

Exploring the Chemical Subspace of RPLC: a Data Driven Approach

Denice van Herwerden,^{*,†} Alexandros Nikolopoulos,[†] Leon P. Barron,^{‡,†} Jake W. O'Brien,^{¶,†} Bob W. J. Pirok,[†] Kevin V. Thomas,[¶] and Saer Samanipour^{*,†,§,¶}

[†]*Van 't Hoff Institute for Molecular Sciences (HIMS), University of Amsterdam, Amsterdam*

[‡]*MRC Centre for Environment and Health, Environmental Research Group, School of Public Health, Faculty of Medicine, Imperial College London, W12 0BZ, United Kingdom*

[¶]*Queensland Alliance for Environmental Health Sciences (QAEHS), The University of Queensland, Australia*

[§]*UvA Data Science Center, University of Amsterdam, Amsterdam*

E-mail: d.vanherwerden@uva.nl; s.samanipour@uva.nl

Abstract

The chemical space is comprised of a vast number of possible structures, of which an unknown portion comprises the human and environmental exposome. Such samples are frequently analyzed using non-targeted analysis via liquid chromatography (LC) coupled to high-resolution mass spectrometry often employing a reversed phase (RP) column. However, prior to analysis, the contents of these samples are unknown and could be comprised of thousands of known and unknown chemical constituents. Moreover, it is unknown which part of the chemical space is sufficiently retained and eluted using RPLC. Therefore, we present a generic framework that uses a data driven approach to predict whether molecules fall ‘inside’, ‘maybe’ inside, or ‘outside’ of the

11 RPLC subspace. Firstly, three retention index random forest (RF) regression models
12 were constructed that showed that molecular fingerprints are able to predict RPLC
13 retention behavior. Secondly, these models were used to setup the dataset for building
14 a RPLC RF classification model. The RPLC classification model was able to cor-
15 rectly predict whether a chemical belonged to the RPLC subspace with an accuracy
16 of 92% for the testing set. Finally, applying this model to the 91737 small molecules
17 (i.e., ≤ 1000 Da) in NORMAN SusDat showed that 19.1% fall ‘outside’ of the RPLC
18 subspace. Knowing which chemicals are outside of the RPLC subspace can assist in
19 reducing potential candidates for library searching and avoid screening for chemicals
20 that will not be present in RPLC data.

21 Introduction

22 The chemical space refers to a collection of all possible organic structures - for example,
23 the GBD-17 database includes 116 billion possible organic molecules with a maximum of 17
24 atoms, which is only a fraction of the chemical space.¹⁻⁸ Increasing the number of atoms
25 only drastically increases these numbers and shows how vast the chemical space actually is.
26 Even though these are possible structures, not all of them are likely to be present in the
27 human and environmental exposome.⁸ When evaluating the exposome, the main difficulty is
28 that the contents of the samples taken are unknown prior to analysis and may comprise of
29 thousands of both known and unknown constituents, particularly for small molecules (i.e.,
30 molecular weight ≤ 1000 Da).⁹⁻¹⁶ A frequently used approach for analyzing such samples
31 is non-targeted analysis (NTA) via liquid chromatography (LC) coupled to high-resolution
32 mass spectrometry (HRMS), for which a reversed phase (RP) LC selectivity is often used.⁸
33 However, it is not yet known what part of the chemical space is covered by RPLC. The
34 knowledge of the covered subspace also contains crucial information on chemicals that might
35 not be visible in the final data even though they were present in the sample.³

37 Knowing what is separable with RPLC can have an improved outcome for both NTA
38 and suspect screening. For NTA, the aim is to identify as much as possible of the potentially
39 thousands of chemicals present in samples coming from, for example, biological or environ-
40 mental backgrounds. Eliminating the potential candidates that fall outside of the chemical
41 subspace of the selectivity (e.g., RPLC), reduces the number of false positive identifications.
42 On the other hand, suspect screening is also a frequently used approach, where samples are
43 screened for lists or even databases of compounds. Defining the subspace of a selectivity can
44 reduce the number of potential candidates in these compound lists, reducing the computa-
45 tional time required and the false positive matches with chemicals that cannot possibly be
46 measured with this technique.

47

48 Separation data is usually limited to the mere assessment of whether the analyte retention
49 time could fit in the range of the candidate's chemical class.¹⁷⁻²⁰ To take better advantage
50 of the LC data, retention times are required to be initially converted to retention indices
51 (r_i), since the former are significantly influenced by the chromatography conditions, such as
52 temperature, mobile phase composition, and gradients.^{20,21} On the other hand, r_i values pro-
53 vide a robust and highly reproducible way to express retention in liquid chromatography.²⁰
54 High reproducibility makes inter-laboratory results comparable, enabling both m/z and r_i
55 comparison with a reference and resulting in more confident suspect shortlisting.

56

57 As for any r_i system, different chromatography conditions should have negligible influence
58 on the r_i value of the analytes, suggesting that there is a correlation between the r_i values
59 and structural properties, expressed as molecular descriptors. This is the main principle
60 used by the quantitative structure-retention relationship (QSRR) based models,²² enabling
61 the construction of QSRR models that either use all or a selection of descriptors to predict r_i
62 values.²³⁻²⁶ However, difficulties arise when calculating descriptors due to convergence issues
63 related to calculation time-out or local minima.²⁵⁻²⁷ Moreover, descriptors can often be diffi-

64 cult to interpret, since they contain mathematical representations of the molecular structure.
65 Alternatively, molecular fingerprints directly encode the molecular structure, making them
66 more descriptive/understandable to interpret in relation to the chemical and do not require
67 structural optimization (i.e., only uses 2D structural information), making them a potential
68 alternative to descriptors.

69

70 In this paper, we present a data driven approach for a generic framework that enables
71 quick screening of the RPLC chemical space, assuming that the molecules are in solution and
72 can be injected into a system. A set of regression and classification models were built to assess
73 whether a structure can theoretically be analyzed via RPLC. To build the RPLC classifica-
74 tion model, firstly, we show the potential of using fingerprints for the prediction of r_i values
75 for three retention index series, confirming that molecular fingerprints contain information on
76 RPLC retention behavior. Three commonly used scales, namely: the n-alkylamide system,
77 containing the n-alkylamide homologous series from n-propanamide to n-tetradecanamide
78 (C3-C14)²⁸, the r_i system developed by Aalizadeh et al. from the University of Athens re-
79 ferred to as UoA, comprising of 18 reference compounds that were computationally selected in
80 order to achieve a broad and reliable r_i reference system²⁹, and the cocamide diethanolamine
81 homologous series that is comprised of C(n = 0-23)-DEA chemicals³⁰ were employed for our
82 model building. Secondly, we show the performance of the RPLC classification model and
83 apply the model on a set of 91737 small molecules (i.e., molecular weight \leq 1000 Da) from
84 the NORMAN substance database (SusDat).

85 **Experimental Section**

86 **Overall Workflow**

87 The overall workflow for this work can be found in figure 1 and the details are explained
88 in the following sections. In brief, a total of four random forest (RF) models were built, of

89 which three were r_i RF regression models (Figure 1A) and the fourth a RPLC RF classifi-
90 cation model (Figure 1B). For building these models, a type of molecular fingerprint needed
91 to be selected and the dataset obtained before model optimization and performance testing
92 (Figure 1C). These models were used for evaluating the potential of using molecular finger-
93 prints for prediction of retention behavior in RPLC and for setting up two of the classes
94 for the fourth RF classification model. The latter refers to the ‘inside’ and ‘maybe’ inside
95 class. Here, the ‘maybe’ class represents the chemicals that are poorly retained (i.e., close
96 to t_0) or require relatively high amounts of organic modifier to elute. All chemicals in be-
97 tween the ‘maybe’ regions are classified as ‘inside’. For the RPLC classification model, a
98 dataset with chemicals that were ‘inside’, ‘maybe’ inside, and ‘outside’ of the RPLC sub-
99 space was constructed (Figure 1B). Finally, the application of the RPLC classification model
100 was showcased by applying it on the NORMAN SusDat database, which is a collection of
101 expert curated environmentally relevant chemicals that have been actively used for screening
102 of complex samples. All datasets for constructing the models and the NORMAN SusDat
103 database can be found on Figshare.³¹

104

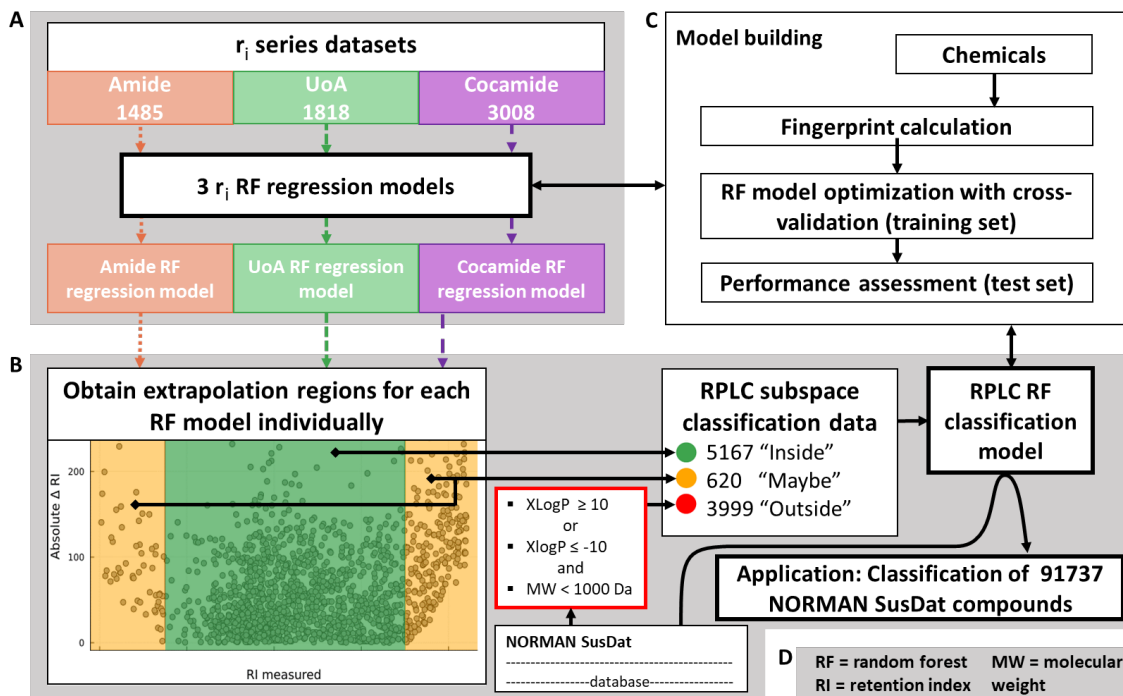


Figure 1: Workflow for construction of the RPLC classification model, comprising of the construction of three r_i RF regression models (A) and the construction the RPLC dataset for the RPLC RF classification model, which was applied to NORMAN SusDat (B). Finally C shows the model setup and D contains an overview of the abbreviations.

105 Fingerprint Calculations

106 The RF models were built using a combination of two different fingerprint series as inputs,
 107 which included the AtomPairs2DFingerprintCount (2DAPC) and PubChem fingerprints,³²
 108 obtained through PaDEL.³³ The 2DAPC fingerprints counted the number of times two atoms
 109 were present with a certain distance between themselves. For example, the molecule with
 110 the SMILES 'NC(CC)CN' contains two times a distance of 3 between a C and N atom (i.e.,
 111 C-x-x-N in the 2D molecular structure). The distances included ranges from 1 to 10 and
 112 the elements considered were C, N, O, Cl, I, Br, F, P, S, Si, B, and X, where X represents
 113 all halogens, yielding a total of 780 2DAPC fingerprints. As for the PubChem fingerprints,
 114 only the portion of fingerprints containing ring information was used (i.e., PubChem finger-
 115 print 115 - 262). These fingerprints were converted and reduced to a total of 10 additional

116 variables, which were the number of rings with a size of 3, 4, 5, 6, 7, 8, 9, 10, the number of
117 aromatic rings, and the number of hetero-aromatic rings. Since the PubChem fingerprints
118 are binary, there were multiple columns describing the same information but only differing
119 in the number of a ring of a certain size. For example, for a ring size of 3, there were 2
120 fingerprints, namely PubChem fingerprint 115 and 122, which were described as more than
121 1 ring with a size of 3 or more than 2 rings with a size of 3, respectively. In case a molecule
122 contained 2 rings with a size of 3, the PubChem fingerprints 115 would be 0 and 122 would
123 be 1, which was converted to a single variable for our model containing the number of rings
124 with a size of 3, meaning that this variable would be equal to 2 for this example case. An
125 overview of which PubChem fingerprints were used for each of the 10 reduced PubChem
126 variables can be found in table S2.

127

128 **Retention Index Random Forest Regression Models**

129 To show that fingerprints can be used to describe retention behavior in RPLC and for set-
130 ting up the dataset for the RPLC classification model, random forest (RF) regression models
131 were built using three different retention index series (Figure 1A). The three series used for
132 this, were the amide²⁸, University of Athens (UoA)²⁹, and cocamide series.³⁰ For each of
133 the series, the measured r_i were obtained from their respective articles, yielding 1485, 1818,
134 and 3008 unique chemicals with measured r_i values for the amide, UoA, and cocamide series,
135 respectively. For all chemicals, the 2DAPC and PubChem fingerprints were calculated ac-
136 cording to Section ‘Fingerprint Calculations’. For each r_i series, data was split into a training
137 and test set, at random, with a ratio of 0.85:0.15, ensuring similar coverage of the r_i range
138 in both sets. The test set was only used for testing and thus never used for training. For
139 optimization of the RF regression models, the training set was used with a 0.8:0.2 split for
140 training and cross-validation, respectively. This ratio of split has been shown to be effective
141 in such data sets.^{25,26,34,35} The RF regression models used a third of the features (i.e., 264)

142 for training each tree. The parameters that were optimized were the minimum number of
143 samples per leaf and the number of trees. The minimum number of samples per leaf tested
144 were 4, 6, 8, 10, 15, and 20. The tested number of trees were 50, 100, 150, 200, 250, 300,
145 350, 400, 500, 600, 700, 800, 900, and 1000. In addition, the random state for splitting the
146 cross-validation set and selection of the features in the RF models for each tree was also
147 varied with values of 1, 2, and 3. The accuracy of the cross-validation set for each possible
148 combination of the minimum number of samples per leaf, number of trees, and random state
149 was used for the optimization of the RF models. After obtaining the optimized models for
150 the amide, UoA, and cocamide series, the applicability domains were assessed according to
151 Section ‘Applicability Domain Calculations’. Finally, for each r_i series, the optimized model
152 and applicability domain assessment were applied on the test set to evaluate the performance
153 of the model on unseen data.

154

155 **RPLC Random Forest Classifier**

156 The dataset for building the RPLC classifier model was comprised of three classes: ‘inside’,
157 ‘maybe’, and ‘outside’ the RPLC subspace (Figure 1B). The ‘outside’ chemicals were ob-
158 tained from the NORMAN SusDat database based on their extreme XLogP values. Here,
159 the XLogP was chosen rather than the logD due to the fact that it is easier to predict, more
160 stable, and more accurate.³⁶ For the ‘outside’ case, a total of 3999 compounds with a XLogP
161 value above 10 or below -10 and with a molecular weight below 1000 Da were obtained. As
162 for the ‘inside’ and ‘maybe’ chemicals, these were obtained from the experimentally defined
163 r_i values by the three r_i series. For each of the series, the absolute difference between the
164 predicted and measured r_i (i.e., the residuals) versus the measured r_i values were plotted
165 and the regions of extrapolation were identified. These regions were obtained based on the
166 increasing residuals that were caused by the inherent over estimation and under estimation
167 of a RF regression model, which are associated with either extremely low or extremely high

168 r_i values, respectively. These regions correspond to chemicals that elute close to t_0 or are
169 very difficult to elute from the column (i.e., require a relatively high percentage of organic
170 modifier). The chemicals with a measured r_i in these extrapolation regions were labeled
171 as ‘maybe’ and the remaining chemicals were labeled as ‘inside’ the RPLC subspace. This
172 yielded a total of 620 ‘maybe’ and 5167 ‘inside’ compounds. Whenever a chemical was found
173 in multiple classes (i.e., it was present in multiple datasets of the r_i models), it was removed
174 from the lower ranking RPLC classes and kept in the highest ranking RPLC class (i.e., ‘in-
175 side’ > ‘maybe’ > ‘outside’ RPLC class rank). For example, if a chemical was found in the
176 ‘maybe’ region for UoA and in the ‘inside’ for Cocamide, it would be classified as ‘inside’.
177 More details on the division between the ‘inside’ and ‘maybe’ classification can be found
178 in Section ‘RPLC Classification Model’ as these are based on the results of the three RF
179 regression models.

180

181 The dataset described above was used for building the RPLC classifier model with a
182 training set/test set split of 0.85:0.15, ensuring equal distribution of each class in both sets.
183 The optimized RF classifier model was obtained using the same approach as for the RF re-
184 gression models (see Section ‘Retention Index Random Forest Regression Models’). For this
185 model, the applicability domain was also obtained as described below. Finally, the optimized
186 RPLC classification model and applicability domain assessment was applied to the test set
187 and the performance was evaluated.

188

189 **RPLC Space Prediction for NORMAN SusDat**

190 To showcase the model’s potential, it was applied to the NORMAN SusDat database.⁵ For
191 this, the 2DAPC and reduced PubChem fingerprints for a total of 91737 chemicals with a
192 molecular weight below 1000 Da from SusDat were calculated. These fingerprints were then
193 used to calculate the leverage of each chemical with the RPLC classifier training set, as

194 explained in the next section ‘Applicability Domain Calculations’, and to apply the RPLC
195 classifier model to each of the SusDat chemicals. To visualize the coverage of each class
196 (i.e., ‘inside’, ‘maybe’, and ‘outside’ the RPLC subspace), the molecular weight was plotted
197 against the XLogP, which were obtained from the descriptor calculations of PaDEL.

198

199 **Applicability Domain Calculations**

200 Applicability domain calculations were used to assess whether the training data, used in the
201 random forest models, sufficiently covered the variable space for new chemicals on which the
202 models need to be applied.^{25,37} This was done through leverage calculations of a chemical
203 with the entire training set, yielding a distance of that chemical to the training set. Equation
204 1 shows how the leverage is calculated, where X is the training data matrix and x_i is the
205 sample vector, both containing the 2DAPC and reduced PubChem fingerprints for our mod-
206 els. To set a threshold for this, the leverage was calculated for all training samples with the
207 entire training set of a model, yielding values between 0 and 1. Then, a leverage threshold
208 was obtained that covered 95% of the training data. If a chemical, compared to the training
209 set of the model in question, had a value lower than the leverage threshold, the compound
210 was within the applicability domain, and, if the value was above the leverage threshold, the
211 results should be taken with care as the training data might not be sufficiently describing
212 the variable space for the new compound.

213

$$l_{ii} = x_i(X^T X)^{-1}x_i \quad (1)$$

214 **Calculations and Code Availability**

215 The calculations and development of the models were executed on a personal computer with
216 12 CPUs and 32 GB of RAM, using Windows 10. The r_i regression and RPLC classifi-

217 cation models were developed and evaluated with the Julia programming language (v1.6).
218 The code for using the r_i regression models and RPLC space prediction model is available
219 at: https://bitbucket.org/Denice_van_Herwerden/riprediction/src/main/. This Ju-
220 lia package contains functions for obtaining the required 2DAPC and reduced PubChem
221 fingerprints and for using the r_i regression models and RPLC sub space classification model.
222

223 Results and discussion

224 Retention Index Random Forest Regression Models

225 All three r_i regression models obtained an accuracy of 81% for the training set and, for the
226 test set. The amide, UoA, and cocamide models had an accuracy of 68%, 70%, and 67%,
227 respectively. The r_i regression models were built and optimized for the amide, UoA and
228 cocamide series. Grid optimization of each of these models showed that the number of trees
229 did not influence the performance of the model (Figures S1, S2, and S3). Therefore, to
230 keep the model light, 200 trees were selected. As for the minimum number of samples per
231 leaf, 8 was found to be the optimum, based on the training and cross-validation accuracy.
232 When evaluating the predicted versus the measured r_i values for these models a trend of over
233 prediction for lower r_i values and under prediction of higher r_i values was found (Figures S4,
234 S6, and S8), corresponding to the regions where the RF regression models were extrapolat-
235 ing. These regions were used for establishing the ‘maybe’ areas for the RPLC classification
236 dataset.

237

238 Most compounds (i.e., 88.5%) in our test set appeared to be within the applicability
239 domain of each model. To obtain the applicability domains of these models, a 95% leverage
240 threshold of 0.189 for amide, 0.652 for UoA, and 0.424 for cocamide was found for the train-
241 ing sets. For the training set the leverage values range between 0 and 1, meaning that the

242 lower threshold for the amide model showed how similar most of the amide compounds were
243 to each other, while for the UoA and cocamide models, the higher thresholds corresponded
244 with the larger variety of chemical structures found in the dataset. When the leverage cal-
245 culations were applied on the test sets for these models, a total of 22, 34, and 54 compounds
246 were found to be outside of the applicability domain for the amide, UoA, and cocamide r_i
247 models, respectively. This does not necessarily mean that the predicted outcome for these
248 cases was wrong, as can be seen in figures S4, S6, and S8. Here, most chemicals outside the
249 applicability domain still follow the trend of the other data points. However, the outcome
250 should be taken with care as the model might insufficiently cover the chemical space for a
251 new compound in question, especially for leverage values > 1 . It should be noted that the
252 largest training set leverage value obtained from our applicability domain calculations was 1.

253

254 The cocamide RF regression model used the most fingerprints for the prediction of the
255 r_i indices (i.e., 215 fingerprints), while the UoA and amide r_i models used 165 and 61, re-
256 spectively. The low number of fingerprints used for amide was not surprising due to the
257 fact that the compounds in this r_i series are only comprised of C, H, N, and O. Hence, the
258 amide r_i model only used the 2DAPC fingerprint counts with a certain distance between C,
259 N, and O atoms. At first sight, this was also noticeable when comparing the top 20 most
260 important fingerprints for the three r_i models (S3). The most contributing fingerprints for
261 the amide r_i model were the distances 1 till 7 between two C atoms with importance ranging
262 between 27% and 4%. As for the UoA r_i model, C-Cl and C-X distance begin to contribute
263 more to the model and the most important fingerprint (i.e., distance 7 between C-C) only
264 contributes 9.6%, having an overall more divided importance between a larger group of con-
265 tributing features than the amide model. Finally, a similar trend was also observed for the
266 cocamide model, except that the C-X distances start to play a more important role than the
267 C-Cl distances, which could be explained by the higher number of halogens present in the
268 compounds from the cocamide dataset. This variability in important features used in each

269 r_i regression model shows that different structures may be better captured by one r_i model
270 vs another, due to the diversity of training set in terms of chemical structures. This, also,
271 further indicates the need for a more generic model incorporating the information from all
272 three r_i models.

273

274 Overall, these models show that a combination of the 2DAPC fingerprints and the re-
275 duced PubChem fingerprints can be used to predict r_i values. All three models performed
276 almost equally well with negligible deviations for the training set accuracy. However, de-
277 pending on the chemicals for which r_i would be predicted, it is advised to evaluate which
278 model would be most suitable based on the leverage applicability domain calculations.

279

280 **RPLC Classification Model**

281 To build the RPLC classification model, it was assumed that the chemicals are in solution
282 and that the chemicals can be injected into a system. Additionally, the model focuses on
283 whether an analyte could be analyzed with RPLC regardless of experimental parameters or
284 sample pretreatment. The dataset for this was comprised of 5167 ‘inside’, 620 ‘maybe’ in-
285 side, and 3999 ‘outside’ chemicals for the RPLC subspaces. The ‘outside’ cases were obtained
286 from NORMAN SusDat with extreme XLogP values, while the ‘inside’ and ‘maybe’ cases
287 came from the three r_i regression models. In figures S10, S11, and S12 the extrapolation
288 limits for each of the models are defined. For r_i range for the ‘inside’ RPLC subspace for
289 the amide, UoA, and cocamide series were 350-900, 100-900, and 250-1300, respectively. All
290 compounds that had a higher or lower r_i value for the corresponding range of the model it
291 was coming from, were classified as ‘maybe’ inside the RPLC subspace, due to the fact that
292 these chemicals either elute close to t_0 or require high percentages of organic eluent to be
293 eluted.

294

295 The final optimized classification model resulted in an accuracy of 94% and 92% for the
296 training and test set, respectively (Figures 2, and S15). In this case 200 trees and 8 minimum
297 samples per leaf was found to be the optimum for the model (Figure S13). For the training
298 and test set, 90.8% and 87.7% of the ‘inside’ and ‘maybe’ cases were correctly classified, 7.4%
299 and 9.3% of the ‘inside’ and ‘maybe’ cases were wrongly classified as a ‘maybe’ or ‘inside’
300 case, respectively, and 1.7% and 3.0% of the ‘inside’ and ‘maybe’ cases were wrongly classi-
301 fied as ‘outside’. For the ‘outside’ cases, 0.7% and 1.5% of the cases were wrongly classified
302 as an ‘inside’ or ‘maybe’ case and 99.3% and 98.5% of the cases was correctly classified as
303 an ‘outside’ case for the training and test set, respectively. Overall, considering that the
304 wrongly classified ‘inside’ and ‘maybe’ cases as ‘maybe’ and ‘inside’, respectively, still are
305 considered part of the RPLC subspace, the performance of the model was very good with
306 only 2.4% of all cases being wrongly classified as ‘inside’ or ‘maybe’ while being an ‘outside’
307 or vice versa for the test set.

308

309 As for the applicability domain of the RPLC classification model, the 95% leverage
310 threshold of the training set was 0.209 (Figure S14). In total, 102 compounds from the test
311 set (i.e., 6.9%) had a leverage with the training set that was higher than 0.209, of which 31
312 cases had leverage values above 1. Out of these 102 cases only 10 were wrongly classified
313 and had leverage values ranging between 0.209 to the most extreme (i.e., 809.255), showing
314 that in this case higher leverage values did not necessarily mean that the model would have
315 a higher error. However, it should be noted that cases with a very large leverage should be
316 considered with extra care, as they may have a higher level of uncertainty.

317

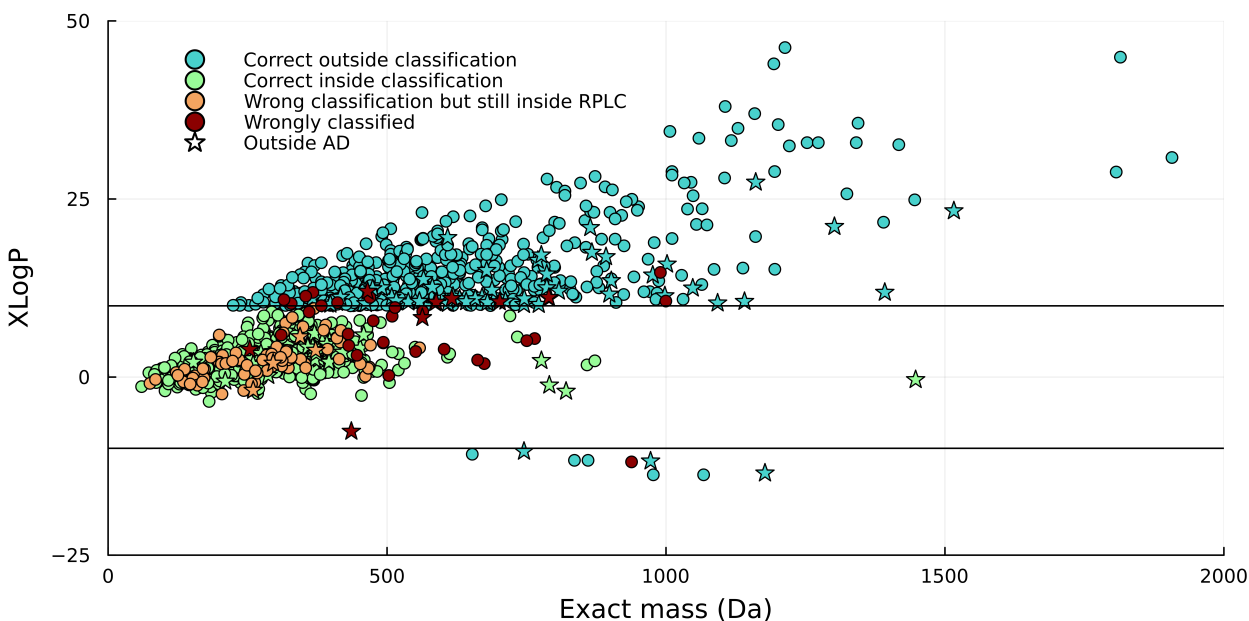


Figure 2: XLogP values versus the molecular weight for the RPLC classification test set. In blue are the correctly classified ‘outside’ cases, in green are the correctly classified ‘inside’ and ‘maybe’ cases, in orange are the wrongly classified ‘inside’ cases as ‘maybe’ and vice versa, in red the wrongly classified ‘inside’ and ‘maybe’ cases as ‘outside’ and the wrongly classified ‘outside’ cases as ‘inside’. The star markers show the compounds that were outside the 95% applicability domain of the RPLC classification training set

318 A total of 280 features were contributing to the RPLC classification model. This is more
 319 than for each of the three r_i regression models, which was expected due to the higher variety
 320 in chemical structures used in the RPLC classification model. The 20 most contributing fea-
 321 tures are mainly described by ring related features and distances between combinations C,
 322 N, and O atoms. A previous version of the model that was tested, using only the 2DAPC fin-
 323 gerprints, frequently wrongly classified ‘inside’ as ‘outside’ due to the high degree of cyclicity
 324 in the chemical structures (e.g., InChIKey: IUKLSMSEHKDIIP-BZMYINFQSA-N). Hence,
 325 the addition of the reduced PubChem fingerprints better captures these chemical properties.
 326 As a result, the number of rings with a size of 6, the minimum number of aromatic rings, and
 327 the number of rings with a size of 5 were also part of the top 20 most contributing features.

328

329 In total, considering the extreme misclassifications, 9 out of 599 ‘outside’ chemicals were

330 wrongly classified as ‘inside’ or ‘maybe’ inside the RPLC subspace and 14 out of the 767
331 ‘inside’ and 12 out of the 102 ‘maybe’ cases were classified as ‘outside’ the RPLC subspace.
332 Two of the nine wrongly classified ‘outside’ cases were organic complexes that, in the mobile
333 phase, would be analyzed as multiple smaller molecules (e.g., Gadopentetic acid dimeglu-
334 mine salt). Also, another case was a surfactant containing a positive and negative charge
335 (i.e., 4-Dodecyl-2-[(2-nitrophenyl)azo]phenol). This case was a chemical that falls ‘outside’
336 of the RPLC space due to its predicted XLogP value of 10.452. However, the charges on
337 this molecule would make it difficult to calculate this value accurately. Lexidronam was one
338 of the ‘maybe’ cases that was classified as ‘outside’, due to a large leverage value of 26.0
339 and the fact that it elutes at t_0 (i.e., amide scale r_i of 206 versus urea $r_i = 200$), indicating
340 the need for special gradients to be able to retain such a chemical. As for the ‘inside’ cases
341 that were wrongly classified as ‘outside’, generally larger, branched (e.g., SCHEMBL312614),
342 or hydrolyzing (e.g., Bis[2-(perfluorohexyl)ethyl] Phosphate, respectively) chemicals showed
343 higher likelihood of such misclassifications. Again these are structures that may require very
344 specific adjustment of experimental condition (e.g., pH of mobile phase) to fit them within
345 the RPLC analyzable chemical subspace.

346

347 Overall, our RPLC classification model was highly successful in identifying the chemical
348 structures that are easily analyzable via RPLC (i.e., ‘inside’ cases) as well as the ‘maybe’ and
349 ‘outside’ cases. The classification model used a combination of similar molecular fingerprints
350 as those used by the three r_i models, taking advantage of all the structural information.

351 **NORMAN SusDat Chemical Space Prediction**

352 Finally, the RPLC classification model was applied to a set of small molecules (i.e., molecular
353 weight < 1000) from the NORMAN SusDat database. In total, 80503 chemicals were within
354 the applicability domain with leverage values ≤ 0.209 , 6570 compounds had leverage values
355 between 0.209 and 1, and 4664 compounds had even larger leverages. This showed that the

356 RPLC classification model was suitable for a large variety, 87.8%, of compounds present in
357 SusDat. The model predicted that 79.0% of the compounds would fit ‘inside’ the RPLC
358 subspace, 2.0% was ‘maybe’ in this space, and 19.1% was ‘outside’ of the RPLC subspace.
359 Examples of molecules classified as ‘inside’, ‘maybe’, and ‘outside’ were carbamazepine, su-
360 dan I, and coronene, respectively. When comparing the relationship between XlogP and
361 r_i , it is clearly observable that these parameters, even though relatively linear, are insuffi-
362 cient to determine if a chemical fits the RPLC subspace, figure 3. In figures S16, S17, and
363 S18, the XlogP values of the chemicals with the same r_i range vary between -10 to +10 units.

364

365 Using the developed classification models implies that for screening RPLC samples against
366 databases such as SusDat, 1/5 of the overall time can be saved, which becomes even more
367 significant when applying it to larger sample sets. Additionally, this will result in higher
368 confidence identifications when performing database matching for an RPLC NTA method
369 with SusDat, by reducing the overall number of potential candidates and thus false positive
370 identifications.

371

372 The amide r_i model is the least suited scale based on its applicability domain coverage
373 since only 44500 (i.e., 48.5%) chemicals fell within the applicability domain. For the chem-
374 icals that were outside the applicability domain, 18988 had a leverage value between 0.189
375 and 1 (i.e., similar to the full training set) and 28249 had an even higher leverage value. As
376 for the UoA and cocamide r_i models, 71022 (i.e., 77.4%) and 74252 (i.e., 80.9%) compounds
377 were within the applicability domain. For the UoA model, 3421 and 17294 chemicals had a
378 leverage value below and above 1, respectively, and the cocamide model had 5947 chemicals
379 with a leverage value below 1 and 11538 chemicals with higher leverage values. Figures S16,
380 S17, and S18 show the coverage of the ‘inside’, ‘maybe’, and ‘outside’ RPLC classes in terms
381 of the XLogP values versus the predicted r_i values for the amide, UoA, and cocamide series.
382 As expected the chemicals classified as ‘maybe’ inside RPLC are mainly clustering around

383 the lower and higher r_i values. While the chemicals classified as ‘outside’ the RPLC space
384 span the entire r_i range for each of the three r_i series, suggesting that r_i prediction would
385 also be insufficient to define the boundaries of the RPLC subspace.

386

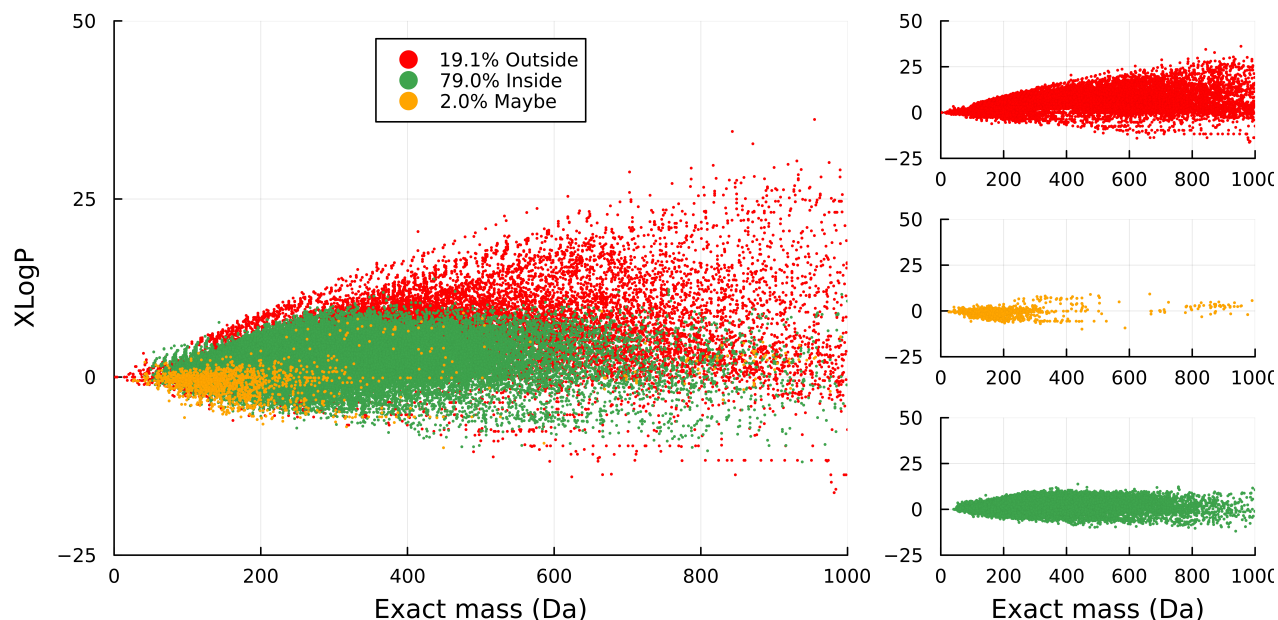


Figure 3: XLogP values versus the molecular weight for the NORMAN SusDat database compounds with a molecular weight below 1000 Da. In red, orange, and green are the compounds that were classified as ‘outside’, ‘maybe’, and ‘inside’ the RPLC chemical space, respectively. The subplots on the left show the coverage of the individual classes.

387 Potentials and Limitations

388 Overall, we developed four models for exploration of the RPLC subspace. The r_i regression
389 models showed that fingerprints can be used for describing RPLC retention indices. Con-
390 sequently, these fingerprints were used for RPLC classification model building. This model
391 was able to predict whether chemicals were ‘inside’, ‘maybe’ inside, or ‘outside’ of RPLC
392 chemical subspace with an accuracy of 92% on the test set. Applying the RPLC classification
393 model on NORMAN SusDat showed that 19.1% of the compounds were classified as ‘out-
394 side’ the RPLC subspace. This means that, when performing identification on NTA RPLC

395 samples, candidates classified as ‘outside’ compounds are unlikely to be the true structure of
396 the chemical and can be removed to reduce the number of false positive identifications. In
397 terms of suspect screening, it can save computational time since the ‘outside’ chemicals fall
398 ‘outside’ of the RPLC subspace and thus should not be screened for. Additionally, 87.8%
399 of NORMAN SusDat was within the applicability domain of the RPLC classifier, showing
400 good coverage of a variety of compounds. The RPLC classification model also showed that
401 the XLogP or r_i values alone are not sufficient to define the RPLC subspace.

402

403 The RPLC classification model overall did have more difficulties with regard to more
404 bulky and branched or surfactant-like chemicals. Additionally, the model was not able to
405 properly predict the RPLC subspace class of chemicals that are organic complexes, due to
406 the fact that in solution those are dissociated into multiple individual structures. The latter
407 is not a major limitation for the model itself, since, using expert knowledge, they can be
408 easily identified. Generally, as knowledge on analyzable chemicals with RPLC grows, the
409 model could easily be rebuilt and expanded for the range of analytes. Ideally, when sufficient
410 data becomes available, selectivity classification models could be constructed for other se-
411 lectivities (e.g., HILIC). This allows for further understanding of what part of the chemical
412 space is actually covered by the selectivities used in NTA and what we are missing.

413

414 Moreover, the RPLC classification model uses a data driven approach and is intended
415 for quick screening of the RPLC chemical space. The model assumes that compounds are
416 analyzable with RPLC regardless of the chemicals solubility, experimental parameters, or
417 pretreatment steps taken. This means that it cannot be assumed that chemicals ‘inside’ the
418 RPLC space will be analyzable with every RPLC method. Here, the method subspace plays
419 a major role when looking at what individual NTA methods can cover, becoming an even
420 more complex issue due to the fact that sample pretreatment, gradient program’s, and RP
421 column selectivities play a large influence on this. Defining the method chemical space would

422 be the next step in understanding what part of the vast chemical space we are covering and,
423 more importantly, excluding with our current NTA methods.

424

425 **Acknowledgement**

426 The authors thank the Environmental Monitoring and Computational Mass Spectrometry
427 (www.emcms.info) group for their insights and feedback. The Queensland Alliance for En-
428 vironmental Health Sciences. Finally, the University of Queensland gratefully acknowledges
429 the financial support from the Queensland Department of Health. J.W.O is the recipient of
430 an NHMRC Emerging Leadership Fellowship (EL1 2009209).

431 **Supporting Information Available**

432 Overview of performance for using different types of molecular fingerprints, composition of
433 reduced PubChem fingerprints, optimization, prediction, leverage, and feature importance
434 results for the 3 RF regression models and the RPLC classification model, and the RPLC
435 classification of NORMAN SusDat visualized by plotting the XLogP values versus the pre-
436 dicted r_i values for the three r_i regression models.

437 **Author Information**

438 Corresponding Author:

439 Saer Samanipour

440 Van 't hoff institute for molecular sciences (HIMS),

441 University of Amsterdam,

442 the Netherlands

443 Email: s.samanipour@uva.nl

444

445 Denice van Herwerden

446 Van 't hof institute for molecular sciences (HIMS),

447 University of Amsterdam,

448 the Netherlands

449 Email: d.vanherwerden@uva.nl

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562 **TOC Graphic**

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