

Title

Overpotential-derived thermogenesis in mitochondrial respiratory chain

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Thermogenesis, which is associated with intracellular aerobic respiration, is a fundamental function that controls the internal temperatures of living organisms. Proton leakage is considered to be correlated with thermogenesis through aerobic respiration. It is widely known that in electrochemical cells such as fuel cells, overpotentials applied to redox reactions generate heat as energy loss. Even in the electrochemical reaction system of the mitochondrial respiratory chain, a considerable amount of heat is generated by overpotential. However, the physical mechanism of thermogenesis is not yet clear. We propose a thermogenesis model based on the electrochemical overpotential of the mitochondrial respiratory chain. As a result of quantitatively estimating the value of the overpotential applied in each reaction of the mitochondrial respiratory chain, we found that 39-63% of the initial free energy in the respiratory chain was converted into heat, and the rate of thermogenesis changed depending on respiratory activity. Furthermore, that heat was intensively produced in complex IV. The overpotential-derived thermogenesis model is expected to open a research field for electrochemically elucidating mitochondrial functions.

Main text

Intracellular thermogenesis is one of the most basic biological processes required to sustain life¹⁻⁵. However, the physical mechanism of the thermogenesis has not yet been clarified experimentally or theoretically. The widely accepted theory that thermogenesis originates from proton leakage across the inner mitochondrial membrane has little experimental or theoretical basis⁵⁻⁷. However, energy loss (overpotential) in the electrochemical reaction that proceeds like a fuel cell in the mitochondrial respiratory chain is likely the cause of thermogenesis. In enzyme complexes I-IV in the respiratory chain, electrons flow constantly from the oxidation of NADH

43 to the reduction of oxygen molecules. Each enzyme generates heat because redox reactions
44 require an overpotential. The oxygen reduction reaction in the respiratory chain is similar to
45 that in fuel cells, where the overpotentials are converted to heat corresponding to electrical
46 energy loss (see Fig. 1a, b, c). Research on the mitochondrial respiratory chain reaction as a
47 fuel cell system was conducted by Bockris^{8,9} and Berry¹⁰ in the 1980s. However, the detailed
48 structure of the respiratory chain was unknown, and Mitchell's chemiosmosis hypothesis^{11,12}
49 became widespread. However, electrochemical reactions accompanied by fuel cell-like
50 overpotentials occur in the mitochondrial respiratory chain, and it is necessary to re-examine
51 the thermogenesis mechanism derived from the overpotential. The oxygen reduction reaction
52 at the fuel cell cathode requires an overpotential of 0.3 V or more to drive the reaction, even if
53 the most advanced platinum-based catalyst is used, and after the reaction is driven, it is
54 dissipated as Joule heat.

55 The theoretical treatment of fuel cell models can be regarded as an analysis of nonequilibrium
56 phenomena. In other words, it is not about dividing the free energy change of a reaction based
57 on simple thermodynamics, because the analysis of thermogenesis is related to the reaction
58 kinetics at the steady state, which corresponds to non-equilibrium thermodynamic theory. In
59 this paper, we propose an "overpotential-derived electrochemical thermogenesis model" in
60 which an electrochemical driving force or Gibbs free energy drop is used as an overpotential to
61 drive a reaction and is then dissipated as heat. Electrocatalytic reactions at the steady state when
62 an overpotential is applied are kinetic or non-equilibrium phenomena, and the generation of
63 heat is non-equilibrium energetics. In other words, the thermogenesis is related to the reaction
64 rate. In the field of chemical reactions, it is generally understood that most of the energy used
65 during a reaction is distributed during the rate-determining process. The above discussion also
66 applies to redox reaction systems in living organisms. The basic principles of the non-
67 equilibrium theory of reactions in living organisms have been developed by Prigogine,
68 Kacharski, and others⁹. However, considering that energy transfer based on specific biological
69 reaction systems remains a major problem, it is also a way to elucidate the physics of life
70 phenomena.

71 In this study, we verified that the majority of thermogenesis in the mitochondrial respiratory
72 chain is derived from the overpotential. Furthermore, we verified that the energy balance
73 between overpotential-induced thermogenesis and ATP production is modulated by the reaction
74 rate of the respiratory chain, that is, respiratory activity. Specifically, we used literature data
75 from electrochemical experiments in our proposed overpotential-thermogenesis model to
76 estimate the amount of thermogenesis, and the calculated ratio of thermogenesis relative to the
77 Gibbs free energy of oxygen reduction reaction was compared with the generally reported ratio
78 of thermogenesis (40–60 %) ^{13,14}. We also clarify whether it is necessary to consider frictional
79 heat due to proton movement within the membrane. Electrochemical experimental data are now
80 available regarding the enzymatic reaction system of the mitochondrial respiratory chain.
81 Therefore, it is possible to discuss the magnitude of the overpotential at any current value and
82 the thermogenesis derived from the overpotential. In this study, we estimated the relationship
83 between the overpotential and current for the steady state of respiratory chains I, III, and IV,
84 and estimated the sum of the total overpotential at a certain current value by considering the

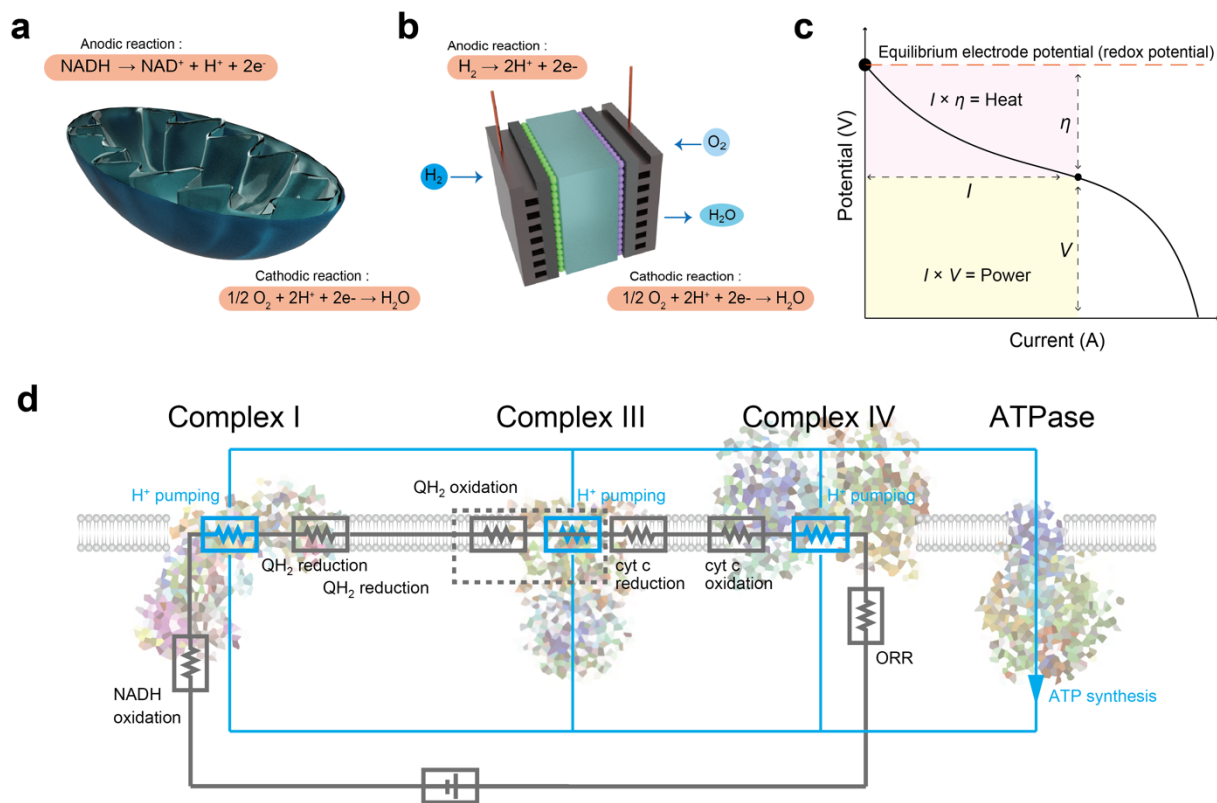


Fig. 1 | Thermogenesis model based on the electrochemical overpotential of the mitochondrial respiratory chain. Analogy of reactions in **a**, mitochondrial respiratory chain and **b**, hydrogen fuel cell. **c**, typical voltage-current curve for hydrogen fuel cell. The overpotential applied to drive the reaction is converted to heat. **d**, Reaction circuit model of a mitochondrial respiratory chain.

85 series circuit shown in Fig. 1d. This determines the relationship between the current and the
 86 sum of the overpotentials. Furthermore, the current and overpotential values were converted
 87 numerically into respiration rate and heat, respectively.

88 First, it is necessary to understand the electrochemical properties of enzymes in the
 89 respiratory chain, that is, the relationship between current j and overpotential η including the
 90 transmission coefficient α . As shown in equation (1), the relationship among η , j , α , and
 91 exchange current value j_0 follows the Butler-Volmer equation, so if α and j_0 are known, the
 92 relational expression between η and j can be derived. By substituting the current value j in the
 93 steady state into the derived equation, the overpotential η can be estimated.

94
 95

$$j = j_0 \left\{ e^{\frac{(1-\alpha)zF}{RT}\eta} - e^{-\frac{\alpha zF}{RT}\eta} \right\} \quad (1)$$

96

97 In this study, α and j_0 in complexes I, III, and IV were estimated from literature data^{15–26}.
 98 That is, α and j_0 were determined by fitting the voltammogram measured for the enzyme or

99 enzyme model system. Specifically, for the linear sweep voltammogram, a simulation was
100 performed using the Butler-Volmer equation (1), and for the cyclic voltammogram, a simulation
101 was performed using a simulator that used a combination of the Butler-Volmer and diffusion
102 equations²⁷. The fitting results are shown in Supplementary Figs. 3–16. It is important to note
103 that to perform quantitative electrochemical analysis of the mitochondrial respiratory chain, the
104 electron transfer frequency (ETF) was adopted in this study as the current value j divided by
105 the number of active sites of the enzyme. The unit of the ETF is $e^- \text{ site}^{-1} \text{ s}^{-1}$, which facilitates
106 the analysis of the oxygen consumption rate per active site of the enzyme. The values of j_0 and
107 ETF_0 are shown in Supplementary Table 3; j_0 (10^{-7} – 10^{-10} A cm^{-2}) and ETF_0 (10^{-7} – 10^{-2} $e^- \text{ site}^{-1}$
108 s^{-1}) in complex IV were quite small compared with those of the other complexes. The
109 electrochemical exchange current refers to the activity of the electrode catalyst and
110 quantitatively depends on the activation energy of the catalytic reaction. Therefore, the small
111 exchange current value of complex IV indicates that the overpotential is large and that the
112 driving force of the redox reactions in the respiratory chain system is largely complex IV.

113 Since the exchange current value ETF_0 per active site has been determined, the relationship
114 between ETF and η for each enzyme has been found. Once the ETF is determined, η for each
115 enzyme can be determined. Thus, we estimated the typical ETF values for mitochondrial
116 respiration from the literature. Supplementary Table 1 lists the average oxygen consumption
117 data of the cells obtained from the literature²⁸. This oxygen consumption was divided by the
118 number of complex IV molecules contained in each cell (Supplementary Table 2), and the
119 reaction rate per complex IV molecule was converted to the electron transfer rate per unit time
120 or ETF. Supplementary Figure 1 shows ETFs for various mitochondrial respiratory chains,
121 which were distributed at 1–10 $e^- \text{ site}^{-1} \text{ s}^{-1}$. Details of the method used to calculate the oxygen
122 consumption rate are provided in the Supplementary Material.

123 Assuming a reaction circuit consisting of complexes I, III, and IV of the mitochondrial
124 respiratory chain (Fig. 1d) and a typical ETF of 1–10 $e^- \text{ site}^{-1} \text{ s}^{-1}$, we estimated the overpotential
125 and thermogenesis of the entire circuit. Figure 1d can be regarded as a series circuit; therefore,
126 the steady-state current values flowing through each enzyme in complexes I, III, and IV are
127 equal, and the sum of these overpotentials is the overpotential of the entire circuit. Figure 2a
128 shows the reaction overpotential and the sum of overpotentials for each enzyme at $\text{ETF} = 4 e^- \text{ site}^{-1} \text{ s}^{-1}$.
129 As relatively large overpotentials, 0.4 ± 0.1 V is used for the oxygen reduction reaction
130 of complex IV, and 0.2 ± 0.1 V is used for the ubiquinone reduction of complex I. The sum of
131 the overpotentials is approximately 0.60 V. That is, approximately 54% of the net energy of the
132 respiratory chain (1.1 V) is used as the overpotential to drive the reaction, which is ultimately
133 converted to heat. Figure 2b shows the relationship between ETF and total overpotential. This
134 relationship is a function of the Butler-Volmer equation; however, as the ETF increases, the
135 overpotential and the proportion of thermogenesis to total energy also increase. An important
136 finding of this study is that the rate of thermogenesis increases with increasing oxygen
137 consumption.

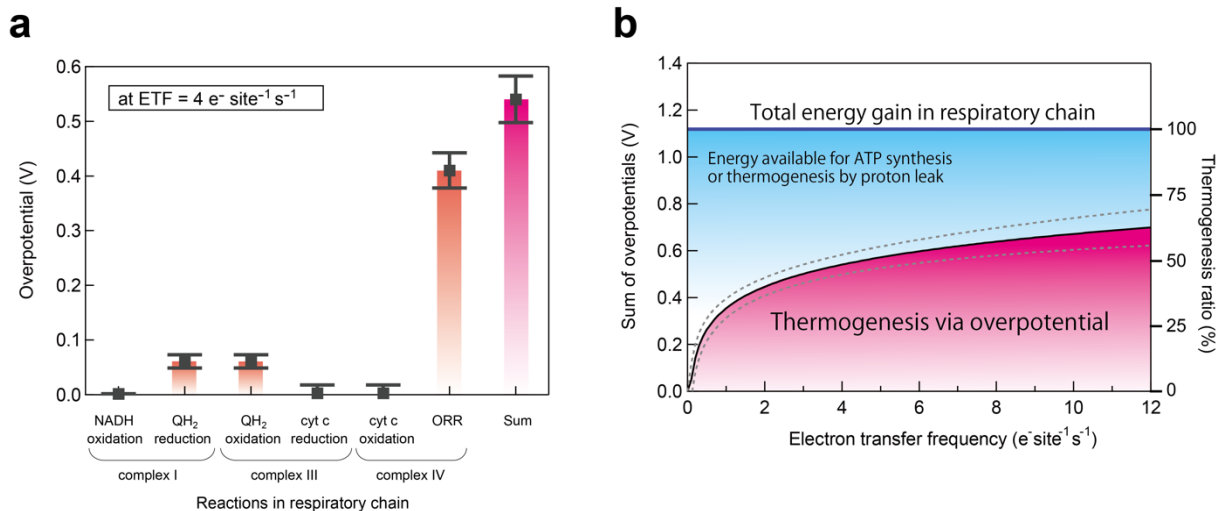


Fig. 2 | Estimated overpotentials and thermogenesis in a mitochondrial respiratory chain. **a**, Calculated overpotential in each reaction at respiration activity of $4 \text{ e}^- \text{ site}^{-1} \text{ s}^{-1}$ for NADH oxidation in complex I, ubiquinone reduction in complex I, ubiquinol oxidation in complex III, cytochrome c reduction in complex III, cytochrome c oxidation in complex IV, and oxygen Reduction Reaction (ORR) in complex IV. **b**, Estimated total overpotential applied in a respiratory chain as a function of respiration activity. The dotted lines reflect the error bqr in **a**.

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139 The first important result shown in Figure 2 is that the proportion of thermogenesis in the total
 140 respiratory chain energy gain, 39–63 % ($\text{ETF} = 1\text{--}10 \text{ e}^- \text{ site}^{-1} \text{ s}^{-1}$), estimated in the present study
 141 is in good agreement with the literature data of 40–60%^{13,14} (see the example of values for
 142 various cells in Supplementary Fig. 2). That is, the remaining 37–61 % of the Gibbs free energy
 143 change in ORR becomes available energy for ATP synthesis. Second, it was clearly shown that
 144 the amount of heat produced changed depending on the oxygen consumption rate corresponding
 145 to the ETF. It was also quantitatively demonstrated that the rate of thermogenesis increased
 146 with oxygen consumption rate. Thirdly, thermogenesis occurred mostly (more than 70%) in
 147 complex IV. Identification of heat spots is important for future molecular research on
 148 mitochondria. Fourth, it was suggested that thermogenesis due to proton transfer was negligible.
 149 In this study, we assumed that the resistance to proton transfer was zero, and that the work
 150 gained by proton pumping was equal to the energy required for ATP synthesis, that is, the
 151 assumption of zero activation energy for proton pumping. The fact that thermogenesis due to
 152 enzymatic reactions matches the actual thermogenesis indicates that this assumption is correct.
 153 Although the activation energy required for proton transfer is finite and thermogenesis is not
 154 zero, it is considered to be much smaller than the activation energy of the reactions in the
 155 respiratory chain. It should be noted that the work obtained by proton transfer and the activation
 156 energy required for proton transfer have different meanings: the former is an equilibrium value
 157 and the latter is a non-equilibrium value.

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159 Based on the results shown in Fig. 2, we estimated the distribution of work gained by
 160 the proton pumps and energy loss as thermogenesis in complexes I, III, and IV. Figure 3 shows
 161 the polarization curves of each enzyme reaction in complexes I, III, and IV plotted against the
 potential. The standard redox potential, exchange current density, and Tafel slope used for each

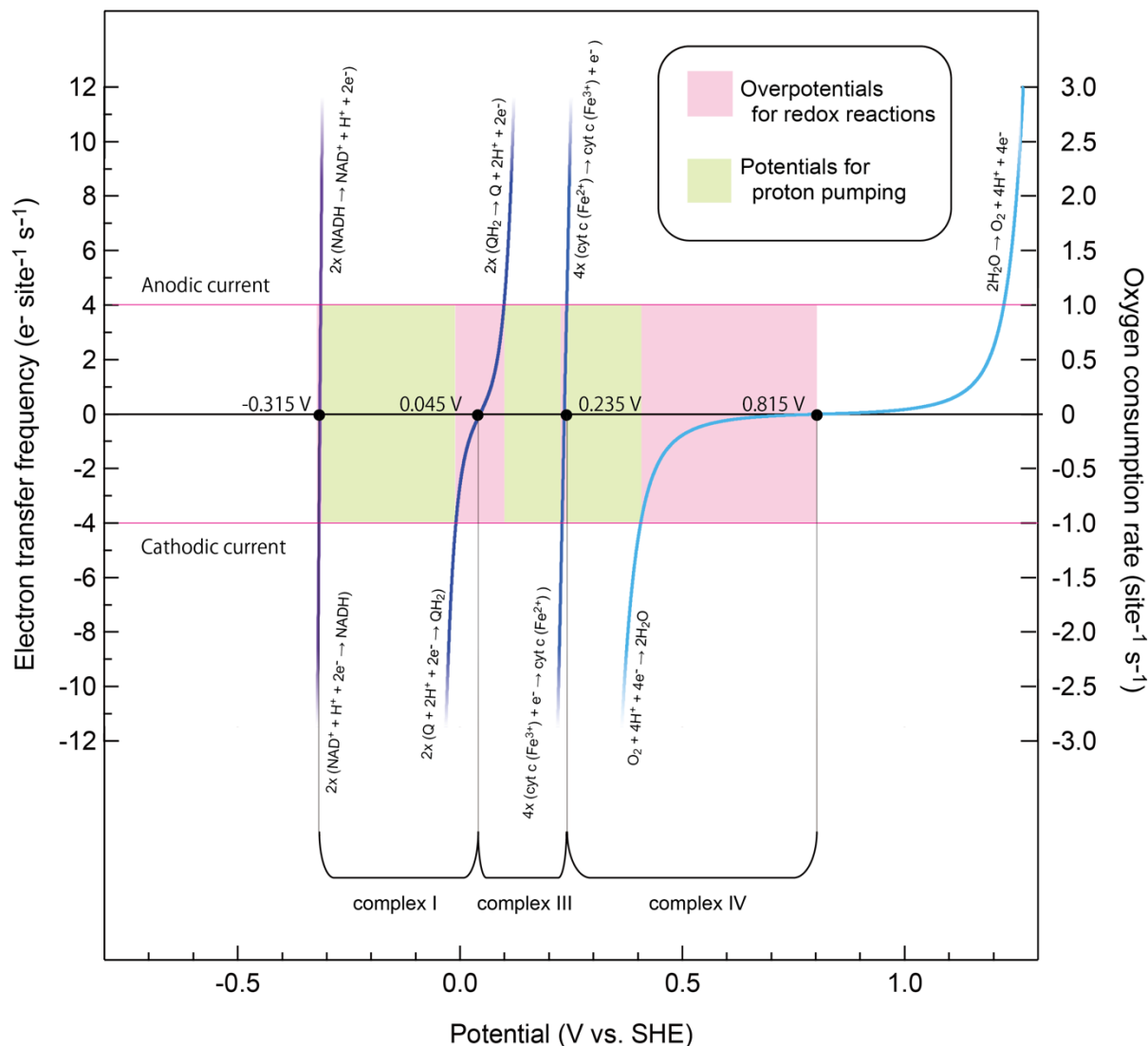


Fig. 3. | Electrochemical energy partitioning in a respiratory chain. Polarization curves of each enzymatic reaction in complex I, III, and IV based on kinetic parameters estimated in the present study. The pink and light green areas correspond to the power used for overpotential followed by heat dissipation and for proton pumping for ATP synthesis, respectively.

162 polarization curve were the average values listed in Supplementary Table 3. The positive
 163 current is the anode current resulting from an oxidation reaction, and the negative current is the
 164 cathode current resulting from a reduction reaction. In the steady state, the anode and cathode
 165 currents have the same magnitude but opposite signs. For example, $\text{NADH} \rightarrow \text{NAD}^+ + \text{H}^+ + 2\text{e}^-$
 166 in complex I corresponds to an anodic reaction, wherein a positive current flows in the
 167 polarization curve. On the other hand, $\text{Q} + 2\text{H}^+ + 2\text{e}^- \rightarrow \text{QH}_2$ in complex I is a cathodic reaction
 168 and corresponds to a negative current. In Fig. 3, the thermogenesis due to overpotential when

169 ETF = 4 e⁻ site⁻¹s⁻¹ is marked in pink, and the work gained by the proton pump or ATP
170 production is marked in yellow-green. It can be observed that heat is not significantly produced
171 in complexes I and III, and the heat part is about 20 % compared with the sum of driving forces
172 (0.235 V – (–0.315 V) = 0.55 V). In complex IV, on the other hand, the heat part was about
173 80%, indicating that most of the heat was produced in complex IV. Looking at the total area,
174 the pink and yellow-green parts were almost the same, about 50 %. That is, when ETF = 4 e⁻
175 site⁻¹ s⁻¹, 50% of the energy gain of the respiratory chain reaction is used for overpotential or
176 thermogenesis, and 50% is available for ATP production. More interestingly, Figure 3 explains
177 the literature values for the proton transfer numbers in complexes I, III, and IV. It is believed
178 that the proton transfers over the 0.15 V intermembrane potential difference. The energy gain
179 for proton pumping (pink part) was shown to be approximately 0.3 eV, 0.12 eV, and 0.12 eV
180 for complexes I, III, and IV, respectively. Therefore, the energy required to pump one proton
181 was approximately 0.15 eV. ETF = 4 e⁻ site⁻¹ s⁻¹ means that a four-electron process occurs once,
182 O₂ + 4 H⁺ + 4 e⁻ → 2 H₂O, which means that one oxygen molecule is consumed in the respiratory
183 chain. Therefore, it is possible to do work four times the potential difference in the yellow part.
184 In other words, each time one oxygen molecule is consumed, work of 1.2 eV, 0.48 eV, and 0.48
185 eV is done in respiratory chains I, III, and IV, respectively, corresponding to 8, 3.2, and 3.2
186 protons transported through the membrane. These numbers of protons are close to those
187 reported in the literature. However, these numbers varied slightly depending on the respiration
188 rate.

189 In this study, we propose an overpotential-derived thermogenesis model in which the
190 Gibbs free energy change of NADH oxidation through aerobic respiration is used to drive the
191 reaction of the respiratory chain and is then dissipated as heat. The kinetic parameters of the
192 enzymatic reactions in the respiratory chain were analyzed using previously reported data, and
193 the sum of the overpotentials in a series circuit of the respiratory chain was quantitatively
194 estimated. As a result, the ratio of thermogenesis to total energy was 39-63 %, which is
195 consistent with literature values. We also found a relationship between the rate of oxygen
196 consumption, the rate of thermogenesis, and the division of energy used for thermogenesis and
197 proton pumping in each complex. The above results indicate that the function of the respiratory
198 chain in mitochondria can be analyzed in detail, and it is expected that the mechanism of
199 respiratory function will be further elucidated from the perspective of chemical reactions in the
200 future.

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269 **Funding:**

270 This work was supported by a JSPS Grant-in-Aid for Scientific Research (KAKENHI) Grant
271 Number 23H05459, JST establishment of university fellowships for the creation of science
272 technology innovation Grant Number JPMJFS2106, and the TRiSTAR Program, a Top Runner
273 Development Program Engaging Universities, National Labs, and Companies. K.T.
274 acknowledges Dr. Naohito Otani, Mr. Kennichi Kazama, Mr. Yuichi Enomoto, and Dr. Satoshi
275 Harashima for fruitful discussions and funding aid through Academist crowdfunding.

276

277 **Author contribution**

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279 Author contributions: J.N. conceived thermogenesis by overpotential. K.T. conceived an
280 electrochemical circuit model for the respiratory chain. J.N. and K.T. designed analytical
281 protocols. N.A.P.N. performed the analyses and the calculations. N.A.P.N., M.Y., J.N. and K.T.
282 discussed the results and prepared the manuscript for publication.

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284 **Competing interests:** The authors declare no conflict of interest.

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