

High Pressure Homogenization – An Update on its Usage and Understanding

Pavan Inguva¹, Silvia Grasselli^{2*}, Paul W.S. Heng^{3*}

¹ Department of Chemical Engineering, Massachusetts Institute of Technology, 25 Ames Street, Cambridge, Massachusetts 02142, United States

² GEA Mechanical Equipment Italia S.p.A. Via A.M. da Erba Edoari 29, Parma 43123, Italy

³ GEA-NUS Pharmaceutical Processing Research Laboratory, Department of Pharmacy, National University of Singapore, 18 Science Drive 4, Singapore 117543

* Correspondence:

Silvia Grasselli, Silvia.Grasselli@gea.com

Paul W.S. Heng, paulwsheng@outlook.com

Abstract

Despite being a technology of several decades, high pressure homogenization (HPH) remains widely used in food and pharmaceutical industries, often as an essential unit operation in liquid product processing. Continual advances in the technology are made on multiple fronts, on equipment innovations by the manufacturers, new applications by users and advances in process understanding by multidisciplinary scientists alongside subject matter experts amongst industry practitioners. While HPH is comparatively simple conceptually, the homogenization process involves complex engineering physics which is influenced by the varied processing conditions and highly diverse inputs with each use-case requiring its own treatment. The successful application of a HPH process indubitably

requires practitioners to draw upon insights from multiple domains and the optimization for each case. Thus, this timely review aims to outline the more recent trends and advancements in HPH process understanding and novel applications involving HPH from both academic and industrial perspectives.

1. Introduction

High pressure homogenization (HPH) was first introduced in the early 20th century by Auguste Gaulin (Gaulin, 1904) for processing milk using pressures up to 30 MPa to improve product stability. The basic operational principle of HPH remains unchanged since and it involves using a high-pressure pump to force the fluid through a small orifice. The early successes expanded the user-base of HPH and it has become an integral unit operation in liquid product processing in several industries such as food and beverages (F&B) (Harte, 2016; Levy et al., 2020; Patrignani and Lanciotti, 2016), pharmaceuticals (Kluge et al., 2012; Yadav and Kale, 2020), waste water treatment (Zhang et al., 2012), material production and processing (Azoubel and Magdassi, 2010; Phanthong et al., 2018; Xu et al., 2011) and biotechnological processing (Kelly and Muske, 2004; Samarasinghe et al., 2012). This list is certainly non-exhaustive and will likely continue to expand.

In conjunction with the introduction of new HPH applications, much work from both industry and academia is required to better understand the impact of the HPH process. Various research studies have considered the theoretical aspects of the homogenization process such as how fluid flow in the HPH valve (Håkansson et al., 2010; Taghinia et al., 2016) and the mechanistic appreciation of complex processes like HPH assisted emulsification (Gupta et al., 2016a; Håkansson et al., 2013). Other studies were directed

at the operational aspects of HPH such as exploring methods for process optimization (Davoudpour et al., 2015; Dopp and Reuel, 2018) and improved process monitoring and characterization (Besseling et al., 2019; Ralbovsky et al., 2022). Concomitant equipment innovations such as the design of new homogenization valve geometries (Donsi et al., 2012; Gall et al., 2016; Yadav and Kale, 2020) and ultra-high pressure homogenization (UHPH) which employs homogenization pressures between 300-400 MPa (Georget et al., 2014). These incremental innovations reflect the continued dynamism and ingenuity of the equipment manufacturers. These efforts by various stakeholders facilitate improvements for existing applications as well as exploring new opportunities.

This review will provide a concise overview of the latest developments and applications involving HPH with the requisite contemporary explanation of the HPH process and its uses. The update by the authors will therefore mainly focus on publications of the last one to two decades. It should be noted that in many of the applications mentioned, HPH forms only part of the overall processing strategies. Often, published studies involved lab or pilot-scale facilities where such operations are adequate for the task. This can lead to the situation where for a given application, there might be comparatively fewer studies in the literature that specifically use HPH even though it is a viable option. In a sense, this presents an exciting opportunity as HPH is a well-established, robust and highly-scalable process option that allows for routine use and continually tested for possibilities to accelerate the development and commercialization of new processes or products.

2. Recent Applications of HPH

2.1. Food and Beverage Applications

Having its roots in the F&B sector, HPH continues to remain an integral unit operation in many well-established applications such as ensuring food safety and improving physico-chemical properties of foods such as fruit juices and dairy products (Comuzzo et al., 2014; Levy et al., 2020; Patrignani and Lanciotti, 2016; Salehi, 2020). Thus, many publications still explored areas for improvement or advancement in these more traditional F&B applications. For brevity, this review will avoid dwelling in these areas but emphasize instead on new and emerging applications.

Alongside the concerns about food sustainability and climate change, developments into alternative foods such as plant-based dairy and meats (Aschemann-Witzel et al., 2021), lab-cultured meats (Chriki and Hocquette, 2020), precision-fermented proteins (Lawton, 2021; Teng et al., 2021; Zollman Thomas and Bryant, 2021) and insect proteins (Loveday, 2019) are gaining significant consumer and commercial interest. The application of HPH in processing alternative foods is functionally identical to traditional foods as similar process objectives are desired such as improved product stability and sensory properties (Codina-Torrella et al., 2017; Levy et al., 2021), enhanced structural and physico-chemical properties (Dong et al., 2011; Song et al., 2013) and improved processability (Gul et al., 2017). The similarities in process objectives and implementation are encouraging as considerable literature and industrial know-how in processing traditional foods can be relevant in facilitating process development and scale-up of alternative foods production. Similarly, HPH has also been found to be useful in the valorization of meat scraps and waste into nutritional and functional products (Chen et al., 2020) which serves to improve sustainability for the meat industry.

HPH has also contributed to the development and production of innovative foods with improved health benefits such as functional foods, often with enhanced sensory properties and stability. Nanoemulsions, dispersions of two immiscible liquids emulsified with droplets of around 100nm (Gupta et al., 2016b), have facilitated this trend. These nanoemulsions benefit relevant food products by incorporating lipophilic ingredients as an optically transparent form which can improve product appearance and stability, protect sensitive compounds and enhance bioavailability of functional ingredients (Aswathanarayan and Vittal, 2019; Donsì et al., 2010; Yang et al., 2012). HPH has been used for preparing nanoemulsions of functional ingredients such as fatty acids and carotenoids in food products (Silva et al., 2012), flavoring and coloring agents (Aswathanarayan and Vittal, 2019), and various cannabinoids (Banerjee et al., 2021; Lewińska, 2021). An interesting recent application is the use of essential oil nanoemulsions as a natural antimicrobial additive to foods (Aswathanarayan and Vittal, 2019; Donsì and Ferrari, 2016) which can be leveraged to meet increasing consumer desire for foods with natural ingredients.

2.2. Pharmaceutical and Biotechnological Applications

Recent pharmaceutical uses of HPH have relied on established capabilities of HPH related to comminution by high shear, in novel applications and improving existing products or processes. Broadly, pharmaceutical usage of HPH can be divided into two categories: excipients, particularly polymeric types, processing and active pharmaceutical ingredients (APIs), especially as nanoparticulate systems, for enhanced drug delivery. More recently, HPH uses in the biotechnology and biopharmaceutical industries are also more common.

2.2.1. Processing Excipients

Polymers and gums such as alginates and cellulosic derivatives are common pharmaceutical excipients used for drug delivery systems. These polymeric systems are widely used as thickeners or stabilizers in vehicle liquid for drug delivery and tissue engineering (Curvello et al., 2019), suspensions (Lin and Dufresne, 2014) or transdermal products (Raghav et al., 2021). HPH is often used in the production and/or modification of these polymers. In oral solid dosage forms, polymeric excipients are also used as fillers, binders and disintegrants among others (Debotton and Dahan, 2017).

HPH is often applied in the modification of polymers and gums as it can alter the characteristics of polymers by the intense high shear and turbulence. Examples of such modifications include changes to rheological properties (Eren et al., 2015; Porto et al., 2015), viscosity reduction (Harte and Venegas, 2010; Inguva et al., 2015), zeta potential (Prateepchanachai et al., 2017), molecular weight and hydrodynamic radius reduction (Belmiro et al., 2018). These modifications can improve processability and enable new formulation strategies such as incorporating excessively viscous polymer gels into formulations (Inguva et al., 2015) or enhancing product quality by improved film strength and water resistance (Shahbazi et al., 2018). However, the HPH process is also recognized to have the potential to adversely impact certain excipients such as protein emulsifiers (Ali et al., 2018) and such a possibility should be considered.

2.2.2. Processing of APIs

Size reduction capabilities of HPH on APIs confer several advantageous properties to the final drug product. A particular application is in the formation of nanocrystals and

nanosuspensions of APIs. By preparing the API as a nanosuspension, the dissolubility is increased which improves the bioavailability of poorly water-insoluble drugs. By improving drug solubility, commercialization of drugs would be possible as previously, they were deemed unviable due to poor solubility (Rabinow, 2004). While this paradigm of thought is not new, recent literature reports focused on developing a better understanding between the product and process such as the impact of formulation choices (Paredes et al., 2016; Sun et al., 2011; Van Eerdenbrugh et al., 2008), HPH operating conditions (Kluge et al., 2012; Oktay et al., 2019), and process type and configuration e.g. HPH vs other techniques for producing nanoparticulates (Kakran et al., 2012; Y. Li et al., 2015; Salazar et al., 2012; Zhou et al., 2018).

Another noteworthy HPH application for the processing of APIs is in the formation of nanoemulsions. The hydrophobic drug is dissolved in an oil phase which is then emulsified as an oil-in-water (O/W) nanoemulsion for use in a variety of drug delivery systems such as sprays, creams and capsules (L. Chen et al., 2020; Singh et al., 2017). The benefits of pharmaceutical nanoemulsions include improved bioavailability, stability and for controlled release (Gué et al., 2016; Singh et al., 2017). Examples of pharmaceutical nanoemulsions processed using HPH include nanoemulsions of various APIs (Kotta et al., 2015; Sharma et al., 2015; Tagne et al., 2008; Tran et al., 2017), DNA/nanoemulsion complexes for gene therapy (Schuh et al., 2018) and vaccines (Fox et al., 2013; Haensler, 2017; Peshkovsky and Bystryak, 2014).

Outside the pharmaceutical sector, recent studies exploring various aspects of the nanoemulsification process have yielded better process and product understanding by optimizing emulsifier type and use concentration (Qian and McClements, 2011; Silva et

al., 2015; Uluata et al., 2016), HPH operating conditions and equipment design (Donsi et al., 2012; Silva et al., 2015; Wan et al., 2019), process type and configuration (Calligaris et al., 2018, 2016) and the development of semi-empirical scaling laws (Gupta et al., 2016a). Many insights from these studies provided relevance of HPH use to prepare nanoemulsions for both the pharmaceutical and allied industries.

2.2.3. Nanoparticle Systems

Nanoparticulate systems in the pharmaceutical industry form an important platform for achieving a range of clinical and therapeutic objectives to introduce improved and/or targeted drug delivery or efficacious and safer imaging agents (Bobo et al., 2016). This review avoids the discussion of the various types of nanoparticulate systems and their different therapeutic and clinical functions as there are already some excellent reviews (Bhatia, 2016; Bobo et al., 2016; Mitchell et al., 2021). In summary, HPH has emerged as a viable mechanism for the processing of three classes of nanoparticulate systems: lipid-based nanoparticles (Pardeike et al., 2009), polymeric nanoparticles (Vinchhi et al., 2021), and inorganic/hybrid-type nanoparticles (Grüttner et al., 2007; Qi et al., 2014).

The involvement of HPH in processing therapeutically relevant nanoparticles has grown incrementally. Most literature reports focused on developing nanoparticulate products and formulations for different APIs (Vinchhi et al., 2021; Soni et al., 2020), using lipid-based and polymeric nanoparticles gene therapy (del Pozo-Rodríguez et al., 2016)), molecular targeting agents (Grüttner et al., 2007; Natarajan et al., 2008), and imaging agents (Lee et al., 2004; Yang et al., 2019). The HPH process has been explored within the context of nanoparticle production in order to develop improved process insights for

scale-up, process control, and optimization (Homayouni et al., 2014; Hu et al., 2016; Lomis et al., 2016; Qi et al., 2014; Soni et al., 2020).

2.2.4. Biopharmaceutical and Biotechnology Applications

Most recent biopharmaceutical and biotechnology applications involving HPH have relied on its well-established cell disruption/lysis capability. The applications include the production of cell-free protein synthesis systems using various cell types (Carlson et al., 2012; Failmezger et al., 2018; L. Zhang et al., 2020), downstream processing for recombinant protein and virus-like particle production, by batch or continuous operation (Effio and Hubbuch, 2015; Tam et al., 2012; Wilken and Nikolov, 2012) along with extraction of oils and other products from algae (Günerken et al., 2015; Samarasinghe et al., 2012; Spiden et al., 2013).

Further analysis of process and product specific nuances and avenues for improvement such as the interplay between upstream and downstream unit operations on product quality (Eggenreich et al., 2020; Hutterer et al., 2013; Slouka et al., 2018), improved cell disruption process monitoring strategies (Eggenreich et al., 2017), energy density of algal cell disruption (Yap et al., 2015), and strategies for process optimization (Dopp and Reuel, 2018; Metzger et al., 2020; Pekarsky et al., 2019).

2.3. Advanced Materials and Chemical Applications

2.3.1. Advanced Polymer Production and Processing

Natural biopolymers are again being explored as alternative and sustainable materials for many uses such as synthetic polymer replacements (Vinod et al., 2020), therapeutic applications, and electronic materials (de Amorim et al., 2020). HPH, sometimes with

other techniques like enzymatic hydrolysis, has been successfully applied to the production of biopolymers such as nanocellulose (Kawee et al., 2018; Lenhart et al., 2020; Li et al., 2012; Wang et al., 2019), starch nanoparticles (Ahmad et al., 2020; Apostolidis and Mandala, 2020; Shi et al., 2011; Wang et al., 2021), chitin nanofibers (Mushi et al., 2019; Ono et al., 2020; Salaberria et al., 2015; Satam and Meredith, 2021), and silk nanofibers (Uddin et al., 2020).

Often, native biopolymers do not possess the requisite physico-chemical properties for subsequent processing or for the final product. Structural modifications by chemical treatment to alter rheological properties or adding modifiers like plasticizers and similar materials to form composites may be necessary (Aaliya et al., 2021; George et al., 2020; Harish Prashanth and Tharanathan, 2007; Kaur et al., 2012). In most cases, HPH is used primarily for the production of the material of interest and subsequent processing steps may utilize other techniques (Rizal et al., 2021; Rocca-Smith et al., 2019). In certain cases, HPH was also used in subsequent modification and processing steps such as in the modification of starch nanoparticles (Apostolidis and Mandala, 2020; Shahbazi et al., 2018), rheological and structural modifications of polymer dispersions (Fu et al., 2011), and formation of nanofibrillated cellulose composites with nanoclays (Garusinghe et al., 2018; Shanmugam et al., 2021; Yong et al., 2018).

Nanocellulose deserves particular attention as a cellulosic material of nanoscale dimension and with three main forms in the literature, nanocrystalline cellulose (NCC), bacterial nanocellulose (BNC), and nanofibrillated cellulose (NFC) (Phanthong et al., 2018; Thomas et al., 2020). The different types of nanocellulose have attracted significant interest from academia and industry due to the many advantageous properties, being a

biodegradable natural product of high strength and stiffness along with the possibility of surface chemical modifications, among many others (Phanthong et al., 2018; Salas et al., 2014). Nanocellulose has been utilized in many sectors such as foods (Gómez H. et al., 2016), composite materials and packaging (Kargarzadeh et al., 2017; F. Li et al., 2015), electronics (Sabo et al., 2016), biomedical and pharmaceuticals (Jorfi and Foster, 2015; Kamel et al., 2020), and environmental remediation (Mahfoudhi and Boufi, 2017). As previously mentioned, HPH has been established as the convenient, scalable and comparatively environmentally-friendly technology to process and produce nanocellulose and its derivatives.

HPH has also found application in the broader sustainability trend of upcycling and valorizing agri-food remains or waste into usable materials like bioplastics (Otoni et al., 2021). Examples of HPH use in this context include the production of nanocellulose and other nanofibers from agricultural wastes and other sources (Hongrattanaichit and Aht-Ong, 2020; Li et al., 2012; Pacaphol and Aht-Ong, 2017; Salaberria et al., 2015), bio-based adhesives from soybean meal (Y. Zhang et al., 2020), and biopolymer films from fruit and vegetable wastes (Kang and Min, 2010; Wu et al., 2020).

2.3.2. Nanoscale Materials and Fluids

HPH has been applied to the scalable production of many exciting nanomaterials through liquid-phase exfoliation such as boron nitride nanosheets (Guerra et al., 2018; Shang et al., 2016), transition metal dichalcogenide nanosheets (Piao et al., 2018; Shang et al., 2016), and graphene and its variants (Arao et al., 2016; Chen et al., 2020; Qi et al., 2017; Shang et al., 2015). Subsequent application of these nanomaterials often requires the dispersion of the nanostructured material in some media and in some cases, the

dispersion produced from the liquid-phase exfoliation step can be used directly (Backes et al., 2020; Johnson et al., 2015). Correspondingly, HPH is suitable for use to directly produce a wide variety of dispersions and has been demonstrated as an effective technique for dispersing and potentially modifying nanomaterials (Azoubel and Magdassi, 2010; Li et al., 2019; Schlüter et al., 2014; Tölle et al., 2012). Many studies have reported successful use of HPH as a processing step to produce advanced materials such as graphene films and inks (Tölle et al., 2012), polymer composites incorporating nanomaterials for improved mechanical, thermal and electrical properties (Appel et al., 2012; Chatterjee et al., 2012; Clausi et al., 2020; Shayganpour et al., 2019; Wu et al., 2021; Xu et al., 2020), and nanofluids which will be discussed.

Nanofluids contain various types of nanoparticles like metal nanoparticles and their oxides or carbon nanotubes/graphite in a base fluid have emerged as a promising advanced thermal fluid to improve the heat transfer performance and thermal efficiency in systems for engine cooling or building temperature management (Saidur et al., 2011). HPH has successfully been used to produce nanofluids containing carbon nanoparticles like carbon nanotubes, diamond, graphite, or graphene (Fontes et al., 2015; Oliveira et al., 2017; Sica et al., 2021) and metal oxides (Fedele et al., 2011). Similarly, homogenization has been applied for the production of phase change material emulsions which are currently being researched for use as advanced thermal fluids and energy storage media (Wang et al., 2019).

3. Advances in Process Understanding

3.1. Mechanistic Modelling of HPH

The HPH process involves complex fluid mechanics (primarily shear, turbulence, cavitation, and impact) which can be engineered to obtain desired process and product outcomes. With improvements in computational and experimental resources available to researchers, extensive computational fluid dynamics (CFD) simulations and experiments have been performed to elucidate theoretical features of fluid flow in the homogenizer, to better understand how HPH operation relates to product characteristics and seek avenues for process improvement. Despite these advances, it is often helpful to first consider the governing physical laws and theory prior to numerical simulations / experiments. Meaningful insights in the form of scaling relationships and qualitative understanding of flow features relevant to the process of interest can be obtained with significantly less effort (Håkansson, 2019; Jahnke, 1998; Vinchhi et al., 2021; Yadav and Kale, 2020).

Applying CFD to study HPH is complicated as there are multiple complex fluid processes that occur within the homogenization valve and in many applications, often two or more phases such as solid-liquid, liquid-liquid and/or liquid-gas phases are involved and needed to be accounted for (Jahnke, 1998). Due to these complexities, direct numerical simulation (DNS) of the governing fluid mechanics equations e.g., Navier-Stokes equation for Newtonian flows, are often intractable for real operating conditions. As such, researchers have often employed the Reynolds averaged Navier-Stokes (RANS) model with different turbulence models or large eddy simulations. Håkansson et al., (2012) provide an excellent overview of the various CFD studies on homogenization valves using the RANS model by different groups. In that study, it was reported that while RANS with some variants of the $k - \varepsilon$ turbulence model can reasonably model the inlet section of the

valve but failed to adequately describe flow in the outlet chamber (post constriction) when compared with experimental data. In an effort to improve the accuracy of CFD simulations, recent studies have employed large eddy simulations to study flow in homogenizer valves with much greater success (Taghinia et al., 2016, 2015).

CFD has also been applied to study specific phenomena of interest such as droplet breakage and emulsion formation or the impact of equipment design and operation both by academia (Håkansson, 2022, 2018) and industry (see Section 4). The emulsification process in homogenizers has been modelled using the governing equations of fluid dynamics that are typically coupled with the dynamics of the dispersed phase using a population balance model (Becker et al., 2014; Dubbelboer et al., 2014; Håkansson et al., 2013, 2009; Inguva et al., 2022; Raikar et al., 2010) or a discrete phase model (Casoli et al., 2010; Zahari et al., 2018). These models are able to track the evolution of the droplet size distribution by accounting for effects such as breakage, coalescence (Maindarkar et al., 2012) and adsorption of an emulsifier (Håkansson et al., 2013). These models can be implemented in commercially available software such as ANSYS or in open-source codes such as OpenFOAM (da Rosa and Braatz, 2018; Passalacqua et al., 2018). The implementation of these models conventionally requires a parameter estimation step which reduces the general applicability of these models to some extent. It is possible to employ multiphase flow CFD techniques such as the volume-of-fluid interface capturing method (Malekzadeh and Roohi, 2015; Mukherjee et al., 2018) or smooth particle hydrodynamics (Wieth et al., 2016) to model droplet and emulsion formation. This approach is far more predictive as only a handful of physical constants such as the interfacial tension and viscosities are required to formulate the model but it

can be significantly more computationally expensive if not currently intractable, considering the highly turbulent flow present in HPH valves.

As a final point, many newer applications of HPH involve molecular length scales like the production of nanofibrillated cellulose (Abdul Khalil et al., 2014) or graphene-based materials (Shang et al., 2015). While CFD and other continuum-scale techniques might be able to provide some insights into the process (Stafford et al., 2021), it may not be adequate to provide a complete picture of the process. At these length scales, mesoscale and molecular-scale methods such as dissipative particle dynamics (J. Zhang et al., 2020) and molecular dynamics (Choi et al., 2014; Li et al., 2017) can be used in conjunction with continuum-scale methods like CFD to study HPH process in a multiscale manner. However, despite the existence of several mesoscale/molecular-scale studies for specific phenomena / system such as emulsions and colloids, there is a paucity of literature for multiscale modelling and simulations of HPH processes and this presents an exciting opportunity for new research.

3.2. Data-Driven / Empirical Modelling

In many cases, it may be impractical or even intractable to develop an adequately detailed mechanistic model for HPH processes. Thus, empirical/ semi-empirical and data-driven models that relate input and output variables are often modelled for product development and optimization (Garcia-Ortega et al., 2015). These models are also very useful for developing digital twins and process simulations for the whole process where HPH is an important unit operation. The diversity of HPH applications does mean that there is no universal model to describe the relationship between key input and output variables as the specific application determines important variables and factors.

Models attempted involve variables related to particle size distribution for emulsification or nanoparticle production processes (Dubbelboer et al., 2014; Durán-Lobato et al., 2013; Lebaz and Sheibat-Othman, 2019), viscosity reduction of polymers (Harte and Venegas, 2010; Inguva et al., 2015), cell disruption for intracellular product recovery and sludge disintegration (Ekpeni et al., 2015; Kelly and Muske, 2004; Tam et al., 2012; Y. Zhang et al., 2012), and nanocellulose production (Davoudpour et al., 2015). Two modelling strategies are of particular interest; response surface model (RSM) which employs a multi-dimensional first order or second order polynomial to curve-fit experimental data conducted as part of a design of experiment (Khuri and Mukhopadhyay, 2010). The first approach is conceptually straightforward and can be implemented easily as it is already commonly used to model HPH processes (Davoudpour et al., 2015; Gul et al., 2018). The second approach is machine learning based approaches e.g., neural network / gaussian processes / k-nearest neighbors that are being applied in HPH settings (Bhilare et al., 2019; Patil et al., 2016). Machine learning approaches offer the potential for better regression and predictive performance than RSM in addition to facilitating the construction of more complex models that integrate more data-streams.

3.3. Process Analytical Technology

While process analytical technology (PAT) as a concept has mainly gained prominence in the pharmaceutical and fine chemicals industries (Chew and Sharratt, 2010; Simon et al., 2015), many of the principles and techniques such as the real-time monitoring of process and product parameters and multivariate data collection and analysis methods are relevant and useful for the broader manufacturing sector (Jerome et al., 2019; van den Berg et al., 2013). The use of analytical tools for process control and monitoring for

HPH is highly application specific as each product and process will require the monitoring and evaluation of different properties of the system using different platforms and technologies. Therefore, two sub-sections will describe the topic under (a) monitoring and control of the HPH process, and (b) analysis and characterization of products. A discussion of the relevant data collection, mathematical and statistical methods will not be covered as these topics have been extensively discussed in the PAT literature.

3.3.1. Monitoring and Control of the HPH Process

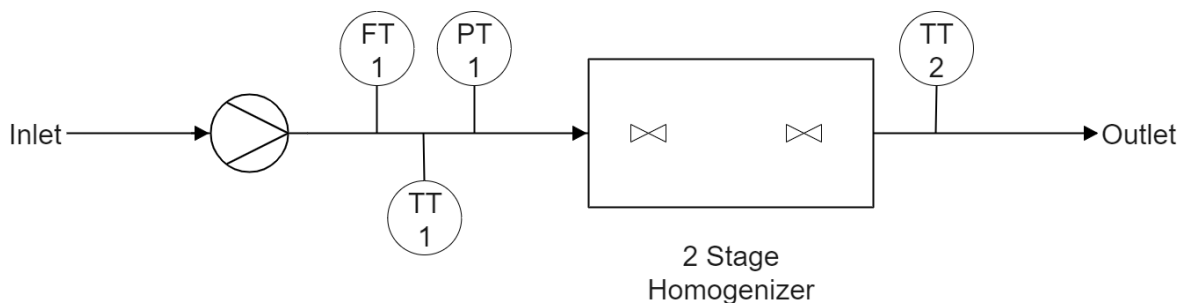


Figure 1. Schematic of 2-Stage HPH process with temperature (TT), pressure (PT) and flow (FT) sensors/transmitters.

As shown in figure 1, HPH is a conceptually simple unit operation. Various conventional sensors such as temperature, flow and pressure sensors can and are typically integrated into the equipment for standard process control and monitoring. For a given process configuration such as a single 2-stage homogenizer, the main manipulated variables are the homogenization pressure of the different stages and number of passes. The pressure can be automatically controlled with the use of a pressure controller such as in GEA XStream Lab homogenizer as an example. Another parameter that can be easily

monitored for additional process insight is the power consumption of the homogenizer. While not as extensively discussed in the HPH literature, power consumption measurement is a common PAT tool in many unit operations such as granulation (Hansuld and Briens, 2014) and mixing (Bowler et al., 2020) to characterize different stages of the process and determine the process end-point. While such analysis may not be directly applicable to the continuous nature of HPH operations, power measurement can help to monitor the process for deviations. The HPH equipment may be monitored by its acoustic emission for improved process insights. The use of acoustic measurement to better understand the occurrence of cavitation during operation and its impact on the emulsification process has been expounded (Håkansson et al., 2010; Schlender et al., 2015b). While not explored thus far, possible incorporation of acoustic emissions and other monitored variables as output variables in a multivariate data-driven model of the process which can then be leveraged for enhanced process monitoring, control, and optimization.

3.3.2. Analysis and Characterization of Products

The trend to further develop and incorporate PAT for better product and process understanding has facilitated significant technological advancements in ancillary areas such as new or improved sensor technologies for inline process integration and multivariate data analytic methods to handle complex sensor data streams (Grassi and Alamprese, 2018; Rüdts et al., 2017; van den Berg et al., 2013; Zhao et al., 2015). Recently, several noteworthy studies explored new technologies and the integration of PAT for process monitoring and control (Besseling et al., 2019; Eggenreich et al., 2017; Ralbovsky et al., 2022). Table 1 summarizes the various PAT tools employed for HPH.

Considering the diversity and increasing sophistication of HPH applications that require precise control, additional work by all stakeholders is necessary to holistically fuel the exciting challenge of advancing the development and incorporation of PAT in HPH processes.

Table 1. Summary of process/product variables of interest and analytical technologies available for PAT. Where possible, examples provided are applied within the context of HPH, but some examples are drawn from other areas such as chemical synthesis.

Process/Product Variable of Interest	Available Analytical Technologies (Process Integration Format)	Comments	Examples
Temperature	Thermocouple (inline)	Sensors can be integrated at both the inlet and outlet to monitor temperature change.	(Martínez-Monteagudo et al., 2017; Poliseli-Scopel et al., 2013)
Pressure	Pressure transducer (inline)	Pressure gauges typically come standard with the HPH equipment.	
Flowrate	Flowmeter (inline)	Various flowmeters with different operating principles are available. Select a flowmeter type best suited for the application.	(Poliseli-Scopel et al., 2013; Schlender et al., 2015a)
Acoustic emissions	Microphone / Acoustics emission sensor	Signal processing such as amplification may be necessary. Currently used to characterize cavitation patterns.	(Håkansson et al., 2010; Schlender et al., 2015b)
Power consumption	Power meter		See various citations in Hansuld and Briens, (2014)
Imaging / Visualization of fluid flow	1. Particle image velocimetry (inline) 2. High-speed imaging (inline)	High-speed imaging and particle image velocimetry to characterize fluid flow features such as the velocity profile, cavitation patterns and droplet breakage.	See various citations in Bisten and Schuchmann, (2016)

Imaging of products	<ol style="list-style-type: none"> 1. Inline imaging 2. Optical / Electron / Atomic force microscopy (offline) 	<p>Inline imaging of the HPH process remains relatively unexplored with many studies using scaled models. Offline imaging using various microscopy techniques is well-established.</p>	(Ralbovsky et al., 2022)
Turbidity	<p>Turbidity sensor (inline / at-line / offline), Spectrophotometer (inline / at-line / offline)</p>		(Linke and Drusch, 2016; Spiden et al., 2013)
Particle Size	<ol style="list-style-type: none"> 1. Dynamic light scattering (DLS) (at-line / offline) 2. Spatially resolved DLS (inline) 3. Laser scattering (at-line / offline) 4. Focused beam reflectance measurement (FBRM) (inline) 		<p>DLS: (Qian and McClements, 2011; Yu et al., 2013) Spatially resolved DLS: (Besseling et al., 2019) Laser scattering: (Samarasinghe et al., 2012) FBRM: (Ralbovsky et al., 2022)</p>
Viscosity / Rheological properties	<ol style="list-style-type: none"> 1. Viscometer (inline / at-line / offline) 2. Rheometer (offline) 	<p>Rheometers are better at characterizing rheological properties than viscometers; choice depends on process / product requirements.</p>	<p>Viscometer: (Inguva et al., 2015; Pu et al., 2015) Rheometer: (Fu et al., 2011; Qian and McClements, 2011)</p>

Chemical properties / Composition	<ol style="list-style-type: none"> 1. X-ray diffraction (XRD) (offline) 2. Spectroscopic techniques (Raman, UV-Vis, NIR etc.) (inline / at-line / offline) 3. Chromatographic techniques (HPLC, size-exclusion etc.) (at-line / offline) 4. Nuclear magnetic resonance (NMR) (online / at-line / offline) 	The choice of analytical technique and integration format depends on the application and process requirements. Cost can also be an important factor.	<p>XRD: (Apostolidis and Mandala, 2020; Li et al., 2012)</p> <p>Spectroscopic: (Ralbovsky et al., 2022; Shang et al., 2016, 2015)</p> <p>Chromatographic: (Eggenreich et al., 2017; Salaberria et al., 2015)</p> <p>NMR: (T. Li et al., 2015; Ono et al., 2020)</p>
-----------------------------------	---	--	---

4. Industrial Insights and Perspectives

4.1. Industrial Outlook

The HPH industry is pushed by innovation and technological advances, pulled by the need for scale-up of successful “startup” products, and inspired by an increasing focus on sustainability at lowest possible total cost of ownership. While HPH will remain integral to multiple traditional applications, several high growth areas in the HPH market have recently emerged and include complex foodstuff products, pharmaceutical/nutraceuticals, and new industrial products related to the transition to a more sustainable and circular economy. In this regard, many of the active areas of research involving homogenizers are closely aligned with industrial trends.

Developments in traditional, high-volume, and low-complexity applications (e.g., fresh milk) are ongoing and will continue to be an important driver to meet demands for increased throughput at lower homogenization pressures while yielding the same or better homogenization effect. Simultaneously, major global dairy and food players are investing in differentiating some of their products by adding supplements like vitamins and fibers to turn them into healthier and more profitable products. These higher-end products typically require higher homogenization pressures. Key developments to take note of in this space include improved product and process understanding and more efficient homogenizer valves designed with the latest CFD tools.

Some of the emerging applications (e.g., certain novel biotech applications, new foods, battery materials, and NFC/PHA) will either fizzle out while others will grow and mature into main-stream applications with large capacity, 24/7 production plants on a global

scale. Consequently, HPH machine designs and materials will evolve to better suit these emerging applications in an industrial setting.

HPH manufacturers are actively developing designs and solutions to better meet customers' expectations and to align processes with increasingly challenging regulatory guidance for simpler, more robust cleanability and achieving and maintaining sterility for longer time than presently possible. Improvements in materials of construction and design of wear parts in contact with the product are important to reducing maintenance frequency and costs. In-line measurements of product quality attributes, other than what is currently commonly available (e.g., temperature, flow, density, viscosity, turbidity etc.) will become viable even for lower margin products and will allow application of quality-by-design concepts in future homogenizer design and process control.

4.2. Case Studies from Industry

With widespread industrial application of HPH, the following case studies are presented to convey noteworthy points drawn from various recent industrial applications at the different stages of operation. Multiple empirical observations and “rules of thumb” are also provided as a reference to the reader. It is hoped that both industry practitioners and R&D scientists alike will find some of the information presented in this section relevant when working HPH. In some cases, specific details are omitted as this information is proprietary and/or commercially sensitive. All identifying information has also been removed.

4.2.1. Process Scale-Up

Introduction

Scale-up of HPH processes can often be very challenging as optimal operating conditions identified at a laboratory/pilot scale may not directly translate to satisfactory product quality and/or process conditions in scale-up runs. In this case study, a pharmaceutical company scaled-up (~30X the capacity of a table-top HPH) production of an API dispersion with 1~3 μm particles. The scale-up process was challenging but succeeded with inputs from both the pharmaceutical company and equipment manufacturer.

Case Study and Results

A table-top HPH system was used to identify the optimal process operating conditions for a product of the desired quality, quantified by size and size distribution determinations. As part of the scale-up procedure, the equipment manufacturer performed CFD studies and relied on prior experience to scale up the radial diffuser homogenizing valve size and profile. Initial trials conducted on the production scale equipment failed to achieve the desired particle size reduction as the particles were too large, even with the same operating pressure and number of passes. Increasing the operating pressure was not an option as the identified operating conditions were already near the limit of the machine and higher pressures increased the polydispersity of the particle size distribution. To resolve the issue, experiments were carried out on the production machine to characterize the relationship between product flowrate and particle size distribution. The CFD model was updated using the experimental data which enabled the equipment manufacturer to modify the size and shape of the homogenizing valve, yielding the desired outcome. Shortly after, an even larger production scale HPH system (~6X larger) was ordered and the scale-up was not problematical as the updated CFD model and

process insights gained from the first experience proved to remain valid at the larger scale and the first batch of product met quality specifications.

Conclusions

Scale-up, especially for complex products and mixtures, remains a highly empirical exercise, drawing upon deep process and product know-how from both the user and equipment manufacturer. The following points should be considered in scale-up:

- Modelling techniques like CFD can facilitate scale-up but remains relatively immature and needs to be used in conjunction with product and process know-how.
- Radial diffuser type homogenizer valves are much more industrially validated and successfully scaled-up when compared to other types such as fixed orifice or opposing jet valves.
- Radial diffuser type homogenizer valves are much more amenable to scale up compared to other valve types. There are many variants, with multiple adjustable design parameters (Figure 2) to suit a desired application at a given production scale

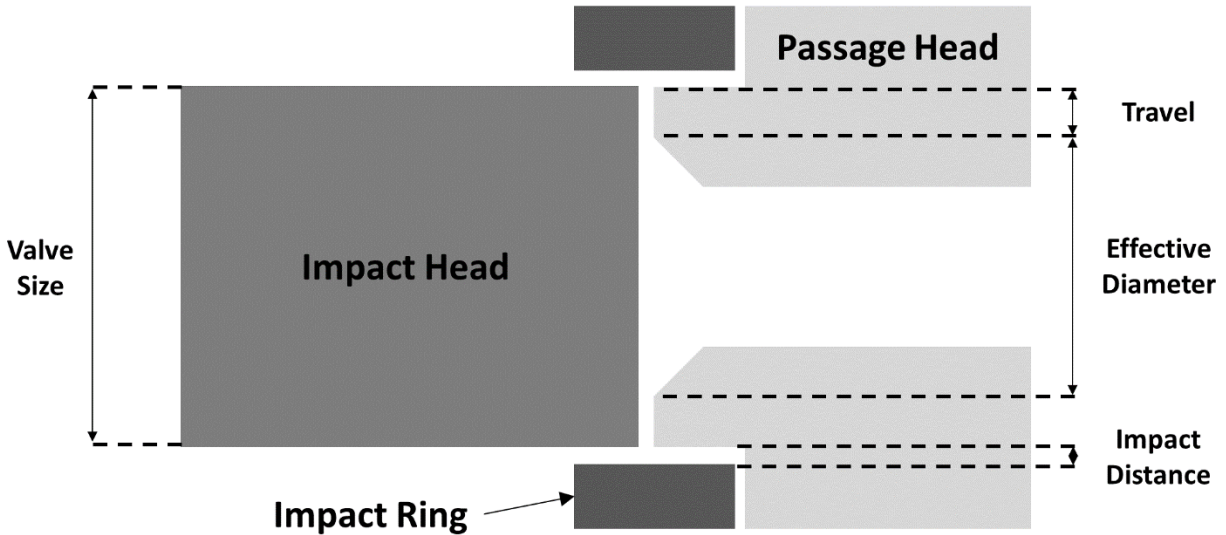


Figure 2. Schematic of a radial diffuser type homogenizer valve. The valve diameter, effective diameter, travel, and impact distance are important valve design parameters that can be adjusted to achieve the desired fluid dynamic effect for a specific application.

4.2.2. Process Optimization

Introduction

An industrial-scale process for the production of NFC was being implemented. While the process conditions identified and successfully transferred from R&D to industrial-scale, the production process was deemed commercially unviable as the operating expenses (OPEX) was excessively high. Process optimization was necessary to reduce OPEX as much as possible along with capital expense (CAPEX) to acceptable levels.

Case Study

The process originally specified a 5,000 L/hr homogenizer rated for 150 MPa. The energy consumption per pass at 150 MPa was 46 kWh/1000L of product. The cellulose raw material had natural variations and contained abrasive contaminants. The removal of these contaminants was neither necessary (the contaminants did not impact final product

quality) nor practicable (the process would then be too expensive and complicated). At these operating conditions, the machine experienced unacceptable levels of wear which resulted in significant cost for parts replacement and labor alongside production downtime.

To address this issue, multiple avenues of optimization were undertaken by both the equipment manufacturer and process development team. The equipment manufacturer carried out CFD studies to optimize the homogenizing valve geometry to enhance the desired fluid dynamic effect necessary for NFC fibrillation at gentler operating conditions. The process development team also optimized the pre-processing steps which typically consist of hydrolysis / enzymatic reduction steps.

Results

The combined efforts of the equipment manufacturer and process development team resulted in the process being able to produce NFC of acceptable quality at 60 MPa which reduced energy consumption per pass to 18.3 kWh/1000L of product. The gentler operating conditions also reduced parts wear, required labor, and frequency of maintenance by ~15X. Furthermore, as lower pressures were used, a larger machine (14,000L/hr machine rated at 70 MPa) could be supplied at lower price, thus, reducing the CAPEX per unit capacity by ~5X. These cost savings derived from optimizing both the HPH and upstream processes made the operation commercially viable and successful.

Conclusion

The following rules of thumb can be used when considering avenues for process optimization and better economies:

- At the same rated operating pressure, machine price ratio changes to the capacity ratio to the power of 0.5 or even lower: $\frac{C_2}{C_1} \sim \left(\frac{\dot{V}_2}{\dot{V}_1}\right)^{0.5}$, where C_i is the cost of machine i rated for capacity \dot{V}_i .
- At the same rated capacity, machine price ratio changes to the rated pressure ratio to the power of 1.5 or even higher: $\frac{C_2}{C_1} \sim \left(\frac{P_2}{P_1}\right)^{1.5}$, where P_i is the rated pressure of machine i .
- The power consumption of the HPH process can be estimated using the following formula: Power Consumption (kW) = $\frac{\dot{V}\left(\frac{L}{hr}\right) \times P(\text{bar})}{36000 \times \eta}$, where η is an efficiency factor and can range from ~0.50 to 0.92.
- Consider possible pre-processing steps to more efficiently utilize HPH. One common example is to use a lower-energy dissipation method (e.g., rotor-stator high shear mixer) to perform pre-dispersion/crude emulsion formation as extensively as possible prior to using a high-energy dissipation device. This strategy generally helps improve product quality and reduce OPEX and CAPEX.
- Actively consider process economics at the R&D stage to better align R&D with manufacturing.

4.2.3. Process Troubleshooting

Introduction

HPH is often used as an intermediate unit operation in the process to achieve a specific goal such as cell disruption or emulsification. Correspondingly, the final product quality in relation to HPH performance is dependent on both successful HPH operation and upstream and/or downstream operations. Two case studies are presented here which illustrate how factors upstream or downstream of the HPH process can adversely impact the quality of product and efficiency of process.

Case Study

In the first case, a manufacturer had over many years successfully produced bovine growth hormones using an *E. coli* culture process with HPH for cell disruption and achieved ~95% cell rupture efficiency in two passes. However, the cell rupture efficiency dropped to < 60% from one batch to the next which seriously affected productivity. Technicians from the HPH manufacturer first repaired and/or replaced all relevant mechanical parts and certified the machine as good as new. However, this did not improve cell rupture efficiency which remained < 60%. The manufacturer organized an audit led by a senior process specialist to further diagnose the problem and identified a process change upstream (initially assumed as non-material) that caused the drop in cell rupture efficiency.

In the second case, a high-fat, high-viscosity non-Newtonian emulsion food product was produced using HPH. The product had a comparatively lower surfactant loading without adverse impact on product stability as the high viscosity impeded coalescence of fat globules. The manufacturer experienced a recurring problem where emulsion failure occurred in the packaging and free oil would appear on top of the product in the package

which resulted in significant costs arising from recalls and waste handling. The problem was only recently addressed during an audit and fixes were recommended.

Results

In the first case study, a new storage tank (without a cooling jacket) was installed between the fermentation vessel and downstream processes (centrifugal separation and HPH cell disruption) to reduce the time taken to empty the fermentation vessel (with cooling jacket) from ~3 h to ~15 min, enabling higher fermenter utilization. As a result, the *E. coli* cells were held in the storage tank at the fermentation temperature in a low nutrient environment for up to an additional ~3 h, during which cell wall morphology significantly changed, impacting cell rupture efficiency. As a diagnostic test, the new storage tank was bypassed and cell rupture efficiency was immediately restored. The long-term solution involved installing an inline cooler to chill the cell culture to $< 10^{\circ}\text{C}$ enroute to the storage tank. With the intervention, cell rupture efficiency (a slight increase in efficiency was observed) improved and there was more efficient fermenter utilization.

During the audit in the second case study, it was determined that the issue arose downstream of the homogenizer as product samples were taken directly from the outlet of the homogenizer did not experience emulsion failure. The outlet of the homogenizer is connected to the fill-heads of the packaging machine by a jacketed pipe of about 50 m in length. The flow in the pipe is highly unsteady due to the filling process with intermittent start-stop breaks after every few seconds when switching between fill heads. However, the flow would also have frequent start-stop pauses over longer periods when there are packing line issues. The unsteady flow introduces sharp velocity gradients in the pipe, which when coupled with the comparatively lower surfactant content of the product,

resulted in coalescence of oil droplets and emulsion failure. Changes to the process to maintain a steady flow of product in the pipe and to shorten the distance between the homogenizer and the filling line are recommended.

Conclusion

Understanding the complex interactions between HPH operation, upstream/downstream processes and the product is vital for effective operation of the entire process and to ensure continuous good product quality. Correspondingly, process design, modification and troubleshooting decisions should be made with inputs from subject matter experts (e.g., process/product technologists) to ensure holistic decision making with the equipment manufacturer. Failure to do so can result in significant costs, from lost in time and output-related issues like efficiency and quality.

5. Conclusions and Outlook

Even though HPH is a mature technology platform and has become a standard unit operation in industry, many challenges either persist or are on the horizon. From a theoretical perspective, many knowledge gaps such as a detailed understanding of fluid flow in the valve remain. Emerging applications, in particular biological and nanoscale applications, contribute many open questions and exciting research and/or commercial opportunities. The push for sustainability and economies in more conventional applications will drive innovation in process and equipment design. Continued research and development by academia, equipment manufacturers, and industry is necessary to address these challenges going forward and collaborative efforts between stakeholders will become increasingly important. Advances in adjacent fields such as PAT,

computational simulations, manufacturing, and materials have been and will continue to be significant in progressing HPH technology and its use.

6. Acknowledgements

The authors would like to express thanks to Mr Domenico Gambarelli, GEA Italy, and Mr Sarma Inguva, GEA Singapore, for sharing their time and invaluable expertise on the industrial applications of high-pressure homogenization. Thanks also go to Dr Frando Van der Pas, InProcess-LSP, The Netherlands, for sharing about the NanoFlowSizer PAT platform.

References

- Aaliya, B., Sunooj, K.V., Lackner, M., 2021. Biopolymer composites: a review. *Int. J. Biobased Plast.* 3, 40–84. <https://doi.org/10.1080/24759651.2021.1881214>
- Abdul Khalil, H.P.S., Davoudpour, Y., Islam, M.N., Mustapha, A., Sudesh, K., Dungani, R., Jawaid, M., 2014. Production and modification of nanofibrillated cellulose using various mechanical processes: A review. *Carbohydr. Polym.* 99, 649–665. <https://doi.org/10.1016/j.carbpol.2013.08.069>
- Ahmad, A.N., Lim, S.A., Navaranjan, N., Hsu, Y.-I., Uyama, H., 2020. Green sago starch nanoparticles as reinforcing material for green composites. *Polymer (Guildf)*. 202, 122646. <https://doi.org/10.1016/j.polymer.2020.122646>
- Ali, A., Le Potier, I., Huang, N., Rosilio, V., Cheron, M., Faivre, V., Turbica, I., Agnely, F., Mekhloufi, G., 2018. Effect of high pressure homogenization on the structure and the interfacial and emulsifying properties of β -lactoglobulin. *Int. J. Pharm.* 537, 111–121. <https://doi.org/10.1016/j.ijpharm.2017.12.019>
- Apostolidis, E., Mandala, I., 2020. Modification of resistant starch nanoparticles using high-pressure homogenization treatment. *Food Hydrocoll.* 103, 105677. <https://doi.org/10.1016/j.foodhyd.2020.105677>
- Appel, A.-K., Thomann, R., Mülhaupt, R., 2012. Polyurethane nanocomposites prepared from solvent-free stable dispersions of functionalized graphene nanosheets in polyols. *Polymer (Guildf)*. 53, 4931–4939. <https://doi.org/10.1016/j.polymer.2012.09.016>

Arao, Y., Mizuno, Y., Araki, K., Kubouchi, M., 2016. Mass production of high-aspect-ratio few-layer-graphene by high-speed laminar flow. *Carbon N. Y.* 102, 330–338. <https://doi.org/10.1016/j.carbon.2016.02.046>

Aschemann-Witzel, J., Gantriis, R.F., Fraga, P., Perez-Cueto, F.J.A., 2021. Plant-based food and protein trend from a business perspective: markets, consumers, and the challenges and opportunities in the future. *Crit. Rev. Food Sci. Nutr.* 61, 3119–3128. <https://doi.org/10.1080/10408398.2020.1793730>

Aswathanarayan, J.B., Vittal, R.R., 2019. Nanoemulsions and Their Potential Applications in Food Industry. *Front. Sustain. Food Syst.* 3. <https://doi.org/10.3389/fsufs.2019.00095>

Azoubel, S., Magdassi, S., 2010. The formation of carbon nanotube dispersions by high pressure homogenization and their rapid characterization by analytical centrifuge. *Carbon N. Y.* 48, 3346–3352. <https://doi.org/10.1016/j.carbon.2010.05.024>

Backes, C., Abdelkader, A.M., Alonso, C., Andrieux-Ledier, A., Arenal, R., Azpeitia, J., Balakrishnan, N., Banszerus, L., Barjon, J., Bartali, R., Bellani, S., Berger, C., Berger, R., Ortega, M.M.B., Bernard, C., Beton, P.H., Beyer, A., Bianco, A., Bøggild, P., Bonaccorso, F., Barin, G.B., Botas, C., Bueno, R.A., Carriazo, D., Castellanos-Gomez, A., Christian, M., Ciesielski, A., Ciuk, T., Cole, M.T., Coleman, J., Coletti, C., Crema, L., Cun, H., Dasler, D., De Fazio, D., Díez, N., Drieschner, S., Duesberg, G.S., Fasel, R., Feng, X., Fina, A., Forti, S., Galiotis, C., Garberoglio, G., García, J.M., Garrido, J.A., Gibertini, M., Götzhäuser, A., Gómez, J., Greber, T., Hauke, F., Hemmi, A., Hernandez-Rodriguez, I., Hirsch, A., Hodge, S.A., Huttel, Y.,

Jepsen, P.U., Jimenez, I., Kaiser, U., Kaplas, T., Kim, H., Kis, A., Papagelis, K., Kostarelos, K., Krajewska, A., Lee, K., Li, C., Lipsanen, H., Liscio, A., Lohe, M.R., Loiseau, A., Lombardi, L., Francisca López, M., Martin, O., Martín, C., Martínez, L., Martin-Gago, J.A., Ignacio Martínez, J., Marzari, N., Mayoral, Á., McManus, J., Melucci, M., Méndez, J., Merino, C., Merino, P., Meyer, A.P., Miniussi, E., Miseikis, V., Mishra, N., Morandi, V., Munuera, C., Muñoz, R., Nolan, H., Ortolani, L., Ott, A.K., Palacio, I., Palermo, V., Parthenios, J., Pasternak, I., Patane, A., Prato, M., Prevost, H., Prudkovskiy, V., Pugno, N., Rojo, T., Rossi, A., Ruffieux, P., Samorì, P., Schué, L., Setijadi, E., Seyller, T., Speranza, G., Stampfer, C., Stenger, I., Strupinski, W., Svirko, Y., Taioli, S., Teo, K.B.K., Testi, M., Tomarchio, F., Tortello, M., Treossi, E., Turchanin, A., Vazquez, E., Villaro, E., Whelan, P.R., Xia, Z., Yakimova, R., Yang, S., Yazdi, G.R., Yim, C., Yoon, D., Zhang, X., Zhuang, X., Colombo, L., Ferrari, A.C., Garcia-Hernandez, M., 2020. Production and processing of graphene and related materials. *2D Mater.* 7, 022001.

<https://doi.org/10.1088/2053-1583/ab1e0a>

Banerjee, A., Binder, J., Salama, R., Trant, J.F., 2021. Synthesis, characterization and stress-testing of a robust quillaja saponin stabilized oil-in-water phytocannabinoid nanoemulsion. *J. Cannabis Res.* 3, 43. <https://doi.org/10.1186/s42238-021-00094-w>

Becker, P.J., Puel, F., Dubbelboer, A., Janssen, J., Sheibat-Othman, N., 2014. Coupled population balance–CFD simulation of droplet breakup in a high pressure homogenizer. *Comput. Chem. Eng.* 68, 140–150.
<https://doi.org/10.1016/j.compchemeng.2014.05.014>

- Belmiro, R.H., Tribst, A.A.L., Cristianini, M., 2018. Application of high-pressure homogenization on gums. *J. Sci. Food Agric.* 98, 2060–2069.
<https://doi.org/10.1002/jsfa.8695>
- Besseling, R., Damen, M., Wijgergangs, J., Hermes, M., Wynia, G., Gerich, A., 2019. New unique PAT method and instrument for real-time inline size characterization of concentrated, flowing nanosuspensions. *Eur. J. Pharm. Sci.* 133, 205–213.
<https://doi.org/10.1016/j.ejps.2019.03.024>
- Bhatia, S., 2016. Nanoparticles Types, Classification, Characterization, Fabrication Methods and Drug Delivery Applications, in: *Natural Polymer Drug Delivery Systems*. Springer International Publishing, Cham, pp. 33–93.
https://doi.org/10.1007/978-3-319-41129-3_2
- Bhilare, K.D., Patil, M.D., Tangadpalliwar, S., Shinde, A., Garg, P., Banerjee, U.C., 2019. Machine learning modelling for the ultrasonication-mediated disruption of recombinant *E. coli* for the efficient release of nitrilase. *Ultrasonics* 98, 72–81.
<https://doi.org/10.1016/j.ultras.2019.06.006>
- Bisten, A., Schuchmann, H., 2016. Optical Measuring Methods for the Investigation of High-Pressure Homogenisation. *Processes* 4, 41.
<https://doi.org/10.3390/pr4040041>
- Bobo, D., Robinson, K.J., Islam, J., Thurecht, K.J., Corrie, S.R., 2016. Nanoparticle-Based Medicines: A Review of FDA-Approved Materials and Clinical Trials to Date. *Pharm. Res.* 33, 2373–2387. <https://doi.org/10.1007/s11095-016-1958-5>
- Bowler, A.L., Bakalis, S., Watson, N.J., 2020. A review of in-line and on-line

measurement techniques to monitor industrial mixing processes. *Chem. Eng. Res. Des.* 153, 463–495. <https://doi.org/10.1016/j.cherd.2019.10.045>

Calligaris, S., Plazzotta, S., Bot, F., Grasselli, S., Malchiodi, A., Anese, M., 2016. Nanoemulsion preparation by combining high pressure homogenization and high power ultrasound at low energy densities. *Food Res. Int.* 83, 25–30. <https://doi.org/10.1016/j.foodres.2016.01.033>

Calligaris, S., Plazzotta, S., Valoppi, F., Anese, M., 2018. Combined high-power ultrasound and high-pressure homogenization nanoemulsification: The effect of energy density, oil content and emulsifier type and content. *Food Res. Int.* 107, 700–707. <https://doi.org/10.1016/j.foodres.2018.03.017>

Carlson, E.D., Gan, R., Hodgman, C.E., Jewett, M.C., 2012. Cell-free protein synthesis: Applications come of age. *Biotechnol. Adv.* 30, 1185–1194. <https://doi.org/10.1016/j.biotechadv.2011.09.016>

Casoli, P., Vacca, A., Berta, G.L., 2010. A numerical procedure for predicting the performance of high pressure homogenizing valves. *Simul. Model. Pract. Theory* 18, 125–138. <https://doi.org/10.1016/j.simpat.2009.09.014>

Chatterjee, S., Wang, J.W., Kuo, W.S., Tai, N.H., Salzmann, C., Li, W.L., Hollertz, R., Nüesch, F.A., Chu, B.T.T., 2012. Mechanical reinforcement and thermal conductivity in expanded graphene nanoplatelets reinforced epoxy composites. *Chem. Phys. Lett.* 531, 6–10. <https://doi.org/10.1016/j.cplett.2012.02.006>

Chen, L., Cheng, L., Doyle, P.S., 2020. Nanoemulsion-Loaded Capsules for Controlled Delivery of Lipophilic Active Ingredients. *Adv. Sci.* 7, 2001677.

<https://doi.org/10.1002/advs.202001677>

Chen, S., Wang, Q., Zhang, M., Huang, R., Huang, Y., Tang, J., Liu, J., 2020. Scalable production of thick graphene film for next generation thermal management application. *Carbon N. Y.* 167, 270–277.

<https://doi.org/10.1016/j.carbon.2020.06.030>

Chen, X., Liang, L., Xu, X., 2020. Advances in converting of meat protein into functional ingredient via engineering modification of high pressure homogenization. *Trends Food Sci. Technol.* 106, 12–29. <https://doi.org/10.1016/j.tifs.2020.09.032>

Chew, W., Sharratt, P., 2010. Trends in process analytical technology. *Anal. Methods* 2, 1412. <https://doi.org/10.1039/c0ay00257g>

Choi, S. Bin, Yoon, H.M., Lee, J.S., 2014. Multi-scale approach for the rheological characteristics of emulsions using molecular dynamics and lattice Boltzmann method. *Biomicrofluidics* 8, 052104. <https://doi.org/10.1063/1.4892977>

Chriki, S., Hocquette, J.-F., 2020. The Myth of Cultured Meat: A Review. *Front. Nutr.* 7. <https://doi.org/10.3389/fnut.2020.00007>

Clausi, M., Grasselli, S., Malchiodi, A., Bayer, I.S., 2020. Thermally conductive PVDF-graphene nanoplatelet (GnP) coatings. *Appl. Surf. Sci.* 529, 147070.

<https://doi.org/10.1016/j.apsusc.2020.147070>

Codina-Torrella, I., Guamis, B., Ferragut, V., Trujillo, A.J., 2017. Potential application of ultra-high pressure homogenization in the physico-chemical stabilization of tiger nuts' milk beverage. *Innov. Food Sci. Emerg. Technol.* 40, 42–51.

<https://doi.org/10.1016/j.ifset.2016.06.023>

Comuzzo, P., Calligaris, S., 2019. Potential Applications of High Pressure

Homogenization in Winemaking: A Review. *Beverages* 5, 56.

<https://doi.org/10.3390/beverages5030056>

Curvello, R., Raghuwanshi, V.S., Garnier, G., 2019. Engineering nanocellulose

hydrogels for biomedical applications. *Adv. Colloid Interface Sci.* 267, 47–61.

<https://doi.org/10.1016/j.cis.2019.03.002>

da Rosa, C.A., Braatz, R.D., 2018. openCrys: Open-Source Software for the Multiscale

Modeling of Combined Antisolvent and Cooling Crystallization in Turbulent Flow.

Ind. Eng. Chem. Res. 57, 11702–11711. <https://doi.org/10.1021/acs.iecr.8b01849>

Davoudpour, Y., Hossain, S., Khalil, H.P.S.A., Haafiz, M.K.M., Ishak, Z.A.M., Hassan, A., Sarker, Z.I., 2015. Optimization of high pressure homogenization parameters for

the isolation of cellulosic nanofibers using response surface methodology. *Ind.*

Crops Prod. 74, 381–387. <https://doi.org/10.1016/j.indcrop.2015.05.029>

de Amorim, J.D.P., de Souza, K.C., Duarte, C.R., da Silva Duarte, I., de Assis Sales

Ribeiro, F., Silva, G.S., de Farias, P.M.A., Stingl, A., Costa, A.F.S., Vinhas, G.M.,

Sarubbo, L.A., 2020. Plant and bacterial nanocellulose: production, properties and

applications in medicine, food, cosmetics, electronics and engineering. A review.

Environ. Chem. Lett. 18, 851–869. <https://doi.org/10.1007/s10311-020-00989-9>

Debotton, N., Dahan, A., 2017. Applications of Polymers as Pharmaceutical Excipients

in Solid Oral Dosage Forms. *Med. Res. Rev.* 37, 52–97.

<https://doi.org/10.1002/med.21403>

del Pozo-Rodríguez, A., Solinís, M.Á., Rodríguez-Gascón, A., 2016. Applications of lipid nanoparticles in gene therapy. *Eur. J. Pharm. Biopharm.* 109, 184–193.

<https://doi.org/10.1016/j.ejpb.2016.10.016>

Dong, X., Zhao, M., Yang, B., Yang, X., Shi, J., Jiang, Y., 2011. Effect of High-Pressure Homogenization on the Functional Property of Peanut Protein. *J. Food Process Eng.* 34, 2191–2204. <https://doi.org/10.1111/j.1745-4530.2009.00546.x>

<https://doi.org/10.1111/j.1745-4530.2009.00546.x>

Donsì, F., Ferrari, G., 2016. Essential oil nanoemulsions as antimicrobial agents in food. *J. Biotechnol.* 233, 106–120. <https://doi.org/10.1016/j.jbiotec.2016.07.005>

<https://doi.org/10.1016/j.jbiotec.2016.07.005>

Donsì, F., Sessa, M., Ferrari, G., 2012. Effect of Emulsifier Type and Disruption Chamber Geometry on the Fabrication of Food Nanoemulsions by High Pressure Homogenization. *Ind. Eng. Chem. Res.* 51, 7606–7618.

<https://doi.org/10.1021/ie2017898>

Donsì, F., Wang, Y., Li, J., Huang, Q., 2010. Preparation of Curcumin Sub-micrometer Dispersions by High-Pressure Homogenization. *J. Agric. Food Chem.* 58, 2848–

2853. <https://doi.org/10.1021/jf903968x>

Dopp, J.L., Reuel, N.F., 2018. Process optimization for scalable E. coli extract preparation for cell-free protein synthesis. *Biochem. Eng. J.* 138, 21–28.

<https://doi.org/10.1016/j.bej.2018.06.021>

Dubbelboer, A., Janssen, J., Hoogland, H., Mudaliar, A., Maindarkar, S., Zondervan, E., Meuldijk, J., 2014. Population balances combined with Computational Fluid

Dynamics: A modeling approach for dispersive mixing in a high pressure

homogenizer. *Chem. Eng. Sci.* 117, 376–388.

<https://doi.org/10.1016/j.ces.2014.06.047>

Durán-Lobato, M., Enguix-González, A., Fernández-Arévalo, M., Martín-Banderas, L., 2013. Statistical analysis of solid lipid nanoparticles produced by high-pressure homogenization: a practical prediction approach. *J. Nanoparticle Res.* 15, 1443. <https://doi.org/10.1007/s11051-013-1443-6>

Effio, C.L., Hubbuch, J., 2015. Next generation vaccines and vectors: Designing downstream processes for recombinant protein-based virus-like particles. *Biotechnol. J.* 10, 715–727. <https://doi.org/10.1002/biot.201400392>

Eggenreich, B., Rajamanickam, V., Wurm, D.J., Fricke, J., Herwig, C., Spadiut, O., 2017. A combination of HPLC and automated data analysis for monitoring the efficiency of high-pressure homogenization. *Microb. Cell Fact.* 16, 134. <https://doi.org/10.1186/s12934-017-0749-y>

Eggenreich, B., Wurm, D.J., Rajamanickam, V., Klausser, R., Slouka, C., Spadiut, O., 2020. High pressure homogenization is a key unit operation in inclusion body processing. *J. Biotechnol.* 324, 100022. <https://doi.org/10.1016/j.btecx.2020.100022>

Ekpeni, L.E.N., Benyounis, K.Y., Nkem-Ekpeni, F.F., Stokes, J., Olabi, A.G., 2015. Underlying factors to consider in improving energy yield from biomass source through yeast use on high-pressure homogenizer (hph). *Energy* 81, 74–83. <https://doi.org/10.1016/j.energy.2014.11.038>

Eren, N.M., Santos, P.H.S., Campanella, O., 2015. Mechanically modified xanthan gum: Rheology and polydispersity aspects. *Carbohydr. Polym.* 134, 475–484.

<https://doi.org/10.1016/j.carbpol.2015.07.092>

Failmezger, J., Scholz, S., Blombach, B., Siemann-Herzberg, M., 2018. Cell-Free Protein Synthesis From Fast-Growing *Vibrio natriegens*. *Front. Microbiol.* 9. <https://doi.org/10.3389/fmicb.2018.01146>

Fedele, L., Colla, L., Bobbo, S., Barison, S., Agresti, F., 2011. Experimental stability analysis of different water-based nanofluids. *Nanoscale Res. Lett.* 6, 300. <https://doi.org/10.1186/1556-276X-6-300>

Fontes, D.H., Ribatski, G., Bandarra Filho, E.P., 2015. Experimental evaluation of thermal conductivity, viscosity and breakdown voltage AC of nanofluids of carbon nanotubes and diamond in transformer oil. *Diam. Relat. Mater.* 58, 115–121. <https://doi.org/10.1016/j.diamond.2015.07.007>

Fox, C.B., Barnes V, L., Evers, T., Chesko, J.D., Vedvick, T.S., Coler, R.N., Reed, S.G., Baldwin, S.L., 2013. Adjuvanted pandemic influenza vaccine: variation of emulsion components affects stability, antigen structure, and vaccine efficacy. *Influenza Other Respi. Viruses* 7, 815–826. <https://doi.org/10.1111/irv.12031>

Fu, Z., Wang, L., Li, D., Wei, Q., Adhikari, B., 2011. Effects of high-pressure homogenization on the properties of starch-plasticizer dispersions and their films. *Carbohydr. Polym.* 86, 202–207. <https://doi.org/10.1016/j.carbpol.2011.04.032>

Gall, V., Runde, M., Schuchmann, H., 2016. Extending Applications of High-Pressure Homogenization by Using Simultaneous Emulsification and Mixing (SEM)—An Overview. *Processes* 4, 46. <https://doi.org/10.3390/pr4040046>

- Garcia-Ortega, X., Reyes, C., Montesinos, J.L., Valero, F., 2015. Overall Key Performance Indicator to Optimizing Operation of High-Pressure Homogenizers for a Reliable Quantification of Intracellular Components in *Pichia pastoris*. *Front. Bioeng. Biotechnol.* 3. <https://doi.org/10.3389/fbioe.2015.00107>
- Garusinghe, U.M., Varanasi, S., Raghuwanshi, V.S., Garnier, G., Batchelor, W., 2018. Nanocellulose-montmorillonite composites of low water vapour permeability. *Colloids Surfaces A Physicochem. Eng. Asp.* 540, 233–241. <https://doi.org/10.1016/j.colsurfa.2018.01.010>
- Gaulin, A., 1904. System for intimately mixing milk. US756953A.
- George, A., Sanjay, M.R., Srisuk, R., Parameswaranpillai, J., Siengchin, S., 2020. A comprehensive review on chemical properties and applications of biopolymers and their composites. *Int. J. Biol. Macromol.* 154, 329–338. <https://doi.org/10.1016/j.ijbiomac.2020.03.120>
- Georget, E., Miller, B., Callanan, M., Heinz, V., Mathys, A., 2014. (Ultra) High Pressure Homogenization for Continuous High Pressure Sterilization of Pumpable Foods - A Review. *Front. Nutr.* 1. <https://doi.org/10.3389/fnut.2014.00015>
- Gómez H., C., Serpa, A., Velásquez-Cock, J., Gañán, P., Castro, C., Vélez, L., Zuluaga, R., 2016. Vegetable nanocellulose in food science: A review. *Food Hydrocoll.* 57, 178–186. <https://doi.org/10.1016/j.foodhyd.2016.01.023>
- Grassi, S., Alamprese, C., 2018. Advances in NIR spectroscopy applied to process analytical technology in food industries. *Curr. Opin. Food Sci.* 22, 17–21. <https://doi.org/10.1016/j.cofs.2017.12.008>

- Grüttner, C., Müller, K., Teller, J., Westphal, F., Foreman, A., Ivkov, R., 2007. Synthesis and antibody conjugation of magnetic nanoparticles with improved specific power absorption rates for alternating magnetic field cancer therapy. *J. Magn. Magn. Mater.* 311, 181–186. <https://doi.org/10.1016/j.jmmm.2006.10.1151>
- Gué, E., Since, M., Ropars, S., Herbinet, R., Le Pluart, L., Malzert-Fréon, A., 2016. Evaluation of the versatile character of a nanoemulsion formulation. *Int. J. Pharm.* 498, 49–65. <https://doi.org/10.1016/j.ijpharm.2015.12.010>
- Guerra, V., Wan, C., Degirmenci, V., Sloan, J., Presvytis, D., McNally, T., 2018. 2D boron nitride nanosheets (BNNS) prepared by high-pressure homogenisation: structure and morphology. *Nanoscale* 10, 19469–19477. <https://doi.org/10.1039/C8NR06429F>
- Gul, O., Atalar, I., Saricaoglu, F.T., Yazici, F., 2018. Effect of multi-pass high pressure homogenization on physicochemical properties of hazelnut milk from hazelnut cake: An investigation by response surface methodology. *J. Food Process. Preserv.* 42, e13615. <https://doi.org/10.1111/jfpp.13615>
- Gul, O., Saricaoglu, F.T., Mortas, M., Atalar, I., Yazici, F., 2017. Effect of high pressure homogenization (HPH) on microstructure and rheological properties of hazelnut milk. *Innov. Food Sci. Emerg. Technol.* 41, 411–420. <https://doi.org/10.1016/j.ifset.2017.05.002>
- Günerken, E., D'Hondt, E., Eppink, M.H.M., Garcia-Gonzalez, L., Elst, K., Wijffels, R.H., 2015. Cell disruption for microalgae biorefineries. *Biotechnol. Adv.* 33, 243–260. <https://doi.org/10.1016/j.biotechadv.2015.01.008>

- Gupta, A., Eral, H.B., Hatton, T.A., Doyle, P.S., 2016a. Controlling and predicting droplet size of nanoemulsions: scaling relations with experimental validation. *Soft Matter* 12, 1452–1458. <https://doi.org/10.1039/C5SM02051D>
- Gupta, A., Eral, H.B., Hatton, T.A., Doyle, P.S., 2016b. Nanoemulsions: formation, properties and applications. *Soft Matter* 12, 2826–2841. <https://doi.org/10.1039/C5SM02958A>
- Haensler, J., 2017. Manufacture of Oil-in-Water Emulsion Adjuvants. pp. 165–180. https://doi.org/10.1007/978-1-4939-6445-1_12
- Håkansson, A., 2022. Effect of inlet chamber design and operation conditions on laminar drop deformation in a production-scale high-pressure homogenizer—A hydrodynamic investigation. *Chem. Eng. Res. Des.* 180, 333–345. <https://doi.org/10.1016/j.cherd.2022.02.033>
- Håkansson, A., 2019. Emulsion Formation by Homogenization: Current Understanding and Future Perspectives. *Annu. Rev. Food Sci. Technol.* 10, 239–258. <https://doi.org/10.1146/annurev-food-032818-121501>
- Håkansson, A., 2018. Flow pulsation plays an important role for high-pressure homogenization in laboratory-scale. *Chem. Eng. Res. Des.* 138, 472–481. <https://doi.org/10.1016/j.cherd.2018.09.015>
- Håkansson, A., Fuchs, L., Innings, F., Revstedt, J., Bergenståhl, B., Trägårdh, C., 2010. Visual observations and acoustic measurements of cavitation in an experimental model of a high-pressure homogenizer. *J. Food Eng.* 100, 504–513. <https://doi.org/10.1016/j.jfoodeng.2010.04.038>

- Håkansson, A., Fuchs, L., Innings, F., Revstedt, J., Trägårdh, C., Bergenståhl, B., 2012. Experimental validation of k - ϵ RANS-CFD on a high-pressure homogenizer valve. Chem. Eng. Sci. 71, 264–273. <https://doi.org/10.1016/j.ces.2011.12.039>
- Håkansson, A., Innings, F., Trägårdh, C., Bergenståhl, B., 2013. A high-pressure homogenization emulsification model—Improved emulsifier transport and hydrodynamic coupling. Chem. Eng. Sci. 91, 44–53. <https://doi.org/10.1016/j.ces.2013.01.011>
- Håkansson, A., Trägårdh, C., Bergenståhl, B., 2009. Dynamic simulation of emulsion formation in a high pressure homogenizer. Chem. Eng. Sci. 64, 2915–2925. <https://doi.org/10.1016/j.ces.2009.03.034>
- Hansuld, E.M., Briens, L., 2014. A review of monitoring methods for pharmaceutical wet granulation. Int. J. Pharm. 472, 192–201. <https://doi.org/10.1016/j.ijpharm.2014.06.027>
- Harish Prashanth, K.V., Tharanathan, R.N., 2007. Chitin/chitosan: modifications and their unlimited application potential—an overview. Trends Food Sci. Technol. 18, 117–131. <https://doi.org/10.1016/j.tifs.2006.10.022>
- Harte, F., 2016. Food Processing by High-Pressure Homogenization. Springer, New York, NY, pp. 123–141. https://doi.org/10.1007/978-1-4939-3234-4_7
- Harte, F., Venegas, R., 2010. A Model for Viscosity Reduction in Polysaccharides Subjected to High-Pressure Homogenization. J. Texture Stud. 41, 49–61. <https://doi.org/10.1111/j.1745-4603.2009.00212.x>

Homayouni, A., Sadeghi, F., Varshosaz, J., Garekani, H.A., Nokhodchi, A., 2014.

Comparing various techniques to produce micro/nanoparticles for enhancing the dissolution of celecoxib containing PVP. *Eur. J. Pharm. Biopharm.* 88, 261–274.

<https://doi.org/10.1016/j.ejpb.2014.05.022>

Hongrattanavichit, I., Aht-Ong, D., 2020. Nanofibrillation and characterization of

sugarcane bagasse agro-waste using water-based steam explosion and high-pressure homogenization. *J. Clean. Prod.* 277, 123471.

<https://doi.org/10.1016/j.jclepro.2020.123471>

Hu, C., Qian, A., Wang, Q., Xu, F., He, Y., Xu, J., Xia, Y., Xia, Q., 2016. Industrialization

of lipid nanoparticles: From laboratory-scale to large-scale production line. *Eur. J.*

Pharm. Biopharm. 109, 206–213. <https://doi.org/10.1016/j.ejpb.2016.10.018>

Hutterer, K.M., Hong, R.W., Lull, J., Zhao, X., Wang, T., Pei, R., Le, M.E., Borisov, O.,

Piper, R., Liu, Y.D., Petty, K., Apostol, I., Flynn, G.C., 2013. Monoclonal antibody disulfide reduction during manufacturing. *MAbs* 5, 608–613.

<https://doi.org/10.4161/mabs.24725>

Inguva, P.K., Ooi, S.M., Desai, P.M., Heng, P.W.S., 2015. Encapsulation of volatiles by

homogenized partially-cross linked alginates. *Int. J. Pharm.* 496, 709–716.

<https://doi.org/10.1016/j.ijpharm.2015.11.017>

Inguva, P.K., Schickel, K.C., Braatz, R.D., 2022. Efficient numerical schemes for

population balance models. *Comput. Chem. Eng.* 162, 107808.

<https://doi.org/10.1016/j.compchemeng.2022.107808>

Jahnke, S., 1998. The theory of high-pressure homogenization, in: Muller, R., Benita,

S., Bohm, H.L. (Eds.), *Emulsions and Nanosuspensions for the Formulation of Poorly Soluble Drugs*. Medpharm, Stuttgart, pp. 177–200.

Jerome, R.E., Singh, S.K., Dwivedi, M., 2019. Process analytical technology for bakery industry: A review. *J. Food Process Eng.* 42. <https://doi.org/10.1111/jfpe.13143>

Johnson, D.W., Dobson, B.P., Coleman, K.S., 2015. A manufacturing perspective on graphene dispersions. *Curr. Opin. Colloid Interface Sci.* 20, 367–382.
<https://doi.org/10.1016/j.cocis.2015.11.004>

Jorfi, M., Foster, E.J., 2015. Recent advances in nanocellulose for biomedical applications. *J. Appl. Polym. Sci.* 132, n/a-n/a. <https://doi.org/10.1002/app.41719>

Kakran, M., Shegokar, R., Sahoo, N.G., Al Shaal, L., Li, L., Müller, R.H., 2012. Fabrication of quercetin nanocrystals: Comparison of different methods. *Eur. J. Pharm. Biopharm.* 80, 113–121. <https://doi.org/10.1016/j.ejpb.2011.08.006>

Kamel, R., El-Wakil, N.A., Dufresne, A., Elkasabgy, N.A., 2020. Nanocellulose: From an agricultural waste to a valuable pharmaceutical ingredient. *Int. J. Biol. Macromol.* 163, 1579–1590. <https://doi.org/10.1016/j.ijbiomac.2020.07.242>

Kang, H.J., Min, S.C., 2010. Potato peel-based biopolymer film development using high-pressure homogenization, irradiation, and ultrasound. *LWT - Food Sci. Technol.* 43, 903–909. <https://doi.org/10.1016/j.lwt.2010.01.025>

Kargarzadeh, H., Mariano, M., Huang, J., Lin, N., Ahmad, I., Dufresne, A., Thomas, S., 2017. Recent developments on nanocellulose reinforced polymer nanocomposites: A review. *Polymer (Guildf)*. 132, 368–393.

<https://doi.org/10.1016/j.polymer.2017.09.043>

Kaur, B., Ariffin, F., Bhat, R., Karim, A.A., 2012. Progress in starch modification in the last decade. *Food Hydrocoll.* 26, 398–404.

<https://doi.org/10.1016/j.foodhyd.2011.02.016>

Kawee, N., Lam, N.T., Sukyai, P., 2018. Homogenous isolation of individualized bacterial nanofibrillated cellulose by high pressure homogenization. *Carbohydr. Polym.* 179, 394–401. <https://doi.org/10.1016/j.carbpol.2017.09.101>

Kelly, W.J., Muske, K.R., 2004. Optimal operation of high-pressure homogenization for intracellular product recovery. *Bioprocess Biosyst. Eng.* 27, 25–37.

<https://doi.org/10.1007/s00449-004-0378-9>

Khuri, A.I., Mukhopadhyay, S., 2010. Response surface methodology. *Wiley Interdiscip. Rev. Comput. Stat.* 2, 128–149. <https://doi.org/10.1002/wics.73>

Kluge, J., Muhrer, G., Mazzotti, M., 2012. High pressure homogenization of pharmaceutical solids. *J. Supercrit. Fluids* 66, 380–388.

<https://doi.org/10.1016/j.supflu.2012.01.009>

Kotta, S., Khan, A.W., Ansari, S.H., Sharma, R.K., Ali, J., 2015. Formulation of nanoemulsion: a comparison between phase inversion composition method and high-pressure homogenization method. *Drug Deliv.* 22, 455–466.

<https://doi.org/10.3109/10717544.2013.866992>

Lawton, G., 2021. Brewing milk. *New Sci.* 251, 46–49. [https://doi.org/10.1016/S0262-4079\(21\)01431-7](https://doi.org/10.1016/S0262-4079(21)01431-7)

- Lebaz, N., Sheibat-Othman, N., 2019. Modeling Emulsification in Static Mixers: Equilibrium Correlations versus Population Balance Equations. *Chem. Eng. Technol.* ceat.201900109. <https://doi.org/10.1002/ceat.201900109>
- Lee, S.-J., Jeong, J.-R., Shin, S.-C., Kim, J.-C., Chang, Y.-H., Chang, Y.-M., Kim, J.-D., 2004. Nanoparticles of magnetic ferric oxides encapsulated with poly(D,L lactide-co-glycolide) and their applications to magnetic resonance imaging contrast agent. *J. Magn. Magn. Mater.* 272–276, 2432–2433. <https://doi.org/10.1016/j.jmmm.2003.12.416>
- Lenhart, V., Quodbach, J., Kleinebudde, P., 2020. Fibrillated Cellulose via High Pressure Homogenization: Analysis and Application for Orodispensible Films. *AAPS PharmSciTech* 21, 33. <https://doi.org/10.1208/s12249-019-1593-7>
- Levy, R., Okun, Z., Davidovich-Pinhas, M., Shpigelman, A., 2021. Utilization of high-pressure homogenization of potato protein isolate for the production of dairy-free yogurt-like fermented product. *Food Hydrocoll.* 113, 106442. <https://doi.org/10.1016/j.foodhyd.2020.106442>
- Levy, R., Okun, Z., Shpigelman, A., 2020. High-Pressure Homogenization: Principles and Applications Beyond Microbial Inactivation. *Food Eng. Rev.* <https://doi.org/10.1007/s12393-020-09239-8>
- Lewińska, A., 2021. Optimizing the Process Design of Oil-in-Water Nanoemulsion for Delivering Poorly Soluble Cannabidiol Oil. *Processes* 9, 1180. <https://doi.org/10.3390/pr9071180>
- Li, F., Mascheroni, E., Piergiovanni, L., 2015. The Potential of NanoCellulose in the

Packaging Field: A Review. *Packag. Technol. Sci.* 28, 475–508.

<https://doi.org/10.1002/pts.2121>

Li, J., Bing, C., Jun, W., Xiaoying, L., Ailin, H., Qi, J., 2019. High pressure homogenization treatment on graphene oxide and its electrochemical energy storage performance. *Appl. Surf. Sci.* 493, 441–447.

<https://doi.org/10.1016/j.apsusc.2019.07.046>

Li, J., Wei, X., Wang, Q., Chen, J., Chang, G., Kong, L., Su, J., Liu, Y., 2012.

Homogeneous isolation of nanocellulose from sugarcane bagasse by high pressure homogenization. *Carbohydr. Polym.* 90, 1609–1613.

<https://doi.org/10.1016/j.carbpol.2012.07.038>

Li, N., Panagiotopoulos, A.Z., Nikoubashman, A., 2017. Structured Nanoparticles from the Self-Assembly of Polymer Blends through Rapid Solvent Exchange. *Langmuir* 33, 6021–6028. <https://doi.org/10.1021/acs.langmuir.7b00291>

Li, T., Rui, X., Wang, K., Jiang, M., Chen, X., Li, W., Dong, M., 2015. Study of the dynamic states of water and effects of high-pressure homogenization on water distribution in tofu by using low-field nuclear magnetic resonance. *Innov. Food Sci. Emerg. Technol.* 30, 61–68. <https://doi.org/10.1016/j.ifset.2015.03.008>

Li, Y., Zhao, X., Zu, Y., Zhang, Y., 2015. Preparation and characterization of paclitaxel nanosuspension using novel emulsification method by combining high speed homogenizer and high pressure homogenization. *Int. J. Pharm.* 490, 324–333. <https://doi.org/10.1016/j.ijpharm.2015.05.070>

Lin, N., Dufresne, A., 2014. Nanocellulose in biomedicine: Current status and future

prospect. *Eur. Polym. J.* 59, 302–325.

<https://doi.org/10.1016/j.eurpolymj.2014.07.025>

Linke, C., Drusch, S., 2016. Turbidity in oil-in-water-emulsions — Key factors and visual perception. *Food Res. Int.* 89, 202–210.

<https://doi.org/10.1016/j.foodres.2016.07.019>

Lomis, N., Westfall, S., Farahdel, L., Malhotra, M., Shum-Tim, D., Prakash, S., 2016.

Human Serum Albumin Nanoparticles for Use in Cancer Drug Delivery: Process Optimization and In Vitro Characterization. *Nanomaterials* 6, 116.

<https://doi.org/10.3390/nano6060116>

Loveday, S.M., 2019. Food Proteins: Technological, Nutritional, and Sustainability

Attributes of Traditional and Emerging Proteins. *Annu. Rev. Food Sci. Technol.* 10, 311–339. <https://doi.org/10.1146/annurev-food-032818-121128>

Mahfoudhi, N., Boufi, S., 2017. Nanocellulose as a novel nanostructured adsorbent for environmental remediation: a review. *Cellulose* 24, 1171–1197.

<https://doi.org/10.1007/s10570-017-1194-0>

Maindarkar, S.N., Raikar, N.B., Bongers, P., Henson, M.A., 2012. Incorporating

emulsion drop coalescence into population balance equation models of high pressure homogenization. *Colloids Surfaces A Physicochem. Eng. Asp.* 396, 63–

73. <https://doi.org/10.1016/j.colsurfa.2011.12.041>

Malekzadeh, S., Roohi, E., 2015. Investigation of Different Droplet Formation Regimes

in a T-junction Microchannel Using the VOF Technique in OpenFOAM. *Microgravity Sci. Technol.* 27, 231–243. <https://doi.org/10.1007/s12217-015-9440-2>

- Martínez-Monteagudo, S.I., Kamat, S., Patel, N., Konuklar, G., Rangavajla, N., Balasubramaniam, V.M., 2017. Improvements in emulsion stability of dairy beverages treated by high pressure homogenization: A pilot-scale feasibility study. *J. Food Eng.* 193, 42–52. <https://doi.org/10.1016/j.jfoodeng.2016.08.011>
- Metzger, K.F.J., Padutsch, W., Pekarsky, A., Kopp, J., Voloshin, A.M., Kühnel, H., Maurer, M., 2020. IGF1 inclusion bodies: A QbD based process approach for efficient USP as well as early DSP unit operations. *J. Biotechnol.* 312, 23–34. <https://doi.org/10.1016/j.jbiotec.2020.02.014>
- Mitchell, M.J., Billingsley, M.M., Haley, R.M., Wechsler, M.E., Peppas, N.A., Langer, R., 2021. Engineering precision nanoparticles for drug delivery. *Nat. Rev. Drug Discov.* 20, 101–124. <https://doi.org/10.1038/s41573-020-0090-8>
- Mukherjee, S., Zarghami, A., Haringa, C., van As, K., Kenjereš, S., Van den Akker, H.E.A., 2018. Simulating liquid droplets: A quantitative assessment of lattice Boltzmann and Volume of Fluid methods. *Int. J. Heat Fluid Flow* 70, 59–78. <https://doi.org/10.1016/j.ijheatfluidflow.2017.12.001>
- Mushi, N.E., Nishino, T., Berglund, L.A., Zhou, Q., 2019. Strong and Tough Chitin Film from α -Chitin Nanofibers Prepared by High Pressure Homogenization and Chitosan Addition. *ACS Sustain. Chem. Eng.* 7, 1692–1697. <https://doi.org/10.1021/acssuschemeng.8b05452>
- Natarajan, A., Gruettner, C., Ivkov, R., DeNardo, G.L., Mirick, G., Yuan, A., Foreman, A., DeNardo, S.J., 2008. NanoFerrite Particle Based Radioimmunonanoparticles: Binding Affinity and In Vivo Pharmacokinetics. *Bioconjug. Chem.* 19, 1211–1218.

<https://doi.org/10.1021/bc800015n>

Oktay, A.N., Ilbasimis-Tamer, S., Celebi, N., 2019. The effect of critical process parameters of the high pressure homogenization technique on the critical quality attributes of flurbiprofen nanosuspensions. *Pharm. Dev. Technol.* 24, 1278–1286. <https://doi.org/10.1080/10837450.2019.1667384>

Oliveira, G.A., Cardenas Contreras, E.M., Bandarra Filho, E.P., 2017. Experimental study on the heat transfer of MWCNT/water nanofluid flowing in a car radiator. *Appl. Therm. Eng.* 111, 1450–1456. <https://doi.org/10.1016/j.applthermaleng.2016.05.086>

Ono, Y., Ogura, K., Kaku, Y., Fujisawa, S., Isogai, A., 2020. Structural changes in α -chitin through nanofibrillation by high-pressure homogenization in water. *Polym. J.* 52, 813–818. <https://doi.org/10.1038/s41428-020-0322-0>

Otoni, C.G., Azeredo, H.M.C., Mattos, B.D., Beaumont, M., Correa, D.S., Rojas, O.J., 2021. The Food–Materials Nexus: Next Generation Bioplastics and Advanced Materials from Agri-Food Residues. *Adv. Mater.* 33, 2102520. <https://doi.org/10.1002/adma.202102520>

Pacaphol, K., Aht-Ong, D., 2017. Preparation of hemp nanofibers from agricultural waste by mechanical defibrillation in water. *J. Clean. Prod.* 142, 1283–1295. <https://doi.org/10.1016/j.jclepro.2016.09.008>

Pardeike, J., Hommoss, A., Müller, R.H., 2009. Lipid nanoparticles (SLN, NLC) in cosmetic and pharmaceutical dermal products. *Int. J. Pharm.* 366, 170–184. <https://doi.org/10.1016/j.ijpharm.2008.10.003>

- Paredes, A.J., Llabot, J.M., Sánchez Bruni, S., Allemandi, D., Palma, S.D., 2016. Self-dispersible nanocrystals of albendazole produced by high pressure homogenization and spray-drying. *Drug Dev. Ind. Pharm.* 42, 1564–1570.
<https://doi.org/10.3109/03639045.2016.1151036>
- Passalacqua, A., Laurent, F., Madadi-Kandjani, E., Heylmun, J.C., Fox, R.O., 2018. An open-source quadrature-based population balance solver for OpenFOAM. *Chem. Eng. Sci.* 176, 306–318. <https://doi.org/10.1016/j.ces.2017.10.043>
- Patil, M.D., Patel, G., Surywanshi, B., Shaikh, N., Garg, P., Chisti, Y., Banerjee, U.C., 2016. Disruption of *Pseudomonas putida* by high pressure homogenization: a comparison of the predictive capacity of three process models for the efficient release of arginine deiminase. *AMB Express* 6, 84. <https://doi.org/10.1186/s13568-016-0260-6>
- Patrignani, F., Lanciotti, R., 2016. Applications of High and Ultra High Pressure Homogenization for Food Safety. *Front. Microbiol.* 7.
<https://doi.org/10.3389/fmicb.2016.01132>
- Pekarsky, A., Spadiut, O., Rajamanickam, V., Wurm, D.J., 2019. A fast and simple approach to optimize the unit operation high pressure homogenization - a case study for a soluble therapeutic protein in *E. coli*. *Prep. Biochem. Biotechnol.* 49, 74–81. <https://doi.org/10.1080/10826068.2018.1536988>
- Peshkovsky, A.S., Bystryak, S., 2014. Continuous-flow production of a pharmaceutical nanoemulsion by high-amplitude ultrasound: Process scale-up. *Chem. Eng. Process. Process Intensif.* 82, 132–136. <https://doi.org/10.1016/j.cep.2014.05.007>

- Phanthong, P., Reubroycharoen, P., Hao, X., Xu, G., Abudula, A., Guan, G., 2018. Nanocellulose: Extraction and application. *Carbon Resour. Convers.* 1, 32–43. <https://doi.org/10.1016/j.crcon.2018.05.004>
- Piao, M., Chu, J., Wang, X., Chi, Y., Zhang, H., Li, C., Shi, H., Joo, M.-K., 2018. Hydrothermal synthesis of stable metallic 1T phase WS₂ nanosheets for thermoelectric application. *Nanotechnology* 29, 025705. <https://doi.org/10.1088/1361-6528/aa9bfe>
- Poliseli-Scopel, F.H., Hernández-Herrero, M., Guamis, B., Ferragut, V., 2013. Characteristics of soymilk pasteurized by ultra high pressure homogenization (UHPH). *Innov. Food Sci. Emerg. Technol.* 20, 73–80. <https://doi.org/10.1016/j.ifset.2013.06.001>
- Porto, B.C., Augusto, P.E.D., Terekhov, A., Hamaker, B.R., Cristianini, M., 2015. Effect of dynamic high pressure on technological properties of cashew tree gum (*Anacardium occidentale* L.). *Carbohydr. Polym.* 129, 187–193. <https://doi.org/10.1016/j.carbpol.2015.04.052>
- Prateepchanachai, S., Thakhiew, W., Devahastin, S., Soponronnarit, S., 2017. Mechanical properties improvement of chitosan films via the use of plasticizer, charge modifying agent and film solution homogenization. *Carbohydr. Polym.* 174, 253–261. <https://doi.org/10.1016/j.carbpol.2017.06.069>
- Pu, Y., Chaudhry, S., Parikh, M., Berry, J., 2015. Application of in-line viscometer for in-process monitoring of microcrystalline cellulose-carboxymethylcellulose hydrogel formation during batch manufacturing. *Drug Dev. Ind. Pharm.* 41, 28–34.

<https://doi.org/10.3109/03639045.2013.845837>

- Qi, D., Cao, Z., Ziener, U., 2014. Recent advances in the preparation of hybrid nanoparticles in miniemulsions. *Adv. Colloid Interface Sci.* 211, 47–62.
<https://doi.org/10.1016/j.cis.2014.06.001>
- Qi, X., Zhang, H.-B., Xu, J., Wu, X., Yang, D., Qu, J., Yu, Z.-Z., 2017. Highly Efficient High-Pressure Homogenization Approach for Scalable Production of High-Quality Graphene Sheets and Sandwich-Structured $\alpha\text{-Fe}_2\text{O}_3$ /Graphene Hybrids for High-Performance Lithium-Ion Batteries. *ACS Appl. Mater. Interfaces* 9, 11025–11034. <https://doi.org/10.1021/acsami.7b00808>
- Qian, C., McClements, D.J., 2011. Formation of nanoemulsions stabilized by model food-grade emulsifiers using high-pressure homogenization: Factors affecting particle size. *Food Hydrocoll.* 25, 1000–1008.
<https://doi.org/10.1016/j.foodhyd.2010.09.017>
- Rabinow, B.E., 2004. Nanosuspensions in drug delivery. *Nat. Rev. Drug Discov.* 3, 785–796. <https://doi.org/10.1038/nrd1494>
- Raghav, N., Sharma, M.R., Kennedy, J.F., 2021. Nanocellulose: A mini-review on types and use in drug delivery systems. *Carbohydr. Polym. Technol. Appl.* 2, 100031.
<https://doi.org/10.1016/j.carpta.2020.100031>
- Raikar, N.B., Bhatia, S.R., Malone, M.F., McClements, D.J., Almeida-Rivera, C., Bongers, P., Henson, M.A., 2010. Prediction of emulsion drop size distributions with population balance equation models of multiple drop breakage. *Colloids Surfaces A Physicochem. Eng. Asp.* 361, 96–108.

<https://doi.org/10.1016/j.colsurfa.2010.03.020>

Ralbovsky, N.M., Soukup, R.J., Lomont, J.P., Lauro, M.L., Gulasarian, A., Saha-Shah, A., Winters, M.A., Richardson, D.D., Wang, S.-C., Mangion, I., Smith, J.P., 2022. In situ real time monitoring of emulsification and homogenization processes for vaccine adjuvants. *Analyst*. <https://doi.org/10.1039/D1AN01797G>

Rizal, S., Sadasivuni, K.K., Atiqah, M.S.N., Olaiya, N.G., Paridah, M.T., Abdullah, C.K., Alfatah, T., Mistar, E.M., Khalil, H.P.S.A., 2021. The role of cellulose nanofibrillated fibers produced with combined supercritical carbon dioxide and high-pressure homogenization process as reinforcement material in biodegradable polymer. *Polym. Compos.* 42, 1795–1808. <https://doi.org/10.1002/pc.25935>

Rocca-Smith, J.R., Pasquarelli, R., Lagorce-Tachon, A., Rousseau, J., Fontaine, S., Aguié-Béghin, V., Debeaufort, F., Karbowiak, T., 2019. Toward Sustainable PLA-Based Multilayer Complexes with Improved Barrier Properties. *ACS Sustain. Chem. Eng.* 7, 3759–3771. <https://doi.org/10.1021/acssuschemeng.8b04064>

Rüdt, M., Briskot, T., Hubbuch, J., 2017. Advances in downstream processing of biologics – Spectroscopy: An emerging process analytical technology. *J. Chromatogr. A* 1490, 2–9. <https://doi.org/10.1016/j.chroma.2016.11.010>

Sabo, R., Yermakov, A., Law, C.T., Elhajjar, R., 2016. Nanocellulose-Enabled Electronics, Energy Harvesting Devices, Smart Materials and Sensors: A Review. *J. Renew. Mater.* 4, 297–312. <https://doi.org/10.7569/JRM.2016.634114>

Saidur, R., Leong, K.Y., Mohammed, H.A., 2011. A review on applications and challenges of nanofluids. *Renew. Sustain. Energy Rev.* 15, 1646–1668.

<https://doi.org/10.1016/j.rser.2010.11.035>

Salaberria, A.M., Fernandes, S.C.M., Diaz, R.H., Labidi, J., 2015. Processing of α -chitin nanofibers by dynamic high pressure homogenization: Characterization and antifungal activity against *A. niger*. *Carbohydr. Polym.* 116, 286–291.
<https://doi.org/10.1016/j.carbpol.2014.04.047>

Salas, C., Nypelö, T., Rodriguez-Abreu, C., Carrillo, C., Rojas, O.J., 2014. Nanocellulose properties and applications in colloids and interfaces. *Curr. Opin. Colloid Interface Sci.* 19, 383–396. <https://doi.org/10.1016/j.cocis.2014.10.003>

Salazar, J., Ghanem, A., Müller, R.H., Möschwitzer, J.P., 2012. Nanocrystals: Comparison of the size reduction effectiveness of a novel combinative method with conventional top-down approaches. *Eur. J. Pharm. Biopharm.* 81, 82–90.
<https://doi.org/10.1016/j.ejpb.2011.12.015>

Salehi, F., 2020. Physico-chemical and rheological properties of fruit and vegetable juices as affected by high pressure homogenization: A review. *Int. J. Food Prop.* 23, 1136–1149. <https://doi.org/10.1080/10942912.2020.1781167>

Samarasinghe, N., Fernando, S., Lacey, R., Faulkner, W.B., 2012. Algal cell rupture using high pressure homogenization as a prelude to oil extraction. *Renew. Energy* 48, 300–308. <https://doi.org/10.1016/j.renene.2012.04.039>

Satam, C.C., Meredith, J.C., 2021. Increasing efficiency of the homogenization process for production of chitin nanofibers for barrier film applications. *Carbohydr. Polym.* 274, 118658. <https://doi.org/10.1016/j.carbpol.2021.118658>

- Schlender, M., Minke, K., Spiegel, B., Schuchmann, H.P., 2015a. High-pressure double stage homogenization processes: Influences of plant setup on oil droplet size. *Chem. Eng. Sci.* 131, 162–171. <https://doi.org/10.1016/j.ces.2015.03.055>
- Schlender, M., Spengler, A., Schuchmann, H.P., 2015b. High-pressure emulsion formation in cylindrical coaxial orifices: Influence of cavitation induced pattern on oil drop size. *Int. J. Multiph. Flow* 74, 84–95. <https://doi.org/10.1016/j.ijmultiphaseflow.2015.04.004>
- Schlüter, B., Mülhaupt, R., Kailer, A., 2014. Synthesis and Tribological Characterization of Stable Dispersions of Thermally Reduced Graphite Oxide. *Tribol. Lett.* 53, 353–363. <https://doi.org/10.1007/s11249-013-0275-y>
- Schuh, R.S., de Carvalho, T.G., Giugliani, R., Matte, U., Baldo, G., Teixeira, H.F., 2018. Gene editing of MPS I human fibroblasts by co-delivery of a CRISPR/Cas9 plasmid and a donor oligonucleotide using nanoemulsions as nonviral carriers. *Eur. J. Pharm. Biopharm.* 122, 158–166. <https://doi.org/10.1016/j.ejpb.2017.10.017>
- Shahbazi, M., Majzoobi, M., Farahnaky, A., 2018. Physical modification of starch by high-pressure homogenization for improving functional properties of κ -carrageenan/starch blend film. *Food Hydrocoll.* 85, 204–214. <https://doi.org/10.1016/j.foodhyd.2018.07.017>
- Shang, J., Xue, F., Ding, E., 2015. The facile fabrication of few-layer graphene and graphite nanosheets by high pressure homogenization. *Chem. Commun.* 51, 15811–15814. <https://doi.org/10.1039/C5CC06151B>
- Shang, J., Xue, F., Fan, C., Ding, E., 2016. Preparation of few layers hexagonal boron

nitride nanosheets via high-pressure homogenization. *Mater. Lett.* 181, 144–147.

<https://doi.org/10.1016/j.matlet.2016.05.154>

Shanmugam, K., Ang, S., Maliha, M., Raghuwanshi, V., Varanasi, S., Garnier, G.,

Batchelor, W., 2021. High-performance homogenized and spray coated nanofibrillated cellulose-montmorillonite barriers. *Cellulose* 28, 405–416.

<https://doi.org/10.1007/s10570-020-03515-w>

Sharma, S., Sahni, J.K., Ali, J., Baboota, S., 2015. Effect of high-pressure

homogenization on formulation of TPGS loaded nanoemulsion of rutin – pharmacodynamic and antioxidant studies. *Drug Deliv.* 22, 541–551.

<https://doi.org/10.3109/10717544.2014.893382>

Shayganpour, A., Naderizadeh, S., Grasselli, S., Malchiodi, A., Bayer, I.S., 2019.

Stacked-Cup Carbon Nanotube Flexible Paper Based on Soy Lecithin and Natural Rubber. *Nanomaterials* 9, 824. <https://doi.org/10.3390/nano9060824>

Shi, A., Li, D., Wang, L., Li, B., Adhikari, B., 2011. Preparation of starch-based

nanoparticles through high-pressure homogenization and miniemulsion cross-linking: Influence of various process parameters on particle size and stability.

Carbohydr. Polym. 83, 1604–1610. <https://doi.org/10.1016/j.carbpol.2010.10.011>

Sica, L.U.R., Contreras, E.M.C., Bandarra Filho, E.P., Parise, J.A.R., 2021. An

experimental viscosity investigation on the use of non-Newtonian graphene heat transfer nanofluids at below-ambient temperatures. *Int. J. Energy Res.* 45, 14530–

14546. <https://doi.org/10.1002/er.6675>

Silva, H.D., Cerqueira, M.A., Vicente, A.A., 2015. Influence of surfactant and processing

conditions in the stability of oil-in-water nanoemulsions. *J. Food Eng.* 167, 89–98.

<https://doi.org/10.1016/j.jfoodeng.2015.07.037>

Silva, H.D., Cerqueira, M.Â., Vicente, A.A., 2012. Nanoemulsions for Food Applications: Development and Characterization. *Food Bioprocess Technol.* 5, 854–867.

<https://doi.org/10.1007/s11947-011-0683-7>

Simon, L.L., Pataki, H., Marosi, G., Meemken, F., Hungerbühler, K., Baiker, A., Tummala, S., Glennon, B., Kuentz, M., Steele, G., Kramer, H.J.M., Rydzak, J.W., Chen, Z., Morris, J., Kjell, F., Singh, R., Gani, R., Gernaey, K. V., Louhi-Kultanen, M., O'Reilly, J., Sandler, N., Antikainen, O., Yliruusi, J., Froberg, P., Ulrich, J., Braatz, R.D., Leysens, T., von Stosch, M., Oliveira, R., Tan, R.B.H., Wu, H., Khan, M., O'Grady, D., Pandey, A., Westra, R., Delle-Case, E., Pape, D., Angelosante, D., Maret, Y., Steiger, O., Lenner, M., Abbou-Oucherif, K., Nagy, Z.K., Litster, J.D., Kamaraju, V.K., Chiu, M.-S., 2015. Assessment of Recent Process Analytical Technology (PAT) Trends: A Multiauthor Review. *Org. Process Res. Dev.* 19, 3–62.

<https://doi.org/10.1021/op500261y>

Singh, Y., Meher, J.G., Raval, K., Khan, F.A., Chaurasia, M., Jain, N.K., Chourasia, M.K., 2017. Nanoemulsion: Concepts, development and applications in drug delivery. *J. Control. Release* 252, 28–49.

<https://doi.org/10.1016/j.jconrel.2017.03.008>

Slouka, C., Kopp, J., Hutwimmer, S., Strahammer, M., Strohmmer, D., Eitenberger, E., Schwaighofer, A., Herwig, C., 2018. Custom made inclusion bodies: impact of classical process parameters and physiological parameters on inclusion body

quality attributes. *Microb. Cell Fact.* 17, 148. <https://doi.org/10.1186/s12934-018-0997-5>

Song, X., Zhou, C., Fu, F., Chen, Z., Wu, Q., 2013. Effect of high-pressure homogenization on particle size and film properties of soy protein isolate. *Ind. Crops Prod.* 43, 538–544. <https://doi.org/10.1016/j.indcrop.2012.08.005>

Soni, G., Kale, K., Shetty, S., Gupta, M.K., Yadav, K.S., 2020. Quality by design (QbD) approach in processing polymeric nanoparticles loading anticancer drugs by high pressure homogenizer. *Heliyon* 6, e03846. <https://doi.org/10.1016/j.heliyon.2020.e03846>

Spiden, E.M., Yap, B.H.J., Hill, D.R.A., Kentish, S.E., Scales, P.J., Martin, G.J.O., 2013. Quantitative evaluation of the ease of rupture of industrially promising microalgae by high pressure homogenization. *Bioresour. Technol.* 140, 165–171. <https://doi.org/10.1016/j.biortech.2013.04.074>

Stafford, J., Uzo, N., Farooq, U., Favero, S., Wang, S., Chen, H.-H., L'Hermitte, A., Petit, C., Matar, O.K., 2021. Real-time monitoring and hydrodynamic scaling of shear exfoliated graphene. *2D Mater.* 8, 025029. <https://doi.org/10.1088/2053-1583/abdf2f>

Sun, W., Mao, S., Shi, Y., Li, L.C., Fang, L., 2011. Nanonization of Itraconazole by High Pressure Homogenization: Stabilizer Optimization and Effect of Particle Size on Oral Absorption. *J. Pharm. Sci.* 100, 3365–3373. <https://doi.org/10.1002/jps.22587>

Taghinia, J., Rahman, M., Tse, T.K.T., Siikonen, T., 2016. CFD modeling of homogenizer valve: A comparative study. *Chem. Eng. Res. Des.* 106, 327–336.

<https://doi.org/10.1016/j.cherd.2015.12.014>

Taghinia, J., Rahman, M.M., Siikonen, T., 2015. Large eddy simulation of a high-pressure homogenizer valve. *Chem. Eng. Sci.* 131, 41–48.

<https://doi.org/10.1016/j.ces.2015.03.041>

Tagne, J.-B., Kakumanu, S., Ortiz, D., Shea, T., Nicolosi, R.J., 2008. A Nanoemulsion Formulation of Tamoxifen Increases Its Efficacy in a Breast Cancer Cell Line. *Mol. Pharm.* 5, 280–286. <https://doi.org/10.1021/mp700091j>

Tam, Y.J., Allaudin, Z.N., Lila, M.A.M., Bahaman, A.R., Tan, J.S., Rezaei, M.A., 2012. Enhanced cell disruption strategy in the release of recombinant hepatitis B surface antigen from *Pichia pastoris* using response surface methodology. *BMC Biotechnol.* 12, 70. <https://doi.org/10.1186/1472-6750-12-70>

Teng, T.S., Chin, Y.L., Chai, K.F., Chen, W.N., 2021. Fermentation for future food systems. *EMBO Rep.* 22. <https://doi.org/10.15252/embr.202152680>

Thomas, P., Duolikun, T., Rumjit, N.P., Moosavi, S., Lai, C.W., Bin Johan, M.R., Fen, L.B., 2020. Comprehensive review on nanocellulose: Recent developments, challenges and future prospects. *J. Mech. Behav. Biomed. Mater.* 110, 103884. <https://doi.org/10.1016/j.jmbbm.2020.103884>

Tölle, F.J., Fabritius, M., Mülhaupt, R., 2012. Emulsifier-Free Graphene Dispersions with High Graphene Content for Printed Electronics and Freestanding Graphene Films. *Adv. Funct. Mater.* 22, 1136–1144. <https://doi.org/10.1002/adfm.201102888>

Tran, L.T.C., Gueutin, C., Frebourg, G., Burucoa, C., Faivre, V., 2017. Erythromycin

encapsulation in nanoemulsion-based delivery systems for treatment of *Helicobacter pylori* infection: Protection and synergy. *Biochem. Biophys. Res. Commun.* 493, 146–151. <https://doi.org/10.1016/j.bbrc.2017.09.060>

Uddin, M.G., Batchelor, W., Allardyce, B.J., Byrne, N., Barrow, C.J., Wang, X., Rajkhowa, R., 2020. Preparing *Bombyx mori* Silk Nanofibers Using a Sustainable and Scalable Approach. *ACS Sustain. Chem. Eng.* 8, 1155–1162. <https://doi.org/10.1021/acssuschemeng.9b06138>

Uluata, S., Decker, E.A., McClements, D.J., 2016. Optimization of Nanoemulsion Fabrication Using Microfluidization: Role of Surfactant Concentration on Formation and Stability. *Food Biophys.* 11, 52–59. <https://doi.org/10.1007/s11483-015-9416-1>

van den Berg, F., Lyndgaard, C.B., Sørensen, K.M., Engelsen, S.B., 2013. Process Analytical Technology in the food industry. *Trends Food Sci. Technol.* 31, 27–35. <https://doi.org/10.1016/j.tifs.2012.04.007>

Van Eerdenbrugh, B., Van den Mooter, G., Augustijns, P., 2008. Top-down production of drug nanocrystals: Nanosuspension stabilization, miniaturization and transformation into solid products. *Int. J. Pharm.* 364, 64–75. <https://doi.org/10.1016/j.ijpharm.2008.07.023>

Vinchhi, P., Patel, J.K., Patel, M.M., 2021. High-Pressure Homogenization Techniques for Nanoparticles, in: *Emerging Technologies for Nanoparticle Manufacturing*. Springer International Publishing, Cham, pp. 263–285. https://doi.org/10.1007/978-3-030-50703-9_11

Vinod, A., Sanjay, M.R., Suchart, S., Jyotishkumar, P., 2020. Renewable and

sustainable biobased materials: An assessment on biofibers, biofilms, biopolymers and biocomposites. *J. Clean. Prod.* 258, 120978.

<https://doi.org/10.1016/j.jclepro.2020.120978>

Wan, J., Zhong, S., Schwarz, P., Chen, B., Rao, J., 2019. Physical properties, antifungal and mycotoxin inhibitory activities of five essential oil nanoemulsions: Impact of oil compositions and processing parameters. *Food Chem.* 291, 199–206.

<https://doi.org/10.1016/j.foodchem.2019.04.032>

Wang, B., Lin, X., Zheng, Y., Zeng, M., Huang, M., Guo, Z., 2021. Effect of homogenization-pressure-assisted enzymatic hydrolysis on the structural and physicochemical properties of lotus-seed starch nanoparticles. *Int. J. Biol. Macromol.* 167, 1579–1586. <https://doi.org/10.1016/j.ijbiomac.2020.11.113>

<https://doi.org/10.1016/j.ijbiomac.2020.11.113>

Wang, F., Lin, W., Ling, Z., Fang, X., 2019. A comprehensive review on phase change material emulsions: Fabrication, characteristics, and heat transfer performance. *Sol. Energy Mater. Sol. Cells* 191, 218–234.

Sol. Energy Mater. Sol. Cells 191, 218–234.

<https://doi.org/10.1016/j.solmat.2018.11.016>

Wang, H., Zuo, M., Ding, N., Yan, G., Zeng, X., Tang, X., Sun, Y., Lei, T., Lin, L., 2019. Preparation of Nanocellulose with High-Pressure Homogenization from Pretreated Biomass with Cooking with Active Oxygen and Solid Alkali. *ACS Sustain. Chem. Eng.* 7, 9378–9386. <https://doi.org/10.1021/acssuschemeng.9b00582>

<https://doi.org/10.1021/acssuschemeng.9b00582>

Wieth, L., Kelemen, K., Braun, S., Koch, R., Bauer, H.-J., Schuchmann, H.P., 2016.

Smoothed Particle Hydrodynamics (SPH) simulation of a high-pressure

homogenization process. *Microfluid. Nanofluidics* 20, 42.

<https://doi.org/10.1007/s10404-016-1705-6>

Wilken, L.R., Nikolov, Z.L., 2012. Recovery and purification of plant-made recombinant proteins. *Biotechnol. Adv.* 30, 419–433.

<https://doi.org/10.1016/j.biotechadv.2011.07.020>

Wu, H., Xiao, D., Lu, J., Jiao, C., Li, Shasha, Lei, Y., Liu, D., Wang, J., Zhang, Z., Liu, Y., Shen, G., Li, Shanshan, 2020. Effect of high-pressure homogenization on microstructure and properties of pomelo peel flour film-forming dispersions and their resultant films. *Food Hydrocoll.* 102, 105628.

<https://doi.org/10.1016/j.foodhyd.2019.105628>

Wu, H., Zhou, W., Liu, Q., Cai, X., Qu, Z., Li, P., Hu, D., Jia, X., 2021. High pressure homogenization of graphene and carbon nanotube for thermal conductive polyethylene composite with a low filler content. *J. Appl. Polym. Sci.* 51838.

<https://doi.org/10.1002/app.51838>

Xu, J., Zhang, J., Wang, Q., Wang, A., 2011. Disaggregation of palygorskite crystal bundles via high-pressure homogenization. *Appl. Clay Sci.* 54, 118–123.

<https://doi.org/10.1016/j.clay.2011.07.020>

Xu, N., Chen, J., Wei, Q., Ding, E., Zeng, X., Xue, F., Zhang, N., Shang, J., 2020. Preparation of polyvinyl alcohol/two-dimensional transition metal dichalcogenides composites by high-pressure homogenization. *J. Appl. Polym. Sci.* 137, 48487.

<https://doi.org/10.1002/app.48487>

Yadav, K.S., Kale, K., 2020. High Pressure Homogenizer in Pharmaceuticals: Understanding Its Critical Processing Parameters and Applications. *J. Pharm.*

Innov. 15, 690–701. <https://doi.org/10.1007/s12247-019-09413-4>

Yang, Q., Xiao, Y., Yin, Y., Li, G., Peng, J., 2019. Erythrocyte Membrane-Camouflaged IR780 and DTX Coloading Polymeric Nanoparticles for Imaging-Guided Cancer Photo-Chemo Combination Therapy. *Mol. Pharm.* 16, 3208–3220. <https://doi.org/10.1021/acs.molpharmaceut.9b00413>

Yang, Y., Marshall-Breton, C., Leser, M.E., Sher, A.A., McClements, D.J., 2012. Fabrication of ultrafine edible emulsions: Comparison of high-energy and low-energy homogenization methods. *Food Hydrocoll.* 29, 398–406. <https://doi.org/10.1016/j.foodhyd.2012.04.009>

Yap, B.H.J., Dumsday, G.J., Scales, P.J., Martin, G.J.O., 2015. Energy evaluation of algal cell disruption by high pressure homogenisation. *Bioresour. Technol.* 184, 280–285. <https://doi.org/10.1016/j.biortech.2014.11.049>

Yong, C., Mei, C., Guan, M., Wu, Q., Han, J., Sun, X., 2018. A comparative study of different nanoclay-reinforced cellulose nanofibril biocomposites with enhanced thermal and mechanical properties. *Compos. Interfaces* 25, 301–315. <https://doi.org/10.1080/09276440.2018.1400271>

Yu, Z., Reid, J.C., Yang, Y.-P., 2013. Utilizing Dynamic Light Scattering as a Process Analytical Technology for Protein Folding and Aggregation Monitoring in Vaccine Manufacturing. *J. Pharm. Sci.* 102, 4284–4290. <https://doi.org/10.1002/jps.23746>

Zahari, N.M., Zawawi, M.H., Sidek, L.M., Mohamad, D., Itam, Z., Ramli, M.Z., Syamsir, A., Abas, A., Rashid, M., 2018. Introduction of discrete phase model (DPM) in fluid flow: A review. p. 020234. <https://doi.org/10.1063/1.5066875>

Zhang, J., Chen, L., Wang, A., Yan, Z., 2020. Dissipative Particle Dynamics Simulation of Ionic Liquid-Based Microemulsion: Quantitative Properties and Emulsification Mechanism. *Ind. Eng. Chem. Res.* 59, 763–773.

<https://doi.org/10.1021/acs.iecr.9b05660>

Zhang, L., Liu, W.-Q., Li, J., 2020. Establishing a Eukaryotic *Pichia pastoris* Cell-Free Protein Synthesis System. *Front. Bioeng. Biotechnol.* 8.

<https://doi.org/10.3389/fbioe.2020.00536>

Zhang, S., Zhang, P., Zhang, G., Fan, J., Zhang, Y., 2012. Enhancement of anaerobic sludge digestion by high-pressure homogenization. *Bioresour. Technol.* 118, 496–501.

<https://doi.org/10.1016/j.biortech.2012.05.089>

Zhang, Y., Shi, R., Xu, Y., Chen, M., Zhang, J., Gao, Q., Li, J., 2020. Developing a stable high-performance soybean meal-based adhesive using a simple high-pressure homogenization technology. *J. Clean. Prod.* 256, 120336.

<https://doi.org/10.1016/j.jclepro.2020.120336>

Zhang, Y., Zhang, P., Ma, B., Wu, H., Zhang, S., Xu, X., 2012. Sewage sludge disintegration by high-pressure homogenization: A sludge disintegration model. *J. Environ. Sci.* 24, 814–820.

[https://doi.org/10.1016/S1001-0742\(11\)60834-6](https://doi.org/10.1016/S1001-0742(11)60834-6)

Zhao, L., Fu, H.-Y., Zhou, W., Hu, W.-S., 2015. Advances in process monitoring tools for cell culture bioprocesses. *Eng. Life Sci.* 15, 459–468.

<https://doi.org/10.1002/elsc.201500006>

Zhou, Y., Fang, Q., Niu, B., Wu, B., Zhao, Y., Quan, G., Pan, X., Wu, C., 2018.

Comparative studies on amphotericin B nanosuspensions prepared by a high

pressure homogenization method and an antisolvent precipitation method. *Colloids Surfaces B Biointerfaces* 172, 372–379.

<https://doi.org/10.1016/j.colsurfb.2018.08.016>

Zollman Thomas, O., Bryant, C., 2021. Don't Have a Cow, Man: Consumer Acceptance of Animal-Free Dairy Products in Five Countries. *Front. Sustain. Food Syst.* 5.

<https://doi.org/10.3389/fsufs.2021.678491>