Versatile Biaryls and Fused Aromatics through Oxidative Coupling of Hydroquinones with (Hetero)Arenes

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Abstract: Hydroquinones bearing an electron-withdrawing group at the C2-position effectively underwent oxidative coupling with indoles or arenes in the presence of 2,3-dichloro-5,6-dicyano p-benzoquinone (DDQ) and FeCl₃ to give the corresponding biaryls. Indole-based products were further converted into tetracyclic aromatics using DDQ and FeCl₃. Thiophene derivatives were also applicable to give the tetracyclic aromatics, possessing luminescent properties.

Introduction

Biaryls (compounds with benzene-benzene, benzene-indole, etc. units) and multi-fused aromatic heterocycles are the basic backbones of biologically active substances, natural products, and functional materials such as organic light-emitting diodes (OLEDs) (Figure 1).¹⁻⁴ For example, fluvastatin¹ (a HMG-CoA reductase inhibitor that is used to treat hypercholesterolemia) and azilsartan² (an angiotensin II receptor blocker used to treat hypertension) have a biaryl moiety in their structure. Azonazine,³ isolated from a fungus in the Hawaiian marine sediments, has a tetracyclic fused dihydrobenzofuran-indoline moiety. Benzofuran-indole-fused tetracycle A^4 is expected to be a raw material for OLEDs. Therefore, it is important to develop efficient and systematic synthetic methods to construct these highly functionalized aromatic derivatives.



Figure 1. Structures of some useful compounds bearing biaryl and multi-fused aromatic backbones.

Hydroquinone can be easily modified by the Friedel-Crafts type reaction to the corresponding C2-functionalized hydroquinone (e.g., electron-withdrawing group substituted at the C2 position; 1, Scheme 1C).⁵ Moreover, benzoquinones (2), which are the oxidized forms of hydroquinones, can undergo nucleophilic addition on their a,bunsaturated carbonyl moieties to give the corresponding benzene-fused products in a stepwise manner from hydroquinone.⁶ On the other hand, tandem reactions are valuable as environmentally friend methods, as they do not require isolation and purification of reaction intermediates, thereby reducing the amount of wastes generated during the isolation of these intermediates.⁷ Particularly, one-pot oxidative functionalizations of hydroquinones can be a powerful and straightforward tool to synthesize versatile aromatic products. Masson⁸ and Jørgensen⁹ have reported the asymmetric and oxidative one-pot reactions of hydroquinones with enamines and aliphatic aldehydes to construct dihydrobenzofuran derivatives (Scheme 1A). Furthermore, Zhong et al. have recently developed the one-pot synthesis of tetracyclic aromatics from 2-methoxycarbonyl hydroquinone (1a) and indoles, without the isolation of any reaction intermediates, in the presence of copper and cobalt co-catalysts under atmospheric molecular oxygen (Scheme 1B).¹⁰ This transformation is realized by well-designed co-catalytic system, and thus considerably environmentally benign method to obtain cyclic compounds. However, to the best of our knowledge, there are no reports on the oxidative one-pot synthesis of biaryls from hydroquinones, bearing some electron-withdrawing groups (2-methoxycarbonyl, 2acetyl and 2-foymyl).

Herein, we report a novel oxidative coupling reaction of hydroquinones 1 with indole and electron-rich benzene derivatives to construct highly functionalized biaryls 3 and 4 in the presence of 2,3-dichloro-5,6-dicyano-*p*-benzoquinone (DDQ) and FeCl₃ as an oxidant and Lewis acid, respectively (Scheme 1C). Benzene-indole type biaryls **3** underwent further oxidative cyclization to benzofuran-indole-fused tetracyclic aromatics **5** in a stepwise manner. Benzofuran-thiophene derivative-fused tetracycles **6** and **7** could be directly constructed from benzoquinone (**2**) in a one-pot manner. Additionally, tetracyclic products **5**–**7** exhibited luminescence.



Scheme 1. Oxidative couplings of hydroquinones.

Results and discussion

First, the oxidative coupling of 1a with indole was investigated (eq. 1). The oxidation of 1a with DDQ produced the corresponding 2-methoxycarbonyl benzoquinone intermediate 2a, which underwent the FeCl₃-catalyzed site-selective nucleophilic addition of indole at the C3 position of 2a to give the desired biaryl product 3a in 97% yield. This site-selectivity was attributed to the increased electrophilicity at the C3 position of 2a owing to the electron-withdrawing ester group substituted at the C2 position. Phenyliodine (III) diacetate (PIDA) also acted as an effective oxidant to give 3a in 95% yield. The effects of other Lewis acids and oxidants are described in Table S1.



Next, the substrate scope of the indole nucleophiles and hydroquinones was investigated in the presence of DDQ (or PIDA)¹¹ and FeCl₃, (Scheme 2). When using *N*-methyl-, *N*-tosyl-, *N*-benzyl-, 5-methoxy-, 5-fluoro-, and 5-bromo indoles as nucleophiles with **1a**, the corresponding biaryl products **3b**–**3g** were obtained in good to excellent yields. 2-Methoxycarbonyl indole was also applicable to this reaction, affording biaryl **3k** that could be transformed into indole-fused 2-chromanone **8** as an important skeleton bearing bioactivity^{12,13} by intramolecular cyclization between a hydroxy group and ester moiety under basic conditions. Furthermore, 2-acetyl- and 2-formyl hydroquinones **1b** and **1c** underwent oxidative coupling with indole to give the corresponding biaryls **3h** and **3i**, respectively. On the other hand, hydroquinone **1d** was not converted to biaryl **3j** because of the poor electrophilicity at the C3 position. Notably, electron-rich benzene derivatives could also be used instead of indole in the present oxidative coupling of **1a**. Anisole, 2-hydroxynaphthalene, 1,3,5-trimethoxybenzene, 1-bromo-3,5-dimethoxybenzene, and 1,4-dimethoxybenzene acted as nucleophiles to afford biaryls **4a**–**4e** in moderate to good yields.



Scheme 2. Investigation of substrate scope. ^aPIDA (1.0 equiv.) was used instead of DDQ. ^bTHF was used instead of CH₂Cl₂.

Indole-based biaryl 3a was successfully converted to tetracyclic aromatic product 5a in 91% yield in the presence of DDQ and catalytic FeCl₃ (Scheme 3A; direct path). This transformation can proceed via the oxidation of 3a to benzoquinone 9, followed by the FeCl₃-catalyzed cyclization of 9 to 5a (stepwise path). The transformation of 9 to 5a can be facilitated by the coordination of FeCl₃ as a Lewis acid to the two carbonyl moieties at the C1 position and the ester moiety at the C2 position of 9 (Scheme 3B). Reaction intermediate **B** was formed subsequently by the donation of the lone pair of electrons on the N atom of indole. The subsequent intramolecular nucleophilic attack of the carbonyl oxygen at the C4 position of **B** to the iminium moiety produced **C**. Finally, aromatization of **C** gave 5a. Compounds 3d, 3e, and 3g were also applicable as substrates in this reaction, affording the corresponding tetracyclic aromatic products 5b-5d in good yields (Scheme 3C). Using the present oxidative coupling methods, versatile biaryls and tetracyclic aromatics could be constructed. Although 5a-5d could be directly constructed by Zhong's

method in Scheme 1-B¹⁰, our methodology has the benefit to apply the coupling reaction using thiophene derivatives instead of indoles, shown in next section.



Scheme 3. Transformation of 3 to tetracyclic arene 5. LA denotes Lewis acid.

The developed method was next applied for coupling using thiophene derivatives. The oxidative coupling of **1a** with thieno[3,2-*b*]thiophene in the presence of 1.0 equiv. of DDQ and catalytic FeCl₃ directly gave tetracyclic product **6**¹⁴ in 19% yield, without the generation of biaryl **9**, unlike the case using indole (Scheme 4A). When the DDQ increased to 2.2 equiv., a complex mixture was obtained (Scheme 4B). Meanwhile, the reaction using benzoquinone **2a** as a substrate furnished **6** in 43% yield. The addition of K₂CO₃ suppressed the cyclization to give 3-thienothiophene-substituted benzoquinone **10** in 43% yield. This is because K₂CO₃ lowered the Lewis acidity of FeCl₃. The cyclization of **10** was catalyzed by FeCl₃ to afford **6** in 40% yield. Furthermore, the use of benzothiophene gave another type of tetracyclic aromatic product, **7**,¹⁵ in 67% yield. Although low to moderate yields were obtained, novel tetracyclic aromatics bearing a thiophene skeleton could be synthesized using the developed oxidative coupling reactions.





Scheme 4. Oxidative coupling with thiophene derivatives. n.d denotes 'not detected.'

Because Zhong, et al. have reported that a solution of 5 (2 × 10⁻⁵ M in toluene) shows blue-light emission at a wavelength of *ca*. 426 nm,¹⁰ we also turned our attention to the photophysical properties of newly prepared compounds 6 and 7 (Figure. 2). Therefore, we investigated the photophysical properties of **5a**, 6 and 7 in the solution (CHCl₃ and toluene) and solid states. Figure 2A shows the fluorescence spectra in CHCl₃ as a representative (the fluorescence spectra in toluene are shown in Fig. S3). The fluorescence maximum decreased in the order **5a** ($\lambda_{\text{fl}} = 441 \text{ nm}$) > 6 ($\lambda_{\text{fl}} = 420 \text{ nm}$) > and 7 ($\lambda_{\phi \text{l}} = 410 \text{ nm}$). The relative fluorescence quantum yields of **5a**, 6, and 7 in CHCl₃ were 47%, 26%, and 6%, respectively (see absorption spectra of **5a**, 6, and 7 in CH2Cl₂ in Figure S4). Among the three compounds, the longest fluorescence maximum wavelength was observed for 6 in the solid state. Compound 6 exhibited green fluorescence, with a fluorescence maximum at 520 nm (Figure 2B). The photophysical data of these compounds are summarized in Figure 2C. The results indicate that the incorporation of thienothiophene units into benzofuran extends the π -conjugation, endowing unique optical properties in the solid state.



Figure 2. (A) Normalized fluorescence spectra of 5a, 6, and 7 in CHCl₃. (B) Solid-state fluorescence spectra of 5a, 6, and 7. Insets show the photographs of 5a, 6, and 7 under 365-nm irradiation. (C) Photophysical data of 5a, 6, and 7 in solution and solid state. ^{*a*}The relative fluorescence quantum yield (Φ_{fl}) was measured upon excitation at 366 nm using quinine sulfate ($\Phi_{fl} = 55\%$ in 0.1 M H₂SO₄) as a reference material.

Conclusions

We have developed the oxidative coupling of hydroquinones bearing an electronwithdrawing group at the C2 position with (hetero)aromatics to afford biaryl products as pharmaceutically useful backbones. Furthermore, tetracyclic aromatics derived from indole and thiophene derivatives were constructed. The developed synthetic methodology can be a powerful tool for the flexible design of various polycyclic aromatics that have applications as functional luminescent materials.

Conflicts of interest

The authors declare no competing financial interest.

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Author Contributions

T.A., K.N., H.N., K.M., H.S. and S.A. contributed to organic synthesis. R.U. and S.K analyzed the luminescent property. T.N., F.M., Y.O., G.K. and K.N. performed MicroED analysis. Y.S. directed the project. T.A. and Y.S. wrote the manuscript.

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- 14 The substitution position of thienothiophene to form **6** was determined from its nanocrystals by MicroED (see SI), because a single crystal of **6** large enough for our inhouse X-ray diffractometer could not be obtained. The microcrystals of **6** were bent and plate-like, and the data quality was limited by streaking of the diffraction spots and twinning. Nonetheless, the resulting map showed the structure of **6** unambiguously. (Crystallographic coordinates of the compound **6** are deposited to CCDC (ID 2294744) and COD (3000464). MicroED raw diffraction images are available at XRDa (ID 162).) The assignment by MicroED was also supported by chemical modification of thienothiophene to di-deuterated thienothiophene and the following similar reaction as shown in Scheme 5 (see SI). Nucleophilicity of the C2 position on a thiophene is known to be higher than that of the C3 position. Probably, the nucleophilic properties of thienothiophene and thiophene are similar.
- 15 The structure was determined by NOE experiments (SI). Nucleophilicity of the C3 position on benzothiophene is known to be high, similar to the case of indole.

Electronic Supplementary Information

Versatile Biaryls and Fused Aromatics through Oxidative Coupling of Hydroquinones with (Hetero)Arenes

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1. General information.

All reactions were carried out in dry solvents under argon atmosphere. Unless otherwise noted, all substrates and solvents were purchased from commercial sources and were used without further purification. Flash column chromatography was performed with 40–50 or 63–210 μ m Silica Gel 60 N (Kanto Chemical Co., Inc.). Melting points were measured on SANSHO SMP-300 or Yanaco MP-S3. IR spectra were recorded on SHIMADZU IRAffinity-1S as a thin film on NaCl or Bruker FT-IR ALPHA/SHIMADZU IRAffinity-1 as an ATR method. ¹H and ¹³C NMR spectra were recorded on JEOL JNM-ECZ400 (¹H NMR: 400 MHz, ¹³C NMR: 100 MHz) or JEOL JNM-ECA500 (¹H NMR: 500 MHz, ¹³C NMR: 125 MHz, ²H NMR: 77 MHz) with chemical shifts reported in δ (ppm) relative to an internal standard or the residual solvent signal for ¹H (tetramethylsilane: δ = 0.00 ppm, CDCl₃: δ = 77.0 ppm, (CD₃)₂CO: δ = 29.8 ppm). High resolution mass spectra were measured on JEOL JMS-S3000 (MALDI), JEOL JMS-700 (EI, FAB) or Shimadzu hybrid IT-TOF (ESI).

2. Optimization of reaction conditions.

Table S1. Optimization in the reaction of 1a with indole

OH CO ₂ Me	1) oxidant CH ₂ Cl ₂ ,	(1.0 equiv.) rt, 1 h	OH CO ₂ Me OH 3a		
ОН 1а	2) <mark>indole</mark> (Lewis a 0°C to r	(1.5 equiv.) acid (5 mol%) t, 24 h			
Entry	oxidant	Lewis acid	NMR yield (%) ^a		
1	DDQ	FeCl ₃	97 ^b		
2	DDQ	-	25		
3 c	hloranil	FeCl ₃	25		
4	PIFA FeCl ₃		42		
5	Ag ₂ O FeCl ₃		33		
6	PIDA	$FeCl_3$	95		
7	DDQ	ZnCl ₂	80		
8	DDQ	InCl ₃	82		
9	DDQ	$BF_3 \cdot Et_2O$	31		

^{*a*} Determined by ¹H NMR using 1,1,2,2-tetrachloroethane as an internal standard. ^{*b*} Isolated yield.



Methyl 3,6-dihydroxy-2-(1*H*-indol-3'-yl)benzoate (3a)

(Entry 1)

DDQ (182 mg, 0.80 mmol) was added to a solution of methyl 2,5-dihydroxybenzoate (1a, 135 mg, 0.80 mmol) in CH₂Cl₂ (4.0 mL) at 0 °C. After being stirred for 1 h at room temperature, indole (140 mg, 1.20 mmol), FeCl₃ (6,4 mg, 0.040 mmol), and CH₂Cl₂ (2.0 mL) were successively added to the reaction mixture at 0 °C. The reaction mixture was stirred for 24 h at 0 °C to room temperature. After that, the reaction was quenched with aqueous NaHCO₃ solution. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 2/1) to afford **3a** (220 mg, 0.78 mmol) in 97% yield.

Pale yellow solid; ¹H NMR (500 MHz, CDCl₃): δ 10.36 (s, 1H), 8.50 (brs, 1H), 7.41 (d, J = 8.0 Hz, 1H), 7.30 (d, J = 8.0 Hz, 1H), 7.24 (dd, J = 8.0, 7.0 Hz, 1H), 7.17 (d, J = 9.0 Hz, 1H), 7.16 (s, 1H), 7.10 (dd, J = 8.0, 7.0 Hz, 1H), 6.99 (d, J = 9.0 Hz, 1H), 5.24 (s, 1H), 3.23 (s, 3H). Spectroscopic data of ¹H NMR was identical to that reported in the reference 1.

(Entry 2)

DDQ (45.4 mg, 0.20 mmol) was added to a solution of methyl 2,5-dihydroxybenzoate (**1a**, 33.6 mg, 0.20 mmol) in CH₂Cl₂ (1.5 mL) at 0 °C. After being stirred for 1 h at room temperature, indole (35.1 mg, 0.30 mmol) was added to the mixture at 0 °C. The reaction mixture was stirred for 24 h at 0 °C to room temperature. After that, the reaction was quenched with saturated aqueous NaHCO₃ solution. The resulting mixture was extracted with CHCl₃. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The yield was calculated by crude ¹H NMR using 1,1,2,2-tetrachloroethane (21 mL, 0.20 mmol) as an internal standard.

(Entries 3–9)

DDQ (45.4 mg, 0.20 mmol) was added to a solution of methyl 2,5-dihydroxybenzoate (**1a**, 33.6 mg, 0.20 mmol) in CH_2Cl_2 (1.5 mL) at 0 °C. After being stirred for 1 h at room temperature, indole (35.1 mg, 0.30 mmol), and Lewis acid (5 mol%) were successively added to the mixture at 0 °C. The reaction mixture was stirred for 24 h at 0 °C to room temperature. After that, the reaction was

quenched with saturated aqueous NaHCO₃ solution. The resulting mixture was extracted with CHCl₃. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The yield was calculated by crude ¹H NMR using 1,1,2,2-tetrachloroethane (21 mL, 0.20 mmol) as an internal standard.

3. Procedure in Schemes 2–4.

Experimental procedure A for Scheme 2:

DDQ (45.4 mg, 0.20 mmol) or PIDA (64.4 mg, 0.20 mmol) was added to a solution of methyl 2,5dihydroxybenzoate (**1a**, 33.6 mg, 0.20 mmol) in CH₂Cl₂ (1.0–1.5 mL) at 0 °C. After being stirred for 1–1.5 h at room temperature, indoles or arenes (0.30 mmol), and FeCl₃ (1.6 mg, 0.010 mmol) were successively added to the mixture at 0 °C. The reaction mixture was stirred for 24 h at 0 °C to room temperature. After that, the reaction was quenched with saturated aqueous NaHCO₃ solution. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel to afford biaryl **3b–3k** and **4a–4e**.



Methyl 3,6-dihydroxy-2-(1'-methyl-1*H*-indol-3'-yl)benzoate (3b)

According to Experimental procedure A, DDQ (45.4 mg, 0.20 mmol), 1-methylindole (37 μ L, 0.30 mmol), and CH₂Cl₂ (1.0 mL) were used. The rection time of oxidation step was 1.5 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 3/1) to afford **3b** (52.3 mg, 0.176 mmol) in 88% yield.

Purple oil; ¹H NMR (500 MHz, CDCl₃): δ 10.27 (s, 1H), 7.39 (d, J = 8.0 Hz, 1H), 7.30 (d, J = 7.5 Hz, 1H), 7.28 (dd, J = 8.0, 7.5 Hz, 1H), 7.17 (d, J = 9.5 Hz, 1H), 7.10 (t, J = 8.0 Hz, 1H), 7.06 (s, 1H), 6.98 (d, J = 9.5 Hz, 1H), 5.24 (s, 1H), 3.88 (s, 3H), 3.24 (s, 3H). Spectroscopic data of ¹H NMR was identical to that reported in the reference 1.



Methyl 3,6-dihydroxy-2-(1'-tosyl-1*H*-indol-3'-yl)benzoate (3c)

According to Experimental procedure A, DDQ (45.4 mg, 0.20 mmol), 1-tosylindole (81.4 mg, 0.30 mmol), and CH_2Cl_2 (1.0 mL) were used. The reaction time of oxidation step was 1.5 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 3/1) to afford **3c** (52.5 mg, 0.120 mmol) in 60% yield.

White solid; m.p. 210.8–212.2 °C; IR(ATR) cm⁻¹: 3508, 3111, 2950, 1666, 1593, 1445, 1369, 1445, 1369, 1332, 1174, 1113, 1090; ¹H NMR (500 MHz, CDCl₃): δ 10.54 (s, 1H), 8.11 (d, *J* = 9.0 Hz, 1H), 7.82 (d, *J* = 9.0 Hz, 2H), 7.55 (s, 1H), 7.39 (dd, *J* = 9.0, 8.0 Hz, 1H), 7.27 (d, *J* = 9.0 Hz, 2H), 7.22 (dd, *J* = 8.0, 7.5 Hz, 1H), 7.18 (d, *J* = 9.0 Hz, 1H), 7.16 (d, *J* = 7.5 Hz, 1H), 7.03 (d, *J* = 9.0 Hz, 1H), 4.74 (s, 1H), 2.91 (s, 3H), 2.36 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 170.4, 156.2, 146.7, 145.3, 135.0, 134.9, 130.7, 129.9, 126.8, 125.5, 124.3, 123.9, 123.1, 119.8, 119.4, 117.9, 116.5, 113.8, 112.1, 51.3, 21.5; ESI-HRMS (*m*/*z*) calcd. for C₂₃H₁₉NO₆SNa [M+Na]⁺ 460.0825, found 460.0808.



Methyl 3,6-dihydroxy-2-(1'-benzyl-1*H*-indol-3'-yl)benzoate (3d)

According to Experimental procedure A, PIDA (64.4 mg, 0.20 mmol), 1-benzylindole (62.2 mg, 0.30 mmol), and CH_2Cl_2 (1.5 mL) were used. The reaction time of oxidation step was 1 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 4/1) to afford **3d** (69.2 mg, 0.185 mmol) in 93% yield.

Pale yellow solid; m.p. 130–131 °C; IR (ATR) cm⁻¹: 3483, 3029, 2950, 1664, 1608, 1453, 1439, 1332, 1226, 1176, 1126, 1025; ¹H NMR (500 MHz, CDCl₃): δ 10.35 (s, 1H), 7.99 (d, *J* = 7.5 Hz, 1H), 7.33 (m, 5H), 7.27 (m, 5H), 5.45 (d, *J* = 16.0 Hz, 1H), 5.35 (d, *J* = 16.0 Hz, 1H), 5.18 (s, 1H), 3.22 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 171.2, 155.5, 147.2, 137.2, 136.6, 128.9, 127.9, 126.6, 122.7, 122.1, 120.4, 119.5, 119.3, 118.2, 113.1, 110.0, 109.5, 51.7, 50.1; ESI-HRMS (*m/z*)

calcd. for $C_{23}H_{20}NO_4$ [M+H]⁺ 374.1387, found 374.1384.



Methyl 3,6-dihydroxy-2-(5'-methoxy-1*H*-indol-3'-yl)benzoate (3e)

According to Experimental procedure A, DDQ (45.4 mg, 0.20 mmol), 5-methoxyindole (44.2 mg, 0.30 mmol), and CH_2Cl_2 (1.0 mL) were used. The reaction time of oxidation step was 1.5 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 2/1) to afford **3e** (55.7 mg, 0.178 mmol) in 89% yield.

White solid; ¹H NMR (500 MHz, CDCl₃): δ 10.30 (s, 1H), 8.43 (s, 1H), 7.32 (d, J = 8.7 Hz, 1H), 7.20–7.16 (m, 2H), 7.01 (d, J = 8.7 Hz, 1H), 6.91 (dd, J = 8.7, 2.3 Hz, 1H), 6.71 (d, J = 2.3 Hz, 1H), 3.76 (s, 3H), 3.29 (s, 3H). Spectroscopic data of ¹H NMR was identical to that reported in the reference 1.



Methyl 3,6-dihydroxy-2-(5'-fluoro-1*H*-indol-3'-yl)benzoate (3f)

According to Experimental procedure A, DDQ (45.4 mg, 0.20 mmol), 5-fluoroindole (40.5 mg, 0.30 mmol), and CH_2Cl_2 (1.0 mL) were used. The rection time of oxidation step was 1.5 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 2/1) to afford **3f** (53.0 mg, 0.176 mmol) in 88% yield.

Gray solid; ¹H NMR (500 MHz, CDCl₃): δ 10.42 (s, 1H), 8.53 (brs, 1H), 7.36 (dd, J = 9.0, 4.5 Hz, 1H), 7.22 (d, J = 2.5 Hz, 1H), 7.17 (d, J = 9.0 Hz, 1H), 7.01–6.97 (m, 2H), 6.95 (dd, J = 9.0, 2.5 Hz, 1H), 5.13 (s, 1H), 3.29 (s, 3H). Spectroscopic data of ¹H NMR was identical to that reported in the reference 1.



Methyl 3,6-dihydroxy-2-(5'-bromo-1*H*-indol-3'-yl)benzoate (3g)

According to Experimental procedure A, DDQ (45.4 mg, 0.20 mmol), 5-bromoindole (58.5 mg, 0.30 mmol), and CH_2Cl_2 (1.0 mL) were used. The rection time of oxidation step was 1.5 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 2/1) to afford **3f** (55.1 mg, 0.152 mmol) in 76% yield.

Gray solid; ¹H NMR (500 MHz, CDCl₃): δ 10.45 (s, 1H), 8.45 (brs, 1H), 7.43 (s, 1H), 7.35–7.32 (m, 2H), 7.21–7.17 (m, 2H), 7.00 (d, J = 9.0 Hz, 1H), 4.99 (s, 1H), 3.30 (s, 3H). Spectroscopic data of ¹H NMR was identical to that reported in the reference 1.



1-(3,6-dihydroxy-2-(1*H*-indol-3'-yl)phenyl)ethan-1-one (3h)

According to Experimental procedure A, 2',5'-dihydroxyacetophenone (30.4 mg, 0.20 mmol), DDQ (45.4 mg, 0.20 mmol), indole (35.1 mg, 0.30 mmol), and THF (1.5 mL) were used. The reaction time of oxidation step was 1 h. The crude product was purified by flash column chromatography on silica gel (hexane/Et₂O = 3/2) to afford **3h** (33.1 mg, 0.124 mmol) in 62% yield.

Yellow solid; m.p. 228–229 °C; IR(NaCl) cm⁻¹: 3353, 1687, 1615, 1459, 1286, 1207; ¹H NMR (500 MHz, CDCl₃): δ 10.62 (brs, 1H), 10.45 (s, 1H), 7.51 (d, *J* = 8.0 Hz, 1H), 7.37–7.35 (m, 2H), 7.20–7.17 (m, 1H), 7.11 (d, *J* = 8.9 Hz, 1H), 7.09–7.06 (m, 2H), 6.87 (d, *J* = 8.9 Hz, 1H), 1.76 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 206.9, 153.5, 149.2, 137.6, 128.5, 126.9, 125.7, 122.9, 122.3, 121.6, 120.7, 120.3, 117.7, 112.6, 111.3, 30.7; MALDI-HRMS (*m*/*z*) calcd. for C₁₆H₁₃NO₃ [M]⁺ 267.0890, found 267.0884.



3,6-dihydroxy-2-(1*H*-indol-3'-yl)benzaldehyde (3i)

According to Experimental procedure A, 2,5-dihydroxybenzaldehyde (27.6 mg, 0.20 mmol), DDQ (45.4 mg, 0.20 mmol), indole (35.1 mg, 0.30 mmol), and CH₂Cl₂ (1.5 mL) were used. The reaction time of oxidation step was 1 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 2/1) to afford **3i** (32.3 mg, 0.128 mmol) in 64% yield. Orange solid; m.p. 248 °C (decomp.); IR(NaCl) cm⁻¹: 3384, 1640, 1579, 1458, 1277, 1178; ¹H NMR (500 MHz, (CD₃)₂CO): δ 11.44 (s, 1H), 10.70 (brs, 1H), 9.71 (s, 1H), 7.55 (d, *J* = 2.3 Hz, 1H), 7.53 (d, *J* = 8.6 Hz, 1H), 7.36 (brs, 1H), 7.33 (d, *J* = 8.0 Hz, 1H), 7.29 (d, *J* = 8.9 Hz, 1H), 7.21–7.18 (m, 1H), 7.08 (td, *J* = 7.4, 1.1 Hz, 1H), 6.88 (d, *J* = 8.9 Hz, 1H); ¹³C NMR (125 MHz, (CD₃)₂CO): δ 198.7, 157.2, 148.9, 137.4, 129.4, 127.9, 126.5, 125.1, 122.9, 120.8, 120.0, 117.4, 112.6, 107.0; MALDI-HRMS (*m/z*) calcd. for C₁₅H₁₁NO₃ [M]⁺ 253.0733, found 253.0735.



Methyl 3-(3',6'-dihydroxy-2'-(methoxycarbonyl)phenyl)-1H-indole-2-carboxylate (3k)

According to Experimental procedure A, DDQ (45.4 mg, 0.20 mmol), methyl 1*H*-indole-2carboxylate (52.5 mg, 0.30 mmol), and CH_2Cl_2 (1.0 mL) were used. The reaction time of oxidation step was 1.5 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 1/1) to afford **3k** (61.4 mg, 0.180 mmol) in 90% yield.

Pink solid; m.p. 188.7–189.6 °C; IR(ATR) cm⁻¹: 3381, 3312, 2946, 1694, 1465, 1438, 1319, 1278, 1243, 1191, 1146, 1064; ¹H NMR (500 MHz, CDCl₃): δ 10.55 (s, 1H), 9.17 (brs, 1H), 7.47 (d, *J* = 8.0 Hz, 1H), 7.38 (t, *J* = 8.0 Hz, 1H), 7.32 (d, *J* = 8.0 Hz, 1H), 7.21 (d, *J* = 9.0 Hz, 1H), 7.14 (t, *J* = 8.0 Hz, 1H), 7.05 (d, *J* = 9.0 Hz, 1H), 4.87 (brs, 1H), 3.77 (s, 3H), 3.26 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 170.9, 162.1, 156.0, 146.7, 135.9, 127.5, 126.3, 123.9, 123.1, 121.5, 121.5, 121.1, 118.9, 116.7, 112.5, 111.9, 52.2, 51.8; ESI-HRMS (*m*/*z*) calcd. for C₁₈H₁₅NO₆Na [M+Na]⁺ 364.0792, found 364.0776.



Methyl 2-hydroxy-6-oxo-6,7-dihydrochromeno[3,4-b]indole-1-carboxylate (8)

 K_2CO_3 (69.1 mg, 0.50 mmol) was added to a solution of **3k** (34.1 mg, 0.10 mmol) in THF (1.0 mL) at room temperature. After being stirred for 12 h at room temperature, distilled water was added to the mixture. The resulting mixture was filtered through a pad of Celite. The filtrate was concentrated in vacuo to afford **8** (13.9 mg, 0.0449 mmol) in 45% yield.

Pale yellow solid; m.p. 248.6–250.8 °C; IR(ATR) cm⁻¹: 3258, 2951, 1707, 1672, 1620, 1591, 1571, 1522, 1435, 1380, 1331, 1286, 1212, 1012; ¹H NMR (500 MHz, (CD₃)₂SO): δ 7.79 (d, *J* = 8.5 Hz, 1H), 7.62 (d, *J* = 8.5 Hz, 1H), 7.45 (t, *J* = 8.5 Hz, 1H), 7.37 (d, *J* = 9.2 Hz, 1H), 7.26 (t, *J* = 8.5 Hz, 1H), 6.95 (d, *J* = 9.2 Hz, 1H), 3.88 (s, 3H); ¹³C NMR (125 MHz, (CD₃)₂SO): δ 169.8, 156.9, 155.4, 141.3, 139.9, 126.3, 123.1, 122.9, 121.8, 120.9, 119.5, 118.3, 116.6, 116.5, 115.5, 113.6, 52.0; ESI-HRMS (*m/z*) calcd. for C₁₇H₁₁NO₅Na [M+Na]⁺ 332.0529, found 332.0504.



Methyl 3,6-dihydroxy-4'-methoxy-[1,1'-biphenyl]-2-carboxylate (4a)

According to Experimental procedure A, DDQ (45.4 mg, 0.20 mmol), anisole (32.4 mg, 0.30 mmol), and CH_2Cl_2 (1.0 mL) were used. The reaction time of oxidation step was 1.5 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 1/1) to afford **4a** (41.6 mg, 0.152 mmol) in 76% yield.

Orange solid; m.p. 86.9–87.9 °C. IR(ATR) cm⁻¹: 3443, 2953, 2838, 1667, 1609, 1514, 1456, 1336, 1246, 1177, 1129, 830; ¹H NMR (500 MHz, CDCl₃): δ 10.45 (s, 1H), 7.13 (d, *J* = 8.2 Hz, 2H), 7.12 (d, *J* = 9.2 Hz, 1H), 7.00 (d, *J* = 8.2 Hz, 2H), 6.96 (d, *J* = 9.2 Hz, 1H), 4.62 (s, 1H), 3.87 (s, 3H), 3.46 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 170.9, 159.2, 155.6, 146.1, 130.4, 127.8, 127.0, 122.5, 118.1, 114.3, 111.8, 55.2, 51.8; ESI-HRMS (*m*/*z*) calcd. for C₁₅H₁₄O₅Na [M+Na]⁺ 297.0733, found 297.0731.



Methyl 3,6-dihydroxy-2-(2'-hydroxynaphthalen-1'-yl)benzoate (4b)

According to Experimental procedure A, DDQ (45.4 mg, 0.20 mmol), 2-naphthol (43.2 mg, 0.30 mmol), and CH_2Cl_2 (1.0 mL) were used. The reaction time of oxidation step was 1.5 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 1/1) to afford **4b** (37.8 mg, 0.122 mmol) in 61% yield.

White solid; ¹H NMR (500 MHz, CDCl₃): δ 10.81 (s, 1H), 7.86 (d, J = 9.5 Hz, 1H), 7.84–7.82 (m, 1H), 7.37–7.35 (m, 2H), 7.30 (d, J = 9.0 Hz, 1H), 7.27 (d, J = 9.5 Hz, 1H), 7.18–7.16 (m, 1H), 7.15 (d, J = 9.0 Hz, 1H), 3.20 (s, 3H). Spectroscopic data of ¹H NMR was identical to that reported in the reference 2.



Methyl 3,6-dihydroxy-2',4',6'-trimethoxy-[1,1'-biphenyl]-2-carboxylate (4c)

According to Experimental procedure A, PIDA (64.4 mg, 0.20 mmol), 1,3,5-trimethoxybenzene (50.5 mg, 0.30 mmol), and CH_2Cl_2 (1.5 mL) were used. The reaction time of oxidation step was 1 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 1/1) to afford **4c** (47.2 mg, 0.141 mmol) in 71% yield.

White solid; m.p. 133.4–135.3 °C; IR(ATR) cm⁻¹: 3466, 2950, 2839, 1663, 1607, 1582, 1506, 1453, 1335, 1224, 1121, 1053; ¹H NMR (500 MHz, CDCl₃): δ 10.41 (s, 1H), 7.13 (d, *J* = 9.3 Hz, 1H), 6.94 (d, *J* = 9.3 Hz, 1H), 6.23 (s, 2H), 4.90 (s, 1H), 3.88 (s, 3H), 3.70 (s, 6H), 3.50 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 171.1, 161.7, 158.3, 155.6, 146.4, 122.7, 120.1, 117.9, 113.0, 105.3, 90.9, 55.9, 55.3, 51.7; ESI-HRMS (*m*/*z*) calcd. for C₁₇H₁₉O₇ [M+H]⁺ 335.1125, found 335.1104.



Methyl 2'-bromo-3,6-dihydroxy-4',6'-dimethoxy-[1,1'-biphenyl]-2-carboxylate (4d)

According to Experimental procedure A, PIDA (64.4 mg, 0.20 mmol), 1-bromo-3,5dimethoxybenzene (65.1 mg, 0.30 mmol), and CH_2Cl_2 (1.5 mL) were used. The reaction time of oxidation step was 1 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 2/1) to afford **4d** (44.4 mg, 0.116 mmol) in 58% yield.

Colorless oil; m.p. 107.1–110.2 °C; IR(ATR) cm⁻¹: 3448, 3008, 2951, 1665 1602, 1561, 1496, 1462, 1436, 1212, 1152, 1033; ¹H NMR (500 MHz, CDCl₃): δ 10.72 (s, 1H), 7.16 (d, *J* = 9.0 Hz, 1H), 7.00 (d, *J* = 9.0Hz, 1H), 6.85 (d, *J* = 2.5 Hz, 1H), 6.51 (d, *J* = 2.5 Hz, 1H), 4.47 (s, 1H), 3.86 (s, 3H), 3.68 (s, 3H), 3.52 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 170.6, 160.9, 158.8, 156.3, 145.8, 125.3, 123.1, 122.8, 118.9, 118.1, 111.7, 108.9, 98.2, 56.0, 55.6, 52.2; ESI-HRMS (*m/z*) calcd. for C₁₆H₁₅O₆BrNa [M+Na]⁺ 404,9944, found 404,9947.



Methyl 3,6-dihydroxy-2',5'-dimethoxy-[1,1'-biphenyl]-2-carboxylate (4e)

According to Experimental procedure A, DDQ (45.4 mg, 0.20 mmol), 1,4-dimethoxybenzene (41.4 mg, 0.30 mmol), and CH_2Cl_2 (1.0 mL) were used. The reaction time of oxidation step was 1.5 h. The crude product was purified by flash column chromatography on silica gel (hexane/EtOAc = 1/1) to afford **4b** (28.4 mg, 0.0933 mmol) in 47% yield.

Brown solid; m.p. 122.6–124.2 °C; IR(ATR) cm⁻¹: 3437, 2952, 2834, 1667, 1605, 1459, 1438, 1271, 1221, 1044; ¹H NMR (500 MHz, CDCl₃): δ 10.35 (s, 1H), 7.15 (d, *J* = 9.0 Hz, 1H), 6.97 (d, *J* = 9.0 Hz, 1H), 6.95 (d, *J* = 9.5 Hz, 1H), 6.91 (dd, *J* = 9.5, 3.0 Hz, 1H), 6.67 (d, *J* = 3.0 Hz, 1H), 4.95 (s, 1H), 3.77 (s, 3H), 3.72 (s, 3H), 3.47 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 170.8, 155.6, 153.9, 150.7, 145.8, 123.8, 123.2, 118.4, 116.1, 114.3, 112.5, 112.2, 56.4, 55.7, 51.8; ESI-HRMS (*m/z*) calcd. for C₁₆H₁₇O₆ [M+H]⁺ 305.1020, found 305.1022.

Experimental procedure for Scheme 3:



Methyl 2-hydroxy-6*H*-benzofuro[2,3-*b*]indole-1-carboxylate (5a)

FeCl₃ (1.4 mg, 9.0 µmol), DDQ (40 mg, 0.18 mmol), and CH₂Cl₂ (1.0 mL) were successively added to a solution of **3a** (50 mg, 0.18 mol) in CH₂Cl₂ (1.0 mL) at 0 °C. After being stirred for 24 h at 0 °C to room temperature, saturated aqueous NaHCO₃ solution was added to the mixture. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 2/1) to afford **5a** (46 mg, 0.164 mmol) in 91% yield. Yellow solid; ¹H NMR; (500 MHz, CDCl₃): δ 11.10 (s, 1H), 8.40 (s, 1H), 7.93–7.89(m, 1H), 7.56

(d, J = 8.5 Hz, 1H), 7.44–7.42 (m, 1H), 7.31–7.27 (m, 2H), 6.80 (d, J = 8.5 Hz, 1H), 4.23 (s, 3H). Spectroscopic data of ¹H NMR was identical to that reported in the reference 3.



Methyl 2-(1*H*-indol-3'-yl)-3,6-dioxocyclohexa-1,4-diene-1-carboxylate (9)

DDQ (23 mg, 0.10 mmol) was added to a solution of **3a** (28 mg, 0.10 mmol) in CH_2Cl_2 (1.0 mL) at room temperature. After being stirred for 24 h at room temperature, saturated aqueous NaHCO₃ solution was added to the mixture. The resulting mixture was extracted with CH_2Cl_2 . The combined organic layers were concentrated in vacuo to afford **9** (35 mg, 0.12 mmol) in quantitatively yield.

Purple solid; ¹H NMR (500 MHz, CDCl₃): δ 8.67 (brs, 1H), 7.51–7.47 (m, 2H), 7.39 (d, J = 8.2 Hz, 1H), 7.25–7.16 (m, 2H), 6.95–6.87 (m, 2H), 3.68 (s, 3H). Spectroscopic data of ¹H NMR was identical to that reported in the reference 4.



Methyl 2-hydroxy-6*H*-benzofuro[2,3-*b*]indole-1-carboxylate (5a)

FeCl₃ (0.75 mg, 5.0 μ mol) was added to a solution of **9** (26 mg, 0.090 mmol) in CH₂Cl₂ (0.5 mL) at 0 °C. After being stirred for 4 h at 0 °C to room temperature, saturated aqueous NaHCO₃ solution was added to the mixture. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 1/1) to afford **5a** (20 mg, 0.70 mmol) in 78% yield.



Methyl 6-benzyl-2-hydroxy-6H-benzofuro[2,3-b]indole-1-carboxylate (5b)

FeCl₃ (0.80 mg, 5.0 μ mol) and DDQ (22.7 mg, 0.10 mmol) were added to a solution of **3d** (37.3 mg, 0.10 mmol) in CH₂Cl₂ (1.0 mL) at 0 °C. After being stirred for 24 h at 0 °C to room temperature, saturated aqueous NaHCO₃ solution was added to the mixture. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 3/1) to afford **5b** (26.9 mg, 0.072 mmol) in 72% yield.

Yellow solid; m.p. 155–156 °C; IR (ATR) cm⁻¹: 3030, 2952, 1659, 1622, 1572, 1553, 1511, 1495, 1474, 1453, 1440, 1359, 1203, 1112, 1026; ¹H NMR (500 MHz, CDCl₃): δ 11.1 (s, 1H), 7.90 (d, J = 8.0 Hz, 1H), 7.52 (d, J = 8.6 Hz, 1H), 7.33–7.23 (m, 8H) , 6.76 (d, J = 8.6 Hz, 1H), 5.43 (s, 2H), 4.19 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 171.0, 159.1, 158.4, 149.9, 137.6, 128.9, 127.9, 126.8, 125.2, 121.5, 121.3, 120.9, 118.3, 110.4, 109.5, 103.8, 98.6, 52.8, 46.5; ESI-HRMS (*m/z*) calcd. for C₂₃H₁₈NO₄ [M+H]⁺ 372.1230, found 372.1228.



Methyl 2-hydroxy-9-methoxy-6H-benzofuro[2,3-b]indole-1-carboxylate (5c)

FeCl₃ (0.80 mg, 5.0 µmol) and DDQ (22.7 mg, 0.10 mmol) were added to a solution of **3e** (31.3 mg, 0.10 mmol) in CH₂Cl₂ (1.0 mL) at 0 °C. After being stirred for 24 h at 0 °C to room temperature, saturated aqueous NaHCO₃ solution was added to the mixture. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 2/1) to afford **5c** (23.3 mg, 0.075 mmol) in 75% yield.

Yellow solid; m.p. 162–163 °C; IR (ATR) cm⁻¹: 3302, 2952, 1658, 1626, 1579, 1550, 1519, 1461, 1438, 1287, 1205, 1112, 1104; ¹H NMR (500 MHz, CDCl₃): δ 11.1 (s, 1H), 8.29 (s, 1H), 7.53 (d, J = 8.6 Hz, 1H), 7.44 (d, J = 2.3 Hz 1H), 7.30 (d, J = 9.2 Hz, 1H), 6.90 (dd, J = 8.6, 2.3 Hz, 1H), 6.78 (d, J = 9.2 Hz, 1H), 4.19 (s, 3H), 3.93 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 170.9, 159.0, 158.3, 154.9, 149.8, 131.6, 124.8, 122.7, 118.3, 112.4, 109.8, 109.7, 106.0, 103.9, 100.1, 56.0, 52.9; ESI-HRMS (*m*/*z*) calcd. for C₁₆H₁₄NO₅ [M+H]⁺ 312.0866, found 312.0866.



Methyl 9-bromo-2-hydroxy-6H-benzofuro[2,3-b]indole-1-carboxylate (5d)

FeCl₃ (0.80 mg, 5.0 μ mol) and DDQ (22.7 mg, 0.10 mmol) were added to a solution of **3g** (36.2 mg, 0.10 mmol) in CH₂Cl₂ (1.0 mL) at 0 °C. After being stirred for 24 h at 0 °C to room temperature, saturated aqueous NaHCO₃ solution was added to the mixture. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 2/1) to afford **5d** (21.3 mg, 0.059 mmol) in 59% yield.

Yellow solid; m.p. 260–263 °C, IR (ATR) cm⁻¹: 3247, 1651, 1623, 1461, 1438, 1419, 1359, 1257, 1212, 1208, 1178, 1156; 1H NMR (500 MHz, (CD₃)₂SO): δ 10.30 (s, 1H), 7.99 (d, *J* = 2.3 Hz, 1H), 7.66 (d, *J* = 8.6 Hz, 1H), 7.39 (d, *J* = 8.6 Hz, 1H), 7.29 (dd, *J* = 8.6, 1.7 Hz, 1H), 6.73 (d, *J* = 8.6 Hz, 1H), 7.99 (dz, *J* = 8.6 Hz, 1H), 7.29 (dz, *J* = 8.6, 1.7 Hz, 1H), 6.73 (dz, *J* = 8.6 Hz, 1H), 7.80 (dz, *J* = 8.6 Hz, 1H), 7.29 (dz, *J* = 8.6, 1.7 Hz, 1H), 6.73 (dz, *J* = 8.6 Hz, 1H), 7.80 (dz, *J* = 8.6 Hz, 1H), 7.90 (dz, J = 8.6 Hz

= 9.2 Hz, 1H), 4.05 (s, 3H); ¹³C NMR (125 MHz, (CD₃)₂SO): δ 169.3, 159.0, 156.0, 149.9, 136.3, 124.9, 124.0, 123.5, 123.0, 117.5, 114.8, 113.3, 110.4, 107.0, 97.9, 53.1; ESI-HRMS (*m/z*) calcd. for C₁₆H₁₁NO₄Br [M+H]⁺ 359.9866, found 359.9872.

Experimental procedure for Scheme 4:



Methyl 5-hydroxy-thieno[2',3':4,5]thieno[3,2-b]benzofuran-4-carboxylate (6)

Thienothiophene (42 mg, 0.30 mmol), FeCl₃ (3.2 mg, 0.020 mmol), DDQ (45.4 mg, 0.20 mmol), and CH₂Cl₂ (1.0 mL) were added to a solution of **1a** (34 mg, 0.20 mmol) in CH₂Cl₂ (1.0 mL) at 0 °C. After being stirred for 24 h at 0 °C to room temperature, saturated aqueous NaHCO₃ solution was added to the mixture. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 5/1) to afford **6** (12 mg, 0.039 mmol) in 20% yield.

Yellow solid; m.p. 198–205 °C; IR (NaCl) cm⁻¹: 2922, 1679, 1610, 1491, 1464, 1426, 1355, 1304, 1256, 1232, 1222, 1112; ¹H NMR (500 MHz, CDCl₃): δ 11.01 (s, 1H), 7.66 (d, *J* = 9.0 Hz, 1H), 7.47 (d, *J* = 5.0 Hz, 1H), 7.34 (d, *J* = 5.0 Hz, 1H), 6.94 (d, *J* = 9.0 Hz, 1H), 4.16 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 170.4, 159.4. 152.4, 151.4, 144.1, 127.8, 124.3, 122.4, 122.1, 121.1, 119.5, 113.5, 102.5, 52.3; MALDI-HRMS (*m/z*) calcd. for C₁₄H₈O₄S₂ [M]⁺ 303.9859, found 303.9857.



Thienothiophene (21 mg, 0.15 mmol), FeCl₃ (1.6 mg, 0.010 mmol), DDQ (22.7 mg, 0.10 mmol), and CH₂Cl₂ (0.5 mL) were added to a mixture of **2a** (17 mg, 0.10 mmol) in CH₂Cl₂ (0.5 mL) at 0 °C. After being stirred for 24 h at 0 °C to room temperature, saturated aqueous NaHCO₃ was added to the mixture. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash

column chromatography on silica gel (hexane/EtOAc = 5/1) to afford **6** (13 mg, 0.043 mmol) in 43% yield.



Methyl 3,6-dioxo-2-(thieno[3,2-b]thiophen-2-yl)cycloxexa-1,4-diene-1-carboxylate (10)

Thienothiophene (63 mg, 0.45 mmol), DDQ (68 mg, 0.30 mmol), and CH_2Cl_2 (1.5 mL) were added to a solution of **2a** (48 mg, 0.30 mmol) and K_2CO_3 (81 mg, 0.60 mmol) in CH_2Cl_2 (1.5 mL) at 0 °C. After being stirred for 24 h at 0 °C to room temperature, saturated aqueous NaHCO₃ solution was added to the mixture. The resulting mixture was extracted with CH_2Cl_2 . The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 4/1) to afford **10** (40 mg, 0.13 mmol) in 43% yield.

Dark red solid; m.p. 135–140 °C; IR (NaCl) cm⁻¹:1735, 1667, 1650, 1560, 1486, 1448, 1435, 1419, 1379, 1308, 1280, 1230, 1099, 1046; ¹H NMR (500 MHz, CDCl₃): δ 7.70 (s, 1H), 7.54 (d, *J* = 5.5 Hz, 1H), 7.28 (d, *J* = 5.5 Hz, 1H), 6.84–6.91 (m, 2H), 3.90 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 185.7, 183.5, 165.2, 146.0, 139.8, 136.4, 136.0, 134.5, 133.0, 131.4, 124.9, 119.5, 53.2; MALDI-HRMS (*m*/*z*) calcd. for C₁₄H₈O₄S₂ [M]⁺ 303.9853, found 303.9858.



FeCl₃ (1,5 mg, 9.2 μ mol) was added to a mixture of **10** (28 mg, 0.092 mmol) in CH₂Cl₂ (1.0 mL) at room temperature. After being stirred for 5 h at room temperature, water was added to the mixture. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The yield was calculated by crude ¹H NMR using dimethyl sulfone (8.7 mg, 0.092 mmol) as an internal standard.



Methyl 2-hydroxybenzo[4,5]thieno[2,3-b]benzofuran-1-carboxylate (7)

Benzothiophene (20 mg, 0.15 mmol), FeCl₃ (1.6 mg, 0.010 mmol), DDQ (22.7 mg, 0.10 mmol), and CH₂Cl₂ (0.5 mL) were successively added to a solution of **2a** (17 mg, 0.10 mmol) in CH₂Cl₂ (0.5 mL) at 0 °C. After being stirred for 24 h at 0 °C to room temperature, saturated aqueous NaHCO₃ solution was added to the mixture. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 5/1) to afford 7 (20 mg, 0.067 mmol) in 67% yield.

Pale yellow solid; m.p. 136–140 °C; IR (NaCl) cm⁻¹: 2952, 1667, 1612, 1577, 1493, 1457, 1439, 1433, 1421, 1327, 1276, 1255, 1255, 1213, 1117, 1032; ¹H NMR (400 MHz, CDCl₃): δ 10.53 (s, 1H), 7.92 (d, J = 8.2 Hz, 1H), 7.82 (d, J = 8.2 Hz, 1H), 7.68 (d, J = 9.1 Hz, 1H), 7.49–7.46 (m, 1H), 7.38–7.35 (m, 1H), 6.98 (d, J = 9.1 Hz, 1H), 4.13 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 170.3, 162.1, 158.3, 154.2, 137.8, 131.1, 124.9, 124.1, 124.0, 123.8, 122.8, 120.0, 118.6, 113.1, 104.9, 52,4; MALDI-HRMS (m/z) calcd. for C₁₆H₁₀O₄S [M]⁺ 298.0294, found 298.0294.

4. Structure determination of compound 6.

4-1. Determination by MicroED.

MicroED data collection;

The microcrystals of 6, as obtained in S14 or recrystallized as follows, were used to prepare two MicroED grids. Crystals in the two grids were isomorphic and merged during data processing. For recrystallization, the compound 6 was dissolved in a small amount of hot MeOH. After 12 h at room temperature, resulting precipitates were filtrated and dried in the air.

A Quantifoil grid (Cu R 1.2/1.3) containing the crystals was loaded into a CRYO ARM 300 II (JEOL Ltd.) equipped with an XF416 camera (TVIPS). The CRYO ARM 300 II had an omega filter set at a slit width of 20 eV and was operated at an acceleration voltage of 300 kV while being cooled at liquid nitrogen temperature. The emission and illumination conditions were as follows: CL aperture at 100 μ m, spot 7, alpha 1, emission at 2.0 μ A and A2 at 4.8 kV for parallel beam illumination, leading to the beam diameter of *ca*. 2.0 μ m. The virtual camera distance was set to nominal 600 mm, calibrated to be 715 mm. Automatic data collection was carried out by SerialEM⁵ with a strategy similar to those reported in the references 6–8. Crystals identified on montage maps were centered by stage shifts and brought to the eucentric height at a view magnification of x2500. Continuous rotation diffraction patterns were then collected at a rotation rate of 1°/sec, 1 frame/sec, with an electron flux of 0.05 electron/Å²/sec, spanning 60° per crystal. To mitigate preferred orientations of crystals on a grid, the starting angles of rotation were varied from -50° to -20° among crystals. Images were recorded at 2x binning (2048 x 2048 pixels) with a pedestal of 100 and the image correction parameter LC_ROW 1⁹.

Diffraction patterns from more than 550 crystals were measured. Typical images of crystals and diffraction patterns are shown in Supplementary Figure S2 The crystals were thin plates and tended to be bent and curly. Diffraction spots were streaky.

MicroED data processing;

Diffraction patterns were processed with DIALS^{10,11} with parallelization provided by GNU parallel¹². 26 crystals that diffracted to high resolutions and showed similar unit cell parameters were scaled with dials.scale¹³. Merging statistics are shown in Supplementary Table S2. Structure solution by SHELXD¹⁴ and kinematical refinement by SHELXL¹⁵ were carried out in the Olex2 GUI¹⁶. Twin refinement with a twinning operator h,-k,-h-l was employed. Anisotropic atomic displacement factor (ADP) refinement was unstable, leading to many non-positive definite ADP

tensors. Thus we refined only isotropic ADP. Probably this is caused by the limited data quality (diffraction spot streaking and unit cell variability), crystal twinning and/or multiple scattering. The resulting MicroED structure should be interpreted qualitatively, not quantitatively.



Figure S1: MicroED structure and map of 6 showing the bc plane. The figure was rendered by PyMOL 2.5.0 (Schrödinger LLC).

Supplementary Figure S2:

(A) A square montage showing microcrystals of **6**. Note the crystal bending.

(B) A diffraction pattern of compound 6.

(C) Raw distributions of the unit cell parameters for crystals diffracted better than 1.2 Å. These are from an early step of processing before using prior cell information in the indexing routine and include bad crystals and mis-indexed crystals. Note the broad angle distributions, probably due to streaky diffraction spots.









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d_max	d_min	#obs	#uniq	mult.	%comp	<i></i>	<i si=""></i>	r_mrg	r_meas	r_pim	cc1/2
8.15	2.29	1752	179	9.79	94.71	68.60	33.40	0.12	0.12	0.04	0.992*
2.29	1.83	1644	164	10.02	94.25	33.40	20.80	0.22	0.24	0.07	0.977*
1.83	1.60	2848	208	13.69	97.20	28.30	18.70	0.26	0.27	0.07	0.968*
1.60	1.45	2225	186	11.96	95.38	19.10	11.10	0.33	0.34	0.10	0.945*
1.45	1.35	1874	167	11.22	95.43	12.50	7.30	0.42	0.44	0.13	0.880*
1.35	1.27	1711	158	10.83	95.76	11.50	6.10	0.45	0.47	0.14	0.842*
1.27	1.21	2622	183	14.33	94.82	11.40	6.20	0.47	0.49	0.13	0.923*
1.21	1.15	2954	195	15.15	96.53	10.20	5.40	0.48	0.50	0.13	0.917*
1.15	1.11	2829	197	14.36	95.63	12.70	6.20	0.47	0.49	0.13	0.893*
1.11	1.07	2552	198	12.89	96.59	9.40	4.50	0.52	0.54	0.14	0.832*
1.07	1.04	2163	175	12.36	96.69	6.90	3.30	0.57	0.60	0.16	0.859*
1.04	1.01	2162	171	12.64	96.07	6.10	2.70	0.59	0.61	0.17	0.817*
1.01	0.98	2016	171	11.79	94.48	5.80	2.30	0.60	0.63	0.17	0.743*
0.98	0.96	1871	167	11.20	95.43	4.60	1.80	0.66	0.69	0.20	0.801*
0.96	0.94	1876	162	11.58	95.86	3.90	1.50	0.71	0.74	0.21	0.450*
0.94	0.92	2621	190	13.79	96.94	3.60	1.40	0.71	0.74	0.20	0.554*
0.92	0.90	3263	199	16.40	96.60	3.30	1.40	0.73	0.76	0.18	0.577*
0.90	0.88	2874	189	15.21	94.50	2.80	1.10	0.74	0.77	0.19	0.674*
0.88	0.86	2766	200	13.83	96.62	2.40	0.90	0.73	0.76	0.19	0.375*
0.86	0.85	2514	176	14.28	96.17	2.70	1.00	0.76	0.79	0.19	0.310*
8.15	0.85	47137	3635	12.97	95.83	12.90	6.90	0.35	0.37	0.10	0.982*

Supplementary Table S2: Merging statistics of the MicroED dataset for compound 6.

Supr	olementary	Table S3:	Crystallog	graphic ref	inement sta	atistics of c	compound 6.

Compound	6
Chemical formula	$C_{14} H_8 O_4 S_2$
Formula wt	304.34
Cryst syst	Triclinic
Space group	<i>P</i> -1
Т, К	79
a, Å	3.711
b, Å	13.550
c, Å	22.244
α, deg	95.74
β, deg	92.86
γ, deg	91.59
Ζ	4
V, Å ³	1110.8
$D_{calc}, g cm^{-3}$	1.820
R1[I > 2(I)]	0.2074
wR2 (all)	0.4785
GOF	1.639
Data collection	Talos Arctica microscope
High resolution limit for refinement, Å	0.90
COD ID.	3000464
CCDC no.	2294744
XRDa ID	162

4-2. Determination by chemical modification.



The reaction with 2a and thieno[3,2-*b*]thiophene-2,5-*d*₂ provided mono-deuterated $10-d_1$ as the reaction intermediate of compound $6-d_1$ (see page S12). If di-duterated product S1 are obtained during this step, iso- $6-d_1$ would be produced. These results indirectly proved the structure of 6.

Thieno[3,2-b]thiophene-2,5-d2

n-BuLi (2.64 M in *n*-hexane; 758 mL, 2.00 mmol) was added to a mixture of thieno[3,2*b*]thiophene (140 mg, 1.00 mmol) in THF (5.0 mL) at -78 °C. After being stirred for 2 h at -78 °C to room temperature, D₂O was added to the mixture. The resulting mixture was extracted with hexane. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo to afford thieno[3,2-*b*]thiophene-2,5-*d*₂ (120 mg, 0.86 mmol) in 86% yield.

Gray solid; ¹H NMR (500 MHz, CDCl₃): δ 7.39 (d, J = 5.0 Hz, 0.08 H), 7.27 (s, 2H); ²H NMR (77 MHz, CDCl₃): δ 7.44 (brs).



$10-d_1$

Thieno[3,2-*b*]thiophene-2,5-*d*₂ (42 mg, 0.40 mmol), FeCl₃ (3.2 mg, 0.020 mmol), DDQ (45.4 mg, 0.20 mmol), and CH₂Cl₂ (1.0 mL) were added to a mixture of **2a** (33 mg, 0.20 mmol) in CH₂Cl₂ (1.0 mL) at 0 °C. After being stirred for 24 h at 0 °C to room temperature, saturated aqueous NaHCO₃ was added to the mixture. The resulting mixture was extracted with CH₂Cl₂. The combined organic layers were dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (hexane/EtOAc = 4/1) to afford **S1** (23 mg, 0.075 mmol) in 38% yield.

Dark red solid; ¹H NMR (500 MHz, CDCl3): δ 7.71 (s, 1H), 7.29 (s. 1H), 6.92–6.85 (m, 2H), 3.91 (s, 3H); ²H NMR (77 MHz, CDCl3): δ 7.44 (brs).

5. Fluorescence test of compounds 5a, 6, and 7.

Fluorescence and absorption measurement. All solvents for spectrophotometry were purchased from Nacalai Tesque. The solution- and solid-state fluorescence spectra were collected on a Hitachi F-7100 fluorescence spectrophotometer. To obtain an accurate spectrum, spectrum correction was carried out with a concentrated solution of rhodamine B. The solid-state fluorescence spectra were measured by surface photometric method. A cut filter was used to eliminate multiple lights due to the effects of light scattering. The absolute fluorescence quantum yields in the solid state were determined by Hitachi F-7100 fluorescence spectrophotometer by using a calibrated integrating sphere system. UV/Vis spectra were recorded on a HITACHI UH-5700 spectrophotometer at room temperature using a 1 mm quartz cuvette.

Computational details. All calculations were carried with the Gaussian 16 program¹⁷. Structural optimizations were carried out at the B3LYP level in the gas phase using the 6-31G(d) basis set. The vibrational frequencies were computed at the same level to check whether each optimized structure is an energy minimum (no imaginary frequency). Excitation wavelengths and oscillator strengths were obtained at the density functional level using time-dependent perturbation theory (TDDFT) approach.



Figure. S3. Normalized fluorescence spectra of 5a, 6, and 7 in toluene.



Figure S4. Absorption spectra of (A) 6, (B) 5a, and (C) 7 in CHCl₃.

All generated absorption bands in the 300 to 400 nm UV region. Compound 6 had a wide and large absorption band, whereas 5a and 7 had a weak absorption band. Regarding the prediction by time-dependent DFT calculations, the large absorption band of 6 corresponded to the HOMO \rightarrow LUMO transition and was attributable to the π - π * transition (See Figure S5).



Figure S5. (A) Calculated excitation wavelength (λ_{abs}), oscillator strength (*f*) and major contribution for **6**. (B) Calculated absorption spectra. (C) Frontier molecular orbitals of **6**.

Cartesian Coordination

6

Sum of electronic and thermal Free Energies = -1635.40999 A.U _____ -0.14341031 С -1.47393609 -2.20522482 С -0.09817779 -2.54911158 -0.15844407 С 0.92249001 -1.62355729 -0.05533917 С 0.55682240 -0.28792318 0.06523415 С -0.79438611 0.10845538 0.07622161 С -0.83751594 -1.84217686 -0.02874223 1.96227449 Η -1.93261086 -0.06961212 Η 1.32750599 0.47474178 0.15159541 С -3.25551908 -0.38577800 -0.02766041 -3.64948936 0.75811951 Ο -0.10407248 0 -4.11749446 -1.43811313 0.07479680 -5.50832282 -1.08333239 0.07482245 С 0.15740873 -6.04896967 -2.02652581 Η 0.92279427 Η -5.73613152 -0.43245679 Η -5.77364253 -0.56557865 -0.85060119 Ο 0.10474850 -3.90907152 -0.29208949 -0.27863643 С -2.13860569 -3.47340679 С -1.14656410-4.43600625 -0.36429221 S -3.73621123 -4.18959121 -0.37764453 С -3.01032474 -5.78561809 -0.53951827 -5.75855810 С -1.61481536 -0.51248642 S -0.94174103 -7.35473706 -0.66959794 -0.68704780 С -3.54423870 -7.09844950 С -8.03626561 -0.76960986 -2.55212790 Η -4.60052907 -7.33784496 -0.72974320 -2.66335417 -9.10616743 -0.88516124 Η 0 -1.10518981 1.42518959 0.20213082 1.92734615 0.28094589 Η -0.27865065 _____

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7. ¹H and ¹³C NMR spectra of newly synthesized compounds.



¹H NMR of methyl 3,6-dihydroxy-2-(1*H*-indol-3'-yl)benzoate (**3a**)



¹H NMR of methyl 3,6-dihydroxy-2-(1'-methyl-1*H*-indol-3'-yl)benzoate (**3b**)

¹H NMR of methyl 3,6-dihydroxy-2-(1'-tosyl-1*H*-indol-3'-yl)benzoate (**3c**)



¹³C NMR of methyl 3,6-dihydroxy-2-(1'-tosyl-1*H*-indol-3'-yl)benzoate (**3c**)



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¹H NMR of methyl 3,6-dihydroxy-2-(1'-benzyl-1*H*-indol-3'-yl)benzoate (**3d**)



¹³C NMR of methyl 3,6-dihydroxy-2-(1'-benzyl-1*H*-indol-3'-yl)benzoate (**3d**)





¹H NMR of methyl 3,6-dihydroxy-2-(5'-methoxy-1*H*-indol-3'-yl)benzoate (**3e**)



¹H NMR of methyl 3,6-dihydroxy-2-(5'-fluoro-1*H*-indol-3'-yl)benzoate (**3f**)



¹H NMR of methyl 3,6-dihydroxy-2-(5'-bromo-1*H*-indol-3'-yl)benzoate (**3g**)



¹H NMR of 1-(3,6-dihydroxy-2-(1*H*-indol-3'-yl)phenyl)ethan-1-one (**3h**)

¹³C NMR of 1-(3,6-dihydroxy-2-(1*H*-indol-3'-yl)phenyl)ethan-1-one (**3h**)



¹H NMR of 3,6-dihydroxy-2-(1*H*-indol-3'-yl)benzaldehyde (**3i**)



¹³C NMR of 3,6-dihydroxy-2-(1*H*-indol-3'-yl)benzaldehyde (3i)



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¹H NMR of methyl 3-(3',6'-dihydroxy-2'-(methoxycarbonyl)phenyl)-1*H*-indole-2-carboxylate (**3k**)

¹³C NMR of methyl 3-(3',6'-dihydroxy-2'-(methoxycarbonyl)phenyl)-1*H*-indole-2-carboxylate (**3**k)







¹³C NMR of methyl 2-hydroxy-6-oxo-6,7-dihydrochromeno[3,4-*b*]indole-1-carboxylate (8)



¹H NMR of methyl 3,6-dihydroxy-4'-methoxy-[1,1'-biphenyl]-2-carboxylate (4a)



¹³C NMR of methyl 3,6-dihydroxy-4'-methoxy-[1,1'-biphenyl]-2-carboxylate (4a)



¹H NMR of methyl 3,6-dihydroxy-2-(2'-hydroxynaphthalen-1'-yl)benzoate (4b)





¹H NMR of methyl 3,6-dihydroxy-2',4',6'-trimethoxy-[1,1'-biphenyl]-2-carboxylate (4c)

¹³C NMR of methyl 3,6-dihydroxy-2',4',6'-trimethoxy-[1,1'-biphenyl]-2-carboxylate (4c)





¹H NMR of methyl 2'-bromo-3,6-dihydroxy-4',6'-dimethoxy-[1,1'-biphenyl]-2-carboxylate (4d)

¹³C NMR of methyl 2'-bromo-3,6-dihydroxy-4',6'-dimethoxy-[1,1'-biphenyl]-2-carboxylate (4d)







¹³C NMR of methyl 3,6-dihydroxy-2',5'-dimethoxy-[1,1'-biphenyl]-2-carboxylate (4e)





¹H NMR of methyl 2-hydroxy-6*H*-benzofuro[2,3-*b*]indole-1-carboxylate (5a)



¹H NMR of methyl 2-(1*H*-indol-3'-yl)-3,6-dioxocyclohexa-1,4-diene-1-carboxylate (9)

¹H NMR of methyl 6-benzyl-2-hydroxy-6*H*-benzofuro[2,3-*b*]indole-1-carboxylate (**5b**)



¹³C NMR of methyl 6-benzyl-2-hydroxy-6*H*-benzofuro[2,3-*b*]indole-1-carboxylate (**5b**)







¹³C NMR of methyl 2-hydroxy-9-methoxy-6*H*-benzofuro[2,3-*b*]indole-1-carboxylate (5c)







¹³C NMR of methyl 9-bromo-2-hydroxy-6*H*-benzofuro[2,3-*b*]indole-1-carboxylate (5d)





¹H NMR of methyl 5-hydroxy-thieno[2',3':4,5]thieno[3,2-*b*]benzofuran-4-carboxylate (6)

¹³C NMR of methyl 5-hydroxy-thieno[2',3':4,5]thieno[3,2-*b*]benzofuran-4-carboxylate (6)





¹H NMR of methyl 3,6-dioxo-2-(thieno[3,2-*b*]thiophen-2-yl)cycloxexa-1,4-diene-1-carboxylate (10)

¹³C NMR of methyl 3,6-dioxo-2-(thieno[3,2-*b*]thiophen-2-yl)cycloxexa-1,4-diene-1-carboxylate (10)





¹H NMR of methyl 2-hydroxybenzo[4,5]thieno[2,3-*b*]benzofuran-1-carboxylate (7)

¹³C NMR of methyl 2-hydroxybenzo[4,5]thieno[2,3-*b*]benzofuran-1-carboxylate (7)





NOESY of methyl 2-hydroxybenzo[4,5]thieno[2,3-b]benzofuran-1-carboxylate (7)

¹H NMR of thieno[3,2-*b*]thiophene (authentic sample)

¹H NMR (500 MHz, CDCl₃): δ 7.40 (d, *J* = 5.0 Hz, 2H-A), 7.28 (d, *J* = 5.0 Hz, 2H-B).







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²H NMR of thieno[3,2-*b*]thiophene-2,5-*d*₂



¹H NMR of **10-** d_1



²H NMR of $10-d_1$

