Comparison of nanozymatic behavior of protein-protected gold and SiO<sub>2</sub>@gold

nanoparticles

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**Abstract** 

Herein, the nanozymatic behavior of protein-protected gold and SiO<sub>2</sub>@gold nanoparticles

were evaluated and their results compared with each other. The results showed that both

protein-protected gold and SiO<sub>2</sub>@gold nanoparticles reveal intrinsic peroxidase-like

activity. Hence, to precise comparison of nanozymatic behavior of these nanozymes, the

kinetic studies were performed using the Michaelis-Menten steady-state kinetics model

and the velocity and affinity factors were calculated for both nanozyme and then utilized

as a reliable way for comparison of nanozymatic behavior of these nanozymes. The

results showed that the V<sub>max</sub> of protein-protected gold nanoparticles was 12.0-fold higher

than that of SiO<sub>2</sub>@gold nanoparticles, revealing that the catalytic efficiency of protein-

protected gold nanoparticles is 12.0-fold higher than SiO<sub>2</sub>@AuNPs nanocomposite.

Besides, the Km value of SiO<sub>2</sub>@gold nanoparticles was 2-order higher than that of protein-

protected gold nanoparticles, indicating that the substrate affinity toward protein-

protected gold nanoparticles is 2.0-order higher than the SiO<sub>2</sub>@gold nanoparticles. Based

on the results of this work it can be concluded that protein-protected gold nanoparticles

are more efficient nanozymes than SiO<sub>2</sub>@gold nanoparticles.

Keywords: Nanozyme; Artificial enzymes; Protein-protected gold nanoparticles;

SiO<sub>2</sub>@gold nanoparticle

1. Introduction

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Importance and practical application of nanotechnology in modern life lead to design and synthesis of different nanomaterials with optical [1-3], catalytic [4, 5], anti-cancer features [6], medical [7], or anti-bacterial [8, 9] characteristics such as carbon and metalbased nanoparticles [10, 11], quantum dots [12, 13], metal oxide nanoparticle [14], magnetic nanoparticles [15], and metal-organic frameworks [16, 17], etc. Among various nanoparticles with different properties, recently, nanomaterials with enzyme-like properties, called nanozymes have been widely utilized for catalyzing industrial, clinical, and environmental enzyme-mediated reactions under harsh conditions [18-26]. The most significant advantage of these nanozymes compared to the native enzymes is their lower cost, higher efficiency, and especially, their high cycling stability and recyclability [19, 27-30]. Up to now, different nanoparticles with intrinsic peroxidase-like activity were designed and synthesized, for instance, Mn<sub>3</sub> O<sub>4</sub> nanozymes [31], Cu-CuFe<sub>2</sub>O<sub>4</sub> nanozymes [32], BSA-stabilized manganese dioxide nanoparticles [33], BSA-stabilized manganese phosphate nanoflower [34], carbon nanozymes [35], silica-coated-magnetic nanoparticles [36], MnO<sub>2</sub> nanoparticles [37], Fe<sub>3</sub>O<sub>4</sub> nanozymes [38], self-cascade pyrite nanozymes [39], metal-organic frameworks [40], gold nanozymes [41-45], S/N codoped carbon nanozymes [46], and silver nanoparticles [47-52]. Among the different nanomaterials with excellent peroxidase-like activity, gold-based nanozymes have been widely for developing nanozyme-based sensors [53, 54], nanozyme-based cancer treatment [55], and nanozymemediated dye degradation [56]. Moreover, since the first report of patients infected with the new infection disease, COVID-19 in 2019 [57, 58], nanozyme-based methods have been developed for fast clinical diagnosis of this pandemic infection [59-62]. Hence, evaluation of their biochemical features and enzyme-like properties is important for developing nanozyme-based systems with better figures of merit. In this regard, the biochemical behavior of enzyme-like nanosilver was also investigated by our research group [63]. Besides, recently, our research group reported a research article on the investigation of biochemical behaviors of BSA-stabilized gold nanoparticles [64, 65].

Herein, the nanozymatic behavior of protein-protected gold and SiO<sub>2</sub>@gold nanoparticles were evaluated and their results compared with each other. The results showed that both protein-protected gold and SiO<sub>2</sub>@gold nanoparticles reveal intrinsic peroxidase-like activity. Hence, to precise comparison of nanozymatic behavior of these nanozymes, the kinetic studies were performed using the Michaelis–Menten steady-state kinetics model and the velocity and affinity factors were calculated for both nanozyme and then utilized as a reliable way for comparison of nanozymatic behavior of these nanozymes. Based on the results of this work it can be concluded that protein-protected gold nanoparticles are more efficient nanozymes than SiO<sub>2</sub>@gold nanoparticles.

### 2. Experimental

# 2.1. Synthesis of nanomaterials

For the synthesis of protein-protected gold nanoparticles, 10.0 mM HAuCl<sub>4</sub>.4H<sub>2</sub>O (5.0 mL) was introduced to 50 mg mL<sup>-1</sup> bovine serum albumin (BSA; 5.0 mL), followed by stirring at 37 °C and adding 1.0 M NaOH to adjust pH. The solution was incubated at 37 °C for 12 hours to complete the synthesis process. For synthesis, the SiO<sub>2</sub>/gold nanoparticles, in experiment, 1.0 mL of HAuCl<sub>4</sub>, typical tetrakis(hydroxymethyl)phosphonium chloride, and 500.0 µL NaOH (2.0 M) were introduced in 50.0 mL deionized water. The resulting mixture was stirred for about 1.0 hour to prepare the colloidal gold solution. To synthesize the amino-functional SiO<sub>2</sub> NPs, 1.5 mL of TEOS was added into 40.0 mL of pure ethanol, followed by the addition of 3.0 mL of NH<sub>4</sub>OH. The mixture was stirred for about 20.0 hours. Afterward, 20.0 mg of the resulting precipitate was treated with 600.0 µL of APTES for preparation of the final product. Afterward, 10.0 mg of the amino-functional SiO<sub>2</sub> NPs were incubated with a solution of colloidal gold (prepared in section 2.3) for about 12.0 hours. Afterward, the product was separated from the reaction media upon centrifuge at 10000 rpm for 30.0 min. The resulting product was dispersed in 1 mg mL<sup>-1</sup> of PVP (stabilizer) aqueous

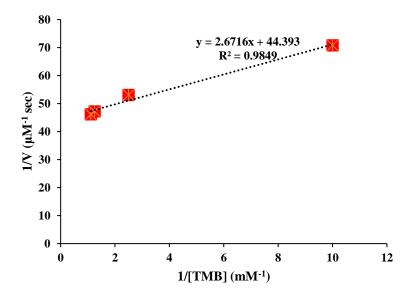
solution, followed by the addition of  $60.0~\mu L$  of HAuCl<sub>4</sub> (10.0~mM) and  $120.0~\mu L$  of 10.0~mM ascorbic acid (reducing agent) into the reaction media. The synthesis process was followed by stirring the above-mentioned mixture for about 5 min. Afterward, the resulting SiO<sub>2</sub>/gold nanoparticles were collected, washed, and then dried at room temperature.

#### 3. Results and discussion

## 3.1. Quantification of maxium velocity velocity and affinity factor

Kinetic studies were carried out to calculate the kinetic parameters of the as-prepared SiO<sub>2</sub>/gold nanoparticles toward 2-electron reversible oxidation of TMB. In fact, the kinetic parameters of an enzyme were previously well defined with numeric values including affinity constant (Km) and maximum enzymatic velocity (Vmax) utilizing the Michaelis-Menten steady-state kinetics model. The V<sub>max</sub> value reflects the intrinsic properties of an enzyme or nanozyme and is defined as the highest possible rate of the nanozymecatalyzed reaction when all enzyme molecules or all nanozyme particles are saturated with the substrate which points to the catalytic efficiency of an enzyme or nanozyme. Hence, the higher value of V<sub>max</sub> for an enzymatic/nanozymatic reaction can be assigned to the higher catalytic efficiency of the enzyme or nanozyme [66-71]. In contrast, the affinity of the substrate of an enzyme or nanozyme for interaction with its active site is represented by the K<sub>m</sub>, as reported [66]. In fact, the lower values of K<sub>m</sub> pointed to the higher affinity of the substrate for binding to the enzyme/nanozyme active site/nodes [17]. Therefore, to evaluate the kinetics parameters of the as-prepared nanocomposite, the Michaelis-Menten model was obtained. For accurate estimation of Km and Vmax of the SiO<sub>2</sub>/gold nanoparticles-mediated oxidation reaction, the Lineweaver–Burk plot was also constructed for the SiO<sub>2</sub>@gold nanocomposite-mediated reaction (Figure 1). The results revealed a V<sub>max</sub> of 0.022 µM min<sup>-1</sup> and a K<sub>m</sub> as very low as 0.06 mM for the SiO<sub>2</sub>@gold nanocomposite. Considering the low K<sub>m</sub> value of the as-synthesized nanocomposite, it

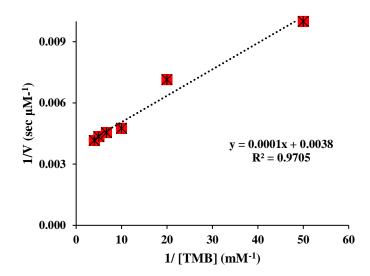
can be concluded that the as-prepared nanocomposite shows high substrate affinity, as reported [17]. Besides, obtaining a  $V_{max}$  of 0.022  $\mu M$  min<sup>-1</sup>, revealed that the as-prepared nanocomposite has a very good catalytic efficiency.



**Figure 1.** Lineweaver–Burk plot for the as-prepared SiO<sub>2</sub>@gold nanoparticles to estimate the kinetic factors of nanozymatic reaction catalyzed by SiO<sub>2</sub>@gold nanoparticles.

Besides, the kinetics studies for exploring more precise on reporting of the peroxidase-like activity and enzymatic power of the as-prepared protein-protected gold nanoparticles were carried out by estimating their activity as a function of substrate concentration and then, the standard Lineweaver–Burk plot was provided by plotting the inverse of the velocity of the nanozymatic reaction (V-1) as a function of [substrate]-1 for estimating the nanozymatic kinetics parameters. To estimate the kinetic parameters of gold nanozymes toward TMB oxidation, the Lineweaver–Burk plot was constructed

(Figure 2). Considering the results obtained in Figure 2, the V<sub>max</sub> and K<sub>m</sub> of the asmentioned gold nanozymes were calculated at about 263 nM sec<sup>-1</sup> and 0.03 mM, in order.



**Figure 2.** Lineweaver–Burk plot for the as-prepared protein-protected gold nanoparticles to estimate the kinetic factors of nanozymatic reaction catalyzed by protein-protected gold nanoparticles

# 2. Comparsion of nanozymatic behavior

According to the experimental results of this work, the V<sub>max</sub> of gold nanoparticles was 12.0-fold higher than that of SiO<sub>2</sub>/Au nanocomposite, revealing that the catalytic efficiency of BSA-stabilized gold nanoparticles is 12.0-fold higher than SiO<sub>2</sub>/Au nanocomposite. Besides, the K<sub>m</sub> value of SiO<sub>2</sub>/Au nanocomposite was 2-order higher than that of gold nanoparticles, indicating that the substrate affinity toward gold nanoparticles is 2.0-order higher than the SiO<sub>2</sub>/Au nanocomposite. Since, the active nodes of both nanozymes are the same (i.e., gold), the difference between their catalytic efficiency and

affinity can be assigned to their different sizes and the ability of the active nodes to bind the substrate. Based on the results of this work, small-size gold nanoparticles are characteristically more efficient peroxidase mimic materials than the SiO<sub>2</sub>/Au nanocomposite.

#### 4. Conclusions

Herein, the nanozymatic behavior of protein-protected gold and SiO2@gold nanoparticles were evaluated and their results compared with each other. The results showed that both protein-protected gold and SiO2@gold nanoparticles reveal intrinsic peroxidase-like activity. Hence, to precise comparison of nanozymatic behavior of these nanozymes, the kinetic studies were performed using the Michaelis–Menten steady-state kinetics model and the velocity and affinity factors were calculated for both nanozyme and then utilized as a reliable way for comparison of nanozymatic behavior of these nanozymes. The results showed that the Vmax of protein-protected gold nanoparticles was 12.0-fold higher than that of SiO2@gold nanoparticles, revealing that the catalytic efficiency of protein-protected gold nanoparticles is 12.0-fold higher than SiO2@AuNPs nanocomposite. Besides, the Km value of SiO2@gold nanoparticles was 2-order higher than that of protein-protected gold nanoparticles, indicating that the substrate affinity toward protein-protected gold nanoparticles is 2.0-order higher than the SiO2@gold nanoparticles. Based on the results of this work it can be concluded that protein-protected gold nanoparticles are more efficient nanozymes than SiO2@gold nanoparticles.

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### Conflict of interest

None.

### 5. References

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