# Flavonoids as dual-target inhibitors against $\alpha$ -glucosidase and $\alpha$ -amylase: a systematic review of *in vitro* studies

Thua-Phong Lam<sup>1,2,†</sup>, Ngoc-Vi Nguyen Tran<sup>1,2,†</sup>, Long-Hung Dinh Pham<sup>1,3</sup>, Nghia Vo-Trong Lai<sup>1</sup>, Bao-Tran Ngoc Dang<sup>1</sup>, Ngoc-Lam Nguyen Truong<sup>1</sup>, Song-Ky Nguyen-Vo<sup>1</sup>, Thuy-Linh Hoang<sup>4</sup>, Tan Thanh Mai<sup>1,\*</sup>, Thanh-Dao Tran<sup>1,\*</sup>

#### **ABSTRACT**

Diabetes mellitus remains a major global health burden and great attention is directed at natural therapeutics. This systematic review aimed to assess the potential of flavonoids as antidiabetic agents by investigating their inhibitory effects on α-amylase and α-glucosidase, two key enzymes involved in starch digestion. Six scientific databases (PubMed, Virtual Health Library, EMBASE, SCOPUS, Web of Science, and WHO Global Index Medicus) were searched until August 21, 2022, for in vitro studies reporting IC<sub>50</sub> values of purified flavonoids on  $\alpha$ -amylase and  $\alpha$ glucosidase, along with corresponding data for acarbose as a control. A total of 339 eligible articles were analyzed, resulting in the retrieval of 1643 flavonoid structures. These structures were rigorously standardized and curated, yielding 974 unique compounds, among which 177 flavonoids exhibited inhibitory activity against both α-amylase and α-glucosidase are presented. Quality assessment was performed following a modified CONSORT checklist. Structure-activity relationship analysis revealed that a double bond at C2=C3 and a keto group at C4=O were crucial for simultaneous inhibition. The presence of a hydroxyl group at C3 was found to enhance α-glucosidase inhibition but had a detrimental effect on  $\alpha$ -amylase inhibition. Additionally, other important features influencing  $\alpha$ -glucosidase and α-amylase inhibition were discussed. The review also addressed several limitations in the current research landscape and proposed potential solutions. In conclusion, this systematic review provides valuable insights into the investigation of flavonoids as dual-target inhibitors of α-glucosidase and α-amylase and offers directions for future research.

Keywords: systematic review, flavonoids, isoflavonoids, chalcones, catechins, flavones, flavonols, aurones, flavanols, flavanols, anthocyanidins, glucosidase, amylase, PRISMA, SAR.

#### 1. INTRODUCTION

Diabetes mellitus (DM), according to the International Diabetes Federation report in 2021, has been one of the leading causes of mortality worldwide, with an estimated 7 million deaths attributed to the disease. The treatment of DM and its complications requires long-term medical management, imposing an enormous expenditure burden on society. Patients with diabetes are prone to life-threatening microvascular and macrovascular complications with subsequent multiple organ dysfunction. These abnormalities stem from inadequate insulin secretion and/or insulin resistance, characterized by the persistent elevation of blood glucose. Type II diabetes mellitus (T2DM) is associated with insulin resistance, accounting for more than 90% of diagnosed patients globally. Pharmacotherapy for T2DM patients is a person-centered approach developed based on diabetic pathogenesis. Particularly, myriads of mechanisms have been discovered, some of which aim to stimulate insulin secretion, recover insulin sensitivity, prevent the absorption of carbohydrates, or decrease gluconeogenesis. Currently, available anti-diabetic therapeutics include sulfonylureas, meglitinides, thiazolidinediones, biguanides, dipeptidyl peptidase-IV inhibitors (DPP-4i), glucagon-like-peptide-1 receptor agonist (GLP1-RA), sodium-glucose transporter-2 inhibitors (SGLT-2i), and α-glucosidase inhibitors. These medications, in addition to lifestyle modification, are crucial for maintaining normal blood glucose levels in T2DM and preventing severe complications in T2DM.

<sup>&</sup>lt;sup>1</sup> Faculty of Pharmacy, University of Medicine and Pharmacy at Ho Chi Minh City, 700000 Ho Chi Minh City, Vietnam

<sup>&</sup>lt;sup>2</sup> Faculty of Pharmacy, Uppsala University, 75105 Uppsala, Sweden

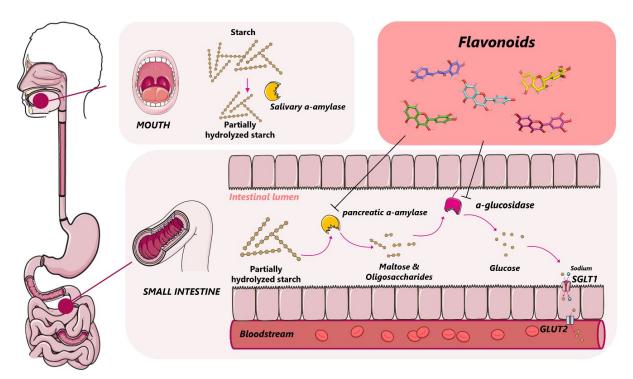
<sup>&</sup>lt;sup>3</sup> Department of Chemistry, Imperial College London, London W12 0BZ, United Kingdom

<sup>&</sup>lt;sup>4</sup> University Medical Center, Ho Chi Minh City, 700000 Ho Chi Minh City, Vietnam

<sup>&</sup>lt;sup>†</sup> These authors contributed equally and are considered co-first authors.

<sup>\*</sup> Corresponding authors: Tan Thanh Mai, <a href="mailto:mthtan@ump.edu.vn">mthtan@ump.edu.vn</a>; Thanh-Dao Tran, <a href="mailto:daott@ump.edu.vn">daott@ump.edu.vn</a>;

One of the key features of patients with T2DM is postprandial hyperglycemia, which is inextricably associated with starch hydrolysis. The starch digestion process commences with the breakdown of polysaccharides into linear and branched malto-oligosaccharides by salivary α-amylase (EC 3.2.1.1). The degradation is intermitted in the stomach and continues in the small intestine with the pancreatic α-amylase, yielding maltose, maltotriose, or smaller oligosaccharides. Following this, four different α-glucosidases, maltase (synonym: α-glucosidase, EC 3.2.1.20), glucoamylase (EC 3.2.1.3), sucrase (EC 3.2.1.48), and isomaltase (EC 3.2.1.10), are responsible for further degradation. It is important to note that, according to the Enzyme Nomenclature (1992),<sup>6</sup> the term "α-glucosidase" does not only refer to a specific enzyme maltase but also a family of hydrolases, yet the term is more commonly used by the scientific community to refer to the former without further clarifications. Human bodies simultaneously produce these four enzymes as two multifunctional complexes, maltase-glucoamylase (MGAM) and sucrase-isomaltase (SI), which cleave α-glycosidic bonds between disaccharides or oligosaccharides to release free monosaccharides.<sup>7,8</sup> The hydrolyzed monomers are then transported into intestinal epithelial cells via specific transporters, such as SGLT1 (for glucose and galactose) or GLUT5 (for fructose), before entering the bloodstream through facilitated diffusion via the GLUT2 transporter in the basolateral membrane.<sup>9</sup> The schematic illustration of starch metabolism is depicted in **Figure 1**.



**Figure 1.** Schematic illustration of starch hydrolysis pathway in humans and potential mechanisms of action of flavonoids as anti-diabetic agents. The figure was partly generated using Servier Medical Art, provided by Servier, licensed under a Creative Commons Attribution 3.0 unported license.

The concept of attenuating the glucose uptake process to maintain blood glucose levels within normal range has become a promising strategy for managing T2DM. Delaying carbohydrate digestion can decrease the portion of glucose entering blood vessels, thereby reducing postprandial glucose levels. Based on the established mode of action, in 1995, the first  $\alpha$ -glucosidase inhibitor (AGI) named acarbose was developed and approved by FDA, followed by miglitol in 1996. However, the use of AGI agents in clinical practice is accompanied by several gastrointestinal side effects, such as flatulence, abdominal distention, and diarrhea. The underlying reason for these events is the substantial inhibitory effects against pancreatic  $\alpha$ -amylases compared to  $\alpha$ -glucosidases. As a result, a considerable amount of complex carbohydrates and starch remain intact throughout the gastrointestinal tract. They are eventually

degraded by bacterial enzymes in the colon, resulting in gas and bloating. Hence, in the search for safer and more effective AGI agents, it should be noted that compounds having modest  $\alpha$ -amylase inhibitory effects are preferable. <sup>11</sup> Flavonoids are natural phenolic compounds characterized by a C6-C3-C6 skeleton consisting of two benzene rings (A and B rings) linked through a three-carbon bridge. In most cases, this three-carbon system forms a heterocyclic pyran ring (C ring). These compounds are well-known for their diverse health-promoting properties, including anti-inflammatory, <sup>12</sup> anti-oxidative, <sup>13</sup> anti-infective, <sup>14</sup> and anti-obesity effects. <sup>15</sup> Notably, several studies have highlighted the potential of flavonoids as anti-diabetic agents due to their strong inhibition of  $\alpha$ -glucosidase and moderate inhibition of  $\alpha$ -amylase, making them promising candidates for the development of anti-diabetic drugs with minimal gastrointestinal side effects. <sup>16-18</sup>

Over the years, the  $\alpha$ -amylase and  $\alpha$ -glucosidase inhibitory effects of flavonoids have been recorded in numerous reviews. <sup>19-24</sup> Nevertheless, to the best of our knowledge, there is no systematic review about the concurrent inhibition of flavonoids against  $\alpha$ -amylase and  $\alpha$ -glucosidase that has been reported until this project commenced (August 2022). Therefore, this systematic review aims to present the current evidence supporting the simultaneous inhibition of flavonoids against the two starch-digestive enzymes and propose a structure-activity relationship (SAR) analysis that could be useful for developing more effective and safer anti-diabetic therapeutic agents in the future.

#### 2. METHODS

## 2.1. Protocol and registration

This study aimed to provide a general overview of flavonoids that could inhibit both  $\alpha$ -glucosidase and  $\alpha$ -amylase. Our study is conducted following The Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) statement.<sup>25</sup> The protocol of this review was uploaded on ResearchGate in August 2022 (DOI: 10.13140/RG.2.2.17980.31368/2). As this study is only concerned with *in vitro* studies, we could not register our protocol on the International Prospective Register of Systematic Reviews (PROSPERO) server. The PRISMA checklist is available in **Table S1**.

# 2.2. Eligibility criteria

A study must meet the following criteria to be included in this systematic review: (1) original *in vitro* studies reporting the inhibitory effects of purified flavonoids on  $\alpha$ -amylase or  $\alpha$ -glucosidase; (2) provide IC<sub>50</sub> results of tested compounds with the concurrent positive control (3) be written in Vietnamese or English. For comparison purposes, we only included the studies which used acarbose as the positive control, as acarbose is the most common substance used in the literature. For exclusion criteria, editorials, case reports, reviews, systematic reviews, conference abstracts, and unpublished papers were excluded. We also eliminated pure *in silico*, *in vivo*, *ex vivo*, or clinical studies. Studies without concurrent positive samples, and those without a clear method description that could present risks of producing unreliable extracted data, were also excluded.

### 2.3. Information sources and search strategies

An electronic search was conducted on six databases until 21 August 2022, including PubMed, Virtual Health Library (VHL), EMBASE, SCOPUS, Web of Science (WOS), and WHO Global Index Medicus (GIM). The search terms (alpha-glucosidase OR maltase OR sucrase OR alpha-amylase) AND (flavonoid OR flavonoids) were customized to fit the corresponding electronic databases. No time restriction, language restriction, and article type restriction were set at this stage. In addition to literature screening using the aforementioned databases, we also searched for structures of flavonoid-enzyme complexes on the RCSB Protein Data Bank (PDB) (https://www.rcsb.org). The search was conducted on April 27<sup>th</sup>, 2023, to systematically evaluate the interaction patterns between flavonoids and two enzymes of interest. The PDB search terms were set to be the Enzyme Commission number (E.C. number) of the corresponding enzymes. The detailed search strategy is available in **Table S2 and Table S3**.

### 2.4. Study selection

During this research, we screened the papers by titles, abstracts, and full texts with the assistance of the online systematic review software Rayyan (<a href="https://rayyan.ai">https://rayyan.ai</a>). Our study selection process consists of three main stages, including duplicate and automatic removal, title/abstract screening, and full-text screening.

The study selection procedure began with eliminating duplicate papers using the Zotero and Rayyan programs, followed by splitting the initial database into three portions for the next stage. Six reviewers who worked in pairs of two (T.P.L. and S.K.N.V.; N.V.T.L. and B.T.N.D.; N.V.N.T. and N.L.N.T.) performed the title/abstract screening for matching articles. Subsequently, full-text screening was conducted to reach the final include or exclude decision. The previously indicated reviewers operated independently and were blinded throughout the screening process. A debate and consensus would be held if there was a dispute, and a third reviewer's viewpoint (T.D.T. and T.T.M.) would be sought to conclude. The summary of the screening results is graphically presented in a PRISMA flow chart (**Figure 2**). In addition, the reasons for full-text exclusion are illustrated in the flow chart.

## 2.5. Data collection process and data items

The following information was collected from each study and stored in a Microsoft Excel workbook. The extracted data consisted of 3 parts: (1) Study determinants, including title, authors, and publication year; (2) Study design (assay protocol, types of enzymes), sample size, intervention, and comparator; (3) Outcomes of the study. Concerning primary outcomes, we searched for the  $\alpha$ -amylase and/or  $\alpha$ -glucosidase inhibitors' chemical structures and their inhibitory efficiency.

## 2.6. Outcomes and prioritization

According to the respective flavonoid subclasses, compounds and derivatives that have one or both inhibitory activities against  $\alpha$ -glucosidase and  $\alpha$ -amylase were classified. The structures of compounds were redrawn using ChemDraw software<sup>27</sup> and stored as Simplified molecular-input line-entry system (SMILES) strings in the same aforementioned workbook. The inhibitory activity outcomes of flavonoids against enzymes were presented as the half-maximal inhibitory concentration (IC<sub>50</sub>).

## 2.7. Quality assessment

A modified version of the CONSORT checklist developed by Faggion<sup>28</sup> was used in the quality assessment process. The checklist consisted of 14 items concerning the assessment of the abstract, introduction, method, results, discussion, and other information to interpret the quality of the research studies. In this study, we only employed Items 1-4, and 10-13 for quality evaluation due to the lack of information on randomization, blinding, and sample size in most of the *in vitro* studies. The detailed checklist and quality evaluation of included studies is provided in **Table S5**.

## 2.8. Data processing

As one natural product may have many synonymous names, it is insufficient to use the compounds' names to organize extracted results from different studies. To overcome this situation, we used SMILES string representation to store the chemical information of the compounds. However, varying SMILES generation algorithms also lead to inconsistent results. To address this, the MolStandardize module in the Python RDKit library<sup>29</sup> was used to standardize the molecular SMILES structures, yielding RDKit canonical SMILES that allowed for deduplication and identification of compounds. Moreover, IC<sub>50</sub> values in mass per volume (m/V) were converted to the standard molar concentration (C<sub>M</sub>/V). The ExactMolWt function from RDKit was employed to calculate the molecular weight of each compound and the acarbose control. An additional column of IC<sub>50</sub> ratio between the test molecules and the control was also calculated and annotated with colors as presented in the Excel Workbook available online at <a href="https://github.com/phonglam3103/Systematic\_review">https://github.com/phonglam3103/Systematic\_review</a>.

#### 3. Results

## 3.1. Systematic search and study selection

The initial search resulted in an accumulation of 9,694 records from the six databases. Following the duplicate removal process, the acquired literature was filtered based on their language and article type characteristics by the Zotero and Rayyan program. The results are 4,408 records that would undergo the title and abstract screening process. We obtained 570 full-text articles and further evaluated them for inclusion eligibility. 231 reports were excluded, thus a total of 339 studies were included in the present review. These studies went through a data extraction process, and flavonoids were grouped based on their SMILES and systematically scrutinized for their capacity to inhibit the two

digestive enzymes. Finally, 177 flavonoids, which have concurrent IC<sub>50</sub> values against both  $\alpha$ -glucosidase and  $\alpha$ -amylase, are presented in the current review.

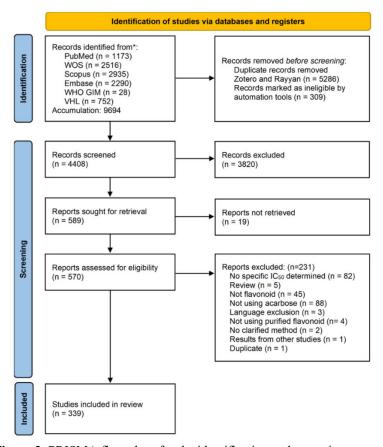


Figure 2. PRISMA flow chart for the identification and screening process.

## 3.2. Study characteristics

A total of 339 research articles, involving at least 1,643 flavonoid structures, were published from 1998 to 2022. Of these, 51 studies conducting both  $\alpha$ -glucosidase and  $\alpha$ -amylase inhibition assays were recorded. Thirty-six studies involved  $\alpha$ -amylase inhibition assay, whereas 252 remaining studies involved only  $\alpha$ -glucosidase inhibition assay. Among the included studies, various flavonoid sources have been documented. As anticipated, most flavonoids were derived from natural sources, with their proportion taking place for 76 percent (257 over 339 studies). Different sources of the enzymes used in the literature were also observed. Yeast  $\alpha$ -glucosidase, especially one derived from *Saccharomyces cerevisiae*, is the most common source of  $\alpha$ -glucosidase used in the literature with the proportion standing at over 70 percent (190 over 303 studies). On the other side, porcine pancreatic  $\alpha$ -amylase is the most popular source used in the  $\alpha$ -amylase involved assay, followed by enzymes from human sources such as human salivary and human pancreatic ones.

A variety of assay methods used in the literature were also documented and characterized based on the substrate and the principle of detection used in the studies. For the  $\alpha$ -glucosidase inhibition assay, a chromogenic method that employed the use of a synthetic substrate *p*-nitrophenyl- $\alpha$ -*D*-glucopyranoside (pNPG) is the most popular method used to evaluate the inhibitory activity of the flavonoids. In this methodology, the pNPG substrate will be hydrolyzed by the enzyme and therefore release a colorimetric *p*-nitrophenol compound. The amount of released *p*-nitrophenol will be measured using the absorbance at around 405 nm. Other assay methods were employed to assess the inhibitory ability of the flavonoids against  $\alpha$ -glucosidase such as the enzymatic method employing glucose oxidase. In the  $\alpha$ -amylase inhibition assay, six different assay principles were recorded, with the most cited technique being the reducing sugar method (51 over 87 studies). In this approach, the liberated sugars from the starch will act as reducing agents

and oxidize the 3,5-dinitrosalicylic acid (DNSA) molecule to yield the deep orange solution which absorbs light strongly at 540 nm.<sup>30</sup> Other assay principles include the chromogenic method (23/87), turbidimetric method (3/87), iodine-starch method (8/87), and enzymatic method (2/87). The detailed characteristics of the included studies are available in **Table S4**.

#### 3.3. Main results

Although our database initially included 1,643 flavonoid structures, only 177 compounds were found to be able to concurrently inhibit both α-glucosidase and α-amylase after merging the duplicates. The original database of 177 presented compounds the full dataset of 1,643 entries available https://github.com/phonglam3103/Systematic review. Unlike other reviews which take a study-by-study approach, 19,23 we used a structure-based approach to summarize results from various studies reporting the same chemical structure. We believe that this approach will provide viewers with a more comprehensive evaluation of compound potency compared to acarbose. To limit variations between studies, we used the median and interquartile range of pIC<sub>50</sub> values and stated inhibitory mechanisms if reported by any of the studies.

## 3.3.1. Flavan – flavanols

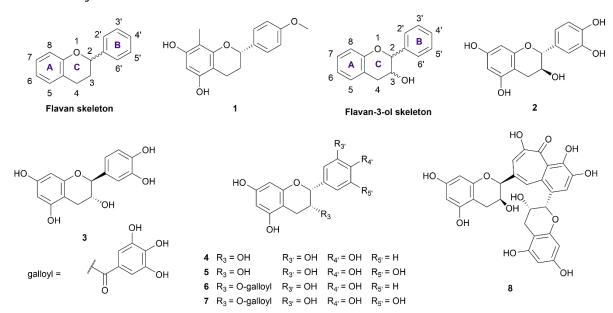


Figure 3. Chemical structures of retrieved flavan and flavan-3-ol derivatives

## 3.3.1.1. Flavans

Flavans are widely distributed in nature and originate from the reduction of flavanone. This flavonoid class is characterized by the absence of a double bond between the  $C_2$  and  $C_3$  positions. However, the most basic flavan skeleton (**Figure 3**) is not usually observed in plant tissues as the  $C_3$  or  $C_4$  positions are usually hydroxylated or ketonized to form other flavan subclasses. In the present review, we only record (2S)-4'-hydroxy-5,7-dimethoxy-8-methylflavan (1), a flavan extracted from the plant of *Dracaena angustifolia*, as a dual-target inhibitor. This compound was able to inhibit  $\alpha$ -glucosidase and  $\alpha$ -amylase at the concentration of 370  $\mu$ M and 6.03 mM, respectively.<sup>31</sup> In comparison with acarbose in the same condition, compound 1 exhibited stronger inhibition on  $\alpha$ -glucosidase but weaker in the  $\alpha$ -amylase inhibition assay.

#### 3.3.1.2. Flavanols (Catechins)

Flavanol inherits the flavan scaffold where the  $C_3$  and/or  $C_4$  position is hydroxylated to form three different subclasses: flavan-3-ol, flavan-4-ol, flavan-3,4-diol, and the most well-known example of which being catechin. In the current review, seven flavan-3-ol structures (2-8) were recorded to have concurrent anti- $\alpha$ -glucosidase and anti- $\alpha$ -amylase activity (**Figure 3**). Based on the two chiral centres  $C_2$  and  $C_3$ , the flavanols can exist in four different stereoisomers

namely (2R,3S), (2S,3R), (2S,3S). **Table S6** summarizes the potency of included Flavan and Flavanol derivatives against  $\alpha$ -glucosidase and  $\alpha$ -amylase.

(+)-Catechin (2) is the most well-studied flavan-3-ol with the IC<sub>50</sub> values against the α-glucosidase and α-amylase ranging from 1.12 to 1276.51 μM and 0.31 μM to at most 70.87 mM, respectively. Considering α-glucosidase inhibition, the current evidence suggests that this compound can inhibit the enzyme as potently as the standard drug in a competitive manner.<sup>32</sup> On the other hand, this compound exhibits two to three times weaker inhibition against the α-amylase than acarbose.<sup>33-37</sup> The wide range of reported results could stem from the various origin of the enzyme and assay protocol. On the contrary, (-)-catechin (3), a 3R isomer of 2, despite achieving low IC<sub>50</sub> against both enzymes, compares far less favorable than that of the standard drug acarbose, indicating a weaker inhibitory activity of this compound.<sup>38</sup> (-)-Epicatechin (4), another (2R,3R) isomer of 2, has also been documented for its inhibitory activity against two digestive enzymes. Most studies suggest compound 4 to be a weak inhibitor, in comparison with acarbose in the same condition.<sup>39,46</sup> However, the results are still controversial, not only in the reported IC<sub>50</sub> value but also in the underlying mechanism. In a study reported by Giang *et al.*,<sup>47</sup> this compound was able to inhibit ten times better than acarbose in both inhibition assays. However, this is the only study that suggests 4 as a strong inhibitor against α-amylase. Thus, further evidence is needed to confirm the potency of this compound. (-)-Epigallocatechin (5) was reported in three studies,<sup>39, 61, 62</sup> all of which consented that this compound was able to inhibit the two enzymes but with weaker activity in comparison to acarbose.

In plant tissues, flavanols are usually esterized with gallic acid to form their gallate derivatives. In this review, two gallate derivatives, namely (-)-epicatechin gallate (ECG, 6) and (-)-epigallocatechin gallate (EGCG, 7) are reported to have concurrent inhibitory activity against  $\alpha$ -glucosidase and  $\alpha$ -amylase. Together with theaflavin (8), these two compounds are famous for their antioxidant, anti-inflammatory, anti-diabetic activity, and abundance in tea tissue (*Camellia sinensis*).<sup>50</sup> The current evidence suggests that both 6 and 7 can inhibit  $\alpha$ -glucosidase stronger than acarbose in the same condition. On the other hand, these two compounds also exhibit weak inhibition against  $\alpha$ -amylase, indicating their potential as promising drug candidates which can minimize the undesirable effects of acarbose. The inhibitory mechanisms of the two compounds were reported to be non-competitive against  $\alpha$ -glucosidase and in a mixed, competitive, or non-competitive manner towards  $\alpha$ -amylase.<sup>51-54</sup> However, the inhibition pattern of theaflavin (8) is the opposite. This compound was reported to be a weak  $\alpha$ -glucosidase inhibitor but a strong  $\alpha$ -amylase inhibitor with the IC<sub>50</sub> of 16.17  $\mu$ M and 0.46  $\mu$ M, respectively.<sup>38</sup>

# 3.3.1.3. Flavanones

Flavanones (also called dihydroflavones) are characterized by the absence of the double bond between  $C_2$  and  $C_3$  in the C-ring and the presence of the ketone group in  $C_4$  (**Figure 4**). In plant tissue, this subclass of flavonoids is biosynthesized from chalcone-like compounds by chalcone isomerase.<sup>55</sup> Flavanones are most famous for their presence in *Citrus* spp. (Rutaceae).<sup>56</sup> In our review, 15 flavanone structures (9-23) that have sufficient evidence for their abilities to inhibit both  $\alpha$ -glucosidase and  $\alpha$ -amylase were included and summarized in **Table S7**.

Naringenin (9) is the most well-studied flavanone with 16 studies employing this compound in their inhibition assay. <sup>38, 44, 53, 67, 78-89</sup> Concerning the  $\alpha$ -glucosidase inhibition, naringenin was recorded to exhibit stronger inhibitory activity than acarbose in the same condition in the majority of the reported results. For  $\alpha$ -amylase inhibition, this compound also exhibited strong inhibition against this enzyme with the IC<sub>50</sub> values ranging from 6.20 to at most 121.50  $\mu$ M. <sup>38, 53, 80</sup> The geranylation of naringenin at the C<sub>8</sub> position forming 8-geranylnaringenin (10) was reported to enhance the inhibitory activity of this compound against both enzymes. <sup>5</sup> Eriodictyol (11), a C<sub>3</sub>' hydroxyl derivative of naringenin, was also reported in the literature for its ability to inhibit both enzymes. <sup>71-75</sup> In comparison with acarbose, this compound was a stronger inhibitor, regardless of the enzyme type employed. Propolin D (12), Propolin H (13), Propolin F (14), Propolin C (15), and Propolin G (16) are five geranylated flavanone aglycones derived from propolis of the Australian honeybees (*Apis mellifera*) reported by Uddin *et al.* <sup>59</sup> In his study, the geranylation at either C<sub>2</sub>· or C<sub>3</sub>· position significantly enhanced the inhibitory activity of flavanone compounds against both enzymes. However, the geranylation or prenylation at the C<sub>6</sub> position reduced such activity against  $\alpha$ -glucosidase but had less impact on the  $\alpha$ -amylase inhibition manner.

Flavanone skeleton 9 
$$R_3 = H$$
  $R_8 = H$  12  $R_2 = geranyl$   $R_3 = OH$   $R_5 = H$   $R_6 = H$  10  $R_3 = H$   $R_6 = geranyl$  13  $R_2 = H$   $R_3 = geranyl$   $R_5 = OH$   $R_6 = H$  11  $R_3 = OH$   $R_6 = H$  14  $R_2 = H$   $R_3 = geranyl$   $R_5 = OH$   $R_6 = H$  15  $R_2 = H$   $R_3 = OH$   $R_5 = H$   $R_6 = geranyl$   $R_6 = OH$   $R_7 = O-geranyl$   $R_7 = OH$   $R_8 = H$   $R_9 = Prenyl$   $R_9 = Prenyl$   $R_9 = H$   $R_9 = OH$   $R_9 = OH$   $R_9 = H$   $R_9 = OH$   $R_9 = OH$ 

Figure 4. Chemical structures of retrieved flavanone derivatives

Isocarthamidin-7-O-glucuronide (17) is a flavanone glycoside derived from the shoot of Scutellaria baicalensis reported by Li et al. <sup>76</sup> This compound was a weak inhibitor against both  $\alpha$ -glucosidase and  $\alpha$ -amylase, in comparison with acarbose, with the respective values being 4.6 mM and 6.3 mM.<sup>76</sup> Different glycoside derivatives of naringenin were also recorded in the literature, with naringin (18) and narirutin (19) being the two representatives. Overall, naringin (18) was recorded to exhibit strong inhibition against both enzymes in the majority of studies involved. 18,68,69,77-79 However, in two studies reported by Kong et al., 8 and Zhang et al., 68 this compound was reported to inhibit α-glucosidase not as well as acarbose in the same condition, as the IC<sub>50</sub> values stood at 27.2 mM and 22.1 mM, respectively. Therefore, further evidence is needed to confirm the potency of this compound. Narirutin (19), a 7-O-rutinose of naringenin (9), showed comparable inhibitory activity to acarbose against  $\alpha$ -glucosidase but far stronger inhibitory activity against  $\alpha$ -amylase. 18,80 Likewise, poncirin (20) also exhibited similar activity against  $\alpha$ glucosidase but was reported to be stronger than acarbose in the  $\alpha$ -amylase inhibition assay. 18,77 Hesperidin (21), a flavanone glycoside that has long been known for its anti-atherogenic and venous protection activity, 81 is also included in our review. In the literature, hesperidin was reported to be a potent antidiabetic therapeutic agent, as it exhibited strong inhibitions against both  $\alpha$ -glucosidase and  $\alpha$ -amylase, reported by at least four studies. <sup>18,63,77,79</sup> Kuwanon L (22) and Sanggenon G (23) are two flavanone compounds derived from the root bark of *Morus alba*. Et In their study, Zhao et al. 82 described these two compounds as strong  $\alpha$ -glucosidase inhibitors and moderate  $\alpha$ -amylase inhibitors.

# 3.3.1.4. Flavanonols

Flavanonol (also referred to as 2,3-dihydroflavonol) is a small flavonoid subclass resulting from the hydroxylation at the  $C_3$  position of flavanone structures (**Figure 5**). In this review, three representatives, namely taxifolin (24), silibinin (25), and dysosmaflavanone (26) are included and their pIC<sub>50</sub> are presented in **Table S8**.

Figure 5. Chemical structures of retrieved flavanonol derivatives

Taxifolin (also called dihydroquercetin, **24**) was reported in six studies for its potency to inhibit both  $\alpha$ -glucosidase and  $\alpha$ -amylase.<sup>35, 101-105</sup> The results showed that this compound exhibited comparable inhibitory activity to that of acarbose in the same condition while inhibiting  $\alpha$ -amylase more weakly. Concerning the underlying mechanism, Su *et al.* <sup>86</sup> reported that this flavonoid was able to inhibit both enzymes competitively. Silibinin (**25**), a flavanonol-lignan hybrid, was reported for its ability to inhibit two carbohydrate-hydrolyzing enzymes in a study conducted by Yang *et al.* <sup>88</sup> In this study, silibinin exhibited strong inhibition against  $\alpha$ -glucosidase, yet moderate inhibition against  $\alpha$ -amylase, in comparison with acarbose. The mechanism of a non-competitive manner was also reported in his study. Additionally, dysosmaflavanone (**26**), a deoxygenated flavanonol derived from *Dysosma difformis*, is also included in this review and has been recorded to exhibit weak inhibition against both enzymes in one study. <sup>89</sup>

## 3.3.2. Flavones

The flavone scaffold constituted a large proportion of our included dataset, with 40 structures (numbered 27-67) recorded of activity on both enzymes of interest (**Figure 6**). Flavone compounds are distinguished from flavanones by the presence of the  $C_2=C_3$  double bond, thus constructing the chromene core (4*H*-1-benzopyran). Hydroxy and methoxy substituents are abundant in this sub-class of flavonoids and vary their positions in all three rings A, B, and C. A summary of flavone inhibitory potency against  $\alpha$ -glucosidase and  $\alpha$ -amylase is available in **Table S9**.

From our curated data, chrysin (27), apigenin (28), and luteolin (30) were widely researched flavones, of which apigenin (28) was the subject of 30 studies.  $^{35, 49, 53, 54, 81, 82, 87, 94, 108-128}$  Despite various origins from numerous plants, there is a general consensus regarding apigenin's activity as a more potent  $\alpha$ -glucosidase inhibitor than acarbose but a weaker  $\alpha$ -amylase antagonist, making apigenin a potential candidate for nutritional and pharmaceutical purposes. However, it should also be noted that 9 out of 30 studies  $^{35, 49, 53, 54, 81, 108, 110, 114, 117}$  disregard apigenin's stronger inhibition on  $\alpha$ -glucosidase than acarbose, but there was no dispute over the activity on  $\alpha$ -amylase. Apigenin's IC<sub>50</sub> on  $\alpha$ -glucosidase was reported in a large range from 9.04  $\mu$ M to 34.3 mM but these results should be taken into account in relation to the acarbose reference and experimental protocols.

Luteolin (30) exhibits a similar inhibition profile on both enzymes but to a lesser extent of consent on  $\alpha$ -glucosidase. 18 out of 31 recorded findings  $^{60,71,94,95,97,100,102,104,107,108,111-118}$  showed stronger activity on  $\alpha$ -glucosidase than acarbose, in contrast with the remaining 13 studies,  $^{39,61,73,90,92,96,99,119-124}$  making a 6:4 conflict. There was greater unity in the results of  $\alpha$ -amylase inhibition, confirming a moderate but generally weaker activity than acarbose on  $\alpha$ -amylase (IC50 ranging from 14.57  $\mu$ M to 1 mM).  $^{39,90,125,126}$  On a side note, 5-deoxyluteolin (29) is structurally related to luteolin (30) but is more limited in experimental data.  $^{39,102,124}$  The currently available results suggest that the absence of the O5 oxygen atom decreased 5-deoxyluteolin's activity on both enzymes compared to acarbose.

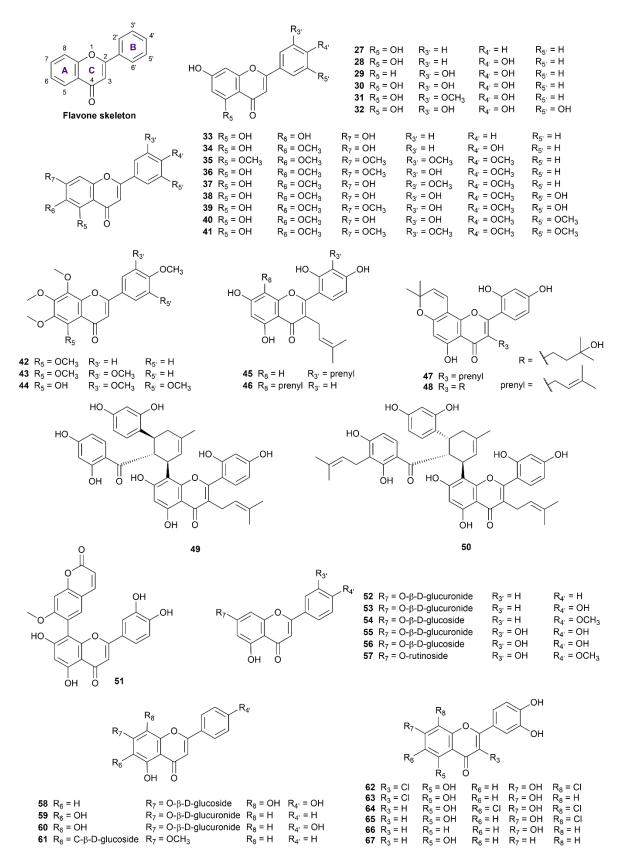


Figure 6. Chemical structures of retrieved flavone derivatives.

Chrysin (27) is next on the list of commonly researched flavones, with eight studies  $^{76,79,111,127-131}$  conducted on  $\alpha$ -glucosidase and one result on  $\alpha$ -amylase. The comparison of activity on  $\alpha$ -glucosidase is inconsistent and equally divided between stronger  $^{79,111,128,129}$  and weaker inhibition than acarbose. Meanwhile, the only reported result on  $\alpha$ -amylase showed weak inhibition at the IC<sub>50</sub> of 1.771  $\mu$ M.

Chrysoeriol (31) and tricetin (32) are closely related to luteolin but lesser researched flavones. Interestingly, there was great consensus in the three studies confirming chrysoeriol's stronger activity on  $\alpha$ -glucosidase than acarbose,  $^{57,102,132}$  and one result reporting on its generally weak inhibition of  $\alpha$ -amylase at 1,270  $\mu$ M,  $^{133}$  making chrysoeriol (31) an attractive ligand for further optimization. Meanwhile, tricetin (32) was the subject of only one paper and was found to weakly inhibit both enzymes.  $^{90}$ 

Proceeding to the next group of  $R_6$ -substituted flavones, baicalein (33) is a major topic of ten studies,  $^{60,73,76,102,119,121,127,130,131,134}$  two out of which  $^{76,119}$  reported  $IC_{50}$  on both  $\alpha$ -glucosidase and  $\alpha$ -amylase. The included literature commonly demonstrated this compound's moderate but weaker inhibition on both enzymes than acarbose.

Hispidulin (34) is another intriguing case for antidiabetic profile optimization. In two studies,  $^{107,135}$  hispidulin was shown to be a stronger inhibitor of  $\alpha$ -glucosidase than acarbose in the same experimental conditions. As for  $\alpha$ -amylase inhibition, hispidulin exhibited a weak activity with the IC  $_{50}$  at 30.08  $\mu$ M, much higher than that of acarbose in the same settings.  $^{105}$ 

Eupatorin (36) and eupatillin (37) are two flavones shown to strongly inhibit both  $\alpha$ -glucosidase and  $\alpha$ -amylase at nanomolar concentrations in a paper by Gulcin *et al.* <sup>136</sup> These activity profiles were significantly stronger than acarbose in the same procedure and conditions, <sup>136</sup> thus should be further studied for their molecular interactions with enzyme targets.

Proceeding to the next group of  $R_6$ -substituted flavones, baicalein (33) is a major topic of ten studies,  $^{60,73,76,102,119,121,127,130,131,134}$  two out of which  $^{76,119}$  reported  $IC_{50}$  on both  $\alpha$ -glucosidase and  $\alpha$ -amylase. The included literature commonly demonstrated this compound's moderate but weaker inhibition on both enzymes than acarbose.

Hispidulin (34) is another intriguing case for antidiabetic profile optimization. In two studies,  $^{124, 155}$  hispidulin was shown to be a stronger inhibitor of  $\alpha$ -glucosidase than acarbose in the same experimental conditions. As for  $\alpha$ -amylase inhibition, hispidulin exhibited a weak activity with the IC<sub>50</sub> at 30.08  $\mu$ M, much higher than that of acarbose in the same settings.  $^{105}$ 

In another notable report,  $^{137}$  four methoxylated flavones at  $R_6$  and  $R_{4^{\circ}}$  (38, 39, 40, 41) all showed an appealing profile with stronger glucosidase inhibition than acarbose (IC<sub>50</sub> ranging from 49 – 77  $\mu$ M) and a simultaneous weak amylase activity (IC<sub>50</sub> varying from 120 – 338  $\mu$ M). Analysis of the structure-activity relationship of  $R_{4^{\circ}}$ -methoxylated flavones can be examined to further enhance these profiles.

Other less data-abundant methoxylated flavones include tangeretin (42), nobiletin (43), and gardenin A (44), whose collective results were not yet conclusive about their activity potency. Promptly, it can be noted that nobiletin (43) inhibited  $\alpha$ -glucosidase stronger than acarbose and moderately inhibited  $\alpha$ -amylase. Meanwhile, gardenin A (44) inhibited both enzymes with a much stronger activity than acarbose in the same conditions. 136

The next group of structural-related compounds includes kuwanon T (45), kuwanon C (46), morusin (47), morusinol (48), kuwanon G (moracenin B) (49), moracenin A (50), and 8-(7-methoxycoumarin-6-yl)luteolin (51). This group is characterized by the presence of a prenyl group at the R<sub>3</sub> position (except for compound 51), and more bulky side chains at the R<sub>8</sub> position for compounds (49), (50), and (51). All seven compounds exhibited inhibitory profiles on both enzymes but noticeably compared much more favorably and selectively on α-glucosidase than acarbose. <sup>99, 161-163</sup> Once again, these gathered results facilitate future efforts in investigating these interesting scaffolds.

The next ten compounds (**52-61**) of this subclass feature flavone glycosides, most commonly glycosylated at the  $R_7$  oxygen. Of these glycosides, luteolin-7-O- $\beta$ -D-glucoside (**56**) is the most well-researched with five studies, <sup>35, 115, 124, 135, 164</sup> outlining a comparable activity to acarbose on  $\alpha$ -glucosidase and a weaker inhibition on  $\alpha$ -amylase. <sup>143</sup> Relatedly, luteolin-7-O- $\beta$ -D-glucuronide (**55**) and diosmin (**57**) all displayed equivalent profiles to acarbose on both enzymes. <sup>164, 165</sup> On a side note, apigenin-7-O- $\beta$ -D-glucuronide (**53**) and baicalin (**59**)'s results on  $\alpha$ -glucosidase vary significantly

between studies (ranging from  $13.63\mu\text{M}$  to 1.217 mM and  $36.3~\mu\text{M}$  to 1.324 mM, respectively),  $^{53,\,94,\,137,\,143}$  thus careful consideration is recommended when interpreting these results. Meanwhile, swertisin (61) is one of the uncommon flavone C-glycosides at the R<sub>6</sub> position and was found by two separate studies to inhibit  $\alpha$ -glucosidase more strongly than acarbose  $^{145}$  while maintaining a negligible activity on  $\alpha$ -amylase (IC<sub>50</sub> at 4.245 mM).  $^{146}$  This interesting structure and attractive activity profile may hence motivate future molecular interaction studies.

The last six compounds (62-67) are flavone derivatives that were synthesized and evaluated for their simultaneous inhibition against  $\alpha$ -glucosidase and  $\alpha$ -amylase by Proença *et al* in two consecutive publications. Overall, these compounds exhibited strong inhibition against the  $\alpha$ -glucosidase and moderate inhibition against the  $\alpha$ -amylase, in comparison with acarbose in the same condition.

#### 3.3.3. Flavonols

Flavonols are probably the most common flavonoid subclasses found across the plant kingdom (except for algae). <sup>148</sup> Flavonols are characterized by the presence of hydroxy substituent at the  $C_3$  position in ring C of flavone structure (**Figure 7**), also known as 3-hydroxy flavones. This subclass of flavonoids received considerable attention in the research of prominent natural substances for the simultaneous inhibitory effect on  $\alpha$ -glucosidase and  $\alpha$ -amylase, with 54 structures (68-121) taken into consideration in the present study (**Table S10**). Quercetin (68), which has been the subject of 75 articles, has garnered the most attention of all the flavonol aglycones, followed by kaempferol (69) and myricetin (70).

Quercetin (68) is a natural polyphenol obtained in more than 50 plant species, as shown in the curated dataset. In terms of  $\alpha$ -glucosidase inhibitory potential, quercetin showed superior efficacy compared to acarbose, as demonstrated by 45 out of 65 in vitro studies,  $^{47,59,60,62,69,83,84,91,101-103,106,109,113,122,124,149-177}$  making it the most well-examined flavonoid against  $\alpha$ -glucosidase. It is worth mentioning that quercetin (68) was even used as a positive control in several studies. Otherwise, compared to the control drug in the same condition, this flavonol exhibited a weak  $\alpha$ -amylase inhibitory effect,  $^{33,39,54,125,126,147,150,166,168,173,178-181}$  with the pIC<sub>50</sub> being 4.07 (3.26-4.83). The mechanism underlying the inhibition of both enzymes remained controversial, which was documented to be in a mixed,  $^{149,157,180}$  non-competitive,  $^{150,163,174}$ , or even competitive manner.  $^{54,102,122,169,180}$ 

Kaempferol (69) is proven to be a strong  $\alpha$ -glucosidase inhibitor, supported by at least 22 studies.  $^{47,60,69,91,94,96,102,106,109,149,152,154,158,162,167,168,177,182-186}$  Meanwhile, the subtle inhibitory ability against  $\alpha$ -amylase promised antidiabetic efficacy while minimizing the adverse effects of kaempferol compared to acarbose. Concerning the inhibitory mechanism, kaempferol was reported to suppress  $\alpha$ -glucosidase via a mixed-type manner while inhibiting  $\alpha$ -amylase via a non-competitive mechanism.  $^{149,150}$  However, those were only two pieces of research evaluating the inhibitory mechanism of kaempferol, which needed further investigation to confirm.

Myricetin (70) displayed similar inhibitory features to those of the previously mentioned aglycones quercetin (68) and kaempferol (69). As one of the major flavonols found in nature, myricetin is also well-known for its ability to inhibit α-glucosidase.  $^{57,60,69,73,83,149,151,161,173,177,179,187-191}$  The IC<sub>50</sub> value on α-glucosidase of myricetin (70) ranged from 0.52 to 3000 μM, as reported in 19 included papers, in which 15 research revealed superior inhibitory effect compared to acarbose in the same experimental condition.  $^{57,60,69,73,83,149,151,161,173,177,179,187-191}$  Additionally, while α-glucosidase inhibitory mechanisms were discordantly suggested, the inhibition of α-amylase was stated competitively. Notably, myricetin (70) is the only flavonoid in our curated database whose inhibitory mechanism against α-amylase has been proved via X-ray crystallography.  $^{192}$ 

A notable finding is that the  $\alpha$ -glucosidase inhibitory activities of quercetin (68), kaempferol (69), and myricetin (70) increase with the number of hydroxyl groups attached to the B ring, as kaempferol (69) < quercetin (68) < myricetin (70). 35, 44, 49, 126, 153, 172, 182, 190 This notion was consistently discussed in multiple articles, and the probable reason was due to their different capabilities to form additional hydrogen bonds with the amino acids residues in the active pocket of  $\alpha$ -glucosidase. 49, 126, 190

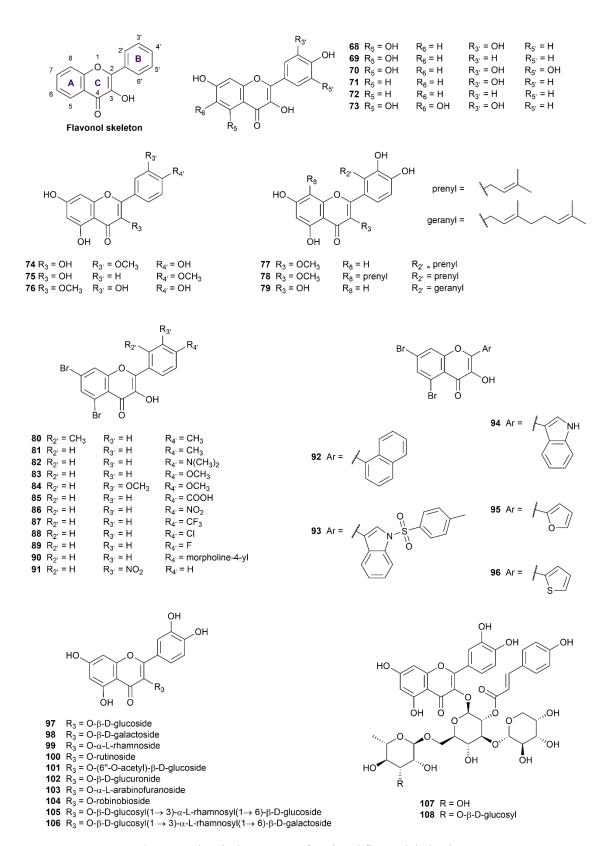


Figure 7. Chemical structures of retrieved flavonol derivatives.

$$\begin{array}{c} \text{HO} \\ \text{OH} \\$$

**Figure 7.** Chemical structures of retrieved flavonol derivatives (cont.)

Fisetin (71) and 3,7,4'-trihydroxyflavone (72) are other natural flavonol aglycones that lack the hydroxyl group at the C<sub>5</sub>-position of the A-ring present in compound **68-70** (Figure 7). Fisetin (71), also known as 7,3',4'-flavon-3-ol, exhibited a better inhibitory potency against  $\alpha$ -glucosidase than acarbose in two studies by Yue *et al.* (2018) and Liu *et al.* (2022),<sup>206, 222</sup> whereas one paper by Jia *et al.* (2019) stated the contrary.<sup>73</sup> Only one included research explored fisetin (71) for its  $\alpha$ -amylase inhibitory potential, which was found to be 20 times weaker than acarbose. Therefore, its dual inhibitory ability is prospective, yet further investigation should be conducted to confirm it. Compound 72, 3,7,4'-trihydroxyflavone was reported to possess inhibitory activity against  $\alpha$ -glucosidase, being six times more potent than acarbose. This compound also poorly inhibited  $\alpha$ - amylase in a competitive manner. However, these claims were only supported by one study for each enzyme. Fisetin (71) and 3,7,4'-trihydroxyflavone (72) seemed to have an inferior  $\alpha$ -glucosidase inhibitory effect than compound **68-70** as a result of the removal of -OH group from the 5-position, which weakened the interaction between flavonoid-enzyme.

Quercetagetin (73), having the additional  $C_6$ -OH in the A ring compared to quercetin (68), speculated a marked affinity to targeted enzymes. Oncerning  $\alpha$ -glucosidase inhibitory potential, two included studies showed opposed results in particular origin of tested enzymes. In the study by Wang *et al.* (2016)  $\alpha$  using *Saccharomyces cerevisiae* are glucosidase, quercetagetin (73) exhibited competence as 4.5 times greater effectiveness than acarbose. Meanwhile, this compound displayed 13 times weaker inhibition towards human sucrase when compared to the positive control. In terms of  $\alpha$ -amylase inhibition, all three curated papers confirmed the weak inhibitory potent of quercetagetin (73). In terms of  $\alpha$ -amylase inhibition, all three curated papers confirmed the weak inhibitory potent of quercetagetin (73).

Isorhamnetin (74) is also known as 3'-methoxy quercetin, a methoxyflavonol found in *Ginkgo biloba* leaf extract. As reported in 10 studies, isorhamnetin (74) demonstrated simultaneous inhibition against  $\alpha$ -glucosidase and  $\alpha$ -amylase. In comparison with acarbose, relevant research revealed isorhamnetin (74) with a superior pattern against both starch hydrolysis enzymes.<sup>38, 126, 173, 175, 181, 206, 210</sup>

Kaempferide (75), the 4'-O-methyl derivative of kaempferol, has been investigated in five relevant studies for its inhibitory activity against α-glucosidase. <sup>49, 53, 67, 89, 123</sup> According to these studies, kaempferide (75) was 0.8 to 20 times more potent than acarbose. <sup>67, 89, 123</sup> However, a study by Tian *et al.* (2021) presented opposite results, claiming that kaempferide (75) was ten times weaker than acarbose. <sup>39</sup> This research also reported the α-amylase inhibitory activity

of kaempferide (75) to be 15 times inferior to the standard drug. Besides, it is worth mentioning that the methoxylation of  $C_4$  weakened the  $\alpha$ -glucosidase inhibitory potency compared to its unmethoxylated form (kaempferol, 69).

Quercetin-3-methyl ether (76) was found in five studies,  $^{49, 53, 67, 89, 123}$  out of which four reported weaker  $\alpha$ -glucosidase inhibition compared to acarbose, with IC<sub>50</sub> values ranging from 10.41 to 120.23  $\mu$ M. Furthermore, compound 76 exhibited nine times weaker inhibition towards  $\alpha$ -amylase as acarbose in the same experimental condition.  $^{39}$ 

As a 2'-prenyl derivative of compound **76**, podoverine A **(77)** could inhibit both enzymes of interest, yet two times weaker than acarbose, reported in one study by Van Thanh *et al.* (2022).<sup>89</sup> This study also extracted and evaluated the digestive enzyme inhibitory effects of 8,2'-diprenylquercetin 3-methyl ether **(78)**, and it was found to be relatively good as acarbose.<sup>89</sup> Solophenol D **(79)** is a 2'-geranyl derivative of quercetin. In a study by Uddin *et al.* (2022),<sup>59</sup> this compound displayed significant inhibitory effects against  $\alpha$ -glucosidase and  $\alpha$ -amylase, respectively, six times and four times more effective than acarbose.

As for the structure-activity relationship of compounds **74-79**, the majority of studies revealed decreased inhibitory activities against  $\alpha$ -glucosidase of methoxyflavonols as compared to unmethoxylated forms. <sup>67, 123, 126, 130, 206</sup> On the other hand, some studies have argued that this phenomenon depended on the position of the substitutions. <sup>49, 89, 192</sup> While the impact of the methoxylation remains disputable, the prenylation of the flavonols appeared to increase the inhibitory potential against  $\alpha$ -glucosidase. <sup>107, 204</sup>

Compounds 80-96 are synthetic 5,7-dibromoflavonol derivatives reported by Ashraf *et al* in 2020.<sup>202</sup> Although achieving impressive  $IC_{50}$  values against both enzymes, these compounds are less potent than the standard drug acarbose, except for compounds 93 and 94, which had the phenyl ring B replaced with the indole ring.

Next is a series of flavonoid glycosides (97-115), each possessing a sugar residue at  $C_3$ . Several SAR studies have shown that the attachment of sugar moieties diminished the  $\alpha$ -glucosidase inhibitory effects of the flavonoids. <sup>35, 38, 44, 49, 67, 101, 126, 153, 172, 182, 190, 206</sup> Moreover, it was suggested that the 3-hydroxyl group in the C ring was essential for proper binding orientation. <sup>109</sup>

Compounds **97-108** are natural *O*-glycoside quercetin derivatives, as displayed in **Figure 7.** Isoquercetin **(97)** or quercetin-3- $\beta$ -D-glucoside has been investigated in 26 research articles, <sup>38, 44, 57, 58, 83, 102, 126, 148, 174, 179, 188, 197-199, 203, 206, 209, 210, 224-233 making it the most extensively studied flavonol glycoside. The majority of included studies have proved the  $\alpha$ -glucosidase inhibitory potential of isoquercetin **(97)** to be better than the standard drug (up to 32 more potent than acarbose). <sup>47</sup> However, there was insufficient data on either the modes of inhibition or  $\alpha$ -amylase inhibitory activity. Hyperoside **(98,** hyperin, or quercetin-3- $\beta$ -D-galactoside) had similar patterns as **97** but was slightly inferior in disaccharide hydrolysis, demonstrated in 13 curated studies. <sup>32, 44, 57, 58, 81, 172, 195, 201, 206, 209, 224, 234, 235</sup></sup>

Quercitrin (99), also known as quercetin-3-rhamnoside, appeared to potently inhibit  $\alpha$ -glucosidase in a mixed manner, <sup>218</sup> noticeably better than acarbose in more than 12 studies. <sup>38, 55, 58, 78, 83, 102, 133, 140, 188, 217, 237, 238</sup> However, only one research examined quercitrin as an  $\alpha$ -amylase inhibitor, which showed quercitrin as roughly 30 times stronger than acarbose. <sup>47</sup> Following this, quercetin-3-rutinoside or rutin (100) had excellent  $\alpha$ -glucosidase inhibitory activity while marginally inhibiting  $\alpha$ -amylase, which could be a promising antidiabetic agent.

Quercetin-3-O-(6"-O-acetyl)-β-D-glucopyranoside (**101**) and quercetin-3-O-glucuronide (**102**) are moderate α-glucosidase and α-amylase inhibitors (comparing to acarbose). A vicularin (**103**) or quercetin-3-O-α-L-arabinofuranoside displayed similar features as **101** and **102** in most studies 58, 172, 191, 197, 206, 217, 239 yet showed strong inhibitory ability towards α-glucosidase in a report by Wang *et al.* (2018). 175

Flavonol glycosides 104-106 are quercetin glycosides isolated from Lu'an GuaPian tea by Fang *et al.* in 2018. <sup>168</sup> These compounds strongly inhibited  $\alpha$ -glucosidase (three to five times better than acarbose) and  $\alpha$ -amylase in the assays. In the same study, compounds 107-108 which are acylated quercetin glycoside derivatives were also reported for their dual target inhibition. These compounds modestly inhibited  $\alpha$ -amylase and showed superior  $\alpha$ -glucosidase inhibitory activities to acarbose.

Compounds 109-115 are kaempferol glycoside derivatives, in which the sugar moieties were attached at the OH 3-position. These derivatives demonstrated moderate to considerable inhibition against  $\alpha$ -glucosidase in comparison

with acarbose. The included studies also revealed that compounds 111-113 were weak to moderate α-amylase inhibitors. Kaempferol-3-*O*-β-D-glucopyranoside, also known as astragalin (109), potently inhibited α-glucosidase as potent as acarbose, proven by at least eight studies. <sup>124, 148, 174, 189, 195, 199, 226, 230</sup> Two out of three studies examined the α-amylase inhibition of 109 as three times weaker than the control agent. <sup>195, 233</sup> Nicotiflorin (110) is a 3-rutinoside derivative, exhibiting controversial results in either inhibitory potential or inhibition mechanisms. Concerning α-glucosidase inhibition, nicotiflorin appeared to have the IC<sub>50</sub> ratio of compound-acarbose ranging from 0.1 to 8, supported in 12 separate studies. <sup>38, 44, 168, 171, 174, 182, 195, 208, 226, 230, 241, 244</sup> Only three papers examined the α-amylase inhibitory potential of 110, revealing 0.32, 6, and 30 times as compared to that of acarbose. <sup>38, 195, 245</sup> Compounds 111-115, which are kaempferol glycosides extracted from Lu'an GuaPian tea, share the same sugar residue as quercetin glycosides 104-108, respectively. <sup>168</sup> Compounds 111-113 had weaker inhibitory activities towards two enzymes of interest than the corresponding quercetin glycosides but were still superior to acarbose. Conversely, compounds 114-115 were slightly better than their respective quercetin derivatives in the abilities to inhibit α-glucosidase and α-amylase. <sup>168</sup>

The remaining compounds (116-121) are glycosides of other flavonols (myricetin, europentin, isorhamnetin, and quercetagetin). Most included papers in our database revealed that myricetin-3-rhamnoside (116, myricitrin) exhibited weaker inhibitory potent against two starch-hydrolyzing enzymes than acarbose and their corresponding aglycones myricetin (70).<sup>35, 44, 202, 218</sup> The IC<sub>50</sub> values against α-amylase of compound 116 lacked compromised results, which required more biological assays.<sup>35, 44, 78, 186, 202, 206, 218, 235, 247</sup>

On the contrary, europetin-3-O-rhamnoside (117), having an additional methyl group at position C-7, potently inhibited  $\alpha$ -amylase eight times more effective than the standard drug. This compound was also 20 times more potent than acarbose for  $\alpha$ -glucosidase inhibition.<sup>161</sup>

Isorhamnetin-3-O-glucoside (118) and isorhamnetin-7-O-β-D-glucopyranuronide (119) have displayed weaker inhibition of α-amylase and α-glucosidase than the positive control in *in vitro* studies.<sup>53, 117, 199</sup> Conversely, isorhamnetin-3-O-rutinoside (120) has shown IC<sub>50</sub> values two to five times less than acarbose,<sup>38, 249, 250</sup> indicating its superiority over other isorhamnetin glycoside derivatives (118, 119).

The last flavonol glycoside, quercetagetin-7-O- $\beta$ -D-glucopyranoside (121), marginally inhibited  $\alpha$ -glucosidase in a mixed mechanism and had little effect on the  $\alpha$ -amylase activity mentioned in a report.<sup>39</sup>

## 3.3.4. Anthocyanidins and anthocyanins

Anthocyanidins and their glycoside forms (anthocyanins) are water-soluble natural pigments and are responsible for the characteristic color of many fruits and flowers. The general skeleton of anthocyanidins is the flavylium ion, with the oxygen atom in the C ring being cationized (**Figure 8**). The color of anthocyanidins depends on the pH of the solution. Some anthocyanidins appear red in low pH conditions (acidic) and turn blue in higher pH conditions. In this review, three anthocyanidins (122-124) and four anthocyanins (125-128) are included and will be introduced briefly for their activity against the two enzymes of interest (**Table S11**).

Pelargonidin (122) was involved in at least two reports for its anti-α-glucosidase and α-amylase capabilities.  $^{142, 159}$  The current evidence shows that pelargonidin could be a potential α-glucosidase and α-amylase inhibitor with the IC<sub>50</sub> being 0.18 μM and 2.07 μM, respectively.  $^{138}$  However, in a study reported by Xiao *et al.*,  $^{126}$  a different inhibitory pattern was observed, with the IC<sub>50</sub> of this compound against α-amylase being 459.15 μM. Such conflict could stem from the difference in the method used in the *in vitro* experiments. Cyanidin (126) is a well-known anthocyanidin derived from red berries and was involved in at least four studies.  $^{142, 252-254}$  Regarding the α-glucosidase inhibition, this compound was able to inhibit the enzyme far better than acarbose in the same condition, and the mechanism was reported to be in a non-competitive pattern.  $^{228-230}$  In terms of α-amylase inhibition, the compound was regarded as a weak inhibitor of this enzyme. However, the retrieved IC<sub>50</sub> values varied significantly between studies.  $^{142, 253, 254}$  Delphinidin (124) was able to inhibit 10 to 14 times as well as acarbose in two studies against the α-glucosidase.  $^{159}$ ,  $^{255}$  The figure was 16 times when it comes to the α-amylase inhibition.  $^{138}$ 

Figure 8. Chemical structures of retrieved anthocyanidin derivatives.

As for the anthocyanins, three cyanidin derivatives (125-127) and a malvidin derivative (128) were included. Some studies suggest that the introduction of sugar moiety may reduce the inhibitory activity toward both enzymes. Although the reported IC<sub>50</sub> values of cyanidin-3-O-glucoside (125) and cyanidin-3-O-rutinoside (127) toward the two enzymes was still in the micromolar range, the activity of these compounds was weaker than that of acarbose in the same experimental conditions. <sup>35, 105, 235, 253, 254, 256, 257</sup> In a study reported by Ho *et al.*, <sup>230</sup> the introduction of sambubioside (126), however, increases dramatically the inhibitory activity of this anthocyanin against  $\alpha$ -glucosidase and  $\alpha$ -amylase, with the respective IC<sub>50</sub> values being 2.80  $\mu$ M and 2.30  $\mu$ M. In addition, malvidin-3-O-glucoside (128) was also reported to exhibit stronger inhibition toward  $\alpha$ -glucosidase while the inhibitory activity of this compound was weaker toward  $\alpha$ -amylase, compared to acarbose. <sup>232-234</sup>

## 3.3.5. Aurone

Regarding the aurone structure, although four studies have been included in our database, we are only able to retrieve one aurone which has concurrent data against the two enzymes of interest (**Figure 9**). Compound **14a** ((*Z*)-6-(2-benzylidene-4,6-dihydroxy-3-oxo-2,3-dihydrobenzofuran-7-yl)-7-methoxy-2*H*-chromen-2-one, numbered **129**) is a semi-synthesized aurone-coumarin hybrid reported by Sun *et al.*<sup>142</sup> This aurone was able to inhibit  $\alpha$ -glucosidase and  $\alpha$ -amylase with the respective IC<sub>50</sub> values being 3.55 and 10.97  $\mu$ M. The respective values for acarbose in the same condition were reported to be 224.7  $\mu$ M and 2.72  $\mu$ M, indicating that compound **129** is a potentially strong  $\alpha$ -glucosidase inhibitor while exhibiting moderate inhibition toward  $\alpha$ -amylase.

## 3.3.6. Chalcones

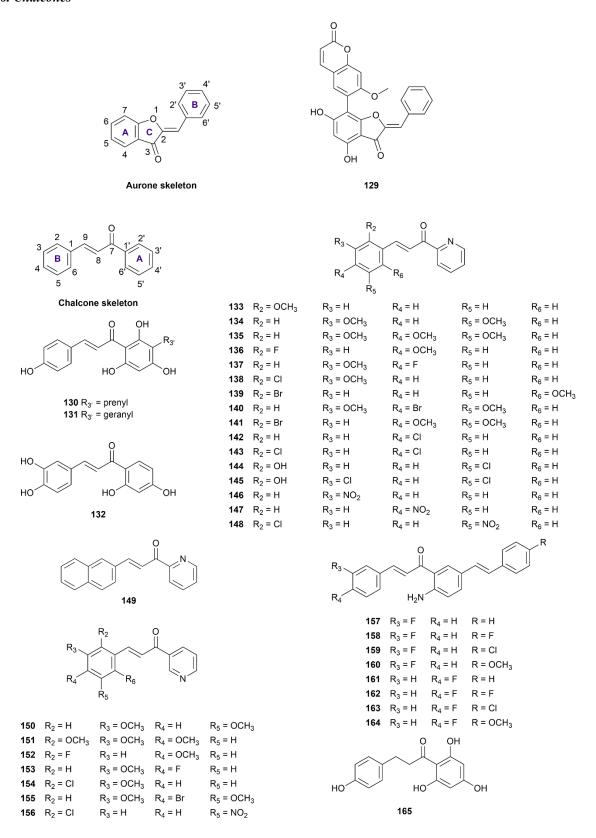


Figure 9. Chemical structures of retrieved aurone and chalcone derivatives.

Chalcones share a common scaffold of 1,3-diaryl-2-propen-1-one, which exists in either *trans* (*E*) or *cis* (*Z*) isomers, with the *trans* (*E*) isomers being more stable and popular (**Figure 9**).<sup>235</sup> These compounds are both biosynthesized in plant tissue or total-synthesized using different approaches, with Claisen-Schmidt condensation being the most famous classical approach.<sup>235,236</sup> In this review, besides the traditional chalcone structure, multiple chalcone mimics (**133-156**), in which the A ring and/or the B ring are substituted by other aromatic rings, are also included. Moreover, one naturally occurring dihydrochalcone called phloretin (**165**) is also included in the study. **Table S12** summarizes the main findings related to the potency of aurone and chalcone derivatives against both enzymes.

Desmethylxanthohumol (130) and 3'-geranylchalconaringenin (131) are two respective prenyl and geranyl derivatives of chalconaringenin which were isolated from hops (*Humulus lupulus*). In a study reported by Sun *et al.* (2018),<sup>65</sup> although chalconaringenin could not achieve a specific IC<sub>50</sub> value toward α-amylase even at the concentration of 100 μM, the introduction of either prenyl or geranyl substituent significantly increased the inhibitory activity of this compound indicating the importance of these groups toward the inhibition manner of this chalcones. Moreover, his study also showed that the introduction of the geranyl group (131) increased the inhibitory potency by 20 times and 4 times, compared with desmethylxanthohumol (123), against α-glucosidase and α-amylase, respectively. As an effort to evaluate the anti-diabetic activities of the chalcone derivatives, 28 chalcones and 13 chalcone analogs have been synthesized and assessed for their IC<sub>50</sub> value toward the two enzymes by Rocha *et al.* (2019).<sup>237</sup> Butein (132) was the only and the most active inhibitor which had sufficient IC<sub>50</sub> data for the two enzymes of interest.

Compounds 133-156 were 24 azachalcone derivatives synthesized by Salem *et al.* which show inhibition against both glucosidase and amylase.<sup>238</sup> In this study, the authors divided the synthesized chalcones into two categories namely categories A and B. The category "A" consists of the chalcone derivatives in which the A ring is a 2-pyridyl aromatic ring (compound 133-149). This ring is the 3-pyridyl ring, when it comes to the "B" category (compound 150-156) and these two categories are cordially playing a vital role in demonstrating the inhibition strength against both enzymes. However, the different substitutions at ring B are accountable for varying these activities. Among the synthesized compounds, compounds 133, 135, 136, 146, and 156 demonstrated excellent inhibition against both enzymes. However, in comparison with acarbose in the same experiment condition, these chalcone derivatives did not perform as well as the standard drug (IC<sub>50</sub> values of 18.67 μM and 18.08 μM for α-glucosidase and α-amylase, correspondingly).

Compounds 157-164 are the seven 5-styryl-2-aminochalcone derivatives that were synthesized and reported by Mphahlele *et al* in 2021.<sup>239</sup> These chalcone hybrids were able to inhibit both digestive hydrolysis enzymes with impressive IC<sub>50</sub> values, ranging from 5.1 to 19.2  $\mu$ M and 1.6 to 15.6  $\mu$ M for  $\alpha$ -glucosidase and  $\alpha$ -amylase, respectively. Although having achieved such low IC<sub>50</sub> values, these compounds were not able to surpass the inhibitory activity of acarbose in the same experiment settings, giving the IC<sub>50</sub> of acarbose against  $\alpha$ -glucosidase and  $\alpha$ -amylase being 0.95  $\mu$ M and 1.03  $\mu$ M, respectively.

Phloretin (**165**) is a dihydrochalcone retrieved from either natural sources <sup>83, 186</sup> or a fully synthetic approach.<sup>35, 157</sup> This is the only dihydrochalcone that comes with sufficient data regarding its inhibitory activity against both enzymes of interest. However, the current evidence differs on whether this flavonoid can inhibit the two enzymes better than the standard drug acarbose or not. Thus, further validations are needed to confirm the potency of this compound.

## 3.3.7. Isoflavonoid

Figure 10. Chemical structures of retrieved isoflavonoid derivatives.

Isoflavonoid is a subclass of flavonoid in which the B ring is attached to the  $C_3$  position instead of  $C_2$ , forming the phenyl-3-chroman skeleton (**Figure 10**). These structures are widely regarded as phytoestrogens due to their ability to bind to estrogen receptors and mimic the activity of  $17\beta$ -estradiol (E2).<sup>240</sup> In our current review, four isoflavonoid derivatives were obtained and showed concurrent inhibitory activity against  $\alpha$ -glucosidase and  $\alpha$ -amylase, with genistein (166) being the most well-studied isoflavonoid (**Table S13**).

Genistein (166) was found to be more potent than acarbose against  $\alpha$ -glucosidase in seven out of eight studies with the inhibiting mechanism being non-competitive, mixed, or uncompetitive type.  $^{35,67,159,167,205,265-267}$  The inconsistency of the inhibiting mechanism could result from the difference in the  $\alpha$ -glucosidase type used in these studies. Additionally, this isoflavonoid was also reported to be more potent than acarbose in the  $\alpha$ -amylase inhibition assay in two references.  $^{159,266}$  On the other hand, the inhibitory potency of formononetin (167) towards  $\alpha$ -glucosidase, is still controversial, with two out of three studies reporting it to be more effective than acarbose with micromolar potency  $^{159,268}$  while the result from Jia *et al.* showed a reverse pattern. Nevertheless, this isoflavonoid exhibited outstanding inhibitory activity against  $\alpha$ -amylase, in comparison with acarbose.  $^{138}$ 

In a study by Ha *et al.* (2018),<sup>242</sup> dalbergioidin (**168**) was extracted from *Desmodium heterophyllum* aerial parts and evaluated for its anti-diabetic properties. According to the results, this substance was found to exhibit stronger inhibition against  $\alpha$ -glucosidase but weaker inhibition against  $\alpha$ -amylase, in comparison with acarbose in the same experimental conditions.<sup>242</sup> Puerarin (**169**) is the only isoflavonoid glycoside included in our current review. The inhibitory activity of puerarin towards  $\alpha$ -glucosidase was found to be comparable to that of acarbose, with the IC<sub>50</sub> values ranging from 75.60 to 524.08  $\mu$ M <sup>61, 89, 188, 267, 269</sup> while its inhibitory activity against  $\alpha$ -amylase was reported to be roughly 74 times lower than that of acarbose, reported in a study by Zhang *et al.*<sup>119</sup>

## 3.3.8. Oligomeric flavonoids

Oligomeric flavonoids or specifically biflavonoids have gained wide attention recently due to their health promotion effects and fascinating flavors with proanthocyanidin being the largest class of flavonoid polymer and found in various appealing-colored fruits and plants. Three subgroups of proanthocyanidin (i.e., procyanidin, prodelphinidin, propelargonidin) differing in their monomers have been identified, yet their simultaneous inhibition against  $\alpha$ -glucosidase and  $\alpha$ -amylase received disproportionated concerns. In the current review, a total of eleven articles and eight oligomeric flavonoids are included (**Figure 11**), with the most well-studied biflavonoid being amentoflavone (**171**). Six out of eight included structures are biflavonoids, demonstrating their potential as anti-diabetic therapeutic agents (**Table S14**).

Figure 11. Chemical structures of retrieved oligomeric flavonoids.

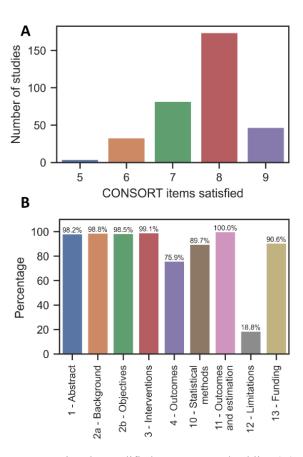
Kuwanon M (170) is the only biflavonoid extracted from the root bark of *Morus alba* L. which exhibited both α-glucosidase and α-amylase inhibition. <sup>82</sup> In his study, Zhao *et al.* reported the IC<sub>50</sub> values of kuwanon M against α-glucosidase and α-amylase as  $0.60 \pm 0.09$  μM and  $1.22 \pm 0.34$  μM, respectively. In the same condition, acarbose only achieved the respective IC<sub>50</sub> of 293.50 μM and 1.51 μM.

Amentoflavone (171) is the most well-studied biflavonoid included, which is formed by the condensation of two apigenin monomers via a  $C_3$ - $C_8$ . linkage. In the  $\alpha$ -glucosidase inhibition assay, amentoflavone exhibited a stronger inhibition against  $\alpha$ -glucosidase in 4 studies, with the IC<sub>50</sub> ranging from 3.28  $\mu$ M to 522.33  $\mu$ M.  $^{40,67,84,91,109,247}$  Regarding the  $\alpha$ -amylase inhibition, although achieving outstanding IC<sub>50</sub> values below 100  $\mu$ M, this compound is 2 times or 4 times less potent than the commercially available drug acarbose, reported by two publications.  $^{263, 264}$  The results on 2,3-dihydroamentoflavone (165) also suggest a similar pattern.  $^{263}$ 

In a study by Ho *et al.*,  $^{230}$  the inhibitory activity of procyanidin C1 (173), procyanidin B2 (174), and procyanidin B5 (175) were also reported. In this study, all these three oligomeric flavonoids outperformed acarbose in the both two  $\alpha$ -glucosidase and  $\alpha$ -amylase inhibition assay, with the IC<sub>50</sub> values varied below 10  $\mu$ M. These results indicate that proanthocyanidins could be promising effective and safe therapeutics in the future.

Lastly, in a report by Josim *et al.* (2022), <sup>137</sup> the anti- $\alpha$ -glucosidase and anti- $\alpha$ -amylase properties of proanthocyanidins from *Ceriscoides campanulata* were documented. In this study, compound **176** and procyanidin A1 (**177**) could inhibit  $\alpha$ -glucosidase more effectively than acarbose (IC<sub>50</sub> of 4.6  $\mu$ M, 6.2  $\mu$ M and 665  $\mu$ M, respectively). The results on  $\alpha$ -amylase inhibition suggested that compound **176** was also active against  $\alpha$ -amylase with an impressive IC<sub>50</sub> value (3.50  $\mu$ M compared to 5.9  $\mu$ M of acarbose). It could be hypothesized that the higher degree of oligomerization allowed this compound to bind and inhibit  $\alpha$ -amylase more effectively, yet further evaluation is needed.

#### 3.4. Quality assessment



**Figure 12.** Result of quality assessment using the modified CONSORT checklist. (A) Overall scores of the included studies and (B) the percentage of studies that satisfied each item in the checklist.

In this study, we employed a modified CONSORT checklist of Faggion *et al.* (2012)<sup>28</sup> for reporting *in vitro* studies. The checklist comprises 14 items, which are divided into six sections (Abstract, Introduction, Methods, Results, Discussion, and Other Information). However, due to the lack of information about sample size, randomization, blinding, and research protocol in most of the *in vitro* studies and the unnecessity of these items in the non-cellular enzymatic assay, we only include items 1-4, 10-13 for quality assessment in our review.<sup>248</sup> The detailed checklist and quality evaluation of included studies are available in **Table S5**, with the overall characteristics being described in **Figure 12**.

As can be seen from **Figure 12**, all the studies satisfied at least five CONSORT items, with most of them could achieve up to eight over nine items. Specifically, it is understandable that most of the studies fulfilled items related to results presentation (Item 11) and provide adequate information for experiment reproduction (items 3, 4, 10). Nevertheless, only 75.9% of the studies indicated how the IC<sub>50</sub> values were determined and 89.7% of the studies represented IC<sub>50</sub> values using statistical methods. Moreover, only 18.8% of studies discussed their limitations, yet up to 90.6% of the studies provided information on the funding. Overall, these figures suggest that the included articles are of moderate to high quality and could be used in our systematic review.

#### 4. Discussions

## 4.1. Structure-activity relationship analysis

## 4.1.1. SAR and mechanism of α-glucosidase inhibition

The abundance of evidence allowed us to outline and summarize the main characteristics required for the inhibition of flavonoids against  $\alpha$ -glucosidase (**Figure 13**). We observed that the decreasing order of the inhibitory activities of flavonoid subgroups against  $\alpha$ -glucosidase could be demonstrated as (1) flavonol > flavono > flavanonol > flavanol > flavanoroids > flavonoids > flavonoid glycoside; and (3) the inhibitory potency of flavonoids is stronger than their corresponding chalcones or isoflavonoids. Nevertheless, the effect of substituents and sugar moieties should also be taken into account and a case-by-case comparison should be conducted to confirm these observations.

Concerning the flavonoid scaffold, generally, the chemical features that are favorable for the inhibition of flavonoids are (1) the presence of  $C_2=C_3$ ; (2) the hydroxyl group at the  $C_3$  position; and (3) the ketone group at the  $C_4$  position. These features help form a large conjugation  $\pi$ -system between the B-ring and the benzopyran system (A and C ring), thereby inducing the near-planarity structure and enhancing the activity of the flavonoids. Furthermore, the number and the position of hydroxyl groups attached to the flavonoids also highly affect the activity of flavonoids. An increase in the number of hydroxyl groups in either ring A (5,6,7,8-positions) and B (3',4',5'-positions) seems to be beneficial for the inhibitory activity and the replacement of these hydroxyl groups by alkyl or glycosyl groups generally decreases the activity of the flavonoids. However, the influence of hydroxyl groups also depends on the substitution position.

As regards the B-ring, compounds with o-dihydroxyl groups (catechol moiety) are more active than those with m-dihydroxyl groups (resorcinol moiety), and the addition of a third adjacent -OH group to form a pyrogallol moiety would significantly increase the activity of the flavonoids. Furthermore, the introduction of several highly polar functional groups such as -NO<sub>2</sub>, -NH-SO<sub>2</sub>-, -N(CH<sub>3</sub>)<sub>2</sub>, or -COOH at the 3' or 4'-positions of the B-ring also yields stronger inhibitors while the introduction of hydrophobic or halogen atoms such as -CH<sub>3</sub>, -CF<sub>3</sub>, -Cl, -F results in a decrease in activity. Interestingly, although the free 4'-OH group is considered an important feature for  $\alpha$ -glucosidase inhibition, several studies showed that the esterification of this OH group with the carboxylic group of p-hydroxybenzoic acid and the etherification of this OH group with a polyethylene glycol chain (up to dimer) or N-tosyl group seems to enhance the activity of the flavonoids.

As for the A ring, hydroxyl groups at the C<sub>5</sub> or C<sub>7</sub> position are abundantly presented in nature and usually listed as important features for α-glucosidase inhibition, but the alkylation of these hydroxyl groups with the alkyl chains (2-7 carbons) seems to benefit the activity while the methylation possibly exerts the opposite. Additionally, the introduction of the prenyl group and geranyl group at several positions is beneficial for the activity. Relatively, the induced effect of the geranyl group on the flavonoids' activities is stronger than that of the prenyl group and the substitution at the 6-position affords a stronger inhibitor than that at the 8-position. Several attempts to attach the halogen atoms to the flavonoid scaffold have been reported, with the substitution of bromine atoms at 6,8,5',3'-positions and the

substitution of chlorine atoms at 3,6,8-positions resulting in strong inhibitors. Surprisingly, the replacement of the hydroxyl group at  $C_3$  with the chlorine atom does not significantly affect the inhibitory activity of the flavonoids. Lastly, the attachment of the hydroxybenzyl moiety to the 6,8,3'-positions and the formation of the fourth  $\delta$ -lactone ring at  $C_7$  and  $C_8$  also enhance the inhibitory effect.

As previously mentioned, the glycosylation of flavonoids appears not to favor the inhibitory capability of the flavonoid. Several hypotheses for this phenomenon have been proposed, including the increase of steric hindrance, or an increase in molecular size, polarity, and the presence of a non-planar structure, all of which could affect the binding capability of the flavonoids. The impact of different sugar molecules on flavonoid glycosides is not well understood, and it may vary depending on the compound. Nevertheless, several naturally occurring substituents on sugar moiety that may recover the activity of flavonoid glycosides have been reported, including the oxidation from glucose to form glucuronic acid, or the addition of the galloyl units, *p*-caffeoyl, *p*-coumarin or acetyl groups at 2",3", or 6"-positions of the sugar part.

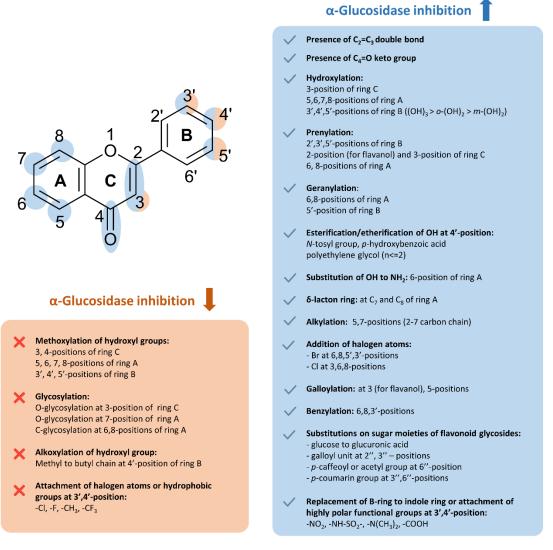


Figure 13. Structure-activity relationship of flavonoids against  $\alpha$ -glucosidase

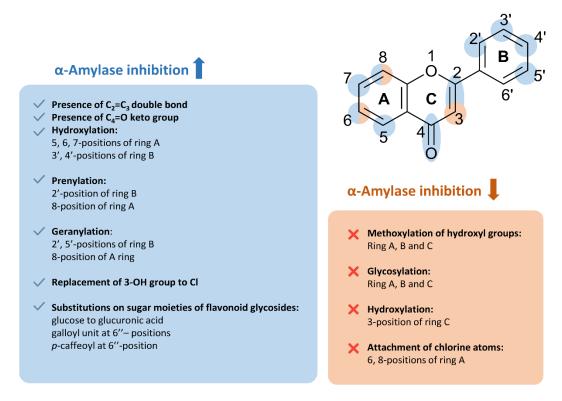
The inhibitory mechanism of flavonoids towards  $\alpha$ -glucosidase is a subject of ongoing debate. Although utilizing the same enzyme and substrate, different inhibitory mechanisms have been reported for the same flavonoid molecule, such as epicatechin (4), luteolin (30), quercetin (68), isoquercetin (105), and genistein (166). These discrepancies

could be due to variations in the assay protocols, such as differences in the concentration of enzyme and substrate, and incubation time used by various research groups. After the analysis of numerous studies, there is still no consensus on the inhibitory mechanism of flavonoids against  $\alpha$ -glucosidases. Flavonoids could inhibit  $\alpha$ -glucosidase competitively or non-competitively or even only when the enzyme-substrate complex has been formed (uncompetitively). Thus, a case-by-case evaluation is needed to confirm the inhibitory mechanism.

It is worth mentioning that despite nearly four decades of research on the anti- $\alpha$ -glucosidase activity of flavonoids (since 1986 – the article is not included due to the absence of acarbose), no co-crystallized structure of flavonoids- $\alpha$ -glucosidase from either origin has been reported to the date the manuscript was written. While molecular modeling techniques, such as homology modeling, molecular docking, and molecular dynamics simulation, have been utilized to overcome this limitation, experimental three-dimensional structures are still needed to gain insights into how flavonoids inhibit  $\alpha$ -glucosidase at an atomistic level.

# 4.1.2. SAR and mechanism of α-amylase inhibition

The evidence supporting the inhibition of flavonoids against  $\alpha$ -amylase is not as abundant as for  $\alpha$ -glucosidase, but it is still sufficient to establish a preliminary SAR for flavonoids against this digestive enzyme (**Figure 14**). In general, the activity of aglycone flavonoids is stronger than their corresponding glycosides and isoflavonoids.



**Figure 14.** Structure-activity relationship of flavonoids against  $\alpha$ -amylase

Several common features have been considered favorable for the inhibition against  $\alpha$ -amylase, including (1) the double bond of  $C_2=C_3$ ; (2) the keto group at the  $C_4$ -position; and (3) hydroxyl groups at 5,6,7,3',4'-positions. Generally, the higher number of hydroxyl groups leads to a higher activity of flavonoids, yet the position at which the hydroxyl groups are introduced is also important. In fact, the hydroxyl group at  $C_3$ -position, which is crucial for the inhibitory activity against  $\alpha$ -amylase and the replacement of this OH-group with a chlorine atom may increase the latter activity of the flavonoids. Similar to  $\alpha$ -glucosidase inhibition, the catechol moiety at the B-ring is responsible for the strong  $\alpha$ -amylase inhibitory activity while the addition of another adjacent OH-group at  $C_5$  only enhances the effect modestly. Prenylation and geranylation at several specific

positions also similarly enhance inhibitory activity. It is observed that the induced effect of the geranyl group is stronger than that of the prenyl group at the same position.

With respect to the reducing factors, methylation or glycosylation of hydroxyl groups is also found to weaken the inhibitory effects. The effect is more pronounced when it comes to crucial positions such as  $C_3$ ,  $C_4$ , and  $C_7$ . Nonetheless, the recovery in the activity of flavonoid glycosides whose sugar moiety has been substituted with several polyphenol groups is also recorded. In addition, we also observe that the presence of chlorine substituent at either 6 or 8-positions does not favor the anti- $\alpha$ -amylase activity.

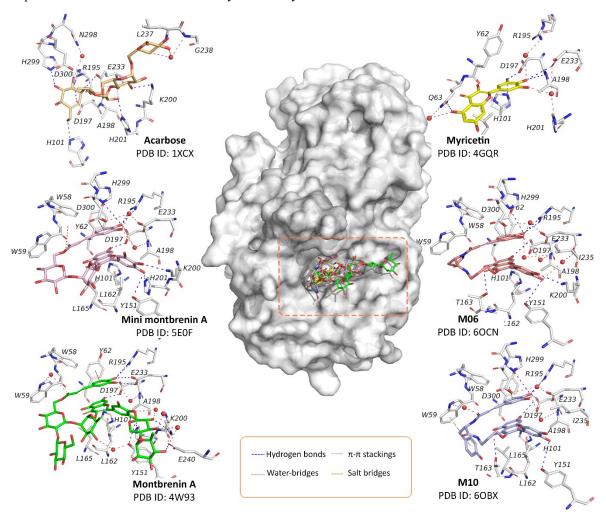


Figure 15. Interactions between the flavonoid derivatives with human pancreatic α-amylases retrieved from the Protein Data Bank as of April 2023. The small molecules with their corresponding colors and PDB IDs are:

Acarbose (orange – PDB ID: 1XCX);<sup>249</sup> Myricetin (yellow - PDB ID: 4GQR);<sup>192</sup> Montbretin A (Green - PDB ID: 4W93);<sup>250</sup> "Mini-montbretin A" (Pink - PDB ID: 5E0F);<sup>250</sup> M06 (Salmon - PDB ID: 6OCN);<sup>251</sup> and M10 (Blue - PDB ID: 6OBX).<sup>251</sup>

Similar to findings on  $\alpha$ -glucosidase, the inhibitory mechanism of flavonoids against  $\alpha$ -amylase exhibits a wide range of reported outcomes. Within our curated database, we have observed that flavonoids can inhibit  $\alpha$ -amylase competitively, non-competitively, or through a combination of both mechanisms. However, no instances of uncompetitive inhibition have been documented. It is important to acknowledge that although certain compounds show consistent mechanisms of inhibition, there are still discrepancies within specific compounds. These variations may arise due to variations in assay protocols and detection methods employed across different studies. However, the structural information from our search on the Protein Data Bank as of April 2023 suggests a more homogenous result,

leaning toward the competitive mechanism (**Figure 15**). These structures provide valuable insights into the binding site and protein-ligand interactions that underlie the mechanism of action of flavonoids, facilitating future rational drug design efforts. In our manuscript, we hence provide a concise discussion of these protein-ligand interactions and their potential implications for drug discovery.

As can be seen from Figure 15, all the available structures suggest that flavonoid derivatives occupied the same binding site of acarbose, which is also the active site responsible for the hydrolysis of starch.<sup>249</sup> However, upon taking a closer look at the interaction between the small molecules and the enzyme, there is a transition between the conformation of myricetin to other 3-O-glycosylated and 3-O-caffeate flavonoid derivatives. The myricetin-α-amylase complex shows that the B-ring headed toward the catalytic triad (Asp197, Glu233, and Asp300) and formed multiple hydrogen bonds and water-meditated bridges between the pyrogallol moiety and the residues in the cavity pocket. Additionally, the hydroxyl group at C<sub>5</sub> of the A-ring could form hydrogen bonds with the Gln63 residue, thereby stabilizing the ligand in the binding pocket. These observations confirm the SAR that the OH at C<sub>3</sub> is not necessary for the binding capability, but several features such as the hydroxyl groups at the B and the A-ring are required. However, in flavonoid glycosides and caffeate flavonoid derivatives, the aglycone part is oriented in the opposite direction, with the B-ring headed toward the solvent site and forming several hydrogen bonds with residues such as Tyr151 and Lys200. In this case, the hydroxyl group at the C7-position is responsible for the interaction with catalytic residues such as Asp197 and Glu233. The addition of the caffeate moiety (as in the case of M06 and M10) allowed these flavonoids to form more hydrogen bonds with the enzyme residues and additional  $\pi$ - $\pi$  contacts with the Tyr62, thereby inducing inhibitory activity. On the other hand, the addition of sugar moiety (as of montbrenin A and minimontbrenin A) did not significantly increase the contact with the enzyme residues, which is consistent with our SAR observations. In summary, it could be postulated that the binding conformation of flavonoid aglycones within the  $\alpha$ amylase pocket is similar to that of myricetin, while the opposite conformation of the aglycone part is expected for flavonoid glycosides or caffeate flavonoids. Even so, as mentioned previously, multiple mechanisms of flavonoid inhibition towards α-amylase have been reported. Therefore, further validation is necessary to confirm the binding patterns of new flavonoids with  $\alpha$ -amylase.

#### 4.2. Limitations

Our work has highlighted the concurrent  $\alpha$ -glucosidase and  $\alpha$ -amylase inhibitory effects of flavonoid compounds from 339 included studies, yet several limitations should be considered.

The inconsistent assay methods and enzyme origins are two limitations that could impact the interoperability of the data on  $\alpha$ -glucosidase inhibition. This can lead to variations in IC<sub>50</sub> values and inhibitory mechanisms being reported among studies, making it difficult to compare results. A lack of a common recommendation has hindered researchers in replicating experiments and verifying findings. In addition, it is interesting to note that although the ultimate therapeutic targets are human enzymes, yeast or rat proteins are often preferred in *in vitro* studies due to their availability and affordability. However, using yeast enzymes may not accurately reflect the inhibition of human MGAM and SI enzymes, which is an important consideration when designing experiments. Therefore, a consensual protocol is needed for *in vitro* assays using human-sourced enzymes to improve predictive capacity and reduce potential variations in results.

Recent efforts to develop quantitative methods for the inhibitory activity evaluation of flavonoids on human enzymes have been reported. For example, studies by Barber *et al.* (2021)<sup>194</sup> and Pyner *et al.* (2017),<sup>252</sup> employed human intestinal Caco-2 cells to extract human-sourced maltase and sucrase have been conducted. Additionally, although not being included in our literature database due to the absence of inhibitory data of acarbose, the study by Lim *et al.* in 2021<sup>253</sup> presents a promising model for future evaluation of the anti-α-glucosidase activity. In this study, the researchers utilized human Ct-MGAM and Nt-MGAM expressed on the baculovirus Sf9 system and *Drosophila* S2 cells, respectively. This approach could improve the reliability of *in vitro* results thereby enhancing the success rate of drug candidates in subsequent evaluations. These studies provide valuable insights and could be served as guiding references for future research on evaluating the activity of flavonoids toward human-sourced α-glucosidases.

Furthermore, there is a gap in our understanding of the interaction between flavonoids and human MGAM or SI at an atomistic level, which could provide valuable insights into their inhibitory mechanism and be used for future rational drug design. Therefore, future studies should focus on filling these gaps to explore the mechanism of flavonoids' inhibitory effects on human  $\alpha$ -glucosidases.

The results of  $\alpha$ -amylase inhibition, in contrast, show less variation and support a competitive mechanism. This is likely due to the fact that most research groups have conducted experiments using porcine  $\alpha$ -amylase, which shares a high degree of sequence identity with human  $\alpha$ -amylase. This enables more accurate extrapolation from the inhibition of flavonoids on porcine enzymes to human-sourced counterparts. It is worth noting that commercial human  $\alpha$ -amylase is also commercially available, allowing for more effective and cost-efficient inhibition assays. Additionally, the availability of flavonoid- $\alpha$ -amylase complexes enables the identification of the core interactions involved in inhibition and confirms SAR observations, thus providing a valuable tool for drug optimization before any further investigation.

There were several limitations encountered during our review process. Firstly, our database was limited to papers published in English, which meant that we were unable to retrieve data from a number of studies in other languages such as Chinese, Japanese, Korean, Portuguese, and German. This language barrier prevented us from accessing potentially relevant research articles. Additionally, although had intended to cover studies in Vietnamese, the shortage of a Vietnamese journal database also hindered us from retrieving research articles in Vietnamese. Furthermore, our systematic review has taken a considerable period due to the extended data analysis process, initiated in August 2022 and culminated in May 2023. We can only accumulate data until August 21, 2022, and thus, some of the latest articles published after this date may not have been included in our analysis. Hence, future follow-up reviews may be necessary to update our findings with the latest research articles.

## 5. Conclusions and future perspectives

Diabetes has become a challenging health problem with rapidly increasing incidents and mortality yearly. Especially, sustainable management of diabetes has faced the consistent development of drug resistance, which recognized a significant necessity for novel treatments with better pharmacological characteristics. In recent years, researchers have been increasingly turning to safe and effective phytochemicals, particularly flavonoids, because of their impressive ability to interfere with starch digestion.

The ongoing data on flavonoids *in vitro* inhibition on  $\alpha$ -glucosidase and  $\alpha$ -amylase have grown substantially. However, to the best of our knowledge, there has yet to be any exhaustive data collection efforts and discussion to provide this insight. Our review established a comprehensive overview of the inhibition potential of flavonoids against two starch-digestive enzymes, covering six databases and 974 standardized unique flavonoid structures, each with corresponding IC<sub>50</sub> values for the test substance and acarbose. In addition to the inhibition data, we also recorded the origin of flavonoids, testing methods, and the underlying mechanism of inhibition. The curated database can lay a foundation for future ligand-based QSAR studies, facilitating natural products drug discovery efforts aimed at  $\alpha$ -glucosidase inhibition while minimizing side effects related to  $\alpha$ -amylase. A number of remarks have been discussed thoroughly, including structure characteristics that would impact the activity of interest. Additionally, considerable efforts have been made to curate and assess the molecular-level interactions between flavonoids and two enzymes. Subsequently, we wish to provide a balance of evidence for the advance in the discovery of new antidiabetic candidates.

On a side note, we identified exciting prospects that should be brought to the attention of investigators.

- 1. As for *in vitro* assays, the inhibitory effects of flavonoids varied significantly with respect to the methods, substrates, and enzyme origins, thus impeding the reproducibility of consistent results. Hence, future studies should focus on developing consensual protocols for *in vitro* enzyme activity evaluation and coming up with unambiguous instructions to improve the replication of results.
- It should be noted that while porcine α-amylase could be used provisionally in replacement of human α-amylase
  in vitro due to its sequential and structural identity with human α-amylase, yeast or rat-sourced α-glucosidases
  diverged considerably from human MGAM and SI enzymes. Taking account of these distinctions, it is crucial to

- establish *in vitro* assays using human-sourced enzymes to improve the reliability of the data for successive stages on living models.
- 3. The molecular interactions between flavonoids and enzymes are of great importance in the drug development pipeline. Despite recent advances in scientific approaches and modeling techniques, little research has been reported about the mechanisms underlying the inhibitory effects of flavonoids on α-amylase and α-glucosidases. Thus, this area remained a research gap that required further investigation.

## **Author contributions**

Thua-Phong Lam: data curation, formal analysis, investigation, methodology, project administration, validation, visualization, writing-original draft; Ngoc-Vi Nguyen Tran: data curation, formal analysis, investigation, methodology, validation, visualization, writing-original draft; Long-Hung Dinh Pham: formal analysis, validation, visualization, writing-original draft; Nghia Vo-Trong Lai: data curation, investigation; Bao-Tran Ngoc Dang: data curation, investigation; Ngoc-Lam Nguyen Truong: data curation, investigation; Song-Ky Nguyen-Vo: data curation, investigation; Thuy-Linh Hoang: validation, writing-review and editing; Tan Thanh Mai: conceptualization, funding acquisition, methodology, project administration, resources, validation, writing-review and editing; Thanh-Dao Tran: conceptualization, funding acquisition, methodology, project administration, resources, validation, supervision, writing-review and editing.

#### **Conflicts of interest**

The authors declare no conflict of interest.

#### Acknowledgments

This research is funded by the University of Medicine and Pharmacy at Ho Chi Minh City under grant number 162/2019/HD-DHYD for Thanh-Dao Tran. The funding organization does not affect the transparency and the findings of the review. We would like to thank Phuc Quang Vu (School of Global Public Health, New York University) for the aid with numerical data processing and presentation and Lam-Duy Pham (Faculty of Pharmacy, University of Medicine and Pharmacy at Ho Chi Minh City) for the manuscript formatting.

#### Availability of data

The original data of 177 presented compounds and the original dataset of 1643 entries from the included studies are available online at: https://github.com/phonglam3103/Systematic review.

## References

- 1. International Diabetes Federation. *IDF Diabetes Atlas, 10th edition.* 2021.
- 2. ElSayed NA, Aleppo G, Aroda VR, et al. 2. Classification and Diagnosis of Diabetes: Standards of Care in Diabetes—2023. *Diabetes Care*. 2023;46(Supplement\_1):S19-S40. doi:10.2337/dc23-S002
- 3. Dahlén AD, Dashi G, Maslov I, et al. Trends in antidiabetic drug discovery: FDA approved drugs, new drugs in clinical trials and global sales. *Front Pharmacol*. 2022;12:4119. doi:10.3389/fphar.2021.807548
- 4. Davies MJ, Aroda VR, Collins BS, et al. Management of hyperglycemia in type 2 diabetes, 2022. A consensus report by the American Diabetes Association (ADA) and the European Association for the Study of Diabetes (EASD). *Diabetes Care*. 2022;45(11):2753-2786. doi:10.2337/dci22-0034
- 5. ElSayed NA, Aleppo G, Aroda VR, et al. 9. Pharmacologic approaches to glycemic treatment: Standards of Care in diabetes—2023. *Diabetes Care*. 2023;46(Supplement\_1):S140-S157. doi:10.2337/dc23-S009
- 6. Webb EC. Enzyme nomenclature 1992. Recommendations of the Nomenclature Committee of the International Union of Biochemistry and Molecular Biology on the Nomenclature and Classification of Enzymes. Academic Press; 1992.
- 7. Dhital S, Lin AH-M, Hamaker BR, Gidley MJ, Muniandy A. Mammalian Mucosal α-Glucosidases Coordinate with α-Amylase in the Initial Starch Hydrolysis Stage to Have a Role in Starch Digestion beyond Glucogenesis. *PLoS One*. 2013;8(4):e62546. doi:10.1371/journal.pone.0062546
- 8. Ren L, Qin X, Cao X, et al. Structural insight into substrate specificity of human intestinal maltase-glucoamylase. *Protein Cell*. 2011;2(10):827-836. doi:10.1007/s13238-011-1105-3
- 9. Koepsell H. Glucose transporters in the small intestine in health and disease. *Pflug Arch Eur J Physiol*. 2020;472(9):1207-1248. doi:10.1007/s00424-020-02439-5

- 10. Ghani U. Chapter one Introduction, rationale and the current clinical status of oral α-glucosidase inhibitors. In: Ghani U, ed. *Alpha-Glucosidase Inhibitors*. Elsevier; 2020:1-15.
- 11. Yee HS, Fong NT. A review of the safety and efficacy of acarbose in diabetes mellitus. *Pharmacotherapy*. 1996;16(5):792-805. doi:10.1002/j.1875-9114.1996.tb02997.x
- 12. Maleki SJ, Crespo JF, Cabanillas B. Anti-inflammatory effects of flavonoids. *Food Chem.* 2019;299:125124. doi:10.1016/j.foodchem.2019.125124
- 13. Agati G, Brunetti C, Fini A, et al. Are Flavonoids Effective Antioxidants in Plants? Twenty Years of Our Investigation. *Antioxidants*. 2020;9(11):1098. doi:10.3390/antiox9111098
- 14. Lam T-P, Tran V-H, Mai TT, et al. Identification of Diosmin and Flavin Adenine Dinucleotide as Repurposing Treatments for Monkeypox Virus: A Computational Study. *Int J Mol Sci.* 2022;23(19):11570. doi:10.3390/ijms231911570
- 15. Vo C-VT, Nguyen LC, Le TTA, et al. Natural mimetic 4-benzyloxychalcones as potent pancreatic lipase inhibitors: Virtual screening, synthesis and biological evaluation. *Phytochem Lett.* 2022;51:28-33. doi:10.1016/j.phytol.2022.06.011
- 16. Collado-González J, Grosso C, Valentão P, et al. Inhibition of α-glucosidase and α-amylase by Spanish extra virgin olive oils: The involvement of bioactive compounds other than oleuropein and hydroxytyrosol. *Food Chem.* 2017;235:298-307. doi:10.1016/j.foodchem.2017.04.171
- 17. Sun L, Warren FJ, Netzel G, Gidley MJ. 3 or 3'-Galloyl substitution plays an important role in association of catechins and theaflavins with porcine pancreatic α-amylase: The kinetics of inhibition of α-amylase by tea polyphenols. *J Funct Foods*. 2016;26:144-156. doi:10.1016/j.jff.2016.07.012
- 18. Tundis R, Bonesi M, Sicari V, et al. Poncirus trifoliata (L.) Raf.: Chemical composition, antioxidant properties and hypoglycaemic activity via the inhibition of α-amylase and α-glucosidase enzymes. *J Funct Foods*. 2016;25:477-485. doi:10.1016/j.jff.2016.06.034
- 19. Proença C, Ribeiro D, Freitas M, Fernandes E. Flavonoids as potential agents in the management of type 2 diabetes through the modulation of α-amylase and α-glucosidase activity: a review. *Crit Rev Food Sci Nutr*. 2022;62(12):3137-3207. doi:10.1080/10408398.2020.1862755
- 20. Zhu J, Chen C, Zhang B, Huang Q. The inhibitory effects of flavonoids on α-amylase and α-glucosidase. *Crit Rev Food Sci Nutr.* 2020;60(4):695-708. doi:10.1080/10408398.2018.1548428
- Şöhretoğlu D, Sari S. Flavonoids as alpha-glucosidase inhibitors: mechanistic approaches merged with enzyme kinetics and molecular modelling. *Phytochem Rev.* 2020;19(5):1081-1092. doi:10.1007/s11101-019-09610-6
- 22. Mahapatra DK, Asati V, Bharti SK. Chalcones and their therapeutic targets for the management of diabetes: Structural and pharmacological perspectives. *Eur J Med Chem.* 2015;92:839-865. doi:10.1016/j.ejmech.2015.01.051
- 23. Rocha S, Ribeiro D, Fernandes E, Freitas M. A Systematic Review on Anti-diabetic Properties of Chalcones. *Curr Med Chem.* 2020;27(14):2257-2321. doi:10.2174/0929867325666181001112226
- 24. Xiao J, Ni X, Kai G, Chen X. A Review on Structure–Activity Relationship of Dietary Polyphenols Inhibiting α-Amylase. *Crit Rev Food Sci Nutr.* 2013;53(5):497-506. doi:10.1080/10408398.2010.548108
- 25. Page MJ, McKenzie JE, Bossuyt PM, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ*. 2021;372:n71. doi:10.1136/bmj.n71
- 26. Ouzzani M, Hammady H, Fedorowicz Z, Elmagarmid A. Rayyan—a web and mobile app for systematic reviews. *Syst Rev.* 2016;5(1):210. doi:10.1186/s13643-016-0384-4
- 27. Perkin Elmer. ChemDraw The Gold Standard for Communicating Chemistry Research. Accessed Dec 13 2022, <a href="https://perkinelmerinformatics.com/products/research/chemdraw/">https://perkinelmerinformatics.com/products/research/chemdraw/</a>
- 28. Faggion CM, Jr. Guidelines for reporting pre-clinical in vitro studies on dental materials. *J Evid Based Dent Pract*. 2012;12(4):182-9. doi:10.1016/j.jebdp.2012.10.001
- 29. Landrum G. RDKit: Open-Source Cheminformatics Software. Accessed Oct 09, 2022, http://www.rdkit.org
- 30. Visvanathan R, Qader M, Jayathilake C, Jayawardana BC, Liyanage R, Sivakanesan R. Critical review on conventional spectroscopic α-amylase activity detection methods: merits, demerits, and future prospects. 10.1002/jsfa.10315. *J Sci Food Agric*. 2020;100(7):2836-2847. doi:10.1002/jsfa.10315
- 31. Yi J, Zhao T, Zhang Y, et al. Isolated compounds from Dracaena angustifolia Roxb and acarbose synergistically/additively inhibit alpha-glucosidase and alpha-amylase: an in vitro study. *BMC Complement Altern Med*. 2022;22(1):177. doi:10.1186/s12906-022-03649-3
- 32. Zhang LL, Han L, Yang SY, Meng XM, Ma WF, Wang M. The mechanism of interactions between flavan-3-ols against a-glucosidase and their in vivo antihyperglycemic effects. *Bioorg Chem.* 2019;85:364-372. doi:10.1016/j.bioorg.2018.12.037

- 33. Lima Júnior JPD, Franco RR, Saraiva AL, Moraes IB, Espindola FS. Anacardium humile St. Hil as a novel source of antioxidant, antiglycation and α-amylase inhibitors molecules with potential for management of oxidative stress and diabetes. *J Ethnopharmacol*. 2021;268:113667. doi:10.1016/j.jep.2020.113667
- 34. Nazir N, Zahoor M, Ullah R, Ezzeldin E, Mostafa GAE. Curative Effect of Catechin Isolated from Elaeagnus Umbellata Thunb. Berries for Diabetes and Related Complications in Streptozotocin-Induced Diabetic Rats Model. *Molecules*. 2020;26(1):137. doi:10.3390/molecules26010137
- Jiang P, Zhao Y, Xiong J, et al. Extraction, Purification, and Biological Activities of Flavonoids from Branches and Leaves of Taxus cuspidata S. et Z. *BioResources*. 2021;16(2):2655-2682. doi:10.15376/biores.16.2.2655-2682
- 36. Wu Q, Min Y, Xiao J, et al. Liquid state fermentation vinegar enriched with catechin as an antiglycative food product. *Food Funct*. 2019;10(8):4877-4887. doi:10.1039/c8fo01892h
- 37. Liu J, Lu JF, Kan J, Wen XY, Jin CH. Synthesis, characterization and in vitro anti-diabetic activity of catechin grafted inulin. *Int J Biol Macromol*. 2014;64:76-83. doi:10.1016/j.ijbiomac.2013.11.028
- 38. Ray S, Samanta T, Mitra A, De B. Effect of extracts and components of black tea on the activity of beta-glucuronidase, lipase, alpha-amylase, alpha-glucosidase: An in vitro study. *Curr Nutr Food Sci.* 2014;10(3):181-186. doi:10.2174/1573401310666140529205646
- 39. Tian JL, Si X, Wang YH, et al. Bioactive flavonoids from Rubus corchorifolius inhibit α-glucosidase and α-amylase to improve postprandial hyperglycemia. *Food Chem.* 2021;341:128149. doi:10.1016/j.foodchem.2020.128149
- 40. Zhou H, Li HM, Du YM, et al. C-geranylated flavanones from YingDe black tea and their antioxidant and α-glucosidase inhibition activities. *Food Chem*. 2017;235:227-233. doi:10.1016/j.foodchem.2017.05.034
- 41. Numonov S, Edirs S, Bobakulov K, et al. Evaluation of the antidiabetic activity and chemical composition of geranium collinum root extracts—computational and experimental investigations. *Molecules*. 2017;22(6):983. doi:10.3390/molecules22060983
- 42. Deutschlander MS, Lall N, Van De Venter M, Hussein AA. Hypoglycemic evaluation of a new triterpene and other compounds isolated from Euclea undulata Thunb. var. myrtina (Ebenaceae) root bark. *J Ethnopharmacol*. 2011;133(3):1091-1095. doi:10.1016/j.jep.2010.11.038
- 43. Zhao L, Wen L, Lu Q, Liu R. Interaction mechanism between alpha-glucosidase and A-type trimer procyanidin revealed by integrated spectroscopic analysis techniques. *Int J Biol Macromol*. 2020;143:173-180. doi:10.1016/j.ijbiomac.2019.12.021
- 44. Zhang H, Yerigui, Yang Y, Ma C. Structures and antioxidant and intestinal disaccharidase inhibitory activities of A-type proanthocyanidins from peanut skin. *J Agric Food Chem.* 2013;61(37):8814-20. doi:10.1021/jf402518k
- 45. He XF, Chen JJ, Huang XY, et al. The antidiabetic potency of Amomum tsao-ko and its active flavanols, as PTP1B selective and α-glucosidase dual inhibitors. *Ind Crops Prod.* 2021;160:112908. doi:10.1016/j.indcrop.2020.112908
- 46. Gong T, Yang X, Bai F, et al. Young apple polyphenols as natural α-glucosidase inhibitors: In vitro and in silico studies. *Bioorg Chem.* 2020;96:103625. doi:10.1016/j.bioorg.2020.103625
- 47. Giang Thanh Thi H, Kase ET, Wangensteen H, Barsett H. Effect of Phenolic Compounds from Elderflowers on Glucose- and Fatty Acid Uptake in Human Myotubes and HepG2-Cells. *Molecules*. 2017;22(1):90. doi:10.3390/molecules22010090
- 48. Wang X, Liu Q, Zhu H, et al. Flavanols from the Camellia sinensis var. assamica and their hypoglycemic and hypolipidemic activities. *Acta Pharm Sin B*. 2017;7(3):342-346. doi:10.1016/j.apsb.2016.12.007
- 49. Dat NT, Dang NH, Thanh le N. New flavonoid and pentacyclic triterpene from Sesamum indicum leaves. *Nat Prod Res.* 2016;30(3):311-5. doi:10.1080/14786419.2015.1057730
- 50. Li S, Lo C-Y, Pan M-H, Lai C-S, Ho C-T. Black tea: chemical analysis and stability. 10.1039/C2FO30093A. *Food Funct*. 2013;4(1):10-18. doi:10.1039/C2FO30093A
- 51. Wu X, Hu M, Hu X, Ding H, Gong D, Zhang G. Inhibitory mechanism of epicatechin gallate on α-amylase and α-glucosidase and its combinational effect with acarbose or epigallocatechin gallate. *J Mol Liq*. 2019;290:111202. doi:10.1016/j.molliq.2019.111202
- 52. Chen Y, Ye X, Wang L, et al. Three flavanols delay starch digestion by inhibiting alpha-amylase and binding with starch. *Int J Biol Macromol*. 2021;172:503-514. doi:10.1016/j.ijbiomac.2021.01.070
- 53. Xu L, Li W, Chen Z, et al. Inhibitory effect of epigallocatechin-3-O-gallate on alpha-glucosidase and its hypoglycemic effect via targeting PI3K/AKT signaling pathway in L6 skeletal muscle cells. *Int J Biol Macromol*. 2019;125:605-611. doi:10.1016/j.ijbiomac.2018.12.064

- 54. Su J, Tang Z. Effects of (-)-epigallocatechin gallate and quercetin on the activity and structure of alpha-amylase. *Trop J Pharm Res.* 2019;18(3):585-590. doi:10.4314/tjpr.v18i3.20
- 55. Jez JM, Bowman ME, Dixon RA, Noel JP. Structure and mechanism of the evolutionarily unique plant enzyme chalcone isomerase. *Nat Struct Biol*. 2000;7(9):786-791. doi:10.1038/79025
- 56. Cassidy A, Kay C. Phytochemicals: Classification and Occurrence. In: Caballero B, ed. *Encyclopedia of Human Nutrition (Third Edition)*. Academic Press; 2013:39-46.
- 57. Thuy NTL, Thuy PT, Tung BT, et al. A new flavone glycoside from lumnitzera littorea with in vitro α-glucosidase inhibitory activity. *Nat Prod Commun*. 2019;doi:10.1177/1934578X19851361
- 58. Choi CI, Lee SR, Kim KH. Antioxidant and α-glucosidase inhibitory activities of constituents from Euonymus alatus twigs. *Ind Crops Prod.* 2015;76:1055-1060. doi:10.1016/j.indcrop.2015.08.031
- 59. Uddin S, Brooks PR, Tran TD. Chemical Characterization, α-Glucosidase, α-Amylase and Lipase Inhibitory Properties of the Australian Honey Bee Propolis. *Foods*. 2022;11(13):1964. doi:10.3390/foods11131964
- 60. Tang H, Huang L, Sun C, Zhao D. Exploring the structure-activity relationship and interaction mechanism of flavonoids and alpha-glucosidase based on experimental analysis and molecular docking studies. *Food Funct*. 2020;11(4):3332-3350. doi:10.1039/c9fo02806d
- 61. Zhang X, Liu Z, Bi X, Liu J, Li W, Zhao Y. Flavonoids and its derivatives from Callistephus chinensis flowers and their inhibitory activities against α-glucosidase. *EXCLI J.* 2013;12:956-966.
- 62. Zhang Y, Xiao Z, Zhang X, Sun H. Hypoglycemic and hypolipidemic dual activities of extracts and flavonoids from Desmodium caudatum and an efficient synthesis of the most potent 8-prenylquercetin. *Fitoterapia*. 2022;156:105083. doi:10.1016/j.fitote.2021.105083
- 63. Fang HL, Liu ML, Li SY, et al. Identification, potency evaluation, and mechanism clarification of alpha-glucosidase inhibitors from tender leaves of Lithocarpus polystachyus Rehd. *Food Chem.* 2022;371:131128. doi:10.1016/j.foodchem.2021.131128
- 64. Priscilla DH, Roy D, Suresh A, Kumar V, Thirumurugan K. Naringenin inhibits alpha-glucosidase activity: A promising strategy for the regulation of postprandial hyperglycemia in high fat diet fed streptozotocin induced diabetic rats. *Chem Biol Interact*. 2014;210(1):77-85. doi:10.1016/j.cbi.2013.12.014
- 65. Sun H, Wang D, Song X, et al. Natural Prenylchalconaringenins and Prenylnaringenins as Antidiabetic Agents: alpha-Glucosidase and alpha-Amylase Inhibition and in Vivo Antihyperglycemic and Antihyperlipidemic Effects. *J Agric Food Chem.* 2017;65(8):1574-1581. doi:10.1021/acs.jafc.6b05445
- 66. Qin NB, Jia CC, Xu J, et al. New amides from seeds of Silybum marianum with potential antioxidant and antidiabetic activities. *Fitoterapia*. 2017;119:83-89. doi:10.1016/j.fitote.2017.04.008
- 67. Ren D, Meng FC, Liu H, et al. Novel biflavonoids from Cephalotaxus oliveri Mast. *Phytochem Lett.* 2018;24:150-153. doi:10.1016/j.phytol.2018.02.005
- 68. Zhang K, Ding Z, Duan W, et al. Optimized preparation process for naringenin and evaluation of its antioxidant and α-glucosidase inhibitory activities. *J Food Process Preserv*. 2020;44(12):e14931. doi:10.1111/jfpp.14931
- Xu J, Wang X, Yue J, Sun Y, Zhang X, Zhao Y. Polyphenols from Acorn Leaves (Quercus liaotungensis) Protect Pancreatic Beta Cells and Their Inhibitory Activity against α-Glucosidase and Protein Tyrosine Phosphatase 1B. Molecules. 2018;23(9):2167. doi:10.3390/molecules23092167
- 70. Zhao BT, Duc Dat L, Phi Hung N, et al. PTP1B, alpha-glucosidase, and DPP-IV inhibitory effects for chromene derivatives from the leaves of Smilax china L. *Chem Biol Interact*. 2016;253:27-37. doi:10.1016/j.cbi.2016.04.012
- 71. Nguyen TP, Le TD, Minh PN, et al. A new dihydrofurocoumarin from the fruits of Pandanus tectorius Parkinson ex Du Roi. *Nat Prod Res.* 2016;30(21):2389-95. doi:10.1080/14786419.2016.1188095
- 72. Zhou Q, Lei X, Niu J, Chen Y, Shen X, Zhang N. A new hemiacetal chromone racemate and α-glucosidase inhibitors from Ficus tikoua Bur. *Nat Prod Res*. 2022;doi:10.1080/14786419.2022.2068544
- 73. Jia Y, Ma Y, Cheng G, Zhang Y, Cai S. Comparative Study of Dietary Flavonoids with Different Structures as alpha-Glucosidase Inhibitors and Insulin Sensitizers. *J Agric Food Chem.* 2019;67(37):10521-10533. doi:10.1021/acs.jafc.9b04943
- 74. Habtemariam S. The anti-obesity potential of sigmoidin A. *Pharm Biol.* 2012;50(12):1519-1522. doi:10.3109/13880209.2012.688838
- 75. Luyen NT, Tram LH, Hanh TTH, et al. Inhibitors of a-glucosidase, a-amylase and lipase from Chrysanthemum morifolium. *Phytochem Lett.* 2013;6(3):322-325. doi:10.1016/j.phytol.2013.03.015
- 76. Li K, Yao F, Xue Q, et al. Inhibitory effects against α-glucosidase and α-amylase of the flavonoids-rich extract from Scutellaria baicalensis shoots and interpretation of structure-activity relationship of its eight flavonoids by a refined assign-score method. *Chem Cent J.* 2018;12(1):82. doi:10.1186/s13065-018-0445-y

- 77. Sahnoun M, Trabelsi S, Bejar S. Citrus flavonoids collectively dominate the α-amylase and α-glucosidase inhibitions. *Biologia*. 2017;72(7):764-773. doi:10.1515/biolog-2017-0091
- 78. Kong F, Ding Z, Zhang K, et al. Optimization of extraction flavonoids from Exocarpium Citri Grandis and evaluation its hypoglycemic and hypolipidemic activities. *J Ethnopharmacol*. 2020;262:113178. doi:10.1016/j.jep.2020.113178
- 79. Taslimi P, Caglayan C, Gulcin I. The impact of some natural phenolic compounds on carbonic anhydrase, acetylcholinesterase, butyrylcholinesterase, and alpha-glycosidase enzymes: An antidiabetic, anticholinergic, and antiepileptic study. *J Biochem Mol Toxicol*. 2017;31(12):e21995. doi:10.1002/jbt.21995
- 80. Qurtam AA, Mechchate H, Es-Safi I, et al. Citrus flavanone narirutin, in vitro and in silico mechanistic antidiabetic potential. *Pharmaceutics*. 2021;13(11):1818. doi:10.3390/pharmaceutics13111818
- 81. Pyrzynska K. Hesperidin: A Review on Extraction Methods, Stability and Biological Activities. *Nutrients*. 2022;14(12):2387. doi:10.3390/nu14122387
- 82. Zhao Y, Kongstad KT, Jager AK, Nielsen J, Staerk D. Quadruple high-resolution alpha-glucosidase/alpha-amylase/PTP1B/radical scavenging profiling combined with high-performance liquid chromatography-high-resolution mass spectrometry-solid-phase extraction-nuclear magnetic resonance spectroscopy for identification of antidiabetic constituents in crude root bark of Morus alba L. *J Chromatogr A*. 2018;1556:55-63. doi:10.1016/j.chroma.2018.04.041
- 83. Bui TT, Nguyen KPT, Nguyen PPK, Le DT, Nguyen TLT. Anti-Inflammatory and α-Glucosidase Inhibitory Activities of Chemical Constituents from Bruguiera parviflora Leaves. *J Chem.* 2022;doi:10.1155/2022/3049994
- 84. Jing P, Xiaomin Y, Shujuan Z, et al. Bioactive phenolics from mango leaves (Mangifera indica L.). *Ind Crops Prod.* 2018;111:400-406. doi:10.1016/j.indcrop.2017.10.057
- 85. Khalid MF, Rehman K, Irshad K, Chohan TA, Akash MSH. Biochemical Investigation of Inhibitory Activities of Plant-Derived Bioactive Compounds Against Carbohydrate and Glucagon-Like Peptide-1 Metabolizing Enzymes. *Dose-Response*. 2022;(2)doi:10.1177/15593258221093275
- 86. Su H, Ruan YT, Li Y, Chen JG, Yin ZP, Zhang QF. In vitro and in vivo inhibitory activity of taxifolin on three digestive enzymes. *Int J Biol Macromol*. 2020;150:31-37. doi:10.1016/j.ijbiomac.2020.02.027
- 87. Yoon KD, Lee JY, Kim TY, et al. In Vitro and in Vivo Anti-Hyperglycemic Activities of Taxifolin and Its Derivatives Isolated from Pigmented Rice (Oryzae sativa L. cv. Superhongmi). *J Agric Food Chem*. 2020;68(3):742-750. doi:10.1021/acs.jafc.9b04962
- 88. Yang J, Li H, Wang X, Zhang C, Feng G, Peng X. Inhibition Mechanism of alpha-Amylase/alpha-Glucosidase by Silibinin, Its Synergism with Acarbose, and the Effect of Milk Proteins. *J Agric Food Chem*. 2021;69(36):10515-10526. doi:10.1021/acs.jafc.1c01765
- 89. Van Thanh B, Van Anh NT, Ha CTT, et al. A new 2,3-dioxygenated flavanone and other constituents from dysosma difformis. *Rec Nat Prod.* 2022;16(1):92-97. doi:10.25135/rnp.256.21.03.2017
- 90. Wu S, Tian L. A new flavone glucoside together with known ellagitannins and flavones with anti-diabetic and anti-obesity activities from the flowers of pomegranate (Punica granatum). *Nat Prod Res.* 2019;33(2):252-257. doi:10.1080/14786419.2018.1446009
- 91. Nguyen TTH, Nguyen VT, Van Cuong P, et al. A new flavonoid from the leaves of Garcinia mckeaniana Craib and α-glucosidase and acetylcholinesterase inhibitory activities. *Nat Prod Res.* 2021;36(19):5074-5080. doi:10.1080/14786419.2021.1916019
- 92. Chang CC, Ho SL, Lee SS. Acylated glucosylflavones as α-glucosidase inhibitors from Tinospora crispa leaf. *Bioorg Med Chem.* 2015;23(13):3388-96. doi:10.1016/j.bmc.2015.04.053
- 93. Vi LNT, Tuan NN, Hung QT, et al. alpha-glucosidase Inhibitory Activity of Extracts and Compounds from the Leaves of Ruellia tuberosa L. *Nat Prod J.* 2022;12(5):63-68. doi:10.2174/2210315511666210218214955
- 94. Choi CI, Eom HJ, Kim KH. Antioxidant and α-glucosidase inhibitory phenolic constituents of Lactuca indica L. *Russ J Biographic Chem.* 2016;42(3):310-315. doi:10.1134/S1068162016030079
- 95. Fidelis QC, Faraone I, Russo D, et al. Chemical and Biological insights of Ouratea hexasperma (A. St.-Hil.) Baill.: a source of bioactive compounds with multifunctional properties. *Nat Prod Res.* 2019;33(10):1500-1503. doi:10.1080/14786419.2017.1419227
- 96. Tian X, Guo S, Zhang S, et al. Chemical characterization of main bioactive constituents in Paeonia ostii seed meal and GC-MS analysis of seed oil. *J Food Biochem*. 2020;44(1):e13088. doi:10.1111/jfbc.13088
- 97. Nguyen DH, Le DD, Ma ES, Min BS, Woo MH. Development and validation of an HPLC-PDA method for quantitation of ten marker compounds from Eclipta prostrata (L.) and evaluation of their protein tyrosine phosphatase 1B, alpha-glucosidase, and acetylcholinesterase inhibitory activities. *Nat Prod Sci.* 2020;26(4):326-333. doi:10.20307/nps.2020.26.4.326

- 98. Su ZR, Fan SY, Shi WG, Zhong BH. Discovery of xanthine oxidase inhibitors and/or α-glucosidase inhibitors by carboxyalkyl derivatization based on the flavonoid of apigenin. *Bioorg Med Chem Lett.* 2015;25(14):2778-81. doi:10.1016/j.bmcl.2015.05.016
- 99. Li M, Bao X, Zhang X, et al. Exploring the phytochemicals and inhibitory effects against α-glucosidase and dipeptidyl peptidase-IV in Chinese pickled chili pepper: Insights into mechanisms by molecular docking analysis. *LWT*. 2022;162:113467. doi:10.1016/j.lwt.2022.113467
- 100. Dao TBN, Nguyen TMT, Nguyen VQ, et al. Flavones from combretum quadrangulare growing in vietnam and their alpha-glucosidase inhibitory activity. *Molecules*. 2021;26(9):2531. doi:10.3390/molecules26092531
- 101. Abbasi MA, Hussain G, Aziz ur R, Ahmad VU. Flavonoids from Rhynchosia pseudo-cajan as suitable alpha-glucosidase inhibitors and free radical scavengers. *Int Res J Pharm*. 2014;5(8):636-641. doi:10.7897/2230-8407.0508130
- 102. Proença C, Freitas M, Ribeiro D, et al. α-Glucosidase inhibition by flavonoids: an in vitro and in silico structure-activity relationship study. *J Enzyme Inhib Med Chem*. 2017;32(1):1216-1228. doi:10.1080/14756366.2017.1368503
- 103. Duong TH, Nguyen HT, Nguyen CH, et al. Identification of Highly Potent alpha-Glucosidase Inhibitors from Artocarpus integer and Molecular Docking Studies. *Chem Biodivers*. 2021;18(12):e2100499. doi:10.1002/cbdv.202100499
- 104. Assefa ST, Yang EY, Asamenew G, Kim HW, Cho MC, Lee J. Identification of α-glucosidase inhibitors from leaf extract of pepper (Capsicum spp.) through metabolomic analysis. *Metabolites*. 2021;11(10):649. doi:10.3390/metabol1100649
- 105. Wang X, Yang J, Li H, Shi S, Peng X. Mechanistic study and synergistic effect on inhibition of α-amylase by structurally similar flavonoids. *J Mol Liq*. 2022;360:119485. doi:10.1016/j.molliq.2022.119485
- 106. Quan YS, Zhang XY, Yin XM, Wang SH, Jin LL. Potential alpha-glucosidase inhibitor from Hylotelephium erythrostictum. *Bioorg Med Chem Lett.* 2020;30(24):127665. doi:10.1016/j.bmcl.2020.127665
- 107. Lianwu X, Qiachi F, Shuyun S, Jiawei L, Xinji Z. Rapid and comprehensive profiling of alpha-glucosidase inhibitors in Buddleja Flos by ultrafiltration HPLC-QTOF-MS/MS with diagnostic ions filtering strategy. *Food Chem.* 2021;344:128651-128651. doi:10.1016/j.foodchem.2020.128651
- 108. Li H, Song F, Xing J, Tsao R, Liu Z, Liu S. Screening and Structural Characterization of α-Glucosidase Inhibitors from Hawthorn Leaf Flavonoids Extract by Ultrafiltration LC-DAD-MSn and SORI-CID FTICR MS. *J Am Soc Mass Spectrom*. 2009;20(8):1496-1503. doi:10.1016/j.jasms.2009.04.003
- 109. Wu B, Song HP, Zhou X, et al. Screening of minor bioactive compounds from herbal medicines by in silico docking and the trace peak exposure methods. *J Chromatogr A*. 2016;1436:91-99. doi:10.1016/j.chroma.2016.01.062
- 110. Wang YM, Zhao JQ, Yang JL, Tao YD, Mei LJ, Shi YP. Separation of antioxidant and alpha-glucosidase inhibitory flavonoids from the aerial parts of Asterothamnus centrali-asiaticus. *Nat Prod Res.* 2017;31(12):1365-1369. doi:10.1080/14786419.2016.1247083
- 111. Cheng N, Yi WB, Wang QQ, Peng SM, Zou XQ. Synthesis and α-glucosidase inhibitory activity of chrysin, diosmetin, apigenin, and luteolin derivatives. *Chin Chem Lett.* 2014;25(7):1094-1098. doi:10.1016/j.cclet.2014.05.021
- 112. Huang Q, Chen JJ, Pan Y, et al. Chemical profiling and antidiabetic potency of Paeonia delavayi: Comparison between different parts and constituents. *J Pharm Biomed Anal.* 2021;198:113998. doi:10.1016/j.jpba.2021.113998
- 113. Silva EL, Lobo JFR, Vinther JM, Borges RM, Staerk D. High-resolution alpha-glucosidase inhibition profiling combined with HPLC-HRMS-SPE-NMR for identification of anti-diabetic compounds in Eremanthus crotonoides (Asteraceae). *Molecules*. 2016;21(6):782. doi:10.3390/molecules21060782
- 114. Vonia S, Hartati R, Insanu M. In Vitro Alpha-Glucosidase Inhibitory Activity and the Isolation of Luteolin from the Flower of Gymnanthemum amygdalinum (Delile) Sch. Bip ex Walp. *Molecules*. 2022;27(7):2132. doi:10.3390/molecules27072132
- 115. Djeujo FM, Ragazzi E, Urettini M, et al. Magnolol and Luteolin Inhibition of alpha-Glucosidase Activity: Kinetics and Type of Interaction Detected by In Vitro and In Silico Studies. *Pharmaceuticals*. 2022;15(2):205. doi:10.3390/ph15020205
- 116. Yang Y, Gu L, Xiao Y, et al. Rapid identification of α-glucosidase inhibitors from phlomis tuberosa by sepbox chromatography and thin-layer chromatography bioautography. *PLoS One.* 2015;10(2):e0116922. doi:10.1371/journal.pone.0116922
- 117. Ablat A, Halabi MF, Mohamad J, et al. Antidiabetic effects of Brucea javanica seeds in type 2 diabetic rats. *BMC Complement Altern Med*. 2017;17(1):94. doi:10.1186/s12906-017-1610-x

- 118. Hlila MB, Majouli K, Ben Jannet H, Mastouri M, Aouni M, Selmi B. Antioxidant and anti alpha-glucosidase of luteolin and luteolin 7-O-glucoside isolated from Scabiosa arenaria Forssk. *J Coast Life Med.* 2017;5(7):317-320. doi:10.12980/jclm.5.2017J7-66
- 119. Bo-wei Z, Xia L, Wen-long S, et al. Dietary flavonoids and acarbose synergistically inhibit alpha-glucosidase and lower postprandial blood glucose. *J Agric Food Chem.* 2017;65(38):8319-8330. doi:10.1021/acs.jafc.7b02531
- 120. Yan J, Zhang G, Pan J, Wang Y. α-Glucosidase inhibition by luteolin: kinetics, interaction and molecular docking. *Int J Biol Macromol*. 2014;64:213-23. doi:10.1016/j.ijbiomac.2013.12.007
- 121. Kuroda M, Iwabuchi K, Mimaki Y. Chemical constituents of the aerial parts of Scutellaria lateriflora and their alpha-glucosidase inhibitory activities. *Nat Prod Commun*. 2012;7(4):471-474. doi:10.1177/1934578x1200700413
- 122. Park MJ, Kang Y-H. Isolation of Isocoumarins and Flavonoids as α-Glucosidase Inhibitors from Agrimonia pilosa L. *Molecules*. 2020;25(11):2572. doi:10.3390/molecules25112572
- 123. Matsui T, Kobayashi M, Hayashida S, Matsumoto K. Luteolin, a flavone, does not suppress postprandial glucose absorption through an inhibition of alpha-glucosidase action. *Biosci Biotechnol Biochem*. 2002;66(3):689-692. doi:10.1271/bbb.66.689
- 124. Dej-Adisai S, Rais IR, Wattanapiromsakul C, Pitakbut T. Alpha-glucosidase inhibitory assay-screened isolation and molecular docking model from bauhinia pulla active compounds. *Molecules*. 2021;26(19):5970. doi:10.3390/molecules26195970
- 125. Lo Piparo E, Scheib H, Frei N, Williamson G, Grigorov M, Chou CJ. Flavonoids for controlling starch digestion: Structural requirements for inhibiting human alpha-amylase. *J Med Chem.* 2008;51(12):3555-3561. doi:10.1021/jm800115x
- 126. Xiao Z, Hou X, Zhang T, et al. Starch-digesting product analysis based on the hydrophilic interaction liquid chromatography coupled mass spectrometry method to evaluate the inhibition of flavonoids on pancreatic α-amylase. *Food Chem.* 2022;372:131175. doi:10.1016/j.foodchem.2021.131175
- 127. Yang JR, Luo JG, Kong LY. Determination of α-glucosidase inhibitors from ScutScutellaria baicalensis using liquid chromatography with quadrupole time of flight tandem mass spectrometry coupled with centrifugal ultrafiltration. *Chin J Nat Med.* 2015;13(3):208-214. doi:10.1016/S1875-5364(15)30006-6
- 128. Meesakul P, Richardson C, Pyne SG, Laphookhieo S. α-Glucosidase Inhibitory Flavonoids and Oxepinones from the Leaf and Twig Extracts of Desmos cochinchinensis. *J Nat Prod.* 2019;82(4):741-747. doi:10.1021/acs.jnatprod.8b00581
- 129. Do LTM, Sichaem J. New Flavonoid Derivatives from Melodorum fruticosum and Their α-Glucosidase Inhibitory and Cytotoxic Activities. *Molecules*. 2022;27(13):4023. doi:10.3390/molecules27134023
- 130. Hari Babu T, Rama Subba Rao V, Tiwari AK, et al. Synthesis and biological evaluation of novel 8-aminomethylated oroxylin A analogues as alpha-glucosidase inhibitors. *Bioorg Med Chem Lett.* 2008;18(5):1659-62. doi:10.1016/j.bmcl.2008.01.055
- 131. Kumar GS, Tiwari AK, Rao VRS, Prasad KR, Ali AZ, Babu KS. Synthesis and biological evaluation of novel benzyl-substituted flavones as free radical (DPPH) scavengers and-glucosidase inhibitors. *J Asian Nat Prod Res*. 2010;12(11):978-984. doi:10.1080/10286020.2010.511190
- 132. Ha K-N, Nguyen T-V-A, Mai D-T, et al. Alpha-glucosidase inhibitors from Nervilia concolor, Tecoma stans, and Bouea macrophylla. *Saudi J Biol Sci.* 2022;29(2):1029-1042. doi:10.1016/j.sjbs.2021.09.070
- 133. Nickavar B, Abolhasani L. Bioactivity-Guided Separation of an α-Amylase Inhibitor Flavonoid from Salvia virgata. *Iran J Pharm Res.* 2013;12(1):57-61.
- 134. Gao H, Kawabata J. alpha-Glucosidase inhibition of 6-hydroxyflavones. Part 3: Synthesis and evaluation of 2,3,4-trihydroxybenzoyl-containing flavonoid analogs and 6-aminoflavones as alpha-glucosidase inhibitors. *Bioorg Med Chem.* 2005;13(5):1661-1671. doi:10.1016/j.bmc.2004.12.010
- 135. Nguyen MTT, Nguyen NT, Nguyen HX, Huynh TNN, Min BS. Screening of alpha-glucosidase inhibitory activity of vietnamese medicinal plants: Isolation of active principles from Oroxylum indicum. *Nat Prod Sci.* 2012;18(1):47-51.
- 136. Gulcin I, Taslimi P, Aygun A, et al. Antidiabetic and antiparasitic potentials: Inhibition effects of some natural antioxidant compounds on alpha-glycosidase, alpha-amylase and human glutathione S-transferase enzymes. *Int J Biol Macromol.* 2018;119:741-746. doi:10.1016/j.ijbiomac.2018.08.001
- 137. Uddin MJ, Faraone I, Haque MA, et al. Insights into the leaves of Ceriscoides campanulata: Natural proanthocyanidins alleviate diabetes, inflammation, and esophageal squamous cell cancer via in vitro and in silico models. *Fitoterapia*. 2022;158:105164. doi:10.1016/j.fitote.2022.105164

- 138. Demir Y, Durmaz L, Taslimi P, Gulçin İ. Antidiabetic properties of dietary phenolic compounds: Inhibition effects on α-amylase, aldose reductase, and α-glycosidase. *Biotechnol Appl Biochem.* 2019;66(5):781-786. doi:10.1002/bab.1781
- 139. Sadasivam M, Kumarasamy C, Thangaraj A, et al. Phytochemical constituents from dietary plant Citrus hystrix. *Nat Prod Res.* 2018;32(14):1721-1726. doi:10.1080/14786419.2017.1399386
- 140. Ha MT, Seong SH, Nguyen TD, et al. Chalcone derivatives from the root bark of Morus alba L. act as inhibitors of PTP1B and α-glucosidase. *Phytochemistry*. 2018;155:114-125. doi:10.1016/j.phytochem.2018.08.001
- 141. Kwon RH, Thaku N, Timalsina B, Park SE, Choi JS, Jung HA. Inhibition Mechanism of Components Isolated from Morus alba Branches on Diabetes and Diabetic Complications via Experimental and Molecular Docking Analyses. *Antioxidants*. 2022;11(2):383. doi:10.3390/antiox11020383
- 142. Sun H, Song X, Tao Y, et al. Synthesis & α-glucosidase inhibitory & glucose consumption-promoting activities of flavonoid–coumarin hybrids. *Future Med Chem.* 2018;10(9):1055-1066. doi:10.4155/fmc-2017-0293
- 143. Asghari B, Salehi P, Sonboli A, Ebrahimi SN. Flavonoids from Salvia chloroleuca with alpha-amylsae and alpha-glucosidase inhibitory effect. *Iran J Pharm Res*. 2015;14(2):609-615.
- 144. Dubey K, Dubey R, Gupta R, Gupta A. Exploration of diosmin to control diabetes and its complications-an in vitro and in silico approach. *Curr Comput Aided Drug Des.* 2021;17(2):307-313. doi:10.2174/1573409916666200324135734
- 145. Wu C, Shen J, He P, et al. The alpha-glucosidase inhibiting isoflavones isolated from belamcanda chinensis leaf extract. *Rec Nat Prod.* 2012;6(2):110-120.
- 146. Sadeghi M, Miroliaei M, Ghanadian M. Inhibitory effect of flavonoid glycosides on digestive enzymes: In silico, in vitro, and in vivo studies. *Int J Biol Macromol*. 2022;217:714-730. doi:10.1016/j.ijbiomac.2022.07.086
- 147. Proença C, Freitas M, Ribeiro D, et al. Evaluation of a flavonoids library for inhibition of pancreatic α-amylase towards a structure–activity relationship. *J Enzyme Inhib Med Chem*. 2019;34(1):577-588. doi:10.1080/14756366.2018.1558221
- 148. Crozier A, Jaganath IB, Clifford MN. Phenols, Polyphenols and Tannins: An Overview. In: Crozier A, Clifford MN, Ashihara H, eds. *Plant Secondary Metabolites*. Blackwell Publishing; 2006:1-24.
- 149. Liu Y, Zhan L, Xu C, et al. α-Glucosidase inhibitors from Chinese bayberry (: Morella rubra Sieb. et Zucc.) fruit: Molecular docking and interaction mechanism of flavonols with different B-ring hydroxylations. *RSC Adv*. 2020;10(49):29347-29361. doi:10.1039/d0ra05015f
- 150. Sun J, Dong S, Wu Y, Zhao H, Li X, Gao W. Inhibitor discovery from pomegranate rind for targeting human salivary alpha-amylase. *Med Chem Res.* 2018;27(6):1559-1577. doi:10.1007/s00044-018-2164-2
- 151. Phuong NH, Thuy NTL, Duc NT, Tuyet NTA, Mai NTT, Phung NKP. A new glycoside and in vitro evalution of alpha-glucosidase inhibitory activity of constituents of the mangrove Lumnitzera racemosa. *Nat Prod Commun*. 2017;12(11):1751-1754. doi:10.1177/1934578x1701201125
- 152. Habtemariam S. alpha-Glucosidase inhibitory activity of kaempferol-3-O-rutinoside. *Nat Prod Commun*. 2011;6(2):201-203. doi:10.1177/1934578x1100600211
- 153. Wang Y, Xiang L, Wang C, Tang C, He X. Antidiabetic and Antioxidant Effects and Phytochemicals of Mulberry Fruit (Morus alba L.) Polyphenol Enhanced Extract. *PLoS One*. 2013;8(7):e71144. doi:10.1371/journal.pone.0071144
- 154. Vu NK, Kim CS, Ha MT, et al. Antioxidant and Antidiabetic Activities of Flavonoid Derivatives from the Outer Skins of Allium cepa L. *J Agric Food Chem.* 2020;68(33):8797-8811. doi:10.1021/acs.jafc.0c02122
- 155. Jibril S, Sirat HM, Basar N. Bioassay-guided isolation of antioxidants and alpha-glucosidase inhibitors from the root of Cassia sieberiana D.C. (Fabaceae). *Rec Nat Prod.* 2017;11(4):406-410.
- 156. Adhikari-Devkota A, Elbashir SMI, Watanabe T, Devkota HP. Chemical constituents from the flowers of Satsuma mandarin and their free radical scavenging and alpha-glucosidase inhibitory activities. *Nat Prod Res.* 2019;33(11):1670-1673. doi:10.1080/14786419.2018.1425856
- 157. Li YQ, Zhou FC, Gao F, Bian JS, Shan F. Comparative evaluation of quercetin, isoquercetin and rutin as inhibitors of alpha-glucosidase. *J Agric Food Chem.* 2009;57(24):11463-11468. doi:10.1021/jf903083h
- 158. Lyu Q, Kuo T-H, Sun C, Chen K, Hsu C-C, Li X. Comprehensive structural characterization of phenolics in litchi pulp using tandem mass spectral molecular networking. *Food Chem.* 2019;282:9-17. doi:10.1016/j.foodchem.2019.01.001
- 159. Praparatana R, Maliyam P, Barrows LR, Puttarak P. Flavonoids and Phenols, the Potential Anti-Diabetic Compounds from Bauhinia strychnifolia Craib. Stem. *Molecules*. 2022;27(8):2393. doi:10.3390/molecules27082393

- 160. Devkota HP, Kurizaki A, Tsushiro K, et al. Flavonoids from the leaves and twigs of Lindera sericea (Seibold et Zucc.) Blume var. sericea (Lauraceae) from Japan and their bioactivities. *Funct Foods Health Dis.* 2021;11(1):34-43. doi:10.31989/ffhd.v11i1.769
- 161. Manaharan T, Appleton D, Cheng HM, Palanisamy UD. Flavonoids isolated from Syzygium aqueum leaf extract as potential antihyperglycaemic agents. *Food Chem.* 2012;132(4):1802-1807. doi:10.1016/j.foodchem.2011.11.147
- 162. Shan-Shan Z, Niu-Niu Z, Sen G, et al. Glycosides and flavonoids from the extract of Pueraria thomsonii Benth leaf alleviate type 2 diabetes in high-fat diet plus streptozotocin-induced mice by modulating the gut microbiota. *Food Funct.* 2022;13(7):3931-3945. doi:10.1039/d1fo04170c
- 163. Jin DX, He JF, Zhang KQ, Luo XG, Zhang TC. α-Glucosidase Inhibition Action of Major Flavonoids Identified from Hypericum Attenuatum Choisy and Their Synergistic Effects. *Chem Biodivers*. 2021;18(10):e2100244. doi:10.1002/cbdv.202100244
- 164. Flores-Bocanegra L, Pérez-Vásquez A, Torres-Piedra M, Bye R, Linares E, Mata R. α-Glucosidase inhibitors from Vauquelinia corymbosa. *Molecules*. 2015;20(8):15330-15342. doi:10.3390/molecules200815330
- 165. Tasnuva ST, Qamar UA, Ghafoor K, et al. α-glucosidase inhibitors isolated from Mimosa pudica L. *Nat Prod Res*. 2019;33(10):1495-1499. doi:10.1080/14786419.2017.1419224
- 166. Wang W, Xu H, Chen H, Tai K, Liu F, Gao Y. In vitro antioxidant, anti-diabetic and antilipemic potentials of quercetagetin extracted from marigold (Tagetes erecta L.) inflorescence residues. *J Food Sci Technol*. 2016;53(6):2614-24. doi:10.1007/s13197-016-2228-6
- 167. Lian-Xin P, Li-Juan W, Qian Y, et al. In vitro potential of flavonoids from tartary buckwheat on antioxidants activity and starch digestibility. *Int J Food Sci Technol*. 2019;54(6):2209-2218. doi:10.1111/ijfs.14131
- 168. Fang H, Peng Z, Hao-Yue W, Gang-Xiu C, Zhong-Wen X, Guan-Hu B. Inhibition of alpha-glucosidase and alpha-amylase by flavonoid glycosides from Lu'an GuaPian tea: molecular docking and interaction mechanism. *Food Funct.* 2018;9(8):4173-4183. doi:10.1039/c8fo00562a
- 169. Hyun TK, Eom SH, Kim JS. Molecular docking studies for discovery of plant-derived α-glucosidase inhibitors. *Plant OMICS*. 2014;7(3):166-170.
- 170. Li N, Zhu HT, Wang D, Zhang M, Yang CR, Zhang YJ. New Flavoalkaloids with Potent α-Glucosidase and Acetylcholinesterase Inhibitory Activities from Yunnan Black Tea 'Jin-Ya'. *J Agric Food Chem*. 2020;68(30):7955-7963. doi:10.1021/acs.jafc.0c02401
- 171. Kim TH, Lee J, Kim HJ, Jo C. Plasma-Induced Degradation of Quercetin Associated with the Enhancement of Biological Activities. *J Agric Food Chem.* 2017;65(32):6929-6935. doi:10.1021/acs.jafc.7b00987
- 172. Islam MN, Jung HA, Sohn HS, Kim HM, Choi JS. Potent alpha-glucosidase and protein tyrosine phosphatase 1B inhibitors from Artemisia capillaris. *Arch Pharm Res.* 2013;36(5):542-552. doi:10.1007/s12272-013-0069-7
- 173. Arumugam B, Palanisamy UD, Chua KH, Kuppusamy UR. Potential antihyperglycaemic effect of myricetin derivatives from Syzygium malaccense. *J Funct Foods*. 2016;22:325-336. doi:10.1016/j.jff.2016.01.038
- 174. Qu X, Li J, Yan P, et al. Quercetin of Potentilla bifurca 3-Glycosylation Substitution Impact the Inhibitory Activity on alpha-glucosidase. *Pharmacogn Mag.* 2022;18(78):458-462. doi:10.4103/pm.pm 522 21
- 175. Wang L, Liu Y, Luo Y, Huang K, Wu Z. Quickly Screening for Potential α-Glucosidase Inhibitors from Guava Leaves Tea by Bioaffinity Ultrafiltration Coupled with HPLC-ESI-TOF/MS Method. *J Agric Food Chem*. 2018;66(6):1576-1582. doi:10.1021/acs.jafc.7b05280
- 176. Sun H, Li Y, Zhang X, et al. Synthesis, α-glucosidase inhibitory and molecular docking studies of prenylated and geranylated flavones, isoflavones and chalcones. *Bioorg Med Chem Lett.* 2015;25(20):4567-71. doi:10.1016/j.bmcl.2015.08.059
- 177. Liu Y, Wang R, Ren C, et al. Two Myricetin-Derived Flavonols from Morella rubra Leaves as Potent alpha Glucosidase Inhibitors and Structure-Activity Relationship Study by Computational Chemistry. *Oxid Med Cell Longev*. 2022;2022:9012943. doi:10.1155/2022/9012943
- 178. Kashchenko NI, Chirikova NK, Olennikov DN. Acylated Flavonoids from Spiraea Genus as Inhibitors of α-Amylase. *Russ J Bioorganic Chem.* 2018;44(7):876-886. doi:10.1134/S1068162018070051
- 179. Wang H, Du YJ, Song HC. alpha-Glucosidase and alpha-amylase inhibitory activities of guava leaves. *Food Chem.* 2010;123(1):6-13. doi:10.1016/j.foodchem.2010.03.088
- 180. Zhou Y, Jiang Q, Ma S, Zhou X. Effect of quercetin on the in vitro Tartary buckwheat starch digestibility. *Int J Biol Macromol.* 2021;183:818-830. doi:10.1016/j.ijbiomac.2021.05.013
- 181. Nickavar B, Amin G. Enzyme Assay Guided Isolation of an α-Amylase Inhibitor Flavonoid from Vaccinium arctostaphylos Leaves. *Iran J Pharm Res*. 2011;10(4):849-53.
- 182. Varghese GK, Bose LV, Habtemariam S. Antidiabetic components of Cassia alata leaves: Identification through alpha-glucosidase inhibition studies. *Pharm Biol.* 2013;51(3):345-349. doi:10.3109/13880209.2012.729066

- 183. Sohretoglu D, Sari S, Barut B, Ozel A. Discovery of potent alpha-glucosidase inhibitor flavonols: Insights into mechanism of action through inhibition kinetics and docking simulations. *Bioorg Chem.* 2018;79:257-264. doi:10.1016/j.bioorg.2018.05.010
- 184. Li R, Wang Q, Zhao M, Yang P, Hu X, Ouyang D. Flavonoid glycosides from seeds of Hippophae rhamnoides subsp. Sinensis with alpha-glucosidase inhibition activity. *Fitoterapia*. 2019;137:104248. doi:10.1016/j.fitote.2019.104248
- 185. Peng X, Zhang G, Liao Y, Gong D. Inhibitory kinetics and mechanism of kaempferol on alpha-glucosidase. *Food Chem.* 2016;190:207-215. doi:10.1016/j.foodchem.2015.05.088
- 186. Ajish KR, Antu KA, Riya MP, et al. Studies on alpha-glucosidase, aldose reductase and glycation inhibitory properties of sesquiterpenes and flavonoids of Zingiber zerumbet Smith. *Nat Prod Res.* 2015;29(10):947-952. doi:10.1080/14786419.2014.956741
- 187. Li S, Wang R, Hu X, Li C, Wang L. Bio-affinity ultra-filtration combined with HPLC-ESI-qTOF-MS/MS for screening potential α-glucosidase inhibitors from Cerasus humilis (Bge.) Sok. leaf-tea and in silico analysis. *Food Chem.* 2022;373:131528. doi:10.1016/j.foodchem.2021.131528
- 188. Li Q, Zhang X, Cao J, et al. Depside derivatives with anti-hepatic fibrosis and anti-diabetic activities from Impatiens balsamina L. flowers. *Fitoterapia*. 2015;105:234-239. doi:10.1016/j.fitote.2015.07.007
- 189. Chen J, Wu Y, Zou J, Gao K. α-Glucosidase inhibition and antihyperglycemic activity of flavonoids from Ampelopsis grossedentata and the flavonoid derivatives. *Bioorg Med Chem.* 2016;24(7):1488-94. doi:10.1016/j.bmc.2016.02.018
- 190. Li Z, Guowen Z, Suyun L, Deming G. Inhibitory mechanism of apigenin on alpha-glucosidase and synergy analysis of flavonoids. *J Agric Food Chem.* 2016;64(37):6939-6949. doi:10.1021/acs.jafc.6b02314
- 191. Yu Z, Jian-Nan MA, Chun-Li MA, Zhi QI, Chao-Mei MA. Simultaneous quantification of ten constituents of Xanthoceras sorbifolia Bunge using UHPLC-MS methods and evaluation of their radical scavenging, DNA scission protective, and α-glucosidase inhibitory activities. *Chin J Nat Med.* 2015;13(11):873-880. doi:10.3724/SP.J.1009.2015.00873
- 192. Williams LK, Li C, Withers SG, Brayer GD. Order and Disorder: Differential Structural Impacts of Myricetin and Ethyl Caffeate on Human Amylase, an Antidiabetic Target. *J Med Chem.* 2012;55(22):10177-10186. doi:10.1021/jm301273u
- 193. Yue Y, Chen Y, Geng S, Liang G, Liu B. Antioxidant and α-Glucosidase Inhibitory Activities of Fisetin. *Nat Prod Commun.* 2018;13(11):1489-1492. doi:10.1177/1934578x1801301119
- 194. Barber E, Houghton MJ, Williamson G. Flavonoids as human intestinal  $\alpha$ -glucosidase inhibitors. *Foods*. 2021;10(8):1939. doi:10.3390/foods10081939
- 195. Ahmed S, Al-Rehaily AJ, Alam P, et al. Antidiabetic, antioxidant, molecular docking and HPTLC analysis of miquelianin isolated from Euphorbia schimperi C. Presl. *Saudi Pharm J.* 2019;27(5):655-663. doi:10.1016/j.jsps.2019.03.008
- 196. Renda G, Sari S, Barut B, et al. α-Glucosidase inhibitory effects of polyphenols from Geranium asphodeloides: Inhibition kinetics and mechanistic insights through in vitro and in silico studies. *Bioorg Chem.* 2018;81:545-552. doi:10.1016/j.bioorg.2018.09.009
- 197. Tao Y, Chen Z, Zhang Y, Wang Y, Cheng Y. Immobilized magnetic beads based multi-target affinity selection coupled with high performance liquid chromatography-mass spectrometry for screening anti-diabetic compounds from a Chinese medicine "Tang-Zhi-Qing". *J Pharm Biomed Anal.* 2013;78:190-201. doi:10.1016/j.jpba.2013.02.024
- 198. Escandon-Rivera S, Gonzalez-Andrade M, Bye R, Linares E, Navarrete A, Mata R. alpha-Glucosidase inhibitors from Brickellia cavanillesii. *J Nat Prod.* 2012;75(5):968-974. doi:10.1021/np300204p
- 199. Olennikov DN, Kashchenko NI. Componential profile and amylase inhibiting activity of phenolic compounds from Calendula officinalis L. leaves. *Sci World J.* 2014;2014:654193. doi:10.1155/2014/654193
- 200. Ye R, Fan YH, Ma CM. Identification and Enrichment of alpha-Glucosidase-Inhibiting Dihydrostilbene and Flavonoids from Glycyrrhiza uralensis Leaves. *J Agric Food Chem*. 2017;65(2):510-515. doi:10.1021/acs.jafc.6b04155
- 201. Fan YH, Ye R, Xu HY, Feng XH, Ma CM. Structures and In Vitro Antihepatic Fibrosis Activities of Prenylated Dihydrostilbenes and Flavonoids from Glycyrrhiza uralensis Leaves. *J Food Sci.* 2019;84(5):1224-1230. doi:10.1111/1750-3841.14592
- 202. Ashraf J, Mughal EU, Sadiq A, et al. Design and synthesis of new flavonols as dual α-amylase and α-glucosidase inhibitors: Structure-activity relationship, drug-likeness, in vitro and in silico studies. *J Mol Struct*. 2020;1218:128458. doi:10.1016/j.molstruc.2020.128458

- 203. Wan C, Yuan T, Cirello AL, Seeram NP. Antioxidant and α-glucosidase inhibitory phenolics isolated from highbush blueberry flowers. *Food Chem.* 2012;135(3):1929-1937. doi:10.1016/j.foodchem.2012.06.056
- 204. Zhang L, Tu ZC, Yuan T, Wang H, Xie X, Fu ZF. Antioxidants and α-glucosidase inhibitors from Ipomoea batatas leaves identified by bioassay-guided approach and structure-activity relationships. *Food Chem.* 2016;208:61-7. doi:10.1016/j.foodchem.2016.03.079
- 205. Hong HC, Li SL, Zhang XQ, Ye WC, Zhang QW. Flavonoids with alpha-glucosidase inhibitory activities and their contents in the leaves of Morus atropurpurea. *Chin Med.* 2013;8(1):19. doi:10.1186/1749-8546-8-19
- 206. López-Angulo G, Miranda-Soto V, López-Valenzuela JA, et al. α-Glucosidase inhibitory phenolics from Echeveria subrigida (B. L. Rob & Seaton) leaves. *Nat Prod Res.* 2022;36(4):1058-1061. doi:10.1080/14786419.2020.1844695
- 207. Monzón Daza G, Meneses Macías C, Forero AM, et al. Identification of α-Amylase and α-Glucosidase Inhibitors and Ligularoside A, a New Triterpenoid Saponin from Passiflora ligularis Juss (Sweet Granadilla) Leaves, by a Nuclear Magnetic Resonance-Based Metabolomic Study. *J Agric Food Chem.* 2021;69(9):2919-2931. doi:10.1021/acs.iafc.0c07850
- 208. Zhang L, Tu ZC, Yuan T, et al. New gallotannin and other phytochemicals from sycamore maple (Acer pseudoplatanus) leaves. *Nat Prod Commun*. 2015;10(11):1977-1980. doi:10.1177/1934578x1501001143
- 209. Nile A, Gansukh E, Park GS, Kim DH, Hariram Nile S. Novel insights on the multi-functional properties of flavonol glucosides from red onion (Allium cepa L) solid waste In vitro and in silico approach. *Food Chem.* 2021;335:127650. doi:10.1016/j.foodchem.2020.127650
- 210. Tan C, Zuo J, Yi X, et al. Phenolic constituents from Sarcopyramis nepalensis and their α-glucosidase inhibitory activity. *Afr J Tradit Complement Altern Med*. 2015;12(3):156-160. doi:10.4314/ajtcam.v12i3.20
- 211. Şöhretoğlu D, Sari S, Šoral M, Barut B, Özel A, Liptaj T. Potential of Potentilla inclinata and its polyphenolic compounds in α-glucosidase inhibition: Kinetics and interaction mechanism merged with docking simulations. *Int J Biol Macromol*. 2018;108:81-87. doi:10.1016/j.ijbiomac.2017.11.151
- 212. Braham N, Phi-Hung N, Bing-Tian Z, Quoc-Hung V, Byung Sun M, Mi Hee W. Protein tyrosine phosphatase 1B (PTP1B) inhibitory activity and glucosidase inhibitory activity of compounds isolated from Agrimonia pilosa. *Pharm Biol.* 2016;54(3):474-480. doi:10.3109/13880209.2015.1048372
- 213. Tao Y, Zhang Y, Cheng Y, Wang Y. Rapid screening and identification of α-glucosidase inhibitors from mulberry leaves using enzyme-immobilized magnetic beads coupled with HPLC/MS and NMR. *Biomed Chromatogr*. 2013;27(2):148-55. doi:10.1002/bmc.2761
- 214. Marmouzi I, Ezzat SM, Mostafa ES, et al. Isolation of secondary metabolites from the mediterranean sponge species; Hemimycale columella and its biological properties. *SN Appl Sci*. 2021;3(2):207. doi:10.1007/s42452-020-04052-8
- 215. Zhang Y, Xiao G, Sun L, Wang Y, Wang Y, Wang Y. A new flavan-3-ol lactone and other constituents from Euonymus alatus with inhibitory activities on a-glucosidase and differentiation of 3T3-L1 cells. *Nat Prod Res.* 2013;27(17):1513-1520. doi:10.1080/14786419.2012.725400
- 216. Fan P, Terrier L, Hay AE, Marston A, Hostettmann K. Antioxidant and enzyme inhibition activities and chemical profiles of Polygonum sachalinensis F.Schmidt ex Maxim (Polygonaceae). *Fitoterapia*. 2010;81(2):124-131. doi:10.1016/j.fitote.2009.08.019
- 217. Yan S, Zhang X, Wen X, et al. Purification of Flavonoids from Chinese Bayberry (Morella rubra Sieb. et Zucc.) Fruit Extracts and alpha-Glucosidase Inhibitory Activities of Different Fractionations. *Molecules*. 2016;21(9):1148. doi:10.3390/molecules21091148
- 218. Sheliya MA, Rayhana B, Ali A, et al. Inhibition of alpha-glucosidase by new prenylated flavonoids from euphorbia hirta L. herb. *J Ethnopharmacol*. 2015;176:1-8. doi:10.1016/j.jep.2015.10.018
- 219. Anh LTT, Son NT, Van Tuyen N, et al. Antioxidative and α-glucosidase inhibitory constituents of Polyscias guilfoylei: experimental and computational assessments. *Mol Divers*. 2022;26(1):229-243. doi:10.1007/s11030-021-10206-6
- 220. Lee HE, Kim JA, Whang WK. Chemical Constituents of Smilax china L. Stems and Their Inhibitory Activities against Glycation, Aldose Reductase, alpha-Glucosidase, and Lipase. *Molecules*. 2017;22(3):451. doi:10.3390/molecules22030451
- 221. Parveen A, Farooq MA, Kyunn WW. A new oleanane type saponin from the aerial parts of nigella sativa with anti-oxidant and anti-diabetic potential. *Molecules*. 2020;25(9):2171. doi:10.3390/molecules25092171
- 222. Astiti MA, Jittmittraphap A, Leaungwutiwong P, et al. LC-QTOF-MS/MS based molecular networking approach for the isolation of α-glucosidase inhibitors and virucidal agents from coccinia grandis (L.) voigt. *Foods*. 2021;10(12):3041. doi:10.3390/foods10123041

- 223. Tan C, Wang Q, Luo C, Chen S, Li Q, Li P. Yeast alpha-glucosidase inhibitory phenolic compounds isolated from Gynura medica leaf. *Int J Mol Sci.* 2013;14(2):2551-2558. doi:10.3390/ijms14022551
- 224. Swilam N, Nawwar MAM, Radwan RA, Mostafa ES. Antidiabetic Activity and In Silico Molecular Docking of Polyphenols from Ammannia baccifera L. subsp. Aegyptiaca (Willd.) Koehne Waste: Structure Elucidation of Undescribed Acylated Flavonol Diglucoside. *Plants*. 2022;11(3):452. doi:10.3390/plants11030452
- 225. Yoshikawa M, Shimada H, Nishida N, et al. Antidiabetic principles of natural medicines. II. Aldose reductase and α-glucosidase inhibitors from brazilian natural medicine, the leaves of Myrcia multiflora DC. (Myrtaceae): Structures of myrciacitrins I and II and myrciaphenones A and B. *Chem Pharm Bull (Tokyo)*. 1998;46(1):113-119. doi:10.1248/cpb.46.113
- 226. Oueslati MH, Bouajila J, Guetat A, Al-Gamdi F, Hichri F. Cytotoxic, alpha-Glucosidase, and Antioxidant Activities of Flavonoid Glycosides Isolated from Flowers of Lotus lanuginosus Vent. (Fabaceae). *Pharmacogn Mag.* 2020;16(68):22-27. doi:10.4103/pm.pm 232 19
- 227. Chen YG, Li P, Li P, et al. α-glucosidase inhibitory effect and simultaneous quantification of three major flavonoid glycosides in microctis folium. *Molecules*. 2013;18(4):4221-4232. doi:10.3390/molecules18044221
- 228. Chen JG, Wu SF, Zhang QF, Yin ZP, Zhang L. α-Glucosidase inhibitory effect of anthocyanins from Cinnamomum camphora fruit: Inhibition kinetics and mechanistic insights through in vitro and in silico studies. *Int J Biol Macromol*. 2020;143:696-703. doi:10.1016/j.ijbiomac.2019.09.091
- 229. Akkarachiyasit S, Charoenlertkul P, Yibchok-Anun S, Adisakwattana S. Inhibitory activities of cyanidin and its glycosides and synergistic effect with acarbose against intestinal α-glucosidase and pancreatic α-amylase. *Int J Mol Sci.* 2010;11(9):3387-96. doi:10.3390/ijms11093387
- 230. Ho GT, Kase ET, Wangensteen H, Barsett H. Phenolic Elderberry Extracts, Anthocyanins, Procyanidins, and Metabolites Influence Glucose and Fatty Acid Uptake in Human Skeletal Muscle Cells. *J Agric Food Chem.* 2017;65(13):2677-2685. doi:10.1021/acs.jafc.6b05582
- 231. Kim JH, Kim HY, Jin CH. Mechanistic investigation of anthocyanidin derivatives as alpha-glucosidase inhibitors. *Bioorg Chem.* 2019;87:803-809. doi:10.1016/j.bioorg.2019.01.033
- 232. Xu Y, Xie L, Xie J, Liu Y, Chen W. Pelargonidin-3-O-rutinoside as a novel α-glucosidase inhibitor for improving postprandial hyperglycemia. *Chem Commun (Camb)*. 2018;55(1):39-42. doi:10.1039/c8cc07985d
- 233. Homoki JR, Nemes A, Fazekas E, et al. Anthocyanin composition, antioxidant efficiency, and α-amylase inhibitor activity of different Hungarian sour cherry varieties (Prunus cerasus L.). *Food Chem.* 2016;194:222-9. doi:10.1016/j.foodchem.2015.07.130
- 234. Nickavar B, Amin G. Bioassay-guided separation of an alpha-amylase inhibitor anthocyanin from Vaccinium arctostaphylos berries. *Z Naturforsch C J Biosci*. 2010;65(9):567-70. doi:10.1515/znc-2010-9-1006
- 235. Zhuang C, Zhang W, Sheng C, Zhang W, Xing C, Miao Z. Chalcone: A Privileged Structure in Medicinal Chemistry. *Chem Rev.* 2017;117(12):7762-7810. doi:10.1021/acs.chemrev.7b00020
- 236. Tran T-D, Nguyen T-T-N, Do T-H, Huynh T-N-P, Tran C-D, Thai K-M. Synthesis and Antibacterial Activity of Some Heterocyclic Chalcone Analogues Alone and in Combination with Antibiotics. *Molecules*. 2012;17(6):6684-6696. doi:10.3390/molecules17066684
- 237. Rocha S, Sousa A, Ribeiro D, et al. A study towards drug discovery for the management of type 2 diabetes mellitus through inhibition of the carbohydrate-hydrolyzing enzymes α-amylase and α-glucosidase by chalcone derivatives. *Food Funct*. 2019;10(9):5510-5520. doi:10.1039/c9fo01298b
- 238. Saleem F, Kanwal, Khan KM, et al. Synthesis of azachalcones, their α-amylase, α-glucosidase inhibitory activities, kinetics, and molecular docking studies. *Bioorg Chem.* 2021;106:104489. doi:10.1016/j.bioorg.2020.104489
- 239. Mphahlele MJ, Agbo EN, Choong YS. Synthesis, Structure, Carbohydrate Enzyme Inhibition, Antioxidant Activity, In Silico Drug-Receptor Interactions and Drug-Like Profiling of the 5-Styryl-2-Aminochalcone Hybrids. *Molecules*. 2021;26(9):2692. doi:10.3390/molecules26092692
- 240. Miadoková E. Isoflavonoids an overview of their biological activities and potential health benefits. *Interdiscip Toxicol*. 2010;2(4):211-218. doi:doi:10.2478/v10102-009-0021-3
- 241. Şöhretoğlu D, Sari S, Özel A, Barut B. α-Glucosidase inhibitory effect of Potentilla astracanica and some isoflavones: Inhibition kinetics and mechanistic insights through in vitro and in silico studies. *Int J Biol Macromol*. 2017;105:1062-1070. doi:10.1016/j.ijbiomac.2017.07.132
- 242. Ha LM, Luyen NT, Phuong NT, et al. Isoflavonoids from desmodium heterophyllum aerial parts. *Nat Prod Commun*. 2018;13(6):699-700. doi:10.1177/1934578x1801300612
- 243. Seong SH, Roy A, Jung HA, Jung HJ, Choi JS. Protein tyrosine phosphatase 1B and alpha-glucosidase inhibitory activities of Pueraria lobata root and its constituents. *J Ethnopharmacol*. 2016;194:706-716. doi:10.1016/j.jep.2016.10.007

- 244. Jiang W, Kan H, Li P, Liu S, Liu Z. Screening and structural characterization of potential α-glucosidase inhibitors from Radix Astragali flavonoids extract by ultrafiltration LC-DAD-ESI-MSn. *Anal Methods*. 2015;7(1):123-128. doi:10.1039/c4ay02081b
- 245. Jeong SY, Nguyen PH, Zhao BT, et al. Chemical constituents of Euonymus alatus (Thunb.) Sieb. and their PTP1B and alpha-glucosidase inhibitory activities. *Phytother Res.* 2015;29(10):1540-1548. doi:10.1002/ptr.5411
- 246. Yue T, Sheng Q, Luo Y, et al. Biflavonoids and Oligomeric Flavonoids from Food. In: Xiao J, Sarker SD, Asakawa Y, eds. *Handbook of Dietary Phytochemicals*. Springer Singapore; 2021:155-203.
- 247. Laishram S, Sheikh Y, Moirangthem DS, et al. Anti-diabetic molecules from Cycas pectinata Griff. traditionally used by the Maiba-Maibi. *Phytomedicine*. 2015;22(1):23-6. doi:10.1016/j.phymed.2014.10.007
- 248. Silva JP, Coelho A, Paula A, et al. The Influence of Irrigation during the Finishing and Polishing of Composite Resin Restorations—A Systematic Review of In Vitro Studies. *Materials*. 2021;14(7):1675. doi:10.3390/ma14071675
- 249. Li C, Begum A, Numao S, Park KH, Withers SG, Brayer GD. Acarbose Rearrangement Mechanism Implied by the Kinetic and Structural Analysis of Human Pancreatic α-Amylase in Complex with Analogues and Their Elongated Counterparts. *Biochemistry*. 2005;44(9):3347-3357. doi:10.1021/bi048334e
- 250. Williams LK, Zhang X, Caner S, et al. The amylase inhibitor montbretin A reveals a new glycosidase inhibition motif. *Nat Chem Biol*. 2015;11(9):691-696. doi:10.1038/nchembio.1865
- 251. Tysoe CR, Caner S, Calvert MB, Win-Mason A, Brayer GD, Withers SG. Synthesis of montbretin A analogues yields potent competitive inhibitors of human pancreatic α-amylase. 10.1039/C9SC02610J. *Chem Sci.* 2019;10(48):11073-11077. doi:10.1039/C9SC02610J
- 252. Pyner A, Nyambe-Silavwe H, Williamson G. Inhibition of Human and Rat Sucrase and Maltase Activities To Assess Antiglycemic Potential: Optimization of the Assay Using Acarbose and Polyphenols. *J Agric Food Chem.* 2017;65(39):8643-8651. doi:10.1021/acs.jafc.7b03678
- 253. Lim J, Ferruzzi MG, Hamaker BR. Structural requirements of flavonoids for the selective inhibition of α-amylase versus α-glucosidase. *Food Chem.* 2022;370:130981. doi:10.1016/j.foodchem.2021.130981