

1                   **Optimization of heterogeneous continuous flow**  
2                   **hydrogenation using FTIR inline analysis: a comparative**  
3                   **study of multi-objective Bayesian optimization and kinetic**  
4                   **modeling**

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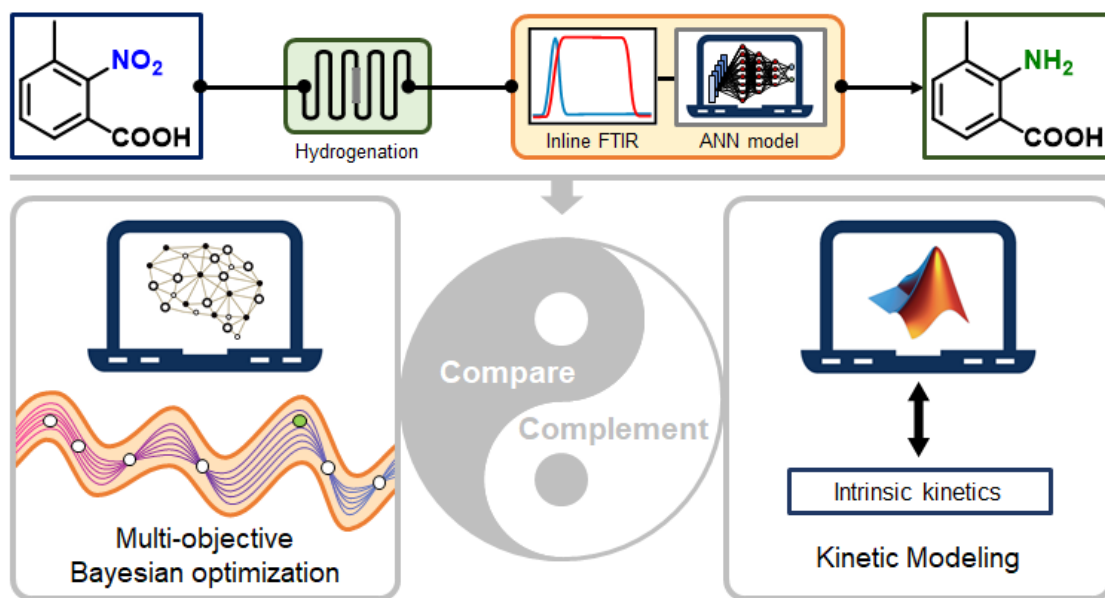
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27 **Graphical Abstract**



28

## 29 **Abstract**

30 The heterogeneous continuous flow hydrogenation is pivotal in chemical research and  
31 production, yet its reaction optimization has historically been intricate and labor-  
32 intensive. This study introduces a heterogeneous continuous flow hydrogenation  
33 system specifically designed for the preparation of 2-amino-3-methylbenzoic acid  
34 (AMA), employing FTIR inline analysis coupled with an artificial neural network  
35 model for monitoring. We explored two distinct reaction optimization strategies: multi-  
36 objective Bayesian optimization (MOBO) and intrinsic kinetic modeling, executed in  
37 parallel to optimize the reaction conditions. Remarkably, the MOBO approach achieved  
38 an optimal AMA yield of 99% and a productivity of 0.64 g/hour within a limited  
39 number of iterations. Conversely, despite requiring extensive experimental data  
40 collection and equation fitting, the intrinsic kinetic modeling approach yielded a similar  
41 optimal AMA yield but a higher productivity of 1.13 g/hour, attributed to increased  
42 catalyst usage. Our findings indicate that while MOBO offers a more efficient route  
43 with fewer required experiments, kinetic modeling provides deeper insights into the  
44 reaction optimization landscape but is limited by its assumptions.

45

## 46 **Keywords**

47 Multi-objective Bayesian optimization; Kinetic modeling; continuous flow;  
48 heterogeneous hydrogenation; Reaction optimization; Inline analysis

## 49 **Introduction**

50 Heterogeneous catalytic hydrogenation is pivotal in organic synthetic chemistry, with  
51 broad applications across the dye, pharmaceutical, and fine chemical industries.[1-3]  
52 Traditional intermittent hydrogenation processes, however, are constrained by critical  
53 safety concerns associated with reaction temperature and pressure, prompting a shift  
54 towards continuous flow hydrogenation as a safer and more efficient alternative.[4, 5]  
55 The reaction optimization within these continuous flow systems has emerged as a  
56 research focus.[6, 7] Historically, the reaction optimization process predominantly  
57 employed the one factor at a time (OFAT) method [8], which relies on a sequential,  
58 factor-based optimization guided by chemical intuition[9-12]. Despite its widespread  
59 use, the OFAT approach is often criticized for its inefficiency and inaccuracy, primarily  
60 due to its inability to account for potential synergistic effects among various factors,  
61 potentially leading to misinterpretation of the chemical processes.[13] Consequently,  
62 there is a pressing need for developing systematic approaches to reaction optimization.

63  
64 Kinetic modeling, grounded in a comprehensive understanding of chemical processes,  
65 represents a classical and crucial strategy for reaction optimization, especially for  
66 heterogeneous hydrogenation reactions.[14-17] For instance, Su et al. demonstrated the  
67 application of kinetic modeling in the heterogeneous flow hydrogenation of  
68 hexafluoroacetone trihydrate, revealing an adsorption-desorption mechanism with  
69 competitive adsorption of H<sub>2</sub> dissociation.[18] Similarly, Yu et al. conducted a  
70 continuous hydrogenation study of 2-(4-nitrophenyl) butanoic acid and kinetics study  
71 in a micropacked-bed reactor, employing kinetic analysis to elucidate the impacts of  
72 internal and external diffusion, as well as salt formation.[19] Despite these advances,  
73 the application of kinetic modeling in heterogeneous catalysis is often hampered by the  
74 complexities associated with non-chemical kinetics phenomena.[20-24]

75  
76 Multi-objective Bayesian Optimization (MOBO) has revolutionized the optimization  
77 of continuous-flow reactions by adeptly handling competing objectives.[25-28] It

78 leverages the surrogate model and acquisition function to systematically explore the  
79 Pareto Front, aiming to strike an optimal balance among objectives. Using probabilistic  
80 models, MOBO predicts outcomes for specific reaction conditions, guiding the  
81 optimization process toward superior performance metrics. Recent significant  
82 implementations of MOBO include Jensen et al.'s optimization of multistep synthetic  
83 routes on an automated platform [25], Bourne et al.'s achievement of an 81% yield  
84 through the simultaneous optimization of telescoped reactions [29], and Lapkin et al.'s  
85 application of MOBO in medicinal chemistry for yield optimization [30]. Despite its  
86 advancements, MOBO's comparison with traditional optimization methods is less  
87 studied.

88

89 Inline analytical techniques, including inline FTIR, NMR, and UV/vis spectroscopy,  
90 are becoming integral to reaction optimization due to their ability to provide real-time  
91 data, which facilitates the creation of self-optimizing systems when combined with  
92 Bayesian optimization.[29, 31, 32] Kappe et al. have notably advanced this field by  
93 integrating four complementary inline analysis instruments and developing advanced  
94 data analysis models, which quantify desired products, intermediates, and impurities  
95 inline across various stages of a multi-step synthetic pathway.[33] Furthermore, they  
96 have successfully applied artificial neural networks (ANN) for processing NMR and  
97 UV/vis spectra of multiple components.[34] Besides enhancing Bayesian optimization,  
98 inline analysis is also anticipated to improve the efficiency of traditional optimization  
99 techniques, such as kinetic modeling.

100

101 In this study, we focused on the synthesis of 2-amino-3-methylbenzoic acid, a key  
102 intermediate for the pesticide chlorantraniliprole, to serve a case study for optimizing  
103 heterogeneous continuous flow hydrogenation. We designed a continuous flow  
104 hydrogenation system equipped with an FTIR for inline monitoring and analysis of  
105 reaction data. Multi-objective Bayesian optimization and kinetic modeling were  
106 conducted in parallel to optimization the reaction process, allowing us to compare the  
107 advantages and limitations of these two approaches directly.

108

## 109 **Materials and Methods**

110 **Materials.** 3-Methyl-2-nitrobenzoic acid (98%, MNA), 2-amino-3-methylbenzoic acid  
111 (98%, AMA), methanol (MeOH, analytical grade), and silica (100-200 mesh) were  
112 purchased from Sinopharm Chemical Reagent Co., Ltd. (SCRC) without further  
113 purification. Hydrogen (H<sub>2</sub>, 99.999%) and nitrogen (N<sub>2</sub>, 99.999%) were purchased from  
114 Hangzhou Jingong Special Gas Co., Ltd. The catalyst Pd@SBA-15 was synthesized  
115 following the method reported in our previous work.[35]

116

117 **Experimental setup.** Metering pump (JJRZ-10004F) was purchased from Hangzhou  
118 Jingjin Technology Co., Ltd. High performance liquid chromatography (HPLC)  
119 column (5 mm in diameter and 50 mm long) was purchased from Dalian Baijia Lida  
120 Technology Co., Ltd. Check valve, T-joint mixer, temperature sensor, pressure gauge,  
121 back pressure regulator, and stainless steel tube (1/8" and 1/16" outside diameter) were  
122 purchased from Beijing Xiongchuan Technology Co., Ltd. Polytetrafluoroethylene  
123 (PTFE) tube (1/8" and 1/16" outside diameter) was purchased from Nanjing Runze  
124 Fluid Control Equipment Co., Ltd. Inline FTIR (ReactIR 702L) was purchased from  
125 Mettler Toledo Technology Co., Ltd. Mass flow controller (D07) was purchased from  
126 Beijing Sevenstar Flow Co., Ltd. Water bath was purchased from Heidolph Instruments  
127 Co., Ltd. Experimental setup was shown in Figure S1a.

128

129 **Heterogeneous continuous flow hydrogenation system.** MNA was dissolved in  
130 MeOH and pumped into the continuous flow system through a metering pump. The  
131 flow rate and pressure of H<sub>2</sub> involved in the hydrogenation were controlled by adjusting  
132 the mass flow controller and the pressure gauge, respectively. The gas and liquid flow  
133 tubes were fitted with check valves to avoid backflow. The hydrogen and MNA solution  
134 were mixed in a T-joint mixer and then flowed into a 1.0-meter pretreatment tube to  
135 reach hydrogenation temperature. The synthesized catalyst Pd@SBA-15 was pre-  
136 loaded in an HPLC column and the remaining space was filled with silica. Then, sieve

137 plates were installed at the ends of the HPLC column as a micro pack-bed reactor  
138 (MPBR). The MPBR was placed vertically in a hot water bath, which facilitates full  
139 contact between the gas-liquid mixture and the catalyst. The gas-liquid mixture  
140 undergoes hydrogenation in the MPBR. The pressure of the entire system was  
141 controlled by a manual back pressure regulator. The water bath and temperature sensor  
142 accurately control the hydrogenation temperature together. The check valve and back  
143 pressure regulator ensured that the continuous flow system was isolated from air. Then,  
144 the gas-liquid mixture flowed into a 1.0-meter pretreatment tube in a cold water bath to  
145 reach 20 °C. Inline FTIR monitored the concentrations of MNA and AMA after the  
146 hydrogenation in real-time. The tubes for solution flow through the inline FTIR were  
147 PTFE tubes and all other tubes were stainless steel tubes. The entire heterogeneous  
148 continuous flow hydrogenation system was controlled by adjusting the reaction  
149 parameters, such as temperature, flow rate, and hydrogen pressure.

150

151 **Concentration determination.** Concentrations of MNA and AMA in the reaction  
152 solution were monitored in real-time by inline FTIR. ANN modeling was performed as  
153 a processing approach for inline FTIR spectra data according to the reported literature  
154 (Figure S2a).[34] Several groups of MNA and AMA in methanol solution with different  
155 concentrations were prepared and their inline FTIR spectra data were collected as  
156 training set and validation set. Here the verification set is equivalent to the test set  
157 (Table S1). Next, 5000 analog spectra are generated from a linear combination of two  
158 pure components, and Gaussian noise is added to augment the training set. Some of the  
159 data simulating the experimental process were also added to the training set and  
160 validation set. To improve the stability and performance of the ANN training phase, all  
161 spectra data were normalized. Next, during the training process, an architecture of one  
162 convolutional layer followed by dense layers was investigated. The spectra data was  
163 processed at the Conv1D convolutional layer for characteristic extraction to screen the  
164 weights of the data. Then, data dimension reduction was performed through different  
165 functions in the dense layers. Finally, the output layer outputs the predicted  
166 concentrations of MNA and AMA. Compared with the known concentration in the test

167 set, the predicted results of the model are basically consistent, indicating that the ANN  
168 model framework developed for this purpose is suitable (Figure S2b). The inline FTIR  
169 spectra data monitored during the experiment were used to quickly predict the  
170 concentrations of MNA and AMA through the ANN model. The conversion of the  
171 MNA ( $\text{Conversion}_{\text{MNA}}$ ), the yield of the AMA ( $\text{Yield}_{\text{AMA}}$ ), and the productivity of the  
172 AMA ( $\text{Productivity}_{\text{MNA}}$ ) in the continuous flow hydrogenation system were calculated  
173 through Equation 1, Equation 2, and Equation 3, respectively:

$$174 \quad \text{Conversion}_{\text{MNA}} = 1 - \frac{C_{\text{MNA}}}{C_{\text{MNA}}^0} \quad (1)$$

$$175 \quad \text{Yield}_{\text{AMA}} = \frac{C_{\text{AMA}}}{C_{\text{MNA}}^0} \quad (2)$$

$$176 \quad \text{Productivity}_{\text{MNA}} = C_{\text{AMA}} F_{\text{MNA}} M_{\text{AMA}} \quad (3)$$

177 where  $C_{\text{MNA}}^0$  was the initial concentration of the MNA in the solution;  $C_{\text{MNA}}$  and  
178  $C_{\text{AMA}}$  were the concentration of the MNA and the AMA in the collected solution,  
179 respectively;  $F_{\text{MNA}}$  was the flow rate of the MNA solution;  $M_{\text{AMA}}$  was the molar mass  
180 of the AMA.

181

182 **Multi-objective Bayesian Optimization.** MOBO was performed according to the  
183 following process in general.[36, 37] The Bayesian optimizer was initialized by the  
184 design of experiments (DoE) or the random collection of initial experimental condition  
185 parameters and results. The expectation and uncertainty of each point were predicted  
186 based on a probabilistic surrogate model generated from the initial experimental results,  
187 with the trade-off between exploration and exploitation of the response space. Among  
188 them, the exploration region had high uncertainty, while the exploitation focuses on the  
189 part with high predictive expectations. New experimental condition parameters were  
190 obtained to perform new experiments after maximizing the acquisition function. The  
191 experimental dataset was then extended and reused to train a more accurate surrogate  
192 model. This process was iterated until obtaining satisfactory reaction yield and  
193 productivity.

194

195 The initial sampling, the surrogate model, and the acquisition function as the three core



196 sections formed the MOBO model.[38, 39] Latin Hypercube Sampling (LHS) was used  
197 as the initial sampling approach to avoid the presence of excessive data aggregation in  
198 simple random sampling. The LHS divided the sampling units into different layers  
199 according to some characterization or some rules, and then extracted samples  
200 independently and randomly from the different layers. Notably, the LHS uses fewer  
201 samples than the traditional OFAT or DoE approach when the same threshold is reached,  
202 thus reducing the complexity of the calculation.

203

204 MOBO was a response surface approach to uncertainty guidance, in which the  
205 performance of the surrogate model represented the predictive accuracy of the  
206 optimizer.[36] The efficiency of the surrogate model could only be recognized if its  
207 estimations of expectation and variance were close enough to the true response  
208 surface.[36] Gaussian process (GP) was an infinite-dimensional extended function  
209 distribution frequently used as the surrogate model.[39] GP allowed the construction of  
210 joint probability distributions of variables for estimating the variance and mean of the  
211 predicted data based on the available data. The Matérn class was a commonly used class  
212 of covariance functions in GP, *via* Equation 4:[40]

$$213 \quad M_{\text{Matérn}}(x, y) = \frac{2^{1-\nu} \sigma^2}{\Gamma(\nu)} (\sqrt{2\nu} \|x - y\|)^\nu K_\nu(\sqrt{2\nu} \|x - y\|) \quad (4)$$

214 Where  $\sigma^2 > 0$  and  $\nu > 0$ ;  $\nu$ ,  $\sigma^2$ ,  $\Gamma(\nu)$ ,  $\|x - y\|$ , and  $K_\nu$  were the non-negative  
215 parameter, the output variance, the gamma function, the distance between two points,  
216 and the Bessel function, respectively.

217

218 Furthermore, the acquisition function was crucial for the desired optimization  
219 performance. Among them, probabilistic improvement (PI), expected improvement  
220 (EI), and upper confidence bound (UCB) are frequently used to tune hyperparameters.  
221 The  $q$ -noisy expected hypervolume improvement ( $q$ NEHVI) function was superior to  
222 other existing acquisition functions for MOBO, for example, it enabled one-step  
223 hypervolume maximization in both noisy and noise-free environments, *via* Equation  
224 5:[41, 42]

225 
$$\alpha_{q\text{NEHVI}}(\chi_{\text{cand}}|\mathcal{P}) = \frac{1}{\tilde{N}} \sum_{t=1}^{\tilde{N}} \text{HVI}(\tilde{f}_t(\chi_{\text{cand}}|\mathcal{P})) \quad (5)$$

226 where  $\tilde{N}$ , HVI,  $\tilde{f}_t$ ,  $\chi_{\text{cand}}$ , and  $\mathcal{P}$  were the number of samples, the hypervolume  
 227 improvement, the posterior sample, the candidate sample, and the Pareto boundary,  
 228 respectively.

229

230 In this work, the  $q\text{NEHVI}$  function was used to optimize the yield of the AMA  
 231 ( $\text{Yield}_{\text{AMA}}$ ) and the productivity of the AMA ( $\text{Productivity}_{\text{MNA}}$ ) in the continuous flow  
 232 hydrogenation system.

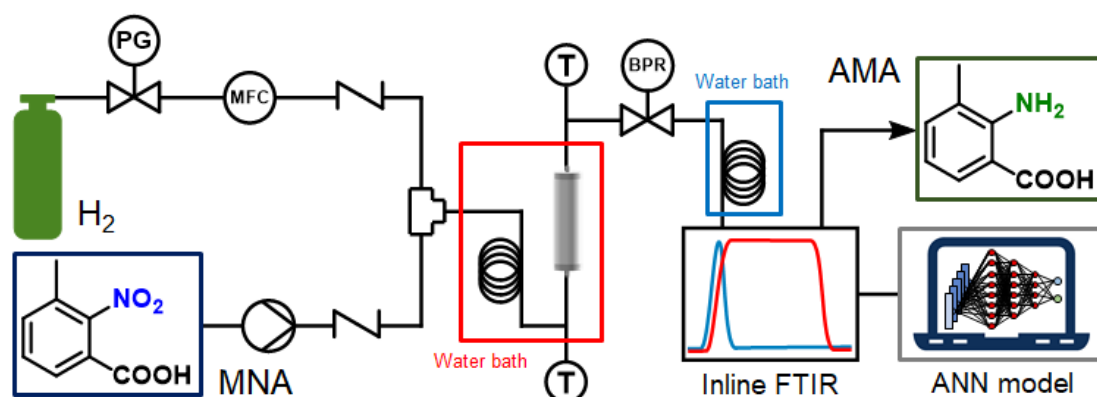
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## 234 Results and discussion

235

236 In this study, a continuous flow hydrogenation system (Figure 1) was constructed to  
 237 perform the hydrogenation of MNA using an MPBR filled with Pd@SBA-15 catalyst.  
 238 Real-time reaction monitoring was achieved through an inline FTIR instrument, the  
 239 ReactIR 702L. The data collected via inline FTIR were processed using an ANN model,  
 240 enabling the acquisition of real-time concentration for MNA and AMA. These data  
 241 points were subsequently utilized in Bayesian optimization and kinetic modeling  
 242 studies to optimize the reaction system.

243



244

245 **Figure 1.** Heterogeneous continuous flow hydrogenation of MNA and its inline analysis

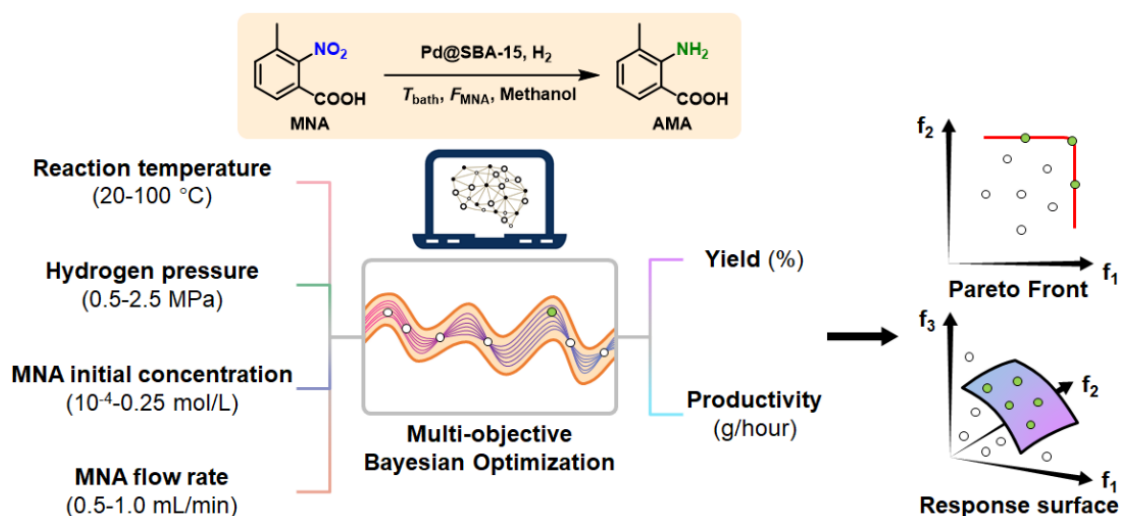
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247

248 **MOBO for reaction optimization.**

249 MOBO was applied to optimize four key reaction parameters (Figure 2): reaction  
250 temperature ( $T_{\text{bath}}$ ), hydrogen pressure ( $P_{\text{H}_2}$ ), initial MNA concentration ( $C_{\text{MNA}}^0$ ), and  
251 MNA solution flow rate ( $F_{\text{MNA}}$ ), to maximize yield and productivity. Initially, Latin  
252 Hypercube Sampling (LHS) was used to create four sets of experimental conditions,  
253 and the outcomes of these experiments formed the initial training data for a Gaussian  
254 Process (GP) surrogate model. Utilizing this GP model, the acquisition function known  
255 as qNEHVI [43] then recommended a new set of experimental conditions. With each  
256 new experiment conducted, the GP model was updated with the results, and qNEHVI  
257 continued to make further recommendations. This process was repeated iteratively until  
258 the desired levels of yield and productivity were reached. Ultimately, this method led  
259 to the identification of the optimal experimental parameters located on the Pareto Front,  
260 achieving a balance between yield and productivity.

261



262

263 **Figure 2.** The MOBO of the continuous flow hydrogenation of MNA

264

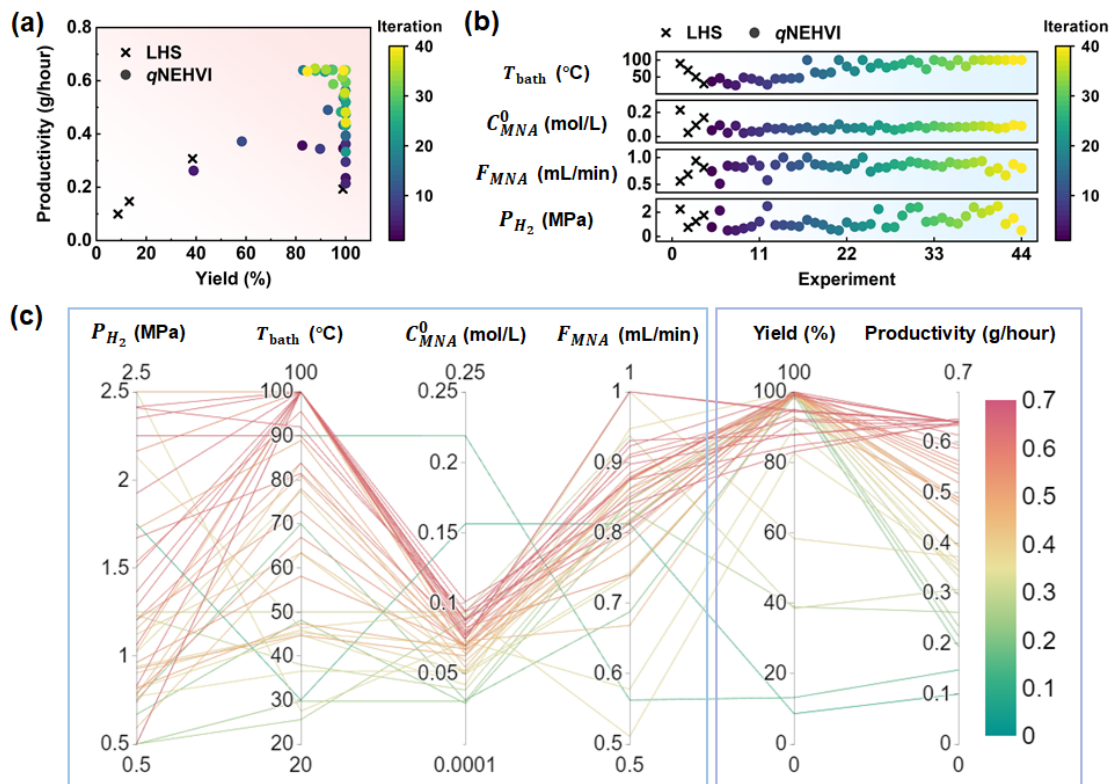
265 The parameter space for the reaction was initially delineated in Figure 2. To ensure  
266 experimental safety, the hydrogen pressure and reaction temperature were capped at 2.5  
267 MPa and 100 °C, respectively. The solubility of MNA in methanol dictated its  
268 concentration limit, setting the upper boundary at 0.25 mL/min. The residence time,  
269 crucial for reaction completion, was adjusted by modulating the solution's flow rate.

270 An excessively high flow rate could result in undue pressure build-up within the reactor  
271 tubes, while a markedly low flow rate might increase the gas phase's proportion,  
272 complicating the monitoring of MNA concentration. Consequently, the flow rate was  
273 confined to 0.5-1.0 mL/min to balance these factors.

274

275 The MOBO results are presented in Figure 3, with Figure 3a highlighting the evolution  
276 of the Pareto Front. Initial data points, marked by black crosses, were derived from LHS.  
277 The colored dots, varying in color based on iteration count, represent the data points  
278 suggested by the acquisition function. Through operation iterations with the GP model  
279 and the qNEHVI function, the data converged to form a compact Pareto Front. The data  
280 points in the upper right corner indicate the best balance between yield and productivity,  
281 showing nearly 99% yield and 0.64 g/hour productivity. The conclusive set of  
282 experimental parameters and results, which includes three sets capable of attaining  
283 these optimal levels, is detailed in Table S2.

284



285

286 **Figure 3.** Results of the MOBO campaign. (a) The Pareto Front of the yield and productivity.

287 (b) Optimization progress: each parameter *versus* experiment number. (c) Parallel coordinate

288 plot showing the interactions between experimental parameters and results.

289

290 Figure 3b demonstrates the evolution of four optimization parameters—reaction  
291 temperature, concentration, flow rate, and hydrogen pressure—throughout the  
292 optimization process. It is observed that the reaction temperature, concentration, and  
293 flow rate values gradually stabilize, indicating that the MOBO successfully identified  
294 their optimal levels. In contrast, hydrogen pressure displayed continuous fluctuations,  
295 implying uncertainty regarding its impact on the optimization goals.

296

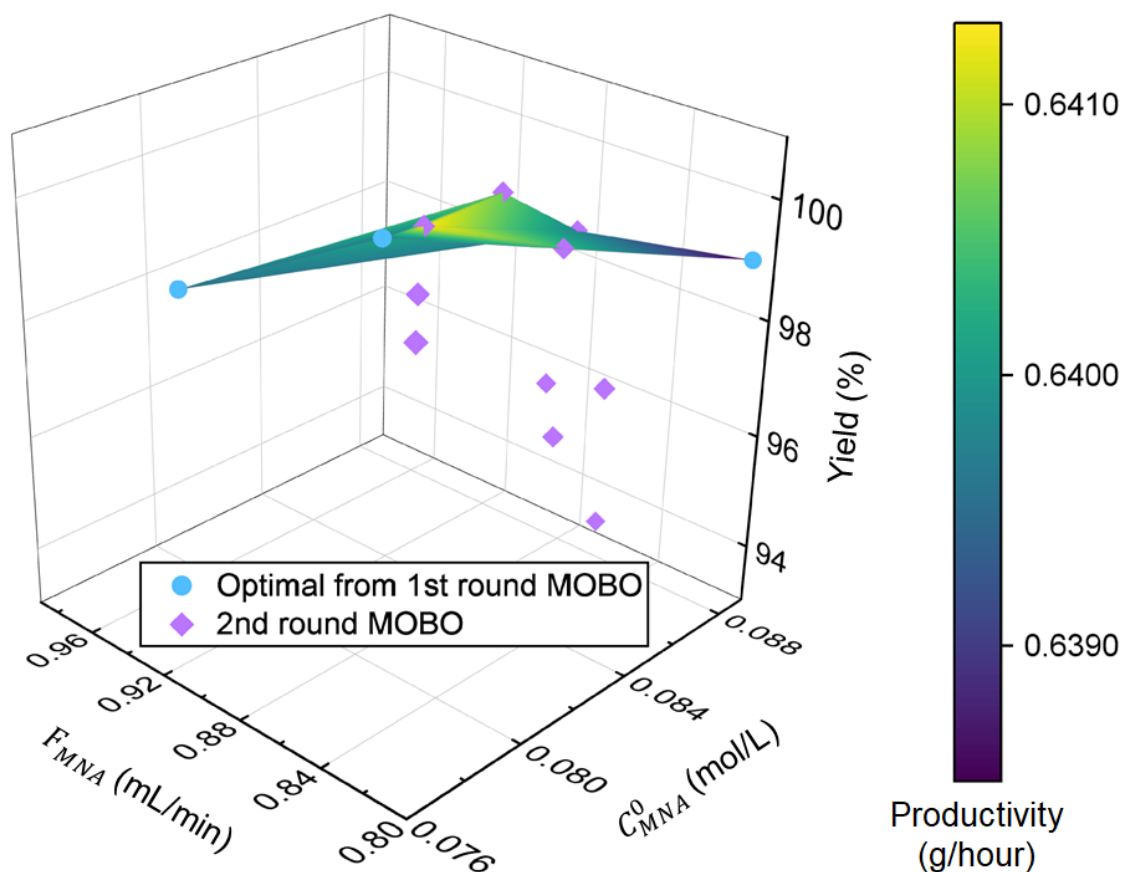
297 Further insights are provided by a parallel coordinate plot (Figure 3c), which delineates  
298 the preferred reaction parameter values as determined by MOBO for enhancing both  
299 yield and productivity. Notably, optimal reaction temperatures are concentrated around  
300 the upper boundary of 100 °C, suggesting a preference for higher temperatures to  
301 maximize yield and productivity while minimizing by-product formation. This  
302 observation highlights the advantage of multi-objective optimization, demonstrating  
303 that a 99% yield is attainable at various temperatures when yield is the sole  
304 consideration. Moreover, the plot shows that the optimal concentration and flow rate  
305 settle at approximately 0.08 mol/L and 0.86 mL/min, respectively, with neither  
306 parameter reaching its maximum or minimum limit. The analysis also indicates that  
307 hydrogen pressure does not significantly influence the optimization objectives, hinting  
308 that even the minimum hydrogen pressure level might suffice for the reaction.

309

310 For the later comparison with kinetic modeling which uses the response surface method  
311 for optimization, we performed another round of MOBO to fully explore the response  
312 surface surrounding the optimal points, with the three sets of optimal experimental  
313 parameters obtained in the previous round as the initial training set. The reaction  
314 temperature and hydrogen pressure were fixed at 100 °C and 0.5 MPa, respectively,  
315 acknowledging the positive correlation of high temperature and the optimization  
316 objectives and the negligible impact of hydrogen pressure. The exploration ranges for  
317 concentration and flow rate were narrowed to 0.0770-0.0884 mol/L and 0.8036-0.9227  
318 mL/min, aiming for a thorough investigation of this refined reaction space. After 10

319 iterations, the results (Table S3) show that an additional four sets of experimental  
320 parameters are able to achieve the same optimal yield and productivity as the original  
321 three sets. Mapping these seven optimal sets of experimental data onto the reaction  
322 space form a response surface with an irregular shape (Figure 4).

323



324

325 **Figure 4.** 3D response surface of objective values formed through fitting the optimal  
326 experimental data underwent two rounds of MOBO. The blue points represent the three  
327 optimal data points from the first round, which were used as the initial training set for the  
328 second round. The purple points indicate all the data points gathered during the second round.

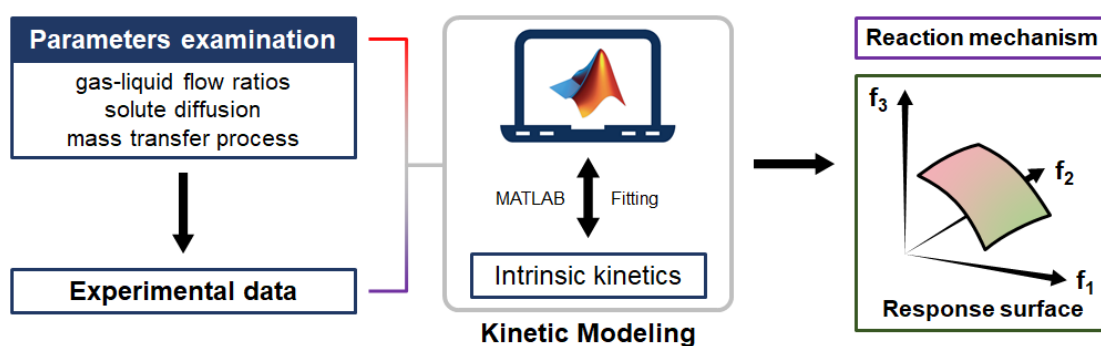
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### 330 **Kinetic modeling for reaction optimization.**

331 Concurrently, kinetic modeling was conducted in alignment with our previously  
332 established intrinsic kinetic modeling methodology[18]. It is important to note that non-  
333 chemical kinetic phenomena, such as gas-liquid interfacial area, fluid kinetics, solute  
334 diffusion, and mass transfer limitations, can significantly impact the accuracy of  
335 intrinsic kinetic modeling.[18] A comprehensive examination of reaction parameters  
336 (e.g., gas-to-liquid flow ratio, mean residence time) was performed to minimize these

337 effects, ensuring that the reaction rate data were obtained under conditions of kinetic  
338 control (see **Supporting Information**). Subsequently, experimental data were  
339 collected on the flow synthesis of AMA across various temperatures and mean  
340 residence times. This data formed the basis for our kinetic modeling efforts, during  
341 which the parameters across all models were estimated using the gPROMS (PSE, UK)  
342 parameter estimation tool [44]. Upon identifying the optimal model, a detailed  
343 examination of the reaction mechanism and the scope of this optimization approach was  
344 conducted (Figure 5).

345



346

347 **Figure 5.** The overall approach for the kinetic modeling

348

349 The Langmuir-Hinshelwood-Hougen-Watson (LHHW) methodology is widely  
350 recognized for its efficacy in the kinetic modeling of heterogeneous catalytic  
351 hydrogenation processes.[44] This approach distinctively elucidates the adsorption and  
352 dissociation stages inherent in catalytic hydrogenation, thereby aiding in the accurate  
353 derivation of the reaction mechanism. The LHHW framework is structured around three  
354 pivotal stages: the Langmuir adsorption of reactants onto the catalyst surface, the  
355 surface reaction involving the adsorbed intermediates, and the subsequent desorption  
356 of the products. Typically, the surface reaction step acts as the rate-determining phase.  
357 Based on the variations in adsorption and dissociation behaviors of the reactants, four  
358 LHHW models, each characterized by unique rate expressions ( $r_{MNA}$ ), have been  
359 formulated (Table 1).

360

361

**Table 1.** The LHHW kinetic models I-IV for MNA hydrogenation

Model <sup>a,b</sup>	Description	Initial rate expression
I	competitive adsorption dissociative adsorbed H <sub>2</sub>	$r_{\text{MNA}} = \frac{-k_1 K_{\text{H}_2} K_{\text{MNA}} C_{\text{MNA}}}{\{1 + \sqrt{K_{\text{H}_2} P_{\text{H}_2}} + K_{\text{MNA}} C_{\text{MNA}} + K_{\text{AMA}} C_{\text{AMA}}\}^3}$
II	competitive adsorption nondissociative adsorbed H <sub>2</sub>	$r_{\text{MNA}} = \frac{-k_1 K_{\text{H}_2} K_{\text{MNA}} C_{\text{MNA}} P_{\text{H}_2}}{\{1 + K_{\text{H}_2} P_{\text{H}_2} + K_{\text{MNA}} C_{\text{MNA}} + K_{\text{AMA}} C_{\text{AMA}}\}^2}$
III	noncompetitive adsorption dissociative adsorbed H <sub>2</sub>	$r_{\text{MNA}} = \frac{-k_1 K_{\text{H}_2} K_{\text{MNA}} C_{\text{MNA}} P_{\text{H}_2}}{\{1 + K_{\text{MNA}} C_{\text{MNA}} + K_{\text{AMA}} C_{\text{AMA}}\} \{1 + \sqrt{K_{\text{H}_2} P_{\text{H}_2}}\}^2}$
IV	noncompetitive adsorption nondissociative adsorbed H <sub>2</sub>	$r_{\text{MNA}} = \frac{-k_1 K_{\text{H}_2} K_{\text{MNA}} C_{\text{MNA}} P_{\text{H}_2}}{\{1 + K_{\text{MNA}} C_{\text{MNA}} + K_{\text{AMA}} C_{\text{AMA}}\} \{1 + K_{\text{H}_2} P_{\text{H}_2}\}}$

<sup>a</sup> $k_1$  is the reaction rate constant of  $r_{\text{MNA}}$ ;  $K_{\text{H}_2}$ ,  $K_{\text{MNA}}$ , and  $K_{\text{AMA}}$  are adsorption equilibrium constants of H<sub>2</sub>, MNA, and AMA, respectively;  $C_{\text{MNA}}$  and  $C_{\text{AMA}}$  are the concentration of MNA and AMA in the collected solution, respectively;  $P_{\text{H}_2}$  is the pressure of H<sub>2</sub>. <sup>b</sup>Experimental conditions for data collection:  $C_{\text{MNA}}^0$  (0.1 mol/L),  $F_{\text{MNA}}$  (0.5 mL/min),  $F_{\text{H}_2}$  (20 mL/min),  $P_{\text{H}_2}$  (0.5 MPa).

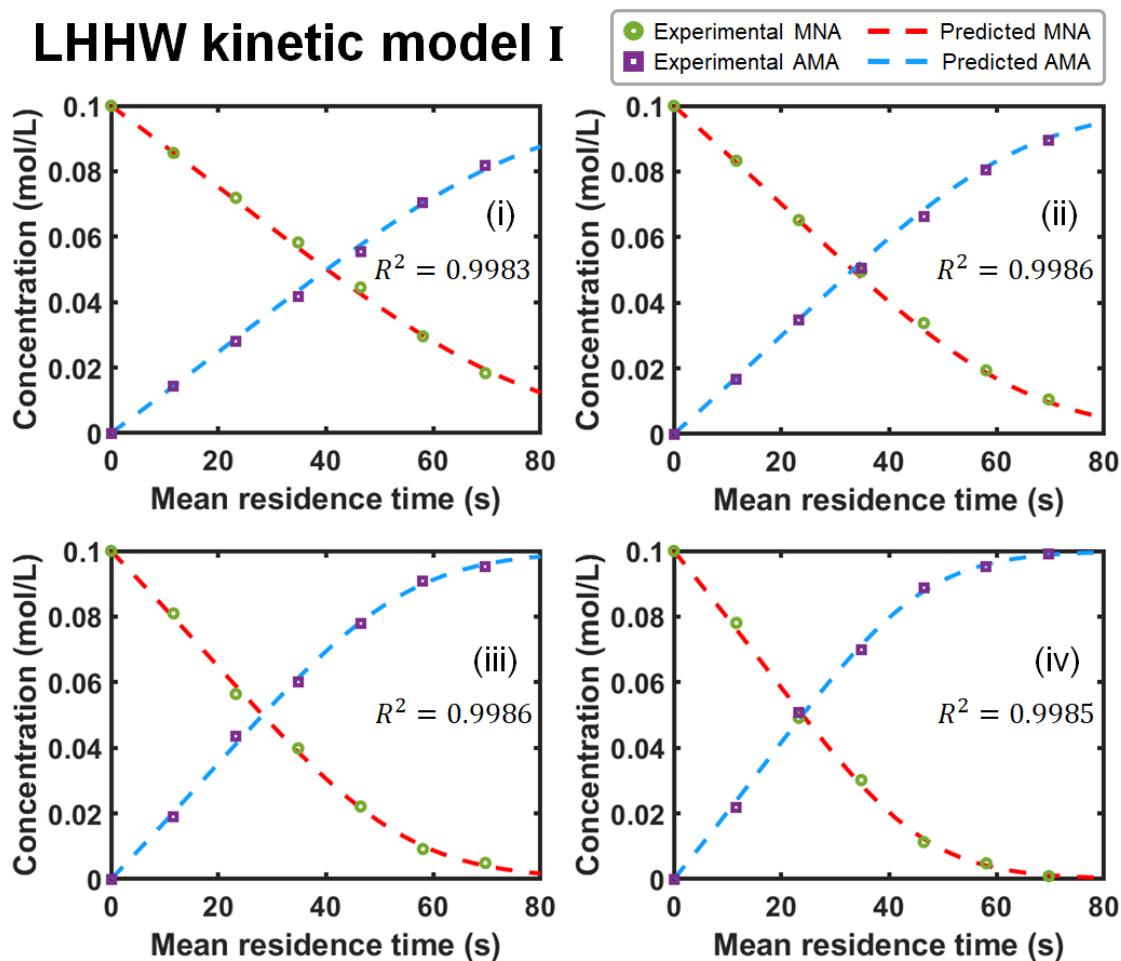
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364 The experimental data obtained were subsequently employed to fit these LHHW rate  
 365 expressions as objective functions in MATLAB, achieved through the fmincon function  
 366 with a Sequential Quadratic Programming (SQP) method. The ordinary differential  
 367 equations (ODEs) were solved using the ODE45 function. We fitted the kinetic models  
 368 to the experimental data obtained at the reaction temperatures of 30 °C (303.15 K), 40  
 369 °C (313.15 K), 50 °C (323.15 K), and 60 °C (333.15 K), with the results of these fittings  
 370 shown in Figure 6 and Figure S3.

371



## LHHW kinetic model I



372

373

**Figure 6.** Fitted curves of the LHHW kinetic model I: (i) 30 °C, (ii) 40 °C, (iii) 50 °C, and (iv) 60 °C.

374

375

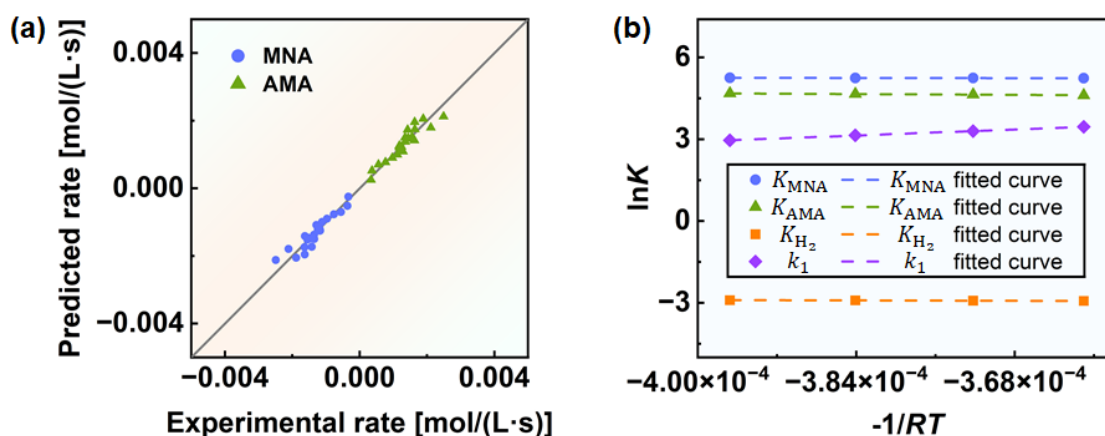
376 Each LHHW kinetic model was evaluated by estimating model parameters using the  
377 gPROMS parameter estimation tool. Based on these estimated parameters, various  
378 statistical indicators were calculated to assess the fitting performance of each model  
379 (see **Supporting Information**). The selection criteria for the optimal model included  
380 the highest value of regression coefficient ( $R^2$ ) alongside the lowest sum of squared  
381 residuals (SSR) and mean relative error (MRE). Upon comparison of these indicators  
382 in Table 2, the LHHW kinetic model I emerged as the most fitting according to these  
383 criteria, thereby establishing it as the optimal kinetic model. Furthermore, the chosen  
384 LHHW kinetic model I was further evaluated by comparing the experimental reaction  
385 rates with those predicted by the model in Figure 7a. This alignment between  
386 experimental and predicted rates underscores the robustness of the LHHW kinetic  
387 model I in simulating the reaction kinetics under study.

388

389 **Table 2.** Statistical evaluations related to experimental and predicted concentrations of MNA  
 390 for different LHHW kinetic models

Model	Stat. param.	30 °C	40 °C	50 °C	60 °C
I	$R^2$	0.9983	0.9986	0.9986	0.9985
	$SSR \times 10^5$	0.874	0.953	1.07	1.29
	MRE (%)	2.23	3.20	5.51	10.2
II	$R^2$	0.9964	0.9984	0.9979	0.9983
	$SSR \times 10^5$	1.90	1.07	1.67	1.54
	MRE (%)	4.32	4.86	6.25	11.9
III	$R^2$	0.9967	0.9982	0.9983	0.9979
	$SSR \times 10^5$	1.72	1.18	1.32	1.89
	MRE (%)	3.95	4.94	7.71	12.4
IV	$R^2$	0.9969	0.9982	0.9978	0.9979
	$SSR \times 10^5$	1.64	1.19	1.72	1.88
	MRE (%)	3.80	4.28	8.19	12.7

391



392

393 **Figure 7.** (a) Comparison of experimental reaction rate and predicted reaction rate using the  
 394 LHHW kinetic model I. (b) Fitted curves of Arrhenius equation for LHHW model I.

395

396 The kinetic and thermodynamic parameters of the LHHW model I are determined and  
 397 shown in Table 3. The  $k_1$ ,  $K_{H_2}$ ,  $K_{MNA}$ , and  $K_{AMA}$  were obtained through kinetic  
 398 model fitting in MATLAB. The activation energy ( $E$ ) and the pre-exponential factor  
 399 ( $k_0$ ) were obtained from the Arrhenius equation (Equation 6) and its variant (Equation

400 7) for the surface reaction:

$$401 \quad k = k_0 e^{-\frac{E}{RT}} \quad (6)$$

$$402 \quad \ln k = -\frac{E}{RT} + \ln k_0 \quad (7)$$

403 where  $k$  is the reaction rate constant;  $k_0$  is the pre-exponential factor;  $E$  is the  
404 activation energy;  $R$  is the universal gas constant;  $T$  the temperature in K.

405

406 Table 3. Various kinetic and thermodynamic parameters of the LHHW model I

Para.	Temperature (K)				Thermodynamic parameters	
	303.15	313.15	323.15	333.15	$E$ (kJ mol <sup>-1</sup> )	$k_0$ (mol L <sup>-1</sup> s <sup>-1</sup> )
$k_1$ (mol L <sup>-1</sup> s <sup>-1</sup> )	19.290	23.072	26.972	31.590	13.740	$4.503 \times 10^3$

Para.	Temperature (K)				Thermodynamic parameters	
	303.15	313.15	323.15	333.15	$E_a$ (kJ mol <sup>-1</sup> )	$K_0$
$K_{MNA}$ (L mol <sup>-1</sup> )	190.988	189.882	188.768	187.866	-0.465	158.823 (L mol <sup>-1</sup> )
$K_{AMA}$ (L mol <sup>-1</sup> )	107.645	105.101	102.990	100.703	-1.849	51.685 (L mol <sup>-1</sup> )
$K_{H_2}$ (MPa <sup>-1</sup> )	0.0548	0.0543	0.0538	0.0533	-0.776	0.0403 (MPa <sup>-1</sup> )

407

408 The activation energy of the surface reaction is 13.740 kJ/mol through the linear fitting  
409 method, and the adsorption heats ( $E_a$ ) of H<sub>2</sub>, MNA, and AMA are all negative,  
410 signifying that the adsorption process is exothermic (Figure 7b). The adsorption heat  
411 ( $E_a$ ) and the pre-exponential factor ( $K_0$ ) were also estimated using the Arrhenius  
412 equation (Equation 8) for the adsorption process:

$$413 \quad \ln K = -\frac{E_a}{RT} + \ln K_0 \quad (8)$$

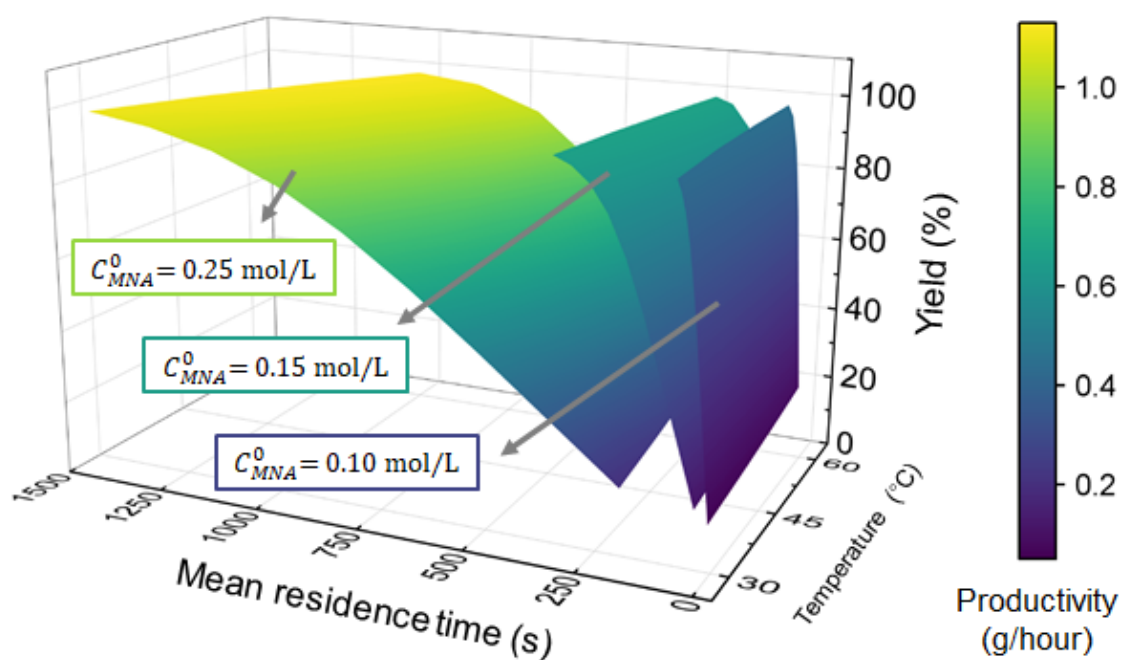
414 where  $K$  is the adsorption equilibrium constant;  $K_0$  is the pre-exponential factor;  $E_a$   
415 is the adsorption heat;  $R$  is the universal gas constant;  $T$  the temperature in K.

416

417 To achieve reaction optimization via intrinsic kinetic modeling, response surfaces of  
418 the kinetic model were generated across various MNA concentrations, with yield  
419 represented on the Z axis and the productivity illustrated through a color map (Figure

420 8). While it is feasible to attain a yield nearing 99% at a lower concentration and reduced  
421 mean residence time, optimizing both productivity and yield simultaneously—targeting  
422 the yellow area in Figure 8—requires an increase in both the mean residence time and  
423 MNA concentration. Under these optimized conditions, it is possible to achieve both a  
424 yield and productivity of up to 99% and 1.13 g/hour, respectively. The results predicted  
425 by the kinetic model at the optimized conditions were all confirmed by experimental  
426 results (Table S4).

427



428

429 **Figure 8.** 3D response surface of objective values formed through the LHHW kinetic model I

430

### 431 **Comparison of MOBO and kinetic modeling.**

432 Table 4 presents the optimal reaction parameters and results from the two optimization  
433 approaches. The MOBO achieved an optimal yield of approximately 99% and  
434 productivity of 0.64 g/hour, respectively. Conversely, kinetic modeling achieved a  
435 similar optimal yield (~99%) but attained higher productivity (1.13 g/hour). The  
436 variation in optimal productivity was attributed to the distinct strategies for  
437 manipulating the residence time of the two approaches. MOBO modulated the  
438 residence time by adjusting the solution's flow rate, keeping the catalyst mass constant

439 within the MPBR. This approach was favored because MOBO was implemented in an  
 440 automated manner, discouraging altering the catalyst quantity in the MPBR. Manually  
 441 increasing the catalyst quantity could further increase productivity. Conversely, the  
 442 intrinsic kinetic modeling methodology, which presumes that non-chemical kinetics  
 443 phenomena (e.g., mass transfer, solute diffusion, flow dynamics) minimally impact the  
 444 reaction, does not permit free variation of the flow rate but requires adjustment of the  
 445 catalyst mass to maintain a constant solution flow rate.

446

447 Table 4. The optimal reaction parameters and results of MOBO and kinetic modeling

Data type <sup>a</sup>	$C_{MNA}^0$ (mol/L)	$F_{MNA}$ (mL/min)	Yield (%)	Productivity (g/hour)
MOBO	0.0770	0.9227	99.25	0.6396
	0.0805	0.8763	99.99	0.6397
	0.0884	0.8036	99.09	0.6385
	0.0854	0.8547	99.18	0.6404
	0.0868	0.8648	99.08	0.6398
	0.0846	0.8741	99.99	0.6407
	0.0817	0.8733	99.99	0.6413

Data type <sup>b</sup>	$T_{bath}$ (°C)	MRT (s)	Predicted data		Experimental data	
			Yield (%)	Productivity (g/hour)	Yield (%)	Productivity (g/hour)
Kinetic modeling	60	980	99.16	1.1242	99.98	1.1335
	55	1050	99.02	1.1226	99.92	1.1328
	50	1150	99.02	1.1226	99.95	1.1332
	45	1280	99.11	1.1237	99.93	1.1329
	40	1400	99.06	1.1231	99.91	1.1327

<sup>a</sup>Experimental conditions for data collection:  $T_{bath}$  (100 °C),  $P_{H_2}$  (0.5 MPa). <sup>b</sup>Experimental conditions for data collection:  $C_{MNA}^0$  (0.25 mol/L),  $F_{MNA}$  (0.5 mL/min),  $F_{H_2}$  (20 mL/min),  $P_{H_2}$  (0.5 MPa).

448

449 Regarding optimization efficiency, MOBO excels in swiftly identifying the optimal  
 450 reaction space by utilizing the Pareto Front. With an additional round of optimization,  
 451 it effectively narrowed down the search to the seven promising parameter combinations  
 452 (Figure 4). These selected points are subsequently used to construct the response  
 453 surface for the optimization objectives, effectively mapping out the optimal reaction  
 454 space from a broader set of possibilities. The integration of inline FTIR with MOBO  
 455 enhances its capability to optimize reactions rapidly and with high precision. In contrast,

456 kinetic modeling requires significant time and resources. The process involves the  
457 collection of large amounts of experimental data, the fitting of kinetic equations, and  
458 the derivation of scientific models. This approach is more labor intensive and slower  
459 than the simplified and data-saving process of MOBO.

460

461 Although kinetic modeling is may not be as efficient as MOBO for reaction  
462 optimization, it offers a distinct advantage in terms of understanding reaction kinetics.  
463 While Bayesian Optimization is engineered for rapid convergence to optimal solutions,  
464 it lacks the capability to generate a comprehensive response surface across the full  
465 reaction space (Figure 4). In contrast, kinetic modeling allows for the calculation of a  
466 spectrum of optimal reaction parameters by leveraging predictions from scientific  
467 models. The response surfaces derived from kinetic modeling (Figure 8) provide  
468 valuable insights into reaction optimization by offering a visual representation of the  
469 entire reaction space, thereby highlighting trends and delineating the boundaries of  
470 optimization possibilities [7, 45].

471

## 472 **Conclusions**

473 In this work, we developed a heterogeneous continuous flow hydrogenation system  
474 enhanced with real-time inline infrared monitoring utilizing an ANN model for  
475 optimizing the synthesis of the crucial pesticide intermediate, AMA. This work  
476 systematically undertook and compared MOBO and kinetic modeling approaches to  
477 optimize the reaction process. In only 40 iterations, MOBO efficiently identified the  
478 Pareto optimal parameter combinations within the vast and intricate reaction space. The  
479 compromise between yield and productivity was impressively achieved. Conversely,  
480 intrinsic kinetic modeling revealed the activation energy of this hydrogenation reaction  
481 and characterized the hydrogen adsorption as competitive dissociative adsorption. This  
482 approach provided a comprehensive understanding of how variations in parameters  
483 affect the reaction results. Simultaneous optimization of yield and productivity was also  
484 achieved by leveraging the response surfaces generated from the kinetic model.

485

486 While MOBO is efficient and accurate in determining the optimal reaction conditions,  
487 kinetic modeling provides insight into the mechanistic details of the reaction and offers  
488 comprehensive optimization possibilities over the entire reaction space. Thus, the in-  
489 depth study of heterogeneous continuous flow hydrogenation systems benefits from the  
490 synergistic application of MOBO and kinetic modeling. This dual approach enriches  
491 our understanding of reaction optimization and provides a deeper exploration of  
492 continuous-flow hydrogenation systems, offering valuable prospects for future research  
493 and development in this field.

494

### 495 **Declaration of Competing Interest**

496 The authors declare that they have no known competing financial interests or personal  
497 relationships that could have appeared to influence the work reported in this paper.

498

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504

### 505 **Appendix A. Supplementary data**

506 Supplementary data to this article can be found online at XXXXXXXXXXXXXXXXXXXX.

507

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