Molecular Gas Phase Conformational Ensembles

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Abstract. Accurately determining the global minima of a molecular structure is important in diverse scientific fields, including drug design, materials science, and chemical synthesis. Conformational search engines serve as valuable tools for exploring the extensive conformational space of molecules and identifying energetically favorable conformations. In this study, we present a comprehensive comparison of Auto3D, CREST, Balloon, and RDKit, which are freely available conformational search engines, to evaluate their effectiveness in locating the global minima. These engines employ distinct methodologies, including machine learning (ML) potential-based, semiempirical, and force field (FF) based approaches. Through rigorous analyses and employing novel approaches for validation, including the utilization of a unique physical property known as collisional cross section (CCS), which characterizes the molecular shape, size, and charge, we thoroughly assess the capabilities of these engines in generating conformation ensembles that effectively capture the global minima. To accomplish this, we created the gas-phase conformation library (GPCL) which currently consists of the full ensembles of 20 small molecules, which can be used by the community to validate any conformational search engine. Further members of the GPCL can be readily created for any molecule of interest using our standard workflow used to compute CCS values expanding the ability of the GPCL in validation exercises. These innovative validation techniques enhance our understanding of the conformational landscape and provide valuable insights into the performance of conformation generation engines. Our findings shed light on the strengths and limitations of each search engine, enabling informed decisions for their utilization in various scientific fields, where accurate molecular structure determination is crucial for understanding biological activity and designing targeted interventions. By facilitating the identification of reliable conformations, this study significantly contributes to enhancing the efficiency and accuracy of molecular structure determination, with a particular focus on metabolite

structure elucidation. The findings of this research also provide valuable insights for developing effective workflows in predicting the structures of unknown compounds with high precision.

Keywords: Conformational search engines, Global minima, Conformational sampling, Molecular structure determination, Collisional Cross Section.

Introduction:

Accurately determining molecular ensembles is crucial in computational chemistry for understanding molecular behavior and properties. However, predicting the global minima, which represents the most stable conformation, presents a significant challenge due to the size of conformational spaces for flexible molecules. Moreover, to correctly rank order all low energy conformations poses another significant technical challenge. To overcome these challenges, efficient and reliable conformational search algorithms are necessary to explore this space. Conformer generation plays a pivotal role in various computational analyses, including computational drug design[1,2], 3D QSAR modeling[3,4,] protein-ligand docking[5–8], and structure elucidation of unknown compounds[9–11]. Different methods exist for generating conformers, ranging from obtaining a single low-energy conformation to generating ensembles that encompass biologically relevant low-energy conformational space. The choice of conformational sampling technique directly influences the subsequent analysis's reliability and speed.

Multiple conformational search engines are available including, for example, Balloon[12,13], RDKit[14–16], Confab[17], Frog2[7,18], MacroModel[19,20], OMEGA[21,22], CREST[23–25], and Auto3D[26]. These tools offer diverse methods and algorithms for conformation generation, ranging from force field-based approaches to semiempirical and machine learning potential-based methods. Force field-based methods are utilized by Balloon, RDKit, Confab, Frog2, and MacroModel to generate conformation ensembles. They combine systematic and random sampling techniques within the framework of a force field to explore conformational space. Balloon combines systematic and random sampling techniques to explore the conformational space of molecules. By combining systematic and random sampling techniques, it covers a wide range of molecular conformations, including both low-energy conformations and higher-energy regions. This comprehensive exploration of conformational space gives a more complete picture of the conformational landscape. RDKit, an open-source cheminformatics toolkit, employs a distance geometry algorithm along with distance constraints derived from a force field.[16,27,28] Confab, provided by the Molecular Operating Environment (MOE) software package, integrates systematic and random sampling methods within a force field

framework.[29–35] Frog2, developed by Certara, utilizes a proprietary algorithm based on force field methods to sample low-energy conformations of drug-like molecules.[7,18,36–43] MacroModel, offered by Schrödinger, employs molecular mechanics force fields such as OPLS-AA to explore conformational space.[44–53] Omega, developed by OpenEye Scientific Software, is a conformation generation tool that combines distance geometry, systematic search, and random perturbation methods to generate diverse conformations.[54–64] CREST (Conformer-Rotamer Ensemble Sampling Tool) utilizes an extended semiempirical tight-binding model, GFN2-xTB, a broadly parametrized self-consistent tight-binding (TB) quantum chemical method with multipole electrostatics and density-dependent dispersion contributions to calculate energy profiles and explore conformational space.[65–71] Auto3D employs machine learning potential-based methods, utilizing artificial neural networks trained on a large dataset of molecular structures to predict energetically favorable conformations.[72–80]

In addition to these engines, there are several other notable conformation generation tools available. The BioChemical Library, BCL::Conf is a conformational sampling tool developed by Meiler *et. al* that utilizes a combination of systematic search, stochastic optimization, and diversity analysis methods.[81] The Experimental-Torsion Distance Geometry with basic Knowledge (ETKDG) is a stochastic search method that uses distance geometry and knowledge from experimental crystal structures to explore the conformational space.[82] Conformator is a conformation search engine provided by the NAOMI ChemBio Suite that generates conformer ensembles using an incremental construction approach.[83] The CSD conformer generator is a tool specifically designed for generating conformations of small organic molecules using information from the Cambridge Structural Database (CSD).[84] ConfGen, developed by Schrödinger, is a conformation generation tool that combines systematic search and molecular dynamics simulations to explore conformational space.[62] CORINA is a conformational search tool that utilizes a combination of stochastic search algorithms, distance geometry, and energy minimization to generate low-energy conformations.[85] MOE (Molecular Operating Environment), a software package from Chemical Computing Group, provides a suite of conformational search algorithms and methods for generating conformational ensembles.[86] Other conformation generation engines include iCon, which employs an incremental construction approach to systematically explore the conformational space[73], and CAESAR, a tool that combines genetic algorithms with energy minimization to generate low-energy conformations.[55]

Apart from the above mentioned conformation generation software, there exists a diverse range of algorithms specifically designed for generating conformer ensembles. These algorithms facilitate comprehensive conformational

sampling in both gas and solution phases, allowing for a more thorough exploration of molecular flexibility. These include Confort[87], ROTATE[88], CONFECT[89], Catalyst[90,91], MED-3DMC[92], Multiconf-DOCK[93], CONFECT[94], BRIKARD[95], ForceGen[96], TCG (TrixX Conformer Generator)[97] and Cxcalc (ChemAxon)[98]. These tools utilize a range of algorithms and methodologies to explore the conformational space of molecules and generate conformational ensembles. ROTATE employs a systematic search algorithm based on molecular flexibility, while Catalyst utilizes a stochastic search algorithm with a focus on energy optimization. Confort incorporates a distance geometry approach to generate low-energy conformations, while MED-3DMC utilizes a Monte Carlo-based method. Multiconf-DOCK utilizes a systematic search approach for exploring ligand flexibility within the DOCK5 program. It extends multiple anchor segments stepwise and generates conformations by systematically rotating single, nonterminal, acyclic bonds at specified increments, while CONFECT employs an evolutionary algorithm. BRIKARD utilizes a knowledge-based approach, and ForceGen incorporates force field-based methods. TCG utilizes a systematic torsion angle search algorithm. Cxcalc utilizes a fragment fusion method and the Dreiding force field for the calculation and optimization of conformers. These tools aid in the exploration of potential binding modes and interactions.[98,99] Finally, it's important to note that all of these methods generate conformations in the gas-phase and not solution or the crystalline phase.

The primary objective of these conformation generation tools is to identify the global minima or a list of low-energy conformers from a large ensemble of generated conformations. The accuracy, speed, and computational reliability of these tools are achieved through different algorithmic approaches.[100,101] However, it is crucial to validate the results obtained from these tools with experimental findings. The validation of ligand conformations often involves comparing the generated conformers with experimentally determined structures, typically obtained from protein-bound ligand conformations extracted from the Protein Data Bank (PDB).[102–111] A shortcoming of these so-called "bioactive conformers" for the validation of conformation generation software is the limited number an diversity of experimentally determined protein-ligand structures and questions surrounding whether these "bioactive" conformers represent the global minimum or local minimum or high energy structures in the conformational ensemble.[85,112–127] Additionally, X-ray structures in the PDB represent static snapshots of molecules in crystalline states, which may not fully capture their dynamic behavior in solution or other environments. The resolution of X-ray structures is used as a quality criterion and low-resolution structures may lack precision and atomic-level details necessary for accurate conformation determination. It is crucial to consider these limitations and explore alternative validation approaches,

such as benchmark datasets or comparison with other experimental data.[102–104,128–131] In addition to the Protein Data Bank (PDB), another widely used validation dataset for ligand conformations is the Cambridge Structural Database (CSD).[132] The CSD primarily consists of small organic molecule crystal structures obtained from X-ray crystallography experiments. It offers a large collection of experimentally determined structures, providing valuable insights into the three-dimensional arrangements and intermolecular interactions of small molecules in the solid state. The use of the CSD as a validation dataset complements the information obtained from the PDB, expanding the scope of ligand conformation validation and contributing to a more comprehensive understanding of ligand behavior in different environments.

In this work, we propose a novel approach to evaluate and compare gas-phase conformational search engines based on their ability to characterize the gas-phase conformational ensemble and identify the global minima using a quantum mechanics (QM) based workflow whose outcome are compared against experimental information.[9,10] Specifically, we have developed a QM-based method to calculate Collisional Cross Sections (CCS), which is an accurate indicator of the global minima for molecular structures in the gas-phase[10]. Our CCS calculations have been validated against experimental data, demonstrating their reliability in capturing the most stable conformations.[133–138] To conduct our comparative analysis, we employed four different freely available conformational search engines: Auto3D, CREST, Balloon, and RDKit. These engines utilize diverse methodologies, including force field-based conformation generation (RDKit, Balloon), semi-empirical methods (CREST), and machine learning potentials (Auto3D). By generating conformations using each engine and comparing them with the ensemble and global minima validated through CCS calculations, we aimed to identify the most effective conformational search engine for accurate global minima prediction. By evaluating and comparing the performance of different engines in the gas phase, our study aims to provide valuable insights into the selection of the optimal conformational search approach for improved molecular structure determination and related applications. Moreover, the resultant data set can be used to validate other gas-phase conformational search engines.

Computational Methods:

In this study, we focused on 20 metabolites and employed a DFT based workflow to compute their Collisional Cross Section values. Our workflow has demonstrated good accuracy in CCS prediction, with an error rate of less than 3% compared to experiment (experimental error is ~3%). Our established workflow encompasses the following steps to predict accurate CCS values: First, the conformations of each metabolite were generated using the

RDKit tool.[14,15] A maximum number of generated conformers is set to 1000 for the small molecules systems. Each generated conformer was then geometry optimized using the ANI QM-ML model.[75,77,139] The optimized structures were subsequently clustered using our in-house automated clustering code called AutoGraph, enabling the identification of chemically unique conformations.[140–142] Geometry optimization and Mulliken atomic charge calculations were performed on representative conformations of each identified cluster using B3LYP/6-31+G(d,p) and B3LYP/6-311++G(d,p) level of theory, respectively employing a GPU enabled, in-house developed QM engine called QUICK.[143–145] The CCS values were computed using the trajectory method (TM) as implemented in the HPCCS code developed by Zanotto et al.[146,147] The inclusion of an unsupervised clustering method in our workflow reduces the potential for human bias and error in cluster selection, while the QM-ML model and clustering technique contribute to its computational efficiency.

To assess the accuracy of conformational search engines in predicting the global minima, we compared the generated conformations from Auto3D, CREST, Balloon, and RDKit, with the most stable conformation determined by our QM based workflow. Conformations were ranked based on increasing relative energies, computed using the respective potential energy functions employed by each conformation generation tool. We performed RMSD calculations between the generated conformations and the QM optimized most stable conformation using the LS-align algorithm, a high-throughput virtual screening atom-level structural alignment method developed by Zhang et al.[148] The conformation with the lowest RMSD and energy values was considered the global minima for that particular molecule using the specific conformation generation engine. If no conformation matched these criteria, it was deemed that the engine failed to find the global minima for that molecule.

Furthermore, we calculated the Boltzmann average CCS values using the conformations generated by the conformation search engines and compared them with experimental values. The percentage error in predicting the CCS was reported and an error range within ± 3% was considered indicative of a good CCS prediction as the experimental uncertainty of CCS values within ± 3%.

System setup. In our previous study, we extensively investigated various ionization models (protonation/deprotonation) and their impact on CCS prediction accuracy for metabolites.[10] The predicted CCS values were compared to experimental results to identify the charge model that exhibited the lowest error percentage. In the current study, focusing on finding global minima based on CCS values, we selected the protonation state that yielded the best predicted CCS values (lowest error percentage) for further analysis. For instance, in the case of the carnosine

molecule, five models were considered (model 1, model 2, model 3, model 4, and model 5) with corresponding CCS errors of 9.4%, 9.9%, 8.3%, 0.1%, and 31.0%, respectively. Model 4, exhibiting the lowest error percentage, was chosen as the representative carnosine model for the present investigation. Figure 1 presents an overview of the metabolites included, with their respective ionization sites highlighted in red.

Results and Discussions:

Conformation generation and global minima search. In this study, we examined the performance of various conformation generation engines, including Auto3D, CREST, Balloon, and RDKit, in generating conformations and identifying global minima. Table 1 provides an overview of the number of conformations generated by each engine for the selected metabolites. In the case of QM results, the conformation generation process involved using RDKit to initially generate conformations, followed by clustering and subsequent QM geometry optimization.

Among the engines, Auto3D and CREST had the capability to perform clustering as part of their conformation generation process, whereas Balloon and RDKit did not include this clustering step. Consequently, after generating conformations using Balloon and RDKit, we applied the Autograph clustering algorithm to cluster the resulting conformational ensemble. This allowed for a comprehensive analysis of the conformations and their subsequent evaluation in terms of capturing global minima. The number of conformations generated by Balloon and RDKit prior to the clustering step can be found in Table S1-S64 in the SI. The inclusion of a clustering step in Auto3D and CREST eliminated high energy conformations giving a short list of conformations to consider. On the other hand, Balloon and RDKit produced a significantly higher number of conformations due to the lack of this pruning step. It is worth noting that the number of conformations generated by Balloon was lower than that of RDKit.

To determine the global minima for each molecule, we employed a root-mean-square deviation (RMSD) matrix to compare the conformations generated by the conformation search engines with the lowest-energy QM conformation. The RMSD values for all conformations can be found in the SI specifically Table S1-S102. The conformations were ranked based on both RMSD and energy values, and those achieving the top rank (ranked as 1, lowest RMSD, lowest energy) in both categories were considered global minima and are highlighted in green in Table 2. On the other hand, if the lowest-energy conformation did not correspond to the lowest RMSD value, it indicated that the engines failed to identify the global minima, and these instances are highlighted in red. For instance, in the case of carnosine, Auto3D successfully identified the global minima with a rank of 1 out of 13 conformations, while

RDKit achieved a rank of 1 out of 9 conformations. However, CREST and Balloon were unable to find the global minima, as their lowest RMSD conformations ranked 7 out of 7 and 6 out of 10 in terms of energy, respectively. The details of the RMSD values, relative energies, and corresponding rank of Carnosine conformations are given in Table 3. It is important to note that the range of relative energies obtained from the conformation generation engines exhibits significant variation. Our analysis revealed that the relative energies generated by Auto3D span a wide range, while the relative energies produced by CREST are relatively compressed. For the carnosine system, all seven conformations generated by CREST exhibited relative energies within 5 kcal/mol, whereas none of the 13 conformations generated by Auto3D fell within this range. Notably, Conformation 9 (viz. Conf_9) was the second lowest in energy among the Auto3D conformations, but it had a higher energy by 15.9 kcal/mol. The highest relative energies obtained from Balloon and RDKit were 6.3 kcal/mol and 17.8 kcal/mol, respectively. In comparison, the highest energy conformations generated by Auto3D and CREST were 31.9 kcal/mol and 5.7 kcal/mol, respectively. Detailed energy values for all the molecules can be found in the Supporting Information (Table S1-S102). Out of the 20 metabolites considered in this study, Auto3D successfully identified the global minima for 8 metabolites, while CREST, Balloon, and RDKit each succeeded for 3 metabolites. The success rate of Auto3D in finding global minima was 42%, while CREST, Balloon, and RDKit achieved success rates of 16%, indicating a relatively lower performance.

CCS Prediction and Comparison. We further evaluated the accuracy of CCS predictions for the generated conformational ensembles using all the engines, as summarized in Table 4. The calculated CCS values were compared with experimental CCS values, and predictions within 3% of the experimental values were considered accurate and highlighted in green. Conversely, predictions with errors exceeding 3% were considered inaccurate and highlighted in red.

Our results showed that out of the 20 metabolites, the QM method achieved accurate CCS predictions for 13 metabolites, resulting in a success rate of 65%. Among the conformation generation engines, Auto3D demonstrated accurate CCS predictions for 8 molecules, yielding a success rate of 40%. CREST performed well, achieving accurate CCS predictions for 9 metabolites with a success rate of 50%. However, Balloon and RDKit exhibited lower accuracy, correctly predicting CCS values for only 5 and 6 metabolites, respectively, with success rates of 25% and 30%. The average errors in CCS predictions for the QM, Auto3D, CREST, Balloon, and RDKit generated conformational ensembles were found to be 2.5%, 4.9%, 3.7%, 5.9%, and 6.0%, respectively. Notably, the QM method achieved the

highest accuracy in CCS prediction, highlighting its superiority in capturing the conformational behavior of the metabolites. The semi-empirical-based engine CREST demonstrated a notable success rate of 50% in accurately predicting CCS values. However, the ML-based engine Auto3D exhibited a slightly lower accuracy rate of 40%. The force field-based engines Balloon and RDKit yielded the lowest accuracy rates of 25% and 30%, respectively.

These results underscore the importance of selecting appropriate conformation generation engines for accurate prediction of global minima and CCS values in molecular gas phase conformational ensembles. The QM method, with its ability to capture fine structural details and accurately calculate CCS values, emerges as the most reliable approach. The findings also highlight the promising performance of the semi empirical based engine CREST and the ML based engine Auto3D, while indicating the limitations of FF based tools such as Balloon and RDKit in accurately representing the conformational space and predicting CCS values.

Conclusion:

In this study, we investigated the performance of different conformation generation engines, namely Auto3D, CREST, Balloon, and RDKit, in generating molecular gas phase conformational ensembles. This was accomplished utilizing the GPCL database, which currently encompasses a comprehensive assessment of the conformational ensembles of 20 small molecules. Our aim was to identify the most effective engine for accurate global minima prediction and reliable computational workflows in the fields of drug design and metabolite structure prediction. We utilized a comprehensive computational workflow that encompassed conformer generation, clustering, and analysis of global minima and accurate CCS prediction. The conformations were generated using the respective engines, and we compared them with the global minima obtained through extensive QM computation. We also compared the predicted CCS values of the generated conformations with experimental values to assess their accuracy. Based on the analysis of global minima and accurate CCS prediction, we observed that ML based algorithm, Auto3D achieved the highest success rate in identifying global minima, followed by RDKit, CREST, and Balloon. In terms of CCS prediction accuracy, QM methods yielded the most accurate results, followed by CREST, Auto3D, RDKit, and Balloon. It is noteworthy that while Auto3D demonstrated a higher success rate in global minima identification, CREST exhibited relatively higher accuracy in CCS prediction among the engines considered.

This study provides valuable insights into the performance of different conformation generation engines and their impact on global minima identification and accurate CCS prediction. Moreover, the findings will contribute to the development of more reliable computational workflows for conformational search and related applications in drug

design. Based on our present observations conformational search tools have significant room for improvement for gas-phase ensemble prediction. Further investigations can focus on optimizing the parameters of the conformation generation engines and integrating ML techniques to enhance the accuracy and efficiency of global minima prediction and CCS prediction. Our study emphasizes the significance of selecting appropriate conformation generation engines for the accurate prediction of molecular gas phase conformational ensembles, which has broad implications for drug design and metabolite structure prediction.

ASSOCIATED CONTENT

Data Availability Statement. The data associated with this study are available at a Zenodo (10.5281/zenodo.8247755) repository. The repository includes all the computational results. Researchers can access and download the data to reproduce or analyze the results presented in this manuscript.

Supporting Information. Comprehensive details regarding the conformation generation process, Boltzmann averaging data, RMSD and relative energy ranking, as well as the coordinates of all the molecules can be found in the Supporting Information (SI). The Supporting Information is available free of charge on the ACS Publications website.

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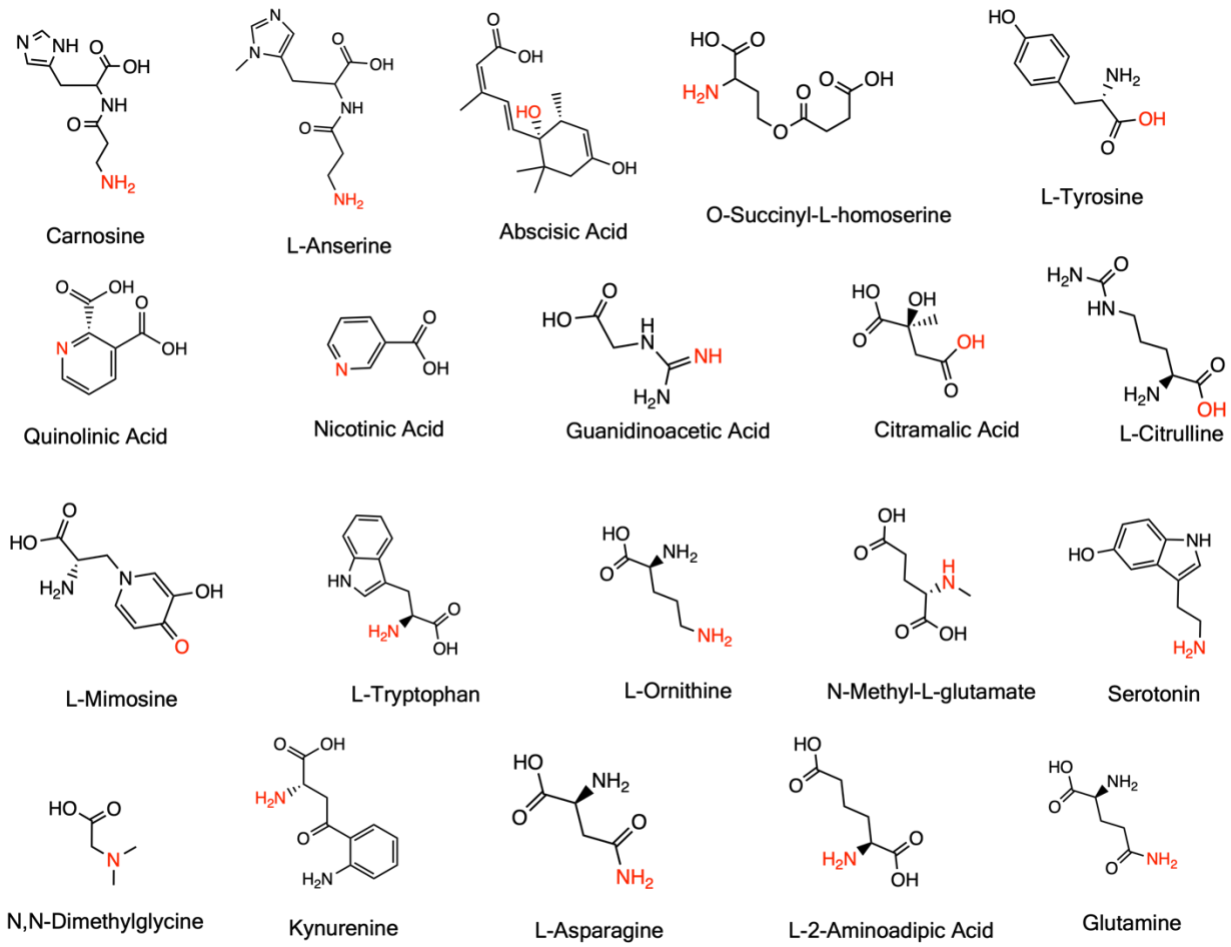


Figure 1. Metabolites examined in the study, highlighting the ionization site in red.

Table 1. Conformation generation results and number of rotatable bonds (N.R.B) for the metabolites. (*The QM geometry optimized conformations after clustering using the standard workflow.)

Number	Metabolites	N.R.B	QM*	Auto3D	CREST	Balloon	RDKit
1	Carnosine	6	12	13	7	10	9
2	O-succinyl-L-homoserine	8	17	13	16	14	23
3	L-tyrosine	3	10	5	11	3	10
4	L-mimosine	3	9	7	15	5	8
5	Citramalic acid	3	9	8	2	3	10
6	N-methyl-L-glutamate	5	17	9	5	12	13
7	L-ornithine	4	12	5	4	4	12
8	Abscisic Acid	3	9	13	9	7	10
9	L-tryptophan	3	10	9	6	4	9
10	L-asparagine	3	8	7	2	4	8
11	L-anserine	6	9	10	4	9	10
12	Kynurenine	4	8	8	7	13	12
13	Serotonin	2	4	6	5	2	7
14	N,N-Dimethylglycine	2	6	6	3	4	9
15	L-citrulline	5	17	10	13	5	14
16	Glutamine	4	9	6	4	3	9
17	L-2-Aminoadipic Acid	5	14	8	8	12	13
18	Guanidinoacetic Acid	2	10	6	4	4	11
19	Nicotinic Acid	1	3	2	3	3	4
20	Quinolinic Acid	2	5	4	3	3	5

Table 2. Metabolites and computed CCS using standard workflow (QM), success (green) or failure (Red) in finding global minima with conformation generation engines: Auto3D, CREST, Balloon, and RDKit.

Number	Metabolites	Predicted CCS (% error)	Auto3D	CREST	Balloon	RDKit
1	Carnosine	150.21 (0.11)	YES (1/13)	NO (7/7)	NO (6/10)	YES (1/9)
2	O-succinyl-L-homoserine	145.45 (0.39)	NO (8/13)	NO (6/16)	NO (3/14)	NO (20/23)
3	L-tyrosine	148.53 (4.14)	YES (1/5)	NO (8/11)	NO (3/3)	YES (1/10)
4	L-mimosine	145.36 (1.43)	YES (1/7)	YES (1/15)	NO (3/5)	NO (2/8)
5	citramalic acid	121.58 (0.24)	NO (6/8)	NO (2/2)	YES (1/3)	NO (6/10)
6	N-methyl-L-glutamate	129.30 (1.98)	NO (7/9)	NO (3/5)	NO (10/12)	NO (8/13)
7	L-ornithine	127.39 (0.95)	YES (1/5)	NO (4/4)	NO (2/4)	NO (11/12)
8	Abscisic Acid	162.86 (0.05)	NO (5/13)	NO (6/9)	NO (7/7)	NO (8/10)
9	L-tryptophan	159.69 (6.06)	NO (2/9)	YES (1/6)	NO (3/4)	NO (3/9)
10	L-asparagine	128.93 (0.20)	NO (2/7)	NO (2/2)	NO (3/4)	NO (2/8)
11	L-anserine	159.74 (3.70)	NO (4/10)	NO (4/4)	NO (4/9)	NO (3/10)
12	Kynurenine	146.94 (0.47)	YES (1/8)	NO (3/7)	NO (10/13)	NO (4/12)
13	Serotonin	131.36 (0.38)	NO (3/6)	NO (2/5)	NO (2/2)	NO (2/7)
14	N,N-Dimethylglycine	118.19 (6.23)	YES (1/6)	NO (2/3)	YES (1/1)	NO (6/9)
15	L-citrulline	141.86 (4.59)	NO (4/10)	NO (3/13)	NO (3/5)	NO (10/14)
16	Glutamine	129.92 (0.58)	NO (3/9)	NO (3/11)	NO (6/9)	NO (4/9)
17	L-2-Aminoadipic Acid	129.56 (1.55)	NO (5/8)	NO (3/8)	NO (8/12)	NO (4/13)
18	Guanidinoacetic Acid	130.21 (2.43)	NO (4/6)	NO (2/4)	NO (2/4)	NO (7/11)
19	Nicotinic Acid	132.17 (3.55)	YES (1/2)	NO (3/3)	YES (1/1)	YES (1/4)
20	Quinolinic Acid	142.14 (5.00)	YES (1/4)	YES (1/3)	NO (3/3)	NO (2/5)

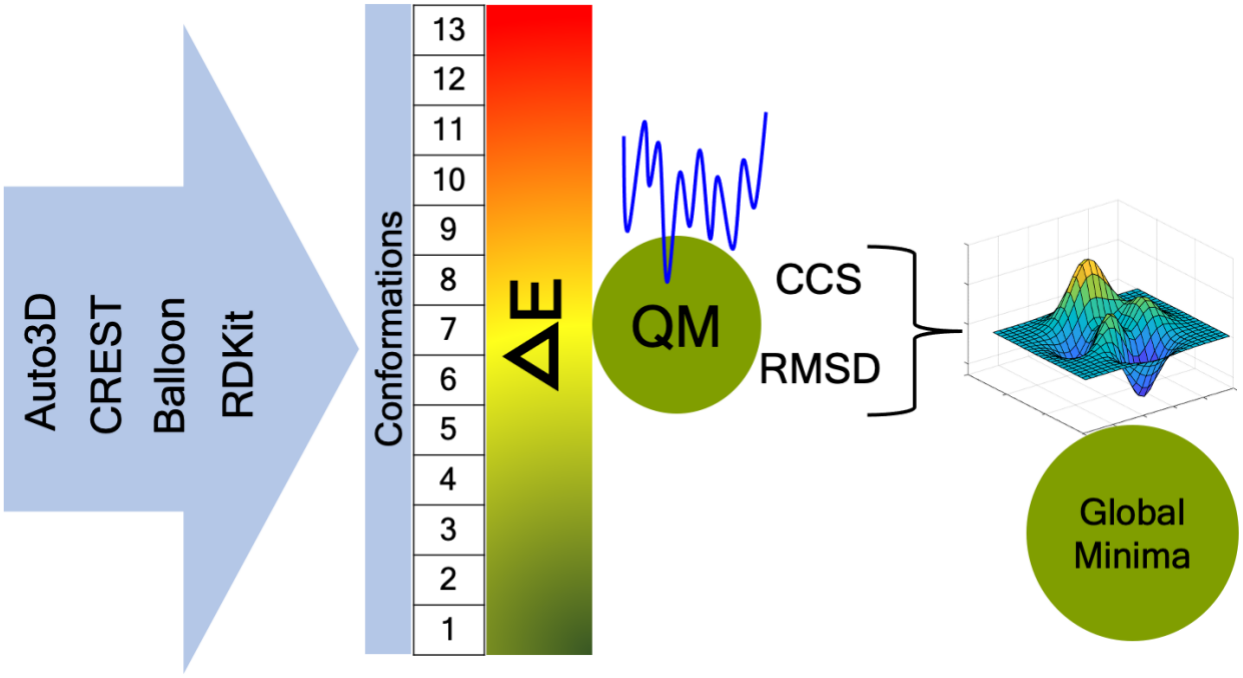
Table 3. RMSD and Relative Energy (kcal/mol) of Carnosine Conformations. The Rank, indicated in parentheses, increases with higher values of RMSD and Relative Energy.

Conformation No.	Auto3D		CREST		Balloon		RDKit	
	RMSD (rank)	Rel_E (rank)	RMSD (rank)	Rel_E (rank)	RMSD (rank)	Rel_E (rank)	RMSD (rank)	Rel_E (rank)
Conf_1	0.07 (1/13)	0.00 (1/13)	0.31 (1/7)	5.72 (7/7)	0.57 (1/10)	3.70 (6/10)	0.49 (1/9)	0.00 (1/9)
Conf_2	0.19 (2/13)	28.84 (10/13)	0.33 (2/7))	3.30 (5/7)	0.72 (2/10)	3.56 (5/10)	0.73 (2/9)	6.82 (3/9)
Conf_3	1.10 (3/13)	26.52 (7/13)	0.57 (3/7)	0.08 (2/7)	0.79 (3/10)	1.65 (3/10)	1.36 (3/9)	9.75 (6/9)
Conf_4	1.54 (4/13)	17.34 (5/13)	0.61 (4/7)	0.00 (1/7)	0.90 (4/10)	2.22 (4/10)	1.54 (4/9)	17.76 (9/9)
Conf_5	1.57 (5/13)	31.99 (13/13)	1.10 (5/7)	2.70 (4/7)	0.90 (5/10)	4.92 (7/10)	1.87 (5/9)	12.47 (8/9)
Conf_6	1.59 (6/13)	17.10 (4/13)	1.49 (6/7)	5.21 (6/7)	1.07 (6/10)	1.53 (2/10)	1.90 (6/9)	6.48 (2/9)
Conf_7	1.67 (7/13)	28.26 (9/13)	1.51 (7/7)	2.25 (3/7)	1.33 (7/10)	0.00 (1/10)	1.92 (7/9)	12.34 (7/9)
Conf_8	1.70 (8/13)	27.89 (8/13)			1.37 (8/10)	5.77 (8/10)	1.96 (8/9)	7.88 (4/9)
Conf_9	1.73 (9/13)	15.90 (2/13)			1.45 (9/10)	6.36 (10/10)	2.01 (9/9)	9.40 (5/9)
Conf_10	1.83 (10/13)	18.28 (6/13)			1.88 (10/10)	5.90 (9/10)		
Conf_11	2.00 (11/13)	16.75 (3/13)						
Conf_12	2.14 (12/13)	29.13 (11/13)						
Conf_13	2.74 (13/13)	29.26 (12/13)						

Table 4. Predicted CCS and error as compared to the experimental value. If the error is within 3%, it is marked as green, indicating accurate CCS prediction. If the error is greater than 3%, it is marked as Red, indicating a poor prediction of CCS value. The CCS values reported here are based on Standard QM results, Auto3D, CREST, Balloon, and RDKit engines, respectively.

Number	Metabolites	QM (% error)	Auto3D (% error)	CREST (% error)	Balloon (% error)	RDKit (% error)
1	Carnosine	150.21 (1.31)	159.60 (5.99)	149.52 (0.35)	161.45 (7.06)	166.49 (9.87)
2	O-succinyl-L-homoserine	145.45 (1.38)	147.86 (2.00)	141.03 (2.47)	147.41 (1.70)	138.52 (4.61)
3	L-tyrosine	148.53 (4.14)	153.78 (7.40)	147.12 (3.21)	153.66 (7.33)	150.41 (5.33)
4	L-mimosine	145.36 (1.43)	146.02 (1.86)	142.49 (0.57)	147.63 (2.93)	147.52 (2.86)
5	citramalic acid	121.58 (0.24)	116.51 (4.11)	118.38 (2.47)	118.38 (1.98)	120.93 (0.31)
6	N-methyl-L-glutamate	129.30 (1.98)	133.98 (1.59)	128.04 (2.98)	135.53 (2.72)	135.93 (3.00)
7	L-ornithine	127.39 (0.95)	119.75 (7.39)	123.48 (4.15)	120.18 (7.01)	133.07 (3.36)
8	Abscisic Acid	162.86 (0.05)	166.06 (1.96)	160.58 (1.38)	174.51 (6.71)	173.12 (5.96)
9	L-tryptophan	159.69 (6.06)	161.97 (10.69)	159.62 (9.38)	161.56 (10.47)	167.83 (13.81)
10	L-asparagine	128.93 (0.20)	124.90 (3.00)	124.20 (3.58)	126.52 (1.68)	125.08 (2.85)
11	L-anserine	159.74 (3.70)	155.98 (1.37)	152.90 (0.62)	166.13 (7.39)	171.23 (10.15)
12	Kynurenine	146.94 (0.47)	156.59 (5.71)	160.65 (8.09)	159.56 (7.46)	157.45 (6.22)
13	Serotonin	131.36 (0.38)	158.29 (6.47)	158.89 (6.82)	159.79 (7.35)	168.18 (11.97)
14	N,N-Dimethylglycine	118.19 (6.23)	113.91 (10.22)	115.02 (9.15)	114.34 (9.80)	116.42 (7.84)
15	L-citrulline	141.86 (4.59)	154.66 (12.49)	135.55 (0.15)	161.73 (16.31)	159.52 (15.15)
16	Glutamine	129.92 (0.58)	126.11 (3.64)	125.95 (3.77)	124.64 (4.86)	132.75 (1.54)
17	L-2-Aminoadipic Acid	129.56 (1.55)	137.00 (3.97)	128.24 (2.58)	138.36 (4.92)	142.40 (7.62)
18	Guanidinoacetic Acid	130.21 (2.43)	120.17 (5.72)	120.23 (5.67)	123.22 (3.11)	122.31 (3.88)
19	Nicotinic Acid	132.17 (3.55)	127.16 (0.27)	126.49 (0.80)	132.29 (3.62)	128.64 (0.89)
20	Quinolinic Acid	142.14 (5.00)	138.84 (2.73)	143.91 (6.16)	140.96 (4.19)	140.84 (4.11)

TOC graphic



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