1	Yeast driven self exfoliation and functionalisation of graphene
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16	We report an eco-friendly, economical and green approach to synthesize few-layers of
17	functionalised graphene from large graphite flakes and chunks without the aid of any external
18	mechanical forces. Here, we demonstrate a fermentation process aided by friendly microbes
19	like yeast to effortlessly exfoliate graphite to graphene with a high yield. The evident flaking
20	and delamination of graphene layers, characterized using scanning electron microscopy (SEM),
21	atomic force microscopy (AFM) and Raman spectroscopy, confirmed the successful
22	exfoliation of graphite to few-layer graphene (~10 nm in thickness). The gate-to-gate life cycle
23	assessment of the developed method identified the environmental hotspots of the process. The
24	biofunctionalized graphene (as suggested by Fourier transform infrared (FTIR) spectroscopy)
25	was explored for its application in NH_4^+ sensing, where it exhibited the limit of detection to be
26	~ 20 ppm. The detection mechanism was explained based on Raman and FTIR spectroscopy
27	which revealed the functional state and lattice changes introduced in the graphene layers by
28	$\mathrm{NH_{4}^{+}}$ ions. Our study demonstrates a novel, cost-effective exfoliation of graphite using off-the-
29	shelf materials in laboratories or kitchens and has the potential to be scaled up to industry
30	levels. The yeast cells also exhibited their potential in flaking off the bulk MoS ₂ structure into
31	few-layer MoS ₂ sheets.
32	Keywords: graphene, economical, green, dry yeast, ammonium ions detection, life cycle

33 assessment

34 Research into two-dimensional (2D) materials has evolved in the last ten years from the labscale proof of concept to pilot-scale demonstrations.^{1,2} One of the key challenges in converting 35 36 the prototype to a product is the energy cost of 2D materials' production; since all of the 37 material synthesis uses external energy sources such as electricity, chemicals, mechanical force 38 etc., and limits to decrease the cost of the production.^{1,3} However, there are several biological 39 processes wherein nature produces compounds without the aid of external energy. For example, 40 collagen in living species is produced without any external energy source.⁴ Previously, there have been attempts to mimic such processes using yeast for the production of chiral centres 41 42 etc.⁵ However, such self-mediated natural processes have never been explored to produce 2D 43 materials. Here, we report the self-exfoliation and functionalization of graphene by using yeast 44 in the presence of glucose without any external energy source. We show that, in such a process, 45 the yeast cells exert much-needed mechanical shear forces to exfoliate graphite from the edges. 46 In addition to the self-exfoliation, this process produces ethanol during fermentation, which 47 helps disperse the exfoliated graphene to form a colloidal dispersion.

48 Fig.1(a) demonstrates the schematic of the process used for the functionalized graphene 49 production. Dry yeast cells were inoculated with graphite powder suspension in glucose 50 solution at room temperature. The solution in the flask turned dark and turbid after one week 51 when visually inspected, as seen in Fig. 1(b). On the other hand, no visible differences were 52 noticed in the reference yeast glucose mixture without graphite and also the reference glucose 53 graphite mixture solution without the yeast (Fig. 1b). The appearance of dark colloidal 54 dispersion when yeast and graphite simultaneously present in glucose solution suggests 55 exfoliation of graphite. As supporting evidence of exfoliation, we anchored thin graphite layers by mechanical exfoliation to an Si/SiO₂ (290nm) samples. As seen in Fig. 1(c), a few layer 56 57 graphene (~10 layers, measured by optical contrast) was successfully seen to be delaminated 58 when immersed inside yeast-glucose solution for 1 week. To further characterise the exfoliation 59 of graphite in yeast-glucose dispersions, the samples were subjected to a differential 60 centrifugation process to remove yeast, and thick graphite flakes (details given in Method 61 section). The obtained suspensions were drop-casted on Si substrates for all further 62 characterization. The AFM micrograph, Fig. 1 (d), showed the uniform distribution of graphene 63 flakes where the thickness profile of the prepared layers, as obtained from the micrograph and 64 histogram, revealed the thickness to be ~ 10 nm, thereby suggesting the exfoliation of bulk 65 graphite. In order to further characterise the exfoliated graphene samples, Raman and TEM 66 analyses were performed. The obtained Raman spectra, Fig. 1(e), suggested the exfoliation of 67 yeast-treated graphite as indicated by the peak at 1350 cm⁻¹ and ~ 2700 cm⁻¹ corresponding to D and 2D bands of few-layer graphene, respectively. The finite D peak for the yeast treated precursor is owed to the functionalization of prepared sheets due to functional groups introduced during yeast mediated exfoliation.⁶ The 2D band at 2700 cm⁻¹, on the other hand, corresponds to the number of layers in the graphene and intensifies with decreasing number of layers.⁷ The TEM micrograph, Fig. 1 (f), of the graphene suspension further demonstrated the presence of a few layer graphene flake in the dispersion.

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76 Fig 1: (a) Schematic of yeast-mediated exfoliation of graphite at room temperature in the presence of glucose, (b) photograph of the graphite 77 in yeast and glucose solution on day 0 and day 7. The bottom image is a photo of the control sample with yeast fermenting glucose solution 78 without graphite. (c) Optical micrograph of an anchored thin graphite flake (light green colour) on an oxidised silicon substrate with a few-79 layer graphene at the edge on Day 0 (pointed by the red dotted oval). After immersing in yeast-glucose solution, the few-layer graphene was 80 found to be disappeared due to exfoliation. (d) Histogram of the thickness distribution of the exfoliated flakes obtained after keeping 81 graphite in yeast and glucose solution for one week measured in AFM. Inset is an AFM image of the drop casted dispersion after differential 82 centrifugation (e) Raman spectrum for graphite and graphene (before and after exfoliation) and (f) TEM image showing the presence of one, 83 two and three layers of graphene at the edge of a few layer flake.

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The exfoliation mechanism was studied using microscopic techniques including AFM and SEM, given in supplementary Fig. 1 and 2. The SEM micrographs suggested the yeast cells' attachment to the carbon layers where the shearing and tearing of the sheets was mediated by the growth and reproduction of the attached cells which occurs asexually through budding in case of *Saccharomyces cerevisiae*.⁸ The separation of a daughter cell from the mother cell is hypothesised to result in layers dissociation and the formation of few-layer graphene structures.

91 The cell growth based AFM micrographs confirmed the exfoliation to be controlled by yeast 92 cell growth where the graphite was considerably thinned down to few layers within few days 93 of incubation. In addition, we suspect that the exfoliation will continue if thinner flakes are 94 subjected to a fresh growing yeast. The thickness-to-time plot revealed a significant reduction 95 in the thickness within 4 hours of incubation which corresponded to the logarithmic growth 96 phase of dry yeast cells (Supplementary Fig. 2 (c)). Further thinning of the sheets was observed 97 where the reduction was observed to reach ~ 10 nm in one week. The uniformity of the 98 exfoliation process was established by estimating the standard deviation in the thickness of the 99 prepared graphene with respect to incubation time where increasing incubation time led to the 100 exfoliated structure with comparable thickness. The action of yeast cells on the edges of the 101 prototype, graphite, was examined by studying the lateral dimension and aspect ratio of the 102 exfoliated structure with respect to increasing incubation time with yeast cells, Supplementary 103 Fig. 2 (d). The lateral dimension of the obtained layers was significantly reduced with 104 increasing time which suggests the tearing the graphite from the edges by the yeast cells as 105 shown in SEM micrographs. The aspect ratio of the obtained exfoliated flakes was observed to 106 be increasing for up to 6 h of incubation. This indicated the formation of extremely thin 107 graphene structure suggesting higher rate of thinning from within the layers of graphite in 108 comparison to rate observed for delayering from the edges (out of plane for [001]). The 109 observation can be attributed to the feasibility in overcoming the Van der Waals bonding in 110 comparison to covalent bonding. The TEM analyses of the prepared few layer graphene has 111 been elaborated in the supplementary section, supplementary Fig 3.

112 The functional and optical characterization of the exfoliated graphene layers was revealed by 113 spectroscopic analyses. The UV absorption study of the prepared graphene, Fig. 2(a), 114 demonstrated a characteristic peak at ~270 nm, assigned to $\pi \rightarrow \pi^*$ transition of aromatic C–C bonds.⁹ The spectra were monitored with respect to the incubation period which showed the 115 116 highest absorption to occur after 1 week of incubation with yeast cells, confirming the AFM 117 analyses. The surface functionalization state of the obtained graphene flake was studied using 118 FTIR and shown in Fig. 2(b). The distinct peaks were obtained at ~ 1630 and 1210 cm⁻¹ which 119 corresponded to the functional groups i.e., carboxylic and phosphate groups arising from yeast 120 cell wall, thus suggesting the yeast cells based functionalization of the exfoliated graphene.¹⁰ 121 Post functional state analyses with FTIR spectroscopy, the exfoliated graphene was further 122 characterized using Raman spectroscopy to identify the impact of yeast cells' growth 123 parameters on its lattice and structural properties. The yeast cells' concentration in the 124 suspension for graphite thinning was optimized by plotting the Raman spectra, Fig. 2 (c), for a 125 range of yeast cells' concentration (0-0.5% (w/v)). The significant change in the integrated area 126 ratio (A_D/A_G) suggests the presence of functionalized graphene sheets owed to the yeast cells 127 mediated exfoliation of graphite into graphene as already been established by FTIR study. 128 However, for increased cells' concentration, the cell suspension was observed to be accrued 129 with yeast cells forming a biofilm on the surface and impeding the aeration thereby limiting the cell growth. Therefore, the yeast cell concentration is considered to be a trade of exfoliation 130 131 efficacy versus functional cell growth. Ordinarily, the transition from bulk to few layer carbon 132 sheets results in the significant changes in the Raman spectroscopy parameters such as D/G, 133 FWHM and band positions which directly correspond to functional and lattice state of the exfoliated sheets. Therefore, the incubation time (cell growth) based Raman spectra were 134 135 obtained with incubation intervals of 0 min, 6 h, 1 day, and 1 week (supplementary file, Figure 136 4). The change in the G peak position and integrated area ratio (A_D/A_G) with respect to incubation time is shown in Fig. 2 (d). 137





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140Fig. 2: (a) Time dependent UV-Vis spectra for (I), (II), (III), (IV), and (V) defined as 0 min, 1h, 6 h, 12 h, and 1 week respectively, (b) FTIR141spectra for obtained graphene suspension, (c) A_D/A_G plot with respect to yeast cells concentration with (I), (II), (III), and (IV) defined as 0,1420.02, 0.2 and 0.5 % respectively, (d) A_D/A_G and G band position plot with respect to yeast cells incubation period, (e) FWHM and L_a with143respect to incubation period with (A), (B), (C), and (D) defined as 0 min, 6 h, 1 day and 1 week respectively, (f) High resolution XPS survey144obtained for C 1s and (g and h) SEM scans of exfoliated graphite, (i and j) UV-Vis and Raman spectrum for graphene obtained at 50° C145Incubation

The red shift in the G band suggests the presence of strained sheets,¹¹ which can be due to the 147 148 stretching of the sheets by the yeast cells leading to elongated C-C bonds (supplementary Fig. 149 2). Further, the A_D/A_G ratio, influenced by the functionalization of the prepared structure, was found to increase with increasing incubation time. The spectra, obtained for 0 min of incubation 150 151 time, exhibited a feeble D peak suggesting the precursors to be free of defects. The 152 intensification of the D band with increasing incubation time is attributed to the 153 functionalization of the edges owing to the yeast cells driven exfoliation.⁶ The A_D/A_G ratio was further explored to evaluate the in plane sp² crystallite size, L_a, according to the relation given 154 by Cançado et al. for graphitic materials, wherein La was reported to be inversely proportional 155 to A_D/A_{G_a} ¹² The relative L_a was plotted with respect to incubation time as shown in Fig. 2 (e). 156 157 The plot suggests the gradually decreasing lateral dimension of the exfoliated graphene for up to 1 week of incubation. The obtained pattern was found to be in agreement with the changes 158 159 observed in the lateral dimension of the layers from AFM micrographs (Supplementary Fig. 2 (e)). The FWHM value for the G band suggested the formation of exfoliated structure as higher 160 G_{FWHM} corresponds to increased interlayer spacing (d₀₀₂).¹³ Fig. 2 (e) shows the change in the 161 162 FWHM with incubation time where the value was found to be gradually increasing with 163 increasing time and hinted at the disruption of graphite for the formation of few layer graphene. 164 The elemental survey of the exfoliated graphene revealed the presence of C, O and N atomic 165 states. The XPS spectra of C 1s, (Fig. 2 (f)), with deconvoluted peaks at ~ 284.8 eV, 286 eV and 288 eV are assigned to sp² C carbon, C-OH and O=C-O.¹⁴ The presence of oxygen 166 containing functional groups in C 1s spectra due to the yeast cells in the graphene suspension. 167 168 Based on the changes observed for A_D/A_G, L_a, the G band position and the FWHM values with respect to incubation time, the exfoliation process can be divided into three stages (attachment, 169 170 division and separation of yeast cells) to elucidate the step wise thinning of graphite. The 171 attachment phase (I), Fig. 2 (g), starts immediately after incubation and as observed from the 172 plots, there are no significant changes in the lattice parameters during this stage. The 173 attachment stage often overlaps with the cell division, where the attached cells begin to divide causing the shearing and tearing of the substrate (graphite), significant changes in the lattice 174 175 parameters were observed at the second stage of exfoliation. Further, the stage III is 176 characterized by the separation stage, which is marked with the cell division phase and is 177 completed within 1 weeks of incubation, Fig. 2(h).

178 In order to detect the impact of elevated temperature on the exfoliation process, the reaction 179 mixture was incubated at 50° C for 1 week. The increased temperature was expected to increase the reaction rate without impacting the yeast cells' growth. The obtained exfoliated structure was examined using spectroscopic techniques to determine its impact on the exfoliation process. The UV-Vis absorption spectrum and the Raman spectrum indicated no significant changes in the exfoliation yield and lattice properties of the obtained exfoliated structure for incubation at higher temperature.

To gain further insight into the mechanism of yeast-assisted exfoliation of graphite as a 185 186 function of time using fully atomistic Molecular Dynamics (MD) simulations. Here, we discussed the role of thickness in exfoliating graphene sheets. To this end, we initially 187 considered two layers of graphite (area = 30 Å x 25 Å) without any defect which were confined 188 189 between two cell membranes of the yeast cell, modelled as a phospholipid bilayer made of 190 phosphatidylcholine (POPC) molecules (see methodology and supplementary section for details) at t=0 ps, as shown in Fig. 3 (a). Within t=16 ps of simulation time (Fig. 3 (b and c)), 191 192 The two-layers separation suggests that graphite-yeast interaction to be sufficiently strong to completely overcome the inter-graphite sheets Van der Waals (VdW) interactions. 193



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195 Fig. 3: Simulated exfoliation of graphene from yeast. Representative MD snapshots of the yeast and graphene layers at different simulation

196 time for (a-c) two layers (d-f) six layers. Initial structure at T=0ps and intermediate structure at T=16ps, 35ps, where graphene starts 197 exfoliating; (g) Comparative life cycle assessment of graphene production routes, see text for discussions

Similarly, we also considered six parallel graphite layers (area = 40 Å x 70 Å) shown in Fig. 3 198 199 (d), to determine whether the layer separation could also occur for thicker structures. As 200 observed for two-layer case, within t=16 ps of simulation time, layers started to separate, 201 showing again that the graphite/yeast interactions are strong enough to overcome the Van der 202 Waals forces of thicker layers. This phenomenon was supported by Fig. 3 (e and f), the 203 concentration profile of the graphene corresponding to the $[0 \ 0 \ 1]$ direction shows that the 204 confined fluid has a higher density closer to the membrane surfaces indicating the sticking to 205 be mediated by shear stress. The yeast sticking phenomena of six layer is slightly higher in 206 comparison to two-layers, this can be attributed to the higher surface area. In summary, the 207 MD results further support that the yeast-membrane interaction with graphite are sufficiently 208 strong to wet its surface and induce exfoliation by shear stress, even considering the thicker 209 graphite structures (further details given in the supplementary file).

210 A stand-alone LCA was performed using the yeast driven-exfoliation process experimental 211 data to identify the potential environmental hotspots by extrapolating the laboratory-scale data 212 to a hypothetical commercial scale (system boundary and other details have been provided in 213 the supplementary section). The outcomes significantly depend upon scale, production routes 214 and energy mix. Environmental profiles for end point characterization of the graphene 215 production are shown in Fig. 3 (g). Overall, de-ionized water (36.11%), electricity production 216 (35.41%) and glucose production (26.57%) were identified as environmental hot-spots in the 217 bio-exfoliation process. The electricity production is identified as the major contributing 218 emission towards human health while de-ionized water production contributed to both climate 219 change and resource depletion. Overall, the bio-exfoliation production route is established as 220 the green process compared to other approaches involving coal tar, rice husk, plastic waste and 221 paper waste. Thus, the shift towards using renewable-based energy sources provides a 222 promising route towards the reduced ecological impacts in the current geographical context.

The acid-base homeostasis in a human body is maintained by the kidneys, which eliminate the 223 224 acid (H⁺) in the form of NH₄⁺ through excretion. Low ammonium excretion is associated with 225 the impaired ability of the kidneys to remove the acids which is classified as chronic kidney 226 disease (CKD and further linked to an elevated risk of end-stage renal disease (ESRD), acidosis and kidney failure.¹⁵ Hence, monitoring the ammonium excretion level in patients at early 227 228 stages is effective to circumventing the occurrence of such unlikely clinical events. Previous 229 studies reported the utilization of graphene for ultra-low detection of ammonia (NH₃), where graphene sheets were functionalized to fabricate a nanocomposite film as sensor.¹⁶⁻¹⁸ 230 231 However, in the present study, the bio-prepared few-layer graphene exhibited the presence of

residual functional groups as shown in the FTIR study, which can effectively provide sites for 232 NH4⁺ absorption or binding, without requiring further post-synthesis modification.¹⁹ Therefore, 233 as-prepared few-layer graphene were explored for their use in low-level sensing of NH4⁺ ions 234 235 (sensing mechanism shown in Fig. 4 (a)). The binding of ammonium ions to graphene sheets 236 was examined using UV-Vis spectroscopic analyses. The change in UV-Vis spectra of 237 graphene was recorded for different concentrations of NH₄Cl solution as shown in Fig. 4 (b). 238 The absorption plots demonstrated a smooth decline in the characteristic absorbance peak at 239 ~ 270 nm with the increasing amount of analyte, NH₄⁺, which suggests the binding of 240 ammonium ions to few-layer graphene. In order to further investigate the UV absorption 241 behaviour of few-layer graphene in the presence of NH4⁺ ions, the absorbance with increasing 242 analyte amount (0.5 mM up to 3.75 mM) was plotted for different graphene amount (G-I to 243 G-V) (Fig. 4 (c and d)). Here, I to V define the increasing concentration of few-layer graphene, i.e., from 0.06% up to 0.15% (v/v). The change in the absorption was quantified and plotted 244 245 with respect to NH₄⁺ ions concentration for elucidating the binding affinity and saturation 246 conditions. The most significant reduction in the absorption was found to be for the graphene 247 concentration of 0.1% (v/v) which suggested the most effective binding and ammonium 248 absorption at the particular concentration. In Fig. 4 (e), G-III showed a linear decline in the 249 absorbance with NH₄⁺ concentration, which was fitted to establish a linear relationship between 250 absorbance changes and ammonium concentration. The calibration curve was further explored 251 to estimate the limit of detection (LoD) and the limit of quantification (LoQ). The detection 252 limit indicated the lowest concentration of ammonium ions which was detected by few-layer 253 graphene and was found to be ~ 20 ppm, while the quantification limit was estimated on the 254 basis of repeatability and precision and was calculated to be 1.8 mM. Further, the association 255 between the analytes and detector i.e. NH_4^+ ions and graphene respectively was studied using 256 the Benesi-Hildebrand equation, Fig. 4 (f), which estimated the binding constant (K_b) to be ~ 257 0.6×10^3 M⁻¹. The low value of the binding constant suggested the moderate binding as reflected by the estimated LoD and LoQ values. The affinity between the prepared few-layer 258 259 graphene and analyte, ammonium ions, is ordinarily governed by a plethora of factors i.e. the 260 number of layers in the graphene, the presence of surface groups, etc. The comparative plot was drawn to collate the ammonium ions detection carried out by graphene sheets. Here, the 261 262 biologically prepared few-layer graphene demonstrated a detection limit of 20 ppm which was comparable to that observed in previous reports, Fig. 4 (g).^{17,18,20} 263

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Fig. 4: (a) schematic for NH₄⁺ ions sensing by graphene layers where (I) and (II) signify the binding and incorporation of N atoms in the carbon hexagonal lattice, (b) change in the UV Vis absorption spectra for graphene with NH₄⁺addition, (c) decreasing absorbance for graphene solution at different concentration (I to V), (d) relative change in absorption plots with respect to graphene concentration, (e) calibration curve for change in absorption with respect to ammonium ions concentration, (f) Benesi–Hildebrand plot for determination of binding constant of graphene for ammonium ions, (g) the comparative plot with ammonium detection for graphene prepared using different methods I: chemiresistive graphene, II: hydrazinium graphene (HG) prepared from exfoliated graphene oxide, III:chemically derived graphene, IV: functionalized graphene (present work), (h) FTIR spectra for I, II, and III i.e. control, ammonium treated, and heated graphene suspension, (i) Raman spectra for I, II, and III i.e. control, NH₄⁺ ions treated, and heated graphene suspension.

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276 In order to gain further insights into the sensing mechanism, control and ammonium treated 277 graphene samples were examined for the changes in their surface chemical state and lattice 278 structure using FTIR and Raman spectroscopy (Fig. 4 (h and i). Here, the reproducibility of the 279 system was checked by heating the ammonium treated graphene and examining the changes 280 using spectroscopic techniques. The FTIR peaks revealed the changes in the surface chemical 281 composition of the graphene sheets upon NH₄Cl application. From the plot, Fig. 4(h), there are 282 significant changes observed in the NH₄⁺ treated graphene, in comparison to control and heat treated graphene-NH₄⁺. The peaks at \sim 3100 and \sim 1400 cm⁻¹, observed for ammonium treated 283 graphene, correspond to N-H stretching and bending respectively. Further, the band at ~1090 284 cm⁻¹ corroborates to the symmetric NH₃ signifying the coordination of NH₃ to Lewis acid sites 285 on the carbon surface (graphene layers).²¹ The peak at \sim 820 cm⁻¹, assigned to the deformation 286 vibration for C-H groups in carbohydrates, i.e., glucose here, was observed to be absent for 287

288 NH4⁺ treated graphene suspension. The absence of peak can be owed to the glucosamine 289 formation, where hydroxyl group at the anomeric carbon can be replaced with an amine group. 290 Therefore, the FTIR examination confirms the binding of ammonium ions to the carbon atoms 291 in the graphene structure. The binding was further validated by Raman spectroscopic analysis 292 where the control and treated samples were studied for the changes in the edge defects, disorder and functional groups. The plots demonstrate the significant changes in the graphene layers 293 294 upon NH4⁺ incorporation, which was inferred by increasing A_D/A_G. The obtained results 295 suggested the formation of C-N bonding and incorporation of nitrogen atom in the hexagonal 296 lattice thereby leading to functionalization. The FTIR and Raman behaviour of the heat treated 297 samples revealed that changes, observed in the ammonium treated graphene samples, were 298 found to be absent in the heated samples, which marks the reproducibility of the bioprepared 299 graphene as ammonium ions sensor. Hence, the biopreparation of graphene layers can be useful 300 for real life application in ammonium ions sensing in human urine to detect the changes due to 301 kidney dysfunctionality.

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303 Conclusion

304 Here, we have presented a method to bio-exfoliate graphite sheets into few-layers graphene, 305 which was elucidated using different structural and spectroscopic analyses. The changes in the 306 thickness reduction were thoroughly examined using AFM, which suggested the reduction to 307 ~10 nm thickness in one week of incubation and Raman spectroscopy also validated the formation of few-layers graphene structures. The IR spectroscopic examination revealed the 308 309 presence of functional groups on the prepared graphene which enabled its application for the ammonium ions sensing via UV-Vis absorption spectroscopy with LoD to be 20 ppm. The 310 311 affinity of the yeast cells for graphene was studied using molecular dynamics simulation which 312 suggested the exfoliation results from shear stress induced by the yeast and the yield to be 313 determined by the thickness and surface area of graphite. In summary, we present a new bio-314 exfoliation method to obtain few-layers graphene from graphite. The method is very simple 315 and it uses widely available yeast cells. It is non-expensive, green and scalable (industrial 316 scale). In principle, it can be used to exfoliate other layered materials (as demonstrated for 317 MoS₂ in the supplementary data), which opens new perspectives to obtain and use few-layer 318 structures.

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322 Methods

323 Yeast mediated exfoliation of graphite

324 Dry yeast cells (0.2% (w/v)) were incubated with graphite containing glucose solution (0.5%325 (w/v)). During the experiment, the yeast cells could reach and remain in the stationary phase 326 of growth. The visual change in the yeast containing solution was the first indication of 327 exfoliation of graphite.

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329 Characterization of the exfoliated graphene

330 Differential centrifugation in the steps of increasing RPMs from 4k (allowing yeast and larger 331 graphite chunks), followed by subjecting the supernatant to 14k RPM in 2 steps to collect the 332 precipitate with smaller exfoliated flakes. The precipitate or the pellet is diluted and rinsed in 333 water. It is further drop-casted on Si substrates with 300 nm SiO2 Height and surface profiles 334 of as prepared graphene sheets were studied using Atomic Force Microscopy (AFM, Agilent Technologies, USA, Model 5100). All the measurements were carried out in air at room 335 336 temperature (25° C) using a Silicon Nitride Tip (PPP-NCL) in the intermittent contact mode. 337 The Scanning Electron Microscope (SEM) micrographs of graphene were obtained with ZEISS 338 EVO 60 Scanning Electron Microscope with an Oxford EDS detector. The phase and lattice 339 characterization of the bio-exfoliated graphene sheets was carried out using powder X-Ray 340 Diffraction (XRD) with monochromatic nickel filtered Cu Ka radiation in the 20 range of 20 341 to 80°. The optical spectra of the prepared samples were recorded using a UV-1800 spectrophotometer, Shimadzu, Japan. The High Resolution- Transmission Electron 342 343 Microscopy (HR-TEM) images were produced using JEOL 2200FS electron microscope. The 344 Fourier Transform Infrared (FTIR) analysis of graphene was carried out using Bruker spectrometer through the range of 4000 to 400 cm⁻¹. The Raman spectra of graphene sheets 345 were acquired with an excitation wavelength of 532 nm (Witec UHTS 300). X-ray 346 347 photoelectron spectroscopy (XPS) measurements were performed with Thermoscientific spectrophotometer (NEXSA) with a micro focused monochromatic Al- K_{α} source (1486.6 eV). 348

349 **Computational study methodology**

To theoretically evaluate the structural stability of few-layers graphene and/or graphene/yeast membrane cells, we carried out fully atomistic Molecular Dynamics (MD) simulations, as implemented in the open-source code LAMMPS.²² The well-known Universal Force-Field (UFF) formalism was used to calculate the interatomic interactions and/or configurational energies.²³ For creating the simulation box, initially, a supercell of 50 Å x 80 Å x 135 Å was created that includes a 20 Å vacuum along with two layers of graphite confined between 356 membrane yeast cells. 20 Å buffer vacuum was used to avoid spurious interactions of the 357 supercell replication. Another supercell was created with the same dimension for the six-layers 358 graphite structure. As discussed above, this was used to determine the thickness relationship 359 with the exfoliation mechanism contributing to yield. The outer cell membrane of the yeast is 360 modeled as a phospholipid bilayer made of phosphatidylcholine (POPC) molecules 361 commensurate with the dimensions of the graphene-layered structures. The structural data of 362 the models are available upon request. For performing the simulations on the layered structures, 363 a variable cell relaxation calculation was performed to equilibrate the yeast cell layers by 364 constraining the graphite layers and to eliminate any bad contacts generated from the 365 layer/membrane building processes. To further avoid any residual structural stresses, NVT 366 equilibration MD runs were performed during 100 ps. Finally, the confined shear simulations were performed to examine the exfoliation mechanism by removing the constraints from 367 368 graphite layers, setting wall velocity (U) to 5Å/ps and simulation time (t) as 100ps. This kind 369 of simulation protocol has been extensively used in the literature.

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371 Life cycle assessment methodology

372 The environmental impacts of graphite exfoliation using biological synthesis were quantified 373 using the life cycle assessment (LCA) approach as per ISO 14040-44 (ISO 14040, 2006; ISO 374 14044, 2006). Four steps were involved in the process (a) goal and scope definition, (b) life 375 cycle inventory (LCI), (c) life cycle impact assessment (LCIA), and (d) life cycle interpretation. 376 This study's primary goal is to evaluate environmental impacts of graphite exfoliation using 377 biological synthesis. The scope of this LCA includes only the exfoliation of graphite into few-378 layers graphene (gate-to-gate). The upstream life cycle stages utilization of graphene and 379 disposal stages were excluded. The energy calculations were performed using specific heats of 380 materials and/or the power rating of the equipment used in the synthesis process. The location 381 of graphite exfoliation plant was considered Kharagpur, India, the source of materials and 382 chemicals was based on the local conditions (Kolkata, India). The eastern India electricity power mix was used for energy consumption. The study's functional unit was the exfoliation 383 384 of 1 ton of graphite. The functional unit was chosen to compare the feasibility of producing 385 graphene from waste-based feedstocks from an environmental perspective at an industrial 386 scale. The parameters and components in synthesis routes for graphite exfoliation were scaled 387 to the industrial level and were normalized to the functional unit. The scaled-up quantities of 388 materials and amount of energy were normalized to the functional unit, assuming that reaction 389 rates remain unaltered at the industrial scale. Environmental impacts were evaluated using

SimaPro 8.0.3 version software and Eco-invent database. IMPACT 2002+® method was used 390 391 for the characterization of end-point impacts. Since we are aiming to study the industrial-scale 392 exfoliation of graphite, we choose the process contributions which are most relevant to the 393 production processes. Sensitivity analysis was performed by varying the country's electricity 394 mix and quantity of deionized water. In the present investigation, a high-voltage electricity grid 395 mix of the Indian geographic location was used for modeling the scenarios. For sensitivity 396 analysis, the electricity mix of China and Norway was modeled to compare and evaluate the 397 impacts associated based on the shift towards reduction in the coal-based electricity mix. 398 Comparative LCA was performed with graphene production routes using (a) coal-tar pitch; (b) 399 Rice-husk; (c) Plastic waste; and (d) Paper Waste.

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401 Ammonium ions sensing experiments

402 The bioprepared graphene sheets were checked for NH_4^+ sensing *via* UV-Vis absorption 403 spectroscopy, where graphene suspension was mixed with NH_4Cl solution at different 404 concentrations (0.5 -3.75 mM). The change in the absorbance was quantified, and the 405 calibration plot was obtained to estimate LoD and LoQ. Further, to comprehend the sensing 406 mechanism, control and ammonium-treated graphene were examined under Raman and FTIR 407 spectroscopy for changes in the surface chemical state and lattice structure introduced by NH_4^+ 408 ions.

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