 μ L of samples from both buffer and plasma chambers were transferred to wells of a 96-well plate. 50 μ L of plasma was added to each buffer sample and an equal volume of PBS was supplemented to the collected plasma sample. 400 μ L of quench solution (acetonitrile containing internal standards 200 nM labetalol, 100 nM tolbutamide and 100 nM ketoprofen) was added to precipitate protein and release compounds. Samples were vortexed for 2 minutes and centrifuged for 30 minutes at 3,220 g. 100 μ L aliquots of the supernatant were diluted by 100 μ L ultra-pure water, and the mixtures were used for LC-MS/MS analysis. The assay was performed in duplicate.

Plasma stability analysis was performed in parallel. 50 μ L of spiked plasma sample was transferred to a new plate. The samples are incubated at 37°C in an incubator with 5% CO₂ for 0 and 6 hours. At designated time points, 50 μ L of PBS was added and mixed thoroughly. 400 μ L of quench solution (acetonitrile containing internal standards 200 nM labetalol, 100 nM tolbutamide and 100 nM ketoprofen) was added to precipitate protein and release compounds. Samples were vortexed for 2 minutes and centrifuged for 30 minutes at 3,220 g. 100 μ L aliquots of the supernatant were diluted by 100 μ L ultrapure water, and the mixtures were used for LC-MS/MS analysis. **53** was stable in plasma.

The concentrations of test compounds in the buffer and plasma chambers were determined from peak area ratios. The percentages of bound compound were calculated as follows:

% Free = (Peak Area Ratio buffer chamber / Peak Area Ratio plasma chamber) × 100%

% Bound = 100% – % Free

% Recovery = (Peak Area Ratio buffer chamber + Peak Area Ratio plasma chamber) / Peak Area Ratio total sample × 100%

% Stability at 6 hours = Peak Area Ratio $_{T=6 \text{ sample}}$ /Peak Area Ratio $_{T=0 \text{ sample}} \times 100 \%$

Lung Protein Binding Assay.

Frozen mouse lung tissue homogenate (Pharmaron, PH-Mouse-20240318, stored at -80°C) was thawed in a 37°C water bath. Working solutions of compound **53** and positive control compound ketoconazole were prepared in DMSO at a concentration of 200 μ M, then spiked into lung tissue homogenate to achieve a final compound concentration of 1 μ M.

Dialysis membranes were soaked in ultra-pure water for 60 minutes to separate strips, then in 20% ethanol for 20 minutes, finally in dialysis buffer for 20 minutes. The dialysis set up was assembled according to the manufacturer's protocol. Each cell containing 150 μ L of lung tissue homogenate sample was dialyzed against equal volume of dialysis buffer (100 mM PBS, pH 7.4). The dialysis plate was sealed and incubated in an incubator at 37°C with 5% CO₂ at 100 rpm for 6 hours. At the end of incubation, 50 μ L of samples from both buffer and lung tissue homogenate were transferred to wells of a 96-well plate. 50 μ L of lung tissue homogenate sample. 400 μ L of precipitation buffer (acetonitrile containing internal standards 200 nM Alprazolam, 200 nM Labetalol, 200 nM Imipramine and 2 μ M Ketoplofen) was added to precipitate protein and release compounds. Samples were vortexed for 2 minutes and centrifuged for 30 minutes at 3,220 g. 100 μ L aliquots of the supernatant were diluted by 100 μ L ultra-pure water, and the mixtures were used for LC-MS/MS analysis. The assay was performed in duplicate.

The concentrations of test compounds in the buffer and lung tissue homogenate chambers were determined from peak area ratios. The percentages of bound compound were calculated as follows:

 $Fu_{measured} = (Peak Area Ratio _{buffer chamber} / Peak Area Ratio _{plasma chamber})$

 $Fu_{undiluted} = \frac{1/8}{\left(\left(\frac{1}{Fu_{measured}}\right) - 1\right) + 1/8}$ % Bound = 100% - Fu_{undiluted} × 100%

Liver Microsomal Stability Assay.

Compounds as 10 mM DMSO stock solutions were diluted to 2.5 mM with DMSO and again to 0.5 mM with MeCN to give a final solution containing a 0.5 mM compound in 1:4 DMSO/ MeCN. Liver microsomes from male CD-1 mice were sourced from Xenotech (Kansas City, KS). A reaction plate was prepared by adding 691.25 µL and prewarmed (37°C) microsomal solution (0.63 mg/mL protein and 1.3 mM EDTA in potassium phosphate buffer made by mixing ~250mL of 100 mM K₂HPO₄ with ~65mL of KH₂PO₄ until the buffer reaches a pH of 7.4) to an empty well of a 96-well plate and maintained at 37°C. The diluted 0.5 mM compound (8.75 µL) was added to the microsomal solution in the reaction plate and mixed thoroughly by repeated pipetting to give a final assay concentration of 5.0 μ M. The resulting solutions were preincubated for 5 min at 37° C and then dispensed into T = 0 and incubation plates. For the T = 0 plates, an aliguot (160 µL) of each reaction solution was added to an empty well of a 96-well plate as an exact replicate of the reaction plate. Cold (4°C) MeOH (400 µL) was added to each well and mixed thoroughly by repeated pipetting. NADPH regeneration solution (40 μ L) was added to each well and mixed thoroughly by repeated pipetting. For the T = 30 min incubation plate, NADPH $(95 \,\mu L)$ was added to the remaining solution (microsomes + test compound) in each well in the previously prepared reaction plate to initiate the reaction. The plate was sealed and incubated at 37°C for 30 min. An aliquot (100 µL) was removed from each well at the desired time point and dispensed into a well of a 96-well plate. Cold (4°C, 200 µL) MeOH was added to quench the reaction. All plates were sealed, vortexed, and centrifuged at 3000 rpm, 4°C for 15 min, and the supernatants were transferred for analysis by LC-TOFMS. The supernatant (20 µL) was injected onto an AOUASIL C18 column and eluted using a fast-generic gradient program. TOFMS data was acquired using Agilent 6538 Ultra High Accuracy TOF MS in extended dynamic range (m/z 100-1000) using generic MS conditions in positive mode. Following data acquisition, exact mass extraction and peak integration were performed using MassHunter Software (Agilent Technologies). The stability of the compound was calculated as the percent remaining of the unchanged parent at T = 30 min relative to the peak area at T = 0 min.

To determine CL_{int} , aliquots of 50 µL were taken from the reaction solution at 0, 15, 30, 45, and 60 min. The reaction was stopped by the addition of four volumes of cold MeCN with IS (100 nM alprazolam, 200 nM imipramine, 200 nM labetalol, and 2 µM ketoprofen). Samples were centrifuged at 3,220 g for 40 min, and 90 µL of the supernatant was mixed with 90 µL of ultrapure H₂O and then used for LC-MS/MS analysis. Peak areas were determined from extracted ion chromatograms, and the slope value, k, was determined by linear regression of the natural logarithm of the remaining percentage of the parent drug vs incubation time curve. The intrinsic clearance (CL_{int} in µL/min/mg) was calculated using the relationship $CL_{int} = kV/N$ where V is the incubation volume and N is the amount of protein per well.

Hepatocyte Stability Assay.

Human cryopreserved hepatocytes were supplied by BioIVT (lot QZW, 10 pooled donors). Mouse cryopreserved hepatocytes were supplied by BioIVT (lot ZPG, pooled male CD-1). Vials of cryopreserved hepatocytes were removed from storage and thawed in a 37°C water bath with gentle shaking, and then, the contents were poured into a 50 mL thawing medium conical tube. Vials were centrifuged at 100 g for 10 min at room temperature. The thawing medium was aspirated, and hepatocytes were resuspended with a serum-free incubation medium to yield $\sim 1.5 \times 10^6$ cells/mL. Cell viability and density were counted using AO/PI fluorescence staining, and then, cells were diluted with a serum-free incubation medium to a working cell density of 0.5×10^6 viable cells/mL. Aliquots of 198 µL of hepatocytes were dispensed into each well of a 96-well noncoated plate. The plate was placed in an incubator for approximately 10 min. Aliquots of 2 µL of the 100 µM test compound in duplicate and positive control were added into the respective wells of the noncoated 96-well plate to start the reaction. The final concentration of the test compound was 1 µM. The plate was placed in an incubator for the designed time points. Contents (25 μ L) were transferred and mixed with six volumes (150 μ L) of cold MeCN with internal standards (100 nM alprazolam, 200 nM labetalol, 200 nM caffeine, and 200 nM diclofenac) to terminate the reaction at time points of 0, 15, 30, 60, 90, and 120 min. Samples were centrifuged for 45 min at 3,220 g, an aliquot of 100 μ L of the supernatant was diluted with 100 μ L of ultrapure H₂O, and the mixture was used for LC-MS/MS analysis. Peak areas were determined from extracted ion chromatograms, and the slope value, k, was determined by linear regression of the natural logarithm of the remaining percentage of the parent drug vs incubation time curve. The intrinsic clearance (CL_{int} in $\mu L/min/10^6$ cells) was calculated using the relationship $CL_{int} = kV/N$ where V is the incubation volume (0.2 mL) and N is the number of hepatocytes

per well (0.1 \times 10⁶ cells). Scaling factors to convert CL_{int} from μ L/min/10⁶ cells to mL/min/kg were 2540 (human hepatocytes) and 11,800 (mouse hepatocytes).

Pharmacokinetics.

In the 5-hour PK study, male CD-1 mice (6-8 weeks, 20-30 g) were dosed by intravenous (i.v.), oral (p.o.), or intraperitoneal (i.p.) administration of compound **53**. For i.v. administration, a single dose of compound **53** (3 mg/kg) as a formic acid salt was administered as 5 mL/kg of a 0.6 mg/mL solution in NMP/Solutol/PEG-400/normal saline (v/v/v/v, 10:5:30:55) to two mice. For p.o. administration, a single dose of compound **53** (10 mg/kg) was administered as 10 mL/kg of a 1 mg/mL solution in NMP/Solutol/PEG-400/normal saline (v/v/v/v, 10:5:30:55) to two mice. For i.p. administration, a single dose of compound **53** (10 mg/kg) was administered as 10 mL/kg of a 1 mg/mL solution in NMP/Solutol/PEG-400/normal saline (v/v/v/v, 10:5:30:55) to two mice. For i.p. administration, a single dose of compound **53** (10 or 30 mg/kg) was administered as 10 mL/kg of a 1 or 3 mg/mL solution in NMP/Solutol/PEG-400/normal saline (v/v/v/v, 10:5:30:55) to two mice. For i.p. administration, a single dose of compound **53** (10 or 30 mg/kg) was administered as 10 mL/kg of a 1 or 3 mg/mL solution in NMP/Solutol/PEG-400/normal saline (v/v/v/v, 10:5:30:55) to two mice each cohort. The mice had free access to water and food. At 0.5, 1, 3 and 5 hour post-dose, 0.03 mL of blood was collected from the dorsal metatarsal vein of each mice. Blood of each sample was transferred into plastic microcentrifuge tubes containing anticoagulant of EDTA-K₂. Blood samples were centrifuged at 4,000 g for 5 minutes at 4°C to obtain plasma. The samples were stored in a freezer at -75±15°C prior to analysis. Plasma samples from the two mice of each cohort and each time point were pooled together for analysis.

In the 24-hour PK study, male CD-1 mice (6-8 weeks, 20-30 g) were dosed by intraperitoneal (i.p.) administration of compound **53** (10 mg/kg) as a formic acid salt by administration of 10 mL/kg of a 1 mg/mL solution in NMP/Solutol/PEG-400/normal saline (v/v/v/v, 10:5:30:55) to 21 mice. The mice had free access to water and food. At 0.5, 1, 2, 4, 8, 12, and 24 hour post-dose, 0.15 mL of blood was collected by cardiac puncture from three mice at each time point. Blood of each sample was transferred into plastic microcentrifuge tubes containing anticoagulant of EDTA-K₂ and mixed well with anticoagulant, then placed on ice prior to centrifugation at 4000 g for 5 minutes at 4°C to obtain plasma. The samples were stored in a freezer at $-75\pm15^{\circ}$ C prior to analysis. The three mice at each time point were anaesthetized by a rising concentration of CO₂ and lung samples collected. Lung samples were quick frozen in ice box and stored at $-75\pm15^{\circ}$ C. Prior to analysis, all lung samples were weighed and homogenized with phosphate buffered saline (PBS) by lung weight (g) to buffer volume (mL) ratio of 1:3. The final compound concentration was calculated by multiplying the detected concentration by the dilution factor of four.

Concentrations of the test compound in the plasma samples were determined using a Shimadzu LC-MS/MS system with a LC-40D X3 CN pump, DGU-405 degasser, CBM-40 CN system controller, SIL-40C X3CN autosampler, CTO-40C CN column oven with a HALO 160A ES-C18, (2.7μ m, 2.1×50 mm) column, and AB API 5500+ MS instrument. The mobile phase was 5-95% MeCN in H₂O, with 0.1% formic acid. The desired serial concentrations of working solutions were achieved by diluting stock solution of analyte with 50% acetonitrile in water solution. 10 µL of working solutions (1, 2, 5, 10, 50, 100, 500, 1000, 5000 ng/mL) were added to 10 µL of blank plasma or lung homogenate to achieve calibration standards of 1, 2, 5, 10, 50, 100, 500, 1000 ng/mL in a total volume of 20 µL. 5 quality control samples at 2 ng/mL, 5 ng/mL, 10 ng/mL, 100 ng/mL and 4000 ng/mL were prepared independently of those used for the calibration curves in the same manner. 20 µL standards, 20 µL QC samples and 20 µL unknown samples (10 µL plasma or lung homogenate with 10 µL blank solution) were added to 200 µL of acetonitrile containing internal standard mixture for precipitating protein respectively. Then the samples were vortexed for 30 s. After centrifugation at 4°C, 4000 rpm for 15 min, the supernatant was diluted with ultrapure water at a ratio of 1:2 (v/v, 1:2), then 20 µL of diluted supernatant was injected into the LC/MS/MS system for quantitative analysis. PK parameters were calculated from the mean plasma concentration versus time by a noncompartmental model using WinNonlin 8.3 (Phoenix).

In vivo SARS-CoV-2 efficacy.

All mouse studies were conducted under protocols approved by the Institutional Animal Care and Use Committee (IACUC) at the University of North Carolina at Chapel Hill.

CD-1 mice (female, 8-10 week old) received compound **53** dosed 10 mg/kg i.p. in vehicle (NMP/Solutol/PEG-400/normal saline (v/v/v/v, 10:5:30:55). Mice were dosed every 12 hours for a total of 36 hours (3 doses). 12 hours after the first dose, mice were anesthetized with 50 mg/kg ketamine + 5 mg/kg xylazine and intranasally infected with 1x10⁴

plaque forming units (pfu) of mouse adapted coronavirus, SARS-CoV-2 MA10^{20,40}, contained in a 50 μ L volume which was pipetted into the nares of each mouse. Post-challenge, mice were monitored, weighed, and scored for clinical signs and euthanized at 24 hours post-infection. Mice were euthanized by an overdose of isoflurane anesthesia (Baxter), blood was collected by cardiocentesis, and lung lobes collected downstream analysis.

Infectious viral loads were measured by plaque assay. One day prior to assay, Vero cells were seeded at 2×10^5 cells per well in 12-well plates. Titers were measured from superior and middle lung lobes that were homogenized in 0.5 mL of media (DMEM + 5% FBS + 1% L-glutamine) at 6000 rpm for 40 sec using a Roche MagNA Lyser homogenizer. Cell debris was removed by centrifugation for 1 min at full speed. 50 µL of the supernatant of the clarified homogenate was added to 450 µL of dilution media (DMEM + 5% FBS + 1% L-glutamine media). Homogenates were used to create tenfold serial dilutions (10^{-1} to 10^{-6}). Approximately 200 µL of each dilution was pipetted onto the previously plated Vero cells and incubated at 37°C. To ensure even distribution across each well, the plates were rocked every 15 minutes. After 1 hour, 2 mL of overlay (50:50 mixture of 2.5% carboxymethylcellulose and 2X alpha MEM containing 6% FBS + 2% penicillin/streptomycin + 2% L-glutamine + 2% HEPES) was added to each well. After incubation for 4 days in 37°C, 5% CO₂ an equal volume of 4% paraformaldehyde was added to each well and the cells were allowed to fix overnight. The fixative was removed, wells were rinsed with water to remove residual overlay and 0.25% crystal violet was added to each well. Visible plaques were counted and averaged between two technical replicate wells and used to calculate plaque forming units (pfu) per lung tissue. The limit of detection (LOD) for the assay was determined to be 12.5 pfu / lung tissue, and samples that yielded no plaques were assigned a value of 6.25, half of the LOD.

In vivo CK2 inhibition.

All mouse studies were conducted under protocols approved by the Institutional Animal Care and Use Committee (IACUC) at the University of North Carolina at Chapel Hill.

CD-1 mice (female, 8-10 week old) received compound **53** dosed 10 mg/kg i.p. in vehicle (NMP/Solutol/PEG-400/normal saline (v/v/v, 10:5:30:55). Mice were dosed every 12 hours for a total of 36 hours (3 doses). At 36 hours after the first dose, mice were euthanized by an overdose of isoflurane (Baxter), blood collected by cardiac puncture and lung lobes collected for downstream processing and analysis. Recovered blood was placed in EDTA tubes, spun at 5000 xg in a microcentrifuge and the recovered plasma transferred to a clean tube and frozen at -80°C until analysis. The left lung lobe was added to a 2 mL O-ring skirted tube containing 750 μ L of 1×PBS containing Phos Stop (per manufacturers recommendation, Roche) and glass beads. Lungs were homogenized for 60 seconds at 6000 rpm in a Roche Magnalyzer. Homogenates were centrifuged for 5 minutes at 10,000 rpm and 500 μ L of the homogenate transferred to a clean tube and frozen at -80°C until analysis.

Halt Protease Inhibitor cocktail (ThermoFisher, 78429) was added to mouse lung homogenates prior to sonication at 40% for 10 seconds on ice. Samples were quantified using Piece Rapid Gold BCA Protein Assay kit (ThermoFisher, A53226) and 50 µg protein was run on 4-20% Tris-Glycine gels and transferred to PVDF. Blots were blocked in 5% Milk in 1X TBST (0.1% Tween 20) for 1 hour at room temperature, washed with 1X TBST, and incubated with primary antibodies in 5% BSA in 1X TBST for 10 hours at room temperature; 1:10,000 phospho-EIF2S2 P-S2 (provided by Laszlo Gyenis from the David Litchfield group), 1:200 EIF2S2 (Novus Biologicals, H00008894-M09), 1:1000 phospho-AKT Ser129 (Cell Signaling Technology, 13461), 1:1000 AKT (Cell Signaling Technology, 2920), 1:1000 GAPDH (Proteintech, 10494-1-AP), 1:1000 Transferrin (Proteintech, 17435-1-AP). Bands were quantified using Fiji⁴¹ and normalized to GAPDH. Phosphorylated protein was then normalized to its corresponding total protein. GraphPad Prism version 9.4.1 for macOS was used to plot averages (N=3 for each treatment) and assess statistical significance using an unpaired t-test with Welch's correction.

Crystallography.

CSNK2A1 expression and purification were performed as described previously^{19,42,43}. Briefly, transformed BL21(DE3) cells were grown in Terrific Broth medium containing 50 mg/mL kanamycin. Protein expression was induced at an OD₆₀₀ of 2 by using 0.5 mM isopropyl-thio-galactopyranoside (IPTG) at 18°C for 12 hours. Cells expressing His6-tagged

CSNK2A1 were lysed in lysis buffer containing 50 mM HEPES pH 7.5, 500 mM NaCl, 25 mM imidazole, 5% glycerol, and 0.5 mM Tris(2 carboxyethyl)phosphine (TCEP) by sonication. After centrifugation, the supernatant was loaded onto a Nickel-Sepharose column equilibrated with 30 mL lysis buffer. The column was washed with 60 mL lysis buffer. Proteins were eluted by an imidazole step gradient (50, 100, 200, 300 mM). Fractions containing protein were pooled together and dialyzed overnight using 1 L of final buffer (25 mM HEPES pH 7.5, 500 mM NaCl, 0.5 mM TCEP) at 4°C. Additionally, TEV protease was added (protein:TEV 1:20 molar ratio) to remove the tag. The next day the protein solution was loaded onto Nickel-Sepharose column beads again to remove the TEV protease and cleaved Tag. The combined flow through fraction and the wash fraction (25 mM imidazole) containing the protein were concentrated to approximately 4-5 mL and loaded onto Superdex 75 16/60 Hi-Load gel filtration column equilibrated with final buffer. The protein was concentrated to approximately 9 mg/mL.

CSNK2A1 was crystallized using the sitting-drop vapor diffusion method by mixing protein (9 mg/mL) and well solutions in 2:1, 1:1, and 1:2 ratios. The reservoir solution contained 0.2 M ammonia sulphate, 0.1 M bis-tris pH 5.5 and between 23-26% (v/v) PEG 3350. Complex structures were achieved by soaking grown apo crystals for at least 24 h with the desired inhibitor dissolved in reservoir solution. Final concentration of the inhibitor was approximately 0.5 mM.

Diffraction data were collected at beamline X06SA (Villigen, CH) at a wavelength of 1.0 Å at 100 K. The reservoir solution supplemented with 20% ethylene glycol was used as cryoprotectant. Data were processed using XDS⁴⁴ and scaled with aimless⁴⁵. The PDB structure with the accession code 6Z83¹⁹ was used as an initial search MR model using the program MOLREP⁴⁶. The final model was built manually using Coot⁴⁷ and refined with REFMAC5⁴⁸. Data collection and refinement statistics are summarized in Table S2.

General Chemistry Methods.

All chemical reagents were commercially available except those whose synthesis is described below. All reaction mixtures and column eluents were monitored via analytical thin-layer chromatography (TLC) performed on precoated fluorescent silica gel plates, 200 µm with an F254 indicator; visualization was accomplished by UV light (254/365 nm). LC-MS measurements were determined on Shimadzu LC-AB + LCMS-2020, Shimadzu LC-AD + LCMS-2020, Shimadzu LC-AD xR + LCMS-2020, or Agilent 1200 + Infinitylab LC/ MSD instruments. Purity was determined by HPLC measurement using a Shimadzu LC-20 + LCMS-2020 instrument fitted with an Agilent PoroShell 120 EC-C18 column $(45^{\circ}C, 2.7 \,\mu\text{m}, 3.0 \times 50 \,\text{mm})$; 8 min chromatography ran 0.037% TFA in water/MeCN (19:1) (solvent A), 0.018% TFA/MeCN (solvent B), and gradient 0–60% (solvent B) over 6.0 min, held at 60% for 1.0 min, and returned to 0% (solvent B) for 1.0 min at a flow rate of 1.0 mL/min; 4 min chromatography ran 0.037% TFA in water/MeCN (19:1) (solvent A), 0.018% TFA/MeCN (solvent B), and gradient 10-80% (solvent B) over 3.0 min, held at 80% for 0.5 min, and returned to 0% (solvent B) for 0.5 min at a flow rate of 1.0 mL/min. All final compounds were >95% pure unless otherwise stated. Nuclear magnetic resonance (NMR) spectra were obtained on Bruker Avance Neo 400 MHz, Bruker Avance Neo 500 MHz, and Bruker Avance 850 MHz instruments. Chemical shifts are reported in parts per million (ppm, δ), with residual solvent peaks referenced as the internal standard. Coupling constants are reported in Hz. Spin multiplicities are described as s (singlet), br s (broad singlet), d (doublet), t (triplet), q (quartet), p (pentet), and m (multiplet). Data were processed using MestReNova.

Experimental Procedures in Schemes 1 and 2.

5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**). To a solution of 5,7dichloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (**74**) (5.00 g, 23.47 mmol, 1 *eq*) in EtOH (30 mL) was added cyclopropylamine (12.00 g, 211.24 mmol, 14 mL, 9 *eq*) dropwise at 25°C. Then the mixture was stirred at 25°C for 2 hours. The reaction mixture was filtered and the solid was washed with EtOH (4 mL x2). **75** (5.47 g, 23.03 mmol, 98.1% yield) was obtained as a yellow solid. ¹H NMR (400 MHz, MeOD-*d*₄) δ 8.36 (s, 1H), 6.61 (s, 1H), 2.76 (m, 1H), 1.04 - 0.93 (m, 2H), 0.83 - 0.74 (m, 2H). LCMS t_R = 0.500 min in 1 min chromatography, Chromolith @ Flash RP-18e,25-3mm, MS ESI calcd. for C₁₀H₉ClN₅⁺ [M+H]⁺ m/z 234.05, found 234.0. *N*-(*3*-(*ycano*-7-(*cyclopropylamino*)*pyrazolo*[*1*,*5*-*a*]*pyrimidin*-*5*-*y*]*amino*)*phenyl*)*acetamide* (**3**). To a solution of N-(3-aminophenyl)acetamide (**76**) (100 mg, 665.88 μmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-*a*]pyrimidine-3carbonitrile (**75**) (124 mg, 532.70 μmol) in dioxane (4 mL) was added Cs₂CO₃ (651 mg, 2.00 mmol), BINAP (62 mg, 99.88 µmol) and Pd(OAc)₂ (22 mg, 99.88 µmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The residue was purified by flash silica gel chromatography (eluent of 0~4%, MeOH/DCM) to give the product. The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water(NH₄HCO₃)-ACN];B%: 16%-56%,36min). **3** (45.7 mg, 19.7% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-d₆) δ 9.93 (s, 1H), 9.66 (s, 1H), 8.35 (s, 1H), 8.21 (s, 1H), 7.85 – 7.75 (m, 2H), 7.24 (t, *J* = 8.0 Hz, 1H), 7.08 (d, *J* = 8.0 Hz, 1H), 6.06 (s, 1H), 2.59 (tt, *J* = 7.0, 3.7 Hz, 1H), 2.05 (s, 3H), 0.84 – 0.77 (m, 2H), 0.74 – 0.68 (m, 2H). ¹³C NMR (101 MHz, DMSO-d₆) δ 168.32, 156.97, 150.86, 148.31, 145.11, 140.51, 139.65, 128.91, 114.87, 114.52, 113.26, 110.48, 76.45, 76.42, 24.07, 23.32, 6.55. HPLC t_R = 3.332 min in 8 min chromatography, purity 100.0%. LCMS t_R = 1.741 min in 4 min chromatography, MS ESI calcd. for C₁₈H₁₈N₇O⁺ [M+H]⁺ 348.16, found 348.1.

N-(3-((3-cyano-7-(cyclopropylamino)))) pyrazolo[1,5-a]pyrimidin-5-yl)amino) phenyl) methanesulfonamide (4). To a solution of N-(3-aminophenyl)methanesulfonamide (77) (200)mg, 1.07 mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (201 mg, 0.86 mmol) in dioxane (8 mL) was added Cs₂CO₃ (1.40 g, 4.30 mmol), BINAP (100 mg, 0.16 mmol) and Pd(OAc)₂ (36 mg, 0.16 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 40 min. The mixture was filtered and the filtrate was concentrated to dryness. The residue was purified by silica gel chromatography (eluent of 0%~6%, MeOH/DCM) and then purified by prep-HPLC (column: Welch SiO₂ 10u 250*200mm; mobile phase: [Heptane-EtOH (0.1%NH₃H₂O)]; B%: 20%-60%, 15min). The residue was further purified by prep-HPLC (column: Welch Xtimate C18 150*40mm*10um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 16%-56%, 30min). 4 (80 mg, 19.4% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.73 (s, 1H), 9.12 (br s, 1H), 8.37 (s, 1H), 8.27 (s, 1H), 7.65 (dd, J = 8.2, 2.0 Hz, 1H), 7.50 (t, J = 2.1 Hz, 1H), 7.28 (t, J = 8.1 Hz, 1H), 6.82 (dd, J = 8.1, 2.1 Hz, 1H), 6.02 (s, 1H), 3.04 (s, 3H), 2.60 (tt, J = 7.0, 3.6 Hz, 1H), 0.86 – 0.78 (m, 2H), 0.78 – 0.65 (m, 2H). ¹³C NMR (101 MHz, DMSO- d_6) δ 156.78, 150.75, 148.35, 145.18, 141.05, 138.76, 129.54, 114.95, 114.81, 113.88, 110.64, 76.60, 76.58, 39.38, 23.33, 6.56. HPLC $t_R = 3.953$ min in 8 min chromatography, purity 99.6%. LCMS $t_R = 1.780$ min in 4 min chromatography, MS ESI calcd. for C₁₇H₁₈N₇O₂S⁺ [M+H]⁺ 384.12, found 384.3.

7-(cyclopropylamino)-5-((3-((methylsulfonyl)methyl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**5**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (100 mg, 0.43 mmol) and 3-((methylsulfonyl)methyl)aniline (**78**) (79 mg, 0.43 mmol) in dioxane (2 mL) was added Cs₂CO₃ (418 mg, 1.28 mmol), BINAP (40 mg, 0.06 mmol) and Pd(OAc)₂ (14 mg, 0.06 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~3%, MeOH/DCM). The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 26%-56%, 10min). **5** (38.5 mg, 23.1% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.77 (s, 1H), 8.37 (s, 1H), 8.27 (s, 1H), 7.91 (d, *J* = 7.7 Hz, 1H), 7.60 (t, *J* = 1.9 Hz, 1H), 7.37 (t, *J* = 7.9 Hz, 1H), 7.06 (d, *J* = 7.6 Hz, 1H), 6.03 (s, 1H), 4.46 (s, 2H), 2.95 (s, 3H), 2.65 – 2.57 (m, 1H), 0.85 – 0.78 (m, 2H), 0.75 – 0.68 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.84, 150.81, 148.40, 145.12, 140.36, 129.68, 128.99, 124.89, 121.91, 119.58, 114.80, 76.54, 76.40, 59.65, 23.33, 6.54. HPLC t_R = 4.118 min in 8 min chromatography, purity 98.6%. LCMS t_R = 2.028 min in 4 min chromatography, MS ESI calcd. for C₁₈H₁₉N₆O₂S⁺ [M+H]⁺ 383.13, found 383.3.

Methyl(3-nitrobenzyl)sulfane (105). To a solution of 1-(bromomethyl)-3-nitrobenzene (104) (2 g, 9.26 mmol) in MeOH (20 mL) was added NaSMe (510 mg, 7.28 mmol, 463.64 μ L) at 0 °C. The mixture was stirred at 25 °C for 3 h. The resulting mixture was extracted with EtOAc (10 mL x3). The combined organic phase was washed with water (20 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. 105 (1.5 g, 8.19 mmol, 88.4% yield) was obtained as a yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.20 - 8.04 (m, 2H), 7.76 (d, *J* = 7.6 Hz, 1H), 7.65 - 7.57 (m, 1H), 3.78 (s, 2H), 1.95 (s, 3H).

1-((methylsulfinyl)methyl)-3-nitrobenzene (106). To a solution of methyl(3-nitrobenzyl)sulfane (105) (1.3 g, 7.10 mmol) in DCM (15 mL) was added mCPBA (1.44 g, 7.10 mmol, 85% purity) at 0°C. The mixture was stirred at 0°C for 1 h. NaHCO₃ (5 mL) was added to the reaction mixture. The resulting mixture was extracted with DCM (10 mL x3). The combined organic phase was washed with brine (10 mL), NaHCO₃ (10 mL), dried over anhydrous Na₂SO₄, filtered and

concentrated. The residue used into next step without purification. **106** (1.32 g, 6.63 mmol, 93.4% yield) was obtained as a yellow oil. ¹H NMR (400 MHz, DMSO- d_6) δ 8.27 - 8.18 (m, 2H), 7.79 - 7.75 (m, 1H), 7.73 - 7.65 (m, 1H), 4.35 (d, *J* = 12.8 Hz, 1H), 4.11 (d, *J* = 12.8 Hz, 1H), 2.50 (s, 3H).

Tert-butyl N-[methyl-[(3-nitrophenyl)methyl]-oxo-\lambda^6-sulfanylidene]carbamate (107). To a solution of 1-((methylsulfinyl)methyl)-3-nitrobenzene (106) (1.00 g, 5.02 mmol) in DCM (120 mL) was added MgO (809 mg, 20.1 mmol), iodobenzene diacetate (2.43 g, 7.53 mmol) and rhodium(II) acetate (222 mg, 502 µmol). The mixture was added tert-butyl carbamate (1.18 g, 10.0 mmol) and stirred at 25°C for 16 hours. The reaction mixture was filtered and concentrated under reduced pressure to give a residue. The residue was purified by flash silica gel chromatography (Eluent of 0~15% Methanol/Dichloromethane). 107 (836 mg, 2.66 mmol, 53.0% yield) was obtained as a brown oil. LCMS t_R = 0.398 min in 0.8 min chromatography, 5-95AB, LCMS ESI calcd. for C₉H₁₁N₂O₅S⁺ [M+2H-'Bu]⁺ 259.04, found 259.0.

tert-butyl ((3-aminobenzyl)(methyl)(oxo)-\lambda^6-sulfaneylidene)carbamate (79). To a solution of tert-butyl N-[methyl-[(3-nitrophenyl)methyl]-oxo- $\lambda 6$ -sulfanylidene]carbamate (107) (736 mg, 2.34 mmol) in EtOH (30 mL) and H₂O (7.5 mL) was added Fe (1.31 g, 23.4 mmol) and NH₄Cl (501 mg, 9.37 mmol). The mixture was stirred at 65°C for 1 hour. The reaction mixture was filtered and concentrated under reduced pressure to give a residue. 79 (491 mg, 1.73 mmol, 73.7% yield) was obtained as a brown oil. LCMS t_R = 0.242 min in 0.8 min chromatography, 5-95AB, LCMS ESI calcd. for C₁₃H₂₁N₂O₃S⁺ [M+H]⁺ 285.13, found 591.2.

tert-butyl $((3-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)benzyl)(methyl)(oxo)-\lambda^6-sulfaneylidene)carbamate (102). To a solution of tert-butyl ((3-aminobenzyl)(methyl)(oxo)-\lambda^6-sulfaneylidene)carbamate (79) (500 mg, 1.76 mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (411 mg, 1.76 mmol) in 1,4-dioxane (15 mL) was added BINAP (164 mg, 264 µmol), Pd(OAc)₂ (59.2 mg, 264 µmol) and Cs₂CO₃ (2.29 g, 7.03 mmol). The reaction mixture was heated in a microwave reactor at 100°C for 2 hours. The reaction mixture was filtered and concentrated under reduced pressure to give a residue. The residue was purified by flash silica gel chromatography (Eluent of 0~70% Ethyl acetate/Petroleum ether). 102 (436 mg, 905 µmol, 51.5% yield) was obtained as a red oil. LCMS t_R = 0.453 min in 0.8 min chromatography, 5-95AB, LCMS ESI calcd. for C₁₈H₂₀N₇OS⁺ [M+2H-Boc]⁺ 382.14, found 382.0.$

7-(Cyclopropylamino)-5-[3-[(methylsulfonimidoyl)methyl]anilino]pyrazolo[1,5-a]pyrimidine-3-carbonitrile (6 and 7). To a solution of tert-butyl ((3-((3-cyano-7-(cyclopropylamino))))) λ^6 -sulfaneylidene)carbamate (102) (436 mg, 905 µmol) in DCM (10 mL) was added TFA (9.08 g, 79.7 mmol, 5.92 mL). The mixture was stirred at 25°C for 2 hours. The reaction mixture was filtered and concentrated under reduced pressure to give a residue. The residue was purified by prep-HPLC (column: Phenomenex luna C18 150*25mm* 10um; mobile phase: [water (FA)-ACN]; B%: 13%-43%, 10 min). The product was separated by SFC (column: DAICEL CHIRALPAK AS(250mm*30mm, 10 um); mobile phase: [0.1% NH₃H₂O IPA]; B%: 40%-40%, A5.8; 70 min) to give Peak 1 as 6 (53.2 mg, 28.3% yield) and Peak 2 as 7 (52.2 mg, 29.3% yield) both as white solids. 6: ¹H NMR (400 MHz, DMSO- d_6) δ 9.74 (s, 1H), 8.36 (s, 1H), 8.25 (s, 1H), 7.81 (d, J = 8.2 Hz, 1H), 7.66 (s, 1H), 7.35 (t, J = 7.9 Hz, 1H), 7.09 (d, J = 7.6 Hz, 1H), 6.02 (s, 1H), 4.39 (s, 2H), 2.87 (s, 3H), 2.65 – 2.57 (m, 1H), 0.85 – 0.78 (m, 2H), 0.75 – 0.68 (m, 2H). LCMS $t_R = 0.338$ min in 0.8 min chromatography, 5-95AB, MS ESI calcd. For $C_{18}H_{20}N_7OS^+$ [M+H]⁺ 382.14, found 382.0. HPLC t_R = 1.428 min in 4 min chromatography, 10-80AB, purity 96.9%. Chiral SFC: ee% = 100%. 7: ¹H NMR (400 MHz, DMSO- d_6) δ 9.74 (s, 1H), 8.36 (s, 1H), 8.25 (d, J = 1.9 Hz, 1H), 7.81 (dd, J = 8.1, 2.2 Hz, 1H), 7.66 (t, J = 1.9 Hz, 1H), 7.35 (t, J = 7.9 Hz, 1H), 7.08 (d, J = 7.6 Hz, 1H), 6.03 (s, 1H), 4.37 (s, 2H), 2.85 (s, 3H), 2.65 - 2.57 (m, 1H), 0.84 - 0.79 (m, 2H), 0.74 - 0.69 (m, 2H), 0 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.85, 150.83, 148.36, 145.07, 140.16, 131.19, 128.76, 124.96, 122.00, 119.41, 114.80, 76.47, 76.36, 62.40, 40.89, 23.32, 6.52. LCMS $t_R = 0.343$ min in 0.8 min chromatography, 5-95AB, MS ESI calcd. For $C_{18}H_{20}N_7OS^+$ [M+H]⁺ 382.14, found 382.0. HPLC t_R = 1.436 min in 4 min chromatography, 10-80AB, purity 96.9%. Chiral SFC: ee% = 100%.

3-((methylsulfinyl)methyl)aniline (80). A mixture of 1-((methylsulfinyl)methyl)-3-nitrobenzene (106) (200 mg, 1.00 mmol), NH₄Cl (268.49 mg, 5.02 mmol) and Fe (280.33 mg, 5.02 mmol) in EtOH (5 mL) and H₂O (5 mL) was degassed and purged with N₂ for 3 times, and then the mixture was stirred at 90°C for 2 h under N₂ atmosphere. The reaction mixture was filtered. The filtrate was concentrated. The residue was dissolved in water (4 mL). The resulting suspension was extracted with EtOAc (10 mL x3). The combined organic phase was dried over anhydrous Na₂SO₄, filtered and concentrated. The product was used in the next step without further purification. 80 (100 mg, 590.86 µmol, 58.9% yield) was obtained as a

colorless oil. ¹H NMR (400 MHz, chloroform-d) δ 7.15 - 7.10 (m, 1H), 6.65 - 6.60 (m, 3H), 4.05 - 3.96 (m, 1H), 3.80 (d, J = 12.8 Hz, 1H), 2.46 (s, 3H)

7-(cyclopropylamino)-5-((3-((methylsulfinyl)methyl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (8). To a of 3-((methylsulfinyl)methyl)aniline (80) (100)590.86 5-chloro-7solution mg, µmol) and (cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (110.45 mg, 472.69 µmol) in dioxane (2 mL) was added BINAP (55.19 mg, 88.63 µmol), Pd(OAc)₂ (19.90 mg, 88.63 µmol) and Cs₂CO₃ (577.55 mg, 1.77 mmol) at 25°C. Then the mixture was degassed and purged with N_2 . The reaction mixture was heated in microwave at 130°C for 0.5 h. The residue was purified by flash silica gel chromatography (eluent of 0%~2% MeOH/DCM) and prep-HPLC(column: Xtimate C18 150*40mm*10um;mobile phase: [water(NH₄HCO₃)-ACN];gradient:12%-52% B over 1 min). 8 (26.5 mg, 70.57 μmol, 12.3% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.73 (s, 1H), 8.36 (s, 1H), 8.26 (d, J = 1.9Hz, 1H), 7.82 (dd, J = 8.2, 2.1 Hz, 1H), 7.57 (t, J = 1.9 Hz, 1H), 7.34 (t, J = 7.9 Hz, 1H), 6.97 (dt, J = 7.6, 1.3 Hz, 1H), 6.02 (s, 1H), 4.09 (d, J = 12.7 Hz, 1H), 3.94 (d, J = 12.7 Hz, 1H), 2.64 - 2.57 (m, 1H), 2.51 (s, 3H), 0.85 - 0.78 (m, 2H), 0.74 (m, 2H),0.69 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.83, 150.84, 148.38, 145.09, 140.34, 131.72, 128.90, 124.20, 121.33, 118.96, 114.77, 76.52, 76.38, 58.86, 37.41, 23.32, 6.54. HPLC $t_R = 3.857$ min in 8 min chromatography, purity 97.5%. LCMS $t_R = 2.572$ min in 4.0 min chromatography, 10-80AB, LCMS ESI calcd. for $C_{18}H_{19}N_6OS^+$ [M+H]⁺ 367.13, found 367.1.

tert-butyl (3-((dimethylphosphoryl)methyl)phenyl)carbamate (109). To a solution of dimethylphosphine oxide (1.36 g, 17.47 mmol) in THF (10 mL) was degassed and purged with N₂ for 3 times. Then NaHMDS (1 M, 17.47 mL) was added at -30°C under N₂ atmosphere. The mixture was stirred at -30°C for 1 h. Then a solution of tert-butyl (3-(bromomethyl)phenyl)carbamate (108) (1 g, 3.49 mmol) was added dropwise and the mixture was stirred at 25°C for 12 h under N₂ atmosphere. The reaction was quenched with H₂O (8 mL) drop-wise. The resulting mixture was extracted with DCM (30 mL x3). The combined organic phase was washed with brine (15 mL), water (15 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. The crude product was used to the next step without further purification. 109 (1.3 g, crude) was obtained as colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.31 (s, 1H), 7.39 - 7.35 (m, 1H), 7.30 - 7.14 (m, 2H), 6.86 - 6.81 (m, 1H), 3.07 (d, *J* = 14.8 Hz, 2H), 1.47 - 1.45 (m, 9H), 1.35 - 1.33 (m, 3H), 1.31 - 1.30 (m, 3H). LCMS t_R = 0.500 min in 1.0 min chromatography, 5-100AB, LCMS ESI calcd. for C₁₄H₂₃NO₃P⁺ [M+H]⁺284.14 found 284.2.

(3-aminobenzyl)dimethylphosphine oxide **(81)**. То solution а of tert-butyl (3 -((dimethylphosphoryl)methyl)phenyl)carbamate (109) (1.3 g, 4.59 mmol) in DCM (10 mL) was added TFA (4.07 g, 35.72 mmol, 2 mL). The mixture was stirred at 25 °C for 2 h. The reaction mixture was concentrated directly. The reaction mixture was poured into aq. NaHCO₃ (15 mL) slowly. The resulting mixture was then adjusted to pH~8 by aq. NaHCO₃. The resulting mixture was extracted with DCM (30 mL x3). The combined organic phase was washed with brine (15 mL), water (15 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. The residue was used in next step directly. **81** (400 mg, crude) was obtained as brown oil. ¹H NMR (400 MHz, DMSO- d_6) δ 6.94 (t, J = 7.6 Hz, 1H), 6.46 - 6.36 (m, 3H), 5.39 -5.22 (m, 1H), 5.20 - 5.08 (m, 1H), 2.95 (d, J = 14.8 Hz, 2H), 1.33 - 1.32 (m, 3H), 1.30 - 1.29 (m, 3H).

7-(cyclopropylamino)-5-((3-((dimethylphosphoryl)methyl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (9). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (200 mg, 1.09 mmol) and (3-aminobenzyl)dimethylphosphine oxide (81) (204 mg, 873.43 µmol) in dioxane (3 mL) was added BINAP (101 mg, 163.77 µmol), Pd(OAc)₂ (36 mg, 163.77 µmol) and Cs₂CO₃ (1.07 g, 3.28 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in microwave at 130°C for 0.5 h. After being cooled to 25°C, the reaction mixture was concentrated directly. The residue was purified by flash silica gel chromatography (eluent of 0%~19%,EtOAC/PE) and prep-HPLC (column: Xtimate C18 150*40mm*10um;mobile phase: [water(NH₄HCO₃)-ACN];gradient:10%-50% B over 32 min). 9 (31 mg, 81.50 µmol, 7.8% yield) was obtained as was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.67 (s, 1H), 8.35 (s, 1H), 8.22 (s, 1H), 7.69 (d, *J* = 8.1 Hz, 1H), 7.56 (q, *J* = 2.1 Hz, 1H), 7.28 (t, *J* = 7.8 Hz, 1H), 7.04 – 6.83 (m, 1H), 6.01 (s, 1H), 3.12 (d, *J* = 15.1 Hz, 2H), 2.64 – 2.57 (m, 1H), 1.37 (d, *J* = 12.8 Hz, 6H), 0.84 – 0.78 (m, 2H), 0.75 – 0.68 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.90, 150.86, 148.31, 145.08, 140.22 (d, *J* = 2.7 Hz), 134.08 (d, *J* = 7.9 Hz), 128.70 (d, *J* = 68.2 Hz), 6.55. HPLC t_R = 2.118 min in 8 min chromatography, purity 99.2%. LCMS t_R = 1.279 min in 4.0 min chromatography, 10-80AB, LCMS ESI calcd. for C₁₉H₂₂N₆OP⁺ [M+H]⁺381.16, found 381.2.

7-(cyclopropylamino)-5-((3-(2-hydroxyethyl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**10**). То а of 2-(3-aminophenyl)ethan-1-ol (82) (150)1.09 mmol) 5-chloro-7solution mg. and (cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (204 mg, 874.77 µmol) in dioxane (3 mL) was added Cs₂CO₃ (1.07 g, 3.28 mmol), BrettPhos Pd G₃ (148 mg, 164.02 µmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in microwave at 130°C for 0.5 h. No work-up. The residue was purified by flash silica gel chromatography (eluent of $0\sim4\%$ MeOH in DCM). The residue was purified by prep-HPLC (column: Xtimate C₁₈ 150*40mm*10um;mobile phase: [water(NH₄HCO₃)-ACN];gradient:18%-58% B over 1 min). 10 (24.1 mg, 70.35 μmol, 9.41% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.60 (s, 1H), 8.34 (s, 1H), 8.21 (d, J = 1.9Hz, 1H), 7.60 (dd, J = 8.2, 2.2 Hz, 1H), 7.56 (t, J = 1.9 Hz, 1H), 7.23 (t, J = 7.8 Hz, 1H), 6.87 (d, J = 8.0 Hz, 1H), 6.00 (s, 1H), 4.65 (t, J = 5.2 Hz, 1H), 3.64 (td, J = 7.1, 5.1 Hz, 2H), 2.71 (t, J = 7.1 Hz, 2H), 2.63 – 2.55 (m, 1H), 0.85 – 0.78 (m, 1H 2H), 0.75 - 0.69 (m, 2H). ¹³C NMR (101 MHz, DMSO- d_6) δ 156.92, 150.94, 148.31, 145.02, 140.13, 140.05, 128.52, 122.97, 120.16, 117.29, 114.82, 76.40, 76.26, 62.08, 23.31, 6.55. HPLC $t_R = 3.019$ min in 8 min chromatography, purity 97.6%. LCMS $t_R = 2.800$ min in 4.0 min chromatography, 10-80AB, LCMS ESI calcd. for $C_{18}H_{19}N_6O^+$ [M+H]⁺ 335.16, found 335.5.

methyl 2-(3-aminophenyl)acetate (111). To a solution of methyl 2-(3-nitrophenyl)acetate (110) (1 g, 5.12 mmol) in MeOH (10 mL) was added Pd/C (1.00 g, 939.67 µmol, 10% Pd) under N₂ atmosphere. The suspension degassed and purged with H₂ (15 Psi) at 25 °C and stirred for 12 hours. The reaction mixture was filtered. The filtrate was concentrated directly. **111** (756 mg, crude) was obtained as colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 6.98 - 6.91 (m, 1H), 6.47 - 6.41 (m, 2H), 6.37 (d, *J* = 7.2 Hz, 1H), 5.04 (s, 2H), 3.59 (s, 3H), 3.46 (s, 2H).

methyl 2-(3-(((*benzyloxy*)*carbonyl*)*amino*)*phenyl*)*acetate* (**112**). To a solution of methyl 2-(3-aminophenyl)acetate (**111**) (656 mg, 3.97 mmol) in DCM (20 mL) was added DIPEA (769.9 mg, 5.96 mmol, 1.04 mL) at 0°C, then CbzCl (812.9 mg, 4.77 mmol, 680.30 μ L) was added to the mixture slowly. The mixture was stirred at 25°C for 16 hours. The reaction mixture was poured into sat. aq. NaHCO₃ (20 mL). The resulting mixture was extracted with DCM (15 mL x2). The combined organic phase was washed with brine (20 mL), dried over anhydrous Na₂SO₄, filtered and concentrated to dryness. The residue was purified by flash silica gel chromatography (ISCO®; 20g, Sepa Flash® Silica Flash Column, eluent of 0~14% EA/PE@ 35 mL/min). **112** (1.1 g, 3.67 mmol, 92.54% yield) was obtained as colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.77 (s, 1H), 7.44 - 7.30 (m, 7H), 7.22 (t, *J* = 7.6 Hz, 1H), 6.89 (d, *J* = 7.6 Hz, 1H), 5.15 (s, 2H), 3.65 - 3.58 (m, 5H).

benzyl (3-(2-hydroxy-2-methylpropyl)phenyl)carbamate (113). To a solution of methyl 2-(3-(((benzyloxy)carbonyl)amino)phenyl)acetate (112) (1.1 g, 3.67 mmol) in THF (20 mL) was added MeMgBr (3 M in THF, 6.12 mL) at 0 °C. The mixture was stirred at 25°C for 16 hours. The reaction was quenched with aq. NH₄Cl (10 mL) dropwise, then the mixture was extracted with DCM (15 mL x2). The combined organic phase was dried over Na₂SO₄, filtered and concentrated to give crude product. The residue was purified by flash silica gel chromatography (ISCO®; 12g, Sepa Flash® Silica Flash Column, eluent of 0~20% EA/PE@ 35 mL/min). 113 (880 mg, 2.94 mmol, 79.99% yield) was obtained as colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.65 (s, 1H), 7.48 - 7.25 (m, 7H), 7.15 (t, *J* = 7.6 Hz, 1H), 6.84 (d, *J* = 7.6 Hz, 1H), 5.14 (s, 2H), 4.29 (s, 1H), 2.59 (s, 2H), 1.05 (s, 6H).

1-(3-aminophenyl)-2-methylpropan-2-ol (**83**). To a solution of benzyl (3-(2-hydroxy-2-methylpropyl)phenyl)carbamate (**113**) (330 mg, 1.10 mmol) in MeOH (5 mL) was added Pd/C (400 mg, 375.87 μmol, 10% Pd) under N₂ atmosphere. The suspension degassed and purged with H₂ (15 Psi) at 25 °C and stirred for 2 hours. The reaction mixture was filtered. The filtrate was concentrated directly. **83** (165 mg, crude) was obtained as a colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 6.87 (t, *J* = 7.6 Hz, 1H), 6.48 - 6.27 (m, 3H), 4.85 (s, 2H), 4.21 (s, 1H), 2.48 (s, 2H), 1.04 (s, 6H).

7-(cyclopropylamino)-5-((3-(2-hydroxy-2-methylpropyl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (11). To a solution of 1-(3-aminophenyl)-2-methylpropan-2-ol (83) (150 mg, 907.82 µmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (212.1 mg, 907.82 µmol) in dioxane (4 mL) was added BINAP (84.8 mg, 136.17 µmol), Pd(OAc)₂ (30.6 mg, 136.17 µmol) and Cs₂CO₃ (887.3 mg, 2.72 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in microwave at 130°C for 0.5 h. The reaction mixture was concentrated. The residue was purified by flash silica gel chromatography (eluent of 0%~4% MeOH/DCM) and prep-HPLC(column: Xtimate C18 150*40mm*10um;mobile phase: [water(NH₄HCO₃)-ACN];gradient:22%-62% B over 32 min). **11** (22.9 mg, 63.19 µmol, 6.96% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.58 (s, 1H), 8.33 (s, 1H), 8.19 (br s, 1H), 7.61 (dd, *J* = 7.8, 1.8 Hz, 1H), 7.53 (t, *J* = 2.0 Hz, 1H), 7.22 (t, *J* = 7.8 Hz, 1H), 6.87

(dt, J = 7.5, 1.3 Hz, 1H), 6.00 (s, 1H), 4.32 (s, 1H), 2.64 (s, 2H), 2.59 (tt, J = 6.9, 3.6 Hz, 1H), 1.10 (s, 6H), 0.83 – 0.77 (m, 2H), 0.73 – 0.68 (m, 2H). ¹³C NMR (101 MHz, DMSO- d_6) δ 156.97, 150.89, 148.26, 145.01, 139.53, 139.46, 127.89, 124.67, 121.83, 117.34, 114.75, 76.36, 76.23, 69.46, 49.87, 29.24, 23.30, 6.53. HPLC t_R = 4.287 min in 8 min chromatography, purity 98.5%. LCMS t_R = 2.106 min in 4 min chromatography, MS ESI calcd for C₂₀H₂₃N₆O⁺ [M+H]⁺ 363.19, found 363.2.

Tert-butyl 3-((3-cyano-7-(cyclopropylamino)pyrazolo[*1,5-a*]*pyrimidin-5-yl*)*amino*)*benzoate* (*103*). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[*1,5-a*]*pyrimidine-3-carbonitrile* (**75**) (200 mg, 0.86 mmol) and tert-butyl 3-aminobenzoate (**84**) (192 mg, 0.94 mmol) in dioxane (3 mL) was added Cs₂CO₃ (837 mg, 2.57 mmol), BINAP (80 mg, 0.13 mmol) and Pd(OAc)₂ (29 mg, 0.13 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~2%, MeOH/DCM). **103** (400 mg, 95.4%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.86 (s, 1H), 8.39 (s, 1H), 8.32 (d, *J* = 2.0 Hz, 2H), 8.03 (d, *J* = 8.4 Hz, 1H), 7.54 (d, *J* = 8.0 Hz, 1H), 7.46 (t, *J* = 8.0 Hz, 1H), 6.01 (s, 1H), 2.62 (s, 1H), 1.57 (s, 9H), 0.84 - 0.81 (m, 2H), 0.75 - 0.74 (m, 2H). LCMS t_R = 0.575 min in 1 min chromatography, MS ESI calcd. for C₂₁H₂₃N₆O₂⁺ [M+H]⁺ 391.19, found 391.1.

3-((3-Cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)benzoic acid (12). To a solution of Tert-butyl 3-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)benzoate (103) (100 mg, 0.26 mmol) in DCM (3 mL) was added TFA (2.15 g, 18.87 mmol) at 25°C. The mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 0%-30%, 14min). Then the impure product was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 0%-30%, 14min). 12 (8.3 mg, 39.7%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-d₆) δ 9.86 (s, 1H), 8.37 (s, 1H), 8.28 (d, *J* = 1.8 Hz, 1H), 8.24 (s, 1H), 8.12 (dd, *J* = 8.2, 2.3 Hz, 1H), 7.58 (d, *J* = 7.6 Hz, 1H), 7.45 (t, *J* = 7.9 Hz, 1H), 6.01 (s, 1H), 2.63 – 2.57 (m, 1H), 0.87 – 0.79 (m, 2H), 0.75 – 0.69 (m, 2H). ¹³C NMR (101 MHz, DMSO-d₆) δ 167.64, 156.78, 150.77, 148.45, 145.22, 140.55, 128.93, 123.11, 122.89, 120.18, 114.72, 76.75, 76.66, 23.38, 6.60. HPLC t_R = 3.776 min in 8 min chromatography, purity 98.9%. LCMS t_R = 2.114 min in 4 min chromatography, MS ESI calcd. for C₁₇H₁₅N₆O₂⁺ [M+H]⁺ 335.13, found 335.3.

5-((3-(2-Cyanopropan-2-yl)phenyl)amino)-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**13**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (200 mg, 0.85 mmol, 1 *eq*) and 2-(3-aminophenyl)-2-methylpropanenitrile (**85**) (164 mg, 1.03 mmol, 1.2 *eq*) in dioxane (5 mL) was added Cs₂CO₃ (836 mg, 2.57 mmol, 3 *eq*), BINAP (79 mg, 0.13 mmol, 0.15 *eq*) and Pd(OAc)₂ (28 mg, 0.13 mmol, 0.15 *eq*) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was filtered. The solid was washed with DCM (5 mL x2). The combined filtrate was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex Gemini-NX C18 75*30 mm*3um; mobile phase: [water (0.05% NH₃H₂O + 10mM NH₄HCO₃)-ACN]; B%: 39%-79%, 10min). **13** (10 mg, 0.03 mol, 3.2% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.81 (s, 1H), 8.36 (s, 1H), 8.27 (s, 1H), 8.13 (t, *J* = 2.1 Hz, 1H), 7.64 (ddd, *J* = 8.1, 2.1, 1.0 Hz, 1H), 7.38 (t, *J* = 8.0 Hz, 1H), 7.16 (ddd, *J* = 7.8, 2.1, 1.0 Hz, 1H), 6.00 (s, 1H), 2.61 (tt, *J* = 6.9, 3.6 Hz, 1H), 1.72 (s, 6H), 0.87 - 0.78 (m, 2H), 0.77 - 0.70 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.71, 150.80, 148.39, 144.98, 142.05, 140.89, 129.28, 124.71, 118.79, 118.41, 116.07, 114.62, 76.75, 76.56, 36.60, 28.30, 23.32, 6.55. HPLC t_R = 3.407 min in 8 min chromatography, Xtimate C18 2.1*30mm 3um, purity 98.9%. LCMS t_R = 1.665 min in 4 min chromatography, Xtimate C18, 3um, 2.1*30mm, MS ESI calcd. for C₂₀H₂₀N₇+ [M+H]+ 358.18, found 358.4.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (14). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (200 mg, 0.85 mmol, 1 *eq*) and 3-(4H-1,2,4-triazol-4-yl)aniline (86) (164 mg, 1.03 mmol, 1.2 *eq*) in dioxane (5 mL) was added Cs₂CO₃ (836 mg, 2.57 mmol, 3 *eq*) BINAP (79 mg, 0.12 mmol, 0.15 *eq*) and Pd(OAc)₂ (28 mg, 0.12 mmol, 0.15 *eq*) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was filtered. The solid was washed with DCM (5 mL x2). The combined filtrate was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex Gemini-NX C18 75*30mm*3um; mobile phase: [water (0.05% NH₃H₂O + 10mM NH₄HCO₃)-ACN]; B%: 24%-54%, 10min). 14 (11.2 mg, 0.03 mmol, 3.46% yield) was obtained as a light yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.99 (s, 1H), 9.02 (s, 2H), 8.40 (s, 1H), 8.38 (t, *J* = 2.3 Hz, 1H), 8.36 (br s, 1H), 7.62 (dd, *J* = 8.2, 2.0 Hz, 1H), 7.51 (t, *J* = 8.1 Hz, 1H), 7.31 (dd, *J* = 8.0, 2.2 Hz, 1H), 6.03 (s, 1H), 2.63 (tt, *J* = 6.9, 3.6 Hz, 1H), 0.87
-0.80 (m, 2H), 0.76 -0.71 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.58, 150.80, 148.57, 145.04, 141.90, 141.25, 134.22, 130.35, 118.51, 114.75, 114.55, 111.76, 76.97, 76.75, 23.36, 6.59. HPLC t_R = 2.371 min in 8 min chromatography, Xtimate C18 2.1*30mm 3um, purity 94.4%. LCMS t_R = 1.602 min in 4 min chromatography, Xtimate C18, 3um, 2.1*30mm, MS ESI calcd. for C₁₈H₁₆N₉⁺ [M+H]⁺ 358.15, found 358.0.

5-(3-nitrophenyl)-1H-1,2,3-triazole (115). To a solution of 1-ethynyl-3-nitrobenzene (114) (1.00 g, 6.08 mmol) and CuI (65 mg, 0.34 mmol) in DMF (9 mL) and MeOH (1 mL) was added TMSN₃ (1.17 g, 10.20 mmol) under N₂. The mixture was stirred at 100°C for 12 h. Water (30 mL) was added to the residue. The resulting suspension was extracted with DCM (50 mL x 2). The combined organic phase was washed with brine (30 mL), dried over anhydrous with Na₂SO₄, filtered and concentrated. The residue was purified by flash silica gel chromatography (eluent of 0~31%, EtOAc/PE). 115 (1.70 g, crude) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.66-8.62 (m, 2H), 8.32 (d, *J* = 7.6 Hz, 1H), 8.19 (dd, *J* = 1.6, 8.4 Hz, 1H), 7.76 (t, *J* = 8.0 Hz, 1H).

3-(1H-1,2,3-triazol-5-yl)aniline (87). To a solution of 5-(3-nitrophenyl)-1H-1,2,3-triazole (**115**). (686 mg, 3.61 mmol) in EtOH (20 mL) was added Fe (1.01 g, 18.04 mmol). A solution of NH₄Cl (579 mg, 10.82 mmol) in H₂O (5 mL) was added to the mixture. The mixture was stirred at 80°C for 2 hours. The reaction mixture was diluted with MeOH (20 mL) and filtered via a celite pad. The pad was washed with MeOH (15 mL x2). The combined filtrate was concentrated in vacuo. The residue was purified by prep-HPLC column: Xtimate C18 150*40mm*10um; mobile phase: [water (NH₃H₂O)-ACN]; B%: 0%-15%, 10min). **87** (552 mg, 93.0%) was obtained as a yellow solid. LCMS $t_R = 0.413$ min in 4 min chromatography, MS ESI calcd. for C₈H₉N₄⁺ [M+H]⁺ 161.08, found 161.1.

5-((3-(4H-1,2,3-triazol-4-yl)phenyl)amino)-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**15**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (200 mg, 0.86 mmol) and 3-(1H-1,2,3-triazol-5-yl)aniline (**87**) (137 mg, 0.86 mmol) in dioxane (3 mL) was added Cs₂CO₃ (837 mg, 2.57 mmol), BINAP (80 mg, 0.13 mmol) and Pd(OAc)₂ (29 mg, 0.13 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130 °C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~10% MeOH/DCM) to give the product. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 21%-51%, 10min). **15** (16.2 mg, 5.2%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-d₆) δ 15.09 (br s, 1H), 9.82 (s, 1H), 8.43 (d, *J* = 2.5 Hz, 1H), 8.38 (s, 1H), 8.28 (d, *J* = 1.8 Hz, 1H), 8.23 (s, 1H), 7.70 (dd, *J* = 8.1, 2.2 Hz, 1H), 7.51 (d, *J* = 7.6 Hz, 1H), 7.42 (t, *J* = 7.9 Hz, 1H), 6.04 (s, 1H), 2.65 – 2.57 (m, 1H), 0.85 – 0.79 (m, 2H), 0.76 – 0.71 (m, 2H). ¹³C NMR (101 MHz, DMSO-d₆) δ 156.85, 150.93, 148.42, 145.03, 141.02, 131.04, 129.43, 119.31, 118.99, 118.96, 116.49, 116.40, 114.88, 76.68, 76.59, 23.34, 6.59. HPLC t_R = 1.985 min in 4 min chromatography, purity 94.8%. LCMS t_R = 1.655 min in 4 min chromatography, MS ESI calcd. for C₁₈H₁₆N₉+ [M+H]+ 358.15, found 357.9.

5-((3-(1H-1,2,3-triazol-1-yl)phenyl)amino)-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**16**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (200 mg, 0.86 mmol, 1 *eq*) and 3-(1H-1,2,3-triazol-1-yl)aniline (**88**) (137 mg, 0.86 mmol, 1 *eq*) in dioxane (3 mL) was added Cs₂CO₃ (837 mg, 2.57 mmol, 3 *eq*), BINAP (80 mg, 0.13 mmol, 0.15 *eq*) and Pd(OAc)₂ (29 mg, 0.13 mmol, 0.15 *eq*) at 25°C. The mixture was degassed and purged with N₂. Then the reaction mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was filtered. The filter cake was washed with DCM (3 mL x2) and the combined filtrate was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent: 0-11%, MeOH / DCM). The impure product was purified by prep-HPLC (column: Waters Torus 2-PIC 150*19mm*5um; mobile phase: [Heptane-EtOH (0.1%NH₃H₂O)]; B%: 20%-35%, 6min). **16** (11.6 mg, 0.03 mmol, 8.64% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.01 (s, 1H), 8.70 (s, 1H), 8.49 (t, *J* = 2.1 Hz, 1H), 8.39 (s, 1H), 8.35 (s, 1H), 7.99 (d, *J* = 1.1 Hz, 1H), 7.84 (dt, *J* = 8.2, 1.6 Hz, 1H), 7.55 (t, *J* = 8.0 Hz, 1H), 7.53 – 7.48 (m, 1H), 6.04 (s, 1H), 2.66 – 2.59 (m, 1H), 0.88 – 0.80 (m, 2H), 0.77 – 0.70 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.64, 150.74, 148.55, 145.14, 141.78, 137.02, 134.44, 130.24, 122.93, 118.87, 114.69, 113.42, 110.70, 76.97, 76.83, 23.38, 6.59. HPLC t_R = 4.145 min in 8 min chromatography, purity 99.0%. LCMS t_R = 1.729 min in 4 min chromatography, MS ESI calcd. for C₁₈H₁₆N₉⁺ [M+H]⁺ 358.15, found 358.3.

5-Methyl-4-(3-nitrophenyl)-2,4-dihydro-3H-1,2,4-triazole-3-thione (117). To a solution of 1-isothiocyanato-3nitrobenzene (116) (2.00 g, 11.1 mmol, 1 eq) in EtOH (25 mL) was added acetohydrazide (822 mg, 11.1 mmol, 1 eq) and DBU (404 mg, 2.65 mmol). Then the reaction mixture was stirred at 85°C for 10 hours. The reaction mixture was diluted with saturated NH₄Cl (20 mL) and extracted with EtOAc (10 mL x3). The combined organic phase was washed with brine (10 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. **117** (900 mg, 3.01 mmol, 27.12% yield) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.77 (br s, 1H), 8.43 (t, *J* = 2.0 Hz, 1H), 8.40-8.36 (m, 1H), 7.99 - 7.93 (m, 1H), 7.90 - 7.84 (m, 1H), 2.15 (s, 3H). LCMS t_R = 0.418 min in 1 min chromatography, MS ESI calcd. for C₉H₉N₄O₂S⁺ [M+H]⁺ 237.04, found 237.0.

3-Methyl-4-(3-nitrophenyl)-4H-1,2,4-triazole (118). To a solution of 5-methyl-4-(3-nitrophenyl)-2,4-dihydro-3H-1,2,4-triazole-3-thione (117) (700 mg, 2.96 mmol, 1 *eq*) in acetic acid (10 mL) was added NaNO₂ (1.39 g, 12.18 mmol, 2 *eq*) dissolved in H₂O (5 mL) dropwise at 0°C. Then the reaction mixture was stirred at 100°C for 5 hours. The reaction mixture was concentrated in vacuo. Water (10 mL) was added to the residue. The solution was adjusted to pH = 9 with 1M aq. NaOH. The mixture was extracted with DCM : MeOH = 10 : 1 (11 mL x3). The combined organic phase was washed with brine (10 mL) dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. **118** (627 mg, 2.32 mmol, 78.15% yield) was obtained as an orange solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.78 (s, 1H), 8.43 (t, *J* = 2.0 Hz, 1H), 8.39-8.36 (m, 1H), 8.03-8.01 (m, 1H), 7.91 - 7.85 (m, 1H), 2.39 (s, 3H). LCMS t_R = 0.382 min in 1 min chromatography, MS ESI calcd. for C₉H₉N₄O₂⁺ [M+H]⁺ 205.07, found 205.0.

3-(3-Methyl-4H-1,2,4-triazol-4-yl)aniline (89). To a solution of 3-Methyl-4-(3-nitrophenyl)-4H-1,2,4-triazole (118) (990 mg, 4.85 mmol, 1 *eq*) in EtOH (10 mL) was added SnCl₂.H₂O (4.92 g, 21.82 mmol, 4.5 *eq*) dissolved in EtOH (10 mL) and HCl (12 mol/L, 1.5 mL) dropwise at 0°C. Then the reaction mixture was stirred at 70°C for 10 hours. The reaction mixture was cooled down to 0°C and adjusted to pH 11-12 with 50% aq. NaOH. Then the mixture was filtered and the solid was washed with EtOH (10 mL). The filtrate was diluted with brine (10 mL) and extracted with DCM (20 mL x3). The combined organic phase was dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. **89** (789 mg, 2.81 mmol, 57.92% yield) was obtained as brown oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.58 (s, 1H), 7.17 (t, *J* = 8.0 Hz, 1H), 6.69-6.65 (m, 1H), 6.58 - 6.50 (m, 2H), 5.50 (s, 2H), 2.31 (s, 3H). LCMS t_R = 0.404 min in 4 min chromatography, MS ESI calcd. for C₉H₁₁N₄⁺ [M+H]⁺ 175.10, found 175.1.

7-(*Cyclopropylamino*)-5-((3-(3-methyl-4H-1,2,4-triazol-4-yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**17**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (300 mg, 1.28 mmol, 1 eq) and 3-(3-Methyl-4H-1,2,4-triazol-4-yl)aniline (**89**) (268 mg, 1.54 mmol, 1.2 eq) in dioxane (5 mL) was added Cs₂CO₃ (1.25 g, 3.85 mmol, 3 eq), Brettphos Pd G3 (128 mg, 0.141 mmol, 0.1 eq) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was filtered. The solid was washed with DCM (5 mL x2). The combined filtrate was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex luna 30*30 mm*10 um+ YMC AQ 100*30*10 um; mobile phase: [water (0.05%NH₃H₂O)-ACN]; B%: 30%-60%, 20min). **17** (37.6 mg, 0.1 mmol, 7.84% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.98 (s, 1H), 8.68 (s, 1H), 8.39 (s, 1H), 8.36 (s, 1H), 8.17 (t, *J* = 2.1 Hz, 1H), 7.65 (ddd, *J* = 8.3, 2.2, 1.0 Hz, 1H), 7.52 (t, *J* = 8.1 Hz, 1H), 7.13 (ddd, *J* = 7.8, 2.1, 0.9 Hz, 1H), 6.02 (s, 1H), 2.63 (tt, *J* = 6.8, 3.6 Hz, 1H), 2.46 (s, 3H), 0.89 - 0.80 (m, 2H), 0.80 - 0.70 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.57, 150.65, 149.69, 148.49, 145.17, 143.65, 141.56, 134.35, 130.10, 119.07, 118.10, 115.22, 114.64, 76.92, 76.86, 23.35, 10.88, 6.56. HPLC t_R = 3.429 min in 8 min chromatography, Xtimate C18 2.1*30mm 3um, purity 98.7%. LCMS t_R = 1.401 min in 4 min chromatography, Xtimate C18, 3um, 2.1*30mm, MS ESI calcd. for C₁₉H₁₈N₉+ [M+H]+ 372.17, found 372.3.

7-(*Cyclopropylamino*)-5-((3-(2-methyl-1H-imidazol-1-yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**18**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (200 mg, 0.85 mmol, 1 *eq*) and 3-(2-methyl-1H-imidazol-1-yl)aniline (**90**) (177 mg, 1.03 mmol, 1.2 *eq*) in dioxane (5 mL) was added Cs₂CO₃ (836 mg, 2.57 mmol, 3 *eq*) BINAP (79 mg, 0.13 mmol, 0.15 *eq*) and Pd(OAc)₂ (28 mg, 0.13 mmol, 0.15 *eq*) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was filtered. The solid was washed with DCM (5 mL x2). The combined filtrate was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex Gemini-NX C18 75*30mm*3um; mobile phase: [water (0.05% NH₃H₂O + 10mM NH₄HCO₃)-ACN]; B%: 31%-61%, 10min). **18** (24 mg, 0.06 mmol, 7.57% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.92 (s, 1H), 8.38 (s, 1H), 8.33 (s, 1H), 8.13 (t, *J* = 2.1 Hz, 1H), 7.59 (dd, *J* = 8.1, 2.1 Hz, 1H), 7.47 (t, *J* = 8.0 Hz, 1H), 7.29 (d, *J* = 1.4 Hz, 1H), 7.05 (dd, *J* = 7.8, 2.1 Hz, 1H), 6.91 (d, *J* = 1.4 Hz, 1H), 6.01 (s, 1H), 2.62 (tt, *J* = 6.9, 3.6 Hz, 1H), 2.40 (s, 3H), 0.87 – 0.80 (m, 2H), 0.76 – 0.69 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.61, 150.67, 148.44, 145.16, 143.51, 141.32, 137.93, 129.79, 127.28, 120.55, 118.07, 115.42, 114.64, 76.86, 76.82, 23.34, 14.01, 6.55. HPLC t_R = 1.964 min in 8 min chromatography, Xtimate C18 2.1*30mm 3um, purity 99.4%. LCMS t_R

= 1.442 min in 4 min chromatography, Xtimate C18,3um,2.1*30mm, MS ESI calcd. for $C_{20}H_{19}N_8^+$ [M+H]⁺ 371.17, found 371.0.

7-(*Cyclopropylamino*)-5-((3-(4-methyl-4H-1,2,4-triazol-3-yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**19**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (200 mg, 0.85 mmol, 1 *eq*) and 3-(4-methyl-4H-1,2,4-triazol-3-yl)aniline (**91**) (156 mg, 0.89 mmol, 1.05 *eq*) in dioxane (5 mL) was added Cs₂CO₃ (836 mg, 2.57 mmol, 3 *eq*) BINAP (79 mg, 0.13 mmol, 0.15 *eq*) and Pd(OAc)₂ (28 mg, 0.13 mmol, 0.15 *eq*) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was filtered. The solid was washed with DCM (5 mL x2). The combined filtrate was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex Gemini-NX C18 75*30mm*3um; mobile phase: [water (0.05% NH₃H₂O + 10mM NH₄HCO₃)-ACN]; B%: 15%-55%, 11min). **19** (26.4 mg, 0.07 mmol, 8.2% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.89 (s, 1H), 8.60 (s, 1H), 8.38 (s, 1H), 8.34 – 8.26 (m, 2H), 7.76 (dt, *J* = 8.3, 1.4 Hz, 1H), 7.51 (t, *J* = 7.9 Hz, 1H), 7.42 (dt, *J* = 7.7, 1.4 Hz, 1H), 6.03 (s, 1H), 3.86 (s, 3H), 2.62 (tt, *J* = 6.9, 3.6 Hz, 1H), 0.87 – 0.80 (m, 2H), 0.76 – 0.71 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.73, 152.98, 150.75, 148.40, 146.24, 145.17, 140.59, 129.39, 127.39, 122.01, 120.23, 118.57, 114.81, 76.79, 76.69, 32.52, 23.34, 6.56. HPLC t_R = 2.462 min in 8 min chromatography, Xtimate C18 2.1*30mm 3um, purity 99.3%. LCMS t_R = 1.527 min in 4 min chromatography, Xtimate C18,3um,2.1*30mm, MS ESI calcd. for C₁₉H₁₈N₉+ [M+H]+ 372.17, found 372.0.

1-(2,2-Dimethoxyethyl)-3-(3-nitrophenyl)urea (120). To a solution of 2,2-dimethoxyethan-1-amine (704 mg, 6.70 mmol, 1.1 *eq*) in DCM (5 mL) was added 1-isocyanato-3-nitrobenzene (119) (1.00 g, 6.09 mmol, 1 *eq*) dissolved in DCM (5 mL) dropwise at 0°C. Then the reaction mixture was stirred at 0 °C to 25 °C for 3 hours. The reaction mixture was concentrated in vacuo and carried forward without further purification. **120** (1.64 g, crude) was obtained as a light-yellow solid.

1-(3-nitrophenyl)-1,3-dihydro-2H-imidazol-2-one (121). To a solution of 1-(2,2-Dimethoxyethyl)-3-(3-nitrophenyl)urea (120) (1.64 g, 6.09 mmol, 1 *eq*) in DCM (10 mL) was added TFA (1.39 g, 12.18 mmol, 2 *eq*) dissolved in DCM (5 mL) dropwise. Then the reaction mixture was stirred at 25°C for 10 hours. The reaction mixture was concentrated in vacuo. 121 (1.20 g, crude) was obtained as a light yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.53 (s, 1H), 8.82 (t, *J* = 2.0 Hz, 1H), 8.15 (dd, *J* = 1.2, 8.4 Hz, 1H), 8.05 (dd, *J* = 2.0, 8.4 Hz, 1H), 7.72 (t, *J* = 8.0 Hz, 1H), 7.20 (dd, *J* = 2.0, 3.2 Hz, 1H), 6.70 (t, *J* = 2.8 Hz, 1H). LCMS $t_R = 0.425$ min in 1 min chromatography, Chromolith Flash RP-18, 5um, 3.0*25mm, MS ESI calcd. for C₉H₈N₃O₃⁺ [M+H]⁺ 206.06, found 206.0.

1-(3-aminophenyl)-1,3-dihydro-2H-imidazol-2-one (**92**). To a solution of 1-(3-nitrophenyl)-1,3-dihydro-2H-imidazol-2one (**121**) (450 mg, 2.19 mmol, 1 *eq*) in EtOH (4 mL) was added SnCl₂.H₂O (1.24 g, 6.58 mmol, 3 *eq*) dissolved in EtOH (3 mL) and HCl (12 mol/L, 1 mL) dropwise at 0°C. Then the reaction mixture was stirred at 70°C for 10 hours. The reaction mixture was cooled down to 0°C and adjusted to pH 11-12 with 50% aq. NaOH. Then the mixture was filtered and the solid was washed with EtOH (5 mL). The combined filtrate was concentrated in vacuo. **92** (360 mg, 1.26 mmol, 57.59% yield) was obtained as a white solid. LCMS $t_R = 0.243$ min in 1 min chromatography, Chromolith Flash RP-18, 5um, 3.0*25mm, MS ESI calcd. for C₉H₁₀N₃O⁺ [M+H]⁺ 176.08, found 176.1.

7-(*Cyclopropylamino*)-5-((3-(2-oxo-2,3-dihydro-1H-imidazol-1-yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3carbonitrile (**20**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (200 mg, 0.85 mmol, 1 *eq*) and 1-(3-aminophenyl)-1,3-dihydro-2H-imidazol-2-one (**92**) (157 mg, 0.89 mmol, 1.05 *eq*) in dioxane (5 mL) was added Cs₂CO₃ (836 mg, 2.57 mmol, 3 *eq*), Brettphos Pd G3 (78 mg, 0.08 mmol, 0.1 *eq*) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was filtered. The solid was washed with DCM (5 mL x2). The combined filtrate was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex luna 30*30 mm*10 um+YMC AQ 100*30*10 um; mobile phase: [water (0.05%NH₃H₂O)-ACN]; B%: 30%-90%, 20min). **20** (10 mg, 0.02 mmol, 3.0% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.01 (d, *J* = 2.0 Hz, 1H), 8.61 (s, 1H), 7.86 (s, 1H), 7.53 (d, *J* = 3.4 Hz, 1H), 7.19 (d, *J* = 3.4 Hz, 1H), 7.11 (t, *J* = 8.0 Hz, 1H), 6.97 (t, *J* = 2.2 Hz, 1H), 6.78 – 6.70 (m, 1H), 6.56 – 6.46 (m, 1H), 5.33 (br s, 2H), 2.76 – 2.68 (m, 1H), 0.91 – 0.85 (m, 2H), 0.80 – 0.75 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 150.58, 149.94, 149.58, 149.51, 149.48, 146.45, 136.94, 129.46, 114.12, 113.20, 111.91, 108.98, 107.84, 107.26, 78.55, 78.29, 23.63, 6.51. HPLC t_R = 2.204 min in 8 min chromatography, Xtimate C18 2.1*30mm 3um, purity 95.9%. LCMS t_R = 1.529 min in 4 min chromatography, Xtimate C18,3um,2.1*30mm, MS ESI calcd. for C₁₉H₁₇N₈O⁺ [M+H]⁺ 373.15, found 373.0. 4-(3-Nitrobenzyl)-4H-1,2,4-triazole (**122**). A mixture of 1-(bromomethyl)-3-nitrobenzene (**104**) (1.00 g, 4.63 mmol) and 4H-1,2,4-triazole (352 mg, 5.09 mmol) in MeCN (12 mL) was added K₂CO₃ (1.92 g, 13.89 mmol). Then the mixture was stirred at 100°C for 10 h under N₂ atmosphere. The reaction mixture was concentrated in vacuo. Water (45 mL) was added to the residue. The resulting mixture was extracted with EtOAc (20 mL x3). The combined organic phase was dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. **122** (1.70 g, 70.9%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.73 (s, 1H), 8.20 - 8.18 (m, 2H), 8.03 (s, 1H), 7.75 - 7.73 (m, 1H), 7.69 - 7.67 (m, 1H), 5.60 (s, 2H). LCMS t_R = 0.417 min in 1 min chromatography, MS ESI calcd. for C₉H₉N₄O₂⁺ [M+H]⁺ 205.07, found 205.0.

3-((4H-1,2,4-triazol-4-yl)methyl)aniline (93). A solution of 4-(3-Nitrobenzyl)-4H-1,2,4-triazole (122) (200 mg, 0.86 mmol) in EtOH (8 mL) was stirred at 25°C. Then a solution of NH₄Cl (739 mg, 13.81 mmol) in H₂O (1 mL) was added dropwise. And Fe (1.54 g, 27.62 mmol) was added to the reaction mixture at 25°C. The mixture was degassed and purged with N₂ and stirred at 80°C for 3 hours. The reaction mixture was diluted with MeOH (20 mL) and filtered via a celite pad. The pad was washed with MeOH (15 mL x2). The combined filtrate was concentrated in vacuo. The residue was purified by prep-HPLC (column: Welch Ultimate XB-CN 250*50*10um; mobile phase: [Heptane-EtOH (0.1%NH₃H₂O)]; B%: 15%-35%, 9min). **93** (432 mg, 43%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.62 (s, 1H), 7.95 (s, 1H), 6.97 (t, *J* = 7.6 Hz, 1H), 6.53 -6.50 (m, 1H), 6.44 - 6.41 (m, 2H), 5.24 (s, 2H). LCMS t_R = 0.172 min in 1 min chromatography, MS ESI calcd. for C₉H₁₁N₄⁺ [M+H]⁺ 175.10, found 175.1.

5-((3-((4H-1,2,4-triazol-4-yl)methyl)phenyl)amino)-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (21). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (320 mg, 1.37 mmol) and 3-((4H-1,2,4-triazol-4-yl)methyl)aniline (93) (358 mg, 2.05 mmol,) in dioxane (3 mL) was added Cs₂CO₃ (1.34 g, 4.11 mmol), BINAP (128 mg, 0.25 mmol) and Pd(OAc)₂ (46 mg, 0.25 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~7%, MeOH/DCM). The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 23%-53%, 10min). **21** (133.1 mg, 26.0%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-d₆) δ 9.70 (s, 1H), 8.66 (s, 1H), 8.36 (s, 1H), 8.25 (s, 1H), 8.00 (s, 1H), 7.84 (d, *J* = 8.7 Hz, 1H), 7.50 (t, *J* = 1.9 Hz, 1H), 7.33 (t, *J* = 7.9 Hz, 1H), 6.93 (d, *J* = 8.0 Hz, 1H), 5.98 (s, 1H), 5.41 (s, 2H), 2.65 – 2.55 (m, 1H), 0.84 – 0.77 (m, 2H), 0.75 – 0.68 (m, 2H). ¹³C NMR (101 MHz, DMSO-d₆) δ 156.76, 151.78, 150.79, 148.35, 145.08, 144.27, 140.65, 136.90, 129.09, 121.43, 118.84, 118.46, 114.76, 76.59, 76.41, 52.20, 23.30, 6.52. HPLC t_R = 3.712 min in 8 min chromatography, purity 99.5%. LCMS t_R = 1.494 min in 4 min chromatography, MS ESI calcd. for C₁₉H₁₈N₉⁺ [M+H]⁺ 372.17, found 372.3.

5-((3-(1,3,4-Oxadiazol-2-yl)phenyl)amino)-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**22**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (200 mg, 0.86 mmol, 1 *eq*) and 3-(1,3,4-oxadiazol-2-yl)aniline (**94**) (138 mg, 0.86 mmol, 1 *eq*) in dioxane (3 mL) was added Cs₂CO₃ (837 mg, 2.57 mmol, 3 *eq*), BINAP (80 mg, 0.13 mmol, 0.15 *eq*) and Pd(OAc)₂ (29 mg, 0.13 mmol, 0.15 *eq*) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent: 0-10%, MeOH / DCM). The impure product was purified by prep-HPLC (column: Phenomenex luna 30*30mm*10um+YMC AQ 100*30*10um; mobile phase: [water (NH₃H₂O)-ACN]; B%: 40%-80%, 17min). **22** (34.8 mg, 0.09 mmol) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.98 (s, 1H), 9.35 (s, 1H), 8.51 (t, *J* = 2.0 Hz, 1H), 8.40 (s, 1H), 8.35 (s, 1H), 8.11 – 8.04 (m, 1H), 7.66 (dt, *J* = 7.8, 1.4 Hz, 1H), 7.57 (t, *J* = 7.9 Hz, 1H), 6.03 (s, 1H), 2.66 – 2.59 (m, 1H), 0.87 – 0.81 (m, 2H), 0.77 – 0.70 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 163.75, 156.59, 154.49, 150.64, 148.49, 145.20, 141.35, 129.98, 123.67, 122.19, 119.97, 116.88, 114.55, 76.93, 76.78, 23.33, 6.55. HPLC t_R = 4.115 min in 8 min chromatography, purity 98.9%. LCMS t_R = 1.686 min in 4 min chromatography, MS ESI calcd. for C₁₈H₁₅N₈O⁺ [M+H]⁺ 359.14, found 359.3.

3-(*Thiazol-2-yl*)*aniline* (**95**). A mixture of 2-bromothiazole (500 mg, 3.05 mmol, 1 *eq*), 3-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)*aniline* (**123**) (1.34 g, 6.10 mmol, 2 *eq*), Pd(dppf)Cl₂ (299 mg, 0.36 mmol, 0.12 *eq*) and Cs₂CO₃ (2.98 g, 9.15 mmol, 3 *eq*) in dioxane (8 mL) and water (2 mL) was degassed and purged with N₂ for 3 times at 25°C. Then the mixture was stirred at 100°C for 5 h under N₂ atmosphere. The reaction mixture was filtered and the solid was washed with DCM (6 mL x2). The combined filtrate was concentrated in vacuo. The impure product was purified by prep-HPLC (column: column: Xtimate C18 150*40mm*10um; mobile phase: [water (0.05%NH₃H₂O)-ACN]; B%: 20%-50%, 10min). **95** (221 mg, 1.23 mmol) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.86 (d, *J* = 2.4 Hz, 1H), 7.71 (d,

J = 3.2 Hz, 1H), 7.19 (d, J = 1.6 Hz, 1H), 7.14 - 7.06 (m, 2H), 6.65 (dd, J = 0.8, 6.8 Hz, 1H), 5.35 (s, 2H). LCMS t_R = 0.384 min in 1 min chromatography, MS ESI calcd. for C₉H₉N₂S⁺ [M+H]⁺ 177.05, found 177.0.

7-(*cyclopropylamino*)-5-((3-(*thiazol*-2-*yl*)*phenyl*)*amino*)*pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**23**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-*a*]*pyrimidine-3-carbonitrile* (**75**) (200 mg, 0.86 mmol, 1 *eq*) and 3-(Thiazol-2-yl)aniline (**95**) (151 mg, 0.86 mmol, 1 *eq*) in dioxane (3 mL) was added Cs₂CO₃ (837 mg, 2.57 mmol, 3 *eq*), BINAP (80 mg, 0.13 mmol, 0.15 *eq*) and Pd(OAc)₂ (29 mg, 0.13 mmol, 0.15 *eq*) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent: 0-9%, MeOH / DCM). The impure product was purified by prep-HPLC (column: Xtimate C18 150*40mm*10um; mobile phase: [water (NH₃H₂O)-ACN]; B%: 40%-70%, 10min). **23** (13.1 mg, 0.06 mmol) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.94 (s, 1H), 8.61 (s, 1H), 8.38 (s, 1H), 8.29 (s, 1H), 7.93 (t, *J* = 2.8 Hz, 1H), 7.87 – 7.76 (m, 2H), 7.62 (d, *J* = 7.7 Hz, 1H), 7.45 (t, *J* = 7.9 Hz, 1H), 6.05 (s, 1H), 2.65 – 2.57 (m, 1H), 0.87 – 0.79 (m, 2H), 0.76 – 0.70 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 167.50, 156.77, 150.72, 148.44, 145.19, 143.91, 141.32, 133.61, 129.70, 120.52, 120.43, 119.40, 116.82, 114.68, 76.95, 76.86, 23.40, 6.64. HPLC t_R = 4.747 min in 8 min chromatography, purity 99.5%. LCMS t_R = 2.013 min in 4 min chromatography, MS ESI calcd. for C₁₉H₁₆N₇S⁺ [M+H]⁺ 374.12, found 374.3.

3-(1,2,4-Thiadiazol-5-yl)aniline (**96**). A mixture of 5-bromo-1,2,4-thiadiazole (500 mg, 3.03 mmol), 3-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)aniline (**123**) (1.33 g, 6.06 mmol), Pd(dppf)Cl₂ (297 mg, 0.36 mmol) and Cs₂CO₃ (2.96 g, 9.09 mmol) in dioxane (5 mL) and water (2 mL) was degassed and purged with N₂ for 3 times at 25°C. Then the mixture was stirred at 100°C for 5 hours under N₂ atmosphere. The reaction mixture was filtered and the solid was washed with DCM (6 mL x2). The combined filtrate was concentrated in vacuo. The impure product was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₄HCO₃)-ACN]; B%: 12%-42%, 10min). **96** (206 mg, 37.3%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.90 (s, 1H), 7.24-7.22 (m, 1H), 7.20 (d, *J* = 7.6 Hz, 1H), 7.15-7.13 (m, 1H), 6.78 (d, *J* = 8.0 Hz, 1H), 5.50 (s, 2H). LCMS t_R = 0.393 min in 1 min chromatography, MS ESI calcd. for C₈H₈N₃S⁺ [M+H]⁺ 178.04, found 178.0.

5 - ((3-(1,2,4-Thiadiazol-5-yl)phenyl)amino)-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (24). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (200 mg, 0.86 mmol) and 3-(1,2,4-Thiadiazol-5-yl)aniline (96) (136 mg, 0.77 mmol) in dioxane (3 mL) was added Cs₂CO₃ (837 mg, 2.57 mmol), BINAP (80 mg, 0.13 mmol) and Pd(OAc)₂ (29 mg, 0.13 mmol) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent: 0-6%, MeOH / DCM). The product was purified by prep-HPLC (column: Waters Torus 2-PIC 150*19mm*5um; mobile phase: [Heptane-EtOH (0.1%NH₃H₂O)]; B%: 5%-30%, 15min). Then the impure product was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 36%-66%, 10min). **24** (12 mg, 1.9%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.01 (s, 1H), 8.97 (s, 1H), 8.85 (t, *J* = 1.9 Hz, 1H), 8.41 (s, 1H), 8.32 (br s, 1H), 7.84 (dd, *J* = 8.1, 2.2 Hz, 1H), 7.69 (d, *J* = 7.6 Hz, 1H), 7.55 (t, *J* = 7.9 Hz, 1H), 6.03 (s, 1H), 2.63 (tt, *J* = 6.9, 3.6 Hz, 1H), 0.87 - 0.81 (m, 2H), 0.77 - 0.72 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 188.00, 164.35, 156.61, 150.59, 148.48, 145.15, 141.61, 130.12, 130.07, 122.40, 120.09, 117.65, 114.54, 77.04, 76.85, 23.35, 6.56. HPLC t_R = 4.670 min in 8 min chromatography, purity 99.5%. LCMS t_R = 1.847 min in 4 min chromatography, MS ESI calcd. for C₁₈H₁₅N₈S⁺ [M+H]⁺ 375.11, found 375.3.

5-((3-(2H-tetrazol-5-yl)phenyl)amino)-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**25**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-*a*]pyrimidine-3-carbonitrile (**75**) (150 mg, 0.64 mmol) and 3-(2H-tetrazol-5-yl)aniline (**97**) (103 mg, 0.64 mmol) in dioxane (3 mL) was added 'BuOK (216 mg, 1.93 mmol), BINAP (60 mg, 0.10 mmol) and Pd(OAc)₂ (22 mg, 0.10 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~98%, MeOH/DCM). The residue was purified by prep. HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₄HCO₃)-ACN]; B%: 10%-40%, 10min). The residue was purified by SFC (column: DAICEL CHIRALPAK AD (250mm*30mm, 10um); mobile phase: [0.1%NH₃H₂O IPA]; B%: 35%-35%, min).**25**(5.5 mg, 2.3%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d* $₆) <math>\delta$ 9.90 (s, 1H), 8.38 (s, 1H), 8.30 (s, 1H), 8.24 (t, *J* = 1.9 Hz, 1H), 8.12 (d, *J* = 7.4 Hz, 1H), 7.63 (d, *J* = 7.7 Hz, 1H), 7.51 (t, *J* = 7.9 Hz, 1H), 6.05 (s, 1H), 2.64 – 2.59 (m, 1H), 0.86 – 0.80 (m, 3H), 0.76 – 0.71 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 183.77,

156.79, 150.77, 148.46, 145.25, 141.09, 129.71, 123.84, 120.93, 120.32, 117.55, 114.76, 76.74, 76.67, 23.35, 6.60. HPLC $t_R = 1.884 \text{ min}$ in 4 min chromatography, purity 99.0%. LCMS $t_R = 2.239 \text{ min}$ in 7 min chromatography, MS ESI calcd. for $C_{17}H_{15}N_{10}^+$ [M+H]⁺ 359.15, found 359.3.

5-((3-(1H-tetrazol-1-yl)phenyl)amino)-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**26**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (100 mg, 0.43 mmol) and 3-(1H-tetrazol-1-yl)aniline (**98**) (69 mg, 0.43 mmol) in dioxane (3 mL) was added Cs₂CO₃ (418 mg, 1.28 mmol) and Brettphos Pd G₃ (39 mg, 0.04 mmol) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. Water (50 mL) was added to the reaction mixture. The mixture was extracted with DCM (80 mL x2) and the combined organic phase was washed with water (90 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The impure product was purified by prep-HPLC (column: Xtimate C18 150*40mm*10un; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 25%-65%, 10min). **26** (15.7 mg, 2.5%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.12 (s, 1H), 10.02 (s, 1H), 8.52 (t, *J* = 2.1 Hz, 1H), 8.40 (s, 1H), 8.37 (s, 1H), 7.89 (ddd, *J* = 8.3, 2.2, 1.0 Hz, 1H), 7.60 (t, *J* = 8.1 Hz, 1H), 7.51 (ddd, *J* = 8.0, 2.2, 1.0 Hz, 1H), 6.07 (s, 1H), 2.63 (tt, *J* = 6.9, 3.6 Hz, 1H), 0.88 – 0.80 (m, 2H), 0.77 – 0.70 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.59, 150.66, 148.58, 145.20, 142.13, 142.01, 134.07, 130.42, 119.79, 114.60, 114.09, 111.35, 77.07, 76.93, 23.39, 6.59. HPLC t_R = 4.096 min in 8 min chromatography, purity 98.1%. LCMS t_R = 1.615 min in 4 min chromatography, MS ESI calcd. for C₁₇H₁₅N₁₀+ [M+H]⁺ 359.15, found 359.3.

3-Nitrobenzenediazonium tetrafluoroborate (125). To a solution of 3-nitroaniline (124) (500 mg, 3.62 mmol) in EtOH (5 mL) and concentrated HF-BF₃ solution (1.32 g, 7.24 mmol, 48%) was added tert-butyl nitrite (747 mg, 7.24 mmol) at 0°C. The mixture was stirred at 25°C for 1 h. The reaction mixture was diluted with PE (10 mL). The mixture was filtered and the solid was washed with PE (5 mL x2). The solid was dried in vacuo. The crude product was used in the next step directly. 125 (850 mg, 99.1%) was obtained as a light orange solid.

2-(3-Nitrophenyl)-2H-tetrazole (126). To a solution of 3-nitrobenzenediazonium tetrafluoroborate (125) (850 mg, 3.59 mmol) and silver trifluoroacetate (951 mg, 4.31 mmol) in THF (10 mL) was cooled down to -78°C. Then triethylamine (545 mg, 5.38 mmol) was added to the reaction mixture dropwise. After 10 minutes, TMSCHN₂ (2 M, 1.97 mL) was added to the reaction mixture dropwise. After 10 minutes, TMSCHN₂ (2 M, 1.97 mL) was added to the reaction mixture was stirred at -78°C for 1 hour and the mixture was warmed slowly to 25°C. A solution of CsF (1.09 g, 7.18 mmol) in MeOH (10 mL) was added to the reaction mixture dropwise. Then the reaction mixture was stirred at 25°C for 0.5 hour. The reaction mixture was diluted with EtOAc (25 mL) and brine (10 mL). The organic phase was separated, dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The crude product was used in the next step directly. **126** (600 mg, crude) was obtained as a black brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.38 (s, 1H), 8.80 (t, *J* = 2.0 Hz, 1H), 8.59 - 8.57 (m, 1H), 8.47 - 8.45 (m, 1H), 7.99 (t, *J* = 8.4 Hz, 1H)

3-(2H-tetrazol-2-yl)aniline (99). To a solution of 2-(3-Nitrophenyl)-2H-tetrazole (**126**) (600 mg, 3.14 mmol) in EtOH (8 mL) was added NH₄Cl (504 mg, 9.42 mmol) which was dissolved in H₂O (2 mL). Then Fe (1.05 g, 18.83 mmol) was added to the mixture at 25°C. Then the mixture was degassed, purged with N₂ and stirred at 80°C for 3 hours. The reaction mixture was diluted with MeOH (10 mL) and filtered. The solid was washed with MeOH (10 mL x2). The combined filtrate was concentrated in vacuo. The combined filtrate was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₄HCO₃)-ACN]; B%: 15%-45%, 10min). **99** (153 mg, 48.4%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.18 (s, 1H), 7.30 (s, 1H), 7.28 - 7.24 (m, 1H), 7.19 - 7.17 (m, 1H), 6.73 (d, *J* = 7.6 Hz, 1H), 5.70 (s, 2H). LCMS t_R = 0.381 min in 1 min chromatography, MS ESI calcd. for C₇H₈N₅⁺ [M+H]⁺ 162.08, found 162.1.

5-((3-(2H-tetrazol-2-yl)phenyl)amino)-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (27). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (180 mg, 0.77 mmol) and 3-(2H-tetrazol-2-yl)aniline (99) (124 mg, 0.77 mmol) in dioxane (4 mL) was added Cs₂CO₃ (753 mg, 2.31 mmol), BINAP (72 mg, 0.12 mmol) and Pd(OAc)₂ (29 mg, 0.12 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5h. The reaction mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0%~4% MeOH/DCM) to give the product. The impure product was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₄HCO₃)-ACN]; B%: 37%-67%, 10min). **27** (35.8 mg, 12.9%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.08 (s, 1H), 9.25 (s, 1H), 8.72 (t, *J* = 2.2 Hz, 1H), 8.41 (s, 1H), 8.37 (s, 1H), 7.99 – 7.95 (m, 1H), 7.76 – 7.69 (m, 1H), 7.62 (t, *J* = 8.1 Hz, 1H), 6.05 (s, 1H), 2.66

-2.58 (m, 1H), 0.88 - 0.82 (m, 2H), 0.77 - 0.72 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.52, 153.81, 150.54, 148.54, 145.28, 141.94, 136.52, 130.47, 120.04, 114.47, 112.88, 110.02, 77.09, 76.94, 23.35, 6.56. HPLC t_R = 4.643 min in 8 min chromatography, purity 99.7%. LCMS t_R = 1.856 min in 4 min chromatography, MS ESI calcd. for C₁₇H₁₅N₁₀⁺ [M+H]⁺ 359.15, found 359.4.

7-(*Cyclopropylamino*)-5-((*3*-(*pyridin*-2-*yl*)*phenyl*)*amino*)*pyrazolo*[*1*,5-*a*]*pyrimidine*-*3*-*carbonitrile* (**28**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-*a*]*pyrimidine*-3-carbonitrile (**75**) (200 mg, 0.86 mmol, 1 *eq*) and 3-(pyridin 2-yl)aniline (**100**) (146 mg, 0.86 mmol, 1 *eq*) in dioxane (3 mL) was added Cs₂CO₃ (837 mg, 2.57 mmol, 3 *eq*), BINAP (80 mg, 0.13 mmol, 0.15 *eq*) and Pd(OAc)₂ (29 mg, 0.13 mmol, 0.15 *eq*) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent: 0-10%, MeOH / DCM). The impure product was purified by prep-HPLC (column: Phenomenex luna 30*30mm*10um+YMC AQ 100*30*10um; mobile phase: [water (NH₃H₂O)-ACN]; B%: 45%-85%, 17min). **28** (13.4 mg, 0.06 mmol) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.86 (s, 1H), 8.72 (t, *J* = 2.0 Hz, 1H), 8.67 (ddd, *J* = 4.7, 1.8, 0.9 Hz, 1H), 8.38 (s, 1H), 8.27 (d, *J* = 1.8 Hz, 1H), 7.97 (dt, *J* = 8.2, 1.1 Hz, 1H), 7.36 (ddd, *J* = 7.5, 4.8, 1.1 Hz, 1H), 6.05 (s, 1H), 2.65 – 2.57 (m, 1H), 0.86 – 0.80 (m, 2H), 0.76 – 0.71 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.87, 155.96, 150.90, 149.52, 148.40, 145.01, 140.96, 139.17, 137.22, 129.19, 122.70, 120.11, 119.71, 117.52, 114.85, 76.71, 76.61, 23.35, 6.58. HPLC t_R = 3.540 min in 8 min chromatography, purity 99.0%. LCMS t_R = 1.384 min in 4 min chromatography, MS ESI calcd. for C₂₁H₁₈N₇+ [M+H]+ 368.16, found 368.3.

3-(Pyrazin-2-yl) aniline (101). A mixture of 2-bromopyrazine (1.00 g, 6.29 mmol), 3-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)aniline (123) (2.76 g, 12.58 mmol), Pd(dppf)Cl₂ (616 mg, 0.75 mmol) and Cs₂CO₃ (6.15 g, 18.87 mmol) in dioxane (10 mL) and water (3 mL) was degassed and purged with N₂ for 3 times at 25°C. Then the mixture was stirred at 100°C for 5 hours under N₂ atmosphere. The reaction mixture was filtered and the solid was washed with DCM (10 mL x 2). The combined filtrate was concentrated in vacuo. The residue was purified by prep-HPLC (column: Welch Ultimate XB-NH2 250mm*100mm*10u; mobile phase: [Heptane-EtOH (0.1% NH₃H₂O)]; B%: 5%-40%, 12min). The impure product was purified by prep-HPLC (column: Xtimate C18 150*40mm*10ur; mobile phase: [water (NH₃H₂O)-ACN]; B%: 10%-40%, 10min). **101** (627 mg, 57.1%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.10 (s, 1H), 8.68 - 8.67 (m, 1H), 8.56 (d, *J* = 2.4 Hz, 1H), 7.37 (s, 1H), 7.27 - 7.25 (m, 1H), 7.20 - 7.18 (m, 1H), 6.72 (d, *J* = 2.0 Hz, 1H), 5.33 (s, 2H). LCMS t_R = 0.323 min in 1 min chromatography, MS ESI calcd. for C₁₀H₁₀N₃⁺ [M+H]⁺ 172.09, found 172.0.

7-(*Cyclopropylamino*)-5-((*3*-(*pyrazin*-2-*yl*)*phenyl*)*amino*)*pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**29**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-*a*]pyrimidine-3-carbonitrile (**75**) (200 mg, 0.86 mmol) and 3-(pyrazin-2-yl) aniline (**101**) (147 mg, 0.86 mmol) in dioxane (4 mL) was added Cs₂CO₃ (837 mg, 2.57 mmol), BINAP (80 mg, 0.13 mmol) and Pd(OAc)₂ (29 mg, 0.13 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 49%~67%, EtOAC/PE) to give the product. The impure product was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₄HCO₃)-ACN]; B%:37%-67%, 10min). **29** (16.9 mg, 7.3%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.89 (s, 1H), 9.23 (d, *J* = 1.5 Hz, 1H), 8.73 (dd, *J* = 2.5, 1.5 Hz, 1H), 8.68 (t, *J* = 2.0 Hz, 1H), 8.63 (d, *J* = 2.5 Hz, 1H), 8.38 (s, 1H), 8.29 (d, *J* = 1.8 Hz, 1H), 7.93 – 7.87 (m, 1H), 7.83 – 7.76 (m, 1H), 7.51 (t, *J* = 7.9 Hz, 1H), 6.05 (s, 1H), 2.66 – 2.57 (m, 1H), 0.86 – 0.79 (m, 2H), 0.77 – 0.72 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.78, 151.40, 150.81, 148.41, 145.06, 144.29, 143.50, 141.79, 141.18, 136.39, 129.50, 120.55, 120.24, 117.47, 114.71, 76.76, 76.63, 23.33, 6.55. HPLC t_R = 4.595 min in 8 min chromatography, purity 96.7%. LCMS t_R = 1.835 min in 4 min chromatography, MS ESI calcd. for C₂₀H₁₇N₈⁺ [M+H]⁺ 369.16, found 369.4.

Experimental Procedures in Scheme 3.

5-Chloro-7-(cyclobutylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (127). To a solution of 5,7dichloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (74) (350 mg, 1.64 mmol) in EtOH (5 mL) was added cyclobutylamine (234 mg, 3.29 mmol) dropwise at 25°C. Then the mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated in vacuo. The crude compound was used in the next step. **127** (314 mg) was obtained as a light yellow solid. ¹H NMR (DMSO- d_6 , 400 MHz) δ 9.14 (d, J = 6.0 Hz, 1H), 8.67 (s, 1H), 6.50 (s, 1H), 4.39 - 4.17 (m, 1H), 2.40 - 2.18 (m, 4H), 1.79 - 1.57 (m, 2H). LCMS t_R = 0.519 min in 1 min chromatography, Chromolith Flash RP-18, 5um, 3.0*25mm, MS ESI calcd. for C₁₁H₁₁ClN₅⁺ [M+H]⁺ 248.07, found 248.0.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-(cyclobutylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**30**). To a solution of 5-chloro-7-(cyclobutylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**127**) (105 mg, 0.42 mmol) and 3-(4H-1,2,4-triazol-4-yl)aniline (**86**) (68 mg, 0.42 mmol) in dioxane (3 mL) was added 'BuOLi (102 mg, 1.27 mmol), BINAP (40 mg, 0.06 mmol) and Pd(OAc)₂ (14 mg, 0.06 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~2%, MeOH/DCM). The residue was purified by prep. HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃) -ACN]; B%: 26%-56%, 11min). **30** (30 mg, 19.0%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.85 (s, 1H), 9.02 (s, 2H), 8.42 (s, 1H), 8.34 (d, *J* = 1.9 Hz, 1H), 8.26 (d, *J* = 6.2 Hz, 1H), 7.60 (dd, *J* = 8.1, 2.0 Hz, 1H), 7.51 (t, *J* = 8.1 Hz, 1H), 7.31 (dd, *J* = 7.9, 2.1 Hz, 1H), 5.67 (s, 1H), 4.02 (h, *J* = 7.8 Hz, 1H), 2.40 – 2.29 (m, 2H), 2.29 – 2.16 (m, 2H), 1.85 – 1.71 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.59, 150.85, 146.18, 144.94, 141.78, 141.18, 134.18, 130.29, 118.46, 114.64, 114.53, 111.75, 77.02, 75.90, 46.95, 29.06, 14.94. HPLC t_R = 2.721 min in 8 min chromatography, purity 99.2%. LCMS t_R = 1.659 min in 4 min chromatography, MS ESI calcd. for C₁₉H₁₈N₉+ [M+H]⁺ 372.17, found 372.3.

5-chloro-7-(oxetan-3-ylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (128). To a solution of 5,7dichloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (74) (2.00 g, 9.39 mmol, 1 eq) in EtOH (20 mL) was added oxetan-3amine (1.10 g, 15.05 mmol, 14.6 mL, 1.6 eq) dropwise at 25°C. Then the mixture was stirred at 25°C for 2 hours. The reaction mixture was filtered and the solid was washed with EtOH (15 mL x2). The residue was purified by flash silica gel chromatography (eluent 0-4%, MeOH in DCM). **128** (1.80 g, 6.7 mmol, 71.33% yield) was obtained as a light yellow solid. ¹H NMR (400 MHz, DMSO-d₆) δ 9.56 (br s, 1H), 8.71 (s, 1H), 6.48 (s, 1H), 5.09 - 4.95 (m, 1H), 4.87 - 4.72 (m, 4H). LCMS t_R = 0.424 min in 1 min chromatography, MS ESI calcd. for C₁₀H₉ClN₅O⁺ [M+H]⁺ 250.05, found 250.0.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-(oxetan-3-ylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**128**) (100 mg, 0.40 mmol, 1 eq) and 3-(4H-1,2,4-triazol-4-yl)aniline (**86**) (64 mg, 0.40 mmol, 1 eq) in dioxane (5 mL) was added Cs₂CO₃ (391 mg, 1.20 mmol, 3 eq) BINAP (37 mg, 0.06 mmol, 0.15 eq) and Pd(OAc)₂ (13 mg, 0.06 mmol, 0.15 eq) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was concentrated in vacuo. The impure product was purified by prep-HPLC (column: Phenomenex luna 30*30mm*10um+YMC AQ 100*30*10um; mobile phase: [water (0.05%NH₃H₂O) -ACN]; B%: 25%-55%, 20min). **31** (4.3 mg, 0.01 mmol, 2.77% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-d₆) δ 9.88 (s, 1H), 9.02 (s, 2H), 8.71 (d, *J* = 3.5 Hz, 1H), 8.46 (s, 1H), 8.32 (t, *J* = 2.1 Hz, 1H), 7.60 (d, *J* = 8.2 Hz, 1H), 7.52 (t, *J* = 8.0 Hz, 1H), 7.32 (d, *J* = 7.8 Hz, 1H), 5.52 (s, 1H), 4.88 – 4.80 (m, 2H), 4.81 – 4.73 (m, 3H). ¹³C NMR (126 MHz, DMSO-d₆) δ 156.53, 150.91, 146.22, 145.15, 141.65, 141.21, 134.21, 130.35, 118.57, 114.71, 114.58, 111.84, 77.18, 76.00, 75.83, 46.46. HPLC t_R = 3.232 min in 8 min chromatography, purity 96.4%. LCMS t_R = 1.200 min in 4 min chromatography, MS ESI calcd. for C₁₈H₁₆N₉O⁺ [M+H]⁺ 374.15, found 374.3.

5-*Chloro-7-((3,3-difluorocyclobutyl)amino)pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**129**). To a solution of 5,7dichloropyrazolo[1,5-*a*]*pyrimidine-3-carbonitrile* (**74**) (200 mg, 0.94 mmol, 1 *eq*) in EtOH (5 mL) was 3,3difluorocyclobutan-1-amine (404 mg, 2.82 mmol, 3 *eq*, HCl) and Et₃N (304 mg, 3 mmol, 0.42 mL, 3.2 *eq*) dropwise at 25°C. Then the mixture was stirred at 25°C for 2 hours. The reaction mixture was filtered and the solid was washed with EtOH (4 mL x2). **129** (158 mg, 0.55 mmol, 58.49% yield) was obtained as a light yellow solid. ¹H NMR (400 MHz, DMSO*d*₆) δ 9.33 (br s, 1H), 8.71 (s, 1H), 6.66 (s, 1H), 4.35 - 4.16 (m, 1H), 3.20 - 2.89 (m, 4H). LCMS t_R = 0.511 min in 1 min chromatography, Chromolith Flash RP-18, 5um, 3.0*25mm, MS ESI calcd. for C₁₁H₉ClF₂N₅⁺ [M+H]⁺ 284.05, found 284.0.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((3,3-difluorocyclobutyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (32). To a solution of 5-chloro-7-((3,3-difluorocyclobutyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (129) (200 mg, 0.70 mmol, 1 eq) and 3-(4H-1,2,4-triazol-4-yl)aniline (86) (112 mg, 0.70 mmol, 1 eq) in dioxane (5 mL) was added 'BuOLi (169 mg, 2.12 mmol, 3 eq), BINAP (65 mg, 0.10 mmol, 0.15 eq) and Pd(OAc)₂ (23 mg, 0.10 mmol, 0.15 eq) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction

mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent: 0-5%, MeOH / DCM). The impure product was purified by prep-HPLC (column: Phenomenex Gemini-NX C18 75*30mm*3um; mobile phase: [water (10mM NH₄HCO₃)-ACN]; B%: 26%-56%, 10min). **32** (5.5 mg, 0.01 mmol, 1.9% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.88 (s, 1H), 9.03 (s, 2H), 8.50 (s, 1H), 8.45 (s, 1H), 8.34 (t, *J* = 2.2 Hz, 1H), 7.60 (ddd, *J* = 8.3, 2.1, 1.0 Hz, 1H), 7.52 (t, *J* = 8.0 Hz, 1H), 7.32 (ddd, *J* = 7.9, 2.2, 1.0 Hz, 1H), 5.69 (s, 1H), 4.02 (qd, *J* = 8.6, 7.7, 4.8 Hz, 1H), 3.14 – 2.90 (m, 4H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.59, 150.89, 146.60, 145.16, 141.70, 141.27, 134.26, 130.42, 119.42 (dd, *J* = 282.5, 268.2 Hz), 118.55, 114.73, 114.65, 111.79, 77.24, 76.55, 41.50 (t, *J* = 22.5 Hz), 36.58 (dd, *J* = 19.3, 6.2 Hz). HPLC t_R = 3.938 min in 8 min chromatography, purity 99.1%. LCMS t_R = 2.161 min in 4 min chromatography, MS ESI calcd. for C₁₉H₁₆F₂N₉⁺ [M+H]⁺ 408.15, found 408.3.

5-*Chloro-7-(((tetrahydrofuran-3-yl)methyl)amino)pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**130**). To a solution of 5,7dichloropyrazolo[1,5-*a*]pyrimidine-3-carbonitrile (**74**) (500 mg, 2.35 mmol) in EtOH (5 mL) was added (tetrahydrofuran-3-yl)methanamine (475 mg, 4.69 mmol) dropwise at 25°C. Then the mixture was stirred at 25°C for 2 hours. The reaction mixture was filtered. The filter cake was washed with EtOH (4 mL x2). The crude compound was used in next step. **130** (830 mg, crude) was obtained as a light yellow solid. ¹H NMR (DMSO-*d*₆, 400 MHz) δ 9.04 (s, 1H), 8.66 (s, 1H), 6.69 (s, 1H), 3.82 - 3.72 (m, 1H), 3.72 - 3.65 (m, 1H), 3.63 - 3.59 (m, 1H), 3.52 - 3.43 (m, 3H), 2.70 - 2.56 (m, 1H), 2.14 - 1.86 (m, 1H), 1.73 - 1.52 (m, 1H). LCMS t_R = 0.466 min in 1 min chromatography, Chromolith Flash RP-18, 5um, 3.0*25mm, MS ESI calcd. for C₁₂H₁₃ClN₅O⁺ [M+H]⁺ 278.08, found 278.0.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-(((tetrahydrofuran-3-yl)methyl)amino)pyrazolo[1,5-a]pyrimidine-3*carbonitrile* (33). To a solution of 5-chloro-7-(((tetrahydrofuran-3-yl)methyl)amino)pyrazolo[1,5-a]pyrimidine-3carbonitrile (130) (200 mg, 0.72 mmol, 1 eq) and 3-(4H-1,2,4-triazol-4-yl)aniline (86) (115 mg, 0.72 mmol, 1 eq) in dioxane (5 mL) was added 'BuOLi (172 mg, 2.16 mmol, 3 eq) BINAP (67 mg, 0.10 mmol, 0.15 eq) and Pd(OAc)₂ (24 mg, 0.10 mmol, 0.15 eq) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent: 0-11%, MeOH / DCM). The impure product was purified by prep-HPLC (column: Phenomenex Gemini-NX C18 75*30mm*3um; mobile phase: [water (10mM NH₄HCO₃)-ACN]; B%: 26%-56%, 10min). **33** (14.2 mg, 0.03 mmol, 4.8% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.01 (s, 1H), 8.17 (s, 2H), 7.57 (s, 1H), 7.50 (t, J = 2.1 Hz, 1H), 7.41 (t, J = 6.1 Hz, 1H), 6.75 (ddd, J = 8.3, 2.1, 1.0 Hz, 1H), 6.66 (t, J = 8.1 Hz, 1H), 6.46 $(ddd, J = 7.9, 2.2, 1.0 \text{ Hz}, 1\text{H}), 4.93 (s, 1\text{H}), 2.94 (td, J = 8.1, 5.7 \text{ Hz}, 1\text{H}), 2.84 (dd, J = 8.7, 6.8 \text{ Hz}, 1\text{H}), 2.78 (td, J = 8.2, 10 \text{ Hz}, 10 \text{ Hz}), 2.78 (td, J = 8.2, 10 \text{ Hz}), 3.10 \text{ Hz}, 10 \text{ Hz}), 3.10 \text{ Hz}), 3.10 \text{ Hz}, 10 \text{ Hz}), 3.10 \text{ H$ 6.7 Hz, 1H), 2.70 (dd, J = 8.6, 4.7 Hz, 1H), 2.42 (t, J = 6.6 Hz, 2H), 1.82 (dtd, J = 14.2, 7.1, 6.5, 3.6 Hz, 1H), 1.13 (dtd, J = 14.2, 7.1, 6.5, 7.1, 6.5, 7.1) 12.3, 8.0, 5.7 Hz, 1H), 0.88 – 0.75 (m, 1H). ¹³C NMR (101 MHz, DMSO-d₆) δ 156.71, 150.85, 147.33, 145.09, 141.88, 141.27, 134.24, 130.38, 118.48, 114.74, 114.55, 111.72, 77.11, 75.18, 70.40, 66.81, 44.45, 37.56, 29.39. HPLC $t_R = 3.500$ min in 8 min chromatography, purity 97.8%. LCMS $t_{R} = 1.913$ min in 4 min chromatography, MS ESI calcd. for $C_{20}H_{20}N_9O^+$ [M+H]⁺ 402.18, found 402.3.

5-*Chloro-7*-(((*tetrahydro-2H-pyran-4-yl*)*methyl*)*amino*)*pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**131**). To a solution of 5,7-dichloropyrazolo[1,5-*a*]pyrimidine-3-carbonitrile (**74**) (200 mg, 0.94 mmol) in EtOH (5 mL) was added (tetrahydro-2H-pyran-4-yl)methanamine (216 mg, 1.88 mmol) dropwise at 25°C. Then the mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated in vacuo. The crude compound was used in the next step. **131** (273 mg) was obtained as a light yellow solid. ¹H NMR (DMSO-*d*₆, 400 MHz) δ 9.05 - 8.88 (m, 1H), 8.65 (s, 1H), 6.68 (s, 1H), 3.93 - 3.76 (m, 2H), 3.35-3.31 (m, 2H), 3.27 - 3.19 (m, 2H), 2.02 - 1.86 (m, 1H), 1.70 - 1.52 (m, 2H), 1.32 - 1.13 (m, 2H). LCMS t_R = 0.484 min in 1 min chromatography, Chromolith Flash RP-18, 5um, 3.0*25mm, MS ESI calcd. for C₁₃H₁₅ClN₅O⁺ [M+H]⁺ 292.10, found 292.0.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-(((tetrahydro-2H-pyran-4-yl)methyl)amino)pyrazolo[1,5-a]pyrimidine-3carbonitrile (34). To a solution of 5-chloro-7-(((tetrahydro-2H-pyran-4-yl)methyl)amino)pyrazolo[1,5-a]pyrimidine-3carbonitrile (131) (200 mg, 0.68 mmol, 1 eq) and 3-(4H-1,2,4-triazol-4-yl)aniline (86) (109 mg, 0.68 mmol, 1 eq) in dioxane(5 mL) was added 'BuOLi (164 mg, 2.06 mmol, 3 eq), BINAP (64 mg, 0.10 mmol, 0.15 eq) and Pd(OAc)₂ (23 mg, 0.10mmol, 0.15 eq) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at130°C for 0.5 hour. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gelchromatography (eluent: 0-11%, MeOH / DCM). The impure product was purified by prep-HPLC (column: PhenomenexGemini-NX C18 75*30mm*3um; mobile phase: [water (10mM NH₄HCO₃)-ACN]; B%: 26%-56%, 10min).**34**(11.1 mg, 0.02 mmol, 3.6% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.98 (s, 1H), 8.18 (s, 2H), 7.57 (s, 1H), 7.49 (t, *J* = 2.1 Hz, 1H), 7.34 (t, *J* = 6.1 Hz, 1H), 6.76 (ddd, *J* = 8.2, 2.1, 1.0 Hz, 1H), 6.66 (t, *J* = 8.1 Hz, 1H), 6.46 (ddd, *J* = 7.9, 2.2, 1.0 Hz, 1H), 4.92 (s, 1H), 3.01 (ddd, *J* = 11.5, 4.5, 1.9 Hz, 2H), 2.42 (td, *J* = 11.7, 2.0 Hz, 2H), 2.34 (t, *J* = 6.5 Hz, 2H), 1.12 (ddp, *J* = 11.2, 7.3, 3.5 Hz, 1H), 0.86 – 0.72 (m, 2H), 0.40 (qd, *J* = 12.0, 4.4 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.68, 150.83, 147.44, 145.08, 141.89, 141.26, 134.23, 130.38, 118.45, 114.75, 114.51, 111.69, 77.06, 75.31, 66.65, 47.30, 33.78, 30.49. HPLC t_R = 3.644 min in 8 min chromatography, purity 92.3%. LCMS t_R = 2.160 min in 4 min chromatography, MS ESI calcd. for C₂₁H₂₂N₉O⁺ [M+H]⁺ 416.19, found 416.0.

5-*Chloro-7*-(((1-methyl-1H-pyrazol-4-yl)methyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**132**). To a solution of 5,7-dichloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (**74**) (200 mg, 0.94 mmol) in EtOH (5 mL) was added (1-methyl-1H-pyrazol-4-yl)methanamine (209 mg, 1.88 mmol) dropwise at 25°C. Then the mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated in vacuo. The crude compound was used in the next step. **132** (293 mg) was obtained as a light yellow solid. ¹H NMR (DMSO-*d*₆, 400 MHz) δ 8.66 (s, 1H), 7.81 - 7.63 (m, 1H), 7.55 - 7.39 (m, 1H), 6.64 (s, 1H), 4.49 (d, *J* = 6.4 Hz, 2H), 3.82 (s, 1H), 3.77 (s, 3H). LCMS t_R = 0.447 min in 1 min chromatography, Chromolith Flash RP-18, 5um, 3.0*25mm, MS ESI calcd. for C₁₂H₁₁ClN₇⁺ [M+H]⁺ 288.08, found 288.0.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-(((1-methyl-1H-pyrazol-4-yl)methyl)amino)pyrazolo[1,5-a]pyrimidine-3carbonitrile (35). To a solution of 5-chloro-7-(((1-methyl-1H-pyrazol-4-yl)methyl)amino)pyrazolo[1,5-a]pyrimidine-3carbonitrile (132) (200 mg, 0.70 mmol, 1 eq) and 3-(4H-1,2,4-triazol-4-yl)aniline (86) (111 mg, 0.70 mmol, 1 eq) in dioxane (5 mL) was added 'BuOLi (167 mg, 2.09 mmol, 3 eq), BINAP (65 mg, 0.1 mmol, 0.15 eq) and Pd(OAc)₂ (23 mg, 0.10 mmol, 0.15 eq) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent: 0-5%, MeOH / DCM). The residue was purified by flash silica gel chromatography (eluent of 0~5% MeOH/DCM). Then the impure product was purified by prep-HPLC (column: Phenomenex Gemini-NX C18 75*30mm*3um; mobile phase: [water (10mM NH4HCO3)-ACN]; B%: 26%-56%, 10min). The residue was purified by SFC (column: DAICEL CHIRALCEL OJ (250mm*30mm, 10um); mobile phase: [0.1%NH₃H₂O EtOH]; B%: 60%-60%, min). The impure product was purified by prep-HPLC (column: Phenomenex Gemini-NX C18 75*30mm*3um; mobile phase: [water (10mM NH₄HCO₃)-ACN]; B%: 18%-48%, 10min). **35** (13.1 mg, 0.06 mmol) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.84 (s, 1H), 9.01 (s, 2H), 8.45 (s, 1H), 8.41 (s, 1H), 8.31 (t, *J* = 2.1 Hz, 1H), 7.64 (s, 1H), 7.60 (ddd, J = 8.3, 2.1, 1.0 Hz, 1H), 7.50 (t, J = 8.1 Hz, 1H), 7.45 – 7.43 (m, 1H), 7.30 (ddd, J = 7.9, 2.2, 1.0 Hz, 1H), 5.75 (s, 1H), 4.37 (s, 2H), 3.78 (s, 3H). ¹³C NMR (101 MHz, DMSO-d₆) δ 156.56, 150.77, 147.11, 145.09, 141.81, 141.24, 137.82, 134.21, 130.33, 129.34, 118.50, 117.15, 114.70, 114.57, 111.83, 77.05, 75.69, 38.52, 36.05. HPLC $t_R = 3.389$ min in 8 min chromatography, purity 96.2%. LCMS $t_R = 1.285$ min in 4 min chromatography, MS ESI calcd. for $C_{20}H_{18}N_{11}^+$ [M+H]⁺ 412.17, found 412.3.

7-((1-Acetylpiperidin-4-yl)amino)-5-chloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (133). To a solution of 5,7dichloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (74) (220 mg, 1.03 mmol, 1 eq) in EtOH (5 mL) was added 1-(4-(aminomethyl)piperidin-1-yl)ethan-1-one (734 mg, 5.16 mmol, 5 eq) at 25°C. Then the mixture was stirred at 25°C for 2 hours. The reaction mixture was filtered and the solid was washed with EtOH (4 mL x2). 133 (300 mg, 0.90 mmol, 87.16% yield) was obtained as a light yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.66 (s, 1H), 8.57 - 7.78 (m, 1H), 6.79 (s, 1H), 4.47-4.41 (m, 1H), 3.98 (br t, *J* = 10.8 Hz, 1H), 3.87 (br d, *J* = 13.2 Hz, 1H), 3.15 (t, *J* = 12.8 Hz, 1H), 2.63 (t, *J* = 12.4 Hz, 1H), 2.01 (s, 3H), 1.94 - 1.80 (m, 2H), 1.75 - 1.49 (m, 2H). LCMS t_R = 0.447 min in 1 min chromatography, Chromolith Flash RP-18, 5um, 3.0*25mm, MS ESI calcd. for C₁₄H₁₆CIN₆O⁺ [M+H]⁺ 319.11, found 319.1.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((1-acetylpiperidin-4-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**36**). To a solution of 7-((1-acetylpiperidin-4-yl)amino)-5-chloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (**133**) (200 mg, 0.63 mmol, 1 *eq*) and 3-(4H-1,2,4-triazol-4-yl)aniline (**86**) (101 mg, 0.63 mmol, 1 *eq*) in dioxane (3 mL) was added Cs₂CO₃ (613 mg, 1.88 mmol, 3 *eq*) and Brettphos Pd G₃ (57 mg, 0.06 mmol, 0.1 *eq*) at 25°C. The mixture was degassed and purged with N₂. Then the mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent: 0-10%, MeOH/DCM). The impure product was purified by prep-HPLC (column: Waters Torus 2-PIC 150*19mm*5um; mobile phase: [Heptane-EtOH (0.1%NH₃H₂O)]; B%: 5%-60%, 15min). **36** (46.6 mg, 0.10 mmol, 16.07% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.88 (s, 1H), 9.02 (s, 2H), 8.42 (s, 1H), 8.37 (t, *J* = 2.1 Hz, 1H), 7.88 (d, *J* = 8.2 Hz, 1H), 7.59 (dd, *J* = 8.2, 2.0 Hz, 1H), 7.50 (t, J = 8.1 Hz, 1H), 7.31 (dd, J = 7.9, 2.2 Hz, 1H), 5.85 (s, 1H), 4.44 (d, J = 13.2 Hz, 1H), 3.91 (d, J = 13.7 Hz, 1H), 3.66 (dtt, J = 11.6, 8.2, 4.2 Hz, 1H), 3.19 – 3.07 (m, 1H), 2.64 (ddd, J = 14.7, 12.6, 2.8 Hz, 1H), 2.03 (s, 3H), 1.96 (t, J = 14.4 Hz, 2H), 1.69 (qd, J = 12.3, 4.1 Hz, 1H), 1.56 (qd, J = 12.5, 4.3 Hz, 1H). ¹³C NMR (101 MHz, DMSO- d_6) δ 168.10, 156.70, 150.94, 146.33, 144.96, 141.84, 141.21, 134.20, 130.31, 118.42, 114.63, 114.50, 111.67, 77.12, 75.62, 49.58, 44.70, 31.09, 30.34, 28.98, 21.31. HPLC t_R = 3.429 min in 8 min chromatography, purity 95.75%. LCMS t_R = 1.482 min in 4 min chromatography, MS ESI calcd. for C₂₂H₂₃N₁₀O⁺ [M+H]⁺ 443.21, found 443.0.

7-((2-((Tert-butyldimethylsilyl)oxy)ethyl)amino)-5-chloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (134). To a solution of 5,7-dichloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (74) (500 mg, 2.35 mmol) in EtOH (5 mL) was added 2-((tert-butyldimethylsilyl)oxy)ethan-1-amine (412 mg, 2.35 mmol) at 25°C. Then the mixture was stirred at 25°C for 2 h. The residue was purified by flash silica gel chromatography (eluent of DCM) to give the product. 134 (756 mg, 48.9%) was obtained as a white solid. LCMS t_R = 0.753 min in 1.5 min chromatography, MS ESI calcd. for C₁₅H₂₃ClN₅OSi⁺ [M+H]⁺ 352.14, found 352.0.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((2-((tert-butyldimethylsilyl)oxy)ethyl)amino)pyrazolo[1,5-a]pyrimidine-7-((2-((Tert-butyldimethylsilyl)oxy)ethyl)amino)-5-3-carbonitrile (135). То a solution of chloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (134) (250 mg, 0.39 mmol) and 3-(4H-1,2,4-triazol-4-yl)aniline (86) (125 mg, 0.78 mmol) in dioxane (5 mL) was added Cs₂CO₃ (382 mg, 1.17 mmol), BINAP (36 mg, 0.06 mmol) and Pd(OAc)₂ (13 mg, 0.06 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 2 h. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Welch Ultimate XB-CN 250*50*10um; mobile phase: [Heptane-EtOH(0.1%NH₃H₂O)]; B%: 10%-50%, 15min). **135** (227 mg, 34.2%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.84 (s, 1H), 9.02 (s, 2H), 8.42 (s, 1H), 8.34 (s, 1H), 7.91 (t, J = 6.0 Hz, 1H), 7.62-7.56 (m, 1H), 7.50 (t, J = 8.1 Hz, 1H), 7.33-7.29 (m, 1H), 5.83 (s, 1H), 3.82 (t, J = 5.6 Hz, 2H), 3.45-3.39 (m, 2H), 0.80 (s, 9H), 0.03 (s, 6H). LCMS $t_R = 1.633 \text{ min in 1 min chromatography}, MS ESI$ calcd. for $C_{23}H_{30}N_9OSi^+$ [M+H]⁺ 476.23, found 476.2.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((2-hydroxyethyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**37**). To a solution of 5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((2-((tert-butyldimethylsilyl)oxy)ethyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**135**) (100 mg, 0.21 mmol) in DCM (8 mL) was added TFA (2 mL) at 25°C. The mixture was stirred at 35°C for 2 h. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: C18-6 100*30mm*5um; mobile phase: [water (FA)-ACN]; B%: 0%-60%, 15 min). **37** (45.6 mg, 58.7%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.86 (s, 1H), 9.02 (s, 2H), 8.42 (s, 1H), 8.34 (t, *J* = 2.1 Hz, 1H), 7.87 (t, *J* = 6.0 Hz, 1H), 7.66 – 7.56 (m, 1H), 7.51 (t, *J* = 8.1 Hz, 1H), 7.31 (dd, *J* = 8.2, 2.2 Hz, 1H), 5.82 (s, 1H), 4.97 (t, *J* = 5.5 Hz, 1H), 3.66 (q, *J* = 5.7 Hz, 2H), 3.37 (q, *J* = 5.9 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.69, 150.71, 147.58, 145.03, 141.88, 141.24, 134.20, 130.30, 118.52, 114.69, 114.51, 111.75, 76.98, 75.35, 59.03, 44.24. HPLC t_R = 3.021 min in 8 min chromatography, purity 93.3%. LCMS t_R = 1.719 min in 4 min chromatography, MS ESI calcd. for C₁₇H₁₆N₉O⁺ [M+H]⁺ 362.15, found 362.2.

Experimental Procedures in Scheme 4.

1-Methyl-1H-imidazol-4-amine (137). To a solution of 1-methyl-4-nitro-1H-imidazole (136) (312 mg, 2.45 mmol) in MeOH (5 mL) was added Pd/C (0.42 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. The reaction mixture was stirred under H₂ (50 Psi) at 25°C for 3 hours. The reaction mixture was filtered. The filter cake was washed with MeOH (5 mL x 3) and then the combined filtrate was concentrated. 137 (238 mg, 99.8%) was obtained as a black brown oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.34-7.28 (m, 1H), 7.08-7.02 (br s, 1H), 3.56-3.51 (m, 2H), 3.46 (s, 3H). LCMS t_R = 0.142 min in 1.5 min chromatography, MS ESI calcd. for C₈H₁₅N₆⁺ [2M+H]⁺ 195.14, found 195.1.

5-chloro-7-((1-methyl-1H-imidazol-4-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (138). To a solution of 1methyl-1H-imidazol-4-amine (137) (210 mg, 2.16 mmol) in EtOH (5 mL) was added 5,7dichloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (74) (450 mg, 2.11 mmol) under N₂ atmosphere. The reaction mixture was stirred under at 25°C for 5 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0%~78%, EtOAc/PE) to give the product. 138 (520mg, 44.4%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO- d_6) δ 8.98-8.92 (m, 1H), 8.78 (s, 1H), 8.04 (s, 1H), 7.98-7.89 (m, 1H), 7.28-7.26 (m, 1H), 3.73 (s, 3H). LCMS $t_R = 0.714$ min in 2 min chromatography, MS ESI calcd. for $C_{11}H_9ClN_7^+$ [M+H]⁺274.06, found 273.9.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((1-methyl-1H-imidazol-4-yl)amino)pyrazolo[1,5-a]pyrimidine-3carbonitrile (**38**). To a solution of 5-chloro-7-((1-methyl-1H-imidazol-4-yl)amino)pyrazolo[1,5-a]pyrimidine-3carbonitrile (**138**) (300 mg, 1.10 mmol) and 3-(4H-1,2,4-triazol-4-yl)aniline (**86**) (176 mg, 1.10 mmol) in dioxane (5 mL) was added Cs₂CO₃ (1.07 g, 3.29 mmol), BINAP (102 mg, 0.16 mmol) and Pd(OAc)₂ (37.0 mg, 0.16 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 hour. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (MeOH) to give a crude product. The crude product was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water (FA)-ACN]; B%: 0%-38%, 25min). **38** (15.0 mg, 3.30%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.02 (s, 2H), 9.02 (s, 2H), 8.49 (s, 1H), 8.39 (t, *J* = 2.1 Hz, 1H), 7.65 – 7.58 (m, 2H), 7.50 (t, *J* = 8.1 Hz, 1H), 7.30 (dd, *J* = 7.9, 2.2 Hz, 1H), 7.07 (d, *J* = 1.5 Hz, 1H), 6.79 (s, 1H), 3.68 (s, 3H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 145.02, 144.91, 141.92, 141.24, 136.88, 135.15, 134.15, 130.28, 118.56, 118.54, 114.67, 114.55, 111.81, 110.65, 78.85, 77.22, 33.40. HPLC t_R = 2.779 min in 8 min chromatography, purity 95.5%. LCMS t_R = 1.446 min in 4 min chromatography, MS ESI calcd. for C₁₉H₁₆N₁₁⁺ [M+H]⁺ 398.16, found 398.4.

2-(4-Nitro-1H-imidazol-1-yl)ethanol (141). To a solution of 4-nitro-1H-imidazole (139) (3.00 g, 26.53 mmol) and 2-bromoethanol (3.98 g, 31.84 mmol) in MeCN (30 mL) was added K₂CO₃ (11.0 g, 79.59 mmol). Then the reaction mixture was stirred at 60°C for 10 hours. The reaction mixture was filtered. The filtrate was concentrated. 141 (4.55 g, crude) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO- d_6) δ 8.40-8.35 (m, 1H), 7.83-7.80 (m, 1H), 4.11 (t, *J* = 5.2 Hz, 2H), 3.70 (t, *J* = 5.2 Hz, 2H), 3.39 (s, 1H).

1-(2-Chloroethyl)-4-nitro-1H-imidazole (142). To a solution of 2-(4-Nitro-1H-imidazol-1-yl)ethanol (141) (1.50 g, 9.55 mmol) in DCM (20 mL) was added SOCl₂ (3.14 g, 28.64 mmol) and DMF (7 mg, 95.46 µmol). The mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated directly. 142 (1.56 g, 93.1%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO- d_6) δ 8.51-8.45 (m, 1H), 7.94-7.90 (m, 1H), 4.46 (t, *J* = 5.6 Hz, 2H), 4.08-4.04 (m, 2H).

N-methyl-2-(4-nitro-1H-imidazol-1-yl)ethanamine (**143**). To a solution of 1-(2-Chloroethyl)-4-nitro-1H-imidazole (**142**) (1.50 g, 8.54 mmol) and methylamine (1.33 g, 42.72 mmol) in MeCN (8 mL) was added K₂CO₃ (7.08 g, 51.26 mmol) at 25°C. Then NaI (1.54 g, 10.25 mmol) was added to the mixture at 0°C, and the reaction mixture was heated in a microwave reactor at 80°C for 4 hours. The mixture was concentrated in vacuo. **143** (1.49 g, 13.0%) was obtained as a black brown solid. LCMS $t_R = 0.107$ min in 1 min chromatography, MS ESI calcd. for C₆H₁₁N₄O₂⁺ [M+H]⁺ 171.09, found 171.0.

Tert-butyl methyl(2-(4-nitro-1H-imidazol-1-yl)ethyl)carbamate (144). To a solution of N-methyl-2-(4-nitro-1H-imidazol-1-yl)ethanamine (143) (1.48 g, 8.70 mmol) in THF (10 mL) was added K₂CO₃ (3.61 g, 26.09 mmol) in H₂O (5 mL) at 25°C. Then Boc₂O (3.80 g, 17.39 mmol) was added to the mixture and the resulting mixture was stirred at 25°C for 3 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0-80%, EtOAc / PE). 144 (586 mg, 21.8%) was obtained as a black solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.38-8.29 (m, 1H), 7.83-7.78 (m, 1H), 4.19 (t, *J* = 5.2 Hz, 2H), 3.57 (t, *J* = 5.2 Hz, 2H), 2.79 (s, 3H), 1.30-1.11 (m, 9H). LCMS t_R = 0.766 min in 2 min chromatography, MS ESI calcd. for C₁₁H₁₉N₄O₄⁺ [M+H]⁺ 271.14, found 271.0.

Tert-butyl (2-(4-*amino-1H-imidazol-1-yl)ethyl*)(*methyl*)*carbamate* (**147**). To a solution of tert-butyl methyl(2-(4-nitro-1H-imidazol-1-yl)ethyl)carbamate (**144**) (350 mg, 1.29 mmol) in THF (10 mL) was added Pd/C (0.35 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times and it was stirred under H₂ (50Psi) at 40°C for 12 hours. The reaction mixture was filtered. The filter cake was washed with MeOH (10 mL x 3) and then the combined filtrate was concentrated. **147** (283 mg, 61.9%) was obtained as a brown oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.04 (s, 1H), 6.23-5.98 (m, 1H), 3.17 (s, 2H), 2.69-2.62 (m, 7H), 1.33-1.31 (m, 9H). LCMS t_R = 0.825 min in 2 min chromatography, MS ESI calcd. for C₁₁H₂₁N₄O₂⁺ [M+H]⁺ 241.17, found 241.0.

Tert-butyl $(2-(4-((5-chloro-3-cyanopyrazolo[1,5-a]pyrimidin-7-yl)amino)-1H-imidazol-1-yl)ethyl)(methyl)carbamate (149). To a solution of 5,7-dichloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (74) (220 mg, 1.03 mmol) in EtOH (10 mL) was added tert-butyl (2-(4-amino-1H-imidazol-1-yl)ethyl)(methyl)carbamate (147) (273 mg, 1.14 mmol) under N₂ atmosphere. The reaction mixture was stirred under at 25°C for 5 hours. The reaction mixture was concentrated in vacuo. 149 (586 mg, 21.8%) was obtained as a black solid. ¹H NMR (400 MHz, DMSO-d₆) <math>\delta$ 11.30-10.75 (m, 1H), 8.94 (s, 1H),

8.81-8.73 (m, 1H), 8.04 (s, 1H), 7.85-7.64 (m, 1H), 4.20-4.09 (m, 2H), 2.85-2.69 (m, 5H), 1.39-1.30 (m, 9H). LCMS $t_R = 1.301$ min in 2 min chromatography, MS ESI calcd. for $C_{18}H_{22}ClN_8O_2^+$ [M+H]⁺ 417.15, found 417.1.

Tert-butyl (2-(4-((5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-3-cyanopyrazolo[1,5-a]pyrimidin-7-yl)amino)-1Himidazol-1-yl)ethyl)(methyl)carbamate (**151**). To a solution of tert-butyl (2-(4-((5-chloro-3-cyanopyrazolo[1,5-a]pyrimidin-7-yl)amino)-1H-imidazol-1-yl)ethyl)(methyl)carbamate (**149**) (300 mg, 0.72 mmol) and 3-(4H-1,2,4-triazol-4-yl)aniline (**86**) (115 mg, 0.72 mmol) in dioxane (5 mL) was added Cs₂CO₃ (703 mg, 2.16 mmol), BINAP (67 mg, 0.11 mmol) and Pd(OAc)₂ (24 mg, 0.11 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 4 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 4~5%, MeOH/DCM). **151** (397 mg, 69.1%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.07-10.02 (m, 1H), 9.03 (s, 2H), 8.49 (s, 1H), 8.41-8.36 (m, 1H), 7.65-7.61 (m, 1H), 7.60-7.55 (m, 1H), 7.50 (t, *J* = 8.0 Hz, 1H), 7.32-7.29 (m, 1H), 5.48 (s, 1H), 4.14-4.09 (m, 2H), 3.543.47 (m, 2H), 2.75-2.69 (m, 3H), 1.27 (s, 9H). LCMS t_R = 0.797 min in 1.5 min chromatography, MS ESI calcd. for C₂₆H₂₉N₁₂O₂⁺ [M+H]⁺ 541.25, found 541.2.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((1-(2-(methylamino)ethyl)-1H-imidazol-4-

yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**39**). A mixture of tert-butyl (2-(4-((5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-3-cyanopyrazolo[1,5-a]pyrimidin-7-yl)amino)-1H-imidazol-1-yl)ethyl)(methyl)carbamate (**151**) (170 mg, 0.31 mmol) in DCM (3 mL) was added TFA (3 mL) at 25°C. The reaction mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water (FA)-ACN]; B%: 0%-26%, 25min). **39** (40.0 mg, 27.8%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.10 (s, 1H), 9.02 (s, 2H), 8.49 (s, 1H), 8.40 (t, *J* = 2.2 Hz, 1H), 8.20 (s, 1H), 7.67 (s, 1H), 7.63 (dd, *J* = 7.6, 2.0 Hz, 1H), 7.50 (t, *J* = 8.1 Hz, 1H), 7.31 (dd, *J* = 8.0, 2.2 Hz, 1H), 7.15 (d, *J* = 1.5 Hz, 1H), 6.90 (s, 1H), 4.16 (t, *J* = 6.1 Hz, 2H), 3.05 (t, *J* = 6.2 Hz, 2H), 2.42 (s, 3H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 163.55, 156.71, 150.76, 144.99, 144.70, 141.92, 141.22, 137.03, 134.74, 134.13, 130.25, 118.56, 114.53, 111.79, 109.33, 79.00, 77.21, 50.03, 44.80, 34.36. HPLC t_R = 2.532 min in 8 min chromatography, purity 96.3%. LCMS t_R = 1.302 min in 4 min chromatography, MS ESI calcd. for C₂₁H₂₁N₁₂⁺ [M+H]⁺ 441.20, found 441.4.

N,N-dimethyl-2-(4-nitro-1H-imidazol-1-yl)ethanamine (**153**). To a solution of 2-chloro-N,N-dimethylethan-1-amine hydrochloride (1.27 g, 8.84 mmol) in MeCN (8 mL) was added K₂CO₃ (3.67 g, 26.53 mmol) and NaI (1.59 g, 10.61 mmol) at 25°C. Then 4-nitro-1H-imidazole (**139**) (1 g, 8.84 mmol) was added to the mixture and the reaction mixture was stirred at 60 °C for 6 h. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Xtimate C18 150*40mm*10um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 0%-35%, 10min). **153** (300 mg, 17.9% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.39 (s, 1H), 7.85 (s, 1H), 4.15 (t, *J* = 6.0 Hz, 2H), 2.60 (t, *J* = 6.0 Hz, 2H), 2.16 (s, 6H). LCMS t_R = 0.152 min in 1.5 min chromatography, MS ESI calcd. for C₇H₁₃N₄O₂+ [M+H]⁺ 185.10, found 185.1.

1-(2-(Dimethylamino)ethyl)-1H-imidazol-4-amine (159). To a solution of N,N-dimethyl-2-(4-nitro-1H-imidazol-1-yl)ethanamine (153) (250 mg, 1.36 mmol) in MeOH (2 mL) was added Pd/C (100 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 psi) at 35°C for 4 h. The reaction mixture was filtered via a celite pad. The reaction mixture was filtered. The filter cake was washed with MeOH (50 mL x3) and then the combined filtrate was concentrated to dryness. **159** (224 mg) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.12 (s, 1H), 6.15 (d, J = 1.6 Hz, 1H), 4.08 - 3.96 (m, 2H), 3.83 (t, J = 6.4 Hz, 2H), 2.14 - 2.12 (m, 6H)

5-*Chloro-7*-((*1*-(2-(*dimethylamino*)*ethyl*)-*1H-imidazol-4-yl*)*amino*)*pyrazolo*[*1*,5-*a*]*pyrimidine-3-carbonitrile* (**165**). To a solution of 5,7-dichloropyrazolo[1,5-*a*]*pyrimidine-3-carbonitrile* (**74**) (290 mg, 1.36 mmol) in EtOH (5 mL) was added 1-(2-(dimethylamino)ethyl)-1H-imidazol-4-amine (**159**) (210 mg, 1.36 mmol) at 25°C. The reaction mixture was stirred at 25°C for 2 h. The reaction mixture was filtered. The filter cake was the target coarse product and was washed with EtOH (4 mL x2). The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 0%-38%, 36min). **165** (80 mg, 17.3%) was obtained as a light yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.98 - 10.82 (m, 1H), 8.75 (s, 1H), 7.68 (s, 1H), 7.37 (s, 1H), 7.19 (d, *J* = 1.6 Hz, 1H), 4.08 (t, *J* = 6.4 Hz, 2H), 2.57 (t, *J* = 6.4 Hz, 2H), 2.18 (s, 6H). LCMS t_R = 0.560 min in 2.5 min chromatography, MS ESI calcd. for C₁₄H₁₆ClN₈⁺ [M+H]⁺ 331.12, found 331.1.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((1-(2-(dimethylamino)ethyl)-1H-imidazol-4-

yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**40**). To a solution of 5-chloro-7-((1-(2-(dimethylamino)ethyl)-1Himidazol-4-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**165**) (50 mg, 0.15 mmol) and 3-(4H-1,2,4-triazol-4yl)aniline (**86**) (29 mg, 0.18 mmol) in dioxane (8 mL) was added Cs₂CO₃ (147 mg, 0.45 mmol), BINAP (14 mg, 0.02 mmol) and Pd(OAc)₂ (7 mg, 0.03 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 2 h. The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water (HCl)-ACN]; B%: 0%-40%, 10min). The residue was purified by prep-HPLC(column: Phenomenex luna C18 150*25mm* 10um; mobile phase: [water(HCl)-ACN]; B%: 0%-25%, 30min). **40** as a HCl salt (8.6 mg, 11.8%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.40 (s, 1H), 10.26 (s, 1H), 10.23 (s, 1H), 9.11 (s, 2H), 8.52 (s, 1H), 8.42 (s, 1H), 8.00 (s, 1H), 7.76 – 7.62 (m, 1H), 7.52 (t, *J* = 8.1 Hz, 1H), 7.37 – 7.25 (m, 2H), 6.96 (s, 1H), 4.49 (t, *J* = 6.6 Hz, 2H), 3.59 (q, *J* = 5.6 Hz, 2H), 2.82 (d, *J* = 4.6 Hz, 6H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 157.33, 145.83, 142.22, 141.97, 139.90, 135.22, 134.21, 131.70, 130.79, 130.05, 124.39, 121.55, 119.40, 115.62, 112.48, 80.26, 78.10, 55.70, 51.42, 43.00. HPLC t_R = 2.637 min in 8 min chromatography, purity 94.5%. LCMS t_R = 1.375 min in 4 min chromatography, MS ESI calcd. for C₂₂H₂₃N₁₂+ [M+H]⁺ 455.22, found 455.4.

4-Nitro-1-(2-(pyrrolidin-1-yl)ethyl)-1H-imidazole (154). To a solution of 4-nitro-1H-imidazole (139) (1.00 g, 8.84 mmol) and 1-(2-chloroethyl)pyrrolidine (1.50 mg, 8.84 mmol) in DMF (15 mL) was added K₂CO₃ (3.67 g, 26.53 mmol) at 25°C. Then the mixture was stirred at 120°C for 12 h. The reaction mixture was concentrated. Water (50 mL) was added. The resulting mixture was extracted with EtOAc (50 mL x3). The combined organic phase was washed with brine (30 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. 154 (425 mg, 20.0%) was obtained as a black brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.42-8.30 (m, 1H), 7.89-7.81 (m, 1H), 4.47-4.12 (m, 2H), 2.82-2.72 (m, 2H), 2.49-2.44 (m, 4H), 1.70-1.60 (m, 4H). LCMS t_R = 0.437 min in 2.5 min chromatography, MS ESI calcd. for C₉H₁₅N₄O₂⁺ [M+H]⁺ 211.12, found 211.3.

1-(2-(Pyrrolidin-1-yl)ethyl)-1H-imidazol-4-amine (*160*). To a solution of 4-nitro-1-(2-(pyrrolidin-1-yl)ethyl)-1H-imidazole (*154*) (420 mg, 2.00 mmol) in MeOH (8 mL) was added Pd/C (0.33 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 psi) at 20°C for 2 h. The reaction mixture was filtered via a celite pad. The reaction mixture was filtered. The filter cake was washed with MeOH (10 mL x3) and then the combined filtrate was concentrated to dryness. *160* (412 mg, crude) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.35-7.08 (m, 1H), 6.22-5.98 (m, 1H), 4.44-3.80 (m, 4H), 3.17 (s, 1H), 2.67-2.63 (m, 1H), 2.48-2.27 (m, 4H), 1.74-1.56 (m, 4H). LCMS t_R = 0.612 min in 1 min chromatography, MS ESI calcd. for C₉H₁₇N₄⁺ [M+H]⁺ 181.14, found 181.1.

5-*Chloro-7*-((1-(2-(*pyrrolidin-1-yl*)*ethyl*)-1*H-imidazol-4-yl*)*amino*)*pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**166**). To a solution of 5,7-dichloropyrazolo[1,5-*a*]*pyrimidine-3-carbonitrile* (**74**) (400 mg, 1.11 mmol) in EtOH (12 mL) was added 1-(2-(*pyrrolidin-1-yl*)*ethyl*)-1H-imidazol-4-amine (**160**) (406 mg, 2.25 mmol) at 25°C. The reaction mixture was stirred at 25°C for 2 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~9%, MeOH/DCM). **166** (185 mg, 24.7%) was obtained as a black brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.78 (s, 1H), 7.80 (s, 1H), 7.45 (s, 1H), 7.27 (s, 1H), 4.41 (s, 2H), 3.64-3.46 (m, 4H), 3.17 (s, 1H), 3.08-2.94 (m, 2H), 2.02-1.81 (m, 4H). LCMS t_R = 1.039 min in 2.5 min chromatography, MS ESI calcd. for C₁₆H₁₈ClN₈⁺ [M+H]⁺ 357.13, found 357.2.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((1-(2-(pyrrolidin-1-yl)ethyl)-1H-imidazol-4-

yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**41**). To a solution of 5-chloro-7-((1-(2-(pyrrolidin-1-yl)ethyl)-1Himidazol-4-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**166**) (60 mg, 0.17 mmol) and 3-(4H-1,2,4-triazol-4yl)aniline (**86**) (28 mg, 0.17 mmol) in dioxane (3 mL) was added 'BuOLi (40 mg, 0.50 mmol), BINAP (16 mg, 0.03mmol) and Pd(OAc)₂ (6 mg, 0.03 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 2 h. The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water(NH₃H₂O+NH₄HCO₃)-ACN]; B%: 10%-50%, 35min). **41** (6.7 mg, 7.8%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.01 (s, 1H), 9.98 (s, 1H), 9.01 (s, 2H), 8.48 (s, 1H), 8.38 (t, *J* = 2.2 Hz, 1H), 7.67 (d, *J* = 1.5 Hz, 1H), 7.62 (dd, *J* = 7.9, 2.1 Hz, 1H), 7.50 (t, *J* = 8.1 Hz, 1H), 7.30 (dd, *J* = 7.8, 2.2 Hz, 1H), 7.14 (d, *J* = 1.5 Hz, 1H), 6.80 (s, 1H), 4.09 (t, *J* = 6.4 Hz, 2H), 2.76 (t, *J* = 6.5 Hz, 2H), 1.77 – 1.61 (m, 4H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.69, 150.77, 145.02, 144.88, 141.92, 141.22, 136.58, 134.70, 134.14, 130.27, 118.56, 114.66, 114.53, 111.81, 109.74, 78.84, 77.19, 55.85, 53.47, 45.66, 23.15. HPLC $t_R = 2.736$ min in 8 min chromatography, purity 94.0%. LCMS $t_R = 1.591$ min in 4 min chromatography, MS ESI calcd. for $C_{24}H_{25}N_{12}^+$ [M+H]⁺ 481.23, found 481.3.

4-(2-(4-Nitro-1H-imidazol-1-yl)ethyl)morpholine (155). To a solution of 4-nitro-1H-imidazole (139) (2 g, 17.69 mmol) and 4-(2-chloroethyl)morpholine (3.18 g, 21.22 mmol) in MeCN (20 mL) was added K₂CO₃ (7.33 g, 53.06 mmol) at 25°C. The mixture was stirred at 100°C for 5 h. The reaction mixture was concentrated. Water (50 mL) was added. The resulting mixture was extracted with DCM (50 mL x3). The combined organic phase was washed with brine (30 mL), water (30 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. The residue was purified by prep-HPLC (column: Phenomenex luna C18 250*50mm*10 um; mobile phase: [water (NH₃H₂O+NH₄HCO₃) -ACN]; B%: 0%-40%, 20min). **155** (1.2 g, 30.0%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.40 (d, *J* = 1.6 Hz, 1H), 7.85 (d, *J* = 1.2 Hz, 1H), 4.18 (t, *J* = 6.0 Hz, 2H), 3.57-3.49 (m, 4H), 2.66 (t, *J* = 6.0 Hz, 2H), 2.46-2.35 (m, 4H). LCMS t_R = 0.160 min in 1.5 min chromatography, MS ESI calcd. for C₉H₁₅N₄O₃+ [M+H]⁺ 227.11, found 227.0.

1-(2-Morpholinoethyl)-1H-imidazol-4-amine (161). To a solution of 4-(2-(4-nitro-1H-imidazol-1-yl)ethyl)morpholine (155) (1.00 g, 4.42 mmol) in MeOH (15 mL) was added Pd/C (0.84 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 psi) at 35°C for 4 h. The reaction mixture was filtered via a celite pad. The reaction mixture was filtered. The filter cake was washed with MeOH (10 mL x3) and then the combined filtrate was concentrated to dryness. **161** (870 mg, 99.9%) was obtained as a yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.17-7.10 (m, 1H), 6.20-6.15 (m, 1H), 4.33-3.91 (m, 2H), 3.87 (t, *J* = 6.4 Hz, 2H), 3.56-3.52 (m, 4H), 2.53 (t, *J* = 6.4 Hz, 2H), 2.40-2.34 (m, 4H). LCMS t_R = 0.141 min in 1.5 min chromatography, MS ESI calcd. for C₉H₁₇N₄O⁺ [M+H]⁺ 197.14, found 197.1.

Tert-butyl 5-*chloro-7-((1-(2-morpholinoethyl)-1H-imidazol-4-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile* (167). To a solution of 5,7-dichloropyrazolo[1,5-*a*]pyrimidine-3-carbonitrile (74) (785 mg, 3.69 mmol) in EtOH (20 mL) was added 1-(2-morpholinoethyl)-1H-imidazol-4-amine (161) (868 mg, 4.42 mmol) at 25°C. The reaction mixture was stirred at 25°C for 4 h. The reaction mixture was filtered. The filter cake was washed with EtOH (20 mL x2) and then the combined filtrate cake was concentrated to dryness. 167 (1.4 g, 97.1%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.22-10.87 (m, 1H), 8.77 (s, 1H), 7.80 (s, 1H), 7.42 (s, 1H), 7.31-7.25 (m, 1H), 4.48 (s, 2H), 3.84 (s, 4H), 3.47-3.42 (m, 2H), 3.21-2.93 (m, 4H). LCMS t_R = 0.673 min in 1.5 min chromatography, MS ESI calcd. for C₁₆H₁₈ClN₈O⁺ [M+H]⁺ 373.13, found 373.1.

Tert-butyl 5-((3-(4H-1,2,4-triazol-4-vl)phenvl)amino)-7-((1-(2-morpholinoethvl)-1H-imidazol-4yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (42). To a solution of tert-butyl 5-chloro-7-((1-(2-morpholinoethyl)-1H-imidazol-4-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (167) (500 mg, 1.34 mmol) and 3-(4H-1,2,4-triazol-4yl)aniline (86) (258 mg, 1.61 mmol) in dioxane (8 mL) was added Cs₂CO₃ (1.75 g, 5.36 mmol) and Brettphos Pd G3 (182 mg, 0.20 mmol) at 25°C. Then the mixture was degassed and purged with N2. The reaction mixture was heated in a microwave at 130°C for 4 h. The residue was purified by flash silica gel chromatography (eluent of 0%~78%, MeOH/DCM) to give the product. The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water (NH₄HCO₃)-ACN]; B%: 0%-90%, 14min). The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water (HCl)-ACN]; B%: 0%-34%, 30min). 42 as a HCl salt (20 mg, 3.0%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO- d_6) δ 11.65 (br s, 1H), 10.65 (d, J = 7.2 Hz, 1H), 10.37 (s, 1H), 9.40 - 9.17 Hz, 1H), 7.35 (dd, J = 7.9, 2.2 Hz, 1H), 6.94 (s, 1H), 4.67 (t, J = 6.5 Hz, 2H), 4.11 – 3.73 (m, 6H), 3.53 (br s, 2H), 3.20 (br s, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.89, 150.93, 145.53, 141.80, 141.73, 138.02, 134.75, 133.57, 130.42, 119.22, 119.18, 115.43, 114.53, 112.23, 112.18, 79.98, 77.81, 63.05, 54.62, 51.38, 42.49. HPLC $t_R = 2.721$ min in 8 min chromatography, purity 99.7%. LCMS $t_R = 1.302$ min in 4 min chromatography, MS ESI calcd. for $C_{24}H_{25}N_{12}O^+$ [M+H]⁺ 497.23, found 497.4.

Tert-butyl (2-(3-nitro-1H-pyrazol-1-yl)ethyl)carbamate (145). To a solution of 3-nitro-1H-pyrazole (140) (5 g, 44.22 mmol) and tert-butyl (2-bromoethyl)carbamate (11.89 g, 53.06 mmol) in DMF (50 mL) was added K₂CO₃ (18.33 g, 132.66 mmol) at 25°C. The mixture was stirred at 90°C for 3 h. The reaction mixture was poured into ice water (100 mL) slowly. The resulting suspension was filtered. The filter cake was then dried. 145 (7.77 g, 53.0%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.02-7.92 (m, 1H), 7.13-6.90 (m, 2H), 4.25 (t, *J* = 6.0 Hz, 2H), 3.41-3.35 (m, 2H), 1.32 (s, 9H). LCMS t_R = 0.465 min in 1 min chromatography, MS ESI calcd. for C₅H₉N₄O₂⁺ [M+2H-Boc]⁺ 157.07, found 157.0.

Tert-butyl methyl(2-(3-nitro-1*H*-pyrazol-1-yl)*ethyl*)*carbamate* (**146**). To a solution of tert-butyl (2-(3-nitro-1H-pyrazol-1-yl)*ethyl*)*carbamate* (**145**) (2 g, 7.80 mmol) in THF (15 mL) was added NaH (468 mg, 60% in mineral oil) at 0°C for 0.5 h. Then CH₃I (1.11 g, 7.80 mmol) was added to the mixture. The mixture was stirred at 25°C for 10 h. The reaction was quenched with aq.NH₄Cl (10 mL) drop-wise. Water (50 mL) was added. The resulting mixture was extracted with EtOAc (50 mL x3). The combined organic phase was washed with brine (50 mL x2), dried over anhydrous Na₂SO₄, filtered and concentrated. **146** (2.13 g, 98.3%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.99 (d, *J* = 2.4 Hz, 1H), 7.11-6.98 (m, 1H), 4.35 (t, *J* = 5.6 Hz, 2H), 3.65-3.53 (m, 2H), 2.78-2.68 (m, 3H), 1.32-1.16 (m, 9H). LCMS t_R = 1.227 min in 2 min chromatography, MS ESI calcd. for C₆H₁₁N₄O₂⁺ [M+2H-Boc]⁺ 171.09, found 171.3.

Tert-butyl (2-(3-*amino-1H-pyrazol-1-yl)ethyl*)(*methyl*)*carbamate* (**148**). To a solution of tert-butyl methyl(2-(3-nitro-1H-pyrazol-1-yl)ethyl)carbamate (**146**) (2 g, 7.40 mmol) in MeOH (20 mL) was added Pd/C (2.38 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 psi) at 35°C for 4 h. The reaction mixture was filtered via a celite pad. The reaction mixture was filtered. The filter cake was washed with MeOH (20 mL x3) and then the combined filtrate was concentrated to dryness. **148** (1.56 g, 86.0%) was obtained as a yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.20 (s, 1H), 5.37 (d, *J* = 2.4 Hz, 1H), 4.52 (s, 2H), 3.90 (t, *J* = 6.0 Hz, 3H), 3.41 (t, J = 6.0 Hz, 2H), 2.59 (s, 3H), 1.42-1.30 (m, 9H). LCMS t_R = 0.643 min in 1.5 min chromatography, MS ESI calcd. for C₁₁H₂₁N₄O₂⁺ [M+H]⁺ 241.17, found 241.1.

Tert-butyl (2-(3-((5-chloro-3-cyanopyrazolo[1,5-a]pyrimidin-7-yl)amino)-1H-pyrazol-1-yl)ethyl)(methyl)carbamate (**150**). To a solution of 5,7-dichloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (**74**) (1.1 g, 5.16 mmol) in EtOH (20 mL) was added tert-butyl (2-(3-amino-1H-pyrazol-1-yl)ethyl)(methyl)carbamate (**148**) (1.49 g, 6.20 mmol) at 25°C. The reaction mixture was stirred at 25°C for 4 h. The reaction mixture was concentrated. The residue was purified by flash silica gel chromatography (eluent of 0%~32%, EtOAC/PE) to give the product. **150** (1.18 g, 45.5%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.33-11.12 (m, 1H), 8.79 (s, 1H), 7.77-7.57 (m, 2H), 6.38-6.24 (m, 1H), 4.25 (t, *J* = 5.6 Hz, 2H), 3.56 (t, *J* = 5.6 Hz, 2H), 2.72 (s, 3H), 1.33-1.24 (m, 9H) LCMS t_R = 1.665 min in 2.5 min chromatography, MS ESI calcd. for C₁₈H₂₂CIN₈O₂⁺ [M+H]⁺ 417.15, found 417.1.

Tert-butyl (2-(3-((5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-3-cyanopyrazolo[1,5-a]pyrimidin-7-yl)amino)-1Hpyrazol-1-yl)ethyl)(methyl)carbamate (**152**). To a solution of tert-butyl (2-(3-((5-chloro-3-cyanopyrazolo[1,5-a]pyrimidin-7-yl)amino)-1H-pyrazol-1-yl)ethyl)(methyl)carbamate (**150**) (500 mg, 1.20 mmol) and 3-(4H-1,2,4-triazol-4-yl)aniline (**86**) (231 mg, 1.44 mmol) in dioxane (8 mL) was added Cs₂CO₃ (1.56 g, 4.80 mmol), BINAP (112 mg, 0.18 mmol) and Pd(OAc)₂ (40 mg, 0.18 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 4 h. The reaction mixture was concentrated to dryness. The residue was purified by flash silica gel chromatography (eluent of 0%~4%, MeOH/DCM) to give the product. **152** (501 mg, 57.8%) was obtained as a gray solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.40-10.29 (m, 1H), 10.11-9.94 (m, 1H), 9.07-8.98 (m, 2H), 8.51 (s, 1H), 8.40-8.31 (m, 1H), 7.68-7.63 (m, 2H), 7.54 (t, *J* = 8.4 Hz 1H), 7.42-7.29 (m, 2H), 7.27-7.08 (m, 2H), 4.23-4.15 (m, 2H), 3.62-3.52 (m, 2H), 2.66-2.58 (m, 3H), 1.37-1.28 (m, 9H). LCMS t_R = 1.572 min in 2.5 min chromatography, MS ESI calcd. for C₂₆H₂₉N₁₂O₂+ [M+H]+ 541.25, found 541.2.

Tert-butyl $5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((1-(2-(methylamino)ethyl)-1H-pyrazol-3-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile(43).To a solution of tert-butyl<math>(2-(3-((5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-3-cyanopyrazolo[1,5-a]pyrimidin-7-yl)amino)-1H-pyrazol-1-yl)ethyl)(methyl)carbamate(152)(250 mg, 1.20 mg, 1.20 mmol) in DCM (3 mL) was added TFA (5.98 g 52.44 mmol) at 25°C. The mixture was stirred at 35°C for 6 h. The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water (HCl) -ACN];B%: 0%-90%, 36min).43 as a HCl a salt (62 mg, 29.3%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-d_6) <math display="inline">\delta$ 10.83 (d, J = 4.8 Hz, 1H), 10.44 (s, 1H), 9.47 - 9.34 (m, 2H), 9.16 (s, 2H), 8.62 (q, J = 2.0 Hz, 1H), 8.52 (s, 1H), 7.90 (dd, J = 8.3, 2.2 Hz, 1H), 7.78 (d, J = 2.3 Hz, 1H), 7.62 (d, J = 1.8 Hz, 1H), 7.56 (t, J = 8.1 Hz, 1H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 6.31 (d, J = 2.3 Hz, 1H), 4.46 (t, J = 5.9 Hz, 2H), 3.51 (p, J = 6.0 Hz, 2H), 2.58 (t, J = 5.4 Hz, 3H). ¹³C NMR (101 MHz, DMSO-d_6) δ 157.11, 150.67, 147.75, 144.90, 142.87, 142.07, 141.52, 133.60, 131.73, 130.21, 119.14, 115.00, 114.60, 97.57, 81.58, 77.53, 48.00, 47.60, 33.10. HPLC t_R = 2.964 min in 8 min chromatography, purity 96.3%. LCMS t_R = 1.422 min in 4 min chromatography, MS ESI calcd. for C₂₁H₂₁N₁₂+ [M+H]+ 441.20, found 441.4.

N,N-dimethyl-2-(3-nitro-1H-pyrazol-1-yl)ethan-1-amine (156). To a solution of 3-nitro-1H-pyrazole (140) (1 g, 8.84 mmol) in DMF (10 mL) was added NaH (530.57 mg, 60% in mineral oil) in 0°C for 0.5 h, then 2-chloro-N,N-dimethylethan-

1-amine hydrochloride (1.27 g, 8.84 mmol) was added and the mixture was stirred at 15°C for 3.5 h. The reaction was quenched with aq. NH₄Cl (50 mL) drop-wise. The resulting mixture was extracted with EtOAc (20 mL x3). The combined organic phase was washed with brine (50 mL), water (50 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. **156** (1.72 g, 92.0%) was obtained as a yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.03 (s, 1H), 7.03 (s, 1H), 4.33 (t, *J* = 6.4 Hz, 2H), 2.73 (s, 2H), 2.18 (s, 6H). LCMS t_R = 0.857 min in 2 min chromatography, MS ESI calcd. for C₇H₁₃N₄O₂⁺ [M+H]⁺ 185.10, found 185.1.

1-(2-(Dimethylamino)ethyl)-1H-pyrazol-3-amine (162). To a solution of N,N-dimethyl-2-(3-nitro-1H-pyrazol-1-yl)ethan-1-amine (156) (1.6 g, 8.69 mmol) in MeOH (10 mL) was added Pd/C (1.02 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 psi) at 35°C for 2 h. The reaction mixture was filtered via a celite pad. The reaction mixture was filtered. The filter cake was washed with MeOH (20 mL x3) and then the combined filtrate was concentrated to dryness. 162 (1.34 g, 100%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.24-7.23 (m, 2H), 5.37-5.36 (m, 2H), 2.88 (s, 1H), 2.73 (s, 1H), 2.59 (t, *J* = 6.8 Hz, 2H), 2.17 (s, 6H). LCMS t_R = 0.118 min in 1 min chromatography, MS ESI calcd. for C₇H₁₅N₄⁺ [M+H]⁺ 155.13, found 155.0.

5-*Chloro-7*-((*1*-(2-(*dimethylamino*)*ethyl*)-*1H-pyrazol-3-yl*)*amino*)*pyrazolo*[*1*,5-*a*]*pyrimidine-3-carbonitrile* (**168**). To a solution of 5,7-dichloropyrazolo[*1*,5-*a*]*pyrimidine-3-carbonitrile* (**74**) (1.05 g, 4.93 mmol) in EtOH (10 mL) was added 1-(2-(dimethylamino)ethyl)-1H-pyrazol-3-amine (**162**) (912 mg, 5.91 mmol) at 25°C. The reaction mixture was stirred at 25°C for 2 h. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Welch Ultimate XB-CN 250*50*10um; mobile phase: [Heptane-EtOH (0.1%NH3H2O)]; B%: 10%-45%, 15min). The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water(NH₃H₂O+NH₄HCO₃)-ACN]; B%: 8%-48%, 35min). **168** (266 mg, 11.0%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.18 (s, 1H), 8.79 (s, 1H), 7.77-7.75 (m, 1H), 7.54 (s, 1H), 6.27 (d, *J* = 2.4 Hz, 1H), 4.22 (t, *J* = 6.4 Hz, 2H), 2.68 (t, *J* = 6.4 Hz, 2H), 2.19 (s, 6H). LCMS t_R = 1.447 min in 2.5 min chromatography, MS ESI calcd. for C₁₄H₁₆ClN₈⁺ [M+H]⁺ 331.11, found 331.1.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((1-(2-(dimethylamino)ethyl)-1H-pyrazol-3-

yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**44**). To a solution of 5-Chloro-7-((1-(2-(dimethylamino)ethyl)-1Hpyrazol-3-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**168**) (200 mg, 0.60 mmol) and 3-(4H-1,2,4-triazol-4yl)aniline (**86**) (145 mg, 0.91 mmol) in dioxane (8 mL) was added Cs₂CO₃ (591 mg, 1.81 mmol), BINAP (56 mg, 0.09 mmol) and Pd(OAc)₂ (20 mg, 0.09 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 2 h. The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water(HCl)-ACN]; B%: 7%-37%, 10min). **44** as a HCl salt (63.5 mg, 22.7%) was obtained as a yellow solid. ¹H NMR (400 MHz, D₂O) δ 7.94 (s, 2H), 7.78 (s, 1H), 7.31 (s, 1H), 7.16 (s, 1H), 6.50 (s, 1H), 6.30 (s, 1H), 5.99 (s, 1H), 5.94 (s, 1H), 5.51 (s, 1H), 4.19 (s, 2H), 3.53 (s, 2H), 2.82 (s, 6H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 157.15, 157.11, 150.80, 147.70, 145.13, 143.00, 142.18, 141.99, 133.12, 132.21, 130.44, 120.04, 115.57, 114.83, 112.82, 97.85, 77.69, 55.27, 46.49, 42.83. HPLC t_R = 2.981 min in 8 min chromatography, purity 98.3%. LCMS t_R = 1.528 min in 4 min chromatography, MS ESI calcd. for C₂₂H₂₃N₁₂⁺ [M+H]⁺ 455.22, found 455.4.

3-Nitro-1-(2-(pyrrolidin-1-yl)ethyl)-1H-pyrazole (157). To a solution of 3-nitro-1H-pyrazole (140) (500 mg, 4.42 mmol) and 1-(2-chloroethyl)pyrrolidine (1.13 g, 6.63 mmol) in MeCN (10 mL) was added K₂CO₃ (1.83 g, 13.27 mmol) at 25°C. The mixture was stirred at 120°C for 4 h. The reaction mixture was concentrated. Water (50 mL) was added. The resulting mixture was extracted with DCM (50 mL x3). The combined organic phase was washed with brine (30 mL), water (30 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. **157** (234 mg, 14.2%) was obtained as a black brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.05 (d, *J* = 2.0 Hz, 1H), 7.03 (d, *J* = 2.0 Hz, 1H), 4.33 (t, *J* = 6.4 Hz, 2H), 2.85 (t, *J* = 6.4 Hz, 2H), 2.48-2.45 (m, 4H), 1.66-1.63 (m, 4H). LCMS t_R = 1.326 min in 2.5 min chromatography, MS ESI calcd. for C₉H₁₅N₄O₂⁺ [M+H]⁺ 211.11, found 211.1.

1-(2-(Pyrrolidin-1-yl)ethyl)-1H-pyrazol-3-amine (163). To a solution of 3-nitro-1-(2-(pyrrolidin-1-yl)ethyl)-1H-pyrazole (157) (200 mg, 0.95 mmol) in MeOH (5 mL) was added Pd/C (0.4 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 psi) at 25°C for 5 hours. The reaction mixture was filtered via a celite pad. The reaction mixture was filtered. The filter cake was washed with MeOH (10 mL x3) and then the combined filtrate was concentrated to dryness. 163 (99 mg, 49.3%) was obtained as an off-white

oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.29 (d, *J* = 2.0 Hz, 1H), 5.33 (d, *J* = 2.0 Hz, 1H), 4.48 (s, 2H), 3.90 (t, *J* = 6.8 Hz, 2H), 2.70 (t, *J* = 6.8 Hz, 2H), 2.44-2.39 (m, 4H), 1.66-1.62 (m, 4H). LCMS t_R = 0.131 min in 1 min chromatography, MS ESI calcd. for C₉H₁₇N₄⁺ [M+H]⁺ 181.14, found 181.1.

Tert-butyl 5-*chloro-7-((1-(2-(pyrrolidin-1-yl)ethyl)-1H-pyrazol-3-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (169).* To a solution of 5,7-dichloropyrazolo[1,5-*a*]pyrimidine-3-carbonitrile (74) (105 mg, 0.49 mmol) in EtOH (5 mL) was added 1-(2-(pyrrolidin-1-yl)ethyl)-1H-pyrazol-3-amine (163) (98 mg, 0.54 mmol) at 25°C. The reaction mixture was stirred at 25°C for 5 h. The reaction mixture was filtered. The filter cake was washed with EtOH (10 mL x3) and then the combined filtrate cake was concentrated. 169178 mg, 93.4%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.39-10.84 (m, 2H), 8.82 (s, 1H), 7.89 (d, *J* = 2.4 Hz, 1H), 7.60 (s, 1H), 6.39 (d, *J* = 2.4 Hz, 1H), 4.59 (t, *J* = 6.4 Hz, 2H), 3.63 (t, J = 6.0 Hz, 2H), 3.44 (q, J = 7.0 Hz, 2H), 1.91 (s, 4H), 1.05 (t, J = 7.2 Hz, 2H). LCMS t_R = 0.844 min in 2 min chromatography, MS ESI calcd. for C₁₆H₁₈ClN₈⁺ [M+H]⁺ 357.13, found 357.2.

Tert-butyl 5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((1-(2-(pyrrolidin-1-yl)ethyl)-1H-pyrazol-3yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (45). To a solution of tert-butyl 5-chloro-7-((1-(2-(pyrrolidin-1yl)ethyl)-1H-pyrazol-3-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (169) (95 mg, 0.27 mmol) and 3-(4H-1,2,4triazol-4-yl)aniline (86) (51 mg, 0.32 mmol) in dioxane (5 mL) was added Cs₂CO₃ (347 mg, 1.06 mmol), BINAP (25 mg, 0.04 mmol) and Pd(OAc)₂ (9 mg, 0.04 mmol) at 25°C. Then the mixture was degassed and purged with N_2 . The reaction mixture was heated in a microwave at 130°C for 4 h. The residue was purified by flash silica gel chromatography (MeOH) to give the product. The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water (HCl) -ACN]; B%: 0%-90%, 36min). 45 as a HCl salt (28.3 mg, 21.9%) was obtained as a vellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 11.12 – 10.86 (m, 2H), 10.42 (s, 1H), 9.50 (s, 2H), 8.65 (t, *J* = 2.2 Hz, 1H), 8.51 (s, 1H), 7.98 (dd, J = 8.2, 2.1 Hz, 1H), 7.81 (d, J = 2.4 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 6.30 (d, J = 2.3 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 6.30 (d, J = 2.3 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 6.30 (d, J = 2.3 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 6.30 (d, J = 2.3 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 6.30 (d, J = 2.3 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 6.30 (d, J = 2.3 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 6.30 (d, J = 2.3 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.47 (m, 2H), 7.36 (dd, J = 8.0, 2.2 Hz, 1H), 7.61 – 7.48 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.48 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.48 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.48 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.48 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.48 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 – 7.48 (dd, J = 8.0, 2.2 Hz, 1H), 7.68 (dd, J = 8.0, 2.2 Hz, 1H), 4.55 (t, J = 6.3 Hz, 2H), 3.91 (q, J = 6.0 Hz, 2H), 3.44 (td, J = 10.4, 5.1 Hz, 2H), 3.14 - 3.00 (m, 2H), 2.04 - 1.91 (m, 2H), 1.91 – 1.74 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 157.16, 150.67, 147.65, 144.89, 142.92, 142.23, 141.74, 133.13, 131.97, 130.15, 119.73, 115.25, 114.63, 112.59, 97.72, 81.37, 77.52, 53.42, 52.83, 47.55, 22.69. HPLC $t_R = 3.109 \text{ min in } 8$ min chromatography, purity 98.7%. LCMS $t_R = 1.504$ min in 4 min chromatography, MS ESI calcd. for $C_{24}H_{25}N_{12}^+$ [M+H]⁺ 481.23, found 481.5.

4-(2-(3-nitro-1H-pyrazol-1-yl)ethyl)morpholine (158). To a solution of 3-nitro-1H-pyrazole (140) (1.5 g, 13.27 mmol) and 4-(2-chloroethyl)morpholine (2.98 g, 19.90 mmol) in MeCN (20 mL) was added K₂CO₃ (1.83 g, 13.27 mmol) at 25°C. The mixture was stirred at 100°C for 4 h. The reaction mixture was concentrated. Water (50 mL) was added. The resulting mixture was extracted with DCM (50 mL x3). The combined organic phase was washed with brine (20 mL), water (20 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. The residue was purified by prep-HPLC (column: Phenomenex luna C18 250*50mm*10 um; mobile phase: [water(NH₃H₂O+NH₄HCO₃)-ACN]; B%: 0%-30%, 15min). **158** (912 mg, 29.8%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.04 (d, *J* = 2.4 Hz, 1H), 7.03 (d, *J* = 2.4 Hz, 1H), 4.35 (t, *J* = 6.4 Hz, 2H), 3.55-3.50 (m, 4H), 2.74 (t, *J* = 6.4 Hz, 2H), 2.44-2.38 (m, 4H). LCMS t_R = 1.080 min in 2.5 min chromatography, MS ESI calcd. for C₉H₁₅N₄O₃+ [M+H]⁺ 227.11, found 227.2.

1-(2-morpholinoethyl)-1H-pyrazol-3-amine (*164*). To a solution of 4-(2-(3-nitro-1H-pyrazol-1-yl)ethyl)morpholine (**158**) (730 mg, 3.23 mmol) in MeOH (15 mL) was added Pd/C (0.84 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 psi) at 35°C for 4 h. The reaction mixture was filtered via a celite pad. The reaction mixture was filtered. The filter cake was washed with MeOH (10 mL x3) and then the combined filtrate was concentrated to dryness. **164** (815 mg, 99.0%) was obtained as an off-white oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.30 (s, 1H), 5.34 (s, 1H), 4.50 (s, 2H), 3.92 (t, *J* = 6.8 Hz, 2H), 3.53 (t, *J* = 4.4 Hz, 4H), 2.59 (t, *J* = 6.8 Hz, 2H), 2.41-2.31 (m, 4H). LCMS t_R = 0.733 min in 2.5 min chromatography, MS ESI calcd. for C₉H₁₇N₄O⁺ [M+H]⁺ 197.14, found 197.3.

5-chloro-7-((1-(2-morpholinoethyl)-1H-pyrazol-3-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (170). To a solution of 5,7-dichloropyrazolo[1,5-a]pyrimidine-3-carbonitrile (74) (730 mg, 3.43 mmol) in EtOH (20 mL) was added 1-(2-morpholinoethyl)-1H-pyrazol-3-amine (164) (807 mg, 4.11 mmol) at 25°C. The reaction mixture was stirred at 25°C for 5 h. The reaction mixture was concentrated. The resulting mixture was triturated by H₂O (20 mL). The mixture was filtered. The filter cake was washed with EtOH (30 mL x2) and then dried. 170 (1.34 g, 100.0%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO- d_6) δ 11.27 (s, 2H), 8.81 (s, 1H), 7.85 (s, 1H), 7.57 (s, 1H), 6.36 (s, 1H), 4.94-3.42 (m, 10H), 3.21-

2.95 (m, 2H). LCMS $t_R = 1.452$ min in 2.5 min chromatography, MS ESI calcd. for $C_{16}H_{18}ClN_8O^+$ [M+H]⁺ 373.13, found 373.2.

5-((3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-((1-(2-morpholinoethyl)-1H-pyrazol-3-

yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**46**). To a solution of 5-chloro-7-((1-(2-morpholinoethyl)-1H-pyrazol-3-yl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**170**) (300 mg, 0.80 mmol) and 3-(4H-1,2,4-triazol-4-yl)aniline (**86**) (155 mg, 0.97 mmol) in dioxane (8 mL) was added Cs₂CO₃ (1.05 g, 3.22 mmol), BINAP (75 mg, 0.12 mmol) and Pd(OAc)₂ (27 mg, 0.12 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 6 h. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 48%-68%, 36min). The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water (HCl)-ACN]; B%: 0%-50%, 10min). **46** as a HCl salt (26.3 mg, 3.2%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.91 (s, 1H), 10.74 (s, 1H), 10.43 (s, 1H), 9.09 (s, 2H), 8.54 (d, *J* = 18.1 Hz, 2H), 7.91 – 7.79 (m, 2H), 7.54 (t, *J* = 8.2 Hz, 1H), 7.46 (s, 1H), 7.33 (d, *J* = 8.0 Hz, 1H), 6.32 (s, 1H), 4.60 (t, *J* = 6.6 Hz, 2H), 4.02 – 3.90 (m, 2H), 3.87 – 3.74 (m, 4H), 3.44 (d, *J* = 16.9 Hz, 2H), 3.22 (s, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 157.08, 150.73, 147.75, 144.94, 143.02, 142.00, 141.32, 134.03, 131.93, 130.23, 118.79, 114.79, 114.59, 112.00, 97.94, 81.21, 77.52, 63.19, 54.86, 51.59, 45.77. HPLC t_R = 3.013 min in 8 min chromatography, purity 96.4%. LCMS t_R = 1.477 min in 4 min chromatography, MS ESI calcd. for C₂₄H₂₅N₁₂O⁺ [M+H]⁺ 497.23, found 497.5.

Experimental Procedures in Scheme 5.

4-(4-Fluoro-3-nitrophenyl)-4H-1,2,4-triazole (175). To a solution of 4-fluoro-3-nitroaniline (171) (1.00 g, 6.41 mmol) and 1,2-diformylhydrazine (1.69 g, 19.22 mmol) in pyridine (10 mL) was added Et₃N (4.54 g, 44.84 mmol) and Me₃SiCl (10.44 g, 96.05 mmol) at 25°C. Then the mixture was stirred at 100°C for 12 hours. The reaction mixture was concentrated in vacuo. Brine (50 mL) was added. The resulting mixture was extracted with DCM (50mL x3). The combined organic phase was dried over anhydrous Na₂SO₄, filtered and concentrated to dryness. The residue was purified by flash silica gel chromatography (eluent of 0~3%, MeOH/DCM). **175** (667 mg, 49.7%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.22 (s, 2H), 8.61-8.56 (m, 1H), 8.22 - 8.16 (m, 1H), 7.89-7.82 (m, 1H). LCMS t_R = 0.364 min in 1 min chromatography, MS ESI calcd. for C₈H₆FN₄O₂⁺ [M+H]⁺ 209.05, found 208.9.

2-*Fluoro-5-(4H-1,2,4-triazol-4-yl)aniline (179).* To a solution of 4-(4-fluoro-3-nitrophenyl)-4H-1,2,4-triazole (175) (300 mg, 3.28 mmol) in MeOH (10 mL) was added Pd/C (0.60 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (15 mL x3). The combined filtrate was concentrated in vacuo. **179** (252 mg, 80.0%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.86 (s, 2H), 7.13-7.07 (m, 1H), 6.87 (dd, J = 2.8, 7.6 Hz, 1H), 6.70-6.65 (m, 1H), 5.50 (s, 2H). LCMS t_R = 0.644 min in 1 min chromatography, MS ESI calcd. for C₈H₈FN₄⁺ [M+H]⁺ 197.07, found 197.0.

7-(*Cyclopropylamino*)-5-((2-fluoro-5-(4H-1,2,4-triazol-4-yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**47**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (250 mg, 1.40 mmol) and 2-fluoro-5-(4H-1,2,4-triazol-4-yl)aniline (**179**) (300 mg, 1.28 mmol) in dioxane (3 mL) was added Brettphos Pd G3 (175 mg, 0.19 mmol) and 'BuOLi (308 mg, 3.85 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of MeOH). The crude product was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 10%-50%, 28min). The residue was purified by SFC (column: DAICEL CHIRALPAK IC(250mm*30mm, 10um); mobile phase: [0.1%NH₃H₂O-ETOH]; B%: 60%-60%,min). **47** (35.6 mg, 55.6%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.68 (s, 1H), 8.99 (s, 2H), 8.80 (d, *J* = 6.9 Hz, 1H), 8.40 (s, 2H), 7.51 (t, *J* = 9.8 Hz, 1H), 7.41 (dt, *J* = 8.4, 3.5 Hz, 1H), 6.34 (s, 1H), 2.67 – 2.57 (m, 1H), 0.87 – 0.78 (m, 2H), 0.78 – 0.67 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.66, 152.33 (d, *J* = 245.5 Hz), 150.60, 148.71, 145.03, 141.33, 129.99 (d, *J* = 3.4 Hz), 128.95 (d, *J* = 12.5 Hz), 116.50 (d, *J* = 21.8 Hz), 116.07, 115.99, 114.64, 77.11, 76.98, 23.42, 6.58. HPLC t_R = 3.582 min in 8 min chromatography, purity 98.5%. LCMS t_R = 1.978 min in 4 min chromatography, MS ESI calcd. for C₁₈H₁₅FN₉+ [M+H]⁺ 376.14, found 376.3.

4-(2-*Fluoro-5-nitrophenyl*)-4*H*-1,2,4-*triazole* (**176**). To a solution of 2-fluoro-5-nitroaniline (**172**) (5.00 g, 32.03 mmol) and 1,2-diformylhydrazine (8.46 g, 96.08 mmol) in pyridine (15 mL) was added Et₃N (22.69 g, 224.19 mmol, 31 mL) and Me₃SiCl (52.19 g, 480.42 mmol, 61 mL) at 25°C. The mixture was stirred at 100°C for 12 h under N₂ atmosphere. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0%~1% MeOH/DCM). **176** (4.92g, 73.8%) was obtained as a black brown oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.72 (dd, *J* = 2.8, 6.4 Hz, 1H), 8.61 (br d, *J* = 4.4 Hz, 1H), 8.46-8.40 (m, 1H), 7.87-7.82 (m, 1H), 7.48-7.43 (m, 1H). LCMS t_R = 0.358 min in 4 min chromatography, MS ESI calcd. for C₈H₆FN₄O₂⁺ [M+H]⁺ 209.05, found 209.0.

4-Fluoro-3-(4H-1,2,4-triazol-4-yl)aniline (180). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (176) (1 g, 4.80 mmol) in MeOH (3 mL) was added Pd/C (220 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 hours. Then the reaction mixture was stirred under H₂ (15 psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (400 mL). The combined filtrate was concentrated in vacuo. 180 (157 mg, 17.1%) was obtained as a light yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.86 (d, *J* = 1.6 Hz, 2H), 7.15 (dd, *J* = 8.8, 10.4 Hz, 1H), 6.75 - 6.60 (m, 2H), 6.14 - 5.87 (m, 2H). LCMS t_R = 0.229 min in 1 min chromatography, MS ESI calcd. for C₈H₈FN₄⁺ [M+H]⁺ 179.07, found 179.1.

7-(*Cyclopropylamino*)-5-((4-fluoro-3-(4H-1,2,4-triazol-4-yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**48**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (150 mg, 0.64 mmol) and 4-fluoro-3-(4H-1,2,4-triazol-4-yl)aniline (**180**) (158 mg, 0.64 mmol) in dioxane (2 mL) was added Cs₂CO₃ (628 mg, 1.93 mmol), BINAP (60 mg, 0.10 mmol) and Pd(OAc)₂ (22 mg, 0.10 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~4%, MeOH/DCM). The crude product was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 23%-53%, 10min). **48** (14.3 mg, 10.0%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.97 (s, 1H), 8.93 (d, *J* = 1.5 Hz, 2H), 8.39 (s, 1H), 8.37 (s, 1H), 8.26 (dd, *J* = 6.9, 2.7 Hz, 1H), 7.73 (ddd, *J* = 9.1, 4.2, 2.7 Hz, 1H), 7.53 (dd, *J* = 10.3, 9.1 Hz, 1H), 5.98 (s, 1H), 2.62 (tt, *J* = 6.9, 3.6 Hz, 1H), 0.86 – 0.80 (m, 2H), 0.75 – 0.70 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.44, 150.69, 149.33 (d, *J* = 244.2 Hz), 148.53, 145.05, 142.64 (d, *J* = 2.4 Hz), 137.59 (d, *J* = 2.7 Hz), 121.53 (d, *J* = 12.9 Hz), 120.56 (d, *J* = 7.2 Hz), 117.18 (d, *J* = 20.8 Hz), 116.14, 114.57, 76.91, 76.48, 23.33, 6.53. HPLC t_R = 3.904 min in 8 min chromatography, purity 96.6%. LCMS t_R = 2.095 min in 4 min chromatography, MS ESI calcd. for C₁₈H₁₅FN₉+ [M+H]+ 376.14, found 376.3.

2-*Fluoro-4-(4H-1,2,4-triazol-4-yl)pyridine (177)*. To a solution of 2-fluoropyridin-4-amine (173) (300 mg, 2.68 mmol) and 1,2-diformylhydrazine (707 mg, 8.03 mmol) in pyridine (5 mL) was added Me₃SiCl (4.36 g, 40.14 mmol) and Et₃N (1.90 g, 18.73 mmol) at 25°C. The mixture was stirred at 100°C for 12 h under N₂ atmosphere. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0%~2%, MeOH/DCM). **177** (312 mg, 69.7%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.40 (s, 2H), 8.43 (d, *J* = 5.6 Hz, 1H), 7.87-7.81 (m, 1H), 7.78-7.75 (m, 1H). LCMS t_R = 0.465 min in 2.5 min chromatography, MS ESI calcd. for C₇H₆FN₄⁺ [M+H]⁺ 165.06, found 165.2.

4-(4H-1,2,4-triazol-4-yl)pyridin-2-amine (**181**). A mixture of 2-fluoro-4-(4H-1,2,4-triazol-4-yl)pyridine (**177**) (1.00 g, 6.09 mmol) in concentrated aqueous NH₃ (15 mL, 25-28% v/v) was degassed and purged with N₂ for 3 times, and then the mixture was stirred at 100°C for 72 h under N₂ atmosphere. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (FA)-ACN]; B%: 0%-25%, 25 min). **181** (80 mg, 7.85%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.15 (s, 2H), 8.14 (s, 1H), 8.04 (d, *J* = 5.6 Hz, 1H), 6.85 (dd, *J* = 2.0, 5.6 Hz, 1H), 6.65-6.62 (m, 1H), 6.33 (s, 2H). LCMS t_R = 0.242 min in 2.5 min chromatography, MS ESI calcd. for C₇H₈N₅⁺ [M+H]⁺ 162.08, found 162.1.

5-((4-(4H-1,2,4-triazol-4-yl)pyridin-2-yl)amino)-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (49).To a solution of 4-(4H-1,2,4-triazol-4-yl)pyridin-2-amine (181) (50 mg, 0.31 mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (40 mg, 0.17 mmol) in dioxane (3 mL) was added 'BuOLi (41 mg, 0.51 mmol), BINAP (16 mg, 0.03 mmol) and Pd(OAc)₂ (6 mg, 0.03 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Boston Green ODS 150*30mm*5um; mobile phase: [water (NH₄HCO₃)-MeOH]; B%: 30%-90%, 35 min). **49** (18 mg, 28.9%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.56 (s, 1H), 9.16 (s, 2H), 8.57 (d, *J* = 2.1 Hz, 1H), 8.53 (d, *J* = 1.8 Hz, 1H), 8.47 (d, *J* = 5.6 Hz, 1H), 8.46 (s, 1H), 7.43 (dd, *J* = 5.5, 2.1 Hz, 1H), 6.81 (s, 1H), 2.66 – 2.59 (m, 1H), 0.90 – 0.83 (m, 2H), 0.77 – 0.70 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 155.72, 154.91, 150.53, 149.92, 148.94, 145.18, 141.65, 140.62, 114.68, 109.02, 103.71, 77.76, 77.03, 23.50, 6.57. HPLC t_R = 3.059 min in 8 min chromatography, purity 98.5%. LCMS t_R = 1.739 min in 4 min chromatography, MS ESI calcd. for C₁₇H₁₅N₁₀⁺ [M+H]⁺ 359.15, found 359.2.

4-Nitro-2-(4H-1,2,4-triazol-4-yl)pyridine (178). To a solution of 4-nitropyridin-2-amine (174) (800 mg, 5.75 mmol) and 1,2-diformylhydrazine (1.52 g, 17.25 mmol) in pyridine (8 mL) was added Et₃N (4.07 g, 40.26mmol), Me₃SiCl (9.37 g, 86.26 mmol) at 25°C. Then the mixture was stirred at 100°C for 12 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~4%, MeOH/DCM). **178** (904 mg, 27.2%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.53 (s, 2H), 8.95 (d, *J* = 5.2 Hz, 1H), 8.77 (d, *J* = 1.6 Hz, 1H), 8.25 (s, 1H).

2-(4H-1,2,4-triazol-4-yl)pyridin-4-amine (182). To a solution of 4-nitro-2-(4H-1,2,4-triazol-4-yl)pyridine (178) (900 mg, 4.71 mmol) in MeOH (5 mL) was added Pd/C (500 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 hours. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (50 mL). The combined filtrate was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~16%, MeOH/DCM). 182 (241 mg, 29.0%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.09 (s, 2H), 7.93 (d, *J* = 6.0 Hz, 1H), 6.74 (d, *J* = 2.0 Hz, 1H), 6.54 - 6.51 (m, 3H).

5 - ((2 - (4H - 1, 2, 4 - triazol - 4 - yl)pyridin - 4 - yl)amino) - 7 - (cyclopropylamino)pyrazolo[1, 5 - a]pyrimidine - 3 - carbonitrile (**75**) (100 mg, 0.43 mmol) and 2 - (4H - 1, 2, 4 - triazol - 4 - yl)pyridin - 4 - amine (**182**) (69 mg, 0.43 mmol) in dioxane (3 mL) was added 'BuOLi (103 mg, 1.28 mmol, BINAP (40 mg, 0.06 mmol) and Pd(OAc)₂ (14 mg, 0.06 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~6%, MeOH/DCM). Then the impure product was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 22%-52%, 10min).**50**(22.6 mg, 4.8%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d* $₆) <math>\delta$ 10.50 (s, 1H), 9.10 (s, 2H), 8.62 (s, 1H), 8.54 – 8.47 (m, 2H), 8.37 (d, *J* = 5.7 Hz, 1H), 7.64 – 7.51 (m, 1H), 6.13 (s, 1H), 2.74 – 2.62 (m, 1H), 0.93 – 0.80 (m, 2H), 0.80 – 0.70 (m, 2H). ¹³C NMR (214 MHz, DMSO-*d*₆) δ 156.18, 150.42, 149.96, 149.39, 149.00, 147.18, 145.30, 140.14, 114.48, 112.63, 102.37, 77.78, 77.72, 23.41, 6.55. HPLC t_R = 3.999 min in 8 min chromatography, purity 98.4%. LCMS t_R = 1.383 min in 4 min chromatography, MS ESI calcd. for C₁₇H₁₅N₁₀+ [M+H]⁺ 359.15, found 359.4.

Experimental Procedures in Schemes 6 and 7.

Tert-butyl (2-(*methyl*(4-*nitro*-2-(4H-1,2,4-*triazol*-4-*yl*)*phenyl*)*amino*)*ethyl*)*carbamate* (**183**). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (200 mg, 0.96 mmol) and tert-butyl (2-(methylamino)ethyl)carbamate (251 mg, 1.44 mmol) in MeCN (5 mL) was added K₂CO₃ (531 mg, 3.84 mmol) at 25°C. The mixture was stirred at 100°C for 10 h under N₂ atmosphere. The reaction mixture was concentrated. Water (30 mL) was added. The resulting mixture was extracted with DCM (20 mL x3). The combined organic phase was washed with brine (20 mL), water (20 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. The residue was purified by flash silica gel chromatography (eluent of 0%~2%, MeOH/DCM) to give the product. **183** (235 mg, 56.6%) was obtained as a black brown oil. LCMS t_R = 1.368 min in 2.5 min chromatography, MS ESI calcd. for C₁₆H₂₃N₆O₄⁺ [M+H]⁺ 363.18, found 363.1.

Tert-butyl (2-((4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)(methyl)amino)ethyl)carbamate (**201**). To a solution of tertbutyl (2-(methyl(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)amino)ethyl)carbamate (**183**) (235 mg, 0.65 mmol) in MeOH (5 mL) was added Pd/C (0.14 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. The reaction mixture was stirred under H₂ (15 Psi) at 25°C for 2 h. The reaction mixture was filtered. The filter cake was washed with MeOH (30 mL). The combined filtrate was concentrated in vacuo. **201** (189 mg, 84.5%) was obtained as a black brown oil. LCMS t_R = 0.912 min in 2.5 min chromatography, MS ESI calcd. for C₁₆H₂₅N₆O₂⁺ [M+H]⁺ 333.20, found 333.2.

Tert-butyl (2-((4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4yl)phenyl)(methyl)amino)ethyl)carbamate (219). To a solution of tert-butyl (2-((4-amino-2-(4H-1,2,4-triazol-4yl)phenyl)(methyl)amino)ethyl)carbamate (201) (90)0.27 5-chloro-7mg, mmol) and (cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (32 mg, 0.14 mmol) in dioxane (3 mL) was added Cs₂CO₃ (265 mg, 0.81 mmol), BINAP (25 mg, 0.04 mmol) and Pd(OAc)₂ (9 mg, 0.04 mmol) at 25°C. Then the mixture was degased and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~3%, MeOH/DCM). 219 (180 mg, 75.9%) was obtained as a black brown solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.81 (s, 1H), 8.69 (s, 2H), 8.36 (s, 1H), 7.92-7.87 (m, 1H), 7.72-7.66 (m, 1H), 7.35-7.29 (m, 1H), 7.11-7.08 (m, 1H), 5.75 (s, 1H), 3.39-3.38 (m, 1H), 3.34-3.31 (m, 3H), 2.94-2.92 (m, 1H), 2.82-2.79 (m, 2H), 2.72-2.67 (m, 2H), 1.35 (s, 9H), 0.86-0.78 (m, 2H), 0.75-0.69 (m, 2H). LCMS $t_R = 0.913$ min in 1.5 min chromatography, MS ESI calcd. for $C_{26}H_{32}N_{11}O_2^+$ [M+H]⁺ 530.27, found 530.3.

5-((4-((2-aminoethyl)(methyl)amino)-3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(*cyclopropylamino*)*pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**51**). To a solution of tert-butyl (2-((4-((3-cyano-7-(cyclopropylamino))pyrazolo[1,5-*a*]*pyrimidin-5-y*]*amino*)-2-(4H-1,2,4-triazol-4-

yl)phenyl)(methyl)amino)ethyl)carbamate (**219**) (135 mg, 0.25 mmol) in DCM (5 mL) was added TFA (1 mL) at 25°C. The mixture was stirred at 25°C for 2 h. The reaction mixture was concentrated in vacuo. The residue was purified by prep. HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 18%-48%, 11min). **51** (16.8 mg, 14.3%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.82 (s, 1H), 8.86 (s, 2H), 8.36 (s, 1H), 8.32 (s, 1H), 7.88 (d, *J* = 2.5 Hz, 1H), 7.71 (dd, *J* = 8.8, 2.6 Hz, 1H), 7.35 (d, *J* = 8.9 Hz, 1H), 5.96 (s, 1H), 3.01 – 2.76 (m, 2H), 2.71 (t, *J* = 6.6 Hz, 2H), 2.61 (tt, *J* = 6.8, 3.6 Hz, 1H), 2.43 (s, 3H), 0.85 – 0.78 (m, 2H), 0.75 – 0.69 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.53, 150.85, 148.39, 145.00, 143.01, 141.54, 136.21, 128.40, 122.62, 120.32, 117.15, 114.69, 76.65, 76.35, 58.11, 41.42, 39.02, 23.32, 6.53. HPLC t_R = 3.157 min in 8 min chromatography, purity 92.9%. LCMS t_R = 1.768 min in 4 min chromatography, MS ESI calcd. for C₂₁H₂₄N₁₁⁺ [M+H]⁺ 430.22, found 430.4.

Tert-butyl (2-((4-amino-5-fluoro-2-nitrophenyl)(methyl)amino)ethyl)carbamate (**232**). To a solution of 2,4-difluoro-5nitroaniline (**231**) (1 g, 5.74 mmol) and tert-butyl (2-(methylamino)ethyl)carbamate (1.20 g, 6.89 mmol) in MeCN (15 mL) was added K₂CO₃ (2.38 g, 17.23 mmol). Then the reaction mixture was stirred at 60°C for 12 h. The reaction mixture was concentrated directly. The residue was purified by flash silica gel chromatography (eluent of 0~4%, MeOH/DCM). **232** (1.75 g, 83.0%) was obtained as a red solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.21-7.07 (m, 2H), 6.69-6.60 (m, 1H), 5.35 (s, 2H), 3.01-2.94 (m, 2H), 2.93-2.87 (m, 2H), 2.63 (s, 3H), 1.33 (s, 9H). LCMS t_R = 1.295 min in 1 min chromatography, MS ESI calcd. for C₁4H₂₂FN₄O₄⁺ [M+H]⁺ 329.16, found 329.0.

Tert-butyl (2-((4-(((*benzyloxy*)*carbonyl*)*amino*)-5-*fluoro*-2-*nitrophenyl*)(*methyl*)*amino*)*ethyl*)*carbamate* (237). To a solution of tert-butyl (2-((4-amino-5-fluoro-2-nitrophenyl)(methyl)amino)ethyl)carbamate (232) (1.6 g, 4.87 mmol) in THF (20 mL) was added K₂CO₃ (2.02 g, 14.62 mmol) and CbzCl (1.25 g, 7.31mmol) at 25°C under N₂. The mixture was stirred at 25°C for 12 h. The reaction mixture was concentrated directly. The residue was purified by flash silica gel chromatography (eluent of 0~23%, EtOAc/PE). 237 (1.93 g, 84.9%) was obtained as a red oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.53 (s, 1H), 8.19-7.97 (m, 1H), 7.44-7.31 (m, 6H), 7.09 (d, *J* = 13.2 Hz, 1H), 6.78 (t, J = 5.2 Hz, 1H), 5.15 (s, 2H), 3.15-3.09 (m, 4H), 2.77 (s, 3H), 1.31 (s, 9H). LCMS t_R = 0.621 min in 1 min chromatography, MS ESI calcd. for C₂₂H₂₈FN₄O₆⁺ [M+H]⁺ 463.20, found 463.2.

Tert-butyl (2-((2-*amino*-4-(((*benzyloxy*)*carbonyl*)*amino*)-5-*fluorophenyl*)(*methyl*)*amino*)*ethyl*)*carbamate* (242). To a solution of tert-butyl (2-((4-(((*benzyloxy*)*carbonyl*)*amino*)-5-*fluoro*-2-nitrophenyl)(methyl)*amino*)*ethyl*)*carbamate* (237) (1.70 g, 3.68 mmol) in EtOH (15 mL) was added NH₄Cl (590 mg, 11.03 mmol) dissolved in H₂O (3 mL) and Fe (616 mg, 11.03 mmol) at 25°C. Then the mixture was stirred at 100°C under N₂ atmosphere for 2 h. The reaction mixture was filtered and the solid was washed with MeOH (100 mL). The combined filtrate was concentrated in vacuo. The residue was purified by prep. HPLC (column: Welch SiO2 10u 250*300mm; mobile phase: [Heptane-EtOH(0.1%NH3H2O)]; B%: 10%-40%, 10min). **242** (418 mg, 24.7%) was obtained as a gray oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.45-7.31 (m, 6H), 6.91-6.75 (m, 3H), 5.11 (s, 2H), 3.42 (s, 3H), 3.19-3.15 (m, 2H), 3.08-2.99 (m, 2H), 2.81-2.74 (m, 2H), 1.37 (s, 9H). LCMS t_R = 0.867 min in 1.5 min chromatography, MS ESI calcd. for C₂₂H₃₀FN₄O₄⁺ [M+H]⁺ 433.22, found 433.1.

Tert-butyl (2-((((benzyloxy)carbonyl)amino)-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)(methyl)amino)ethyl)carbamate (247). To a solution of tert-butyl (2-((2-amino-4-(((benzyloxy)carbonyl)amino)-

5-fluorophenyl)(methyl)amino)ethyl)carbamate (**242**) (390 mg, 0.90 mmol) and 1,2-diformylhydrazine (237 mg, 2.7 mmol) in pyridine (5 mL) was added Et₃N (639 mg, 6.31 mmol) and Me₃SiCl (1.47 g, 13.53 mmol) at 25°C. The mixture was stirred at 100°C for 12 h. The reaction mixture was concentrated. Saturated NaHCO₃ (50 mL) solution was added. The resulting mixture was extracted with EtOAc (30 mL x 3). The combined organic phase was washed with brine (30 mL), water (30 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. **247** (350 mg, 63.5%) was obtained as a black brown gum. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.71 (s, 2H), 7.44-7.30 (m, 7H), 5.14 (s, 2H), 3.35 (s, 2H), 2.96-2.87 (m, 2H), 2.77-2.69 (m, 2H), 2.43 (s, 3H), 1.33 (s, 9H). LCMS t_R = 0.525 min in 1 min chromatography, MS ESI calcd. for C₂₄H₃₀FN₆O₄⁺ [M+H]⁺ 485.23, found 485.1.

Tert-butyl (2-((4-amino-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)(methyl)amino)ethyl)carbamate (252). To a solution of tert-butyl (2-((4-(((benzyloxy)carbonyl)amino)-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)(methyl)amino)ethyl)carbamate (247) (320 mg, 0.66 mmol) in MeOH (10 mL) was added Pd/C (0.4 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. The reaction mixture was stirred under H₂ (15 psi) at 25°C for 3 h. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (10 mL x3). The combine filtrate was concentrated in vacuo. The residue was purified by prep. HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water(NH4HCO₃)-ACN]; B%: 8%-48%, 36min). 252 (80 mg, 33.1%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-d₆) δ 8.67 (s, 2H), 7.10 (d, *J* = 12.8 Hz, 1H), 6.74-6.64 (m, 2H), 5.24 (s, 2H), 2.85-2.78 (m, 2H), 2.68-2.64 (m, 2H), 2.33 (s, 3H), 1.36-1.33 (m, 9H). LCMS t_R = 0.449 min in 1 min chromatography, MS ESI calcd. for C₁₆H₂₄FN₆O₂+ [M+H]⁺ 351.19, found 351.1.

Tert-butyl (2-((4-((3-cvano-7-(cvclopropylamino)pyrazolo[1,5-a]pvrimidin-5-yl)amino)-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)(methyl)amino)ethyl)carbamate (257). solution of 5-chloro-7-То a (cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (70 mg, 0.20 mmol) and tert-butyl (2-((4-amino-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)(methyl)amino)ethyl)carbamate (252) (42 mg, 0.18 mmol) in dioxane (5 mL) was added Cs₂CO₃ (195 mg, 0.60 mmol), BINAP (19 mg, 0.03 mmol) and Pd(OAc)₂ (7 mg, 0.03 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 6 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~3MeOH/DCM). 257 (100 mg, 38.3%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.48 (s, 1H), 8.74-8.66 (m, 4H), 8.35 (s, 1H), 8.32 (s, 1H), 8.24-8.18 (m, 1H), 6.18 (s, 1H), 2.78-2.74 (m, 2H), 2.69-2.65 (m, 2H), 2.33 (s, 3H), 1.35 (s, 9H), 1.26-1.22 (m, 1H), 0.83-0.79 (m, 2H), 0.73-0.70 (m, 2H). LCMS $t_{R} = 0.880$ min in 4 min chromatography, MS ESI calcd. for C₂₆H₃₁FN₁₁O₂⁺ [M+H]⁺ 548.26, found 548.2.

5-[4-[2-Aminoethyl(methyl)amino]-2-fluoro-5-(1,2,4-triazol-4-yl)anilino]-7-

(*cyclopropylamino*)*pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**52**). A mixture of tert-butyl (2-((4-((3-cyano-7-(cyclopropylamino))pyrazolo[1,5-*a*]*pyrimidin-5-y*]*amino*)-5-fluoro-2-(4H-1,2,4-triazol-4-

yl)phenyl)(methyl)amino)ethyl)carbamate (**257**) (80 mg, 0.15 mmol) in DCM (2 mL) was added TFA (1 mL) at 25°C. The reaction mixture was stirred at 25°C for 3 h. The reaction mixture was concentrated in vacuo. The residue was purified by prep. HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water(HCl)-ACN]; B%: 2%-42%, 36min). **52** as a HCl salt (13 mg, 19.0%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.65 (s, 1H), 9.38 (s, 2H), 8.37 (s, 1H), 8.35 (d, *J* = 8.7 Hz, 1H), 8.20 – 8.01 (m, 3H), 7.44 (d, *J* = 12.5 Hz, 1H), 6.22 (s, 1H), 3.09 (q, *J* = 6.2 Hz, 2H), 2.92 – 2.77 (m, 2H), 2.66 – 2.56 (m, 1H), 2.41 (s, 3H), 0.86 – 0.79 (m, 2H), 0.75 – 0.68 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.85, 154.47 (d, *J* = 250.2 Hz), 150.71, 148.62, 145.10, 143.56, 143.05 (d, *J* = 8.0 Hz), 123.41 (d, *J* = 13.1 Hz), 122.79 (d, *J* = 2.9 Hz), 122.57 (d, *J* = 3.1 Hz), 114.76, 109.75 (d, *J* = 22.4 Hz), 76.67, 76.62, 51.44, 41.50, 35.71, 23.46, 6.62. HPLC t_R = 3.056 min in 8 min chromatography, purity 95.8%. LCMS t_R = 1.430 min in 4 min chromatography, MS ESI calcd. for C₂₁H₂₃FN₁₁+ [M+H]+ 448.21, found 448.4.

Tert-butyl (2-(*ethyl*(4-*nitro*-2-(4H-1,2,4-*triazol*-4-*yl*)*phenyl*)*amino*)*ethyl*)*carbamate* (**184**). To a mixture of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (600 mg, 2.88 mmol) and tert-butyl (2-(ethylamino)ethyl)carbamate (814.0 mg, 4.32 mmol) in MeCN (20 mL) was added K₂CO₃ (1.59 g, 11.53 mmol). The reaction was heated to 100°C and stirred for 12 h. After being cooled to 25°C, the reaction mixture was concentrated to give a crude product. The residue was purified by flash silica gel chromatography (eluent of 0%~5%,MeOH/DCM). **184** (630 mg, 1.12 mmol, 38.96% yield) was obtained as brown oil. LCMS t_R = 0.556 min in 1 min chromatography, MS ESI calcd. for C₁₇H₂₅N₆O₄⁺ [M+H]⁺ 377.19, found 377.2.

Tert-butyl (2-((4-*amino*-2-(4H-1,2,4-*triazol*-4-yl)*phenyl*)(*ethyl*)*amino*)*ethyl*)*carbamate* (**202**). To a solution of tert-butyl (2-(ethyl(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)amino)*ethyl*)*carbamate* (**184**) (630 mg, 1.67 mmol) in MeOH (3 mL) and THF (3 mL) was added Pd/C (310 mg, 291.30 µmol, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. The reaction mixture was stirred under H₂ (15 Psi) at 25°C for 12 h. The reaction mixture was filtered. The filter cake was washed with MeOH (10 mL *3), then the filtrate was concentrated. **202** (500 mg, 1.44 mmol, 86.23% yield) was obtained as a colorless oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.67 (s, 2H), 7.09 (d, *J* = 8.8 Hz, 1H), 6.56 - 6.48 (m, 1H), 5.24 (s, 2H), 2.87 - 2.79 (m, 2H), 2.79 - 2.73 (m, 2H), 2.58 (q, *J* = 7.2 Hz, 2H), 1.35 (s, 9H), 0.69 (t, *J* = 7.2 Hz, 3H).

(2-((4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4-Tert-butyl vl)phenvl)(ethvl)amino)ethvl)carbamate (220). To a solution of tert-butyl (2-((4-amino-2-(4H-1,2,4-triazol-4yl)phenyl)(ethyl)amino)ethyl)carbamate 952.58 (202)(330 mg, µmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (178.1 mg, 762.07 µmol) in dioxane (5 mL) was added BINAP (89 mg, 142.89 µmol), Pd(OAc)₂ (32.1 mg, 142.89 µmol) and Cs₂CO₃ (931.1 mg, 2.86 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in microwave at 130°C for 0.5 hour. The reaction mixture was concentrated. The residue was purified by prep-HPLC (column: Welch Ultimate XB-NH₂ 250*50*10um; mobile phase: [Heptane-EtOH(0.1%NH₃H₂O)]; gradient: 15%-45% B over 15 min). 220 (600 mg, 937.71 μ mol, 98.44% yield) was obtained as a white solid. LCMS t_R = 0.585 min in 1 min chromatography, MS ESI calcd. for $C_{27}H_{34}N_{11}O_2^+$ [M+H]⁺ 544.29, found 544.4.

5-((4-((2-aminoethyl)(ethyl)amino)-3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(*cyclopropylamino*)*pyrazolo*[*1*,5-*a*]*pyrimidine-3-carbonitrile* (**53**). To a solution of tert-butyl (2-((4-((3-cyano-7-(cyclopropylamino)pyrazolo[*1*,5-*a*]*pyrimidin-5-yl*)*amino*)-2-(4H-1,2,4-triazol-4-yl)*phenyl*)(ethyl)*amino*)ethyl)*carbamate* (**220**) (600 mg, 1.10 mmol) in DCM (2 mL) was added TFA (1.54 g, 13.46 mmol, 1 mL). The mixture was stirred at 25 °C for 1 h. The reaction mixture was concentrated directly. The residue was purified by prep-HPLC (column: Xtimate C18 150*40mm*10um;mobile phase: [water(FA)-ACN];gradient:0%-34% B over 25 min). **53** as a formate salt (132 mg, 291.85 µmol, 72.31% yield) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.04 (s, 1H), 8.84 (s, 2H), 8.42 (s, 1H), 8.35 (s, 1H), 7.94 (d, *J* = 2.5 Hz, 1H), 7.78 (dd, *J* = 8.8, 2.6 Hz, 1H), 7.37 (d, *J* = 8.8 Hz, 1H), 6.00 (s, 1H), 3.02 (t, *J* = 7.1 Hz, 2H), 2.71 (t, *J* = 7.1 Hz, 2H), 2.66 (q, *J* = 7.1 Hz, 2H), 2.61 (tt, *J* = 6.9, 3.6 Hz, 1H), 0.85 – 0.80 (m, 2H), 0.77 (t, *J* = 7.1 Hz, 3H), 0.74 – 0.69 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 165.80, 156.57, 150.82, 148.40, 145.01, 143.05, 138.34, 137.00, 129.50, 123.94, 119.94, 117.25, 114.72, 76.73, 76.57, 49.96, 47.61, 36.58, 23.38, 11.40, 6.57. HPLC t_R = 3.181 min in 8 min chromatography, purity 96.1%. LCMS t_R = 1.302 min in 4 min chromatography, MS ESI calcd. for C₂₂H₂₆N₁₁⁺ [M+H]⁺ 444.24, found 444.3.

Tert-butyl (2-((4-amino-5-fluoro-2-nitrophenyl)(ethyl)amino)ethyl)carbamate (233). To a solution of 2,4-difluoro-5nitroaniline (231) (1.35 g, 7.75 mmol) and tert-butyl (2-(ethylamino)ethyl)carbamate (1.46 g, 7.75 mmol) in MeCN (6 mL) was added K₂CO₃ (3.22 g, 23.26 mmol). Then the reaction mixture was stirred at 100°C for 10 h. The reaction mixture was concentrated directly. The residue was purified by flash silica gel chromatography (eluent of 0~21%, EtOAc/PE). 233 (2.08 g, 52.8%) was obtained as a red solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.22 (d, *J* = 12.8 Hz, 1H), 7.09 (d, *J* = 8.8 Hz, 1H), 6.62-6.53 (m, 1H), 5.77-5.68 (m, 1H), 5.50 (s, 2H), 2.93-2.85 (m, 5H), 1.34 (s, 9H), 0.86 (t, *J* = 7.2 Hz, 3H). LCMS t_R = 0.523 min in 1 min chromatography, MS ESI calcd. for C₁₅H₂₄FN₄O₄⁺ [M+H]⁺ 343.18, found 343.0.

Tert-butyl (2-((4-(((*benzyloxy*)*carbonyl*)*amino*)-5-*fluoro*-2-*nitrophenyl*)(*ethyl*)*amino*)*ethyl*)*carbamate* (**238**). To a solution of tert-butyl (2-((4-amino-5-fluoro-2-nitrophenyl)(ethyl)amino)ethyl)*carbamate* (**233**) (2.00 g, 4.87 mmol) in THF (20 mL) was added K₂CO₃ (1.61 g, 11.68 mmol) and CbzCl (1.49 g, 8.76 mmol) at 25°C under N₂. The mixture was stirred at 25°C for 3 h. The residue was washed with water (180 mL) and extracted with DCM (200 mL x2). The organic phase was dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~21%, EtOAc/PE). **238** (2.26 g, 72.4%) was obtained as a orange solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.62 (s, 1H), 8.15-7.98 (m, 1H), 7.46-7.30 (m, 5H), 7.23 (d, *J* = 12.8 Hz, 1H), 6.73-6.62 (m, 1H), 5.25-5.10 (m, 2H), 3.20-3.16 (m, 2H), 3.13 (d, *J* = 7.2 Hz, 1H), 3.08-3.05 (m, 1H), 3.04-2.97 (m, 2H), 1.32 (s, 9H), 1.00 (t, *J* = 7.2 Hz, 3H). LCMS t_R = 0.767 min in 1 min chromatography, MS ESI calcd. for C₂₃H₃₀FN₄O₆⁺ [M+H]⁺ 477.21, found 477.0.

Tert-butyl (2-((2-amino-4-(((benzyloxy)carbonyl)amino)-5-fluorophenyl)(ethyl)amino)ethyl)carbamate (243). To a solution of tert-butyl (2-((4-(((benzyloxy)carbonyl)amino)-5-fluoro-2-nitrophenyl)(ethyl)amino)ethyl)carbamate (238)

(2.00 g, 4.20 mmol) in EtOH (30 mL) was added NH₄Cl (2.25 g, 41.97 mmol) dissolved in H₂O (15 mL) and Fe (1.17 g, 20.99 mmol) at 25°C. Then the mixture was stirred at 100°C under N₂ atmosphere for 12 h. The reaction mixture was filtered and the solid was washed with EtOH (50 mL x2). The combined filtrate was concentrated in vacuo. The residue was purified by prep-HPLC (column: Welch Ultimate XB-CN 250*50*10um; mobile phase: [Heptane-EtOH (0.1%NH3H2O)]; B%: 25%-49%, 9min). **243** (960 mg, 48.4%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.06 (s, 1H), 7.43-7.32 (m, 5H), 6.93-6.73 (m, 3H), 5.12 (s, 2H), 4.76 (s, 2H), 2.97-2.89 (m, 2H), 2.88-2.80 (m, 4H), 1.36 (s, 9H), 0.87 (t, *J* = 6.8 Hz, 3H). LCMS t_R = 0.882 min in 1.5 min chromatography, MS ESI calcd. for C₂₃H₃₂FN₄O₄⁺ [M+H]⁺ 447.24, found 447.2.

Tert-butyl (2-((4-(((benzyloxy)carbonyl)amino)-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)(ethyl)amino)ethyl)carbamate (248).То solution of tert-butyl (2-((2-amino-4-(((benzyloxy)carbonyl)amino)-5а fluorophenyl)(ethyl)amino)ethyl)carbamate (243) (830 mg, 1.86 mmol) and 1,2-diformylhydrazine (491 mg, 5.58 mmol) in pyridine (3 mL) was added Et₃N (1.32 g, 13.01 mmol) at 25°C. The mixture was stirred at 100 °C for 30 min, then Me₃SiCl (3.03 g, 27.88 mmol) was added to the mixture. The mixture was stirred at 100 °C for 11.5 h. The reaction mixture was concentrated. The residue was purified by prep. HPLC (column: Welch Ultimate XB-CN 250*50*10um; mobile phase: [Heptane-EtOH (0.1%NH3H2O)]; B%: 10%-50%, 15min). 248 (200 mg, 19.1%) was obtained as a brown solid. ¹H NMR $(400 \text{ MHz}, \text{DMSO-}d_6) \delta 9.58 \text{ (s, 1H)}, 8.72 \text{ (s, 2H)}, 7.59 \text{ (d, } J = 8.0 \text{ Hz}, 1\text{H}), 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 1H)}, 7.22 \text{ (d, J = 0.0 \text{ Hz}, 1\text{ H})}, 7.44-7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 4H)}, 7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 4H)}, 7.38 \text{ (m, 4H)}, 7.37-7.33 \text{ (m, 4H)}, 7.38 \text{ (m, 4$ J = 12.4 Hz, 1H), 6.81-6.74 (s, 1H), 5.15 (s, 2H), 2.94-2.89 (m, 2H), 2.88-2.81 (m, 2H), 2.64 (d, J = 7.2 Hz, 2H), 1.36-1.32 (m, 9H), 0.75 (t, J = 7.2 Hz, 3H). LCMS $t_{R} = 1.047$ min in 2.5 min chromatography, MS ESI calcd. for $C_{25}H_{32}FN_6O_4^+$ [M+H]⁺ 499.25, found 499.4.

Tert-butyl (2-((4-*amino*-5-*fluoro*-2-(4H-1,2,4-*triazol*-4-*yl*)*phenyl*)(*ethyl*)*amino*)*ethyl*)*carbamate* (**253**). To a solution of tert-butyl (2-((4-(((benzyloxy)carbonyl)amino)-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)(*ethyl*)*amino*)*ethyl*)*carbamate* (**248**) (135 mg, 0.27 mmol) in MeOH (10 mL) was added Pd/C (0.11 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. The reaction mixture was stirred under H₂ (15 psi) at 25°C for 2 h. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (5 mL x3). The combine filtrate was concentrated in vacuo. **253** (125 mg, crude) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.66 (s, 2H), 7.11 (d, *J* = 12.8 Hz, 1H), 6.75-6.88 (m, 2H), 5.27 (s, 2H), 2.85-2.81 (m, 2H), 2.78-2.72 (m, 2H), 2.57 (d, *J* = 7.2 Hz, 2H), 1.35 (s, 9H), 0.70 (t, *J* = 7.2 Hz, 3H). LCMS t_R = 0.572 min in 1 min chromatography, MS ESI calcd. for C₁₇H₂₆FN₆O₂⁺ [M+H]⁺ 365.21, found 365.0.

Tert-butyl (2-((4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)(ethyl)amino)ethyl)carbamate (258).To а solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (110 mg, 0.30 mmol) and tert-butyl (2-((4-amino-5fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)(ethyl)amino)ethyl)carbamate (253) (56 mg, 0.24 mmol) in dioxane (4 mL) was added Cs₂CO₃ (295 mg, 0.91 mmol), BINAP (28 mg, 0.05 mmol) and Pd(OAc)₂ (10 mg, 0.05 mmol) at 25°C. Then the mixture was degassed and purged with N2. The reaction mixture was heated in a microwave at 130°C for 5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of $0 \sim 2\%$, MeOH/DCM). 258 (140 mg, 75.0%) was obtained as a black solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.53-9.47 (m, 1H), 8.82-8.77 (m, 1H), 8.72 (s, 2H), 8.36 (s, 1H), 8.32 (s, 1H), 8.29-8.24 (m, 1H), 7.34-7.26 (m, 2H), 3.17 (s, 1H), 3.16 (s, 1H), 2.96-2.93 (m, 2H), 2.89-2.85 (m, 2H), 2.70-2.68 (m, 1H), 1.37 (s, 3H), 1.35 (s, 9H), 0.80-0.78 (m, 2H), 0.73-0.71 (m, 2H). LCMS $t_R = 0.807$ min in 2 min chromatography, MS ESI calcd. for $C_{27}H_{33}FN_{11}O_2^+$ [M+H]⁺ 562.28, found 562.3.

5-[4-[2-aminoethyl(ethyl)amino]-2-fluoro-5-(1,2,4-triazol-4-yl)anilino]-7-

(*cyclopropylamino*)*pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**54**). A mixture of tert-butyl (2-((4-((3-cyano-7-(cyclopropylamino))pyrazolo[1,5-*a*]*pyrimidin-5-y*]*amino*)-5-fluoro-2-(4H-1,2,4-triazol-4-

yl)phenyl)(ethyl)amino)ethyl)carbamate (**258**) (110 mg, 0.19 mmol) in DCM (2 mL) was added TFA (2 mL) at 25°C. The reaction mixture was stirred at 25°C for 3 h. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (HCl)-ACN]; B%: 4%-44%, 36min). The residue was purified by SFC (column: DAICEL CHIRALPAK AD (250mm*30mm, 10um); mobile phase: [0.1% NH₃H₂O EtOH]; B%: 45%-45%, min). The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (HCl)-ACN]; B%: 0%-38%, 36min). **54** as a HCl salt (6.5 mg, 27.0%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.58 (s, 1H), 8.93 – 8.75 (m, 2H), 8.37 (s, 2H), 8.30 (dd, *J* = 8.4, 2.5 Hz, 1H), 7.74 (s, 3H), 7.39

(dd, J = 12.7, 2.5 Hz, 1H), 6.22 (d, J = 1.9 Hz, 1H), 3.07 (q, J = 8.0, 6.1 Hz, 2H), 2.86 – 2.74 (m, 2H), 2.67 (q, J = 7.0 Hz, 2H), 2.63 – 2.56 (m, 1H), 0.87 – 0.74 (m, 5H), 0.74 – 0.69 (m, 2H). ¹³C NMR (214 MHz, DMSO- d_6) δ 156.73, 153.54 (d, J = 248.3 Hz), 150.65, 148.57, 145.05, 143.12, 140.35 (d, J = 7.6 Hz), 124.53, 123.40 (d, J = 12.7 Hz), 122.04, 114.64, 110.48 (d, J = 21.8 Hz), 76.67, 76.65, 48.01, 47.23, 35.95, 23.40, 11.24, 6.56. HPLC t_R = 3.308 min in 8 min chromatography, purity 96.7%. LCMS t_R = 5.514 min in 10 min chromatography, MS ESI calcd. for C₂₂H₂₅FN₁₁⁺ [M+H]⁺ 462.23, found 462.2.

(*R*)-tert-butyl (1-(methyl(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)amino)propan-2-yl)carbamate (185). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (176) (600 mg, 2.02 mmol) and tert-butyl (R)-(1-(methylamino)propan-2-yl)carbamate (380 mg, 2.02 mmol) in MeCN (7 mL) was added K₂CO₃ (836 mg, 6.05 mmol) at 25°C. The mixture was stirred at 100°C for 10 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~8% MeOH/DCM). 185 (700 mg, 65.4%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.81 (s, 2H), 8.19 - 8.16 (m, 2H), 7.24 - 7.21 (m, 1H), 6.71 (d, *J* = 9.2 Hz, 1H), 3.73 - 3.70 (m, 1H), 2.86 - 2.84 (m, 1H), 2.75 - 2.72 (m, 1H), 2.65 (s, 3H), 1.31 (s, 9H), 0.88 (d, *J* = 6.8 Hz, 3H).

(*R*)-*tert-butyl* (1-((4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)(methyl)amino)propan-2-yl)carbamate (**203**). To a solution of (R)-tert-butyl (1-(methyl(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)amino)propan-2-yl)carbamate (**185**) (686 mg, 1.82 mmol) in MeOH (8 mL) was added Pd/C (500 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (50 mL). The combined filtrate was concentrated in vacuo. **203** (570 mg, 51.4%) was obtained as a purple oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.71 - 8.67 (m, 2H), 7.08 (d, *J* = 8.4 Hz, 1H), 6.65 - 6.63 (m, 2H), 6.51 (d, *J* = 2.4 Hz, 1H), 5.22 (s, 2H), 3.48 - 3.41 (m, 1H), 2.72 - 2.66 (m, 1H), 2.56 - 2.52 (m, 1H), 2.26 (s, 3H), 1.37 - 1.36 (m, 9H), 0.81 (d, *J* = 6.4 Hz, 3H). LCMS t_R = 0.916 min in 2.5 min chromatography, MS ESI calcd. for C₁₇H₂₇N₆O₂⁺ [M+H]⁺ 347.22, found 347.0.

(1-((4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4-(R)-tert-butyl yl)phenyl)(methyl)amino)propan-2-yl)carbamate (221). To a solution of (R)-tert-butyl (1-((4-amino-2-(4H-1,2,4-triazol-4vl)phenvl)(methvl)amino)propan-2-vl)carbamate (203)(265)mg, 0.73 mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (170 mg, 0.73 mmol) in dioxane (3 mL) was added Cs₂CO₃ (711 mg, 2.18 mmol), BINAP (68 mg, 0.11 mmol) and Pd(OAc)₂ (25 mg, 0.11 mmol) at 25°C. Then the mixture was degassed and purged with N_2 . The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~6%, MeOH/DCM). **221** (553 mg, 36.4%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.80 (s, 1H), 8.80 - 8.75 (m, 2H), 8.37 - 8.34 (m, 1H), 8.29 (s, 1H), 7.90 (d, J = 2.4 Hz, 1H), 7.68 (dd, J = 2.0, 8.8 Hz, 1H), 7.30 (d, J = 9.0 Hz, 1H), 5.96 (s, 1H), 3.65 - 3.53 (m, 1H), 2.75 - 2.70 (m, 2H), 2.63 - 2.58 (m, 1H), 2.36 (s, 3H), 1.36 (s, 10H), 0.90 (d, J = 6.4 Hz, 3H), 0.82- 0.80 (m, 2H), 0.72 (d, J = 3.2 Hz, 2H). LCMS t_R = 1.375 min in 2.5 min chromatography, MS ESI calcd. for C₂₇H₃₄N₁₁O₂⁺ [M+H]⁺ 544.29, found 544.3.

(R)-5-((4-((2-aminopropyl)(methyl)amino)-3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(*cyclopropylamino*)*pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**55**). To a solution of (R)-tert-butyl (1-((4-((3-cyano-7-(cyclopropylamino))pyrazolo[1,5-*a*]*pyrimidin-5-y*])*amino*)-2-(4H-1,2,4-triazol-4-y])*phenyl*)(methyl)*amino*)*propan-2-*

yl)carbamate (**221**) (550 mg, 1.01 mmol) in DCM (5 mL) and TFA (2.5 mL) at 25°C. The mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Xtimate C18 150*40mm*10um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 30%-60%, 10min). The crude product was purified by SFC (column: DAICEL CHIRALPAK AD (250mm*30mm, 10um); mobile phase: [0.1%NH₃H₂O ETOH]; B%: 35%-35%, min). **55** (25.5 mg, 62.5%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.83 (s, 1H), 8.86 (s, 2H), 8.36 (s, 1H), 7.88 (d, *J* = 2.5 Hz, 1H), 7.73 (dd, *J* = 8.9, 2.6 Hz, 1H), 7.37 (d, *J* = 8.8 Hz, 1H), 5.96 (s, 1H), 2.84 – 2.71 (m, 1H), 2.65 – 2.55 (m, 2H), 2.47 (d, *J* = 8.3 Hz, 1H), 2.42 (s, 3H), 0.86 – 0.77 (m, 5H), 0.76 – 0.67 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.53, 150.82, 148.38, 145.00, 143.11, 142.00, 136.35, 128.48, 122.93, 120.45, 117.18, 114.68, 76.67, 76.37, 64.17, 44.29, 42.23, 23.32, 21.86, 6.54. HPLC t_R = 2.999 min in 8 min chromatography, purity 99.1%. LCMS t_R = 1.110 min in 4 min chromatography, MS ESI calcd. for C₂₂H₂₆N₁₁⁺ [M+H]⁺ 444.24, found 444.4.

(S)-tert-butyl (1-(methyl(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)amino)propan-2-yl)carbamate (186). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (176) (450 mg, 1.95 mmol) and tert-butyl (S)-(1-(methylamino)propan-2-

yl)carbamate (366 mg, 1.95 mmol) in MeCN (5 mL) was added K₂CO₃ (807 mg, 5.84 mmol) at 25°C. The mixture was stirred at 100°C for 10 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~15% MeOH/DCM). **186** (590 mg, 69.9%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.81 (s, 2H), 8.21 - 8.15 (m, 2H), 7.24 - 7.19 (m, 1H), 6.72 (d, *J* = 8.8 Hz, 1H), 3.78 - 3.65 (m, 2H), 2.86 (dd, *J* = 4.8, 14.0 Hz, 1H), 2.65 (s, 3H), 1.31 (s, 9H), 0.88 (d, *J* = 6.8 Hz, 3H). LCMS t_R = 0.476 min in 1 min chromatography, MS ESI calcd. for C₁₇H₂₅N₆O₄⁺ [M+H]⁺ 377.19, found 377.1.

(*S*)-*tert-butyl* (1-((4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)(methyl)amino)propan-2-yl)carbamate (**204**). To a solution of (S)-tert-butyl (1-(methyl(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)amino)propan-2-yl)carbamate (**186**) (560 mg, 1.49 mmol) in MeOH (6 mL) was added Pd/C (360 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (100 mL). The residue was purified by flash silica gel chromatography (eluent of 0~9%, MeOH/DCM). **204** (253 mg, 44.3%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.69 (s, 2H), 7.09 (d, *J* = 8.8 Hz, 1H), 6.69 - 6.59 (m, 2H), 6.51 (d, *J* = 2.8 Hz, 1H), 5.20 (s, 2H), 3.44 (q, *J* = 6.8 Hz, 1H), 2.73 - 2.64 (m, 1H), 2.55 (dd, *J* = 6.4, 12.4 Hz, 1H), 2.26 (s, 3H), 1.36 (s, 9H), 0.81 (d, *J* = 6.4 Hz, 3H). LCMS t_R = 0.407 min in 1 min chromatography, MS ESI calcd. for C₁₇H₂₇N₆O₂⁺ [M+H]⁺ 347.22, found 347.1.

(1-((4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4-(S)-tert-butyl yl)phenyl)(methyl)amino)propan-2-yl)carbamate (222). To a solution of (S)-tert-butyl (1-((4-amino-2-(4H-1,2,4-triazol-4vl)phenvl)(methvl)amino)propan-2-vl)carbamate (204)0.59 (204)mg, mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (125 mg, 0.53 mmol) in dioxane (3 mL) was added Cs₂CO₃ (523 mg, 1.60 mmol), BINAP (50 mg, 0.08 mmol) and Pd(OAc)₂ (18 mg, 0.08 mmol) at 25°C. Then the mixture was degassed and purged with N_2 . The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~13%, MeOH/DCM). 222 (153 mg, 31.9%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.81 (s, 1H), 8.83 - 8.72 (m, 2H), 8.36 (s, 1H), 8.30 (s, 1H), 7.89 (s, 1H), 7.68 (d, J = 8.0 Hz, 1H), 7.30 (d, J = 8.8 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 4.10 (q, J = 8.8 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 4.10 (q, J = 8.8 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 4.10 (q, J = 8.8 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 4.10 (q, J = 8.8 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 4.10 (q, J = 8.8 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 4.10 (q, J = 8.8 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 4.10 (q, J = 8.8 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 4.10 (q, J = 8.8 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 4.10 (q, J = 8.8 Hz, 1H), 6.73 (d, J = 8.4 Hz, 1H), 4.10 (q, J = 8.8 Hz, 1H), 4.10 (q, J = 8J = 5.2 Hz, 1H), 3.67 - 3.52 (m, 1H), 2.73 (d, J = 6.8 Hz, 2H), 2.61 (s, 1H), 2.35 (s, 3H), 1.36 (s, 9H), 0.90 (d, J = 6.4 Hz, 2H), 2.61 (s, 1H), 2.73 (s, 2H), 2.61 (s, 2H), 2.6 3H), 0.84 - 0.78 (m, 2H), 0.72 (d, J = 3.2 Hz, 2H). LCMS t_R = 0.533 min in 1min chromatography, MS ESI calcd. for $C_{27}H_{34}N_{11}O_2^+$ [M+H]⁺ 544.29, found 544.3.

(S)-5-((4-((2-aminopropyl)(methyl)amino)-3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(*cyclopropylamino*)*pyrazolo*[*1,5-a*]*pyrimidine-3-carbonitrile* (**56**). To a solution of (S)-tert-butyl (1-((4-((3-cyano-7-(cyclopropylamino))pyrazolo[*1,5-a*]*pyrimidin-5-yl*)*amino*)-2-(4H-1,2,4-triazol-4-yl)*phenyl*)(methyl)*amino*)*propan-2-yl*)*carbamate* (**222**) (149 mg, 0.27 mmol) in DCM (1 mL) and TFA (0.5 mL) at 25°C. The mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 20%-59%, 9min). The crude product was purified by SFC (column: DAICEL CHIRALPAK AD (250mm*30mm, 10um); mobile phase: [0.1%NH₃H₂O IPA]; B%: 35%-35%, min). **56** (2.3 mg, 26.1%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.05 (s, 1H), 8.84 (s, 2H), 8.37 (s, 1H), 8.33 (s, 1H), 7.92 (s, 1H), 7.88 – 7.77 (m, 4H), 7.42 (d, *J* = 9.0 Hz, 1H), 6.03 (s, 1H), 3.18 (dq, *J* = 14.0, 7.2, 6.7 Hz, 1H), 3.01 (dd, *J* = 13.1, 6.2 Hz, 1H), 2.80 (dd, *J* = 12.9, 7.1 Hz, 1H), 2.60 (tt, *J* = 6.9, 3.6 Hz, 1H), 2.38 (s, 3H), 1.05 (d, *J* = 6.1 Hz, 3H), 0.86 – 0.78 (m, 2H), 0.74 – 0.67 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.52, 150.77, 148.36, 145.00, 143.15, 143.10, 140.98, 137.05, 122.89, 120.38, 117.45, 114.66, 76.68, 76.53, 58.72, 44.42, 42.60, 23.33, 16.87, 6.52. HPLC t_R = 3.025 min in 8 min chromatography, purity 97.9%. LCMS t_R = 1.722 min in 4 min chromatography, MS ESI calcd. for C₂₂H₂₆N₁₁⁺ [M+H]⁺ 444.24, found 444.5.

Tert-butyl methyl(2-(*methyl*(4-*nitro*-2-(4H-1,2,4-*triazol*-4-*yl*)*phenyl*)*amino*)*ethyl*)*carbamate* (187). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (176) (200 mg, 0.96 mmol) and tert-butyl methyl(2-(methylamino)ethyl)carbamate (271 mg, 1.44 mmol) in MeCN (5 mL) was added K₂CO₃ (531 mg, 3.84 mmol) at 25°C. The mixture was stirred at 100°C for 12 h under N₂ atmosphere. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0%~9%, MeOH/DCM) to give the product. 187 (241 mg, 35.6%) was obtained as a black brown oil. LCMS t_R = 0.875 min in 1.5 min chromatography, MS ESI calcd. for C₁₇H₂₅N₆O₄⁺ [M+H]⁺ 377.19, found 377.1.

Tert-butyl (2-((4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)(methyl)amino)ethyl)(methyl)carbamate (**205**). To a solution of tert-butyl methyl(2-(methyl(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)amino)ethyl)carbamate (**187**) (241 mg, 0.64 mmol) in MeOH (5 mL) was added Pd/C (0.11 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. The reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 h. The reaction mixture was filtered. The filter cake was washed with MeOH (20 mL). The combined filtrate was concentrated in vacuo. **205** (219 mg, 84.6%) was obtained as a black brown oil. LCMS $t_R = 1.035$ min in 2.5 min chromatography, MS ESI calcd. for C₁₇H₂₇N₆O₂⁺ [M+H]⁺ 347.22, found 347.1.

Tert-butyl (2-((4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4yl)phenyl)(methyl)amino)ethyl)(methyl)carbamate (223). To a solution of tert-butyl (2-((4-amino-2-(4H-1,2,4-triazol-4vl)phenyl)(methyl)amino)ethyl)(methyl)carbamate (205)(100)mg, 0.29 mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (34 mg, 0.14 mmol) in dioxane (3 mL) was added Cs₂CO₃ (282 mg, 0.86 mmol), BINAP (27 mg, 0.04 mmol) and Pd(OAc)₂ (10 mg, 0.04 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~5%, MeOH/DCM). **223** (154 mg, 47.7%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.79 (s, 1H), 8.73 (s, 1H), 8.34 (s, 1H), 8.28 (s, 1H), 7.897.84 (m, 1H), 7.73-7.69 (m, 1H), 7.35-7.33 (m, 1H), 5.93 (s, 1H), 5.73 (s, 1H), 3.19-3.06 (m, 4H), 2.66-2.58 (m, 6H), 2.56-2.61 (m, 1H), 1.38-1.31 (m, 9H), 0.80-0.76 (m, 2H), 0.72-0.68 (m, 2H). LCMS t_R = 0.965 min in 1.5 min chromatography, MS ESI calcd. for $C_{27}H_{34}N_{11}O_2^+$ [M+H]⁺ 544.29, found 544.3.

7-(cyclopropylamino)-5-((4-(methyl(2-(methylamino)ethyl)amino)-3-(4H-1,2,4-triazol-4-

yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**57**). To a solution of tert-butyl (2-((4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-*a*]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4-

yl)phenyl)(methyl)amino)ethyl)(methyl)carbamate (**223**) (120 mg, 0.22 mmol) in DCM (5.1 mL) was added TFA (0.9 mL) at 25°C. The mixture was stirred at 25°C for 2 h. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 20%-52%, 9min). **57** (10.8 mg, 10.6%) was obtained as a white solid. ¹H NMR (850 MHz, DMSO-*d*₆) δ 9.84 (s, 1H), 8.86 (s, 2H), 8.36 (s, 1H), 8.29 (s, 1H), 7.89 (dd, *J* = 5.1, 2.6 Hz, 1H), 7.73 (ddd, *J* = 8.7, 5.8, 2.5 Hz, 1H), 7.37 (d, *J* = 8.9 Hz, 1H), 5.97 (s, 1H), 2.83 (t, *J* = 6.7 Hz, 2H), 2.61 (tt, *J* = 7.4, 3.1 Hz, 1H), 2.52 – 2.51 (m, 2H), 2.44 (s, 3H), 2.24 (s, 3H), 0.83 – 0.80 (m, 2H), 0.73 – 0.71 (m, 2H). ¹³C NMR (214 MHz, DMSO-*d*₆) δ 159.67, 153.96, 151.53, 148.14, 146.08, 144.16, 139.55, 131.64, 125.80, 123.37, 120.20, 117.81, 79.81, 79.51, 56.82, 51.19, 44.85, 38.44, 26.46, 9.67. HPLC t_R = 1.972 min in 8 min chromatography, purity 96.4%. LCMS t_R = 2.695 min in 7 min chromatography, MS ESI calcd. for C₂₂H₂₆N₁₁+ [M+H]⁺ 444.24, found 444.4.

Tert-butyl ethyl(2-(*methyl*(4-*nitro*-2-(4H-1,2,4-*triazol*-4-*yl*)*phenyl*)*amino*)*ethyl*)*carbamate* (188). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (176) (400 mg, 1.92 mmol) and tert-butyl ethyl(2-(methylamino)ethyl)carbamate (583 mg, 2.88 mmol) in MeCN (5 mL) was added K₂CO₃ (797 mg, 5.77 mmol) at 25°C. The mixture was stirred at 100°C for 12 h under N₂ atmosphere. The reaction mixture was concentrated in vacuo. Water (20 mL) was added and the resulting mixture was extracted with DCM (20 mL x3). The combined organic phase was dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. **188** (331 mg, 31.5%) was obtained as a black brown oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.94 - 8.64 (m, 2H), 8.19 - 8.15 (m, 1H), 7.34 - 7.5 (m, 1H), 7.30 - 7.26 (m, 1H), 3.19 - 3.13 (m, 4H), 3.04 - 2.92 (m, 3H), 2.73 - 2.61 (m, 2H), 1.36 - 1.33 (m, 9H), 0.99 - 0.90 (m, 3H). LCMS t_R = 0.926 min in 1.5 min chromatography, MS ESI calcd. for C₁₈H₂₇N₆O₄⁺ [M+H]⁺ 391.21, found 391.2.

Tert-butyl (2-((4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)(methyl)amino)ethyl)(ethyl)carbamate (206). To a solution of tert-butyl ethyl(2-(methyl(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)amino)ethyl)carbamate (188) (310 mg, 0.79 mmol) in MeOH (20 mL) was added Pd/C (290 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (20 mL x3). The combined filtrate was concentrated in vacuo. **206** (195 mg, 46.9%) was obtained as a black brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.69 (s, 2H), 7.15 - 7.09 (m, 1H), 6.68 - 6.64 (m, 1H), 6.54 - 6.50 (m, 1H), 5.24 (s, 2H), 3.21 - 3.17 (m, 3H), 3.02 - 2.95 (m, 4H), 2.77 - 2.66 (m, 3H), 1.39 (s, 9H), 0.96 - 0.91 (m, 3H). LCMS t_R = 1.095 min in 2.5 min chromatography, MS ESI calcd. for C₁₈H₂₉N₆O₂⁺ [M+H]⁺ 361.23, found 361.3.

tert-butyl (2-((4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4yl)phenyl)(methyl)amino)ethyl)(ethyl)carbamate (224). To a solution of tert-butyl (2-((4-amino-2-(4H-1,2,4-triazol-4yl)phenyl)(methyl)amino)ethyl)(ethyl)carbamate 0.45 5-chloro-7-(206)(162)mg, mmol) and (cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (70 mg, 0.30 mmol) in dioxane (3 mL) was added Cs₂CO₃ (293 mg, 0.90 mmol), BINAP (28 mg, 0.04 mmol) and Pd(OAc)₂ (67 mg, 0.30 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water ($NH_3H_2O+NH_4HCO_3$)-ACN]; B%: 38%-67%, 10min). **224** (65 mg, 36.8%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.81 (s, 1H), 8.77 (s, 2H), 8.39 - 8.26 (m, 2H), 7.91 - 7.85 (m, 1H), 7.72 (d, *J* = 8.0 Hz, 1H), 7.3 - 7.31 (m, 1H), 5.96 (s, 1H), 3.14 - 2.94 (m, 4H), 2.86 - 2.73 (m, 2H), 2.65 - 2.56 (m, 1H), 2.48 - 2.42 (m, 3H), 1.35 (s, 9H), 0.94 (t, J = 7.2 Hz, 3H), 0.86 - 0.77 (m, 2H), 0.77 - 0.67 (m, 2H). LCMS t_R = 0.539 min in 1 min chromatography, MS ESI calcd. for C₂₈H₃₆N₁₁O₂⁺ [M+H]⁺ 558.30, found 558.4.

7-(cyclopropylamino)-5-((4-((2-(ethylamino)ethyl)(methyl)amino)-3-(4H-1,2,4-triazol-4-

yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**58**). To a solution of tert-butyl (2-((4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-*a*]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4-

yl)phenyl)(methyl)amino)ethyl)(ethyl)carbamate (**224**) (60 mg, 0.11 mmol) in DCM (8.5 mL) was added TFA (1.5 mL) at 25°C. The mixture was stirred at 35°C for 2 hours. Then the mixture was degassed and purged with N₂. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: X timate C18 100*30mm*10um; mobile phase: [water (FA)-ACN]; B%: 15%-35%, 10min). **58** (35 mg, 70.4%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.95 (d, *J* = 11.0 Hz, 1H), 8.87 (s, 2H), 8.37 (s, 1H), 8.32 (br s, 2H), 7.92 (t, *J* = 2.7 Hz, 1H), 7.76 (dd, *J* = 8.9, 2.6 Hz, 1H), 7.38 (d, *J* = 8.9 Hz, 1H), 6.00 (d, *J* = 3.0 Hz, 1H), 2.98 (q, *J* = 6.5, 5.7 Hz, 2H), 2.80 – 2.67 (m, 4H), 2.61 (tt, *J* = 6.7, 3.5 Hz, 1H), 2.42 (s, 3H), 1.08 (td, *J* = 7.3, 2.7 Hz, 3H), 0.86 – 0.78 (m, 2H), 0.74 – 0.67 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.53, 150.82, 148.40, 145.01, 143.02, 140.59, 136.79, 128.56, 122.66, 120.22, 117.20, 114.70, 76.70, 76.47, 52.00, 44.24, 42.43, 41.85, 23.34, 12.32, 6.55. HPLC t_R = 3.400 min in 8 min chromatography, purity 99.0%. LCMS t_R = 1.656 min in 4 min chromatography, MS ESI calcd. for C₂₃H₂₈N₁₁⁺ [M+H]⁺ 458.25, found 458.4.

 N^{l} , N^{l} -diethyl- N^{2} -methyl- N^{2} -(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)ethane-1,2-diamine (**189**). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (200 mg, 0.96 mmol) and N^{1} , N^{1} -diethyl- N^{2} -methylethane-1,2-diamine (188 mg, 1.44 mmol) in MeCN (5 mL) was added K₂CO₃ (531 mg, 3.84 mmol) at 25°C. The mixture was stirred at 100°C for 12 h under N₂ atmosphere. The reaction mixture was concentrated in vacuo. Water (30 mL) was added to the residue. The resulting mixture was extracted with DCM (20 mL x3). The combined organic phase was dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. **189** (187 mg, 34.2%) was obtained as a black brown oil. ¹H NMR (400 MHz, DMSO- d_{6}) δ 8.93 (s, 2H), 8.22 - 8.14 (m, 2H), 7.29 (d, *J* = 9.2 Hz, 1H), 2.66 (s, 3H), 2.46-2.42 (m, 4H), 2.36-2.31 (m, 4H), 0.82 (t, *J* = 7.2 Hz, 6H). LCMS t_R = 0.788 min in 2.5 min chromatography, MS ESI calcd. for C₁₅H₂₃N₆O₂⁺ [M+H]⁺ 319.19, found 319.1.

 N^{l} -(2-(*diethylamino*)*ethyl*)- N^{l} -*methyl*-2-(4H-1,2,4-triazol-4-yl)*benzene*-1,4-*diamine* (207). To a solution of N¹,N¹diethyl-N²-methyl-N²-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)*ethane*-1,2-*diamine* (189) (170 mg, 0.53 mmol) in MeOH (5 mL) was added Pd/C (0.11 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. The reaction mixture was stirred under H₂ (15 Psi) at 25°C for 2 h. The reaction mixture was filtered. The filter cake was washed with MeOH (20 mL). The combined filtrate was concentrated in vacuo. 207 (149 mg, 83.1%) was obtained as a black brown oil. LCMS t_R = 0.488 min in 2.5 min chromatography, MS ESI calcd. for C₁₅H₂₅N₆⁺ [M+H]⁺ 289.21, found 289.1.

7-(cyclopropylamino)-5-((4-((2-(diethylamino)ethyl)(methyl)amino)-3-(4H-1,2,4-triazol-4-

yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (59). To a solution of N¹-(2-(diethylamino)ethyl)-N¹-methyl-2-(4H-1,2,4-triazol-4-yl)benzene-1,4-diamine (207) (145 mg, 0.50 mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (117 mg, 0.50 mmol) in dioxane (4 mL) was added Cs₂CO₃ (491mg, 1.51 mmol), BINAP (47 mg, 0.07 mmol) and Pd(OAc)₂ (16 mg, 0.07 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~12%, MeOH/DCM). The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 42%-72%, 10min). **59** (26 mg, 10.2%) was obtained as a yellow solid. ¹H NMR (850 MHz, DMSO-*d*₆) δ 9.80 (s, 1H), 8.92 (s, 2H), 8.36 (s, 1H), 8.29 (d, *J* = 2.3 Hz, 1H), 7.89 (d, *J* = 2.6 Hz, 1H), 7.70 (dd, *J* = 8.8, 2.6 Hz, 1H), 7.35 (d, *J* = 8.8 Hz, 1H), 5.96 (s, 1H), 2.78 (t, *J* = 6.9 Hz, 2H), 2.63 – 2.58 (m, 1H), 2.46 (s, 3H), 2.38 – 2.33 (m, 4H), 2.30 (t, *J* = 6.8 Hz, 2H), 0.85 (t, *J* = 7.1 Hz, 6H), 0.83 – 0.79 (m, 2H), 0.75 – 0.70 (m, 2H). ¹³C NMR (214 MHz, DMSO-*d*₆) δ 156.54, 150.84, 148.37, 144.98, 142.83, 140.92, 136.15, 128.34, 122.52, 120.08, 116.82, 114.67, 76.64, 76.32, 52.68, 49.97, 46.40, 41.61, 23.31, 11.64, 6.52. HPLC t_R = 3.325 min in 8 min chromatography, purity 96.0%. LCMS t_R = 1.708 min in 7 min chromatography, MS ESI calcd. for C₂₅H₃₂N₁₁⁺ [M+H]⁺ 486.28, found 486.5.

N-(2-((*tert-butyldimethylsilyl*)*oxy*)*ethyl*)-*N*-*methyl*-4-*nitro*-2-(4H-1,2,4-*triazol*-4-*yl*)*aniline* (**190**). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (1.00 g, 3.36 mmol) and 2-((tert-butyldimethylsilyl)oxy)-N-methylethan-1-amine (637 mg, 3.36 mmol) in MeCN (10 mL) was added K₂CO₃ (1.39 g, 10.09 mmol) at 25°C. Then the mixture was stirred at 100°C for 10 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~8%, MeOH/DCM). **190** (1.24 g, 67.0%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.86 (s, 2H), 8.21 - 8.18 (m, 1H), 8.18 - 8.16 (m, 1H), 7.30 (d, *J* = 9.2 Hz, 1H), 3.58 (t, *J* = 5.2 Hz, 2H), 2.91 (t, *J* = 5.2 Hz, 2H), 2.72 (s, 3H), 0.76 (s, 9H), 0.00 (s, 6H). LCMS t_R = 0.560 min in 1 min chromatography, MS ESI calcd. for C₁₇H₂₈N₅O₃Si⁺ [M+H]⁺ 378.20, found 378.1.

 N^{l} -(2-((tert-butyldimethylsilyl)oxy)ethyl)- N^{l} -methyl-2-(4H-1,2,4-triazol-4-yl)benzene-1,4-diamine (**208**). To a solution of N-(2-((tert-butyldimethylsilyl)oxy)ethyl)-N-methyl-4-nitro-2-(4H-1,2,4-triazol-4-yl)aniline (**190**) (1.20 g, 3.18 mmol) in MeOH (12 mL) was added Pd/C (950 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 hours. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (60 mL). The crude compound was used in the next step directly. **208** (1.15 g, 48.9%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.79 (s, 2H), 7.09 (d, *J* = 8.8 Hz, 1H), 6.65 - 6.62 (m, 1H), 6.54 (d, *J* = 2.8 Hz, 1H), 5.22 (s, 2H), 3.45 (t, *J* = 6.0 Hz, 2H), 2.76 (t, *J* = 6.0 Hz, 2H), 2.37 (s, 3H), 0.82 (s, 9H), -0.02 (s, 6H)

5-((4-((2-((tert-butyldimethylsilyl)oxy)ethyl)(methyl)amino)-3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(cvclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (225).To solution of 5-chloro-7а (cyclopropylamino)pyrazolo[1,5-*a*]pyrimidine-3-carbonitrile mg, N¹-(2-((tert-(75) (240 1.03 mmol) and butyldimethylsilyl)oxy)ethyl)-N¹-methyl-2-(4H-1,2,4-triazol-4-yl)benzene-1,4-diamine (208) (376 mg, 1.03 mmol) in dioxane (2 mL) was added Cs₂CO₃ (1.00 g, 1.28 mmol), BINAP (96 mg, 0.15 mmol) and Pd(OAc)₂ (35 mg, 0.15 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~5%, MeOH/DCM). 225 (1.00 g, 56.8%) was obtained as a gray solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.82 (s, 1H), 8.87 (s, 2H), 8.37 (s, 1H), 8.31 (s, 1H), 7.92 (d, J = 2.0 Hz, 1H), 7.71 - 7.68 (m, 1H), 7.35 (d, J = 8.8 Hz, 1H), 5.97 (s, 1H), 3.54 (t, J = 5.6 Hz, 2H), 2.84 (t, J = 6.0 Hz, 2H), 2.62 (d, J = 2.8 Hz, 1H), 2.50 (s, 3H), 0.83 (s, 11H), 0.74 (d, J = 2.8 Hz, 1H), 2.50 (s, 2H), 2.84 (s, 12H), 2.842.8 Hz, 2H), 0.00 (s, 6H). LCMS $t_R = 1.683$ min in 2.5 min chromatography, MS ESI calcd. for $C_{27}H_{37}N_{10}OSi^+$ [M+H]⁺ 545.29, found 545.4.

7-(cyclopropylamino)-5-((4-((2-hydroxyethyl)(methyl)amino)-3-(4H-1,2,4-triazol-4-

yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**60**). To a solution of 5-((4-((2-((tert-butyldimethylsilyl)oxy)ethyl)(methyl)amino)-3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(cyclopropylamino)pyrazolo[1,5-*a*]pyrimidine-3-carbonitrile (**225**) (200 mg, 0.37 mmol) in THF (1 mL) was added TBAF (0.4 mL, 1 M in THF) dropwise at 25°C. The mixture was stirred at 25°C for 10 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~14%, MeOH/DCM). The crude product was purified by prep-HPLC (column: Xtimate C18 100*30mm*10um; mobile phase: [water (FA)-ACN]; B%: 25%-55%, 10min). **60** (11 mg, 12.5%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.83 (s, 1H), 8.89 (s, 2H), 8.36 (s, 1H), 8.28 (s, 1H), 7.90 (d, *J* = 2.6 Hz, 1H), 7.69 (dd, *J* = 8.8, 2.5 Hz, 1H), 7.34 (d, *J* = 8.8 Hz, 1H), 5.97 (s, 1H), 4.57 (s, 1H), 3.37 (d, *J* = 6.1 Hz, 2H), 2.83 (t, *J* = 5.9 Hz, 2H), 2.66 – 2.55 (m, *J* = 3.3 Hz, 1H), 2.43 (s, 3H), 0.86 – 0.75 (m, 2H), 0.78 – 0.67 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.54, 150.85, 148.37, 144.96, 142.80, 140.98, 136.16, 128.32, 122.68, 120.14, 116.86, 114.69, 76.64, 76.35, 58.30, 56.82, 41.76, 23.32, 6.53. HPLC t_R = 3.467 min in 8 min chromatography, purity 98.8%. LCMS t_R = 1.365 min in 4 min chromatography, MS ESI calcd. for C₂₁H₂₃N₁₀O⁺ [M+H]⁺ 431.21, found 431.4.

2-((4-amino-5-fluoro-2-nitrophenyl)(methyl)amino)ethan-1-ol (**262**). To a solution of 2,4-difluoro-5-nitroaniline (**231**) (5.00 g, 28.7 mmol) and 2-(methylamino)ethan-1-ol (4.31 g, 57.4 mmol) in DMF (20 mL) was added K₂CO₃ (11.9 g, 86.2 mmol). Then the reaction mixture was stirred at 60°C for 2 h. The reaction mixture was concentrated and sat. aqueous NaCl (100 mL) was added. The resulting mixture was extracted with EtOAc (20 mL x 3). The combined organic phase was washed with water (30 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. The residue was purified by flash silica gel chromatography (eluent of 0~30%, EtOAc/PE). **262** (3.24 g, 98.1%) was obtained as a red oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.02 (d, *J* = 9.2 Hz, 1H), 6.95 (d, *J* = 13.2 Hz, 1H), 5.12 (s, 2H), 4.35 (t, *J* = 5.2 Hz, 1H), 3.30 (q, J = 6.4 Hz, 2H), 2.79 (t, *J* = 6.4 Hz, 2H), 2.50 (s, 3H). LCMS t_R = 0.289 min in 0.8 min chromatography, 5-95AB, LCMS ESI calcd. for C₉H₁₃FN₃O₃⁺ [M+H]⁺ 230.09, found 230.2.

 N^{1} -(2-((*tert-butyldiphenylsilyl*)*oxy*)*ethyl*)-5-*fluoro*- N^{1} -*methyl*-2-*nitrobenzene*-1,4-*diamine* (**234**). To a solution of 2-((4-amino-5-fluoro-2-nitrophenyl)(methyl)amino)ethan-1-ol (**262**) (3.24 g, 14.1 mmol), TBDPS-Cl (3.89 g, 14.1 mmol) and imidazole (2.12 g, 31.1 mmol) in DMF (11 mL) was stirred at 25°C for 12 h. The reaction mixture was diluted with brine (50 mL) and extracted with CH₂Cl₂ (50 mL). The combined organic phase was washed with water (30 mL), brine (30 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. The residue was purified by flash silica gel chromatography (eluent of 0~20%, EtOAc/PE). **234** (3.44 g, 98.5%) was obtained as a yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.56 - 7.52 (m, 4H), 7.45 - 7.37 (m, 6H), 7.18 (d, *J* = 9.1 Hz, 1H), 7.11 (d, *J* = 13.2 Hz, 1H), 5.29 (s, 2H), 3.66 (t, *J* = 5.6 Hz, 2H), 3.11 (t, *J* = 5.6 Hz, 2H), 2.68 (s, 3H), 0.91 (s, 9H). LCMS t_R = 0.577 min in 0.8 min chromatography, 5-95AB, LCMS ESI calcd. for C₂₅H₃₁FN₃O₃Si⁺ [M+H]⁺ 468.21, found 468.6.

Benzyl (4-((2-((*tert-butyldiphenylsilyl*)*oxy*)*ethyl*)(*methyl*)*amino*)-2-*fluoro-5-nitrophenyl*)*carbamate* (**239**). To a solution of N¹-(2-((*tert-butyldiphenylsilyl*)*oxy*)*ethyl*)-5-fluoro-N¹-methyl-2-nitrobenzene-1,4-diamine (**234**) (1.76 g, 3.75 mmol) in THF (25.0 mL) was added K₂CO₃ (1.56 g, 11.3 mmol). Then Cbz-Cl (960 mg, 5.63 mmol) was added to the mixture at 0°C. Then the mixture was stirred at 25°C for 12 h. The reaction mixture was concentrated under reduced pressure to remove solvent to give a residue. The crude compound was used into the next step without further purification. **239** (2.20 g, 92.7%) was obtained as a red oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.52 - 7.50 (m, 3H), 7.44 - 7.36 (m, 10H), 7.34 (br d, *J* = 6.8 Hz, 1H), 7.31 (d, *J* = 4.4 Hz, 2H), 7.17 (d, *J* = 13.6 Hz, 1H), 5.15 (s, 2H), 4.49 (s, 1H), 3.74 (t, *J* = 5.2 Hz, 2H), 3.34 - 3.31 (m, 2H), 2.80 (s, 3H), 0.89 (s, 9H). LCMS t_R = 0.629 min in 0.8 min chromatography, 5-95AB, LCMS ESI calcd. for C₃₃H₃₇FN₃O₅Si⁺ [M+H]⁺ 602.25, found 602.7.

Benzyl (5-*amino*-4-((2-((*tert-butyldiphenylsilyl*)*oxy*)*ethyl*)(*methyl*)*amino*)-2-*fluorophenyl*)*carbamate* (**244**). To a solution of benzyl (4-((2-((*tert-butyldiphenylsilyl*)*oxy*)*ethyl*)(*methyl*)*amino*)-2-*fluoro*-5-*nitrophenyl*)*carbamate* (**239**) (2.10 g, 3.49 mmol) in EtOH (25.0 mL) was added NH₄Cl (1.12 g, 20.9 mmol) dissolved in H₂O (12.0 mL) and Fe (585 mg, 10.5 mmol) at 25°C. Then the mixture was stirred at 100°C for 12 h. The reaction mixture was filtered and the solid was washed with MeOH (10.0 mL x 3). The residue was purified by flash silica gel chromatography (eluent of 0%~25%, EtOAc/PE) to give the product. **244** (2.03 g, 97.9%) was obtained as a yellow oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.75 - 7.60 (m, 4H), 7.50 - 7.31 (m, 11H), 6.84 (br d, *J* = 7.2 Hz, 1H), 6.77 (d, *J* = 12.0 Hz, 1H), 5.12 (s, 2H), 4.88 (s, 2H), 3.72 (br t, *J* = 5.2 Hz, 2H), 2.97 - 2.92 (m, 2H), 2.56 (s, 3H), 1.17 (t, *J* = 7.2 Hz, 1H), 0.99 (s, 9H). LCMS t_R = 0.565 min in 0.8 min chromatography, 5-95AB, LCMS ESI calcd. for C₃₃H₃₉FN₃O₃Si⁺ [M+H]⁺572.27, found 572.7.

Benzyl (4-((2-((tert-butyldiphenylsilyl)oxy)ethyl)(methyl)amino)-2-fluoro-5-(4H-1,2,4-triazol-4-yl)phenyl)carbamate *(249)*. To solution of benzyl (5-amino-4-((2-((tert-butyldiphenylsilyl)oxy)ethyl)(methyl)amino)-2а fluorophenyl)carbamate (244) (2.03 g, 3.55 mmol) and 1,2-diformylhydrazine (1.56 g, 17.8 mmol) in pyridine (18 mL) was added Et₃N (2.51 g, 24.9 mmol) and Me₃SiCl (5.79 g, 53.3 mmol) at 25°C. Then the reaction mixture was stirred at 100°C for 12 h. The reaction mixture was concentrated under reduced pressure to remove solvent to give a residue. The residue was purified by flash silica gel chromatography (eluent of 0%~5%, MeOH/DCM) to give the product. 249 (1.42 g, 98.6%) was obtained as an off-white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.74 (s, 2H), 7.60 - 7.55 (m, 1H), 7.54 - 7.51 (m, 4H), 7.46 - 7.44 (m, 1H), 7.44 - 7.42 (m, 3H), 7.41 (s, 4H), 7.39 (d, J = 2.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.36 (br d, J = 3.6 Hz, 1H), 7.16 (d, J = 12.4 Hz, 2H), 7.16 (d, J = 3.6 Hz, 1H), 7.16 (d, J = 3 Hz, 1H), 5.15 (s, 2H), 3.54 (br t, J = 5.2 Hz, 2H), 2.82 (br t, J = 5.2 Hz, 2H), 2.53 (s, 3H), 1.17 (s, 1H), 0.93 (s, 9H). LCMS $t_R = 0.588$ min in 0.8 min chromatography, 5-95AB, LCMS ESI calcd. for $C_{35}H_{39}FN_5O_3Si^+$ [M+H]⁺ 624.28, found 624.7.

 N^{l} -(2-((tert-butyldiphenylsilyl)oxy)ethyl)-5-fluoro- N^{l} -methyl-2-(4H-1,2,4-triazol-4-yl)benzene-1,4-diamine (254). To a solution of benzyl (4-((2-((tert-butyldiphenylsilyl)oxy)ethyl)(methyl)amino)-2-fluoro-5-(4H-1,2,4-triazol-4-yl)phenyl)carbamate (249) (800 mg, 1.28 mmol) in MeOH (15 mL) was added Pd/C (800 mg, 10% Pd) under N₂

atmosphere. The suspension was degassed and purged with H₂ for 3 times. The reaction mixture was stirred under H₂ (20 psi) at 25°C for 4 hours. The reaction mixture was filtered, and the filtrate was concentrated under reduced pressure to remove solvent to give a residue. The crude compound was used into the next step without further purification. **254** (600 mg, 96.5%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.54 (d, *J* = 1.6 Hz, 2H), 7.52 (d, *J* = 1.6 Hz, 2H), 7.46 - 7.39 (m, 6H), 7.08 (d, *J* = 12.8 Hz, 1H), 6.74 (d, *J* = 9.2 Hz, 1H), 5.23 (s, 2H), 3.50 (t, *J* = 5.6 Hz, 2H), 3.18 - 3.17 (m, 1H), 3.17 - 3.16 (m, 1H), 2.80 (t, *J* = 5.6 Hz, 2H), 2.41 (s, 3H), 0.95 (s, 9H). LCMS t_R = 0.540 min in 0.8 min chromatography, 5-95AB, LCMS ESI calcd. for C₂₇H₃₃FN₅OSi⁺ [M+H]⁺ 490.24, found 490.6.

5-((4-((2-((tert-butyldiphenylsilyl)oxy)ethyl)(methyl)amino)-2-fluoro-5-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(*cyclopropylamino*)*pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**259**). To a solution of N¹-(2-((tertbutyldiphenylsilyl)oxy)ethyl)-5-fluoro-N¹-methyl-2-(4H-1,2,4-triazol-4-yl)benzene-1,4-diamine (**254**) (600 mg, 1.34 mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-*a*]pyrimidine-3-carbonitrile (**75**) (313 mg, 1.34 mmol) in dioxane (12.0 mL) was added Cs₂CO₃ (1.74 g, 5.35 mmol), BINAP (125 mg, 201 µmol) and Pd(OAc)₂ (45.1 mg, 201 µmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 2 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~44%, EtOAc/PE). **259** (500 mg, 60.1%) was obtained as a yellow oil. LCMS t_R = 0.597 min in 0.8 min chromatography, 5-95AB, LCMS ESI calcd. for C₃₇H₄₀FN₁₀OSi⁺ [M+H]⁺ 687.31, found 687.8.

7-(cyclopropylamino)-5-[2-fluoro-4-[2-hydroxyethyl(methyl)amino]-5-(1,2,4-triazol-4-

yl)anilino]pyrazolo[1,5-a]pyrimidine-3-carbonitrile (61). A mixture of 5-((4-((2-((tert-butyldiphenylsilyl)oxy)ethyl)(methyl)amino)-2-fluoro-5-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(cyclopropylamino)pyrazolo[1,5-*a*]pyrimidine-3-carbonitrile (**259**) (200 mg, 291 μmol) in THF (5 mL) was added TBAF (1.5 mL, 1 mol/L in THF) at 0°C. The reaction mixture was stirred at 25°C for 2 hours. The resulting mixture was quenched by water (10 mL), extracted with EtOAc (20 mL x 3). The mixture was dried over anhydrous Na₂SO₄, filtered and concentrated. The residue was purified by prep-HPLC (column: Phenomenex luna C18 150*25mm*10um; mobile phase: [water (FA)-ACN]; B%: 20%-50%, 8 min). **61** (8.00 mg) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.45 (s, 1H), 8.81 (s, 2H), 8.35 (s, 1H), 8.30 (d, *J* = 1.8 Hz, 1H), 8.16 (d, *J* = 8.3 Hz, 1H), 7.27 (d, *J* = 12.8 Hz, 1H), 6.15 (s, 1H), 4.58 (t, *J* = 5.1 Hz, 1H), 3.37 (q, *J* = 6.0 Hz, 2H), 2.81 (t, *J* = 5.8 Hz, 2H), 2.64 – 2.54 (m, 1H), 2.48 (s, 3H), 0.85 – 0.78 (m, 2H), 0.73 – 0.68 (m, 2H). ¹⁹F NMR (376 MHz, DMSO-*d*₆) δ -121.98. ¹³C NMR (214 MHz, DMSO-*d*₆) δ 160.09, 157.00 (d, *J* = 248.0 Hz), 153.89, 151.66, 148.15, 146.60 (d, *J* = 8.8 Hz), 146.08, 126.26, 125.10, 125.09 (d, *J* = 24.9 Hz), 117.84, 112.17 (d, *J* = 21.8 Hz), 79.65, 79.50, 61.30, 59.44, 44.03, 26.50, 9.68. HPLC t_R = 1.694 min in 4 min chromatography, purity 95.7%. LCMS t_R = 0.384 min in 0.8 min chromatography, 5-95AB, LCMS ESI calcd. for C₂₁H₂₂FN₁₀O⁺ [M+H]⁺449.20, found 449.4.

N-(2-*methoxyethyl*)-4-*nitro*-2-(4H-1,2,4-*triazol*-4-*yl*)*aniline* (**191**). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (1.00 g, 4.80 mmol) and 2-methoxyethan-1-amine (361 mg, 4.80 mmol) in MeCN (10 mL) was added K₂CO₃ (1.99 g, 14.41 mmol) at 25°C. Then the mixture was stirred at 100°C for 10 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~8%, MeOH/DCM). **191** (2.18 g, 54.3%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.73 (s, 2H), 8.20 - 8.17 (m, 1H), 8.01 (d, *J* = 2.8 Hz, 1H), 7.01 (d, *J* = 9.6 Hz, 1H), 6.65 (t, *J* = 5.6 Hz, 1H), 3.45 - 3.43 (m, 2H), 3.40 - 3.36 (m, 2H), 3.24 (s, 3H). LCMS t_R = 0.411 min in 1 min chromatography, MS ESI calcd. for C₁₁H₁₄N₅O₃⁺ [M+H]⁺ 264.11, found 264.0.

 N^{l} -(2-methoxyethyl)-2-(4H-1,2,4-triazol-4-yl)benzene-1,4-diamine (209). To a solution of N-(2-methoxyethyl)-4-nitro-2-(4H-1,2,4-triazol-4-yl)aniline (191) (863 mg, 3.28 mmol) in MeOH (8 mL) was added Pd/C (720 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 hours. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (50 mL). The combined filtrate was concentrated in vacuo. 209 (637 mg, 77.8%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.65 (s, 2H), 6.72 - 6.69 (m, 1H), 6.65 - 6.63 (m, 1H), 6.41 (d, *J* = 2.4 Hz, 1H), 4.74 (s, 2H), 3.92 (t, *J* = 6.0 Hz, 1H), 3.37 - 3.36 (m, 2H), 3.20 (s, 3H), 3.04 - 2.99 (m, 2H).

7-(cyclopropylamino)-5-((4-((2-methoxyethyl)amino)-3-(4H-1,2,4-triazol-4-

yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (62). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (150 mg, 0.64 mmol) and N¹-(2-methoxyethyl)-2-(4H-1,2,4-triazol-4-yl)benzene-1,4-diamine (209) (158 mg, 0.64 mmol) in dioxane (2 mL) was added Cs₂CO₃ (628 mg, 1.93 mmol), BINAP (60 mg, 0.10 mmol) and Pd(OAc)₂ (22 mg, 0.10 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by silica gel chromatography (eluent of 0~5%, MeOH/DCM). The crude product was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 24%-47%, 8min). **62** (79.5 mg, 17.1%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.52 (s, 1H), 8.70 (s, 2H), 8.31 (s, 1H), 8.18 (d, *J* = 1.8 Hz, 1H), 7.61 (dd, *J* = 8.9, 2.5 Hz, 1H), 7.53 (d, *J* = 2.5 Hz, 1H), 6.91 (d, *J* = 8.9 Hz, 1H), 5.88 (s, 1H), 4.73 (t, *J* = 5.8 Hz, 1H), 3.44 (t, *J* = 5.9 Hz, 2H), 3.24 (s, 3H), 3.20 (q, *J* = 5.9 Hz, 2H), 2.62 – 2.54 (m, 1H), 0.83 – 0.76 (m, 2H), 0.73 – 0.66 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.89, 151.05, 148.19, 144.90, 143.42, 139.13, 129.87, 122.71, 119.65, 119.57, 114.84, 112.74, 76.18, 75.69, 70.36, 58.04, 42.63, 23.28, 6.53. HPLC t_R = 3.585 min in 8 min chromatography, purity 99.8%. LCMS t_R = 1.426 min in 4 min chromatography, MS ESI calcd. for C₂₁H₂₃N₁₀O⁺ [M+H]⁺ 431.21, found 431.3.

N-methyl-4-nitro-N-(2-(pyrrolidin-1-yl)ethyl)-2-(4H-1,2,4-triazol-4-yl)aniline (192). To a solution of 4-(2-fluoro-5nitrophenyl)-4H-1,2,4-triazole (**176**) (300 mg, 1.44 mmol) and N-methyl-2-(pyrrolidin-1-yl)ethan-1-amine (185 mg, 1.44 mmol) in MeCN (5 mL) was added K₂CO₃ (598 mg, 4.32 mmol) at 25°C. Then the mixture was stirred at 100°C for 10 hours. Water (150 mL) was added to the reaction mixture. The mixture was extracted with EtOAc (200 mL x2) and the combined organic phase was washed with brine (200 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~21%, MeOH/DCM). **192** (400 mg, 79.7%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.93 (s, 2H), 8.24 - 8.15 (m, 2H), 7.32 (d, *J* = 9.2 Hz, 1H), 3.16 (s, 3H), 2.61 (s, 4H), 2.45 (s, 4H), 1.65 (s, 4H). LCMS t_R = 0.783 min in 2.5 min chromatography, MS ESI calcd. for C₁₅H₂₁N₆O₂⁺ [M+H]⁺ 317.17, found 317.0.

 N^{l} -methyl- N^{l} -(2-(pyrrolidin-1-yl)ethyl)-2-(4H-1,2,4-triazol-4-yl)benzene-1,4-diamine (**210**). To a solution of N-methyl-4-nitro-N-(2-(pyrrolidin-1-yl)ethyl)-2-(4H-1,2,4-triazol-4-yl)aniline (**192**) (400 mg, 1.26 mmol) in MeOH (3 mL) was added Pd/C (220 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (200 mL). The combined filtrate was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~6%, MeOH/DCM). **210** (290 mg, 77.1%) was obtained as a light yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.82 (s, 2H), 7.11 (d, *J* = 8.8 Hz, 1H), 6.64 (dd, *J* = 2.4, 8.8 Hz, 1H), 6.52 (d, *J* = 2.4 Hz, 1H), 5.23 (s, 2H), 3.16 (s, 2H), 2.75 (t, *J* = 6.8 Hz, 2H), 2.36 (s, 3H), 2.33 (s, 4H), 1.63 (s, 4H). LCMS t_R = 0.132 min in 1 min chromatography, MS ESI calcd. for C₁₅H₂₃N₆⁺ [M+H]⁺ 287.20, found 287.2.

7-(cyclopropylamino)-5-((4-(methyl(2-(pyrrolidin-1-yl)ethyl)amino)-3-(4H-1,2,4-triazol-4-

yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (63). To a solution of N¹-methyl-N¹-(2-(pyrrolidin-1-yl)ethyl)-2-(4H-1,2,4-triazol-4-yl)benzene-1,4-diamine (210)(196)mg, 0.68 mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (160 mg, 0.68 mmol) in dioxane (5 mL) was added 'BuOLi (165 mg, 2.05 mmol), and Brettphos Pd G₃ (62 mg, 0.07 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~15%, MeOH/DCM). The crude product was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 30%-62%, 9min). 63 (11.2 mg, 6.0%) was obtained as a light yellow solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.81 (s, 1H), 8.90 (s, 2H), 8.36 (s, 1H), 8.30 (d, J = 1.8 Hz, 1H), 7.89 (d, J = 2.5 Hz, 1H), 7.70 (dd, J = 8.8, 2.6 Hz, 1H), 7.36 (d, J = 8.9 Hz, 1H), 5.96 (s, 1H), 2.82 (t, J = 7.0 Hz, 2H), 2.65 - 2.57 (m, 1H), 2.46 (s, 3H), 2.38 - 2.27 (m, 6H), 1.68 -1.58 (m, 4H), 0.86 - 0.79 (m, 2H), 0.75 - 0.69 (m, 2H). ¹³C NMR (101 MHz, DMSO- d_6) δ 156.54, 150.84, 148.38, 145.01, 142.87, 140.92, 136.27, 128.48, 122.53, 120.13, 116.92, 114.70, 76.66, 76.35, 53.57, 53.40, 52.89, 41.68, 23.32, 23.07, 6.54. HPLC $t_R = 3.061$ min in 8 min chromatography, purity 99.2%. LCMS $t_R = 1.732$ min in 4 min chromatography, MS ESI calcd. for $C_{25}H_{30}N_{11}^+$ [M+H]⁺ 484.27, found 484.5.

N-methyl-N-(2-morpholinoethyl)-4-nitro-2-(4H-1,2,4-triazol-4-yl)aniline (**193**). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (300 mg, 1.30 mmol) and N-methyl-2-morpholinoethan-1-amine (187 mg, 1.30 mmol) in MeCN (5 mL) was added K₂CO₃ (538 mg, 3.89 mmol) at 25°C. Then the mixture was stirred at 100°C for 10 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~5%,

MeOH/DCM). **193** (266 mg, 55.8%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.95 (s, 2H), 8.22 - 8.17 (m, 2H), 7.32 (d, *J* = 9.2 Hz, 1H), 3.45 (t, *J* = 4.4 Hz, 4H), 2.99 (t, *J* = 6.4 Hz, 2H), 2.66 (s, 3H), 2.33 (t, *J* = 6.4 Hz, 2H), 2.26 (s, 4H). LCMS t_R = 0.332 min in 1 min chromatography, MS ESI calcd. for C₁₅H₂₁N₆O₃⁺ [M+H]⁺ 333.17, found 333.1.

 N^{1} -methyl- N^{1} -(2-morpholinoethyl)-2-(4H-1,2,4-triazol-4-yl)benzene-1,4-diamine (211). To a solution of *N*-methyl-N-(2-morpholinoethyl)-4-nitro-2-(4H-1,2,4-triazol-4-yl)aniline (193) (266 mg, 0.80 mmol) in MeOH (12 mL) was added Pd/C (200 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (50 mL). The combined filtrate was concentrated in vacuo. **211** (248 mg, 99.5%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.86 (s, 2H), 7.12 (d, *J* = 8.8 Hz, 1H), 6.65 - 6.53 (m, 2H), 5.23 (s, 2H), 3.52 - 3.45 (m, 4H), 2.77 - 2.74 (m, 2H), 2.36 (s, 3H), 2.22 (s, 4H), 2.14 (t, *J* = 6.4 Hz, 2H). LCMS t_R = 0.132 min in 1 min chromatography, MS ESI calcd. for C₁₅H₂₃N₆O⁺ [M+H]⁺ 303.19, found 303.2.

7-(cyclopropylamino)-5-((4-(methyl(2-morpholinoethyl)amino)-3-(4H-1,2,4-triazol-4-

yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (64). To a solution of N¹-methyl-N¹-(2-morpholinoethyl)-2-(4H-1,2,4-triazol-4-yl)benzene-1,4-diamine (211)(207 mg, 0.69 mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (160 mg, 0.69 mmol) in dioxane (2 mL) was added Cs₂CO₃ (669 mg, 2.05 mmol), BINAP (64 mg, 0.10 mmol) and Pd(OAc)₂ (23 mg, 0.10 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of $0 \sim 7\%$, MeOH/DCM). The crude product was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 21%-50%, 10min). **64** (31.3 mg, 17.1%) was obtained as a white solid. ¹H NMR $(400 \text{ MHz}, \text{DMSO-}d_6) \delta 9.81 \text{ (s, 1H)}, 8.94 \text{ (s, 2H)}, 8.36 \text{ (s, 1H)}, 8.29 \text{ (s, 1H)}, 7.91 \text{ (d, } J = 2.6 \text{ Hz}, 1\text{ H)}, 7.70 \text{ (dd, } J = 8.8, 2.6 \text{ Hz}, 1\text{ H})$ Hz, 1H), 7.36 (d, J = 8.9 Hz, 1H), 5.96 (s, 1H), 3.51 (t, J = 4.6 Hz, 4H), 2.83 (t, J = 6.7 Hz, 2H), 2.65 - 2.57 (m, 1H), 2.46 (s, 3H), 2.31 - 2.18 (m, 6H), 0.85 - 0.78 (m, 2H), 0.74 - 0.69 (m, 2H). ¹³C NMR (101 MHz, DMSO- d_6) δ 156.54, 150.84, 148.38, 144.99, 142.89, 140.89, 136.31, 128.43, 122.63, 120.10, 116.81, 114.70, 76.67, 76.36, 66.15, 55.53, 53.28, 51.35, 41.69, 23.33, 6.55. HPLC t_R = 3.016 min in 8 min chromatography, purity 99.6%. LCMS t_R = 1.665 min in 4 min chromatography, MS ESI calcd. for $C_{25}H_{30}N_{11}O^+$ [M+H]⁺ 500.26, found 500.5.

tert-butyl (R)-(1-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate (194). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (900 mg, 4.32 mmol) and tert-butyl (*R*)-pyrrolidin-3-ylcarbamate (966 mg, 5.19 mmol) in MeCN (12 mL) was added K₂CO₃ (1.79 g, 12.97 mmol) at 25°C. Then the mixture was stirred at 100°C for 10 hours. Water (150 mL) was added to the reaction mixture. The mixture was extracted with EtOAc (180 mL x2) and the combined organic phase was washed with brine (250 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. **194** (1.22 g, 60.1%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.81 (s, 2H), 8.17 (dd, *J* = 2.8, 9.6 Hz, 1H), 8.06 (d, *J* = 2.8 Hz, 1H), 7.18 (d, *J* = 6.4 Hz, 1H), 4.00 (s, 1H), 3.82 (s, 1H), 2.70 - 2.59 (m, 4H), 1.93 (dt, *J* = 5.6, 12.8 Hz, 2H), 1.37 (s, 9H). LCMS t_R = 0.481 min in 1 min chromatography, MS ESI calcd. for C₁₇H₂₃N₆O₄⁺ [M+H]⁺ 375.18, found 375.1.

tert-butyl (R)-(1-(4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate (212). To a solution of tert-butyl (R)-(1-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate (194) (1.2 g, 3.21 mmol) in MeOH (12 mL) was added Pd/C (600 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 hours. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (200 mL). The combined filtrate was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~8%, MeOH/DCM). 212 (385 mg, 33.9%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.74 (s, 2H), 7.04 (d, *J* = 7.2 Hz, 1H), 6.87 (d, *J* = 8.8 Hz, 1H), 6.62 (dd, *J* = 2.4, 8.8 Hz, 1H), 6.48 (d, *J* = 2.4 Hz, 1H), 4.99 (s, 2H), 3.97 - 3.85 (m, 1H), 2.83 (dd, *J* = 6.8, 9.2 Hz, 1H), 2.72 - 2.63 (m, 1H), 2.58 - 2.51 (m, 1H), 2.46 (dd, *J* = 5.2, 8.8 Hz, 1H), 2.00 - 1.89 (m, 1H), 1.59 (qd, *J* = 6.4, 12.8 Hz, 1H), 1.37 (s, 9H). LCMS t_R = 0.404 min in 1 min chromatography, MS ESI calcd. for C₁₇H₂₅N₆O₂+ [M+H]⁺ 345.20, found 345.2.

tert-butyl (*R*)-(1-(4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate (**226**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (190 mg, 0.81 mmol) and tert-butyl (R)-(1-(4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-

yl)carbamate (**212**) (336 mg, 0.96 mmol) in dioxane (5 mL) was added Cs_2CO_3 (795 mg, 2.44 mmol), BINAP (76 mg, 0.12 mmol) and Pd(OAc)₂ (27 mg, 0.12 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~4%, MeOH/DCM). **226** (207 mg, 21.9%) was obtained as a yellow solid. LCMS t_R = 0.524 min in 1 min chromatography, MS ESI calcd. for C₂₇H₃₂N₁₁O₂⁺ [M+H]⁺ 542.27, found 542.2.

(R)-5-((4-(3-aminopyrrolidin-1-yl)-3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(*cyclopropylamino*)*pyrazolo*[*1,5-a*]*pyrimidine-3-carbonitrile* (**65**). To a solution of tert-butyl (R)-(1-(4-((3-cyano-7-(cyclopropylamino))pyrazolo[*1,5-a*]*pyrimidin-5-yl*)*amino*)-2-(4H-1,2,4-triazol-4-yl)*phenyl*)*pyrrolidin-3-yl*)*carbamate* (**26**) (200 mg, 0.37 mmol) in DCM (3 mL) was added TFA (3.10 g, 27.21 mmol) at 25°C. The mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 20%-63%, 10min). Then the impure product was purified by SFC (column: DAICEL CHIRALPAK AD (250mm*30mm, 10um); mobile phase: [0.1%NH₃H₂O ETOH]; B%: 45%-45%, min). **65** (13.6 mg, 17.6%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.62 (s, 1H), 8.76 (s, 2H), 8.32 (s, 1H), 8.22 (br s, 1H), 7.72 – 7.61 (m, 2H), 6.98 (d, *J* = 8.8 Hz, 1H), 5.90 (s, 1H), 3.38 – 3.31 (m, 1H), 2.94 (q, *J* = 7.7 Hz, 1H), 2.86 (dd, *J* = 9.2, 6.0 Hz, 1H), 2.76 (q, *J* = 7.6 Hz, 1H), 2.58 (tt, *J* = 7.1, 3.7 Hz, 1H), 2.42 (dd, *J* = 9.2, 4.9 Hz, 1H), 1.88 (dq, *J* = 16.0, 9.7, 8.2 Hz, 1H), 1.49 (dq, *J* = 12.8, 6.5 Hz, 1H), 0.86 – 0.76 (m, 2H), 0.73 – 0.63 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.72, 150.98, 148.23, 144.92, 144.15, 140.40, 131.62, 121.78, 121.39, 116.50, 114.77, 76.31, 75.88, 58.07, 50.66, 47.88, 34.09, 23.29, 6.52. HPLC t_R = 2.870 min in 8 min chromatography, purity 98.5%. LCMS t_R = 1.597 min in 4 min chromatography, MS ESI calcd. for C₂₂H₂₄N₁₁⁺ [M+H]⁺ 442.22, found 442.4.

tert-butyl (*S*)-(*1*-(*4-nitro-2-*(*4H-1*,2,*4-triazol-4-yl*)*phenyl*)*pyrrolidin-3-yl*)*carbamate* (**195**). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (1 g, 4.80 mmol) and tert-butyl (*S*)-pyrrolidin-3-ylcarbamate (895 mg, 4.80 mmol) in MeCN (12 mL) was added K₂CO₃ (1.99 g, 14.41 mmol) at 25°C. Then the mixture was stirred at 100°C for 10 hours. Water (150 mL) was added to the reaction mixture. The mixture was extracted with EtOAc (180 mL x2) and the combined organic phase was washed with brine (250 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. **195** (1.22 g, 67.8%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.82 (s, 2H), 8.16 (dd, *J* = 2.4, 9.6 Hz, 1H), 8.04 (d, *J* = 2.8 Hz, 1H), 7.18 (d, *J* = 6.4 Hz, 1H), 2.59 - 2.43 (m, 2H), 2.00 - 1.71 (m, 4H), 1.48 (ddd, *J* = 5.2, 7.6, 18.0 Hz, 2H), 1.36 (s, 9H). LCMS t_R = 0.753 min in 1.5 min chromatography, MS ESI calcd. for C₁₇H₂₃N₆O₄⁺ [M+H]⁺ 375.18, found 375.1.

tert-butyl (S)-(1-(4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate (213). To a solution of tert-butyl (S)-(1-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate (195) (1.20 g, 3.21 mmol) in MeOH (12 mL) was added Pd/C (600 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 hours. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (200 mL). The combined filtrate was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~6%, MeOH/DCM). **213** (430 mg, 37.8%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.75 (s, 2H), 7.06 (d, *J* = 7.2 Hz, 1H), 6.87 (d, *J* = 8.8 Hz, 1H), 6.62 (dd, *J* = 2.4, 8.8 Hz, 1H), 6.48 (d, *J* = 2.4 Hz, 1H), 5.00 (s, 2H), 3.96 - 3.85 (m, 1H), 2.83 (dd, *J* = 6.8, 9.2 Hz, 1H), 2.72 - 2.63 (m, 1H), 2.57 - 2.51 (m, 1H), 2.45 (dd, *J* = 5.2, 9.2 Hz, 1H), 2.00 - 1.87 (m, 1H), 1.63 - 1.54 (m, 1H), 1.36 (s, 9H). LCMS t_R = 0.399 min in 1 min chromatography, MS ESI calcd. for C₁₇H₂₅N₆O₂⁺ [M+H]⁺ 345.20, found 345.1.

tert-butyl (S)-(1-(4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate (**227**). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (260 mg, 1.11 mmol) and tert-butyl (S)-(1-(4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate (**213**) (383 mg, 1.11 mmol) in dioxane (5 mL) was added Cs₂CO₃ (1.09 g, 3.34 mmol), BINAP (104 mg, 0.17 mmol) and Pd(OAc)₂ (37 mg, 0.17 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~5%, MeOH/DCM). **227** (440 mg, 67.1%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.66 (s, 1H), 8.79 (s, 2H), 8.33 (s, 1H), 8.25 (s, 1H), 7.70 - 7.61 (m, 2H), 7.12 (d, *J* = 7.1 Hz, 1H), 7.00 (d, *J* = 9.2 Hz, 1H), 5.90 (s, 1H), 4.03 - 3.89 (m, 1H), 2.96 (dd, *J* = 6.8, 9.2 Hz, 1H), 2.90 - 2.83 (m, 1H), 2.76 - 2.69 (m, 1H), 2.62 - 2.54 (m, 2H), 1.94 (td, *J* = 6.0, 12.4 Hz, 1H), 1.66 (qd, *J* = 6.4, 12.4 Hz, 1H), 1.37 (s, 9H), 0.84 - 0.77

(m, 2H), 0.73 - 0.67 (m, 2H). LCMS $t_R = 0.511$ min in 1 min chromatography, MS ESI calcd. for $C_{27}H_{32}N_{11}O_2^+$ [M+H]⁺ 542.27, found 542.2.

(S)-5-((4-(3-aminopyrrolidin-1-yl)-3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(*cyclopropylamino*)*pyrazolo*[*1*,5-*a*]*pyrimidine-3-carbonitrile* (*66*). To a solution of tert-butyl (S)-(1-(4-((3-cyano-7-(cyclopropylamino)pyrazolo[*1*,5-*a*]*pyrimidin-5-yl*)*amino*)-2-(4H-1,2,4-triazol-4-yl)*phenyl*)*pyrrolidin-3-yl*)*carbamate* (*27*) (400 mg, 0.74 mmol) in DCM (3 mL) was added TFA (6.21 g, 54.43 mmol) at 25°C. The mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 20%-50%, 10min). *66* (101.7 mg, 35.5%) was obtained as a gray solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.64 (s, 1H), 8.78 (s, 2H), 8.32 (s, 1H), 8.22 (s, 1H), 7.74 – 7.58 (m, 2H), 7.00 (d, *J* = 8.9 Hz, 1H), 5.91 (s, 1H), 3.42 (p, *J* = 5.6 Hz, 1H), 3.01 – 2.85 (m, 2H), 2.76 (q, *J* = 7.6 Hz, 1H), 2.58 (tt, *J* = 7.0, 3.6 Hz, 1H), 2.48 – 2.45 (m, 1H), 1.93 (dq, *J* = 13.2, 6.8 Hz, 1H), 1.55 (dq, *J* = 12.7, 6.5 Hz, 1H), 0.86 – 0.76 (m, 2H), 0.74 – 0.66 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.70, 150.96, 148.24, 144.92, 144.06, 140.06, 131.97, 121.70, 121.68, 119.80, 116.75, 114.76, 76.34, 75.94, 57.20, 50.36, 33.19, 23.29, 6.52. HPLC t_R = 2.874 min in 8 min chromatography, purity 96.6%. LCMS t_R = 1.570 min in 4 min chromatography, MS ESI calcd. for C₂₂H₂₄N₁₁⁺ [M+H]⁺ 442.22, found 442.4.

tert-butyl (*S*)-(*1*-(*4-amino-5-fluoro-2-nitrophenyl*)*pyrrolidin-3-yl*)*carbamate* (**235**). To a solution of 2,4-difluoro-5nitroaniline (**231**) (2.00 g, 11.49 mmol) and tert-butyl (*S*)-pyrrolidin-3-ylcarbamate (2.57 g, 13.78 mmol) in MeCN (30 mL) was added K₂CO₃ (4.76 g, 34.46 mmol). Then the reaction mixture was stirred at 60°C for 12 hours. The reaction mixture was concentrated directly. The residue was purified by flash silica gel chromatography (eluent of 0~26%, EtOAc/PE). **235** (2.5 g, 59.47%) was obtained as a red solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.29-7.23 (m, 1H), 7.15 (br d, *J* = 5.6 Hz, 1H), 6.86-6.76 (m, 1H), 4.95 (s, 2H), 3.32 (s, 1H), 3.26-3.18 (m, 1H), 3.16-3.07 (m, 2H), 2.81-2.74 (m, 1H), 2.09-1.99 (m, 1H), 1.90-1.79 (m, 1H), 1.37 (s, 9H). LCMS t_R = 1.274 min in 2 min chromatography, MS ESI calcd. for C₁₅H₂₂FN₄O₄+ [M+H]⁺ 341.15, found 340.9.

tert-butyl (*S*)-(*1*-(*4*-(((*benzyloxy*)*carbonyl*)*amino*)-5-*fluoro*-2-*nitrophenyl*)*pyrrolidin*-3-*yl*)*carbamate* (**240**). To a solution of tert-butyl (S)-(1-(4-amino-5-fluoro-2-nitrophenyl)pyrrolidin-3-yl)carbamate (**235**) (2.5 g, 7.35 mmol) in THF (30 mL) was added K₂CO₃ (3.05 g, 22.04 mmol). Then CbzCl (1.88 g, 11.02 mmol) was added to the mixture at 0°C. The mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated directly. The reaction mixture was concentrated. Saturated aqueous NH₄Cl (15 mL) solution was added. The resulting mixture was extracted with EtOAc (50 mL x 3). The combined organic phase was washed with brine (15 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. The residue was purified by flash silica gel chromatography (eluent of 0~31%, EtOAc/PE). **240** (3.4 g, 86.0%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.99 (s, 1H), 7.50-7.31 (m, 5H), 7.26-7.21 (m, 1H), 6.98-6.87 (m, 1H), 5.14 (s, 2H), 4.12-4.00 (m, 1H), 3.34-3.29 (m, 1H), 3.28-3.16 (m, 2H), 2.88-2.77 (m, 1H), 2.15-2.02 (m, 1H), 1.96-1.83 (m, 1H), 1.37 (s, 9H). LCMS t_R = 1.581 min in 2 min chromatography, MS ESI calcd. for C₂₃H₂₈FN₄O₆⁺ [M+H]⁺ 475.20, found 474.9.

tert-butyl (S)-(1-(2-amino-4-(((benzyloxy)carbonyl)amino)-5-fluorophenyl)pyrrolidin-3-yl)carbamate (**245**). To a solution of tert-butyl (S)-(1-(4-(((benzyloxy)carbonyl)amino)-5-fluoro-2-nitrophenyl)pyrrolidin-3-yl)carbamate (**240**) (3.40 g, 7.17 mmol) in EtOH (30 mL) was added NH₄Cl (1.15 g, 21.50 mmol) dissolved in H₂O (6 mL) and Fe (2.40 g, 42.99 mmol) at 25°C. Then the mixture was stirred at 80°C under N₂ atmosphere for 12 hours. The reaction mixture was filtered and the solid was washed with MeOH (150 mL x 2). The residue was purified by flash silica gel chromatography (eluent of 0%~37%, EtOAc/PE) to give the product. **245** (1.9 g, 51.3%) was obtained as a purple solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.99 (s, 1H), 7.45-7.28 (m, 5H), 7.22-7.15 (m, 1H), 6.83-6.73 (m, 1H), 6.68-6.61 (m, 1H), 5.10 (s, 2H), 4.68 (s, 2H), 4.09-4.04 (m, 1H), 3.15-2.99 (m, 2H), 2.84-2.69 (m, 2H), 2.23-2.09 (m, 1H), 1.70-1.61 (m, 1H), 1.39 (s, 9H). LCMS t_R = 0.721 min in 1 min chromatography, MS ESI calcd. for C₂₃H₃₀FN₄O₄⁺ [M+H]⁺ 445.22, found 445.1.

tert-butyl (*S*)-(1-(4-(((*benzyloxy*)*carbonyl*)*amino*)-5-*fluoro*-2-(4H-1,2,4-*triazo*l-4-*yl*)*phenyl*)*pyrrolidin*-3-*yl*)*carbamate* (**250**). To a solution of tert-butyl (S)-(1-(2-amino-4-(((benzyloxy)carbonyl)amino)-5-fluorophenyl)pyrrolidin-3-*yl*)*carbamate* (**245**) (1.00 g, 2.25 mmol) and 1,2-diformylhydrazine (991 mg, 11.25 mmol) in pyridine (5 mL) was added Et₃N (1.59 g, 15.75 mmol). The mixture was stirred at 100°C for 30 min, then Me₃SiCl (3.67 g, 33.75 mmol, 4.28 mL) was added to the mixture. The mixture was stirred at 100°C for 12 hours. Water (5 mL) was added to the resulting mixture was extracted with EtOAc (20 mL x 3). The combined organic phase was washed with brine (5 mL), dried over
anhydrous Na₂SO₄, filtered and concentrated. The residue was purified by flash silica gel chromatography (eluent of 0%~86%, EtOAc/PE) to give the product. **250** (210 mg, 14.4%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.74 (s, 2H), 7.43-7.28 (m, 6H), 7.12-7.06 (m, 1H), 6.83-6.73 (m, 1H), 5.11 (s, 2H), 4.00-3.89 (m, 1H), 2.98-2.86 (m, 2H), 2.81-2.73 (m, 1H), 2.57-2.52 (m, 1H), 1.73-1.62 (m, 1H), 1.36 (s, 9H). LCMS t_R = 0.676 min in 1 min chromatography, MS ESI calcd. for C₂₅H₃₀FN₆O₄⁺ [M+H]⁺ 497.23, found 441.1.

tert-butyl (*S*)-(*1*-(*4-amino-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate* (**255**). To a solution of tert-butyl (S)-(1-(4-(((benzyloxy)carbonyl)amino)-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate (**250**) (210 mg, 422.93 µmol) in MeOH (5 mL) was added Pd/C (0.1 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. The reaction mixture was stirred under H₂ (15 Psi) at 25°C for 2 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (100 mL). The combine filtrate was concentrated in vacuo. **255** (100 mg, 46.9%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.72 (s, 2H), 7.08-7.00 (m, 1H), 6.83-6.76 (m, 1H), 6.70-6.62 (m, 1H), 4.94 (s, 2H), 3.95-3.85 (m, 1H), 2.88-2.79 (m, 1H), 2.78-2.68 (m, 1H), 2.63-2.54 (m, 1H), 2.46-2.39 (m, 1H), 1.98-1.88 (m, 1H), 1.64-1.55 (m, 1H), 1.36 (s, 9H). LCMS t_R = 0.452 min in 1 min chromatography, MS ESI calcd. for C₁₇H₂₄FN₆O₂⁺ [M+H]⁺ 363.19, found 363.0.

(S)-(1-(4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-5-fluoro-2-(4H-1,2,4tert-butyl triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate (260). To a solution of tert-butyl (S)-(1-(4-amino-5-fluoro-2-(4H-1,2,4triazol-4-yl)phenyl)pyrrolidin-3-yl)carbamate (255) (100)mg, 275.94 µmol) 5-chloro-7and (cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (52 mg, 220.75 µmol) in dioxane (3 mL) was added Cs₂CO₃ (270 mg, 827.81 µmol), BINAP (26.0 mg, 41.39 µmol) and Pd(OAc)₂ (10.0 mg, 41.39 µmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~76%, EtOAc/PE). 260 (85 mg, 15.9%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-d₆) δ 7.97-7.95 (m, 2H), 7.90 (s, 2H), 7.65 (s, 3H), 5.76 (s, 2H), 3.98-3.87 (m, 1H), 3.02-2.91 (m, 2H), 2.85-2.74 (m, 2H), 2.69-2.64 (m, 1H), 1.92-1.88 (m, 2H), 1.37-1.36 (m, 9H), 0.92-0.57 (m, 4H). LCMS $t_{R} = 0.559$ min in 1 min chromatography, MS ESI calcd. for $C_{22}H_{23}FN_{11}^{+}$ [M+2H-Boc]⁺ 460.21, found 460.2.

(S)-5-((4-(3-aminopyrrolidin-1-yl)-2-fluoro-5-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(*cyclopropylamino*)*pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**67**). A mixture of tert-butyl (S)-(1-(4-((3-cyano-7-(cyclopropylamino))pyrazolo[1,5-*a*]*pyrimidin-5-yl*)*amino*)-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)pyrrolidin-3-

yl)carbamate (**260**) (85.0 mg, 151.90 µmol) in DCM (5 mL) was added TFA (1 mL) at 25°C. The reaction mixture was stirred at 25°C for 1 hour. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water(FA)-ACN]; B%: 0%-34%, 25min). **67** (4.80 mg, 6.12%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.31 (s, 1H), 8.76 (s, 2H), 8.32 (s, 1H), 8.24 (s, 1H), 7.81 (d, *J* = 8.3 Hz, 1H), 6.93 (d, *J* = 13.5 Hz, 1H), 6.03 (s, 1H), 3.67 – 3.58 (m, 1H), 3.04 – 2.95 (m, 2H), 2.88 (q, *J* = 8.6, 7.9 Hz, 1H), 2.64 (dd, *J* = 10.3, 4.2 Hz, 1H), 2.60 – 2.54 (m, 1H), 2.10 – 1.99 (m, 1H), 1.79 – 1.63 (m, 1H), 0.85 – 0.74 (m, 2H), 0.69 (p, *J* = 4.8, 4.4 Hz, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 159.15 (d, *J* = 239.8 Hz), 157.38, 150.89, 148.42, 145.01, 144.47, 125.61 (d, *J* = 3.2 Hz), 116.77, 114.82, 105.61, 105.09, 103.60 (d, *J* = 24.9 Hz), 76.20, 75.84, 54.44, 49.63, 47.39, 30.63, 23.33, 6.53. HPLC t_R = 1.924 min in 8 min chromatography, purity 89.4%. LCMS t_R = 1.030 min in 4 min chromatography, MS ESI calcd. for C₂₂H₂₃FN₁₁⁺ [M+H]⁺ 460.21, found 460.4.

tert-butyl (S)-(1-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)carbamate (196). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (176) (200 mg, 0.96 mmol) and tert-butyl (*S*)-piperidin-3-ylcarbamate (289 mg, 1.44 mmol) in MeCN (5 mL) was added K₂CO₃ (531 mg, 3.84 mmol) at 25°C. The mixture was stirred at 100°C for 12 h under N₂ atmosphere. The reaction mixture was concentrated in vacuo. Water (40 mL) was added to the residue. The resulting mixture was extracted with DCM (30 mL x3). The combined organic phase was dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. **196** (498 mg, crude) was obtained as a black brown oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.96 (s, 1H), 8.26-8.22 (m, 1H), 7.35 (d, *J* = 9.0 Hz, 1H), 6.97-6.86 (m, 1H), 6.69-6.62 (m, 1H), 5.76 (s, 1H), 3.39-3.38 (m, 1H), 2.90-2.82 (m, 2H), 2.76-2.68 (m, 2H), 1.78-1.71 (m, 2H), 1.59-1.52 (m, 2H), 1.38 (s, 9H). LCMS t_R = 1.293 min in 2.5 min chromatography, MS ESI calcd. for C₁₈H₂₅N₆O₄⁺ [M+H]⁺ 389.19, found 389.0.

tert-butyl (*S*)-(*1*-(*4-amino-2-*(*4H-1,2,4-triazol-4-yl*)*phenyl*)*piperidin-3-yl*)*carbamate* (**214**). To a solution of tert-butyl (S)-(1-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)*carbamate* (**196**) (214 mg, 0.55 mmol) in MeOH (5 mL) was

added Pd/C (0.11 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (10 mL x3). The combined filtrate was concentrated in vacuo. **214** (198 mg, 79.5%) was obtained as a black brown oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.72 (s, 1H), 7.01 (br d, *J* = 8.6 Hz, 1H), 6.75-6.68 (m, 2H), 6.65-6.60 (m, 1H), 6.55-6.52 (m, 1H), 5.23 (s, 1H), 3.51-3.46 (m, 1H), 3.16 (s, 1H), 2.96-2.84 (m, 2H), 2.79-2.73 (m, 2H), 1.73-1.68 (m, 2H), 1.61-1.53 (m, 2H), 1.37 (s, 9H). LCMS t_R = 1.057 min in 2.5 min chromatography, MS ESI calcd. for C₁₈H₂₇N₆O₂⁺ [M+H]⁺ 359.22, found 359.2.

(S)-(1-(4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4tert-butyl yl)phenyl)piperidin-3-yl)carbamate (228). To a solution of tert-butyl (S)-(1-(4-amino-2-(4H-1,2,4-triazol-4-0.53 vl)phenvl)piperidin-3-vl)carbamate (214)(190)mg, mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (74 mg, 0.32 mmol) in dioxane (5 mL) was added Cs₂CO₃ (518 mg, 1.59 mmol) and Brettphos Pd G₃ (74 mg, 0.08 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~1%, MeOH/DCM). 228 (45 mg, 11.1%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.84-8.81 (m, 1H), 8.76 - 8.70 (m, 1H), 8.39-8.35 (m, 1H), 7.86-7.83 (m, 1H), 7.58-7.52 (m, 1H), 7.30-7.21 (m, 1H), 7.07-7.01 (m, 1H), 6.84-6.77 m, 1H), 5.75 (s, 1H), 3.57 (s, 1H), 3.29-3.26 (m, 2H), 2.98-2.90 (m, 2H), 2.83-2.76 (m, 2H), 1.78-1.74 (m, 2H), 1.67-1.62 (m, 2H), 1.38 (s, 9H), 0.99-0.89 (m, 2H), 0.87-0.82 (m, 2H). LCMS $t_R = 0.93$ min in 1.5 min chromatography, MS ESI calcd. for $C_{28}H_{34}N_{11}O_2^+$ [M+H]⁺ 556.29, found 556.3.

(S)-5-((4-(3-aminopiperidin-1-yl)-3-(4H-1,2,4-triazol-4-yl)phenyl)amino)-7-

(*cyclopropylamino*)*pyrazolo*[1,5-*a*]*pyrimidine-3-carbonitrile* (**68**). To a solution of tert-butyl (S)-(1-(4-((3-cyano-7-(cyclopropylamino))pyrazolo[1,5-*a*]*pyrimidin-5-yl*)*amino*)-2-(4H-1,2,4-triazol-4-yl)*phenyl*)*piperidin-3-yl*)*carbamate* (**228**) (38 mg, 0.07 mmol) in DCM (4 mL) was added TFA (1 mL) at 25°C. The mixture was stirred at 25°C for 2 h. Then the mixture was degassed and purged with N₂. The reaction mixture was concentrated in vacuo. The residue was purified by prep. HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 18%-58%, 11min). **68** (11.2 mg, 35.6%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.82 (s, 1H), 8.84 (s, 2H), 8.37 (s, 1H), 8.30 (br s, 1H), 7.93 (d, *J* = 2.5 Hz, 1H), 7.69 (dd, *J* = 8.8, 2.6 Hz, 1H), 7.27 (d, *J* = 8.8 Hz, 1H), 5.97 (s, 1H), 2.81 (dd, *J* = 10.9, 3.8 Hz, 1H), 2.70 – 2.55 (m, 3H), 2.41 (td, *J* = 11.1, 2.8 Hz, 1H), 2.26 (dd, *J* = 10.9, 8.9 Hz, 1H), 1.81 – 1.70 (m, 1H), 1.61 – 1.51 (m, 1H), 1.43 – 1.28 (m, 1H), 1.07 – 0.94 (m, 1H), 0.91 – 0.77 (m, 2H), 0.77 – 0.65 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.55, 150.87, 148.41, 145.04, 142.76, 141.44, 136.39, 128.13, 121.75, 120.27, 116.97, 114.75, 76.68, 76.38, 60.42, 51.87, 47.90, 33.25, 23.82, 23.34, 6.57. HPLC t_R = 3.237 min in 8 min chromatography, purity 96.9%. LCMS t_R = 1.744 min in 4 min chromatography, MS ESI calcd. for C₂₃H₂₆N₁₁+ [M+H]+ 456.24, found 456.4.

tert-butyl (S)-methyl(1-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)carbamate (197). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (300 mg, 1.44 mmol) and tert-butyl (*S*)-methyl(piperidin-3-yl)carbamate (340 mg, 1.59 mmol) in MeCN (2 mL) was added K₂CO₃ (598 mg, 4.32 mmol) at 25°C. Then the mixture was stirred at 100°C for 10 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~17%, MeOH/DCM). **197** (162 mg, 12.5%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.96 (s, 1H), 8.29 - 8.25 (m, 1H), 8.01 (d, *J* = 9.6 Hz, 2H), 7.43 (d, *J* = 9.6 Hz, 1H), 3.63 - 3.58 (m, 2H), 3.14 - 3.11 (m, 1H), 2.92 - 2.74 (m, 2H), 2.70 - 2.65 (m, 3H), 1.76 - 1.74 (m, 4H), 1.42 (s, 9H). LCMS t_R = 0.514 min in 1 min chromatography, MS ESI calcd. for C₁₉H₂₇N₆O₄⁺ [M+H]⁺ 403.21, found 403.2.

tert-butyl (S)-(1-(4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)(methyl)carbamate (215). To a solution of tertbutyl (S)-methyl(1-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)carbamate (197) (977 mg, 2.43 mmol) in MeOH (10 mL) was added Pd/C (600 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 hours. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (8 mL x2). The combined filtrate was concentrated in vacuo. **215** (803 mg, 76.8%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.70 (s, 2H), 7.08 (d, *J* = 8.4 Hz, 1H), 6.65 - 6.62 (m, 1H), 6.54 (d, *J* = 2.4 Hz, 1H), 5.24 (s, 2H), 4.13 (br s, 1H), 3.73 - 3.72 (m, 1H), 3.17 (s, 2H), 2.66 - 2.64 (m, 4H), 1.59 (br d, *J* = 8.4 Hz, 2H), 1.40 (d, *J* = 2.0 Hz, 2H), 1.39 (s, 9H). LCMS t_R = 1.125 min in 2.5 min chromatography, MS ESI calcd. for C₁₉H₂₉N₆O₂⁺ [M+H]⁺ 373.23, found 373.1.

tert-butyl (S)-(1-(4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4*yl)phenyl)piperidin-3-yl)(methyl)carbamate (229)*. То solution of 5-chloro-7а (cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (330 mg, 1.41 mmol) and tert-butyl (S)-(1-(4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)(methyl)carbamate (215) (368 mg, 0.99 mmol) in dioxane (5 mL) was added Cs₂CO₃ (1.38g, 4.24 mmol) and Brettphos Pd G₃ (128 mg, 0.14 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~4%, MeOH/DCM). 229 (669 mg, 20.7%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.83 (s, 1H), 8.81 (s, 2H), 8.36 - 8.35 (m, 1H), 8.30 (s, 1H), 7.89 (d, J = 2.4 Hz, 1H), 7.75 – 7.73 (m, 1H), 7.34 (d, J = 8.8 Hz, 1H), 5.96 (s, 1H), 4.10 (br d, J = 4.8 Hz, 1H), 3.34 (s, 2H), 2.75-2.63 (m, 4H), 2.61 - 2.58 (m, 2H), 1.62 (br s, 2H), 1.52 - 1.48 (m, 2H), 1.39 (s, 9H), 0.82 - 0.79 (m, 2H), 0.78 - 0.72 (m, 2H). LCMS $t_R = 1.528$ min in 2.5 min chromatography, MS ESI calcd. for $C_{29}H_{36}N_{11}O_2^+$ [M+H]⁺ 570.30, found 570.2.

(S)-7-(cyclopropylamino)-5-((4-(3-(methylamino)piperidin-1-yl)-3-(4H-1,2,4-triazol-4-

yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (69). To a solution of tert-butyl (S)-(1-(4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-*a*]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-

yl)(methyl)carbamate (**229**) (610 mg, 1.07 mmol) in DCM (6 mL) was added TFA (1.85 g, 16.21 mmol) at 25°C. The mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Xtimate C18 150*40mm*10um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 5%-55%, 10min). **69** (75.8 mg, 14.3%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.81 (s, 1H), 8.84 (s, 2H), 8.36 (s, 1H), 8.29 (s, 1H), 7.92 (d, *J* = 2.5 Hz, 1H), 7.69 (dd, *J* = 8.8, 2.6 Hz, 1H), 7.26 (d, *J* = 8.8 Hz, 1H), 5.96 (s, 1H), 2.94 – 2.81 (m, 1H), 2.71 – 2.64 (m, 1H), 2.61 (tt, *J* = 6.9, 3.6 Hz, 1H), 2.48 – 2.42 (m, 1H), 2.38 – 2.28 (m, 1H), 2.19 (dd, *J* = 11.0, 9.1 Hz, 1H), 2.13 (s, 3H), 1.83 – 1.71 (m, 1H), 1.61 – 1.53 (m, 1H), 1.45 – 1.32 (m, 1H), 1.07 – 0.94 (m, 1H), 0.86 – 0.78 (m, 2H), 0.74 – 0.67 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.55, 150.86, 148.40, 145.01, 142.76, 141.52, 136.28, 127.98, 121.54, 120.25, 117.02, 114.73, 76.67, 76.37, 57.11, 56.01, 51.86, 33.37, 30.25, 23.78, 23.34, 6.56. HPLC t_R = 3.252 min in 8 min chromatography, purity 95.4%. LCMS t_R = 1.214 min in 4 min chromatography, MS ESI calcd. for C₂₄H₂₈N₁₁⁺ [M+H]⁺ 470.25, found 470.4.

tert-butyl (*R*)-methyl(1-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)carbamate (**198**). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (300 mg, 1.44 mmol) and tert-butyl (*R*)-methyl(piperidin-3-yl)carbamate (340 mg, 1.59 mmol) in MeCN (2 mL) was added K₂CO₃ (598 mg, 4.32 mmol) at 25°C. Then the mixture was stirred at 100°C for 10 hours. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~12%, MeOH/DCM). **198** (367 mg, 42.0%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.94 (s, 2H), 8.26 - 8.25 (m, 2H), 7.42 - 7.40 (m, 1H), 3.17 (d, *J* = 4.4 Hz, 1H), 2.93 - 2.88 (m, 2H), 2.73 (d, *J* = 2.4 Hz, 2H), 2.68 (br s, 3H), 1.77 - 1.74 (m, 2H), 1.65 - 1.59 (m, 2H), 1.40 (s, 9H). LCMS t_R = 0.514 min in 1 min chromatography, MS ESI calcd. for C₁₉H₂₇N₆O₄⁺ [M+H]⁺ 403.21, found 403.1.

tert-butyl (*R*)-(*1*-(*4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)(methyl)carbamate* (**216**). To a solution of tert-butyl (R)-methyl(1-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)carbamate (**198**) (360 mg, 0.89 mmol) in MeOH (3 mL) was added Pd/C (150 mg, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 hours. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (8 mL x2). The combined filtrate was concentrated in vacuo. **216** (274 mg, 41.5%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.69 (s, 2H), 7.99 (d, *J* = 10.0 Hz, 1H), 7.08 (d, *J* = 8.4 Hz, 1H), 6.54 (d, *J* = 2.4 Hz, 1H), 5.25 (s, 2H), 3.81 - 3.64 (m, 2H), 2.73 - 2.71 (m, 3H), 2.66 (br s, 3H), 1.73 (br d, *J* = 8.0 Hz, 2H), 1.59 (br d, *J* = 9.6 Hz, 2H), 1.37 (s, 9H). LCMS t_R = 1.182 min in 2.5 min chromatography, ESI calcd. for C₁₉H₂₉N₆O₂⁺ [M+H]⁺ 373.23, found 373.2.

tert-butyl (R)-(1-(4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)(methyl)carbamate (230). To a solution of 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (80 mg, 0.34 mmol) and tert-butyl (R)-(1-(4-amino-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)(methyl)carbamate (216) (115 mg, 0.31 mmol) in dioxane (5 mL) was added Cs₂CO₃ (335 mg, 1.03 mmol) and Brettphos Pd G₃ (31 mg, 0.03 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in

vacuo. The residue was purified by flash silica gel chromatography (eluent of $0 \sim 5\%$, MeOH/DCM). **230** (189 mg, 29.4%) was obtained as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.82 (s, 1H), 8.80 (s, 2H), 8.36 - 8.35 (m, 1H), 8.29 (s, 1H), 7.88 (d, *J* = 2.4 Hz, 1H), 7.76 - 7.73 (m, 1H), 7.35 (d, *J* = 8.8 Hz, 1H), 5.96 (s, 1H), 3.85 - 3.76 (m, 1H), 2.69 - 2.67 (m, 5H), 2.61 - 2.59 (m, 2H), 2.46 (br s, 1H), 1.62 (br s, 2H), 1.52 - 1.49 (m, 1H), 1.40 (s, 9H), 1.23 (s, 1H), 0.82 - 0.80 (m, 2H), 0.74 - 0. 72 (m, 2H). LCMS t_R = 1.552 min in 2.5 min chromatography, MS ESI calcd. for C₂₉H₃₆N₁₁O₂⁺ [M+H]⁺ 570.30, found 570.3.

(R)-7-(cyclopropylamino)-5-((4-(3-(methylamino)piperidin-1-yl)-3-(4H-1,2,4-triazol-4-

yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**70**). To a solution of tert-butyl (R)-(1-(4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-*a*]pyrimidin-5-yl)amino)-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-

yl)(methyl)carbamate (**230**) (160 mg, 0.28 mmol) in DCM (3 mL) was added TFA (924 mg, 0.01 mmol) at 25°C. The mixture was stirred at 25°C for 2 hours. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Phenomenex C18 75*30mm*3um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 5%-65%, 12min). **70** (22.5 mg, 16.5%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.81 (s, 1H), 8.84 (s, 2H), 8.36 (s, 1H), 8.29 (br s, 1H), 7.91 (d, *J* = 2.5 Hz, 1H), 7.69 (dd, *J* = 8.8, 2.6 Hz, 1H), 7.26 (d, *J* = 8.8 Hz, 1H), 5.96 (s, 1H), 2.88 (dd, *J* = 11.1, 3.6 Hz, 1H), 2.72 – 2.64 (m, 1H), 2.61 (tt, *J* = 6.9, 3.6 Hz, 1H), 2.48 – 2.41 (m, 1H), 2.33 (tt, *J* = 9.2, 3.7 Hz, 1H), 2.18 (dd, *J* = 10.9, 9.0 Hz, 1H), 2.13 (s, 3H), 1.86 – 1.68 (m, 1H), 1.63 – 1.50 (m, 1H), 1.45 – 1.31 (m, 1H), 1.09 – 0.92 (m, 1H), 0.86 – 0.75 (m, 2H), 0.75 – 0.67 (m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 156.54, 150.86, 148.39, 145.00, 142.75, 141.53, 136.27, 127.98, 121.54, 120.25, 117.02, 114.73, 76.66, 76.36, 57.15, 56.02, 51.86, 33.39, 30.28, 23.79, 23.33, 6.56. HPLC t_R = 3.310 min in 8 min chromatography, purity 96.9%. LCMS t_R = 1.244 min in 4 min chromatography, MS ESI calcd. for C₂₄H₂₈N₁₁⁺ [M+H]⁺ 470.25, found 470.4.

tert-butyl (*R*)-(*1*-(*4-amino-5-fluoro-2-nitrophenyl*)*piperidin-3-yl*)(*methyl*)*carbamate* (**236**). To a solution of 2,4difluoro-5-nitroaniline (**231**) (2.00 g, 11.49 mmol) and tert-butyl (*R*)-methyl(piperidin-3-yl)carbamate (2.71 g, 12.64 mmol) in MeCN (20 mL) was added K₂CO₃ (4.76 g, 34.46 mmol). Then the reaction mixture was stirred at 100°C for 12 hours. The reaction mixture was concentrated directly. The residue was purified by flash silica gel chromatography (eluent of 0~18%, EtOAc/PE). **236** (3.70 g, 85.7%) was obtained as a red solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.26-7.12 (m, 2H), 5.46 (s, 2H), 4.03-3.84 (m, 1H), 2.94-2.79 (m, 2H), 2.75 (s, 1H), 2.71 (s, 3H), 2.65-2.52 (m, 1H), 1.78-1.67 (m, 2H), 1.60-1.49 (m, 2H), 1.39 (s, 9H). LCMS t_R = 0.763 min in 1 min chromatography, MS ESI calcd. for C₁₇H₂₆FN₄O₄⁺ [M+H]⁺ 369.19, found 369.1.

tert-butyl (*R*)-(1-(4-(((benzyloxy)carbonyl)amino)-5-fluoro-2-nitrophenyl)piperidin-3-yl)(methyl)carbamate (**241**). To a solution of tert-butyl (R)-(1-(4-amino-5-fluoro-2-nitrophenyl)piperidin-3-yl)carbamate (**236**) (3.7 g, 10.04 mmol) in THF (30 mL) was added K₂CO₃ (4.16 g, 30.13 mmol). Then CbzCl (2.57 g, 15.07 mmol) was added to the mixture at 0°C. The mixture was stirred at 25°C for 12 hours. The reaction mixture was concentrated directly. The reaction mixture was concentrated. Saturated NH₄Cl (100 mL) solution was added. The resulting mixture was extracted with EtOAc (100 mL x3). The combined organic phase was washed with water (30 mL), brine (30 mL), dried over anhydrous Na₂SO₄, filtered and concentrated. The residue was purified by flash silica gel chromatography (eluent of 0~20%, EtOAc/PE). **241** (5.14 g, 99.9%) was obtained as a red oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.45-7.21 (m, 8H), 5.16 (s, 2H), 4.05-3.86 (m, 1H), 3.11-3.04 (m, 1H), 3.03-2.97 (m, 1H), 2.94-2.85 (m, 1H), 2.72 (s, 4H), 1.78-1.70 (m, 2H), 1.65-1.56 (m, 2H), 1.40 (s, 9H). LCMS t_R = 0.830 min in 1 min chromatography, MS ESI calcd. for C₂₅H₃₂FN₄O₆⁺ [M+H]⁺ 503.23, found 503.2.

tert-butyl (R)-(1-(2-amino-4-(((benzyloxy)carbonyl)amino)-5-fluorophenyl)piperidin-3-yl)(methyl)carbamate (246). To a solution of tert-butyl (R)-(1-(4-(((benzyloxy)carbonyl)amino)-5-fluoro-2-nitrophenyl)piperidin-3-yl)carbamate (241) (5.14 g, 10.23 mmol) in EtOH (30 mL) was added NH₄Cl (3.28 g, 61.37 mmol) dissolved in H₂O (15 mL) and Fe (1.71 g, 30.68 mmol) at 25°C. Then the mixture was stirred at 100°C under N₂ atmosphere for 12 hours. The reaction mixture was filtered and the solid was washed with MeOH (50 mL x 3). The residue was purified by flash silica gel chromatography (eluent of 0%~11%, EtOAc/PE) to give the product. 246 (2.92 g, 48.1%) was obtained as a black brown oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.43-7.30 (m, 7H), 6.89-6.82 (m, 1H), 6.77 (d, *J* = 12.4 Hz, 1H), 5.11 (s, 2H), 4.64 (s, 2H), 4.08-3.89 (m, 1H), 2.99-2.92 (m, 1H), 2.73 (s, 3H), 1.82-1.62 (m, 4H), 1.61-1.48 (m, 2H), 1.39 (s, 9H). LCMS t_R = 1.651 min in 2.5 min chromatography, MS ESI calcd. for C₂₅H₃₄FN₄O₄⁺ [M+H]⁺ 473.26, found 473.3.

tert-butyl (*R*)-(1-(4-(((*benzyloxy*)*carbonyl*)*amino*)-5-*fluoro*-2-(4H-1,2,4-*triazol*-4-*yl*)*phenyl*)*piperidin*-3*yl*)(*methyl*)*carbamate* (251). To a solution of tert-butyl (R)-(1-(2-amino-4-(((*benzyloxy*)*carbonyl*)*amino*)-5fluorophenyl)piperidin-3-yl)carbamate (**246**) (2.92 g, 6.18 mmol) and compound **2A** (2.72 g, 30.90 mmol) in pyridine (20 mL) was added Et₃N (4.38 g, 43.25 mmol) and Me₃SiCl (10.07 g, 92.69 mmol) at 25°C. The mixture was stirred at 100°C for 12 hours. The reaction mixture was concentrated. The reaction mixture was concentrated directly. The residue was purified by flash silica gel chromatography (eluent of 0%~35%, EtOAc/PE) to give the product. **251** (1.52 g, 15.8%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.63-9.57 (m, 1H), 8.87 (s, 1H), 8.80 (s, 1H), 7.45-7.38 (m, 7H), 5.15 (s, 3H), 2.71-2.65 (m, 5H), 1.78-1.68 (m, 2H), 1.66-1.52 (m, 4H), 1.43-1.37 (m, 9H). LCMS t_R = 2.298 min in 4 min chromatography, MS ESI calcd. for C₂₇H₃₄FN₆O₄⁺ [M+H]⁺ 525.26, found 525.4.

tert-butyl (*R*)-(1-(4-*amino*-5-*fluoro*-2-(4H-1,2,4-*triazo*l-4-*yl*)*phenyl*)*piperidin*-3-*yl*)(*methyl*)*carbamate* (**256**). To a solution of tert-butyl (R)-(1-(4-(((benzyloxy)carbonyl)amino)-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)*piperidin*-3-yl)carbamate (**251**) (1.51 g, 2.88 mmol) in MeOH (10 mL) was added Pd/C (1.56 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. The reaction mixture was stirred under H₂ (15 psi) at 25°C for 2 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (30 mL x 3). The combine filtrate was concentrated in vacuo. **256** (1.49 g, 36.3%) was obtained as a black brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.78 (s, 1H), 8.71-8.68 (m, 1H), 7.12-7.07 (m, 1H), 6.79-6.74 (m, 1H), 5.31-5.24 (m, 2H), 2.83-2.79 (m, 1H), 2.76-2.69 (m, 3H), 2.66 (s, 4H), 2.61-2.58 (m, 1H), 1.62-1.55 (m, 3H), 1.41-1.35 (m, 9H). LCMS t_R = 1.294 min in 2 min chromatography, MS ESI calcd. for C₁₉H₂₈FN₆O₂⁺ [M+H]⁺ 391.23, found 391.0.

(R)-(1-(4-((3-cyano-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidin-5-yl)amino)-5-fluoro-2-(4H-1,2,4tert-butyl triazol-4-yl)phenyl)piperidin-3-yl)(methyl)carbamate (261). To a solution of tert-butyl (R)-(1-(4-amino-5-fluoro-2-(4H-1,2,4-triazol-4-yl)phenyl)piperidin-3-yl)carbamate (256)(500)0.51 mmol) and 5-chloro-7mg, (cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (108 mg, 0.46 mmol) in dioxane (8 mL) was added Cs₂CO₃ (501 mg, 1.54 mmol), BINAP (48 mg, 0.08 mmol) and Pd(OAc)₂ (17 mg, 0.08 mmol) at 25°C. Then the mixture was degassed and purged with N_2 . The reaction mixture was heated in a microwave reactor at 130°C for 0.5 hour. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of $0 \sim 3\%$, MeOH/DCM). 261 (180 mg, 41.5%) was obtained as a black brown solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.50 (s, 1H), 8.77 (s, 1H), 8.35 (s, 1H), 8.33-8.30 (m, 1H), 8.28-8.21 (m, 1H), 7.44-7.23 (m, 3H), 6.18 (s, 1H), 1.79-1.55 (m, 4H), 1.41-1.38 (m, 9H), 1.34 (s, 2H), 0.87-0.77 (m, 2H), 0.75-0.66 (m, 2H). LCMS $t_R = 0.643$ min in 1 min chromatography, MS ESI calcd. for C₂₉H₃₅FN₁₁O₂⁺ [M+H]⁺ 588.29, found 588.2.

(*R*)-7-(*cyclopropylamino*)-5-((2-*fluoro*-4-(3-(*methylamino*)*piperidin*-1-*yl*)-5-(4*H*-1,2,4-*triazo*l-4*yl*)*phenyl*)*amino*)*pyrazolo*[1,5-*a*]*pyrimidine*-3-*carbonitrile* (**71**). A mixture of tert-butyl (R)-(1-(4-((3-cyano-7-(cyclopropylamino))pyrazolo[1,5-*a*]*pyrimidin*-5-*y*])*amino*)-5-fluoro-2-(4H-1,2,4-triazol-4-*y*])*phenyl*)*piperidin*-3*y*]*carbamate* (**261**) (180 mg, 0.31 mmol) in DCM (3 mL) was added TFA (3 mL) at 25°C. The reaction mixture was stirred at 25°C for 3 hours. The reaction mixture was concentrated in vacuo. The residue was purified by prep. HPLC (column: Welch Xtimate C18 150*30mm*5um; mobile phase: [water (FA)-ACN]; B%: 0%-34%, 25min). **71** as a formate salt (40 mg, 25.2%) was obtained as a gray solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.53 (s, 1H), 8.83 (s, 2H), 8.36 (s, 1H), 8.33 (s, 1H), 8.29 (dd, *J* = 5.1, 2.4 Hz, 1H), 8.25 (d, *J* = 8.4 Hz, 1H), 7.25 (d, *J* = 12.5 Hz, 1H), 6.18 (s, 1H), 3.02 (d, *J* = 11.1 Hz, 1H), 2.71 – 2.62 (m, 2H), 2.63 – 2.56 (m, 1H), 2.48 – 2.36 (m, 2H), 2.30 (s, 3H), 1.89 (d, *J* = 12.1 Hz, 1H), 1.65 – 1.54 (m, 1H), 1.38 (q, *J* = 11.7 Hz, 1H), 1.25 – 1.12 (m, 1H), 0.86 – 0.78 (m, 2H), 0.75 – 0.68 (m, 2H). ¹³C NMR (101 MHz, DMSO*d*₆) δ 164.60, 160.31, 156.86, 153.77 (d, *J* = 247.1 Hz), 150.72, 148.55, 145.03, 142.85, 123.60, 122.94 (d, *J* = 13.9 Hz), 121.80, 114.70, 108.82 (d, *J* = 22.5 Hz), 76.58, 76.51, 55.07, 54.23, 51.26, 31.49, 28.08, 23.38, 23.19, 6.55. HPLC t_R = 3.324 min in 8 min chromatography, purity 95.2%. LCMS t_R = 1.655 min in 4 min chromatography, MS ESI calcd. for C_{24H27}FN₁₁+ [M+H]⁺ 488.24, found 488.5.

4-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)morpholine (**199**). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (500 mg, 2.40 mmol) and morpholine (313.91 mg, 3.60 mmol) in MeCN (15 mL) was added K₂CO₃ (996 mg, 7.21 mmol) at 25°C. Then the mixture was stirred at 100°C for 12 hours. The reaction mixture was concentrated in vacuo. Water (50 mL) was added to the mixture. The residue was extracted with DCM (50 mL x2). The combined organic phase was washed with brine (20 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. **199** (1.16 g, crude) was obtained as a black brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.01 (s, 1H), 8.31-8.26 (m, 1H), 8.02 (s, 1H), 7.48-7.32 (m, 1H), 3.60-3.57 (m, 4H), 3.40-3.35 (m, 4H). LCMS t_R = 0.921 min in 2.5 min chromatography, MS ESI calcd. for C₁₂H₁₄N₅O₃⁺ [M+H]⁺ 276.11, found 276.2.

4-morpholino-3-(4H-1,2,4-triazol-4-yl)aniline (217). To a solution of 4-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)morpholine (199) (750 mg, 2.72 mmol) in MeOH (20 mL) was added Pd/C (0.70 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. Then the reaction mixture was stirred under H₂ (15 Psi) at 25°C for 10 hours. The reaction mixture was filtered via a celite pad. The pad was washed with MeOH (15 mL x3). The residue was purified by flash silica gel chromatography (eluent of 0~6%, MeOH/DCM). **217** (516 mg, 77.2%) was obtained as a black brown oil. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.82-8.76 (m, 1H), 8.02 (s, 1H), 7.16-6.94 (m, 1H), 6.67-6.54 (m, 1H), 5.25 (s, 1H), 3.56-3.55 (m, 2H), 3.38-3.38 (m, 2H), 2.75-2.71 (m, 3H), 2.55-2.53 (m, 2H). LCMS t_R = 0.593 min in 2.5 min chromatography, MS ESI calcd. for C₁₂H₁₆N₅O⁺ [M+H]⁺ 246.13, found 246.1.

7-(cyclopropylamino)-5-((4-morpholino-3-(4H-1,2,4-triazol-4-yl)phenyl)amino)pyrazolo[1,5-a]pyrimidine-3carbonitrile (**72**). To a solution of 4-morpholino-3-(4H-1,2,4-triazol-4-yl)aniline (**217**) (140 mg, 0.34 mmol) and 5-chloro-7-(cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (**75**) (40 mg, 0.17 mmol) in dioxane (3 mL) was added Cs₂CO₃ (167 mg, 0.51 mmol), BINAP (16 mg, 0.03 mmol) and Pd(OAc)₂ (6 mg, 0.03 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave reactor at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash silica gel chromatography (eluent of 0~3%, MeOH/DCM). **72** (15 mg, 19.8%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.85 (s, 1H), 8.89 (s, 2H), 8.37 (s, 1H), 8.31 (d, *J* = 1.9 Hz, 1H), 7.94 (d, *J* = 2.5 Hz, 1H), 7.73 (dd, *J* = 8.8, 2.5 Hz, 1H), 7.32 (d, *J* = 8.8 Hz, 1H), 5.97 (s, 1H), 3.55 (dd, *J* = 5.9, 3.2 Hz, 4H), 2.65 – 2.58 (m, 5H), 0.87 – 0.79 (m, 2H), 0.75 – 0.70 (m, 2H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 156.51, 150.81, 148.41, 145.00, 142.75, 140.46, 136.76, 128.09, 121.46, 120.21, 116.99, 114.66, 76.70, 76.40, 66.10, 51.33, 23.32, 6.53. HPLC t_R = 4.039 min in 8 min chromatography, purity 97.5%. LCMS t_R = 2.172 min in 1 min chromatography, MS ESI calcd. For C₂₂H₂₃N₁₀O⁺ [M+H]⁺ 443.21, found 443.1.

1-methyl-4-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)piperazine (200). To a solution of 4-(2-fluoro-5-nitrophenyl)-4H-1,2,4-triazole (**176**) (1 g, 4.80 mmol) and 1-methylpiperazine (529 mg, 5.28 mmol) in MeCN (10 mL) was added K₂CO₃ (664 mg, 4.80 mmol) at 25°C. The mixture was stirred at 100°C for 12 h under N₂ atmosphere. The reaction mixture was concentrated in vacuo. Water (30 mL) was added to the residue. The residue was purified by flash silica gel chromatography (eluent of 0%~3%, MeOH/DCM) to give the product. **200** (918 mg, 59.1%) was obtained as a brown solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.96 (s, 2H), 8.30-8.22 (m, 2H), 7.39-7.34 (m, 1H), 2.81-2.75 (m, 4H), 2.33-2.30 (m, 4H), 2.18 (s, 3H). LCMS t_R = 0.137 min in 1 min chromatography, MS ESI calcd. for C₁₃H₁₇N₆O₂⁺ [M+H]⁺ 289.14, found 289.1.

4-(4-methylpiperazin-1-yl)-3-(4H-1,2,4-triazol-4-yl)aniline (218). To a solution of 1-methyl-4-(4-nitro-2-(4H-1,2,4-triazol-4-yl)phenyl)piperazine (200) (910 mg, 3.16 mmol) in MeOH (10 mL) was added Pd/C (0.82 g, 10% Pd) under N₂ atmosphere. The suspension was degassed and purged with H₂ for 3 times. The reaction mixture was stirred under H₂ (15 Psi) at 35°C for 2 h. The reaction mixture was filtered. The filter cake was washed with MeOH (20 mL x3). The combined filtrate was concentrated in vacuo. 218 (744 mg, crude) was obtained as a gray solid. ¹H NMR (400 MHz, DMSO-d₆) δ 8.74 (s, 2H), 7.06-7.01 (m, 1H), 6.66-6.61 (m, 1H), 6.56-6.53(m, 1H), 5.22 (s, 2H), 2.57-2.52 (m, 4H), 2.30-2.19 (m, 4H), 2.12 (s, 3H).

7-(cyclopropylamino)-5-((4-(4-methylpiperazin-1-yl)-3-(4H-1,2,4-triazol-4-

vl)phenvl)amino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (73). To solution of 5-chloro-7а (cyclopropylamino)pyrazolo[1,5-a]pyrimidine-3-carbonitrile (75) (300 mg, 1.28 mmol) and 4-(4-methylpiperazin-1-yl)-3-(4H-1,2,4-triazol-4-yl)aniline (218) (498 mg, 1.93 mmol) in dioxane (5 mL) was added K₂CO₃ (532 mg, 3.85 mmol), BINAP (120 mg, 0.19 mmol) and Pd(OAc)₂ (43 mg, 0.19 mmol) at 25°C. Then the mixture was degassed and purged with N₂. The reaction mixture was heated in a microwave at 130°C for 0.5 h. The reaction mixture was concentrated in vacuo. The residue was purified by prep-HPLC (column: Xtimate C18 150*40mm*10um; mobile phase: [water (NH₃H₂O+NH₄HCO₃)-ACN]; B%: 20%-60%, 10min). **73** (139 mg, 23.8%) was obtained as a white solid. ¹H NMR (400 MHz, DMSO- d_6) δ 9.84 (s, 1H), 8.84 (s, 2H), 8.36 (d, J = 0.9 Hz, 1H), 8.20 (br s, 1H), 7.93 (dd, J = 2.5, 1.1 Hz, 1H), 7.70 (dd, J = 8.8, 2.5 Hz, 1H), 7.29 (dd, J = 8.9, 1.5 Hz, 1H), 5.96 (s, 1H), 2.69 - 2.56 (m, 5H), 2.28 (br s, 4H), 2.15 (s, 3H), 0.81(td, J = 7.2, 6.6, 5.0 Hz, 2H), 0.76 - 0.67 (m, 2H). ¹³C NMR (101 MHz, DMSO- d_6) δ 156.53, 150.85, 148.40, 145.02, 142.75, 140.74, 136.56, 127.98, 121.38, 120.18, 116.97, 114.74, 76.70, 76.41, 54.64, 50.83, 45.74, 23.34, 6.57. HPLC t_R = 2.941 min in 8 min chromatography, purity 99.9%. LCMS $t_R = 1.642$ min in 4 min chromatography, MS ESI calcd. for $C_{23}H_{26}N_{11}^{+}$ [M+H]⁺ 456.24, found 456.5.

ASSOCIATED CONTENT

Supporting Information

Eurofins KinaseProfiler radiometric enzymatic assay results; NanoBRET in digitonin-permeabilized cells; phosphorylation of EIF2S2 in A549ACE2 cells in vitro; comparison between CSNK2A2 and CSNK2A1 NanoBRET pIC₅₀s; crystallographic refinement statistics; NanoBRET K192 Selectivity Panel results; ¹H and ¹³C NMR spectra for analogues synthesized; HPLC trace for compound **53** (DOCX).

Molecular formula strings with associated biochemical and biological data (CSV).

Accession Codes

The crystal structures of CSNK2A1 with compounds **14** (PDB ID: 8P07), **50** (PDB ID: 8P06) and **53** (PDB ID: 9EZG) have been deposited in the PDB.

AUTHOR INFORMATION

Corresponding Author

Han Wee Ong – Rapidly Emerging Antiviral Drug Development Initiative (READDI), Chapel Hill, North Carolina 27599, USA; Structural Genomics Consortium (SGC) and Division of Chemical Biology and Medicinal Chemistry, Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA.

Authors

- Xuan Yang Rapidly Emerging Antiviral Drug Development Initiative (READDI), Chapel Hill, North Carolina 27599, USA; Structural Genomics Consortium (SGC) and Division of Chemical Biology and Medicinal Chemistry, Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA.
- Jeffery L. Smith Structural Genomics Consortium (SGC) and Division of Chemical Biology and Medicinal Chemistry, Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, NC 27599, USA.
- Rebekah J. Dickmander Rapidly Emerging Antiviral Drug Development Initiative (READDI), Chapel Hill, North Carolina 27599, USA; Department of Microbiology & Immunology, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA; Lineberger Comprehensive Cancer Center, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA; Department of Chemistry, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA; Department of Chemistry, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA.

Jason W. Brown – Takeda Development Center Americas, Inc., San Diego, California 92121, USA.

- Tammy M. Havener Structural Genomics Consortium (SGC) and Division of Chemical Biology and Medicinal Chemistry, Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA.
- Sharon A. Taft-Benz Rapidly Emerging Antiviral Drug Development Initiative (READDI), Chapel Hill, North Carolina 27599, USA; Department of Genetics, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA.
- Stefanie D. Howell Structural Genomics Consortium (SGC) and Division of Chemical Biology and Medicinal Chemistry, Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA.
- Marcia K. Sanders Rapidly Emerging Antiviral Drug Development Initiative (READDI), Chapel Hill, North Carolina 27599, USA; Department of Genetics, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA.
- Jacob L. Capener Structural Genomics Consortium (SGC) and Division of Chemical Biology and Medicinal Chemistry, Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA.

Rafael M. Couñago – Structural Genomics Consortium (SGC) and Division of Chemical Biology and Medicinal Chemistry, Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA; Centro de Química Medicinal (CQMED), Centro de Biologia Molecular e Engenharia Genética (CBMEG), University of Campinas, Campinas, São Paulo, 13083-886, Brazil.

Edcon Chang - Takeda Development Center Americas, Inc., San Diego, California 92121, USA.

- Andreas Krämer SGC, Institute of Pharmaceutical Chemistry, Goethe University Frankfurt am Main, Max-von-Laue-Str. 9, 60438, Frankfurt am Main, Germany.
- Nathaniel J. Moorman Rapidly Emerging Antiviral Drug Development Initiative (READDI), Chapel Hill, North Carolina 27599, USA; Department of Microbiology & Immunology, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA; Lineberger Comprehensive Cancer Center, University of North Carolina at Chapel Hill, North Carolina 27599, USA;
- Mark T. Heise Rapidly Emerging Antiviral Drug Development Initiative (READDI), Chapel Hill, North Carolina 27599, USA; Department of Genetics, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA.
- Alison D. Axtman Rapidly Emerging Antiviral Drug Development Initiative (READDI), Chapel Hill, North Carolina 27599, USA; Structural Genomics Consortium (SGC) and Division of Chemical Biology and Medicinal Chemistry, Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA.
- David H. Drewry Rapidly Emerging Antiviral Drug Development Initiative (READDI), Chapel Hill, North Carolina 27599, USA; Structural Genomics Consortium (SGC) and Division of Chemical Biology and Medicinal Chemistry, Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA; Lineberger Comprehensive Cancer Center, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, USA.
- Timothy M. Willson Rapidly Emerging Antiviral Drug Development Initiative (READDI), Chapel Hill, North Carolina 27599, USA; Structural Genomics Consortium (SGC) and Division of Chemical Biology and Medicinal Chemistry, Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA.

Author Contributions

H. W. Ong and X. Yang contributed equally to this work.

ACKNOWLEDGEMENTS

The Structural Genomics Consortium (SGC) is a registered charity (no. 1097737) that receives funds from Bayer AG, Boehringer Ingelheim, Bristol Myers Squibb, Genentech, Genome Canada through Ontario Genomics Institute [OGI-196], EU/EFPIA/OICR/McGill/KTH/Diamond Innovative Medicines Initiative 2 Joint Undertaking [EUbOPEN grant 875510], Janssen, Merck KGaA (aka EMD in Canada and United States), Pfizer, and Takeda. Research reported in this publication was supported in part by the NC Biotech Center Institutional support grant 2018-IDG-1030, by the NIH Illuminating the Druggable Genome 1U24DK116204-01, and Department of Defense ALSRP award AL190107. This project was supported by the Rapidly Emerging Antiviral Drug Development Initiative (READDI) at the University of North Carolina at Chapel Hill with funding from the North Carolina Coronavirus State and Local Fiscal Recovery Funds program, appropriated by the North Carolina General Assembly. Additional funding was provided by a grant from Millennium Pharmaceuticals (Takeda). This work was supported by NIH grant S100D032476 for upgrading the 500 MHz NMR spectrometer in the UNC Eshelman School of Pharmacy NMR Facility.

Constructs for NanoBRET measurements of CSNK2A1 and CSNK2A2 were provided by Promega Corporation (Madison, WI). We thank Laszlo Gyenis and David Litchfield for providing us with the EIF2S2 P-S2 antibody. We thank K. Saikatendu Singh (Takeda, San Diego, CA) for facilitating collaborative interactions and constructive criticism throughout the project. WuXi AppTec (Shanghai, China) provided the chemical synthesis support. Analiza, Inc. (Cleveland, OH) performed kinetic solubility and mouse liver microsome studies. Pharmaron (Beijing, China) provided the in vitro and in vivo pharmacokinetic assay support. The kinome tree in Figure 4 was generated using CORAL³⁴.

ABBREVIATIONS USED

AKT, RAC-alpha serine/threonine-protein kinase; b.i.d., twice daily; BINAP, 2,2'-bis(diphenylphosphino)-1,1'binaphthyl; BRET, bioluminescence resonance energy transfer; CME, clathrin-mediated endocytosis; COVID-19, Coronavirus Disease 2019; CSNK2, casein kinase II; CSNK2A1, casein kinase II subunit alpha; CSNK2A2, casein kinase II subunit alpha'; DBU, 1,8-Diazabicyclo(5.4.0)undec-7-ene; DCM, dichloromethane; DIPEA, diisopropylethylamine; DMF, dimethylformamide; DMSO, dimethylsulfoxide; EDTA, ethylenediaminetetraacetic acid; EIF2S2, eukaryotic translation initiation factor 2 subunit 2; HLM, human liver microsome; HPLC, high-performance liquid chromatography; i.p., intraperitoneal; i.v., intraveneous; IPTG, isopropyl-thio-galactopyranoside; LC-MS, liquid chromatography mass spectrometry; LC-TOFMS, liquid chromatography time of flight mass spectrometry; LDH, lactate dehydrogenase; LOD, limit of detection; mBRET, milliBRET; mCPBA, meta-chloroperoxybenzoic acid; MDR1, ATP-dependent translocase ABCB1; MHV, mouse hepatitis virus; MLM, mouse liver microsome; MS, mass spectrometry; NanoBRET, nanoluciferase bioluminescence resonance energy transfer; NLuc, nanoluciferase; NMP, N-methyl-2-pyrrolidone; p.o., oral administration; PBS, phosphate-buffered saline; PFU, plaque forming units; PK, pharmacokinetics; PVDF, polyvinylidene difluoride; RLU, relative light unit; SAR, structure-activity relationship; $S_N 2$, bimolecular nucleophilic substitution; $S_N Ar$, nucleophilic aromatic substitution; TBAF, tetra-n-butylammonium fluoride; TBDMS, tert-butyldimethylsilyl; TBDPS, tertbutyldiphenylsilyl; TEER, transepithelial electrical resistance; TFA, trifluoroacetic acid; THF, tetrahydrofuran; TLC, thinlayer chromatography.

REFERENCES

- (1) World Health Organization. WHO COVID-19 Dashboard. https://covid19.who.int/ (accessed 2024-04-22).
- (2) Payne, S. Family Coronaviridae. In *Viruses*; Elsevier, 2017; pp 149–158.
- (3) Shereen, M. A.; Khan, S.; Kazmi, A.; Bashir, N.; Siddique, R. COVID-19 Infection: Emergence, Transmission, and Characteristics of Human Coronaviruses. *J. Adv. Res.* **2020**, *24*, 91–98.
- (4) Kumar, N.; Sharma, S.; Kumar, R.; Tripathi, B. N.; Barua, S.; Ly, H.; Rouse, B. T. Host-Directed Antiviral Therapy. *Clin. Microbiol. Rev.* **2020**, *33*, 1–36.
- (5) Tripathi, D.; Sodani, M.; Gupta, P. K.; Kulkarni, S. Host Directed Therapies: COVID-19 and Beyond. *Curr. Res. Pharmacol. Drug Discov.* **2021**, *2*, 100058.
- (6) Ji, X.; Li, Z. Medicinal Chemistry Strategies toward Host Targeting Antiviral Agents. *Med. Res. Rev.* 2020, 1–39.
- (7) Keating, J. A.; Striker, R. Phosphorylation Events during Viral Infections Provide Potential Therapeutic Targets. *Rev. Med. Virol.* **2012**, *22*, 166–181.
- (8) Quezada Meza, C. P.; Ruzzene, M. Protein Kinase CK2 and SARS-CoV-2: An Expected Interplay Story. *Kinases and Phosphatases* **2023**, *1*, 141–150.
- Bouhaddou, M.; Memon, D.; Meyer, B.; White, K. M.; Rezelj, V. V.; Correa Marrero, M.; Polacco, B. J.; Melnyk, J. E.; Ulferts, S.; Kaake, R. M.; Batra, J.; Richards, A. L.; Stevenson, E.; Gordon, D. E.; Rojc, A.; Obernier, K.; Fabius, J. M.; Soucheray, M.; Miorin, L.; Moreno, E.; Koh, C.; Tran, Q. D.; Hardy, A.; Robinot, R.; Vallet, T.; Nilsson-Payant, B. E.; Hernandez-Armenta, C.; Dunham, A.; Weigang, S.; Knerr, J.; Modak, M.; Quintero, D.; Zhou, Y.; Dugourd, A.; Valdeolivas, A.; Patil, T.; Li, Q.; Hüttenhain, R.; Cakir, M.; Muralidharan, M.; Kim, M.; Jang, G.; Tutuncuoglu, B.; Hiatt, J.; Guo, J. Z.; Xu, J.; Bouhaddou, S.; Mathy, C. J. P.; Gaulton, A.; Manners, E. J.; Félix, E.; Shi, Y.; Goff, M.; Lim, J. K.; McBride, T.; O'Neal, M. C.; Cai, Y.; Chang, J. C. J.; Broadhurst, D. J.; Klippsten, S.; De wit, E.; Leach, A. R.; Kortemme, T.; Shoichet, B.; Ott, M.; Saez-Rodriguez, J.; TenOever, B. R.; Mullins, R. D.; Fischer, E. R.; Kochs, G.; Grosse, R.; García-Sastre, A.; Vignuzzi, M.; Johnson, J. R.; Shokat, K. M.; Swaney, D. L.; Beltrao, P.; Krogan, N. J. The Global Phosphorylation Landscape of SARS-CoV-2 Infection. *Cell* 2020, *182*, 685–712.
- Gordon, D. E.; Jang, G. M.; Bouhaddou, M.; Xu, J.; Obernier, K.; White, K. M.; O'Meara, M. J.; Rezelj, V. V.; Guo, J. Z.; Swaney, D. L.; Tummino, T. A.; Hüttenhain, R.; Kaake, R. M.; Richards, A. L.; Tutuncuoglu, B.; Foussard, H.; Batra, J.; Haas, K.; Modak, M.; Kim, M.; Haas, P.; Polacco, B. J.; Braberg, H.; Fabius, J. M.; Eckhardt, M.; Soucheray, M.; Bennett, M. J.; Cakir, M.; McGregor, M. J.; Li, Q.; Meyer, B.; Roesch, F.; Vallet, T.; Mac Kain, A.; Miorin, L.; Moreno, E.; Naing, Z. Z. C.; Zhou, Y.; Peng, S.; Shi, Y.; Zhang, Z.; Shen, W.; Kirby, I. T.; Melnyk, J. E.; Chorba, J. S.; Lou, K.; Dai, S. A.; Barrio-Hernandez, I.; Memon, D.; Hernandez-Armenta, C.; Lyu, J.; Mathy, C. J. P.; Perica, T.; Pilla, K. B.; Ganesan, S. J.; Saltzberg, D. J.; Rakesh, R.; Liu, X.; Rosenthal, S. B.; Calviello, L.;

Venkataramanan, S.; Liboy-Lugo, J.; Lin, Y.; Huang, X.-P.; Liu, Y.; Wankowicz, S. A.; Bohn, M.; Safari, M.; Ugur, F. S.; Koh, C.; Savar, N. S.; Tran, Q. D.; Shengjuler, D.; Fletcher, S. J.; O'Neal, M. C.; Cai, Y.; Chang, J. C. J.; Broadhurst, D. J.; Klippsten, S.; Sharp, P. P.; Wenzell, N. A.; Kuzuoglu-Ozturk, D.; Wang, H.-Y.; Trenker, R.; Young, J. M.; Cavero, D. A.; Hiatt, J.; Roth, T. L.; Rathore, U.; Subramanian, A.; Noack, J.; Hubert, M.; Stroud, R. M.; Frankel, A. D.; Rosenberg, O. S.; Verba, K. A.; Agard, D. A.; Ott, M.; Emerman, M.; Jura, N.; von Zastrow, M.; Verdin, E.; Ashworth, A.; Schwartz, O.; D'Enfert, C.; Mukherjee, S.; Jacobson, M.; Malik, H. S.; Fujimori, D. G.; Ideker, T.; Craik, C. S.; Floor, S. N.; Fraser, J. S.; Gross, J. D.; Sali, A.; Roth, B. L.; Ruggero, D.; Taunton, J.; Kortemme, T.; Beltrao, P.; Vignuzzi, M.; García-Sastre, A.; Shokat, K. M.; Shoichet, B. K.; Krogan, N. J. A SARS-CoV-2 Protein Interaction Map Reveals Targets for Drug Repurposing. *Nature* 2020, *583*, 459–468.

- Gordon, D. E.; Hiatt, J.; Bouhaddou, M.; Rezelj, V. V.; Ulferts, S.; Braberg, H.; Jureka, A. S.; Obernier, K.; Guo, J. (11)Z.; Batra, J.; Kaake, R. M.; Weckstein, A. R.; Owens, T. W.; Gupta, M.; Pourmal, S.; Titus, E. W.; Cakir, M.; Soucheray, M.; McGregor, M.; Cakir, Z.; Jang, G.; O'Meara, M. J.; Tummino, T. A.; Zhang, Z.; Foussard, H.; Rojc, A.; Zhou, Y.; Kuchenov, D.; Hüttenhain, R.; Xu, J.; Eckhardt, M.; Swaney, D. L.; Fabius, J. M.; Ummadi, M.; Tutuncuoglu, B.; Rathore, U.; Modak, M.; Haas, P.; Haas, K. M.; Naing, Z. Z. C.; Pulido, E. H.; Shi, Y.; Barrio-Hernandez, I.; Memon, D.; Petsalaki, E.; Dunham, A.; Marrero, M. C.; Burke, D.; Koh, C.; Vallet, T.; Silvas, J. A.; Azumaya, C. M.; Billesbølle, C.; Brilot, A. F.; Campbell, M. G.; Diallo, A.; Dickinson, M. S.; Diwanji, D.; Herrera, N.; Hoppe, N.; Kratochvil, H. T.; Liu, Y.; Merz, G. E.; Moritz, M.; Nguyen, H. C.; Nowotny, C.; Puchades, C.; Rizo, A. N.; Schulze-Gahmen, U.; Smith, A. M.; Sun, M.; Young, I. D.; Zhao, J.; Asarnow, D.; Biel, J.; Bowen, A.; Braxton, J. R.; Chen, J.; Chio, C. M.; Chio, U. S.; Deshpande, I.; Doan, L.; Faust, B.; Flores, S.; Jin, M.; Kim, K.; Lam, V. L.; Li, F.; Li, J.; Li, Y.-L.; Li, Y.; Liu, X.; Lo, M.; Lopez, K. E.; Melo, A. A.; Moss, F. R.; Nguyen, P.; Paulino, J.; Pawar, K. I.; Peters, J. K.; Pospiech, T. H.; Safari, M.; Sangwan, S.; Schaefer, K.; Thomas, P. V.; Thwin, A. C.; Trenker, R.; Tse, E.; Tsui, T. K. M.; Wang, F.; Whitis, N.; Yu, Z.; Zhang, K.; Zhang, Y.; Zhou, F.; Saltzberg, D.; Hodder, A. J.; Shun-Shion, A. S.; Williams, D. M.; White, K. M.; Rosales, R.; Kehrer, T.; Miorin, L.; Moreno, E.; Patel, A. H.; Rihn, S.; Khalid, M. M.; Vallejo-Gracia, A.; Fozouni, P.; Simoneau, C. R.; Roth, T. L.; Wu, D.; Karim, M. A.; Ghoussaini, M.; Dunham, I.; Berardi, F.; Weigang, S.; Chazal, M.; Park, J.; Logue, J.; McGrath, M.; Weston, S.; Haupt, R.; Hastie, C. J.; Elliott, M.; Brown, F.; Burness, K. A.; Reid, E.; Dorward, M.; Johnson, C.; Wilkinson, S. G.; Geyer, A.; Giesel, D. M.; Baillie, C.; Raggett, S.; Leech, H.; Toth, R.; Goodman, N.; Keough, K. C.; Lind, A. L.; Klesh, R. J.; Hemphill, K. R.; Carlson-Stevermer, J.; Oki, J.; Holden, K.; Maures, T.; Pollard, K. S.; Sali, A.; Agard, D. A.; Cheng, Y.; Fraser, J. S.; Frost, A.; Jura, N.; Kortemme, T.; Manglik, A.; Southworth, D. R.; Stroud, R. M.; Alessi, D. R.; Davies, P.; Frieman, M. B.; Ideker, T.; Abate, C.; Jouvenet, N.; Kochs, G.; Shoichet, B.; Ott, M.; Palmarini, M.; Shokat, K. M.; García-Sastre, A.; Rassen, J. A.; Grosse, R.; Rosenberg, O. S.; Verba, K. A.; Basler, C. F.; Vignuzzi, M.; Peden, A. A.; Beltrao, P.; Krogan, N. J.; Owens, T. W.; Gupta, M.; Pourmal, S.; Titus, E. W.; Azumaya, C. M.; Billesbølle, C.; Brilot, A. F.; Campbell, M. G.; Diallo, A.; Dickinson, M. S.; Diwanji, D.; Herrera, N.; Hoppe, N.; Kratochvil, H. T.; Liu, Y.; Merz, G. E.; Moritz, M.; Nguyen, H. C.; Nowotny, C.; Puchades, C.; Rizo, A. N.; Schulze-Gahmen, U.; Smith, A. M.; Sun, M.; Young, I. D.; Zhao, J.; Asarnow, D.; Biel, J.; Bowen, A.; Braxton, J. R.; Chen, J.; Chio, C. M.; Chio, U. S.; Deshpande, I.; Doan, L.; Faust, B.; Flores, S.; Jin, M.; Kim, K.; Lam, V. L.; Li, F.; Li, J.; Li, Y.-L.; Li, Y.; Liu, X.; Lo, M.; Lopez, K. E.; Melo, A. A.; Moss, F. R.; Nguyen, P.; Paulino, J.; Pawar, K. I.; Peters, J. K.; Pospiech, T. H.; Safari, M.; Sangwan, S.; Schaefer, K.; Thomas, P. V.; Thwin, A. C.; Trenker, R.; Tse, E.; Tsui, T. K. M.; Wang, F.; Whitis, N.; Yu, Z.; Zhang, K.; Zhang, Y.; Zhou, F.; Trinidad, D.; Agard, D. A.; Cheng, Y.; Fraser, J. S.; Frost, A.; Jura, N.; Kortemme, T.; Manglik, A.; Southworth, D. R.; Stroud, R. M.; Rosenberg, O. S.; Verba, K. A.; Damas, J.; Hughes, G. M.; Keough, K. C.; Painter, C. A.; Persky, N. S.; Corbo, M.; Kirilenko, B.; Hiller, M.; Koepfli, K.-P.; Kaplow, I.; Wirthlin, M.; Pfenning, A. R.; Zhao, H.; Genereux, D. P.; Swofford, R.; Lind, A.; Pollard, K. S.; Ryderq, O. A.; Nweeia, M. T.; Meadows, J.; Dong, M.; Wallerman, O.; Marinescu, V.; Lindblad-Toh, K.; Ray, D. A.; Power, S.; Teeling, E. C.; Chauhan, G.; Li, S. X.; Karlsson, E. K.; Lewin, H. A. Comparative Host-Coronavirus Protein Interaction Networks Reveal Pan-Viral Disease Mechanisms. Science 2020, 370, eabe9403.
- (12) Ferguson, F. M.; Gray, N. S. Kinase Inhibitors: The Road Ahead. Nat. Rev. Drug Discov. 2018, 17, 353–377.
- (13) Xie, Z.; Yang, X.; Duan, Y.; Han, J.; Liao, C. Small-Molecule Kinase Inhibitors for the Treatment of Nononcologic Diseases. *J. Med. Chem.* **2021**, *64*, 1283–1345.
- (14) García-Cárceles, J.; Caballero, E.; Gil, C.; Martínez, A. Kinase Inhibitors as Underexplored Antiviral Agents. J. *Med. Chem.* **2022**, *65*, 935–954.
- (15) Yang, X.; Dickmander, R. J.; Bayati, A.; Taft-Benz, S. A.; Smith, J. L.; Wells, C. I.; Madden, E. A.; Brown, J. W.; Lenarcic, E. M.; Yount, B. L.; Chang, E.; Axtman, A. D.; Baric, R. S.; Heise, M. T.; McPherson, P. S.; Moorman, N. J.; Willson, T. M. Host Kinase CSNK2 Is a Target for Inhibition of Pathogenic SARS-like β-Coronaviruses. *ACS Chem. Biol.* **2022**, *17*, 1937–1950.
- (16) Ramón, A. C.; Pérez, G. V.; Caballero, E.; Rosales, M.; Aguilar, D.; Vázquez-Blomquist, D.; Ramos, Y.; Rodríguez-

Ulloa, A.; Falcón, V.; Rodríguez-Moltó, M. P.; Yang, K.; Perera, Y.; Perea, S. E. Targeting of Protein Kinase CK2 Elicits Antiviral Activity on Bovine Coronavirus Infection. *Viruses* **2022**, *14*, 1–16.

- (17) Chen, Y.; Wang, Y.; Wang, J.; Zhou, Z.; Cao, S.; Zhang, J. Strategies of Targeting CK2 in Drug Discovery: Challenges, Opportunities, and Emerging Prospects. *J. Med. Chem.* **2023**, *66*, 2257–2281.
- (18) Ong, H. W.; Drewry, D. H.; Axtman, A. D. CK2 Chemical Probes: Past, Present, and Future. *Kinases and Phosphatases* **2023**, *1*, 288–305.
- (19) Wells, C. I.; Drewry, D. H.; Pickett, J. E.; Tjaden, A.; Krämer, A.; Müller, S.; Gyenis, L.; Menyhart, D.; Litchfield, D. W.; Knapp, S.; Axtman, A. D. Development of a Potent and Selective Chemical Probe for the Pleiotropic Kinase CK2. *Cell Chem. Biol.* 2021, 28, 546–558.
- (20) Leist, S. R.; Dinnon, K. H.; Schäfer, A.; Tse, L. V.; Okuda, K.; Hou, Y. J.; West, A.; Edwards, C. E.; Sanders, W.; Fritch, E. J.; Gully, K. L.; Scobey, T.; Brown, A. J.; Sheahan, T. P.; Moorman, N. J.; Boucher, R. C.; Gralinski, L. E.; Montgomery, S. A.; Baric, R. S. A Mouse-Adapted SARS-CoV-2 Induces Acute Lung Injury and Mortality in Standard Laboratory Mice. *Cell* **2020**, *183*, 1070–1085.
- (21) Dowling, J. E.; Chuaqui, C.; Pontz, T. W.; Lyne, P. D.; Larsen, N. A.; Block, M. H.; Chen, H.; Su, N.; Wu, A.; Russell, D.; Pollard, H.; Lee, J. W.; Peng, B.; Thakur, K.; Ye, Q.; Zhang, T.; Brassil, P.; Racicot, V.; Bao, L.; Denz, C. R.; Cooke, E. Potent and Selective Inhibitors of CK2 Kinase Identified through Structure-Guided Hybridization. ACS Med. Chem. Lett. 2012, 3, 278–283.
- (22) Dowling, J. E.; Alimzhanov, M.; Bao, L.; Block, M. H.; Chuaqui, C.; Cooke, E. L.; Denz, C. R.; Hird, A.; Huang, S.; Larsen, N. A.; Peng, B.; Pontz, T. W.; Rivard-Costa, C.; Saeh, J. C.; Thakur, K.; Ye, Q.; Zhang, T.; Lyne, P. D. Structure and Property Based Design of Pyrazolo[1,5-a]Pyrimidine Inhibitors of CK2 Kinase with Activity in Vivo. ACS Med. Chem. Lett. 2013, 4, 800–805.
- (23) Dowling, J. E.; Alimzhanov, M.; Bao, L.; Chuaqui, C.; Denz, C. R.; Jenkins, E.; Larsen, N. A.; Lyne, P. D.; Pontz, T.; Ye, Q.; Holdgate, G. A.; Snow, L.; O'Connell, N.; Ferguson, A. D. Potent and Selective CK2 Kinase Inhibitors with Effects on Wnt Pathway Signaling in Vivo. ACS Med. Chem. Lett. 2016, 7, 300–305.
- Yang, X.; Ong, H. W.; Dickmander, R. J.; Smith, J. L.; Brown, J. W.; Tao, W.; Chang, E.; Moorman, N. J.; Axtman, A. D.; Willson, T. M. Optimization of 3-Cyano-7-Cyclopropylamino-Pyrazolo[1,5-a]Pyrimidines toward the Development of an In Vivo Chemical Probe for CSNK2A. ACS Omega 2023, 8, 39546–39561.
- (25) Körner, R.; Majjouti, M.; Alcazar, M.; Mahabir, E. Of Mice and Men: The Coronavirus MHV and Mouse Models as a Translational Approach to Understand SARS-CoV-2. *Viruses* **2020**, *12*, 880.
- (26) Kumari, S.; Carmona, A. V.; Tiwari, A. K.; Trippier, P. C. Amide Bond Bioisosteres: Strategies, Synthesis, and Successes. J. Med. Chem. 2020, 63, 12290–12358.
- (27) Pennington, L. D.; Aquila, B. M.; Choi, Y.; Valiulin, R. A.; Muegge, I. Positional Analogue Scanning: An Effective Strategy for Multiparameter Optimization in Drug Design. *J. Med. Chem.* **2020**, *63*, 8956–8976.
- (28) Potęga, A. Glutathione-Mediated Conjugation of Anticancer Drugs: An Overview of Reaction Mechanisms and Biological Significance for Drug Detoxification and Bioactivation. *Molecules* **2022**, *27*, 5252.
- (29) Driscoll, J. P.; Yadav, A. S.; Shah, N. R. Role of Glucuronidation and P450 Oxidation in the Bioactivation of Bromfenac. *Chem. Res. Toxicol.* **2018**, *31*, 223–230.
- (30) Evison, B. J.; Sleebs, B. E.; Watson, K. G.; Phillips, D. R.; Cutts, S. M. Mitoxantrone, More than Just Another Topoisomerase II Poison. *Med. Res. Rev.* **2016**, *36*, 248–299.
- (31) Vasta, J. D.; Corona, C. R.; Wilkinson, J.; Zimprich, C. A.; Hartnett, J. R.; Ingold, M. R.; Zimmerman, K.; Machleidt, T.; Kirkland, T. A.; Huwiler, K. G.; Ohana, R. F.; Slater, M.; Otto, P.; Cong, M.; Wells, C. I.; Berger, B.-T.; Hanke, T.; Glas, C.; Ding, K.; Drewry, D. H.; Huber, K. V. M.; Willson, T. M.; Knapp, S.; Müller, S.; Meisenheimer, P. L.; Fan, F.; Wood, K. V.; Robers, M. B. Quantitative, Wide-Spectrum Kinase Profiling in Live Cells for Assessing the Effect of Cellular ATP on Target Engagement. *Cell Chem. Biol.* **2018**, *25*, 206–214.
- (32) Gandin, V.; Masvidal, L.; Cargnello, M.; Gyenis, L.; McLaughlan, S.; Cai, Y.; Tenkerian, C.; Morita, M.; Balanathan, P.; Jean-Jean, O.; Stambolic, V.; Trost, M.; Furic, L.; Larose, L.; Koromilas, A. E.; Asano, K.; Litchfield, D.; Larsson, O.; Topisirovic, I. MTORC1 and CK2 Coordinate Ternary and EIF4F Complex Assembly. *Nat. Commun.* 2016, *7*, 11127.
- (33) Robers, M. B.; Wilkinson, J. M.; Vasta, J. D.; Berger, L. M.; Berger, B. T.; Knapp, S. Single Tracer-Based Protocol for Broad-Spectrum Kinase Profiling in Live Cells with NanoBRET. *STAR Protoc.* **2021**, *2*, 100822.
- (34) Metz, K. S.; Deoudes, E. M.; Berginski, M. E.; Jimenez-Ruiz, I.; Aksoy, B. A.; Hammerbacher, J.; Gomez, S. M.; Phanstiel, D. H. Coral: Clear and Customizable Visualization of Human Kinome Data. *Cell Syst.* **2018**, *7*, 347–350.
- (35) Al Shoyaib, A.; Archie, S. R.; Karamyan, V. T. Intraperitoneal Route of Drug Administration: Should It Be Used in Experimental Animal Studies? *Pharm. Res.* **2020**, *37*, 12.
- (36) Smith, D. A.; Beaumont, K.; Maurer, T. S.; Di, L. Volume of Distribution in Drug Design. J. Med. Chem. 2015, 58, 5691–5698.

- (37) Di Maira, G.; Salvi, M.; Arrigoni, G.; Marin, O.; Sarno, S.; Brustolon, F.; Pinna, L. A.; Ruzzene, M. Protein Kinase CK2 Phosphorylates and Upregulates Akt/PKB. *Cell Death Differ*. **2005**, *12*, 668–677.
- (38) Zhang, X.; Yang, Y.; Grimstein, M.; Liu, G.; Kitabi, E.; Fan, J.; Wang, Y.; Earp, J.; Weaver, J. L.; Zhu, H.; Liu, J.; Reynolds, K. S.; Huang, S.; Wang, Y. Anti–SARS-CoV-2 Repurposing Drug Database: Clinical Pharmacology Considerations. *CPT Pharmacometrics Syst. Pharmacol.* **2021**, *10*, 973–982.
- (39) von Delft, A.; Hall, M. D.; Kwong, A. D.; Purcell, L. A.; Saikatendu, K. S.; Schmitz, U.; Tallarico, J. A.; Lee, A. A. Accelerating Antiviral Drug Discovery: Lessons from COVID-19. *Nat. Rev. Drug Discov.* **2023**, *22*, 585–603.
- (40) Dinnon, K. H.; Leist, S. R.; Schäfer, A.; Edwards, C. E.; Martinez, D. R.; Montgomery, S. A.; West, A.; Yount, B. L.; Hou, Y. J.; Adams, L. E.; Gully, K. L.; Brown, A. J.; Huang, E.; Bryant, M. D.; Choong, I. C.; Glenn, J. S.; Gralinski, L. E.; Sheahan, T. P.; Baric, R. S. A Mouse-Adapted Model of SARS-CoV-2 to Test COVID-19 Countermeasures. *Nature* 2020, *586*, 560–566.
- (41) Schindelin, J.; Arganda-Carreras, I.; Frise, E.; Kaynig, V.; Longair, M.; Pietzsch, T.; Preibisch, S.; Rueden, C.; Saalfeld, S.; Schmid, B.; Tinevez, J.-Y.; White, D. J.; Hartenstein, V.; Eliceiri, K.; Tomancak, P.; Cardona, A. Fiji: An Open-Source Platform for Biological-Image Analysis. *Nat. Methods* **2012**, *9*, 676–682.
- (42) Davis-Gilbert, Z. W.; Krämer, A.; Dunford, J. E.; Howell, S.; Senbabaoglu, F.; Wells, C. I.; Bashore, F. M.; Havener, T. M.; Smith, J. L.; Hossain, M. A.; Oppermann, U.; Drewry, D. H.; Axtman, A. D. Discovery of a Potent and Selective Naphthyridine-Based Chemical Probe for Casein Kinase 2. ACS Med. Chem. Lett. 2023, 14, 432–441.
- (43) Krämer, A.; Kurz, C. G.; Berger, B.-T.; Celik, I. E.; Tjaden, A.; Greco, F. A.; Knapp, S.; Hanke, T. Optimization of Pyrazolo[1,5-a]Pyrimidines Lead to the Identification of a Highly Selective Casein Kinase 2 Inhibitor. *Eur. J. Med. Chem.* 2020, 208, 112770.
- (44) Kabsch, W. XDS. Acta Crystallogr. Sect. D Biol. Crystallogr. 2010, 66, 125–132.
- (45) Evans, P. R.; Murshudov, G. N. How Good Are My Data and What Is the Resolution? *Acta Crystallogr. Sect. D Biol. Crystallogr.* **2013**, *69*, 1204–1214.
- (46) Lebedev, A. A.; Vagin, A. A.; Murshudov, G. N. Model Preparation in MOLREP and Examples of Model Improvement Using X-Ray Data. *Acta Crystallogr. Sect. D Biol. Crystallogr.* **2008**, *64*, 33–39.
- (47) Emsley, P.; Cowtan, K. Coot: Model-Building Tools for Molecular Graphics. Acta Crystallogr. Sect. D Biol. Crystallogr. 2004, 60, 2126–2132.
- (48) Vagin, A. A.; Steiner, R. A.; Lebedev, A. A.; Potterton, L.; McNicholas, S.; Long, F.; Murshudov, G. N. REFMAC 5 Dictionary: Organization of Prior Chemical Knowledge and Guidelines for Its Use. *Acta Crystallogr. Sect. D Biol. Crystallogr.* 2004, 60, 2184–2195.