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Abstract

This review summarizes mainly the activity of our labs. We established more than one laboratory for materials synthesis, characterization, and applications. Our laboratories provide the synthesis of several nanoparticles including metal oxide nanoparticles (e.g., Fe₃O₄, ZnO, ZrOSO₄, MoO_{3-x}, CuO, AgFeO₂, Co₃O₄, SiO₂, and CuFeO₂), metallic nanoparticles (Ag, Au, Pd, and Pt), carbon-based nanomaterials (graphene, graphene oxide, reduced graphene oxide, and carbon dots (CDs)), biopolymers (cellulose, nanocellulose, TOCNF, alginate, and chitosan), organic polymers (conjugated polymers, covalent-organic frameworks (COFs), and intrinsic microporous polymers), and hybrid materials e.g. metal-organic frameworks (MOFs). These materials were applied for energy, environmental, and biomedicine applications. They were applied in several fields such as environmental-based technologies (e.g., water remediation, air purification, gas storage), energy (production of hydrogen, dimethyl ether, solar cells, and supercapacitors), and biomedical sectors (sensing/biosensing, cancer therapy, and drug delivery). They can act as efficient adsorbents and catalysts to remove emerging contaminants such as metals, dyes, drugs, antibiotics, pesticides, and oils in water via adsorption. They can be also used as catalysts for catalytic degradation, reduction, and oxidation of organic pollutants. They can be used as filters for air purification by removing greenhouse gases such as carbon dioxide (CO₂), volatile organic compounds (VOCs), and particulate matter (PMs). They can be used for hydrogen production via water splitting, alcohol oxidation, and hydrolysis of NaBH₄. Biomedical applications such as antibacterial, drug delivery, and biosensing were also involved.

Keywords: Materials; MOFs; Energy; Environmental; Biomedicine.

Introduction

Materials are objects containing a mixture of substances. They can be classified based on several strategies including 1) physical and chemical properties; 2) origin (e.g., natural or synthetic); and 3) biological function. Natural materials can be prepared using raw materials via several procedures including purification, extraction, and shaping. On the other side, synthetic materials can be prepared via several procedures. Human classified their prehistory based on material types into; Stone Age, Bronze Age, and Iron Age. The steel age, plastic age, and silicon age were named for the 19th century, the middle of the 20th century, and the second half of the 20th century, respectively. Materials advanced several applications including energy [1–4], environmental, analytical techniques [5–10], and biomedical applications [11–14].

We have established an advanced multifunctional materials laboratory (**Figure 1**). Our lab, an advanced multifunctional materials laboratory, can synthesize, characterize, and investigate applications for several fields. We can synthesize materials such as:-

- Metal oxide nanoparticles, e.g., Fe₃O₄, ZnO, ZrOSO₄, MoO_{3-x}, CuO, CeO₂, AgFeO₂, Co₃O₄, SiO₂, and CuFeO₂.
- 2) Metallic nanoparticles, e.g., Ag, Au, Pd, and Pt.
- Carbon-based nanomaterials, e.g., graphene, graphene oxide, reduced graphene oxide, and carbon dots (CDs).
- 4) Biopolymers e.g., cellulose, nanocellulose, TOCNF, alginate, and chitosan.
- 5) Organic polymers e.g., conjugated polymers, covalent-organic frameworks (COFs), and intrinsic microporous polymers.
- 6) Ionic liquids (Ils) [15–21].
- 7) Metallodrugs [22].

8) Hybrid materials e.g., metal-organic frameworks (MOFs).

We can do characterization using techniques such as X-ray diffraction (XRD), X-ray photoelectron spectroscopy (XPS), transmission electron microscopy (TEM), high-resolution TEM (HR-TEM), scanning electron microscopy (SEM), energy dispersive X-ray (EDX) analysis and mapping, atomic absorption flame spectroscopy (AAFS), UV-Vis spectroscopy, diffuse reflectance spectroscopy (DRS), and electrochemical measurements (cyclic voltammetry (CV), linear sweep voltammetry (LSV), electrochemical impedance spectroscopy (EIS), galvanostatic charge-discharge, differential pulse voltammetry (DPV), and Mott-Schottky (MS).



Figure 1 Advanced Multifunctional Materials Laboratory established by Dr. Hani Nasser Abdelhamid.

We can test any materials for several applications. Most of these applications are summarized

as shown in Figure 1. We can report full analysis for applications such as:-

- 1. Energy:
 - a) Hydrogen generation via hydrolysis of sodium borohydride (NaBH₄).
 - b) Photocatalytic water splitting for hydrogen generation.
 - c) Photocatalytic alcohol oxidation for hydrogen generation and carbonyl compounds synthesis.
 - d) Supercapacitors.

- e) Lithium-ion Battery.
- f) Dye-sensitizing solar cells (DSSCs) [23].

2. Environmental:-

- a) Water treatment via pollutants removal e.g., adsorption and degradation.
- b) Air purification; removal of greenhouse gases via adsorption.
- c) Adsorption of volatile organic compounds (VOCs).
- d) Photocatalytic degradation of drugs, antibiotics, and pharmaceuticals.
- e) Heavy metal removal via adsorption.
- f) Precious metal recovery.

3. Biomedical Applications:-

- a) Cancer therapy; chemotherapy, photodynamic, and photothermal.
- b) Drug delivery [24,25].
- c) Gene delivery using cell-penetrating peptides (CPPs)[26,27].
- d) Antimicrobial agents; antibacterial, and antifungal [28–33].
- e) Nanotoxicity and Environmental fate for nanoparticles [34–36].
- f) Bone regeneration.
- g) Wound healing.
- h) Tissue Engineering.
- i) Nanozymes and MOFZyme (artificial enzyme based on MOFs materials)
- j) Biosensing of biomarkers, biological heavy metals, enzymes, and proteins.
- k) Detection and analysis of pathogenic bacteria.
- 1) Proteomics and clinical research [37].
- m) Synthesis of biologically active compounds [38,39].
- n) Investigate effective matrix for matrix-assisted laser desorption ionization mass spectrometry [40,41].

Metallic Nanoparticles

Metallic nanoparticles e.g., silver (Ag), gold (Au), palladium (Pd)[42], and platinum (Pt), advanced several applications. Silver nanoparticles have been used for many applications such as catalysis [43], energy [44], biosensing [45], laser desorption/ionization mass spectrometry (LDI-MS) and mass spectrometry imaging (MSI) [46], and others [47]. In our lab, we investigated Ag NPs' antimicrobial activity against bacterial flora of bull semen [48]. AgFeO₂ exhibit high antibacterial activity against several bacteria species [49,50]. Ag NPs were used as a probe for the detection of the freshness of fruits and vegetables via graphene-enhanced Raman spectroscopy (GERS) [51]. Silver nanoparticles can be used as a surface for microextraction proteins and other analytes for the analysis using surface-assisted laser desorption-ionization mass spectrometry (SALDI-MS) [52]. It can be also modified with chitosan for the separation and detection of biothiols [53].

The spermicidal effects of Ag NPs against flora bacteria were reported [48]. Silver salts were mixed with melamine. The mixture was then polymerized at 550 °C to generate graphitic carbon-embedded Ag NPs i.e. Ag@C NPs. Analytical techniques such as XRD, XPS, AAFS, TEM, and HR-TEM confirm the material's phases, composition, morphology, and particle size. Ag@C NPs display a particle size of 1-5 nm with an average particle size of 2.5 nm. The nanoparticles were embedded into carbon. Ag@C NPs were investigated as antimicrobial agents in bacteriospermia of fresh semen collected from five fertile bulls. They exhibited high antibacterial activity against bacteria species found in semen such as *Escherichia coli (E. Coli)*, *Staphylococcus aureus (S. aureus)*, and *Pseudomonas aeruginosa (P. aeruginosa)*. It offered minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) of 3.125-12.5 μ g/mL and 3.125 μ g/mL, respectively. There was no detrimental effect (P > 0.05) on the percentage of sperm motility, plasma membrane integrity, acrosome integrity, and normal sperm morphology at concentrations of 15-30 μ g/mL. Ag@C NPs is a promising

antibiotic agent for bull semen extender during cold storage. It can be used in applications such as the field of artificial insemination [48]. The antibacterial activity of silver ferrite (AgFeO₂) was investigated. AgFeO₂ was modified with polyethylene glycols (PEGs) to render their dispersion high [49,50]. The antibacterial activity against pathogenic bacteria was quantified using plate counting, and the turbidity using optical density at wavelength 600 nm (OD₆₀₀). AgFeO₂ nanoparticles exhibited high antibacterial activity [49,50].

Silver nanoparticles were modified with 1-octadecanethiol (1-ODT)/4-aminothiophenol (4-AMP) and 1-ODT/1-thioglycerol (1-TG) to prepare Ag@ODT/AMP and Ag@ODT/TG, respectively [52]. The materials were used in microextraction as a pseudo-stationary phase via single-drop microextraction (SDME). They can extract proteins and peptides e.g., insulin, ubiquitin, t cytochrome c, cysteine, homocysteine, and lysozyme. The separated proteins can be detected after extraction using matrix-assisted laser desorption/ionization mass spectrometry (MALDI-MS). The method can be used for the analysis of real samples e.g., urine and milk [52]. Silver ferrite iron oxide nanoparticles (AgFeO₂ NPs) were reported for biothiols separation [53]. AgFeO₂ and AgFeO₂ modified chitosan (AgFeO₂@CTS NPs) can be used for the separation of biological thiols e.g., sulfamethizole, thiabendazole, dithiothreitol, and glutathione before the analysis using MALDI-MS and surface assisted laser desorption/ionization mass spectrometry (SALDI–MS) [53].

Au NPs enhanced GERS detection of the freshness of fruits and vegetables [51]. It can be used as a probe for surface-enhanced Raman spectroscopy (SERS, **Figure 2**). Au or Ag nanoparticles were synthesized into reduced graphene oxide nanosheets (e.g., Au@G and Ag@G). The materials can be used as a probe for the analysis of the freshness of fruits and vegetables (e.g., Carrot, Wax apple, Lemon, Red pepper, and Tomato) [51]. One-pot synthesis of Au NPs@carbon dots was reported for the cytosensing of metals in cancer cells [54]. Au NPs enhanced the analysis of simple molecules to intact cells using SALDI-MS [55].

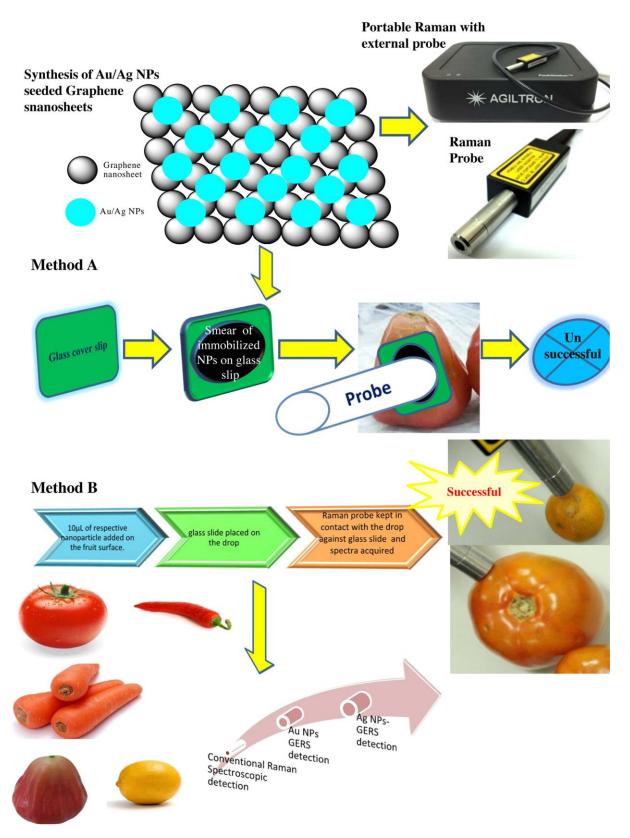


Figure 2 Schematic representation for GERS analysis of fruits and vegetables using Ag and Au NPs. Figure reprinted with permission from Ref. [51].

Carbon Nanomaterials and their Applications

Zero-dimension carbon can be also known as carbon dots (CDs), carbon nanodots (C NDs), or carbon quantum dots (CQDs) [56–62]. CDs were applied for several promising applications such as drug delivery [63], imaging [64–68], sensing [57,69–71], biosensing [78], energy-based applications [77], biomedical [72], and theranostic [73]. Carbon nanodots, including carbon dots and graphene quantum dots, carbon quantum dots (C QDs), or carbon dots (CDs), are emerging new carbon allotropes nanomaterials [74–76]. Carbon nanomaterials have advanced electrochemical-based applications [68,79]. CDs have advanced electrochemical applications [80,81] such as O₂ and H₂O₂ reduction [82], and biosensing of glucose [83–89].

C-dots can be doped with P [90], N [91], S [67], F [92], B [93], nitrogen and sulfur co-doped carbon dots (N, S-CDs) [94,95], and N/B [96]. C dots exhibit good optical properties including photoluminescence in the visible range [97,98], and high quantum yields (QY)[58,99]. The photoluminescence properties of CDs can be tuneable by changing their size, surface modification with functional groups at the graphitic edges of the materials, doping with heteroatoms, or selecting a suitable synthesis method [100,101]. They can be tuned offering fluorescence emission from blue to green [90,102]. It has been used for tackling COVID-19 [103], the virus [104]. It offered naked eye sensors [105]. N-doped CDs especially exhibit remarkable acid-evoked fluorescence enhancement under acidic conditions [106].

Two-dimensional carbon nanomaterials such as graphene, graphene oxide (GO), and reduced graphene oxide were intensively used for several applications. Graphene oxide was used for rare-earth metal adsorption [107]. It can be modified with thymine for selective detection of toxic heavy metals such as mercury (Hg(II)) [108]. The layer structure of GO enables the intercalation of an organic matrix such as sinapinic acid [109]. GO can be modified with SiO₂ for SALDM-MS [110]. It can use for heavy metal detection such as mercury ions [111], lipids

[112], and metallodrugs [113]. It exhibited high efficiency for bone and skin wound regeneration [114] and wound healing [115]. It can use for the drug delivery of in-soluble antibiotics such as gramicidin [116]. It can be used as a co-carrier to enhance the gene transfection of CPPs [117]. GO/cellulose nanocomposite accelerated skin wound healing [118]. Graphene can be used as a surface for SALDI-MS [119].

Metal Oxides

Metal oxides such as CeO₂ enabled the extraction and detection of pathogens proteins [120]. Fe₃O₄@ SiO₂ enabled rapid and direct identification of pathogenic bacteria from blood using [121]. Magnetic nanoparticles modified graphene oxide was reported for separation and preconcentration of pathogenic bacteria for sensitive detection using MALDI-MS [122]. Chitosan magnetic nanoparticles were reported for endotoxin separation and detection using SALDI-MS [123]. ZnO nanoparticle-modified polymethyl methacrylate was used for dispersive liquid–liquid microextraction for rapid analysis of pathogenic bacteria using MALDI-MS [124]. SnO₂@GO exhibited high antibacterial activity [125]

Commercial MoO₃ was used for the exfoliation to synthesize a few layers of MoO_{3-x} (**Figure 3**) [126]. The synthesis procedure involved the reflux of a bulk α -MoO₃ at 80 °C in water for 7 days. The prepared MoO_{3-x} nanosheets displayed infrared plasmonic properties offering localized surface plasmon resonance (LSPR) peaks at 954 and 1160 nm due to the oxygen vacancies upon light excitation. The plasmonic properties of the nanosheets can be enhanced using visible light irradiation for only 10 min. The materials were used as photocatalysts for dye degradation under visible light irradiation [126].

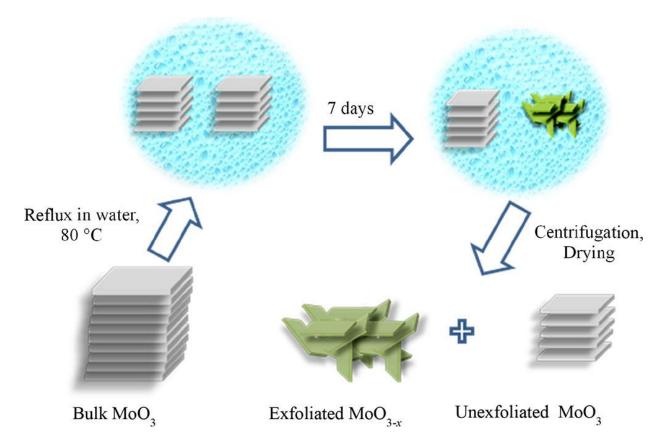


Figure 3 The Exfoliation of α -MoO₃ into MoO_{3-x} Nanosheets. Figure reprinted from Ref. [126]. This is an Open Access Article. Copyright belongs to the American Chemical Society (ACS).

Ruthenium oxide (RuO₂) with mesopore was synthesized via a surfactant-assisted procedure [127]. The materials exhibited higher catalytic oxidation activity of water using ceric ammonium nitrate (CAN).

Magnetic nanoparticles can be synthesized via several procedures including laser techniques [128]. Abdelhamid reviewed the application of delafossite nanoparticles in energy, nanomedicine, and environmental applications [129]. Magnetic nanoparticles of Fe₃O₄ were incorporated into polyplexes of CPPs/oligonucleotides (ONs) for cell transfection [130]. Three different oligonucleotides (e.g., plasmid (pGL3), splicing correcting oligonucleotides (SCO), and small interfering RNA (siRNA)) and six CPPs (e.g. PeptFect220 (denoted PF220), PF221, PF222, PF223, PF224, and PF14) were investigated. Magnetic nanoparticles enhanced the cell

transfection up to 4-fold compared to the noncovalent PF14-SCO complex, which exhibited higher efficiency compared to a commercial vector called Lipofectamine[™]2000 [130].

Quantum Dots (QDs)

Quantum dots (QDs) are nanocrystals with particle sizes less than 10 nm [131]. Cadmium sulfide (CdS) quantum dots were used for selective biosensing of *Staphylococcus aureus* [132] and proteomics [133,134]. It can be used as a surface for SALDI-MS analysis of several analytes [135]. It enabled soft ionization offering the analysis of labile compounds such as metallodrugs [136]. It can also be used for fluorescence spectroscopy[137]. CdS QDs were insitu grown into chitosan (CTS) enabling CdS QDs@CTS [138,139]. The material CdS@CTS exhibited selective interaction with Cu^{2+} due to the formation of $Cd_{1-x}Cu_xS$ [138,139]. The positive charge on chitosan exhibited also high interaction with the negative charge on the bacteria cell membranes [140]. CdS@CTS was also reported as a carrier for drug delivery of a natural anticancer drug called sesamol [141].

Biopolymers

Biopolymers including polysaccharides are intensively applied for biomedical applications [142,143]. Polysaccharides were applied as excipients for tablet formulation, dental implants, bone/tissue engineering, and drug delivery [142,143]. They can also be used for antimicrobial textiles [144–146]. Silver ferrite (AgFeO₂) can be modified with chitosan to render their external surface positive for biothiol separation [53]. Alginate can improve the gene delivery of oligonucleotides [147,148]. Modern technology such as 3D printing enabled simple processing of polylactic acid and hydroxyapatite for water treatment [149].

Cellulose-based advanced several applications such as biomedicine including antifouling [150–153]. They improved bioengineering [154] and water treatment via pollutants adsorption

[155]. Cellulose/ZIF-8 composite was used for water remediation via adsorption and catalytic degradation of organic pollutants such as dyes [156]. Cellulose enabled three-dimensional printing of porous materials such as leaf-like zeolitic imidazolate frameworks (ZIF-L), denoted as CelloZIF-L. Direct ink writing (DIW) or robocasting was used to proceed with the materials. The materials with a ZIF content of 84% were achieved. The materials were used for the adsorption of carbon dioxide (CO₂) and heavy metals offering capacities of 0.64-1.15 mmol/g (at 1 bar, 0 °C) and 554.8±15 mg/g, respectively. The adsorbent exhibited selectivity toward Fe³⁺, Al³⁺, Co²⁺, Cu²⁺, Na⁺, and Ca²⁺ of 86.8%, 6.7%, 2.4%, 0.93%, 0.61%, and 0.19%, respectively [157]. Cellulose enabled also the processing of ZIF materials into filter paper [158,159] and foams [160]. Most of these biopolymers are biodegradable [161,162] compared to synthetic polymers [163]. They can proceed into the membrane for oil separation [161].

Chitosan improved gene delivery [164]. It can stabilize magnetic nanoparticles that enabled high-cell transfection [165]. Magnetic nanoparticles modified chitosan was used for surfactant capture and analysis using SALDI-MS [166]. Chitosan can be modified with thymine to enable specific preconcentration of mercury (II) before analysis using SELDI-MS [167]. It can be used as a porogen for creating mesopores inside microporous materials [168]. The created hierarchical porous materials can be then used for oligonucleotide delivery offering efficient gene treatment. Chitosan mitigates the toxicity of CdS QDs offering efficient drug delivery of the anticancer drug sesamol [141].

Metal-organic frameworks (MOFs)

MOFs advanced several applications including biosensing [169–172]. Lanthanide MOF was reported for the detection of ferric ions and vitamin C [173]. The material was stable and can form high dispersion with high fluorescence emission signals. Fe(III) ions can selectively quench the fluorescence signal enabling a linear relationship in the concentration range of

16.6–167 μ M with a limit of detection (LOD) of 16.6 μ M (S/N ratio of >3) [173]. Explosive materials such as nitroaromatic was detected using Zn-MOF [174].

A composite of hierarchical porous bimetallic of (Co, Zn)-ZIF-8, and semiconductor photocatalyst TiO₂ (Co@ZIF-8/TiO₂) was reported for hydrogen generation via photocatalytic water splitting [175]. Co@ZIF-8/TiO₂ showed a photocatalytic hydrogen generation rate of 13 mmol• h^{-1} •g⁻¹ representing a 151-fold high catalytic performance of pristine TiO₂ [175]. Co@ZIF-8 improved also hydrogen generation via the hydrolysis of NaBH₄ [176]. Carbonized MOF enabled selective dehydrogenation of isopropanol [177].

We reported several procedures to prepare hierarchical porous zeolitic imidazolate frameworks (ZIFs)[178,179]. Template-free and template-based procedures were reported [180]. Dye encapsulation and one-pot synthesis of hierarchical porous (microporous–mesoporous) ZIF-8 were reported for CO₂ sorption and adenosine triphosphate biosensing [181]. A cobalt ZIF material, ZIF-67, was used for hydrogen generation via the hydrolysis of NaBH₄ [182,183]. The generated hydrogen can be used for dye degradation [182]. ZIFs-based materials were reviewed as efficient adsorbents and catalysts for CO₂ removal via adsorption and conversion into value-added compounds [184–188]. ZIF-8 and ZIF-67 can be in-situ grown into cellulosic filter paper that was used as an efficient catalyst for the reduction of water pollutants such as nitrophenols [189]. Our synthesis procedures offered several advantages including the formation of a hierarchical porous structure with fast and potential to use for large-scale production [190].

ZIFs materials including ZIF-8 are biocompatible materials [191]. Thus, ZIF-8 was widely used for biomedical applications [192] including gene delivery [193]. However, our recent study showed the transfer of the metal ions into the environment that caused a significant effect on the colonization and decomposition of shaded outdoor mice carrions by arthropods [194].

A zirconium-based MOF, UiO-66, can enhance bone generation offering induction of bone defects in rabbit femoral condyles [195]. UiO-66 catalyzed the hydrogen formation via the hydrolysis of NaBH₄ [196]. It was also reported as a precursor for the synthesis of ZrOSO₄@C for hydrogen generation [197] and dimethyl ether formation [198].

A cerium MOF (Ce-MOF) exhibited Fenton-like properties that enabled catalytic oxidation of olefins, alcohol, and dyes degradation [199]. It offered 100% and 53% conversion of cinnamyl alcohol and styrene, respectively. It provided high selectivity of 75% and 100% towards styrene oxide and benzaldehyde, respectively. It can catalytically degrade organic pollutants such as dyes [199]. Ce-MOF was also used probe for fluorescence detection of ferric ions and hydrogen peroxide [200], and MOFZyme for the inhibition of fungi [201,202].

A copper-based MOF (Cu and 1,4-benzene dicarboxylic acid as metal nodes and linker, respectively) was in-situ grown into the fiber of cotton textile via a solvothermal procedure [203]. CuBDC@Textile was investigated as a solid sensor and adsorbent for volatile organic compounds (VOCs). It offered selective detection of pyridine via the colorimetric method. Pyridine turned the turquoise color of the prepared materials into deep blue color. It offered a pyridine adsorption capacity of 137.9 mg/g [203]. Lanthanide MOFs were also incorporated into cotton textiles for the photodegradation of stains for smart textiles [204].

Three-dimensional (3D) printing can be used to proceed MOF materials such as leaf-like zeolitic imidazolate frameworks (ZIF-L) into 3D objects with custom porosity and dimension (**Figure 4**)[157]. Direct ink writing (DIW) or robocasting was used to proceed with the materials. The printed materials with a ZIF content of 84% were achieved. The materials can adsorb CO₂ and heavy metals. 3D CelloZIF-L exhibited adsorption capacities of 0.64-1.15 mmol/g for CO₂ gases at 1 bar (0 °C). They showed adsorption capacities of 389.8-554.8 mg/g for Cu²⁺ ions with a selectivity of 86.8% toward Fe³⁺ ions [157]. A filter paper containing

cellulose and ZIF-8 were reported [158,159]. The prepared filter paper, denoted as CelloZIFPaper, was used for heavy metal adsorption. The materials offered adsorption capacities of 66.2–354.0 mg/g. CelloZIFPaper was also tested as a flexible electrode for toxic heavy metal detection [158,159]. The reader can directly go to our recent Review on the topic of cellulose-MOF composite (denoted as CelloMOF) and their applications [205]. CelloMOF enabled multifunctional applications being efficient adsorbents and catalysts [156]. ZIF-8 was also reported for the recovery of rare-earth elements [206].

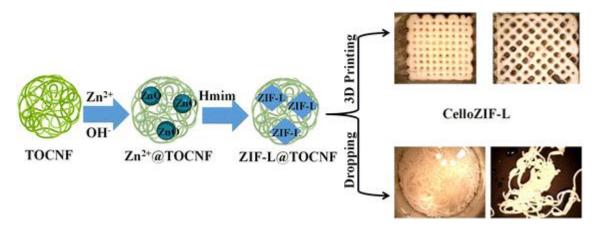


Figure 4 Schematic representation for the synthesis of ZIF-L in TEMPO-oxidized cellulose nanofibers (TOCNF) and 3D printing into cubes and filaments. Figure reprinted with permission from Ref. [157].

Magnetic nanoparticle-modified MOF materials were reported for heavy metal adsorption and removal [207]. Fe₃O₄@ZIF-8 and Fe₃O₄@UiO-66–NH₂) were investigated for the adsorption of Cd²⁺ and Pb²⁺ ions. Fe₃O₄@UiO-66–NH₂ and Fe₃O₄@ZIF-8 offered adsorption capacities of 714.3 mg/g, and 370 mg/g for Cd²⁺, respectively, and 833.3 mg/g, and 666.7 mg/g for Pb²⁺, respectively [207].

CuBDC has used the reduction of nitrophenol into aminophenol [208]. CuBDC was used as a precursor for the synthesis of CuO-embedded C i.e. CuO@C [209,210]. CuO@C exhibits a particle size of 36-123 nm [209]. It can be used as an antifungal agent against *Alternaria alternata*, *Fusarium oxysporum*, *Penicillium digitatum*, and *Rhizopus oryzae* with inhibition

zones of 36, 20.2, 16, and 10.2 mm, respectively [209]. CuO@C was also used as a photocatalyst for pharmaceuticals e.g. paracetamol degradation [211]. It offered an efficiency of 95% within 60 min [211]. It can also used for the reduction of 4-nitrophenol into 4-aminophenol [212]. In the presence of NaBH₄, CuO@C undergo catalytic degradation of organic dyes [213].

ZIF-67 was carbonized into $Co_3O_4@N$ -doped C [214]. The materials after carbonization were used as electroactive material for electrode fabrication. $Co_3O_4@N$ -doped C electrode offered a specific capacitance of 709 F g⁻¹ at 1 A g⁻¹ [214]. It can be also used as co-catalyst to enahnce the photocatalytic water splitting of semiconductor TiO₂ [215]. ZnO@C was prepared via carbonization of ZIF-8 [216]. It was used for supercapacitor [216]. ZIF-8 was used to prepare ZnO@C photocatalyst that can degrade dyes [217,218]. ZnO@C can be also used an efficient catalyst for methanol dehydration forming dimethyl ether that can be used as energy fuel [219].

Covalent Organic Frameworks (COFs)

COFs were used as support for the in-situ growth of palladium nanocrystals (Pd NCs@COF) [220,221]. Pd NCs@COF was used as the catalyst for carbon-carbon coupling reactions with high efficiency and excellent selectivity [220,221]. A composite of COFs material with two-dimensional nanoparticles e.g., graphene oxide, boron nitride, and graphitic carbon nitride (g-C₃N₄) was synthesized via a one-pot procedure [222]. The nanocomposites were used in water treatment via organic pollutants adsorption [222].

COFs have an advanced energy sector [223,224]. A triazine COF was synthesized via in-situ and ex-situ procedures in the presence of graphene oxide (GO, **Figure 5**) [225,226]. The composite was used to synthesize N-doped carbon (N-doped C)/reduced GO (rGO) after carbonization. N-doped C/rGO displayed a specific capacitance of 234 $\text{F} \cdot \text{g}^{-1}$ at the current density of 0.8 $\text{A} \cdot \text{g}^{-1}$. The electrochemical performance of two symmetric supercapacitor devices displayed specific energy and specific power of $14.6 \text{ W}\cdot\text{h}\cdot\text{kg}^{-1}$ and $400 \text{ W}\cdot\text{kg}^{-1}$, respectively (**Figure 5**) [225]. A one-pot synthesis of COFs/graphitic carbon nitride (g-C₃N₄) nanocomposite was also reported in our lab [227,228]. The synthesis procedure involved the polycondensation of melamine and benzene-1,3,5-tricarboxyaldehyde in the presence of g-C₃N₄. COF/g-C₃N₄ was used as a precursor for the synthesis of N-doped carbon and N-doped carbon/g-C₃N₄. The prepared materials were used as electrode materials for supercapacitors and lithium-ion batteries (LIBs). COF, COF/g-C₃N₄, N-doped carbon, and N-doped carbon/g-C₃N₄ exhibited specific capacitance of 211, 257.5, 450, and 835.2 F·g⁻¹, respectively. N-doped carbon/g-C₃N₄ was used to assemble asymmetric devices that offered energy density and power density of 45.97 Wh·kg⁻¹ and 659.3 W·kg⁻¹, respectively [227,228].

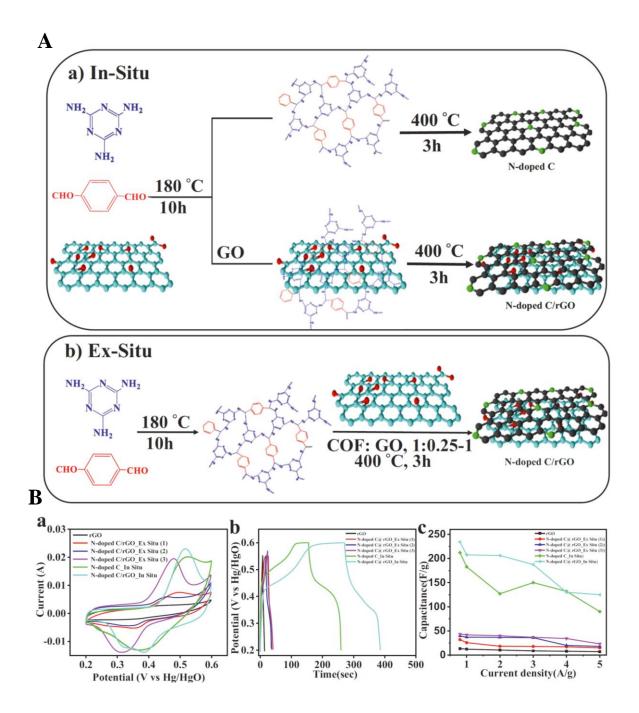


Figure 5 A) Synthesis procedure of the materials and B)electrochemical performance of the prepared electrode using a) CV curves at 50 mV s scan rates b) GCD curves, and c) capacitance over current density. Figure reprinted with permission from Ref.[225].

Conclusions

I summarized the potential of our laboratories to be applied in several fields such as environmental trends e.g., water remediation, air purification, and gas storage; energy e.g., production of hydrogen, dimethyl ether, solar cells, and supercapacitors; and biomedical sectors e.g., sensing/biosensing, cancer therapy, and drug delivery. We can synthesize materials that can be used as efficient adsorbents and catalysts to remove emerging contaminants such as metals, dyes, drugs, antibiotics, pesticides, and oils in water via adsorption. The materials can be also used as catalysts for pollutants degradation, synthesis of new organic compounds, reduction, and oxidation of organic pollutants. They have been applied as filters for air purification by adsorption of greenhouse gases such as carbon dioxide (CO₂), volatile organic compounds (VOCs), and particulate matter (PMs). They can be used for hydrogen production via water splitting, oxidation of alcohol, and hydrolysis of NaBH₄. They can be applied for biomedical applications such as antibacterial, drug delivery, and biosensing.

References

- [1] Abdelhamid HN. A review on hydrogen generation from the hydrolysis of sodium borohydride. Int J Hydrogen Energy 2021;46:726–65. https://doi.org/10.1016/j.ijhydene.2020.09.186.
- [2] Dragan M. Hydrogen Storage in Complex Metal Hydrides NaBH4: Hydrolysis Reaction and Experimental Strategies. Catalysts 2022;12:356. https://doi.org/10.3390/catal12040356.
- [3] Ruslan N, Yahya MS, Siddique MNI, Yengantiwar AP, Ismail M, Awal MR, et al. Review on Magnesium Hydride and Sodium Borohydride Hydrolysis for Hydrogen Production. Crystals 2022;12:1376. https://doi.org/10.3390/cryst12101376.
- [4] Liu BH, Li ZP. A review: Hydrogen generation from borohydride hydrolysis reaction. J Power Sources 2009;187:527–34. https://doi.org/10.1016/j.jpowsour.2008.11.032.
- [5] Abdelhamid HN, Wu H-F. A New Binary Matrix for Specific Detection of Mercury(II) Using Matrix-Assisted Laser Desorption Ionization Mass Spectrometry. J Am Soc Mass Spectrom 2019;30:2617–22. https://doi.org/10.1007/s13361-019-02324-1.
- [6] Abdelhamid HN. Nanoparticle-based surface assisted laser desorption ionization mass spectrometry: a review. Microchim Acta 2019;186:682. https://doi.org/10.1007/s00604-019-3770-5.
- [7] Abdelhamid HN. Nanoparticle assisted laser desorption/ionization mass spectrometry for small molecule analytes. Microchim Acta 2018;185:200. https://doi.org/10.1007/s00604-018-2687-8.
- [8] Abdelhamid HN. Organic matrices, ionic liquids, and organic matrices@nanoparticles assisted laser desorption/ionization mass spectrometry. TrAC Trends Anal Chem 2017;89:68–98. https://doi.org/10.1016/j.trac.2017.01.012.
- [9] Abdelhamid HN, Wu H-F. Soft Ionization of Metallo-Mefenamic Using Electrospray Ionization Mass Spectrometry. Mass Spectrom Lett 2015;6:43–7.
- [10] Khan N, Abdelhamid HN, Yan J-Y, Chung F-TF-T, Wu H-FH-F. Detection of flutamide in pharmaceutical dosage using higher electrospray ionization mass

spectrometry (ESI-MS) tandem mass coupled with Soxhlet apparatus. Anal Chem Res 2015;3:89–97. https://doi.org/10.1016/j.ancr.2015.01.001.

- [11] Abdelhamid HN. Functionalized Materials for Miniaturized Analytical Devices. Miniaturized Anal. Devices, Wiley; 2022, p. 181–95. https://doi.org/10.1002/9783527827213.ch9.
- [12] Kailasa SK, Hussain CM. Miniaturized Analytical Devices: Materials and Technology 2021.
- [13] Abdelhamid HN, Badr G. Nanobiotechnology as a platform for the diagnosis of COVID-19: a review. Nanotechnol Environ Eng 2021. https://doi.org/10.1007/s41204-021-00109-0.
- [14] Acknowledgment of reviewers 2020. Int J Hydrogen Energy 2021;46:1–30. https://doi.org/10.1016/j.ijhydene.2020.12.072.
- [15] Abdelhamid HN. Ionic liquids for nanomaterials recycling. Nanomater. Recycl., Elsevier; 2022, p. 269–87. https://doi.org/10.1016/B978-0-323-90982-2.00024-X.
- [16] Abdelhamid HN. Ionic Liquid-Assisted Laser Desorption/Ionization–Mass Spectrometry: Matrices, Microextraction, and Separation. Methods Protoc 2018;1:23. https://doi.org/10.3390/mps1020023.
- [17] Abdelhamid HN. Ionic liquids for mass spectrometry: matrices, separation and microextraction. TrAC Trends Anal Chem 2015;77:122–38. https://doi.org/10.1016/j.trac.2015.12.007.
- [18] Abdelhamid HN. Physicochemical Properties of Proteomic Ionic Liquids Matrices for MALDI-MS. J Data Mining Genomics Proteomics 2016;07:2153-0602.1000. https://doi.org/10.4172/2153-0602.1000189.
- [19] Abdelhamid HN, Khan MS, Wu H-FF. Design, characterization and applications of new ionic liquid matrices for multifunctional analysis of biomolecules: a novel strategy for pathogenic bacteria biosensing. Anal Chim Acta 2014;823:51–60. https://doi.org/10.1016/j.aca.2014.03.026.
- [20] Abdelhamid HN, Wu H-F. Ionic Liquid Matrices for Mass Spectrometry: Design, Synthesis, and Applications. Ref. Modul. Chem. Mol. Sci. Chem. Eng., 2014. https://doi.org/10.1016/B978-0-12-409547-2.11016-9.
- [21] Abdelhamid HN, Gopal J, Wu H-FF. Synthesis and application of ionic liquid matrices (ILMs) for effective pathogenic bacteria analysis in matrix assisted laser desorption/ionization (MALDI-MS). Anal Chim Acta 2013;767:104–11. https://doi.org/10.1016/j.aca.2012.12.054.
- [22] Abdelhamid HN, Wu H-F. Monitoring metallofulfenamic–bovine serum albumin interactions: a novel method for metallodrug analysis. RSC Adv 2014;4:53768–76. https://doi.org/10.1039/C4RA07638A.
- [23] Abdelhamid HN, El-Zohry AM, Cong J, Thersleff T, Karlsson M, Kloo L, et al. Towards implementing hierarchical porous zeolitic imidazolate frameworks in dyesensitized solar cells. R Soc Open Sci 2019;6:190723. https://doi.org/10.1098/rsos.190723.
- [24] Keservani R, Sharma A, Abdelhamid H. Nanoparticulate Drug Delivery Systems. CRC Press; 2019. https://doi.org/10.1201/9781420008449-6.
- [25] Abdelhamid HN, Wu H-F. Nanoparticles Advanced Drug Delivery for Cancer Cells. In: Keservani RK, Sharma AK, editors. Nanoparticulate Drug Deliv. Syst., 2019, p. 121–44.
- [26] Dowaidar M, Abdelhamid HN, Langel Ü. Improvement of Transfection with PepFects Using Organic and Inorganic Materials, 2022, p. 555–67. https://doi.org/10.1007/978-1-0716-1752-6_35.
- [27] Abdelhamid HN, Dowaidar M, Hällbrink M, Langel Ü. Cell Penetrating Peptides-

Hierarchical Porous Zeolitic Imidazolate Frameworks Nanoparticles: An Efficient Gene Delivery Platform. SSRN Electron J 2019. https://doi.org/10.2139/ssrn.3435895.

- [28] Abdelhamid HN. Nanocytotoxicity using matrix-assisted laser desorption ionization mass spectrometry. Future Microbiol 2020;15:385–7. https://doi.org/10.2217/fmb-2019-0260.
- [29] Nanotoxicity. Elsevier; 2020. https://doi.org/10.1016/C2018-0-05517-6.
- [30] Abdelhamid HN. Metals Linked to Alzheimer's Disease. Front. Clin. Drug Res. -Alzheimer Disord., BENTHAM SCIENCE PUBLISHERS; 2020, p. 213–35. https://doi.org/10.2174/9789811410949120010009.
- [31] Abdelhamid HN, Kumaran S, Wu H-F. One-pot synthesis of CuFeO2 nanoparticles capped with glycerol and proteomic analysis of their nanocytotoxicity against fungi. RSC Adv 2016;6:97629–35. https://doi.org/10.1039/C6RA13396G.
- [32] Kierstead PH, Okochi H, Venditto VJ, Chuong TC, Kivimae S, Fréchet JMJ, et al. The effect of polymer backbone chemistry on the induction of the accelerated blood clearance in polymer modified liposomes. J Control Release 2015;213:1–9. https://doi.org/10.1016/j.jconrel.2015.06.023.
- [33] Abdelhamid HN. Nanoparticles as Pharmaceutical Agents. MJ Anes 2016;1:003–003.
- [34] Abdelhamid HN. General methods for detection and evaluation of nanotoxicity. Nanotoxicity 2020:195–214. https://doi.org/10.1016/B978-0-12-819943-5.00009-9.
- [35] Abdelhamid HN. Nanoparticles Assisted Laser Desorption/Ionization Mass Spectrometry. Handb. Smart Mater. Anal. Chem., Chichester, UK: John Wiley & Sons, Ltd; 2019, p. 729–55. https://doi.org/10.1002/9781119422587.ch23.
- [36] Kumaran S, Abdelhamid HN, Wu H-F. Quantification analysis of protein and mycelium contents upon inhibition of melanin for: Aspergillus Niger: A study of matrix assisted laser desorption/ionization mass spectrometry (MALDI-MS). RSC Adv 2017;7:30289–94. https://doi.org/10.1039/c7ra03741d.
- [37] Abdelhamid HN, Wu H-F. Proteomics analysis of the mode of antibacterial action of nanoparticles and their interactions with proteins. TrAC Trends Anal Chem 2015;65:30–46. https://doi.org/10.1016/j.trac.2014.09.010.
- [38] Zayene M, Algethami FK, Nasser Abdelhamid H, Elamin MR, Abdulkhair BY, Al-Ghamdi YO, et al. New synthetic quinaldine conjugates: Assessment of their anticholinesterase, anti-tyrosinase and cytotoxic activities, and molecular docking analysis. Arab J Chem 2022;15:104177. https://doi.org/10.1016/j.arabjc.2022.104177.
- [39] Algethami FK, Saidi I, Abdelhamid HN, Elamin MR, Abdulkhair BY, Chrouda A, et al. Trifluoromethylated Flavonoid-Based Isoxazoles as Antidiabetic and Anti-Obesity Agents: Synthesis, In Vitro α-Amylase Inhibitory Activity, Molecular Docking and Structure–Activity Relationship Analysis. Molecules 2021;26:5214. https://doi.org/10.3390/molecules26175214.
- [40] Nasser Abdelhamid H, Wu HF. Furoic and mefenamic acids as new matrices for matrix assisted laser desorption/ionization-(MALDI)-mass spectrometry. Talanta 2013;115:442–50. https://doi.org/10.1016/j.talanta.2013.05.050.
- [41] H. N. Abdelhamid. Applications of Nanomaterials and Organic Semiconductors for Bacteria &Biomolecules analysis/ biosensing using Laser Analytical Spectroscopy. National Sun-Yat Sen University, 2013. https://doi.org/etd-0608113-135030.
- [42] Kumaran S, Abdelhamid HN, Hasan N, Wu H-F. Cytotoxicity of Palladium Nanoparticles Against Aspergillus Niger. Nanosci Nanotechnology-Asia 2020;10:80– 5. https://doi.org/10.2174/2210681208666180904113754.
- [43] Lo VK-Y, Chan AO-Y, Che C-M. Gold and silver catalysis: from organic transformation to bioconjugation. Org Biomol Chem 2015;13:6667–80. https://doi.org/10.1039/c5ob00407a.

- [44] Motl NE, Smith AF, DeSantis CJ, Skrabalak SE. Engineering plasmonic metal colloids through composition and structural design. Chem Soc Rev 2014;43:3823–34. https://doi.org/10.1039/c3cs60347d.
- [45] Wu L, Reinhard BM. Probing subdiffraction limit separations with plasmon coupling microscopy: concepts and applications. Chem Soc Rev 2014;43:3884–97. https://doi.org/10.1039/c3cs60340g.
- [46] Sekuła J, Nizioł J, Rode W, Ruman T. Silver nanostructures in laser desorption/ionization mass spectrometry and mass spectrometry imaging. Analyst 2015;140:6195–209. https://doi.org/10.1039/C5AN00943J.
- [47] Dastafkan K, Khajeh M, Bohlooli M, Ghaffari-Moghaddam M, Sheibani N. Mechanism and behavior of silver nanoparticles in aqueous medium as adsorbent. Talanta 2015;144:1377–86. https://doi.org/10.1016/j.talanta.2015.03.065.
- [48] Yousef MS, Abdelhamid HN, Hidalgo M, Fathy R, Gómez-Gascón L, Dorado J. Antimicrobial activity of silver-carbon nanoparticles on the bacterial flora of bull semen. Theriogenology 2021;161:219–27. https://doi.org/10.1016/j.theriogenology.2020.12.006.
- [49] Abdelhamid HN, Talib A, Wu H-F. Facile synthesis of water soluble silver ferrite (AgFeO2) nanoparticles and their biological application as antibacterial agents. RSC Adv 2015;5:34594–602. https://doi.org/10.1039/C4RA14461A.
- [50] Abdelhamid HN, Talib A, Wu H-F. Correction: Facile synthesis of water soluble silver ferrite (AgFeO 2) nanoparticles and their biological application as antibacterial agents. RSC Adv 2015;5:39952–3. https://doi.org/10.1039/C5RA90041G.
- [51] Gopal J, Abdelhamid HN, Huang JH, Wu HF. Nondestructive detection of the freshness of fruits and vegetables using gold and silver nanoparticle mediated graphene enhanced Raman spectroscopy. Sensors Actuators, B Chem 2016;224:413– 24. https://doi.org/10.1016/j.snb.2015.08.123.
- [52] Shastri L, Abdelhamid HN, Nawaz M, Wu H-F. Synthesis, characterization and bifunctional applications of bidentate silver nanoparticle assisted single drop microextraction as a highly sensitive preconcentrating probe for protein analysis. RSC Adv 2015;5:41595–603. https://doi.org/10.1039/C5RA04032A.
- [53] Abdelhamid HN, Wu H-F. Facile synthesis of nano silver ferrite (AgFeO2) modified with chitosan applied for biothiol separation. Mater Sci Eng C 2014;45:438–45. https://doi.org/10.1016/j.msec.2014.08.071.
- [54] Abdelhamid HN, Talib A, Wu H-F. One pot synthesis of gold carbon dots nanocomposite and its application for cytosensing of metals for cancer cells. Talanta 2017;166:357–63. https://doi.org/10.1016/j.talanta.2016.11.030.
- [55] Abdelhamid HN, Wu H-F. Gold nanoparticles assisted laser desorption/ionization mass spectrometry and applications: from simple molecules to intact cells. Anal Bioanal Chem 2016;408:4485–502. https://doi.org/10.1007/s00216-016-9374-6.
- [56] Lim SY, Shen W, Gao Z. Carbon quantum dots and their applications. Chem Soc Rev 2015;44:362–81. https://doi.org/10.1039/C4CS00269E.
- [57] Li M, Chen T, Gooding JJ, Liu J. Review of Carbon and Graphene Quantum Dots for Sensing. ACS Sensors 2019;4:1732–48. https://doi.org/10.1021/acssensors.9b00514.
- [58] Liu ML, Chen B Bin, Li CM, Huang CZ. Carbon dots: synthesis, formation mechanism, fluorescence origin and sensing applications. Green Chem 2019;21:449– 71. https://doi.org/10.1039/C8GC02736F.
- [59] Bag P, Maurya RK, Dadwal A, Sarkar M, Chawla PA, Narang RK, et al. Recent Development in Synthesis of Carbon Dots from Natural Resources and Their Applications in Biomedicine and Multi-Sensing Platform. ChemistrySelect 2021;6:2774–89. https://doi.org/10.1002/slct.202100468.

- [60] Iravani S, Varma RS. Green synthesis, biomedical and biotechnological applications of carbon and graphene quantum dots. A review. Environ Chem Lett 2020. https://doi.org/https://doi.org/10.1007/s10311-020-00984-0.
- [61] Zhang J, Yu S-H. Carbon dots: large-scale synthesis, sensing and bioimaging. Mater Today 2016;19:382–93. https://doi.org/10.1016/j.mattod.2015.11.008.
- [62] Kailasa SK, Hussain CM. Carbon Dots in Analytical Chemistry: Detection and Imaging. 2022. https://doi.org/10.1016/C2021-0-01017-8.
- [63] Nair A, Haponiuk JT, Thomas S, Gopi S. Natural carbon-based quantum dots and their applications in drug delivery: A review. Biomed Pharmacother 2020;132:110834. https://doi.org/10.1016/j.biopha.2020.110834.
- [64] Algarra M, Pérez-Martín M, Cifuentes-Rueda M, Jiménez-Jiménez J, Esteves da Silva JCG, Bandosz TJ, et al. Carbon dots obtained using hydrothermal treatment of formaldehyde. Cell imaging in vitro. Nanoscale 2014;6:9071–7. https://doi.org/10.1039/C4NR01585A.
- [65] Shi H, Wei J, Qiang L, Chen X, Meng X. Fluorescent Carbon Dots for Bioimaging and Biosensing Applications. J Biomed Nanotechnol 2014;10:2677–99. https://doi.org/10.1166/jbn.2014.1881.
- [66] Loo AH, Sofer Z, Bouša D, Ulbrich P, Bonanni A, Pumera M. Carboxylic Carbon Quantum Dots as a Fluorescent Sensing Platform for DNA Detection. ACS Appl Mater Interfaces 2016;8:1951–7. https://doi.org/10.1021/acsami.5b10160.
- [67] Liu H, Zhang Y, Huang C. Development of nitrogen and sulfur-doped carbon dots for cellular imaging. J Pharm Anal 2019;9:127–32. https://doi.org/10.1016/j.jpha.2018.10.001.
- [68] Li H, Yan X, Kong D, Jin R, Sun C, Du D, et al. Recent advances in carbon dots for bioimaging applications. Nanoscale Horizons 2020;5:218–34. https://doi.org/10.1039/C9NH00476A.
- [69] Sun H, Wu L, Wei W, Qu X. Recent advances in graphene quantum dots for sensing. Mater Today 2013;16:433–42. https://doi.org/10.1016/j.mattod.2013.10.020.
- [70] Chen W, Weng W, Niu X, Li X, Men Y, Sun W, et al. Boron-doped Graphene quantum dots modified electrode for electrochemistry and electrocatalysis of hemoglobin. J Electroanal Chem 2018;823:137–45. https://doi.org/10.1016/j.jelechem.2018.06.001.
- [71] Abdelhamid HN. Carbon dots-based fluorescence spectroscopy for metal ion sensing. Carbon Dots Anal. Chem., Elsevier; 2023, p. 87–96. https://doi.org/10.1016/B978-0-323-98350-1.00025-6.
- [72] Feng Z, Adolfsson KH, Xu Y, Fang H, Hakkarainen M, Wu M. Carbon dot/polymer nanocomposites: From green synthesis to energy, environmental and biomedical applications. Sustain Mater Technol 2021;29:e00304. https://doi.org/10.1016/j.susmat.2021.e00304.
- [73] Patel KD, Singh RK, Kim H-W. Carbon-based nanomaterials as an emerging platform for theranostics. Mater Horizons 2019;6:434–69. https://doi.org/10.1039/C8MH00966J.
- [74] Xu X, Ray R, Gu Y, Ploehn HJ, Gearheart L, Raker K, et al. Electrophoretic Analysis and Purification of Fluorescent Single-Walled Carbon Nanotube Fragments. J Am Chem Soc 2004;126:12736–7. https://doi.org/10.1021/ja040082h.
- [75] Li H, Kang Z, Liu Y, Lee S-T. Carbon nanodots: synthesis, properties and applications. J Mater Chem 2012;22:24230. https://doi.org/10.1039/c2jm34690g.
- [76] Esteves da Silva JCG, Gonçalves HMR. Analytical and bioanalytical applications of carbon dots. TrAC Trends Anal Chem 2011;30:1327–36. https://doi.org/10.1016/j.trac.2011.04.009.

- [77] Rasal AS, Yadav S, Yadav A, Kashale AA, Manjunatha ST, Altaee A, et al. Carbon Quantum Dots for Energy Applications: A Review. ACS Appl Nano Mater 2021;4:6515–41. https://doi.org/10.1021/acsanm.1c01372.
- [78] Zulfajri M, Abdelhamid HN, Sudewi S, Dayalan S, Rasool A, Habib A, et al. Plant Part-Derived Carbon Dots for Biosensing. Biosensors 2020;10:68. https://doi.org/10.3390/bios10060068.
- [79] Nekoueian K, Amiri M, Sillanpää M, Marken F, Boukherroub R, Szunerits S. Carbonbased quantum particles: an electroanalytical and biomedical perspective. Chem Soc Rev 2019;48:4281–316. https://doi.org/10.1039/C8CS00445E.
- [80] Campuzano S, Yáñez-Sedeño P, Pingarrón JM. Carbon Dots and Graphene Quantum Dots in Electrochemical Biosensing. Nanomaterials 2019;9:634. https://doi.org/10.3390/nano9040634.
- [81] Abdelhamid HN. Carbon dots for electrochemical analytical methods. Carbon Dots Anal. Chem., Elsevier; 2023, p. 77–86. https://doi.org/10.1016/B978-0-323-98350-1.00023-2.
- [82] Martínez-Periñán E, Bravo I, Rowley-Neale SJ, Lorenzo E, Banks CE. Carbon Nanodots as Electrocatalysts towards the Oxygen Reduction Reaction. Electroanalysis 2018;30:436–44. https://doi.org/10.1002/elan.201700718.
- [83] Li H, Chen L, Wu H, He H, Jin Y. Ionic Liquid-Functionalized Fluorescent Carbon Nanodots and Their Applications in Electrocatalysis, Biosensing, and Cell Imaging. Langmuir 2014;30:15016–21. https://doi.org/10.1021/la503729v.
- [84] Ji H, Zhou F, Gu J, Shu C, Xi K, Jia X. Nitrogen-Doped Carbon Dots as A New Substrate for Sensitive Glucose Determination. Sensors 2016;16:630. https://doi.org/10.3390/s16050630.
- [85] Zheng W, Wu H, Jiang Y, Xu J, Li X, Zhang W, et al. A molecularly-imprintedelectrochemical-sensor modified with nano-carbon-dots with high sensitivity and selectivity for rapid determination of glucose. Anal Biochem 2018;555:42–9. https://doi.org/10.1016/j.ab.2018.06.004.
- [86] Wang Y, Wang Z, Rui Y, Li M. Horseradish peroxidase immobilization on carbon nanodots/CoFe layered double hydroxides: Direct electrochemistry and hydrogen peroxide sensing. Biosens Bioelectron 2015;64:57–62. https://doi.org/10.1016/j.bios.2014.08.054.
- [87] Huang Q, Hu S, Zhang H, Chen J, He Y, Li F, et al. Carbon dots and chitosan composite film based biosensor for the sensitive and selective determination of dopamine. Analyst 2013;138:5417. https://doi.org/10.1039/c3an00510k.
- [88] Zhang L, Han Y, Zhu J, Zhai Y, Dong S. Simple and Sensitive Fluorescent and Electrochemical Trinitrotoluene Sensors Based on Aqueous Carbon Dots. Anal Chem 2015;87:2033–6. https://doi.org/10.1021/ac5043686.
- [89] Lu X, Wang X, Jin J, Zhang Q, Chen J. Electrochemical biosensing platform based on amino acid ionic liquid functionalized graphene for ultrasensitive biosensing applications. Biosens Bioelectron 2014;62:134–9. https://doi.org/10.1016/j.bios.2014.06.036.
- [90] Zhou J, Shan X, Ma J, Gu Y, Qian Z, Chen J, et al. Facile synthesis of P-doped carbon quantum dots with highly efficient photoluminescence. RSC Adv 2014;4:5465. https://doi.org/10.1039/c3ra45294h.
- [91] Teng X, Ma C, Ge C, Yan M, Yang J, Zhang Y, et al. Green synthesis of nitrogendoped carbon dots from konjac flour with "off–on" fluorescence by Fe3+ and l-lysine for bioimaging. J Mater Chem B 2014;2:4631. https://doi.org/10.1039/c4tb00368c.
- [92] Hong D, Deng X, Liang J, Li J, Tao Y, Tan K. One-step hydrothermal synthesis of down/up-conversion luminescence F-doped carbon quantum dots for label-free

detection of Fe3+. Microchem J 2019;151:104217. https://doi.org/10.1016/j.microc.2019.104217.

- [93] Wang F, Hao Q, Zhang Y, Xu Y, Lei W. Fluorescence quenchometric method for determination of ferric ion using boron-doped carbon dots. Microchim Acta 2016;183:273–9. https://doi.org/10.1007/s00604-015-1650-1.
- [94] Sun Y, Shen C, Wang J, Lu Y. Facile synthesis of biocompatible N, S-doped carbon dots for cell imaging and ion detecting. RSC Adv 2015;5:16368–75. https://doi.org/10.1039/C4RA13820A.
- [95] Qie X, Zan M, Miao P, Li L, Chang Z, Ge M, et al. One-step synthesis of nitrogen, sulfur co-doped carbon nanodots and application for Fe 3+ detection. J Mater Chem B 2018;6:3549–54. https://doi.org/10.1039/C8TB00193F.
- [96] Zhang Y, Qin H, Huang Y, Zhang F, Liu H, Liu H, et al. Highly fluorescent nitrogen and boron doped carbon quantum dots for selective and sensitive detection of Fe 3+. J Mater Chem B 2021;9:4654–62. https://doi.org/10.1039/D1TB00371B.
- [97] Zhu S, Meng Q, Wang L, Zhang J, Song Y, Jin H, et al. Highly Photoluminescent Carbon Dots for Multicolor Patterning, Sensors, and Bioimaging. Angew Chemie 2013;125:4045–9. https://doi.org/10.1002/ange.201300519.
- [98] Sachdev A, Gopinath P. Green synthesis of multifunctional carbon dots from coriander leaves and their potential application as antioxidants, sensors and bioimaging agents. Analyst 2015;140:4260–9. https://doi.org/10.1039/C5AN00454C.
- [99] Meiling TT, Schürmann R, Vogel S, Ebel K, Nicolas C, Milosavljević AR, et al. Photophysics and Chemistry of Nitrogen-Doped Carbon Nanodots with High Photoluminescence Quantum Yield. J Phys Chem C 2018;122:10217–30. https://doi.org/10.1021/acs.jpcc.8b00748.
- [100] Sun X, Lei Y. Fluorescent carbon dots and their sensing applications. TrAC Trends Anal Chem 2017;89:163–80. https://doi.org/10.1016/j.trac.2017.02.001.
- [101] Hu C, Li M, Qiu J, Sun Y-P. Design and fabrication of carbon dots for energy conversion and storage. Chem Soc Rev 2019;48:2315–37. https://doi.org/10.1039/C8CS00750K.
- [102] Dong Y, Pang H, Yang H Bin, Guo C, Shao J, Chi Y, et al. Carbon-based dots codoped with nitrogen and sulfur for high quantum yield and excitation-independent emission. Angew Chem Int Ed Engl 2013;52:7800–4. https://doi.org/10.1002/anie.201301114.
- [103] Saraf M, Tavakkoli Yaraki M, Prateek, Tan YN, Gupta RK. Insights and Perspectives Regarding Nanostructured Fluorescent Materials toward Tackling COVID-19 and Future Pandemics. ACS Appl Nano Mater 2021;4:911–48. https://doi.org/10.1021/acsanm.0c02945.
- [104] Belza J, Opletalová A, Poláková K. Carbon dots for virus detection and therapy. Microchim Acta 2021;188:430. https://doi.org/10.1007/s00604-021-05076-6.
- [105] Junaid HM, Solangi AR, Batool M. Carbon dots as naked eye sensors. Analyst 2021;146:2463–74. https://doi.org/10.1039/D0AN02399J.
- [106] Liu Y, Liu Y, Park S-J, Zhang Y, Kim T, Chae S, et al. One-step synthesis of robust nitrogen-doped carbon dots: acid-evoked fluorescence enhancement and their application in Fe 3+ detection. J Mater Chem A 2015;3:17747–54. https://doi.org/10.1039/C5TA05189D.
- [107] Ashour RM, Abdelhamid HN, Abdel-Magied AF, Abdel-khalek AA, Ali M, Uheida A, et al. Rare Earth Ions Adsorption onto Graphene Oxide Nanosheets. Solvent Extr Ion Exch 2017;35:91–103. https://doi.org/10.1080/07366299.2017.1287509.
- [108] Abdelhamid HN, Wu H-F. Reduced graphene oxide conjugate thymine as a new probe for ultrasensitive and selective fluorometric determination of mercury(II) ions.

Microchim Acta 2015;182:1609–17. https://doi.org/10.1007/s00604-015-1461-4.

- [109] Abdelhamid HN, Wu H-F. Synthesis of a highly dispersive sinapinic acid@graphene oxide (SA@GO) and its applications as a novel surface assisted laser desorption/ionization mass spectrometry for proteomics and pathogenic bacteria biosensing. Analyst 2015;140:1555–65.
- [110] Nasser Abdelhamid H, Wu B-S, Wu H-F. Graphene coated silica applied for high ionization matrix assisted laser desorption/ionization mass spectrometry: A novel approach for environmental and biomolecule analysis. Talanta 2014;126:27–37. https://doi.org/10.1016/j.talanta.2014.03.016.
- [111] Abdelhamid HN, Wu H-F. Ultrasensitive, Rapid, and Selective Detection of Mercury Using Graphene Assisted Laser Desorption/Ionization Mass Spectrometry. J Am Soc Mass Spectrom 2014;25:861–8. https://doi.org/10.1007/s13361-014-0825-z.
- [112] Hua P-Y, Manikandan M, Abdelhamid HN, Wu H-F. Graphene nanoflakes as an efficient ionizing matrix for MALDI-MS based lipidomics of cancer cells and cancer stem cells. J Mater Chem B 2014;2:7334–43.
- [113] Abdelhamid HN, Wu H-F. A method to detect metal-drug complexes and their interactions with pathogenic bacteria via graphene nanosheet assist laser desorption/ionization mass spectrometry and biosensors. Anal Chim Acta 2012;751:94–104. https://doi.org/10.1016/j.aca.2012.09.012.
- [114] Hussein KH, Abdelhamid HN, Zou X, Woo H-M. Ultrasonicated graphene oxide enhances bone and skin wound regeneration. Mater Sci Eng C 2019;94:484–92. https://doi.org/10.1016/j.msec.2018.09.051.
- [115] Shahnawaz Khan M, Abdelhamid HN, Wu H-F. Near infrared (NIR) laser mediated surface activation of graphene oxide nanoflakes for efficient antibacterial, antifungal and wound healing treatment. Colloids Surf B Biointerfaces 2015;127C:281–91. https://doi.org/10.1016/j.colsurfb.2014.12.049.
- [116] Abdelhamid HN, Khan MS, Wu HF. Graphene oxide as a nanocarrier for gramicidin (GOGD) for high antibacterial performance. RSC Adv 2014;4:50035–46. https://doi.org/10.1039/c4ra07250b.
- [117] Dowaidar M, Abdelhamid HN, Hällbrink M, Zou X, Langel Ü. Graphene oxide nanosheets in complex with cell penetrating peptides for oligonucleotides delivery. Biochim Biophys Acta - Gen Subj 2017;1861:2334–41. https://doi.org/10.1016/j.bbagen.2017.07.002.
- [118] Soliman M, Sadek AA, Abdelhamid HN, Hussein K. Graphene oxide-cellulose nanocomposite accelerates skin wound healing. Res Vet Sci 2021;137:262–73. https://doi.org/10.1016/j.rvsc.2021.05.013.
- [119] Abdelhamid HN, Wu H-F. Graphene and Its Derivatives as Platforms for MALDI-MS. In: Tobias Stauber, editor. Handb. GRAPHENE 2, Vol. 2 Physics, Chem. Biol., Scrivener Publishing; 2019, p. 273–90.
- [120] Abdelhamid HN, Bhaisare ML, Wu H-F. Ceria nanocubic-ultrasonication assisted dispersive liquid-liquid microextraction coupled with matrix assisted laser desorption/ionization mass spectrometry for pathogenic bacteria analysis. Talanta 2014;120:208–17. https://doi.org/10.1016/j.talanta.2013.11.078.
- [121] Bhaisare ML, Abdelhamid HN, Wu B-S, Wu H-F. Rapid and direct MALDI-MS identification of pathogenic bacteria from blood using ionic liquid-modified magnetic nanoparticles (Fe3O4@SiO2). J Mater Chem B 2014;2:4671–83. https://doi.org/10.1039/C4TB00528G.
- [122] Abdelhamid HN, Wu H-F. Multifunctional graphene magnetic nanosheet decorated with chitosan for highly sensitive detection of pathogenic bacteria. J Mater Chem B 2013;1:3950–61. https://doi.org/10.1039/c3tb20413h.

- [123] Gopal J, Abdelhamid HN, Hua P-Y, Wu H-F. Chitosan nanomagnets for effective extraction and sensitive mass spectrometric detection of pathogenic bacterial endotoxin from human urine. J Mater Chem B 2013;1:2463. https://doi.org/10.1039/c3tb20079e.
- [124] Gedda G, Abdelhamid HN, Khan MS, Wu H-F. ZnO nanoparticle-modified polymethyl methacrylate-assisted dispersive liquid–liquid microextraction coupled with MALDI-MS for rapid pathogenic bacteria analysis. RSC Adv 2014;4:45973–83. https://doi.org/10.1039/C4RA03391D.
- [125] Wu B-S, Abdelhamid HN, Wu H-F. Synthesis and antibacterial activities of graphene decorated with stannous dioxide. RSC Adv 2014;4:3722. https://doi.org/10.1039/c3ra43992e.
- [126] Etman AS, Abdelhamid HN, Yuan Y, Wang L, Zou X, Sun J. Facile Water-Based Strategy for Synthesizing MoO3– x Nanosheets: Efficient Visible Light Photocatalysts for Dye Degradation. ACS Omega 2018;3:2193–201. https://doi.org/10.1021/acsomega.8b00012.
- [127] Iqbal MN, Abdel-Magied AF, Abdelhamid HN, Olsén P, Shatskiy A, Zou X, et al. Mesoporous Ruthenium Oxide: A Heterogeneous Catalyst for Water Oxidation. ACS Sustain Chem Eng 2017;5:9651–6. https://doi.org/10.1021/acssuschemeng.7b02845.
- [128] Abdelhamid HN. Laser Assisted Synthesis, Imaging and Cancer Therapy of Magnetic Nanoparticles. Mater Focus 2016;5:305–23. https://doi.org/10.1166/mat.2016.1336.
- [129] Abdelhamid HN. Delafossite Nanoparticle as New Functional Materials: Advances in Energy, Nanomedicine and Environmental Applications. Mater Sci Forum 2015;832:28–53. https://doi.org/10.4028/www.scientific.net/MSF.832.28.
- [130] Dowaidar M, Abdelhamid HN, Hällbrink M, Kurrikoff K, Freimann K, Zou X, et al. Magnetic Nanoparticle Assisted Self-assembly of Cell Penetrating Peptides-Oligonucleotides Complexes for Gene Delivery. Sci Rep 2017;7:9159. https://doi.org/10.1038/s41598-017-09803-z.
- [131] Abdelhamid HN. Quantum dots hybrid systems for drug delivery. Hybrid Nanomater. Drug Deliv., Elsevier; 2022, p. 323–38. https://doi.org/10.1016/B978-0-323-85754-3.00013-7.
- [132] Abdelhamid HN, Wu H-F. Selective biosensing of Staphylococcus aureus using chitosan quantum dots. Spectrochim Acta Part A Mol Biomol Spectrosc 2018;188:50– 6. https://doi.org/10.1016/j.saa.2017.06.047.
- [133] Chen Z-Y, Abdelhamid HN, Wu H-F. Effect of surface capping of quantum dots (CdTe) on proteomics. Rapid Commun Mass Spectrom 2016;30:1403–12. https://doi.org/10.1002/rcm.7575.
- [134] Wu H-F, Gopal J, Abdelhamid HN, Hasan N. Quantum dot applications endowing novelty to analytical proteomics. Proteomics 2012;12:2949–61. https://doi.org/10.1002/pmic.201200295.
- [135] Abdelhamid HN, Chen Z-Y, Wu H-F. Surface tuning laser desorption/ionization mass spectrometry (STLDI-MS) for the analysis of small molecules using quantum dots. Anal Bioanal Chem 2017;409:4943–50. https://doi.org/10.1007/s00216-017-0433-4.
- [136] Abdelhamid HN, Wu H-F. Synthesis and characterization of quantum dots for application in laser soft desorption/ionization mass spectrometry to detect labile metal– drug interactions and their antibacterial activity. RSC Adv 2015;5:76107–15. https://doi.org/10.1039/C5RA11301F.
- [137] Abdelhamid HN, Wu H-F. Synthesis and multifunctional applications of quantum nanobeads for label-free and selective metal chemosensing. RSC Adv 2015;5:50494– 504. https://doi.org/10.1039/C5RA07069D.
- [138] K. Algethami F, Saidi I, Ben Jannet H, Khairy M, Abdulkhair BY, Al-Ghamdi YO, et

al. Chitosan-CdS Quantum Dots Biohybrid for Highly Selective Interaction with Copper(II) Ions. ACS Omega 2022. https://doi.org/10.1021/acsomega.2c01793.

- [139] Abdelhamid HN, Algethami FK, Saidi I, Jannet H Ben, Khairy M, Abdulkhair BY, et al. Selective Naked-eyes Chemosensing of Cu2+ ions using Chitosan-CdS Quantum Dots Biohybrid. ChemRxiv Cambridge Cambridge Open Engag 2022; 2022. https://doi.org/10.26434/CHEMRXIV-2022-P92XT.
- [140] Abdelhamid HN, Wu H-F. Probing the interactions of chitosan capped CdS quantum dots with pathogenic bacteria and their biosensing application. J Mater Chem B 2013;1:6094. https://doi.org/10.1039/c3tb21020k.
- [141] Abdelhamid HN, El-Bery HM, Metwally AA, Elshazly M, Hathout RM. Synthesis of CdS-modified chitosan quantum dots for the drug delivery of Sesamol. Carbohydr Polym 2019;214:90–9. https://doi.org/10.1016/j.carbpol.2019.03.024.
- [142] Abdelhamid HN. Polysaccharides for biomedical implants. Plant Polysaccharides as Pharm. Excipients, Elsevier; 2023, p. 533–44. https://doi.org/10.1016/B978-0-323-90780-4.00015-2.
- [143] Nayak AK, Hasnain MS, Pal D, Abdelhamid HN. Plant Polysaccharides as Pharmaceutical Excipients. Elsevier; 2023. https://doi.org/10.1016/C2020-0-03919-8.
- [144] Abdelhamid HN. Self-decontaminating antimicrobial textiles. Antimicrob. Text. from Nat. Resour., Elsevier; 2021, p. 259–94. https://doi.org/10.1016/B978-0-12-821485-5.00011-1.
- [145] Rahimi S, Moradi M. Photo-induced antimicrobial agents for textile applications. Antimicrob. Text. from Nat. Resour., Elsevier; 2021, p. 217–58. https://doi.org/10.1016/B978-0-12-821485-5.00015-9.
- [146] Abdelhmaid HN. Antimicrobial Textiles from Natural Resources. n.d.
- [147] Abdelhamid HN. Alginate in Gene and Vaccine Delivery. Alginate Biomater., Singapore: Springer Nature Singapore; 2023, p. 361–88. https://doi.org/10.1007/978-981-19-6937-9_14.
- [148] Abdelhamid, H.N.;Sougata Jana; Subrata Jana. Alginate Biomaterial. Singapore: Springer Nature Singapore; 2023. https://doi.org/10.1007/978-981-19-6937-9.
- [149] Fijoł N, Abdelhamid HN, Pillai B, Hall SA, Thomas N, Mathew AP. 3D-printed monolithic biofilters based on a polylactic acid (PLA) – hydroxyapatite (HAp) composite for heavy metal removal from an aqueous medium. RSC Adv 2021;11:32408–18. https://doi.org/10.1039/D1RA05202K.
- [150] Abdelhamid HN, Mathew AP. Cellulose-Based Nanomaterials Advance Biomedicine: A Review. Int J Mol Sci 2022;23:5405. https://doi.org/10.3390/ijms23105405.
- [151] Abdelhamid HN., Mathew AP. A Review on Cellulose-based Materials for Biomedicine. Preprints 2022:2022010035. https://doi.org/10.20944/preprints202201.0035.v1.
- [152] Abdelhamid HN, Mathew AP. Cellulose-Based Materials for Water Remediation: Adsorption, Catalysis, and Antifouling. Front Chem Eng 2021;3:790314. https://doi.org/10.3389/fceng.2021.790314.
- [153] Aguilar-Sanchez A, Jalvo B, Mautner A, Rissanen V, Kontturi KS, Abdelhamid HN, et al. Charged ultrafiltration membranes based on TEMPO-oxidized cellulose nanofibrils/poly(vinyl alcohol) antifouling coating. RSC Adv 2021;11:6859–68. https://doi.org/10.1039/D0RA10220B.
- [154] Jana S. Micro- and Nanoengineered Gum-Based Biomaterials for Drug Delivery and Biomedical Applications. n.d.
- [155] Georgouvelas D, Abdelhamid HN, Li J, Edlund U, Mathew AP. All-cellulose functional membranes for water treatment: Adsorption of metal ions and catalytic decolorization of dyes. Carbohydr Polym 2021;264:118044.

https://doi.org/10.1016/j.carbpol.2021.118044.

- [156] Nasser Abdelhamid H, Mathew AP. Cellulose-zeolitic imidazolate frameworks (CelloZIFs) for multifunctional environmental remediation: Adsorption and catalytic degradation. Chem Eng J 2021;426:131733. https://doi.org/10.1016/j.cej.2021.131733.
- [157] Abdelhamid HN, Sultan S, Mathew AP. 3D printing of cellulose/leaf-like zeolitic imidazolate frameworks (CelloZIF-L) for adsorption of carbon dioxide (CO2) and heavy metal ions. Dalt Trans 2023. https://doi.org/10.1039/D2DT04168E.
- [158] Nasser Abdelhamid H, Georgouvelas D, Edlund U, Mathew AP. CelloZIFPaper: Cellulose-ZIF Hybrid Paper for Heavy Metal Removal and Electrochemical Sensing. Chem Eng J 2022:136614. https://doi.org/10.1016/j.cej.2022.136614.
- [159] Abdelhamid H, Georgouvelas D, Edlund U, Mathew A. CelloZIFPaper: Cellulose-ZIF Hybrid Paper for Heavy Metal Removal and Electrochemical Sensing. ChemRxiv Cambridge Cambridge Open Engag 2022. https://doi.org/10.26434/chemrxiv-2022gwdxf.
- [160] Valencia L, Abdelhamid HN. Nanocellulose leaf-like zeolitic imidazolate framework (ZIF-L) foams for selective capture of carbon dioxide. Carbohydr Polym 2019;213:338–45. https://doi.org/10.1016/j.carbpol.2019.03.011.
- [161] Abdelhamid HN. Dielectric, Thermal, and Electrical Conductivity Properties of Biodegradable Polymer Nanocomposites. Res Sq 2022. https://doi.org/10.21203/rs.3.rs-2003331/v1.
- [162] Abdelhamid HN. Biodegradable Polymer Nanocomposites: A Review of Properties. ChemRevix 2022. https://doi.org/10.26434/chemrxiv-2022-npnrs.
- [163] Abdelhamid HN, Wu H-F. Polymer dots for quantifying the total hydrophobic pathogenic lysates in a single drop. Colloids Surfaces B Biointerfaces 2014;115:51–60. https://doi.org/10.1016/j.colsurfb.2013.11.013.
- [164] Abdelhamid HN. Chitosan-Based Nanocarriers for Gene Delivery. Nanoeng. Biomater., Wiley; 2022, p. 91–105. https://doi.org/10.1002/9783527832095.ch4.
- [165] Dowaidar M, Nasser Abdelhamid H, Hällbrink M, Langel Ü, Zou X. Chitosan enhances gene delivery of oligonucleotide complexes with magnetic nanoparticles– cell-penetrating peptide. J Biomater Appl 2018;33:392–401. https://doi.org/10.1177/0885328218796623.
- [166] Abdelhamid HN, Lin YC, Wu H-F. Magnetic nanoparticle modified chitosan for surface enhanced laser desorption/ionization mass spectrometry of surfactants. RSC Adv 2017;7:41585–92. https://doi.org/10.1039/C7RA05982E.
- [167] Abdelhamid HN, Lin YC, Wu H-F. Thymine chitosan nanomagnets for specific preconcentration of mercury(II) prior to analysis using SELDI-MS. Microchim Acta 2017;184:1517–27. https://doi.org/10.1007/s00604-017-2125-3.
- [168] Abdelhamid HN, Dowaidar M, Langel Ü. Carbonized chitosan encapsulated hierarchical porous zeolitic imidazolate frameworks nanoparticles for gene delivery. Microporous Mesoporous Mater 2020;302:110200. https://doi.org/10.1016/j.micromeso.2020.110200.
- [169] Abdelhamid HN. Metal-organic frameworks (MOFs) as a unique theranostic nanoplatforms for therapy and imaging. Inorg. Nanosyst., Elsevier; 2023, p. 323–50. https://doi.org/10.1016/B978-0-323-85784-0.00006-6.
- [170] Abdelhamid HN, Wilk-Kozubek M, El-Zohry AM, Bermejo Gómez A, Valiente A, Martín-Matute B, et al. Luminescence properties of a family of lanthanide metalorganic frameworks. Microporous Mesoporous Mater 2019;279:400–6. https://doi.org/10.1016/j.micromeso.2019.01.024.
- [171] Abdelhamid HN. Lanthanide Metal-Organic Frameworks and Hierarchical Porous Zeolitic Imidazolate Frameworks: Synthesis, Properties, and Applications. Stockholm

University, Faculty of Science, 2017. https://doi.org/oai:DiVA.org:su-146398.

- [172] Yao Q, Bermejo Gómez A, Su J, Pascanu V, Yun Y, Zheng H, et al. Series of Highly Stable Isoreticular Lanthanide Metal–Organic Frameworks with Expanding Pore Size and Tunable Luminescent Properties. Chem Mater 2015;27:5332–9. https://doi.org/10.1021/acs.chemmater.5b01711.
- [173] Abdelhamid HN, Bermejo-Gómez A, Martín-Matute B, Zou X. A water-stable lanthanide metal-organic framework for fluorimetric detection of ferric ions and tryptophan. Microchim Acta 2017;184:3363–71. https://doi.org/10.1007/s00604-017-2306-0.
- [174] Yang Y, Shen K, Lin J, Zhou Y, Liu Q, Hang C, et al. A Zn-MOF constructed from electron-rich π-conjugated ligands with an interpenetrated graphene-like net as an efficient nitroaromatic sensor. RSC Adv 2016;6:45475–81. https://doi.org/10.1039/C6RA00524A.
- [175] Saleh MR, El-Bery HM, Abdelhamid HN. Co@ZIF-8/TiO2 Heterojunction for Green Hydrogen Generation. Appl Organomet Chem 2022. https://doi.org/10.1002/aoc.6995.
- [176] Abdelhamid HN. Dehydrogenation of sodium borohydride using cobalt embedded zeolitic imidazolate frameworks. J Solid State Chem 2021;297:122034. https://doi.org/10.1016/j.jssc.2021.122034.
- [177] Abdelhamid HN, Goda MN, Said AE-AA. Selective dehydrogenation of isopropanol on carbonized metal–organic frameworks. Nano-Structures & Nano-Objects 2020;24:100605. https://doi.org/10.1016/j.nanoso.2020.100605.
- [178] Abdelhamid HN. Hierarchical Porous Zeolitic Imidazolate Frameworks: Microporous to Macroporous Regime, 2022, p. 431–47. https://doi.org/10.1007/978-3-030-85397-6_14.
- [179] Abdelhamid HN. Hierarchical porous ZIF-8 for hydrogen production via the hydrolysis of sodium borohydride. Dalt Trans 2020;49:4416–24. https://doi.org/10.1039/D0DT00145G.
- [180] Abdelhamid HN, Zou X. Template-free and room temperature synthesis of hierarchical porous zeolitic imidazolate framework nanoparticles and their dye and CO2 sorption. Green Chem 2018;20:1074–84. https://doi.org/10.1039/c7gc03805d.
- [181] Abdelhamid HN. Dye encapsulation and one-pot synthesis of microporous– mesoporous zeolitic imidazolate frameworks for CO 2 sorption and adenosine triphosphate biosensing. Dalt Trans 2023. https://doi.org/10.1039/D2DT04084K.
- [182] Abdellatif ABA, El-Bery HM, Abdelhamid HN, El-Gyar SA. ZIF-67 and Cobaltbased@heteroatom-doped carbon nanomaterials for hydrogen production and dyes removal via adsorption and catalytic degradation. J Environ Chem Eng 2022;10:108848. https://doi.org/10.1016/j.jece.2022.108848.
- [183] Abdelhamid HN. Zeolitic imidazolate frameworks (ZIF-8, ZIF-67, and ZIF-L) for hydrogen production. Appl Organomet Chem 2021;35:e6319. https://doi.org/10.1002/aoc.6319.
- [184] Abdelhamid HN. A Review on Removal of Carbon Dioxide (CO2) using Zeolitic Imidazolate Frameworks: Adsorption and Conversion via Catalysis. Cambridge Open Engag 2022. https://doi.org/10.26434/chemrxiv-2022-k23gz.
- [185] Abdelhamid HN. Removal of carbon dioxide using zeolitic imidazolate frameworks: Adsorption and conversion via catalysis. Appl Organomet Chem 2022;36:e6753. https://doi.org/10.1002/aoc.6753.
- [186] Abdelhamid HN. Dye encapsulated hierarchical porous zeolitic imidazolate frameworks for carbon dioxide adsorption. J Environ Chem Eng 2020;8:104008. https://doi.org/10.1016/j.jece.2020.104008.
- [187] Abdelhamid HN. Salts Induced Formation of Hierarchical Porous ZIF-8 and Their

Applications for CO2 Sorption and Hydrogen Generation via NaBH4 Hydrolysis. Macromol Chem Phys 2020;221:2000031. https://doi.org/10.1002/macp.202000031.

- [188] Abdelhamid HN. Zinc hydroxide nitrate nanosheets conversion into hierarchical zeolitic imidazolate frameworks nanocomposite and their application for CO2 sorption. Mater Today Chem 2020;15:100222. https://doi.org/10.1016/j.mtchem.2019.100222.
- [189] Abdelhamid HN, Mathew AP. In-situ growth of zeolitic imidazolate frameworks into a cellulosic filter paper for the reduction of 4-nitrophenol. Carbohydr Polym 2021;274:118657. https://doi.org/10.1016/j.carbpol.2021.118657.
- [190] Abdelhamid HN, Huang Z, El-Zohry AM, Zheng H, Zou X. A Fast and Scalable Approach for Synthesis of Hierarchical Porous Zeolitic Imidazolate Frameworks and One-Pot Encapsulation of Target Molecules. Inorg Chem 2017;56:9139–46. https://doi.org/10.1021/acs.inorgchem.7b01191.
- [191] Abdelhamid HN. Biointerface between ZIF-8 and biomolecules and their applications. Biointerface Res Appl Chem 2021;11:8283–97. https://doi.org/10.33263/BRIAC 111.82838297.
- [192] Abdelhamid HN. Zeolitic Imidazolate Frameworks (ZIF-8) for Biomedical Applications: A Review. Curr Med Chem 2021;28:7023–75. https://doi.org/10.2174/0929867328666210608143703.
- [193] Abdelhamid HN, Dowaidar M, Hällbrink M, Langel Ü. Gene delivery using cell penetrating peptides-zeolitic imidazolate frameworks. Microporous Mesoporous Mater 2020;300:110173. https://doi.org/10.1016/j.micromeso.2020.110173.
- [194] Abd El-Aziz FE-ZA, Ebrahem NE, Abdelhamid HN. A comparative study of the toxic effect of ZIF-8 and ZIF-L on the colonization and decomposition of shaded outdoor mice carrions by arthropods. Sci Rep 2022;12:14240. https://doi.org/10.1038/s41598-022-18322-5.
- [195] Sadek AA, Abd-Elkareem M, Abdelhamid HN, Moustafa S, Hussein K. Enhancement of critical-sized bone defect regeneration using UiO-66 nanomaterial in rabbit femurs. BMC Vet Res 2022;18:260. https://doi.org/10.1186/s12917-022-03347-9.
- [196] Abdelhamid HN. UiO-66 as a catalyst for hydrogen production via the hydrolysis of sodium borohydride. Dalt Trans 2020;49:10851–7. https://doi.org/10.1039/D0DT01688H.
- [197] Abdelhamid HN. Solid Acid Zirconium Oxo Sulfate/Carbon-Derived UiO-66 for Hydrogen Production. Energy & Fuels 2021;35:10322–6. https://doi.org/10.1021/acs.energyfuels.1c00516.
- [198] Goda MN, Abdelhamid HN, Said AE-AA. Zirconium Oxide Sulfate-Carbon (ZrOSO4@C) Derived from Carbonized UiO-66 for Selective Production of Dimethyl Ether. ACS Appl Mater Interfaces 2020;12:646–53. https://doi.org/10.1021/acsami.9b17520.
- [199] Shamroukh W, Abdelhamid HN. Fenton-like Cerium Metal–Organic Frameworks (Ce-MOFs) for Catalytic Oxidation of Olefins, Alcohol, and Dyes Degradation. J Clust Sci 2022. https://doi.org/10.1007/s10876-022-02402-7.
- [200] Abdelhamid HN, Sharmoukh W. Intrinsic catalase-mimicking MOFzyme for sensitive detection of hydrogen peroxide and ferric ions. Microchem J 2021;163:105873. https://doi.org/10.1016/j.microc.2020.105873.
- [201] Abdelhamid HN, Mahmoud GA-E, Sharmouk W, Sharmoukh W. Correction: A cerium-based MOFzyme with multi-enzyme-like activity for the disruption and inhibition of fungal recolonization. J Mater Chem B 2020;8:7557–7557. https://doi.org/10.1039/D0TB90139C.
- [202] Abdelhamid HN, Mahmoud GA-E, Sharmouk W. A cerium-based MOFzyme with

multi-enzyme-like activity for the disruption and inhibition of fungal recolonization. J Mater Chem B 2020;8:7548–56. https://doi.org/10.1039/D0TB00894J.

- [203] Abdelhamid HN. MOFTextile: Metal-Organic Frameworks Nanosheets Incorporated Cotton Textile for Selective Vapochromic Sensing and Capture of Pyridine. Appl Organomet Chem 2023:10.1002/aoc.7078. https://doi.org/10.1002/aoc.7078.
- [204] Emam HE, Abdelhamid HN, Abdelhameed RM. Self-cleaned photoluminescent viscose fabric incorporated lanthanide-organic framework (Ln-MOF). Dye Pigment 2018;159:491–8. https://doi.org/10.1016/j.dyepig.2018.07.026.
- [205] Abdelhamid HN, Mathew A. Cellulose-Metal Organic Frameworks (CelloMOFs) Hybrid Materials and their Multifaceted Applications: A Review. Coord Chem Rev 2022;451:214263. https://doi.org/10.1016/j.ccr.2021.214263.
- [206] Abdel-Magied AF, Abdelhamid HN, Ashour RM, Zou X, Forsberg K. Hierarchical porous zeolitic imidazolate frameworks nanoparticles for efficient adsorption of rareearth elements. Microporous Mesoporous Mater 2019;278:175–84. https://doi.org/10.1016/j.micromeso.2018.11.022.
- [207] Abdel-Magied AF, Abdelhamid HN, Ashour RM, Fu L, Dowaidar M, Xia W, et al. Magnetic Metal-Organic Frameworks for Efficient Removal of Cadmium(II), and Lead(II) from Aqueous Solution. J Environ Chem Eng 2022:107467. https://doi.org/10.1016/j.jece.2022.107467.
- [208] Abdelhamid HN. High performance and ultrafast reduction of 4-nitrophenol using metal-organic frameworks. J Environ Chem Eng 2021;9:104404. https://doi.org/10.1016/j.jece.2020.104404.
- [209] Abdelhamid HN, Mahmoud GA. Antifungal and Nanozyme Activities of Metal– Organic Framework-derived CuO@C. Appl Organomet Chem 2023;37. https://doi.org/10.1002/aoc.7011.
- [210] Kassem AA, Abdelhamid HN, Fouad DM, Ibrahim SA. Metal-organic frameworks (MOFs) and MOFs-derived CuO@C for hydrogen generation from sodium borohydride. Int J Hydrogen Energy 2019;44:31230–8. https://doi.org/10.1016/j.ijhydene.2019.10.047.
- [211] Abdelhaleem A, Abdelhamid HN, Ibrahim MG, Chu W. Photocatalytic Degradation of Paracetamol Using Photo-Fenton-Like Metal-Organic Framework-Derived Cuo@C Under Visible Led. SSRN Electron J 2022. https://doi.org/10.2139/ssrn.4157065.
- [212] Kassem AA, Abdelhamid HN, Fouad DM, Ibrahim SA. Catalytic reduction of 4nitrophenol using copper terephthalate frameworks and CuO@C composite. J Environ Chem Eng 2021;9:104401. https://doi.org/10.1016/j.jece.2020.104401.
- [213] Kassem AA, Abdelhamid HN, Fouad DM, Ibrahim SA. Hydrogenation reduction of dyes using metal-organic framework-derived CuO@C. Microporous Mesoporous Mater 2020;305:110340. https://doi.org/10.1016/j.micromeso.2020.110340.
- [214] Al Kiey SA, Abdelhamid HN. Metal-organic frameworks (MOFs)-derived Co3O4@N-doped carbon as an electrode materials for supercapacitor. J Energy Storage 2022;55:105449. https://doi.org/10.1016/j.est.2022.105449.
- [215] El-Bery HM, Abdelhamid HN. Photocatalytic hydrogen generation via water splitting using ZIF-67 derived Co3O4@C/TiO2. J Environ Chem Eng 2021;9:105702. https://doi.org/10.1016/j.jece.2021.105702.
- [216] Abdelhamid HN, Al Kiey SA, Sharmoukh W. A high-performance hybrid supercapacitor electrode based on ZnO/nitrogen-doped carbon nanohybrid. Appl Organomet Chem 2021. https://doi.org/10.1002/aoc.6486.
- [217] Soliman AIA, Abdel-Wahab A-MA, Abdelhamid HN. Hierarchical porous zeolitic imidazolate frameworks (ZIF-8) and ZnO@N-doped carbon for selective adsorption and photocatalytic degradation of organic pollutants. RSC Adv 2022;12:7075–84.

https://doi.org/10.1039/D2RA00503D.

- [218] Soliman AIA, Abdelhamid HN, Aboel-Magd A. Abdel-Wahab. Hierarchical Porous Zeolitic Imidazolate Frameworks (ZIF-8) and ZnO@N-doped Carbon for Selective Adsorption and Photocatalytic Degradation of Organic Pollutants. ChemRxiv Cambridge Cambridge Open Engag 2022; 2022:10.26434/chemrxiv-2022-rwvtp. https://doi.org/10.26434/chemrxiv-2022-rwvtp.
- [219] Goda MN, Said AE-AA, Abdelhamid HN. Highly selective dehydration of methanol over metal-organic frameworks (MOFs)-derived ZnO@Carbon. J Environ Chem Eng 2021;9:106336. https://doi.org/10.1016/j.jece.2021.106336.
- [220] Abdellah AR, El-Adasy A-BA, Atalla AA, Aly KI, Abdelhamid HN. Palladium nanocrystals-embedded covalent organic framework (Pd@COF) as efficient catalyst for Heck cross-coupling reaction. Microporous Mesoporous Mater 2022:111961. https://doi.org/10.1016/j.micromeso.2022.111961.
- [221] Abdellah A, El-Adasy A, Atalla A, Aly K, Abdelhamid H. Palladium Nanocrystalsembedded Covalent Organic Framework (Pd@COF) as Efficient Catalyst for Heck Cross-Coupling Reaction. ChemRxiv Cambridge Cambridge Open Engag 2022. https://doi.org/10.26434/chemrxiv-2022-jhv7t.
- [222] Abdellah AR, Abdelhamid HN, El-Adasy A-BAAM, Atalla AA, Aly KI. One-pot synthesis of hierarchical porous covalent organic frameworks and two-dimensional nanomaterials for selective removal of anionic dyes. J Environ Chem Eng 2020;8:104054. https://doi.org/10.1016/j.jece.2020.104054.
- [223] Nguyen TA, Gupta RK. Covalent Organic Frameworks. Boca Raton: CRC Press; 2022. https://doi.org/10.1201/9781003206507.
- [224] Nguyen TA, Gupta RK, Abdelhamid HN. Covalent Organic Frameworks. Boca Raton: CRC Press; 2022. https://doi.org/10.1201/9781003206507.
- [225] Ibrahim M, Abdelhamid HN, Abuelftooh AM, Mohamed SG, Wen Z, Sun X. Covalent organic frameworks (COFs)-derived nitrogen-doped carbon/reduced graphene oxide nanocomposite as electrodes materials for supercapacitors. J Energy Storage 2022;55:105375. https://doi.org/10.1016/j.est.2022.105375.
- [226] Ibrahim M, Abdelhamid HN, Abuelftooh AM, Mohamed SG, Wen Z, Sun X. Covalent Organic Frameworks-Derived Nitrogen-Doped Carbon/Reduced Graphene Oxide as Electrodes for Supercapacitor. SSRN Electron J 2022. https://doi.org/10.2139/ssrn.4063571.
- [227] Ibrahim M, Fayed MG, Mohamed SG, Wen Z, Sun X, Abdelhamid and HN. High-Performance Lithium-Ion Battery and Supercapacitors Using Covalent Organic Frameworks (COFs)/Graphitic Carbon Nitride (g-C3N4)-Derived Hierarchical N-Doped Carbon. ACS Appl Energy Mater 2022. https://doi.org/https://doi.org/10.1021/acsaem.2c02415.
- [228] Ibrahim M, G. Fayed M, G. Mohamed S, Wen Z, Sun X, Nasser Abdelhamid H. High-Performance Lithium-Ion Battery and Supercapacitors Using Covalent Organic Frameworks (COFs)/Graphitic Carbon Nitride (g-C3N4)-Derived Hierarchical N-Doped Carbon. ACS Appl Energy Mater 2022;5:12828–36. https://doi.org/10.1021/acsaem.2c02415.