

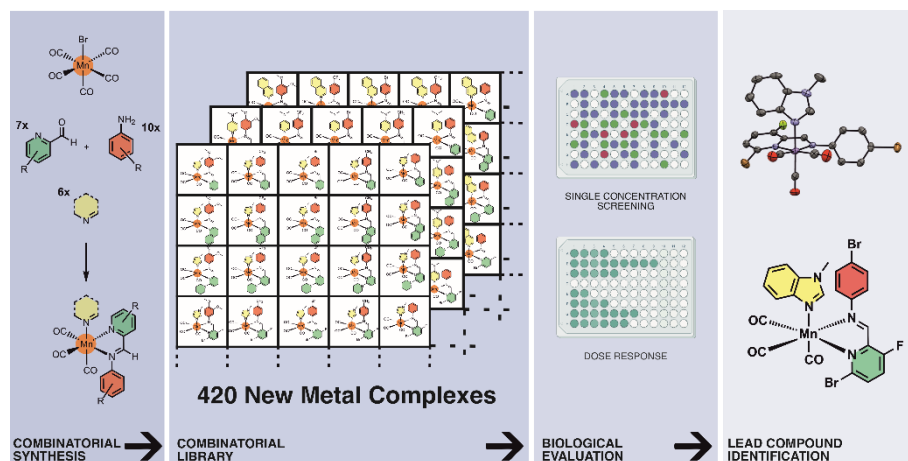
Discovery of Antibacterial Manganese(I) Tricarbonyl Complexes through Combinatorial Chemistry

Mirco Scaccaglia^{1,3}, Silvana Pinelli², Giorgio Pelosi¹, Angelo Frei^{*3}

¹ Department of Chemistry, Life Sciences and Environmental Sustainability, University of Parma, 43124 Parma, Italy

² Department of Medicine and Surgery, University of Parma, Via Gramsci 14, 43126 Parma, Italy

³ Department of Chemistry, Biochemistry & Pharmaceutical Sciences, University of Bern, Freiestrasse 3, 3012 Bern, Switzerland



Abstract

The continuous rise of antimicrobial resistance is a serious threat to human health and already causing hundreds of thousands of deaths each year. While natural products and synthetic organic small molecules have provided the majority of our current antibiotic arsenal, they are falling short in providing new drugs with novel modes of action able to treat multidrug resistant bacteria. Metal complexes have recently shown promising results as antimicrobial agents, but the number of studied compounds is still vanishingly small, making it difficult to identify promising compound classes or elucidate structure-activity relationships. To accelerate the pace of discovery we have applied a combinatorial chemistry approach to the synthesis of metalloantibiotics. Utilizing robust Schiff-base chemistry and combining 7 picolinaldehydes with 10 aniline derivatives, and 5 axial ligands we have prepared a library of 420 novel manganese tricarbonyl complexes. All compounds were evaluated for their antibacterial properties and 10 lead compounds were identified, re-synthesized and fully characterized. All 10 compounds showed high and broad activity against Gram-positive bacteria. The best manganese complex displayed low toxicity against human cells with a therapeutic index of >100. In initial mode of action studies, we show that it targets the bacterial membrane without inducing pore formation or depolarisation. Instead, it releases its carbon monoxide ligands around the membrane and inhibits the bacterial respiratory chain. This work demonstrates that large numbers of metal complexes can be accessed through combinatorial synthesis and evaluated for their antibacterial potential, allowing for the rapid identification of promising metalloantibiotic lead compounds.

Introduction

In 2019, over 1.3 million patients are estimated to have died from infections resistant to our current antibiotic arsenal.¹ Without urgent action and a strong replenishment of the antibiotic drug pipeline, the number of deaths is bound to increase further as antimicrobial resistance (AMR) is on the rise across the world. Unfortunately, the current clinical antibiotic candidates are both few in number and lacking in new targets, limiting their effective shelf-life from the start.² Over the last years, alternatives to the small organic molecule approaches to treat bacterial infections have increasingly been explored.³ We have recently shown that transition metal complexes display promising antimicrobial properties. In a systematic study of over 300,000 tested molecules by the Community for Open Antimicrobial Drug Discovery (CO-ADD), metal-containing compounds had a 10x higher hit-rate against critical ESKAPE (*Enterococcus faecium*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, and *Enterobacter spp.*) pathogens compared to regular organic molecules. Additionally, metal complexes did not show increased rates of cytotoxicity or haemolysis in this comparison.⁴ In the last decade, a few key studies have investigated metal complexes for their antibacterial properties in-depth with several showing promising *in vitro* and *in vivo* activity.^{4–12}

Amongst the only small number of metal compound classes studied for their antibacterial properties, the metal carbonyl core (M(CO)₃) is a commonly occurring motif. Patra *et al.* were the first to highlight that the antibacterial properties of a trimetallic metal compound could be attributed to the presence of a rhenium tricarbonyl moiety in the structure.¹³ Mode of action studies attributed the activity of this compound to its detrimental effect on the bacterial cell membrane, causing depolarization but not pore formation. However, due to unfavorable solubility and cytotoxicity properties, the compounds were not evaluated further. More recently, several groups have investigated metal complexes based on the Re(CO)₃-scaffold. Frei *et al.* reported on dual-mode of action bisquinoline rhenium tricarbonyl complexes which exhibited both light dependent and independent antibacterial activity against drug-resistant Gram-positive and Gram-negative bacteria.¹⁴ The group of Zobi have shown that fac-[Re(CO)₃] diimine compounds show activity against Gram-positive bacteria, including drug-resistant ones. The complexes were shown to be effective in a Zebrafish infection model.^{15,16}

Manganese, the first-row transition metal congener of rhenium, has also been explored by several groups for its antibacterial properties. Due to the higher lability of the carbonyl ligands, manganese tricarbonyl complexes have been studied extensively as CO-releasing molecules (CORM). Mendes *et al.* have synthesized and studied a series of manganese and rhenium tricarbonyl compounds bearing the antifungal drug clotrimazole as an axial ligand. A detailed study of one rhenium complex suggests that part of its mode of action involves inhibition of peptidoglycan synthesis in Gram-positive bacteria by coordinating to lipid I, lipid II and undecaprenyl-pyrophosphate C55PP.¹⁷ The group of Schatzschneider has first reported the antimicrobial activity of a manganese based light-activated CORM (photoCORM) in 2014.¹⁸ This complex and related ones have been extensively studied for their antimicrobial properties in the last decade.^{19–22} The same group also described

Mn(CO)₃ compounds with light independent antibacterial effects *in vitro* and *in vivo*.^{23–25} Ward *et al.* reported a tryptophane-based Mn(CO)₃ photoCORM which showed good antibacterial activity against *N. gonorrhoeae* and *S. aureus* upon light irradiation.^{26,27} A water-soluble Mn(CO)₄-based CORM (CORM-401) was first highlighted by Crook *et al.* and its antimicrobial properties were studied revealed by Wareham *et al.*^{28,29} The group showed that CORM-401 does not inhibit respiration, but instead disrupts cytoplasmic ion balance and induces, amongst other effects, osmotic stress. Of note, the antimicrobial effects were only found at high compound concentrations and toxicity against eukaryotic cells was observed.²⁹

While the potential of metal complexes and the metal-carbonyl scaffold in particular has clearly been established in recent years, current approaches usually focus on the exploration of only a handful of compounds at a time. This makes the exploration of this compound class inefficient as well as time- and resource-intensive. Additionally, while promising molecules have been found, very little is understood about the structure-activity relationships amongst them.

Combinatorial synthesis has been applied successfully in conventional medicinal chemistry to efficiently explore the organic chemical space for promising compounds.³⁰ In particular its combination with DNA-encoded chemical libraries has led to several lead-compounds that have advanced to clinical trials.³¹ The combinatorial concept has been applied to the synthesis of metal complexes as well, albeit only in a limited number of examples. The group of Bernhard has utilized various permutations of bidentate N[^]N and C[^]N-type ligands to prepare large libraries of iridium complexes with promising photophysical properties.^{32–34} The group of Schatzschneider has recently reported the application of so-called iClick reactions to combinatorially prepare [Ru(triazolato)(N[^]N)(terpy)] complexes.³⁵

In the context of medicinally applied combinatorial metal complex synthesis, the group of Ang has pioneered a three-component assembly (3CA) protocol for the synthesis of Schiff-base Arene ruthenium(II) complexes (Figure 1A).³⁶ The group utilised different ruthenium arene precursors in combination with robust Schiff-base ligands formed by the combination of different picolinaldehydes with aniline derivatives to form stable piano-stool type complexes under mild conditions. This approach has been applied to synthesize hundreds of different ruthenium complexes and explore their anticancer, antibacterial and catalytic properties.^{37–40} Konkankit *et al.* have applied a similar approach, albeit only with two components, to the preparation of rhenium carbonyl complexes, identifying some with potent anticancer properties (Figure 1B).^{41,42} Very recently Kench *et al.* reported an approach similar to the Bernhard group for the preparation of polypyridyl iridium complexes. However, in this case the workflow was optimised to obtain photo-cytotoxic compounds with activity against human cancer cells (Figure 1C).⁴³

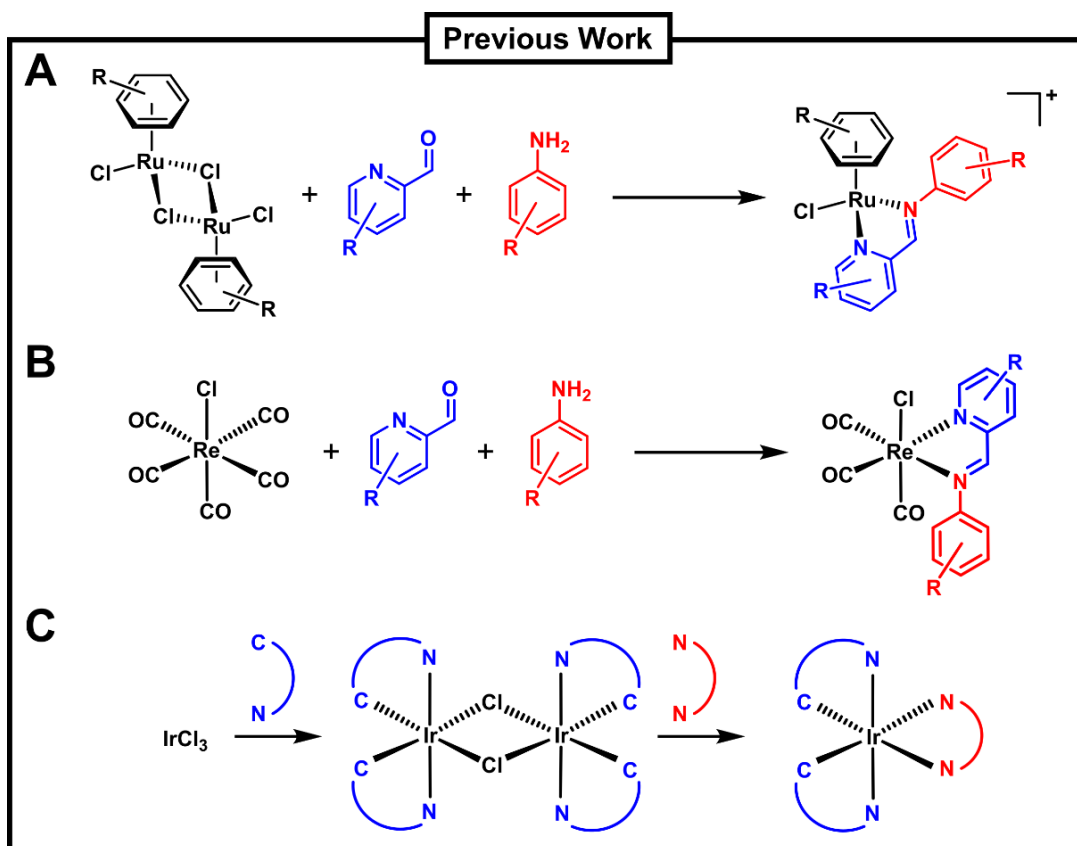


Figure 1. Overview of previous work on the combinatorial synthesis of metal complexes with biological applications by the groups of (A) Ang, (B) Wilson and (C) Vilar.

In light of the promising antibacterial properties reported for carbonyl and specifically manganese tricarbonyl complexes, we have adapted the combinatorial chemistry approach to the systematic synthesis of these compounds. Herein we report the rapid, efficient and economical synthesis of 420 novel manganese tricarbonyl Schiff-base compounds. The complexes were characterised and explored for their antibacterial and toxicity properties. We identified and re-synthesized 10 lead complexes which were isolated and fully characterised. These compounds were further studied for their physicochemical properties and extended biological studies were conducted.

Results and Discussion

Optimization of MnCO_3 combinatorial synthesis and preparation of compound library.

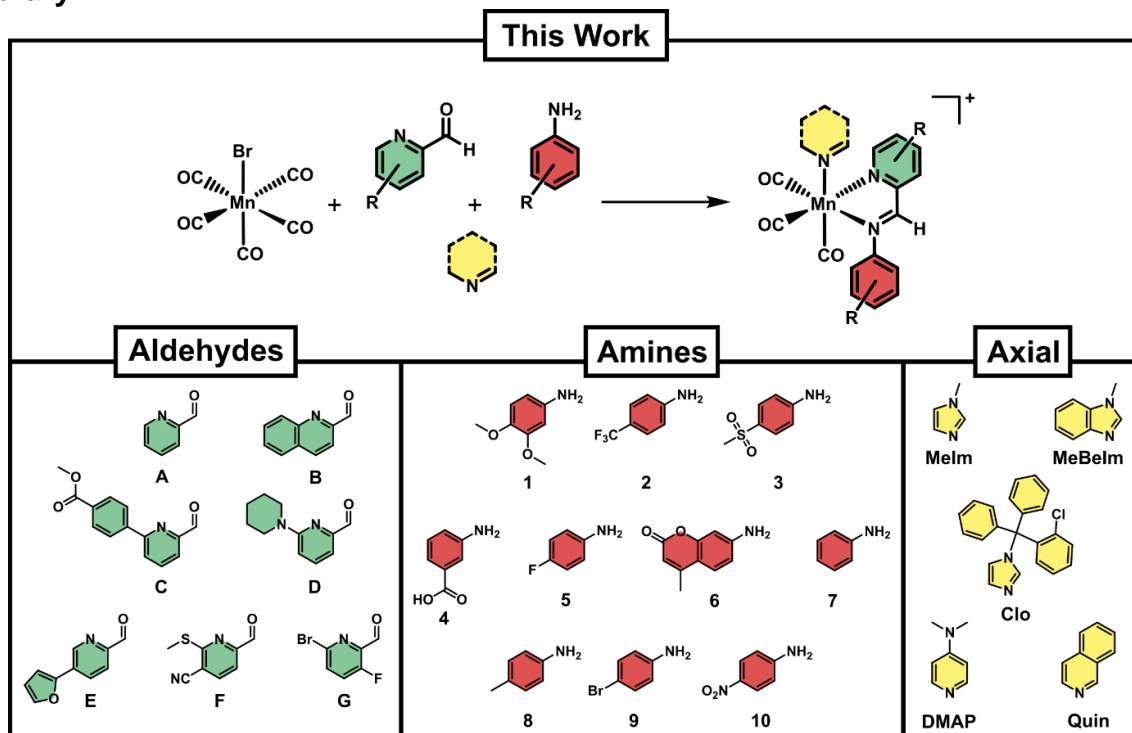


Figure 2. General reaction scheme of the combinatorial synthesis of manganese tricarbonyl Schiff-base complexes and overview of chemical building blocks selected for the synthesis of 420 novel complexes (all 70 possible Schiff-base complexes where also synthesized with no axial ligand).

For a reaction protocol to be applicable to combinatorial chemistry, the conditions need to a) be sufficiently robust to tolerate a variety of building blocks and b) be adaptable to a relative high throughput set up. We chose $\text{MnBr}(\text{CO})_5$ as our starting material due to its commercial availability and its established use as a precursor to $\text{Mn}(\text{CO})_3$ type compounds. The formation of Schiff-base type ligands starting from picolinaldehydes and aniline derivatives has proven a promising strategy as the reaction occurs under mild conditions and in a variety of solvents, including water.^{36,41} In order to add a third dimension to the diversity of synthesized compounds, we introduced an axial ligand to the third available coordination site (Figure 2).

Initial reaction attempts starting directly from $\text{MnBr}(\text{CO})_5$ gave low conversion rates and required long reaction times and high temperatures as determined by LC-MS. Through extensive screening of reaction conditions, we determined that a pre-activation of the $\text{MnBr}(\text{CO})_5$ precursor by reaction with one equivalent silver triflate under inert atmosphere and in dry THF led to a more reactive intermediate. This 'solvated' manganese tricarbonyl could be directly utilised for further reaction with the Schiff-base components and the axial ligand in one pot requiring only 90 min heating at 70 °C under ambient atmosphere. Initially, the reactions were attempted in DMSO and DMSO/ H_2O mixtures to enable direct biological screening of the reaction crudes. However, the conversion rates in these solvents were not satisfactory. Eventually, it was found that THF provided the best compromise of good conversion rates and ease of removal after reaction completion.

Several considerations went into the selection of picolinaldehyde, aniline and axial ligand building blocks. A list of commercially available picolinaldehyde derivatives was obtained from the Reaxys platform. These compounds were further filtered by price and their chemical similarity was determined by Tanimoto indices.⁴⁴ Seven picolinaldehydes with maximized diversity and reasonable price were eventually obtained (Figure 2). In the case of anilines, a large number of these structures were available in our department. From these, a set of 10 maximally diverse (based on their Tanimoto similarity) were chosen for our library. Lastly, several compounds were screened as axial ligands in test-reactions, revealing that even with optimised conditions some axial ligands did not lead to sufficiently high conversion rates. The final selection of axial ligands (Figure 2) is hence a subset of the ones that were coordinating successfully under our reaction conditions. The antifungal clotrimazole (**clo**) has been utilised as a ligand in several other studies and found to convey some level of biological activity that goes beyond the activity of the parent compound.^{17,22,45} We have hence included it in our studies as a means to compare our compounds to others reported in the literature.

With the 10 anilines, 7 picolinaldehydes and 5 axial ligands in hand we proceeded to set up the combinatorial syntheses. Briefly, 70 reactions at a time were set up by combining equimolar amounts of pre-activated manganese carbonyl (20 mM, 100 μ L) together with the different building blocks from pre-prepared stock solutions in THF (40 mM, 50 μ L) in 500 μ L polypropylene tubes. The reaction vessels were sealed and heated in the dark for 120 minutes. The solvent was then removed *in vacuo* and the dried reaction crudes re-dissolved in 100 μ L of DMSO (stock solution 20 mM) and diluted in acetonitrile for LC-MS analysis. The LC-MS spectra were analyzed for both target product formation and conversion percentage (a representative sample of LC-MS spectra are given in the supplementary materials, Figure S1). Altogether 420 novel manganese complexes could be prepared and characterised by LC-MS with minimal use of reagents (less than 1 mg bromopentacarbonylmanganese per reaction).

Antimicrobial activity of combinatorial Mn(CO)₃ library

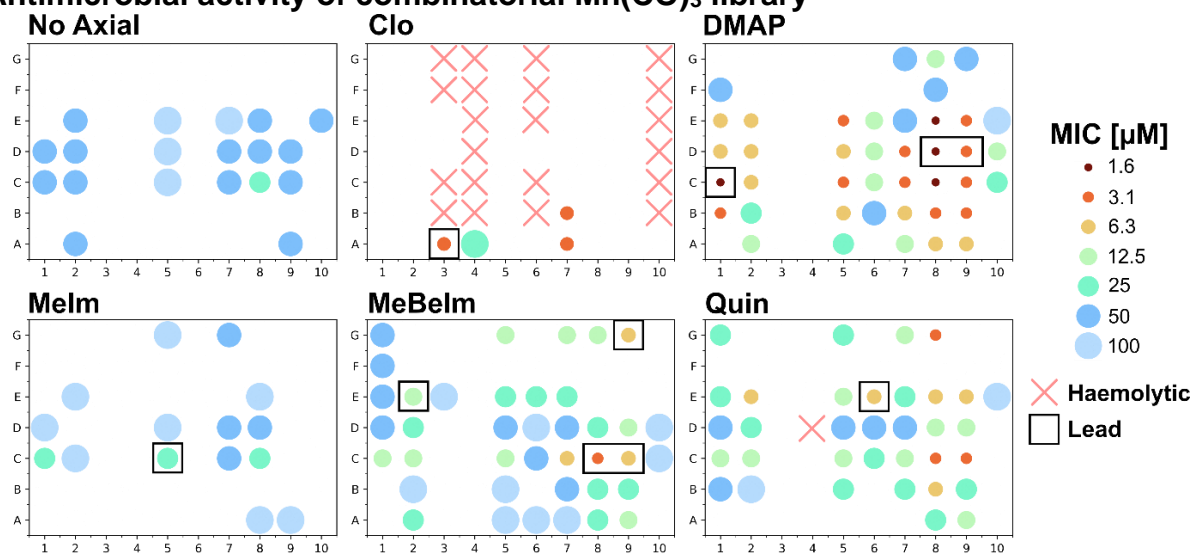


Figure 3. Overview of antibacterial activity of the 420 combinatorial manganese complexes against MRSA. Haemolytic properties are also indicated and the 10 selected lead compounds are highlighted.

To efficiently get a picture of the antibacterial activity profile of all 420 compounds we conducted a single-concentration screen of all crude reaction mixtures at 100 μM against the Gram-positive methicillin resistant *S. aureus* (MRSA) and the Gram-negative *Escherichia coli*. For compounds that showed complete growth inhibition under these conditions, we conducted a microdilution dose response assay to determine a minimum inhibitory concentration (Figure 3 and Table S1).

Firstly, in line with most tricarbonyl complexes reported in the literature, barely any activity was observed against the Gram-negative *E. coli*. While 38 compounds showed some inhibition in the single concentration assay at 100 μM , an MIC could only be determined for 5 compounds. The lowest determined MIC was 50 μM for **MnA7Clo**. Conversely much higher levels of activity were detected against the Gram-positive MRSA. Of all 420 compound crudes tested, an MIC could be determined for 152 (36.2%) of them (Figure S2). Only for 31 compounds where growth inhibition was observed in the single concentration assay at 100 μM no MIC could be detected at that or lower concentrations. If an active compound is defined to have an MIC of 12.5 μM or lower, 64 of the 420 compounds (15.3%) would classify as active. An impressive 39 (9.3%) had an MIC of 6.25 μM or lower and for 4 an MIC of 1.56 μM was found. To verify if the observed activity could stem from the building blocks instead of the complex, we also tested all picolinaldehydes, anilines and axial ligands. No activity was observed for any of the building blocks except for **clo** which showed an MIC of 12.5 μM against MRSA (Table S2), confirming that in most cases the observed activity is likely due to the formed manganese tricarbonyl complex. Significantly more active compounds are observed with the **MeBeIM**, **DMAp** and **Quin** axial ligands. In the picolinaldehydes building blocks we observe that 65% of all actives (i.e. MIC \leq 12.5 μM) contain one of the bicyclic picolinaldehydes i.e. **C**, **D** or **E**. On the other hand in the anilines, polar (**3**, **4** and **10**) and/or bulky (**6**) derivatives seemed to be unfavorable for antibacterial activity.

Overall, this hit-rate is impressive yet not entirely unsurprising considering the literature precedent on this compound class. To obtain a first indication on the potential toxicity of the synthesized complexes against human cells, we also tested all 420 crudes for haemolytic properties against human red blood cells. Only 22 compounds, 21 of which contained the **clo** axial ligand, showed any sign of haemolysis at 20 μM (Figure 3). Additionally, all compounds that displayed haemolysis in this assay did not have any antibacterial activity. Overall, this first toxicity screening indicates that the majority of these manganese Schiff-base compounds are non-haemolytic.

So far, all tested substances were reaction crudes containing the target compound with average purities of $64 \pm 14\%$. All assays so far were conducted with non-purified compounds to enable a significantly higher throughput in much less time. However, the ambiguous purity of the crudes leaves open the possibility that the observed activity could be due to side products or impurities instead of the putative metal complex.

Re-synthesis, purification and characterisation of lead compounds

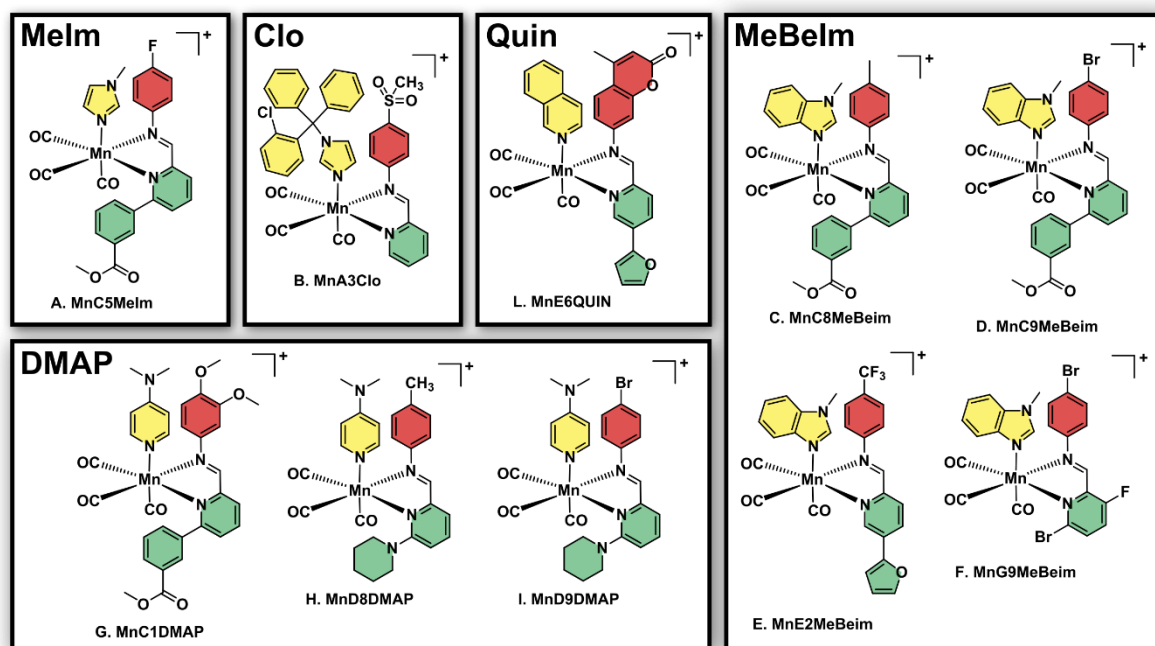


Figure 4. Structures of the 10 manganese lead compounds that were re-synthesized, purified and characterised after the initial combinatorial screening campaign.

After the preliminary biological assessment, we selected ten lead compounds for resynthesis (Figure 4). The selection of lead compounds was based on the determined antibacterial activity. Additionally, we aimed to maximize the chemical diversity of the lead compounds, aiming to have at least one example with each axial ligand included. The selected compounds were resynthesized *via* conventional batch-style chemistry on a ~20 mg/ 72 μmol scale and purified by preparative HPLC with yields ranging from 26-99% (in some cases the pure compound could be isolated directly by precipitation). The pure metal complexes were characterised by ^1H - and ^{13}C NMR, LC-MS, HR-MS and IR spectroscopy (see Supporting Information). Single crystals suitable for X-ray diffraction structural elucidation were obtained for compounds **MnD8DMAP**, **MnC1DMAP**, and **MnG9MeBeim** (Figure 5).

The X-ray structures of these compounds introduce an unexplored family of molecules in the CSD database. Structural analysis unveils the manganese atom's adoption of a pseudopentahedral configuration, wherein a *fac*- $\text{Mn}(\text{CO})_3$ moiety coordinates the bidentate Schiff base and the N-donor axial ligand sourced from either imidazole or pyridine. Notably, bond distances (as detailed in Table S3) exhibit minimal differences across the three structures in this study, aligning with expectations from the parent compounds.⁴⁶ Interestingly, the Schiff base's two rings remarkably deviate from planarity, with unusually large dihedral angles spanning between 37° and 46°.

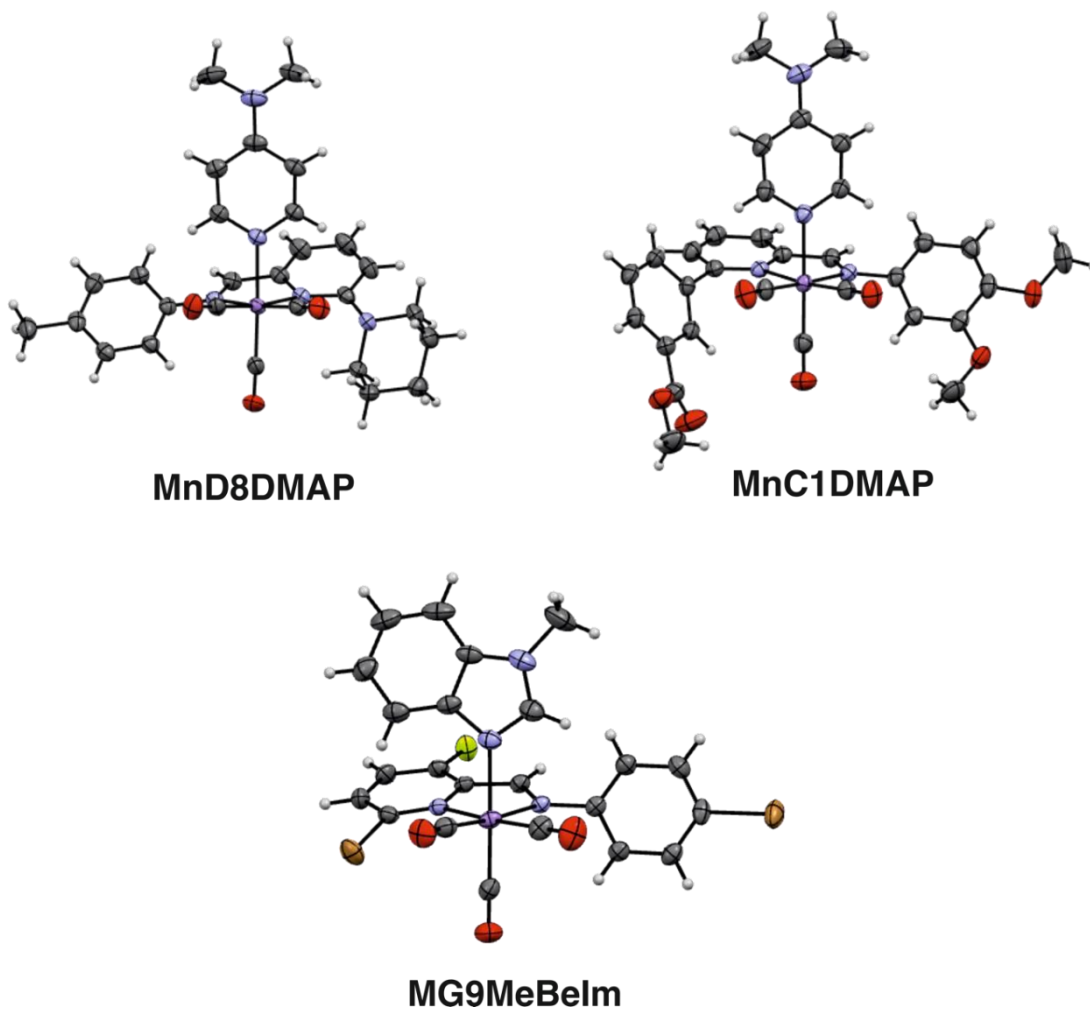


Figure 5. Molecular structure of the cationic moieties of MnD8DMAP, MnC1DMAP and MnG9MeBelm (CCDC No 2280008-10). Atomic displacement ellipsoids are shown at the 50% probability level. The triflate counterion and a co-crystallized solvents molecule are omitted for clarity. Color coding: carbon (grey), hydrogen (white), nitrogen (blue), oxygen (red), bromine (orange), manganese (purple).

Biological valuation of the lead manganese compounds

Table 1. Antibacterial activity against a selection of Gram-positive strains, toxicity data in HuDe cells and human red blood cells and therapeutic indices (TI) for the 10 synthesized lead compounds.

	MIC [μ M] vs. Gram-positive Bacteria						Toxicity [μ M]		
	MRSA	MSSA	Se	Bs	En	En VRE	CC ₅₀	HC ₅₀	TI
MnC5Melm	3.13-6.25	12.5-6.25	12.5	12.5-6.25	25	25	46.1 \pm 0.1	>200	7
MnA3Clo	1.56	1.56	3.13	3.13-1.56	6.25	3.13	37.1 \pm 0.2	68 \pm 15	24
MnC8MeBeim	3.13-1.56	6.25	6.25	6.25	12.5	6.25	18.0 \pm 0.2	77 \pm 10	6
MnC9MeBeim	3.13-1.56	3.13	6.25	3.13	12.5	12.5	16.8 \pm 0.2	95 \pm 13	5
MnE2MeBeim	3.13-1.56	3.13	6.25-3.13	3.13	12.5	12.5	7.2 \pm 0.3	49 \pm 12	2
MnG9MeBeim	0.78	<0.78	<0.78	3.13-1.56	6.25	6.25	85.2 \pm 0.1	117 \pm 12	109
MnC1DMAP	3.13-1.56	3.13-1.56	6.25	6.25-3.13	12.5	6.25	37.0 \pm 0.2	>200	12
MnD8DMAP	1.56	1.56-0.78	1.56	3.13-1.56	12.5	12.5	7.9 \pm 0.2	>200	5
MnD9DMAP	1.56	1.56	3.13	1.56	12.5	6.25	9.4 \pm 0.2	>200	6
MnE6Quin	6.25-3.13	6.25	12.5-6.25	6.25	50	25	36.3 \pm 0.1	>200	6
VAN [μg/mL]	1	1	1	0.25	1	8			

Antibacterial activity is displayed as MIC [μ M]. MRSA – methicillin resistant *S. aureus*; MSSA – methicillin susceptible *S. aureus*; Se – *S. epidermidis*; Bs – *Bacillus subtilis*; En – *Enterococcus spp* En VRE – *Enterococcus casseliflavus* VRE, CC₅₀ – HuDe cells; HC₅₀ – Human red blood cells; TI – therapeutic index, determined by dividing the lowest value between CC₅₀ and HC₅₀ with the lowest MIC value for each compound; VAN – Vancomycin. MIC determined with n=4 across two biological replicates.

With the pure lead compounds in hand, we conducted antibacterial activity assays against a selection of Gram-positive and Gram-negative bacteria (Table 1 and Table S4). The compounds showed generally good activity against a variety of strains, including the ESKAPE pathogens MRSA and vancomycin resistant *Enterococcus* clinical isolate strains. Some of the compounds showed moderate activity against the Gram-negative *A. baumannii* and *E. coli* indicating that structural optimisation might lead to a compound with broad spectrum antibacterial activity (Table S4). To ascertain whether the lead compounds are bacteriostatic or bactericidal the minimum bactericidal concentration (MBC) was determined against MRSA (Table S5). For all compounds except **MnC5Melm**, the MBC was found to be very close to the MIC, suggesting that most of the tested compounds are bactericidal.

To further evaluate the potential toxicity of the lead compounds, we re-measured haemolysis, determining HC₅₀ values and tested them for cytotoxicity against eukaryotic healthy HuDe human epithelial cells. Overall, the activity observed for the crude reaction mixtures translated to activity for the pure compound with high fidelity, validating the pursued combinatorial chemistry approach for the discovery of antibacterial metal complexes pursued in this project. Similarly, the haemolysis values determined with the pure compounds is in agreement with the one measured for the crudes i.e. no compound shows haemolysis under 49 μ M. On the other hand, relatively high levels of cytotoxicity against HuDe cells was observed for some of the compounds. Nevertheless, three of the lead compounds still displayed therapeutic indices (TI) of >10 with **MnG9MeBeim** having a TI of >100. The MICs of

MnG9MeBeim were comparable to the standard of care for Gram-positive infections, vancomycin.

As already mentioned, some manganese tricarbonyl complexes are known for their light-mediated carbon monoxide release. Indeed, we observed rapid decomposition of these compounds when exposed to natural daylight as evidenced by decolorisation of stock solutions. However, no such decolorisation was observed under dim-light conditions, and reproducible data was obtained across multiple biological and technical replicates. To further study the stability of these compounds, we measured their UV/Vis absorbance in different solvents (water, DMSO, HEPES, PBS) over time to see if any changes were visible (Figures S3-12). Over an 18-hour period, no decomposition was observed in DMSO and only small changes in absorbance were detected in water and the buffers.

Based on these extensive biological data, compound **MnG9MeBeim** was chosen as the sole lead compound for further studies due to its high level of antibacterial activity coupled with low haemolytic activity, cytotoxicity and high stability under biological conditions.

Characterisation of CO-releasing properties of **MnG9MeBeim**.

Based on previous studies of manganese tricarbonyl complexes, the involvement of CO release in the antibacterial mode of action is a plausible hypothesis. To assess whether CO is involved in the antibacterial activity of our lead compound we re-measured the MIC of the compound against MRSA in the presence and absence of haemoglobin (Hb), a well-established CO-scavenger. Indeed, we observed a significant 4-8x increase in MIC for **MnG9MeBeim** in the presence of 20 μM Hb. Next, we conducted a checkerboard assay varying both the concentrations of **MnG9MeBeim** and Hb (Figure 6A). A clear antagonistic ($\text{FICI} \gg 4$)⁴⁷ effect of Hb on the antibacterial activity of **MnG9MeBeim** was observed presumably by irreversibly binding the CO released by the manganese compound. However, even in the presence of large concentrations of Hb, the activity of **MnG9MeBeim** was not completely inhibited, suggesting that multiple mechanisms may be involved (the $\text{MIC}_{\text{MnG8MeBeim}}$ in the presence of 160 μM Hb was 50 μM). The release of CO was observed via UV/Vis spectroscopy, monitoring the absorbance changes of Hb in the presence of **MnG9MeBeim** over time. The conversion of deoxy-Hb to Hb-CO was observed over 8 h (25 °C) (Figure 6B). These results combined with the structural similarity and the common MnCO_3 scaffold in our library suggest that CO-release is a factor in the antibacterial effect of all these compounds. However, the distinct levels of antibacterial activity observed across this library suggests that the structure of the compound has a significant effect on the characteristics of the CO release and possibly bacterial uptake which in turn affects their efficacy against bacteria.

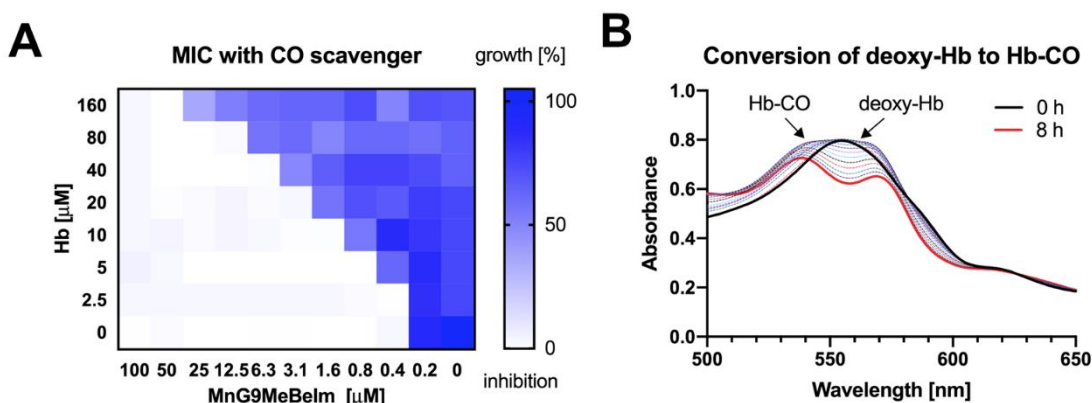


Figure 6. Characterisation of CO-releasing behaviour of **MnG9MeBeim**. (A) Checkerboard assay of the biological activity of the compound against MRSA displayed as growth percentage over the control, in the presence of different concentrations of the CO scavenger hemoglobin (B) Conversion of deoxyhemoglobin (80 μM , HEPES pH 7.4, 25 $^{\circ}\text{C}$) to carboxyhemoglobin in the presence of **MnG9MeBeim** (20 μM) monitored over an 8-hour period.

Bacterial cytological profiling of MnG9MeBeim

To obtain a general assessment of the effect of **MnG9MeBeim** on bacteria, we performed bacterial cytological profiling (BCP) in *Bacillus subtilis*.^{48,49} Growth curve experiments with different concentrations of the compound were conducted to determine the optimal amount and the ideal incubation time, which was found to be 3 μM for 20 min. A *B. subtilis* strain with the *PrpsD-gfp* reporter was incubated with **MnG9MeBeim** for 15 min before the DNA stain DAPI and the membrane stain Nile red were added, and the bacteria were imaged with a fluorescence microscope. No significant differences were observed in the GFP or DAPI images, while the Nile red staining was clearly different, showing multiple hotspots on the bacterial membrane (Figure 7B). Indeed, slightly lower GFP signal is visible at these locations suggesting the presence of small invaginations. As **MnG9MeBeim** seemed to have a distinct effect on the bacterial membrane, we incubated a MinD-GFP reporter with the compound to observe any effects on membrane polarisation.⁴⁹ MinD is involved in cell division regulation and localises at the bacterial cell poles and septa. Membrane depolarisation perturbs the MinD localisation significantly.⁵⁰ With our compound similar 'blob-like' accumulations as with the Nile red were observed. However, the pattern was distinctly different from the positive control nisin (Figure 7B) where complete dispersal of MinD was observed, indicating that **MnG9MeBeim** does not affect membrane polarisation but forms invagination which might disturb MinD localisation. Finally, based on the results indicating that CO release is involved in the mode of action of the compound *B. subtilis* was incubated with both **MnG9MeBeim** and COP-1, a fluorescent probe to detect intracellular CO and stained the cells with Nile red (Figure 7C).⁵¹ From the imaging at the COP-1 emission wavelength it is clear that CO is being released, which triggers the observed fluorescence. Additionally, the blobs observed with Nile red coincide with the peaks in COP-1 fluorescence, indicating that CO is preferentially released in and around the membrane, further supporting the hypothesis that **MnG9MeBeim** seems to have a distinct effect on the latter.

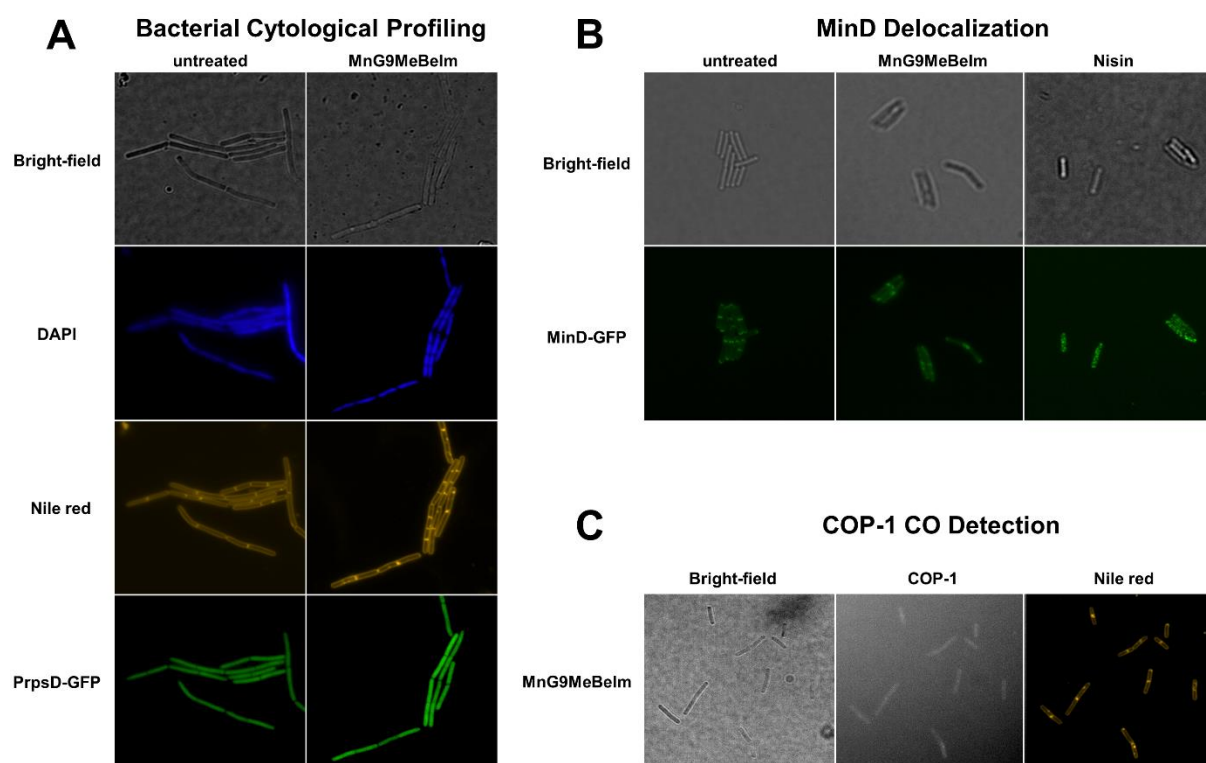


Figure 7. Fluorescence microscopy pictures obtained. **(A)** BCP: *B. subtilis* PrpsD strain incubated with 6.25 μM **MnG9MeBelm** for 20 min and stained with DAPI and Nile red compared with untreated control. **(B)** *B. subtilis* with MinD reporter incubated with 6.25 μM **MnG9MeBelm** for 20 min compared with untreated control. **(C)** *B. subtilis* incubated with 6.25 μM **MnG9MeBelm** for 20 min in the presence of COP-1 and stained with Nile red.

Effect of MnG9MeBelm on the bacterial membrane

Based on the microscopy results, an effect of **MnG9MeBelm** on the bacterial membrane seemed apparent. In previous work, some manganese tricarbonyl compounds were found to induce pore formation and other detrimental effects on the bacterial membrane.²³ To investigate the effect of **MnG9MeBelm** on the membrane, we utilized propidium iodide (PI), a fluorescent agent that is unable to traverse intact membranes but accumulates if the latter becomes compromised by pores. Comparison with sodium dodecyl sulfate (SDS), a surfactant known to form large pores in membranes, showed that **MnG9MeBelm** causes no significant uptake of PI into the bacteria (Figure 8A). A DiOC₂ assay was performed to investigate any effects on membrane polarisation. The absence of any significant fluorescence increase caused by displacement of DiOC₂ from bacterial membranes is in agreement with the distinct MinD distribution pattern described earlier (Figure 7B), suggesting **MnG9MeBelm** does not affect membrane polarisation. Together with the PI assay these experiments demonstrate that any effect the lead compound exerts on the bacterial membrane does not involve pore formation or depolarisation. In previous reports, the inhibition of the respiratory chain was implied as a target for a ruthenium-based CORM (of note it has been questioned whether the antibacterial effect of Ru-CORMs is indeed due to CO-release).^{52,53} To interrogate whether **MnG9MeBelm** affects the respiratory chain we quantified the respiratory activity by monitoring

resazurin reduction.⁵⁴ A dose-dependent decrease in respiratory chain activity was observed (Figure 8C), suggesting that the CO released by the compounds indeed inhibits the electron transport chain. The effect at the highest concentration tested was comparable to that of sodium azide, a well-known inhibitor of the electron transport chain (Figure 8C).⁵⁵ The aerobic respiratory chain is located on the plasma membrane of bacteria and hence the observation that **MnG9MeBelm** causes bacterial death by its inhibition coincides well with the fluorescent microscopy data discussed earlier.

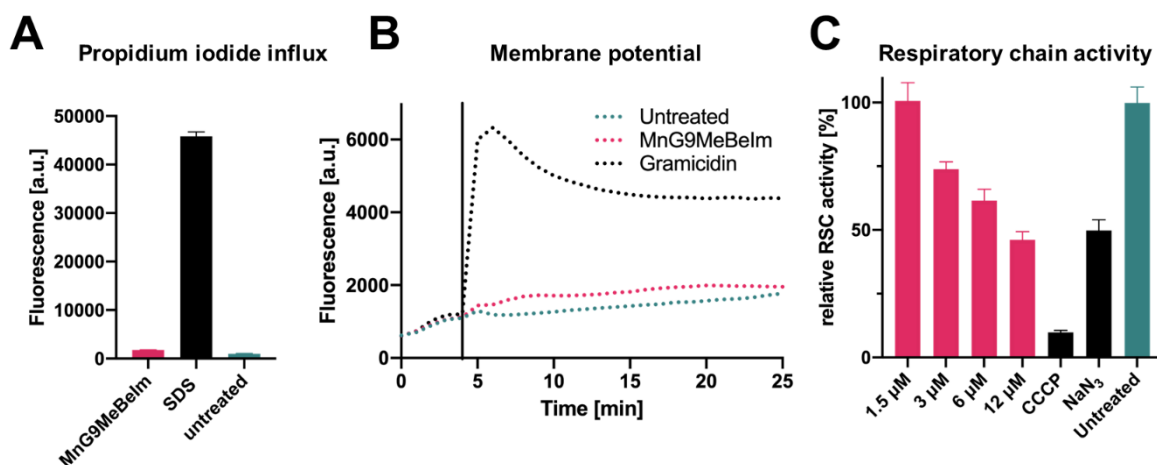


Figure 8. Mode of action characterisation for **MnG9MeBelm**. (A) Quantified PI fluorescence in bacteria after exposure to compounds (3 μ M, 20 min), SDS (positive control, 0.05%, 20 min) or no compound. (B) DiOC₂ fluorescence as a measure of membrane depolarisation after addition of **MnG9MeBelm** (3 μ M), Gramicidin (positive control 1 μ g/mL). The straight black line indicates the time point of antibiotic addition. (C) Relative respiratory chain activity measured by resazurin reduction. MnG8MeBelm was tested at 4 concentrations (after 20 min incubation) and compared to the positive controls CCCP (100 μ M, 20 min) and NaN₃ (15 mM, 20 min).

Conclusion

To accelerate the pace of discovery for novel metalloantibiotics we have applied combinatorial chemistry to the preparation of manganese tricarbonyl compounds for the first time. This enabled the preparation and antibacterial screening of 420 new Schiff-base Mn(CO)₃ complexes. A large fraction of the compounds (64/420, 15.3%) showed significant activity against MRSA (MIC \leq 12.5 μ M) and only inactive compounds displayed haemolysis against human red blood cells. Based on this crude screening data we selected 10 lead compounds, maximizing for activity and structural diversity. These compounds were resynthesized in batch, purified and fully characterised. Re-measuring of antibacterial activity and human cell toxicity revealed **MnG9MeBelm** as lead compound displaying a therapeutic index of over 100. X-ray diffraction unambiguously confirmed the structure of **MnG9MeBelm**. The CO-releasing properties of the complex were characterised, revealing a slow release of the CO-ligands. The presence of Hb reduces the antibacterial activity of the lead compound, implying a significant role of CO in its mode of bacterial killing. Mode of action studies utilising BCP indicate a distinct effect of **MnG9MeBelm** on the bacterial membrane. Common modes of membrane interference such as pore formation and membrane depolarisation could be excluded and we could show that CO is released

in vitro and colocalises with apparent membrane invaginations. Lastly, we show that the compound inhibits the aerobic cell respiratory chain which is located in the bacterial membrane. With this data, the lead compound is primed for initial *in vivo* evaluations for toxicity and efficacy.

Altogether we have efficiently screened a library of 420 new metal compounds for their antibacterial properties. The observed hit-rate is in line and even exceeds that of previous screening initiatives.^{4,56} The approach of directly screening crude reaction mixtures for antibacterial properties is vindicated by the excellent agreement of crude reaction results with the measurements conducted on the purified manganese complexes. Building on this approach and expanding it to other metal-scaffolds together with the automation of the reaction set-up will enable the preparation and screening of orders of magnitude larger metal complex libraries in the near future, enabling a more efficient exploration of the metal complex space for promising new metalloantibiotics.

Author contributions

A.F conceived the project and designed the library of building blocks. M. S. conducted all the synthesis, characterisation and biological assays. S. P. conducted the cytotoxicity studies. G. P. conducted the crystallographic analysis. A.F. and M.S. analyzed the data. A.F. composed the manuscript. All authors discussed, commented and approved the final manuscript.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We thank Prof. J-L. Reymond for generously hosting and supporting our research group. Prof. M. Wenzel and A. Schäfer are acknowledged for their introduction to BCP and providing the GFP-fused *B. subtilis* strains. Prof Cristina Bacci and Dr. Martina Rega are acknowledged for the gift of clinical isolate bacteria strains. We also thank Prof. C. von Ballmoos and N. Dolder for access and introduction to their fluorescence microscope. We thank Çağrı Özsan for helpful comments during the preparation of this manuscript. Acknowledgments to the University of Bern's crystallography support staff for their assistance, and to Chiesi Farmaceutici SpA and Dr. Davide Balestri for their support with the D8 Venture X-ray equipment. A.F. gratefully acknowledges funding from the Swiss National Science Foundation Ambizione grant PZ00P2_202016 and support by the University of Bern.

References

- (1) Murray, C. J.; Ikuta, K. S.; Sharara, F.; Swetschinski, L.; Aguilar, G. R.; Gray, A.; Han, C.; Bisignano, C.; Rao, P.; Wool, E.; Johnson, S. C.; Browne, A. J.; Chipeta, M. G.; Fell, F.; Hackett, S.; Haines-Woodhouse, G.; Hamadani, B. H. K.; Kumaran, E. A. P.; McManigal, B.; Agarwal, R.; Akech, S.; Albertson, S.; Amuasi, J.; Andrews, J.; Aravkin, A.; Ashley, E.; Bailey, F.; Baker, S.; Basnyat, B.; Bekker, A.; Bender, R.; Bethou, A.; Bielicki, J.; Boonkasidecha, S.; Bukosia, J.; Carvalheiro, C.; Castañeda-Orjuela, C.; Chansamouth, V.; Chaurasia, S.; Chiurchiù, S.; Chowdhury, F.; Cook, A. J.; Cooper, B.; Cressey, T. R.; Criollo-Mora, E.; Cunningham, M.; Darboe, S.; Day, N. P. J.; Luca, M. D.; Dokova, K.; Dramowski, A.; Dunachie, S. J.; Eckmanns, T.; Eibach, D.; Emami, A.; Feasey, N.; Fisher-Pearson, N.; Forrest, K.; Garrett, D.; Gastmeier, P.; Giref, A. Z.; Greer, R. C.; Gupta, V.; Haller, S.; Haselbeck, A.; Hay, S. I.; Holm, M.; Hopkins, S.; Iregbu, K. C.; Jacobs, J.; Jarovsky, D.; Javanmardi, F.; Khorana, M.; Kissoon, N.; Kobeissi, E.; Kostyanev, T.; Krapp, F.; Krumkamp, R.; Kumar, A.; Kyu, H. H.; Lim, C.; Limmathurotsakul, D.; Loftus, M. J.; Lunn, M.; Ma, J.; Mturi, N.; Munera-Huertas, T.; Musicha, P.; Mussi-Pinhata, M. M.; Nakamura, T.; Nanavati, R.; Nangia, S.; Newton, P.; Ngoun, C.; Novotney, A.; Nwakanma, D.; Obiero, C. W.; Olivas-Martinez, A.; Olliaro, P.; Ooko, E.; Ortiz-Brizuela, E.; Peleg, A. Y.; Perrone, C.; Plakkal, N.; Ponce-de-Leon, A.; Raad, M.; Ramdin, T.; Riddell, A.; Roberts, T.; Robotham, J. V.; Roca, A.; Rudd, K. E.; Russell, N.; Schnall, J.; Scott, J. A. G.; Shivamallappa, M.; Sifuentes-Osornio, J.; Steenkeste, N.; Stewardson, A. J.; Stoeva, T.; Tasak, N.; Thaiprakong, A.; Thwaites, G.; Turner, C.; Turner, P.; Doorn, H. R. van; Velaphi, S.; Vongpradith, A.; Vu, H.; Walsh, T.; Waner, S.; Wangrangsimakul, T.; Wozniak, T.; Zheng, P.; Sartorius, B.; Lopez, A. D.; Stergachis, A.; Moore, C.; Dolecek, C.; Naghavi, M. Global Burden of Bacterial Antimicrobial Resistance in 2019: A Systematic Analysis. *The Lancet* **2022**, 399 (10325), 629–655. [https://doi.org/10.1016/S0140-6736\(21\)02724-0](https://doi.org/10.1016/S0140-6736(21)02724-0).
- (2) Butler, M. S.; Henderson, I. R.; Capon, R. J.; Blaskovich, M. A. T. Antibiotics in the Clinical Pipeline as of December 2022. *J. Antibiot.* **2023**, 76 (8), 431–473. <https://doi.org/10.1038/s41429-023-00629-8>.
- (3) Bhandari, V.; Suresh, A. Next-Generation Approaches Needed to Tackle Antimicrobial Resistance for the Development of Novel Therapies Against the Deadly Pathogens. *Front. Pharmacol.* **2022**, 13.
- (4) Frei, A.; Zuegg, J.; Elliott, A. G.; Baker, M.; Braese, S.; Brown, C.; Chen, F.; G. Dowson, C.; Dujardin, G.; Jung, N.; King, A. P.; Mansour, A. M.; Massi, M.; Moat, J.; Mohamed, H. A.; Renfrew, A. K.; Rutledge, P. J.; Sadler, P. J.; Todd, M. H.; Willans, C. E.; Wilson, J. J.; Cooper, M. A.; Blaskovich, M. A. T. Metal Complexes as a Promising Source for New Antibiotics. *Chem. Sci.*, **2020**, 11, 2627–2639. <https://doi.org/10.1039/C9SC06460E>.
- (5) Smitten, K. L.; Southam, H. M.; de la Serna, J. B.; Gill, M. R.; Jarman, P. J.; Smythe, C. G. W.; Poole, R. K.; Thomas, J. A. Using Nanoscopy To Probe the Biological Activity of Antimicrobial Leads That Display Potent Activity against Pathogenic, Multidrug Resistant, Gram-Negative Bacteria. *ACS Nano* **2019**, 13 (5), 5133–5146. <https://doi.org/10.1021/acsnano.8b08440>.
- (6) Smitten, K. L.; Fairbanks, S. D.; Robertson, C. C.; Serna, J. B. de la; Foster, S. J.; Thomas, J. A. Ruthenium Based Antimicrobial Theranostics – Using Nanoscopy to Identify Therapeutic Targets and Resistance Mechanisms in *Staphylococcus Aureus*. *Chem. Sci.* **2019**, 11 (1), 70–79. <https://doi.org/10.1039/C9SC04710G>.
- (7) Smitten, K.; Southam, H. M.; Fairbanks, S.; Graf, A.; Chauvet, A.; Thomas, J. A. Clearing an ESKAPE Pathogen in a Model Organism; a Polypyridyl Ruthenium(II) Complex Theranostic That Treats a Resistant *Acinetobacter Baumannii* Infection in *Galleria Mellonella*. *Chem. – Eur. J.* <https://doi.org/10.1002/chem.202203555>.
- (8) Li, F.; Collins, J. G.; Keene, F. R. Ruthenium Complexes as Antimicrobial Agents. *Chem. Soc. Rev.* **2015**, 44 (8), 2529–2542. <https://doi.org/10.1039/C4CS00343H>.
- (9) Frei, A. Metal Complexes, an Untapped Source of Antibiotic Potential? *Antibiotics*, **2020**, 9, 90. <https://doi.org/10.3390/antibiotics9020090>.
- (10) Li, F.; Harry, E. J.; Bottomley, A. L.; Edstein, M. D.; Birrell, G. W.; Woodward, C. E.; Keene, F. R.; Collins, J. G. Dinuclear Ruthenium(II) Antimicrobial Agents That Selectively Target Polysomes in Vivo. *Chem. Sci.* **2013**, 5 (2), 685–693. <https://doi.org/10.1039/C3SC52166D>.
- (11) Li, F.; Feterl, M.; Mulyana, Y.; Warner, J. M.; Collins, J. G.; Keene, F. R. In Vitro Susceptibility and Cellular Uptake for a New Class of Antimicrobial Agents: Dinuclear Ruthenium(II) Complexes. *J. Antimicrob. Chemother.* **2012**, 67 (11), 2686–2695. <https://doi.org/10.1093/jac/dks291>.
- (12) Weber, D. K.; Sani, M.-A.; Downton, M. T.; Separovic, F.; Keene, F. R.; Collins, J. G. Membrane Insertion of a Dinuclear Polypyridylruthenium(II) Complex Revealed by Solid-State NMR and Molecular Dynamics Simulation: Implications for Selective Antibacterial Activity. *J. Am. Chem. Soc.* **2016**, 138 (46), 15267–15277. <https://doi.org/10.1021/jacs.6b09996>.
- (13) Patra, M.; Wenzel, M.; Prochnow, P.; Pierroz, V.; Gasser, G.; Bandow, J. E.; Metzler-Nolte, N. An Organometallic Structure-Activity Relationship Study Reveals the Essential Role of a Re(CO)₃ Moiety in the Activity against Gram-Positive Pathogens Including MRSA. *Chem. Sci.* **2014**, 6 (1), 214–224. <https://doi.org/10.1039/C4SC02709D>.
- (14) Frei, A.; Amado, M.; Cooper, M. A.; Blaskovich, M. A. T. Light-Activated Rhenium Complexes with Dual Mode of Action against Bacteria. *Chem. – Eur. J.*, **2019**, DOI: 10.1002/chem.201904689. <https://doi.org/10.1002/chem.201904689>.

- (15) Sovari, S. N.; Radakovic, N.; Roch, P.; Crochet, A.; Pavic, A.; Zobi, F. Combatting AMR: A Molecular Approach to the Discovery of Potent and Non-Toxic Rhenium Complexes Active against *C. Albicans*-MRSA Co-Infection. *Eur. J. Med. Chem.* **2021**, *226*, 113858. <https://doi.org/10.1016/j.ejmech.2021.113858>.
- (16) Sovari, S. N.; Vojnovic, S.; Bogojevic, S. S.; Crochet, A.; Pavic, A.; Nikodinovic-Runic, J.; Zobi, F. Design, Synthesis and in Vivo Evaluation of 3-Arylcoumarin Derivatives of Rhenium(I) Tricarbonyl Complexes as Potent Antibacterial Agents against Methicillin-Resistant *Staphylococcus Aureus* (MRSA). *Eur. J. Med. Chem.* **2020**, *205*, 112533. <https://doi.org/10.1016/j.ejmech.2020.112533>.
- (17) Mendes, S. S.; Marques, J.; Mesterházy, E.; Straetener, J.; Arts, M.; Pissarro, T.; Reginold, J.; Berscheid, A.; Bornikoel, J.; Kluj, R. M.; Mayer, C.; Oesterhelt, F.; Friães, S.; Royo, B.; Schneider, T.; Brötz-Oesterhelt, H.; Romão, C. C.; Saraiva, L. M. Synergetic Antimicrobial Activity and Mechanism of Clotrimazole-Linked CO-Releasing Molecules. *ACS Bio Med Chem Au* **2022**. <https://doi.org/10.1021/acsbiochemau.2c00007>.
- (18) Nagel, C.; McLean, S.; Poole, R. K.; Braunschweig, H.; Kramer, T.; Schatzschneider, U. Introducing [Mn(CO)₃(Tpa-K3N)]⁺ as a Novel Photoactivatable CO-Releasing Molecule with Well-Defined ICORM Intermediates – Synthesis, Spectroscopy, and Antibacterial Activity. *Dalton Trans.* **2014**, *43* (26), 9986–9997. <https://doi.org/10.1039/C3DT51848E>.
- (19) Betts, J.; Nagel, C.; Schatzschneider, U.; Poole, R.; Ragione, R. M. L. Antimicrobial Activity of Carbon Monoxide-Releasing Molecule [Mn(CO)₃(Tpa-K3N)]Br versus Multidrug-Resistant Isolates of Avian Pathogenic *Escherichia Coli* and Its Synergy with Colistin. *PLOS ONE* **2017**, *12* (10), e0186359. <https://doi.org/10.1371/journal.pone.0186359>.
- (20) Rana, N.; Jesse, H. E.; Tinajero-Trejo, M.; Butler, J. A.; Tarlit, J. D.; von und zur Muhlen, M. L.; Nagel, C.; Schatzschneider, U.; Poole, R. K. A Manganese Photosensitive Tricarbonyl Molecule [Mn(CO)₃(Tpa-K3N)]Br Enhances Antibiotic Efficacy in a Multi-Drug-Resistant *Escherichia Coli*. *Microbiology* **2017**, *163* (10), 1477–1489. <https://doi.org/10.1099/mic.0.000526>.
- (21) Tinajero-Trejo, M.; Rana, N.; Nagel, C.; Jesse, H. E.; Smith, T. W.; Wareham, L. K.; Hippler, M.; Schatzschneider, U.; Poole, R. K. Antimicrobial Activity of the Manganese Photoactivated Carbon Monoxide-Releasing Molecule [Mn(CO)₃(Tpa-K3N)]⁺ Against a Pathogenic *Escherichia Coli* That Causes Urinary Infections. *Antioxid. Redox Signal.* **2016**, *24* (14), 765–780. <https://doi.org/10.1089/ars.2015.6484>.
- (22) Simpson, P. V.; Nagel, C.; Bruhn, H.; Schatzschneider, U. Antibacterial and Antiparasitic Activity of Manganese(I) Tricarbonyl Complexes with Ketoconazole, Miconazole, and Clotrimazole Ligands. *Organometallics* **2015**, *34* (15), 3809–3815. <https://doi.org/10.1021/acs.organomet.5b00458>.
- (23) Güntzel, P.; Nagel, C.; Weigelt, J.; Betts, J. W.; Patrick, C. A.; Southam, H. M.; La Ragione, R. M.; Poole, R. K.; Schatzschneider, U. Biological Activity of Manganese(I) Tricarbonyl Complexes on Multidrug-Resistant Gram-Negative Bacteria: From Functional Studies to in Vivo Activity in *Galleria Mellonella*†. *Metallomics* **2019**, *11* (12), 2033–2042. <https://doi.org/10.1039/c9mt00224c>.
- (24) Betts, J. W.; Roth, P.; Patrick, C. A.; Southam, H. M.; La Ragione, R. M.; Poole, R. K.; Schatzschneider, U. Antibacterial Activity of Mn(i) and Re(i) Tricarbonyl Complexes Conjugated to a Bile Acid Carrier Molecule†. *Metallomics* **2020**, *12* (10), 1563–1575. <https://doi.org/10.1039/d0mt00142b>.
- (25) Betts, J. W.; Cawthraw, S.; Smyth, J. A.; Poole, R. K.; Roth, P.; Schatzschneider, U.; La Ragione, R. M. The Manganese Carbonyl Complex [Mn(CO)₃(Tpa-K3N)]Br: A Novel Antimicrobial Agent with the Potential to Treat Avian Pathogenic *Escherichia Coli* (APEC) Infections. *Vet. Microbiol.* **2023**, *284*, 109819. <https://doi.org/10.1016/j.vetmic.2023.109819>.
- (26) Ward, J. S.; Lynam, J. M.; Moir, J.; Fairlamb, I. J. S. Visible-Light-Induced CO Release from a Therapeutically Viable Tryptophan-Derived Manganese(I) Carbonyl (TryptoCORM) Exhibiting Potent Inhibition against *E. Coli*. *Chem. – Eur. J.* **2014**, *20* (46), 15061–15068. <https://doi.org/10.1002/chem.201403305>.
- (27) Ward, J. S.; Morgan, R.; Lynam, J. M.; Fairlamb, I. J. S.; Moir, J. W. B. Toxicity of Tryptophan Manganese(I) Carbonyl (Trypto-CORM), against *Neisseria Gonorrhoeae*. *MedChemComm* **2017**, *8* (2), 346–352. <https://doi.org/10.1039/C6MD00603E>.
- (28) Crook, S. H.; Mann, B. E.; Meijer, A. J. H. M.; Adams, H.; Sawle, P.; Scapens, D.; Motterlini, R. [Mn(CO)₄(S₂CNMe(CH₂CO₂H))], a New Water-Soluble CO-Releasing Molecule. *Dalton Trans.* **2011**, *40* (16), 4230–4235. <https://doi.org/10.1039/C1DT10125K>.
- (29) Wareham, L. K.; McLean, S.; Begg, R.; Rana, N.; Ali, S.; Kendall, J. J.; Sanguinetti, G.; Mann, B. E.; Poole, R. K. The Broad-Spectrum Antimicrobial Potential of [Mn(CO)₄(S₂CNMe(CH₂CO₂H))], a Water-Soluble CO-Releasing Molecule (CORM-401): Intracellular Accumulation, Transcriptomic and Statistical Analyses, and Membrane Polarization. *Antioxid. Redox Signal.* **2018**, *28* (14), 1286–1308. <https://doi.org/10.1089/ars.2017.7239>.
- (30) Liu, R.; Li, X.; Lam, K. S. Combinatorial Chemistry in Drug Discovery. *Curr. Opin. Chem. Biol.* **2017**, *38*, 117–126. <https://doi.org/10.1016/j.cbpa.2017.03.017>.
- (31) Favalli, N.; Bassi, G.; Scheuermann, J.; Neri, D. DNA-Encoded Chemical Libraries – Achievements and Remaining Challenges. *FEBS Lett.* **2018**, *592* (12), 2168–2180. <https://doi.org/10.1002/1873-3468.13068>.
- (32) Lowry, M. S.; Hudson, W. R.; Pascal, R. A.; Bernhard, S. Accelerated Luminophore Discovery through Combinatorial Synthesis. *J. Am. Chem. Soc.* **2004**, *126* (43), 14129–14135. <https://doi.org/10.1021/ja047156+>.
- (33) Mdluli, V.; Diluzio, S.; Lewis, J.; Kowalewski, J. F.; Connell, T. U.; Yaron, D.; Kowalewski, T.; Bernhard, S. High-Throughput Synthesis and Screening of Iridium(III) Photocatalysts for the Fast and Chemoselective Dehalogenation of Aryl Bromides. *ACS Catal.* **2020**, *10* (13), 6977–6987. <https://doi.org/10.1021/acscatal.0c02247>.

- (34) DiLuzio, S.; Mdluli, V.; Connell, T. U.; Lewis, J.; VanBenschoten, V.; Bernhard, S. High-Throughput Screening and Automated Data-Driven Analysis of the Triplet Photophysical Properties of Structurally Diverse, Heteroleptic Iridium(III) Complexes. *J. Am. Chem. Soc.* **2021**, *143* (2), 1179–1194. <https://doi.org/10.1021/jacs.0c12290>.
- (35) Zach, T.; Geyer, F.; Kiendl, B.; Mößler, J.; Nguyen, O.; Schmidpeter, T.; Schuster, P.; Nagel, C.; Schatzschneider, U. Electrospray Mass Spectrometry to Study Combinatorial iClick Reactions and Multiplexed Kinetics of [Ru(N₃)(N₄N)(Terpy)]PF₆ with Alkynes of Different Steric and Electronic Demand. *Inorg. Chem.* **2023**, *62* (7), 2982–2993. <https://doi.org/10.1021/acs.inorgchem.2c03377>.
- (36) Chow, M. J.; Licon, C.; Yuan Qiang Wong, D.; Pastorin, G.; Gaiddon, C.; Ang, W. H. Discovery and Investigation of Anticancer Ruthenium–Arene Schiff-Base Complexes via Water-Promoted Combinatorial Three-Component Assembly. *J. Med. Chem.* **2014**, *57* (14), 6043–6059. <https://doi.org/10.1021/jm500455p>.
- (37) Juinn Chow, M.; Alfiean, M.; Pastorin, G.; Gaiddon, C.; Han Ang, W. Apoptosis-Independent Organoruthenium Anticancer Complexes That Overcome Multidrug Resistance: Self-Assembly and Phenotypic Screening Strategies. *Chem. Sci.* **2017**, *8* (5), 3641–3649. <https://doi.org/10.1039/C7SC00497D>.
- (38) Weng, C.; Shen, L.; Ang, W. H. Harnessing Endogenous Formate for Antibacterial Prodrug Activation by in Cellulo Ruthenium-Mediated Transfer Hydrogenation Reaction. *Angew. Chem. Int. Ed.* **2020**, *59* (24), 9314–9318. <https://doi.org/10.1002/anie.202000173>.
- (39) Weng, C.; Shen, L.; Teo, J. W.; Lim, Z. C.; Loh, B. S.; Ang, W. H. Targeted Antibacterial Strategy Based on Reactive Oxygen Species Generated from Dioxygen Reduction Using an Organoruthenium Complex. *JACS Au* **2021**, *1* (9), 1348–1354. <https://doi.org/10.1021/jacsau.1c00262>.
- (40) Weng, C.; Yang, H.; Loh, B. S.; Wong, M. W.; Ang, W. H. Targeting Pathogenic Formate-Dependent Bacteria with a Bioinspired Metallo–Nitroreductase Complex. *J. Am. Chem. Soc.* **2023**, *145* (11), 6453–6461. <https://doi.org/10.1021/jacs.3c00237>.
- (41) Konkankit, C. C.; Vaughn, B. A.; MacMillan, S. N.; Boros, E.; Wilson, J. J. Combinatorial Synthesis to Identify a Potent, Necrosis-Inducing Rhenium Anticancer Agent. *Inorg. Chem.* **2019**, *58* (6), 3895–3909. <https://doi.org/10.1021/acs.inorgchem.8b03552>.
- (42) Konkankit, C. C.; Vaughn, B. A.; Huang, Z.; Boros, E.; Wilson, J. J. Systematically Altering the Lipophilicity of Rhenium(I) Tricarbonyl Anticancer Agents to Tune the Rate at Which They Induce Cell Death. *Dalton Trans.* **2020**, *49* (45), 16062–16066. <https://doi.org/10.1039/D0DT01097A>.
- (43) Vilar, R.; Kench, T.; Rahardjo, A.; Bellamkonda, A.; Maher, T. E.; Storch, M. A *Semi-Automated, High-Throughput Approach for the Synthesis and Identification of Highly Photo-Cytotoxic Iridium Complexes*; Chemrxiv, **2023**. <https://doi.org/10.26434/chemrxiv-2023-rt8kr>.
- (44) Bajusz, D.; Rácz, A.; Héberger, K. Why Is Tanimoto Index an Appropriate Choice for Fingerprint-Based Similarity Calculations? *J. Cheminformatics* **2015**, *7* (1), 20. <https://doi.org/10.1186/s13321-015-0069-3>.
- (45) Cortat, Y.; Nedyalkova, M.; Schindler, K.; Kadakia, P.; Demirci, G.; Nasiri Sovari, S.; Crochet, A.; Salentinig, S.; Lattuada, M.; Steiner, O. M.; Zobi, F. Computer-Aided Drug Design and Synthesis of Rhenium Clotrimazole Antimicrobial Agents. *Antibiotics* **2023**, *12* (3), 619. <https://doi.org/10.3390/antibiotics12030619>.
- (46) Orpen, A. G.; Brammer, L.; Allen, F. H.; Kennard, O.; Watson, D. G.; Taylor, R. Supplement. Tables of Bond Lengths Determined by X-Ray and Neutron Diffraction. Part 2. Organometallic Compounds and Co-Ordination Complexes of the d- and f-Block Metals. *J. Chem. Soc. Dalton Trans.* **1989**, No. 12, S1–S83. <https://doi.org/10.1039/DT989000000S1>.
- (47) Huang, R.; Pei, L.; Liu, Q.; Chen, S.; Dou, H.; Shu, G.; Yuan, Z.; Lin, J.; Peng, G.; Zhang, W.; Fu, H. Isobologram Analysis: A Comprehensive Review of Methodology and Current Research. *Front. Pharmacol.* **2019**, *10*.
- (48) Nonejuie, P.; Burkart, M.; Pogliano, K.; Pogliano, J. Bacterial Cytological Profiling Rapidly Identifies the Cellular Pathways Targeted by Antibacterial Molecules. *Proc. Natl. Acad. Sci.* **2013**, *110* (40), 16169–16174. <https://doi.org/10.1073/pnas.1311066110>.
- (49) Schäfer, A.-B.; Wenzel, M. A How-To Guide for Mode of Action Analysis of Antimicrobial Peptides. *Front. Cell. Infect. Microbiol.* **2020**, *10*.
- (50) Strahl, H.; Hamoen, L. W. Membrane Potential Is Important for Bacterial Cell Division. *Proc. Natl. Acad. Sci.* **2010**, *107* (27), 12281–12286. <https://doi.org/10.1073/pnas.1005485107>.
- (51) Michel, B. W.; Lippert, A. R.; Chang, C. J. A Reaction-Based Fluorescent Probe for Selective Imaging of Carbon Monoxide in Living Cells Using a Palladium-Mediated Carbonylation. *J. Am. Chem. Soc.* **2012**, *134* (38), 15668–15671. <https://doi.org/10.1021/ja307017b>.
- (52) Mansour, A. M.; Khaled, R. M.; Khaled, E.; Ahmed, S. K.; Ismael, O. S.; Zeinhom, A.; Magdy, H.; Ibrahim, S. S.; Abdelfatah, M. Ruthenium(II) Carbon Monoxide Releasing Molecules: Structural Perspective, Antimicrobial and Anti-Inflammatory Properties. *Biochem. Pharmacol.* **2022**, *199*, 114991. <https://doi.org/10.1016/j.bcp.2022.114991>.
- (53) Nielsen, V. G. Ruthenium, Not Carbon Monoxide, Inhibits the Procoagulant Activity of Atheris, Echis, and Pseudonaja Venoms. *Int. J. Mol. Sci.* **2020**, *21* (8), 2970. <https://doi.org/10.3390/ijms21082970>.
- (54) Saeloh, D.; Tipmanee, V.; Jim, K. K.; Dekker, M. P.; Bitter, W.; Voravuthikunchai, S. P.; Wenzel, M.; Hamoen, L. W. The Novel Antibiotic Rhodomycytone Traps Membrane Proteins in Vesicles with Increased Fluidity. *PLoS Pathog.* **2018**, *14* (2), e1006876. <https://doi.org/10.1371/journal.ppat.1006876>.
- (55) Tsubaki, M.; Yoshikawa, S. Fourier-Transform Infrared Study of Azide Binding to the Fea3-CuB Binuclear Site of Bovine Heart Cytochrome c Oxidase: New Evidence for a Redox-Linked Conformational Change at the Binuclear Site. *Biochemistry* **1993**, *32* (1), 174–182. <https://doi.org/10.1021/bi00052a023>.

- (56) Frei, A.; Elliott, A. G.; Kan, A.; Dinh, H.; Bräse, S.; Bruce, A. E.; Bruce, M. R.; Chen, F.; Humaidy, D.; Jung, N.; King, A. P.; Lye, P. G.; Maliszewska, H. K.; Mansour, A. M.; Matiadis, D.; Muñoz, M. P.; Pai, T.-Y.; Pokhrel, S.; Sadler, P. J.; Sagnou, M.; Taylor, M.; Wilson, J. J.; Woods, D.; Zuegg, J.; Meyer, W.; Cain, A. K.; Cooper, M. A.; Blaskovich, M. A. T. Metal Complexes as Antifungals? From a Crowd-Sourced Compound Library to the First In Vivo Experiments. *JACS Au* **2022**, *2* (10), 2277–2294. <https://doi.org/10.1021/jacsau.2c00308>.