

From Molecular Descriptors to Intrinsic Fish Toxicity of Chemicals: an Alternative Approach to Chemical Prioritization

Saer Samanipour,^{*,†,‡,¶} Jake W. O'Brien,[¶] Malcolm J. Reid,[§] Kevin V. Thomas,[¶] and Antonia Praetorius^{*,||}

[†]*Van 't Hoff Institute for Molecular Sciences (HIMS), University of Amsterdam (UvA), 1090 GD Amsterdam, the Netherlands*

[‡]*UvA Data Science Center, University of Amsterdam, 1090 GD Amsterdam, the Netherlands*

[¶]*Queensland Alliance for Environmental Health Sciences (QAEHS), The University of Queensland, Brisbane Qld 4072, Australia*

[§]*Norwegian Institute for Water Research (NIVA), NO-0579 Oslo, Norway*

^{||}*Institute for Biodiversity and Ecosystem Dynamics (IBED), University of Amsterdam, 1090 GD Amsterdam, the Netherlands*

E-mail: s.samanipour@uva.nl; a.praetorius@uva.nl

Abstract

The European and US chemical agencies have listed approximately 800k chemicals where knowledge on potential risks to human health and the environment are lacking. Filling these data gaps experimentally is impossible so in-silico approaches and prediction are essential. Many existing models are however limited by assumptions (e.g. linearity and continuity) and small training sets. In this study we present a supervised

7 direct classification model that connects molecular descriptors to toxicity. Categories
8 can be either data-driven (using k-means clustering) or regulatory-defined. This was
9 tested via 907 experimentally defined 96h LC50 values for acute fish toxicity. Our
10 classification model explained $\approx 90\%$ of variance in our data for the training set and \approx
11 80% for the test set. This strategy gave a 5-fold decrease in the incorrect categoriza-
12 tion compared to a QSAR regression model. Our model was subsequently employed to
13 predict the toxicity categories of $\approx 32\text{k}$ chemicals. A comparison between the model-
14 based applicability domain (AD) and the training set AD was performed, suggesting
15 that the training set based AD is a more adequate way to avoid extrapolation when
16 using such models. The better performance of our direct classification model compared
17 to QSAR methods, makes this approach a viable tool for hazard and risk assessment
18 of chemicals.

19 Synopsis

20 In this study an alternative machine learning-based strategy to conventional QSAR models
21 is used for the toxicity categorization of chemicals using molecular descriptors and direct
22 classification.

23 Introduction

24 The chemical space of the human exposome is ever expanding with a wider diversity of chemi-
25 cals from both fate and toxicity points of view.¹⁻⁷ The latest estimates of the environmentally
26 relevant chemicals based on the chemical registries and production volumes are estimated
27 to be between 350k and 800k.^{2,8} For most of these chemicals there is little to no knowledge
28 about their environmental fate nor toxicity.^{1-5,8,9} Since the experimental assessment of the
29 fate and toxicity of such a large number of chemicals is not feasible, modeling approaches to
30 predict hazard indicators play an increasingly important role in chemical prioritization and

31 risk assessment.¹⁰⁻¹³

32

33 Prediction of the physicochemical properties and the biological activity (e.g. aquatic
34 toxicity) has been one of the main approaches to deal with the structural diversity in the
35 chemical space.¹⁰⁻¹³ Most existing modeling strategies employ quantitative structure activ-
36 ity relationship (QSAR) models and rely on building linear and/or non-linear relationships
37 between the structural descriptors and the modeled activity/property.^{10,14-17} These models
38 are often built on very homogeneous training sets (i.e. similar chemical classes), hence the
39 linearity assumption.^{17,18} In fact, recent efforts have been put into using more heterogeneous
40 training sets as well as moving away from the linearity assumption.^{13,14,18,19} Independent
41 from the level of heterogeneity of the training dataset, QSAR models are very limited in the
42 number of measured activities as well as the number of chemicals evaluated (e.g. around
43 1000 chemicals).^{13,14,18,19} The main consequence of this limitation is the fact that the models
44 are used in extrapolation mode when used for prediction. This implies that the new data
45 points are not represented adequately by the chemicals within the training set, thus outside
46 of the model applicability domain. The use of these models for extrapolation may potentially
47 result in very large prediction errors.^{13,19,20}

48

49 For these predicted and measured activities (i.e. toxicity and/or other properties) to
50 be translated into chemical management actions, they are divided into different categories
51 using thresholds based on expert knowledge.^{1,3,21-24} Examples for such categories are environ-
52 mental hazard categories defined by the Globally Harmonized System of Classification and
53 Labelling of Chemicals (GHS) or thresholds for persistence (P), bioaccumulation potential
54 (B) and toxicity (T) defined under the European Registration, Evaluation, Authorization
55 and Restriction of Chemicals (REACH).²⁵ The chemicals that fall within specific categories
56 are then furthered for more active monitoring and eventually for legislation.^{24,26? -28} This
57 process triggers wider experimental evaluation of chemicals within high priority categories,

58 which may result in adjustment of the previously set thresholds, based on the new exper-
59 imental evidence.^{24,26,29} However, for this chemical management strategy to be effective, a
60 more accurate and reliable chemical prioritization (i.e. chemical categorization) approach is
61 warranted.

62

63 In this study we propose an alternative strategy for chemical prioritization on the exam-
64 ple of acute aquatic toxicity, where the QSAR-based activity prediction step is skipped. Our
65 direct classification model directly converts molecular descriptors into chemical categories,
66 avoiding the errors inherent to the activity prediction step. As a proof of concept, this strat-
67 egy was tested with experimentally determined 96h lethal concentration (LC50) values for
68 fish, for 907 organic chemicals. We compared the results of our direct classification strategy
69 with the conventional QSAR approach. Additionally, our modeling strategy was expanded
70 to 32000 chemicals from Norman SusDat.²⁷ Finally, we performed a critical evaluation of
71 applicability domains for all the models in this study.

72 **Methods**

73 **Overall Workflow**

74 The dataset used for our model development, validation, and testing consists of calculated
75 descriptors, monoisotopic mass of each chemical, and experimentally determined LC50 val-
76 ues (96 hours) for acute fish toxicity (see details in Section Dataset). The LC50 values were
77 divided into four categories namely: very low toxicity, low toxicity, moderate toxicity, and
78 high toxicity via k-means clustering. This categorization followed the typical evidence-based
79 effect modeling categorization.³⁰⁻³² Additionally, regulatory-defined toxicity categories were
80 retrieved from the Globally Harmonized System of Classification and Labelling of Chemicals
81 (GHS). We assessed the prediction accuracy of the two types of toxicity categories by em-
82 ploying two different modeling strategies: a conventional QSAR regression model vs direct

83 classification (Figure 1. The QSAR regression model simulated the case where the acute fish
 84 toxicity (as LC50) is predicted based on molecular descriptors via a QSAR model and then
 85 the chemical is assigned a specific toxicity category in a separate step. On the other hand,
 86 the direct classification model skipped the LC50 prediction step and directly classified the
 87 chemical of interest into one of the initially defined toxicity categories. This comparison was
 88 performed for the full dataset (i.e. training set and test set) in order to assess the accuracy
 89 of each approach in acute fish toxicity categorization.

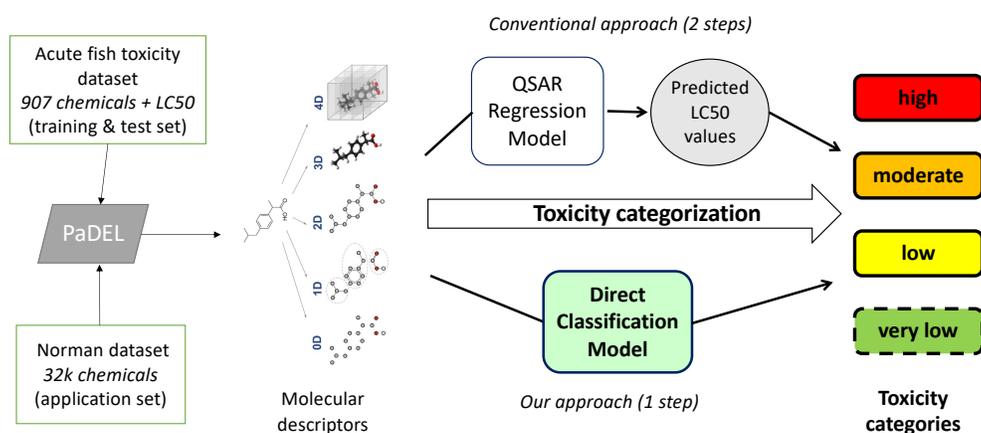


Figure 1: depicts the overall workflow of the study from the raw data to the finally generated models.

90 Datasets

91 We employed two different datasets for our model development¹⁸ and the model applica-
 92 tion.³³ Our modeling dataset consisted of experimental acute fish toxicity values for 907

93 chemicals retrieved from three databases, namely OASIS, ECOTOX and EAT5 and pro-
94 vided by Cassotti et al.¹⁸ The data consisted of the concentrations causing death in 50%
95 of test fathead minnows (*Pimephales promelas*) over a test duration of 96 hours (LC50 96
96 hours). More details regarding the data curation is provided elsewhere.¹⁸ We will refer to
97 this dataset as "acute fish toxicity dataset" here after. The chemicals in this dataset covered
98 different chemical families, including pharmaceuticals, pesticides, conventional persistent or-
99 ganic pollutants (POPs), and industrial chemicals. Throughout this article we refer to the
100 907 chemicals with measured toxicity and curated descriptors as full "acute fish toxicity
101 dataset", the portion used for the model development/validation as training set, and the
102 portion of the data used for additional model testing of the final model as test set.

103

104 The second dataset (hereafter referred to as "Norman dataset") was an extract of around
105 32000 chemicals (31722 chemicals), including their predicted 96h LC50 values for acute fish
106 toxicity (*Pimephales promelas*) from the Norman SusDat database.³⁴ This dataset included
107 only the chemicals that were reported as within the applicability domain of the QSAR model
108 developed by Aalizadeh et al,³⁴ which was used for testing our model applicability, Figure
109 2. This is the model employed by Norman Network for their risk assessment and chemical
110 management. When checking the overlap between the acute fish toxicity dataset and the
111 Norman dataset, we observed around 100 common entries.

112

113 We calculated 2757 1D (i.e. constitutional/count descriptors), 2D (i.e. structural frag-
114 ments), and 3D (i.e. graph invariants) molecular descriptors, and PubChem fingerprints for
115 both datasets using PaDEL software package,³⁵ implemented via a python 3 wrapper called
116 padelpy. Additionally, the name of the chemicals, their SMILES,³⁶ and InChiKeys³⁷ were
117 retrieved from the PubChem database³⁸ via pubchempy API. In order to identify the unsta-
118 ble descriptors—caused by the lack of convergence during the structural optimization—we
119 performed the descriptor calculations for the acute fish toxicity dataset in triplicates. The

120 descriptors were scaled by the maximum of each descriptor in the training set to minimize
121 the impact of the descriptor magnitude on the final models.³⁹ After scaling, the variance of
122 each descriptor in the acute fish toxicity dataset was calculated and only the descriptors that
123 had a variance below 0.1 were kept. We assumed that the stable descriptors for the acute
124 fish toxicity dataset are also stable for the Norman dataset. Therefore, the descriptors for
125 this dataset were calculated only once. Additionally, the maximum of each descriptor in the
126 Norman dataset was compared to those from the training set (from the acute fish toxicity
127 dataset). The descriptors that have this ratio larger than 100 were considered unstable and
128 removed from both datasets, resulting in a total of 2036 final descriptors out of an initial 2780.

129

130 We also evaluated the coverage of the chemical spaces of the datasets by the means of
131 Principal Component Analysis (PCA), Figure 2. The PCA is an unsupervised dimension
132 reduction approach, which enabled us to assess the underlying trends in our datasets by
133 combining several variables into a single principal component.⁴⁰ To perform PCA, we used
134 the curated descriptors matrix and in total two principal components.

135 **Toxicity Categories**

136 To categorize the chemicals based on their acute fish toxicity, we employed two different
137 strategies namely 1) applying k-means clustering to derive four categories from our acute
138 fish toxicity dataset and 2) using predefined categories for acute aquatic hazard as defined
139 in the GHS.⁴¹

140 **K-means Clustering for Toxicity Categorization**

141 The k-means strategy divided the chemicals into four categories consisting of high toxicity,
142 moderate toxicity, low toxicity, and very low toxicity accounting for 96h LC50 values for
143 fish toxicity and monoisotopic mass of the chemicals. The k-means clustering algorithm is
144 an iterative clustering algorithm, where the distances between different measurements from

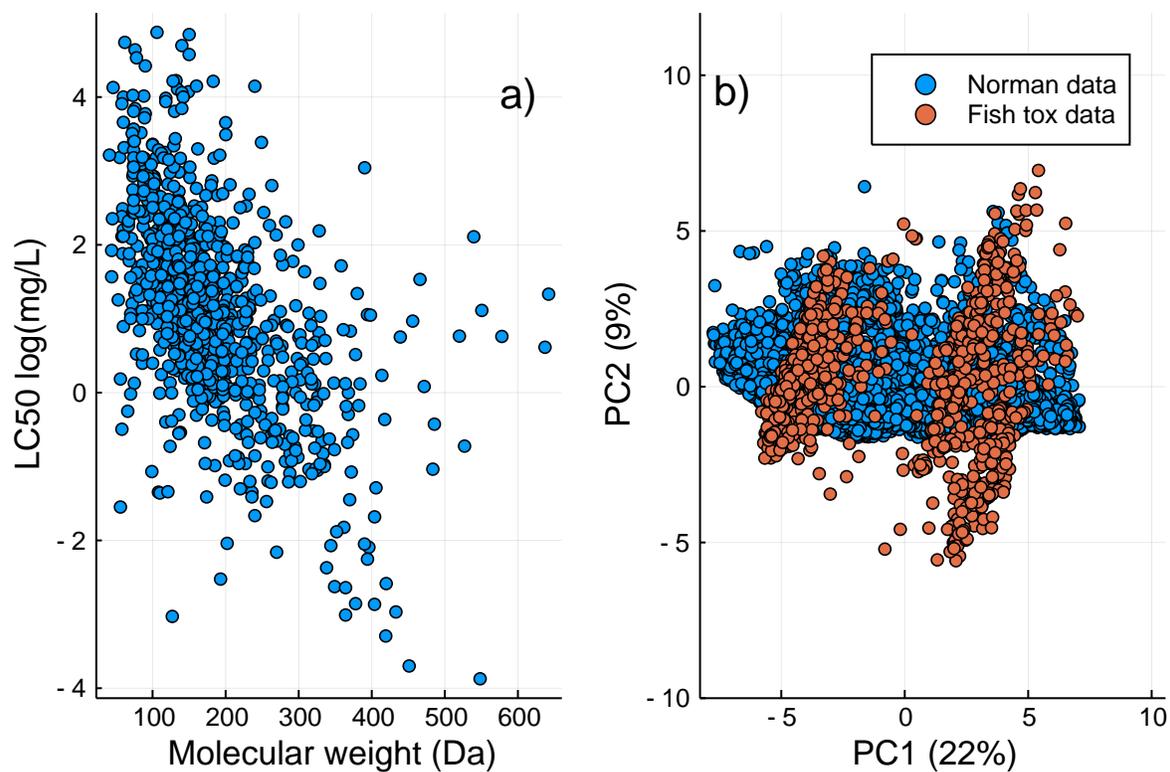


Figure 2: depicts a) the distribution of the experimental LC50 values used for the model development and validation whereas b) shows the chemical space via PCA covered by the acute fish toxicity data (i.e. training and test sets) and the Norman dataset, where the curated descriptors were used for the cluster analysis

145 a set of user defined centers (so called centroides) are used for clustering the data.⁴⁰ This
146 algorithm has the advantage of incorporating more than one parameter, compared to expert
147 manual judgment in the clustering. Additionally, this algorithm, given that it has randomly
148 selected centroides in the first iteration, it requires further validation. Here we employed
149 bootstrapping to assure the selected acute fish toxicity categories (i.e. clusters) are robust
150 enough for predictive purposes. To do that, the fish toxicity data was randomly divided into
151 90% training set and 10% test set. The training set then was bootstrapped with replacement
152 for 500 iterations, to guarantee that each model is built based on a unique dataset. The most
153 commonly identified centroid over 500 iterations was selected as final model and for acute
154 fish toxicity categorization. In the end, the final model was further tested using the test set.
155 During the categorization, we provided the k-means algorithm with two variables namely
156 96h LC50 values and the monoisotopic masses, and the number of clusters of 4, following
157 the category structures adapted by previous studies.³⁰

158 **GHS Categorization for Acute Aquatic Hazards**

159 In addition to the k-mean clustering we also used the three categories for acute aquatic
160 hazards of the GHS, which were hard set thresholds.⁴¹The three GHS-based categories for
161 short-term (acute) aquatic hazard are based on thresholds derived from 96h LC50 values for
162 acute fish toxicity: high toxicity (Category Acute 1: 96h LC50 for fish ≤ 1 mg/L), moderate
163 toxicity (Category Acute 2: $1 \text{ mg/L} < \text{LC50} \leq 10 \text{ mg/L}$), and low toxicity (Category Acute
164 3: $\text{LC50} > 10 \text{ mg/L}$ LC50), Table 4.1.1 in Reference.⁴²

165 **Modeling**

166 In this study, we developed two different models namely: a QSAR regression model and
167 a direct classification model. The details of each model strategy is provided below. Both
168 models, once optimized with the acute fish toxicity dataset, were used with the Norman
169 dataset to further assess their applicability.

170 QSAR Regression Model

171 We developed, optimized, validated, and tested a random forest regression model using the
172 curated descriptors (independent variables) and the experimentally defined LC50 values (de-
173 pendent variable). Random forest is a decision tree based algorithm where several bootstrap
174 data (i.e. training set) are given to several decision trees. This assures that the dataset given
175 to each tree is unique.⁴⁰ Once the model is developed, the most common decision tree model
176 outcome is considered as the random forest model prediction. The main advantage of the
177 random forest modeling strategy is the ability to handle non-linearity and non-continuity
178 in the data, which is highly relevant to toxicity prediction.⁴³ Here, the acute fish toxicity
179 dataset was divided into training set (90% of the full dataset) and test set (10%). The
180 training set was used for the model development and optimization while the test set was
181 for further evaluation of the dataset. For the regression model, the model hyper-parameter
182 optimization was performed with a two dimensional grid with the number of trees ranging
183 from 100 - 1000 whereas the minimum number of points in each leaf varying from 1 - 21.
184 The combination of 3 fold cross-validation and out-of-bag strategy enabled us to generate
185 an optimized regression model while defining the importance of each variable. The variables
186 that had relative levels of importance larger than 1% were considered as essential variables
187 for the model. This strategy enabled us to quickly identify the most relevant variables to
188 our model's accuracy.

189

190 The finally optimized regression model consisted of 600 trees, minimum 4 points in each
191 leaf, and 8 variables. This regression model was employed to predict the 96h LC50 for fish
192 toxicity of the chemicals in the Norman dataset. In a second step, the predicted LC50 values
193 were used to categorize the chemicals into the two types of toxicity categories described
194 above.

195 **Descriptor-Based Direct Classification Model**

196 We developed, validated, and tested a classification model to convert the curated descriptors
197 to the acute fish toxicity categories. For this model, we employed random forest classifica-
198 tion, implemented via ScikitLearn.jl julia package.⁴⁴

199

200 For the direct classification, we split the acute fish toxicity dataset (i.e. curated the de-
201 scriptors and toxicity categories) into training set (90% of the full dataset) and test set (10%).
202 To optimize the main model hyper-parameters, the number of trees, and minimum number
203 of points in each leaf, we generated a grid with 20 steps for each parameter ranging from 200
204 - 2000 and from 1 - 21 for the number of trees and minimum data points in leaf, respectively.
205 For each model, we performed 3 folds of cross-validation to systematically assess the model
206 accuracy. The model with the highest cross validation accuracy (i.e. 73%) was considered
207 as the optimized classification model. This optimized classification model consisted of 1200
208 tress and minimum number of points in each leaf of 4. To avoid overfitting during the train-
209 ing process, when building the model, we set an out-of-bag cross-validation,⁴⁵ where only
210 a randomly selected fraction (i.e. square root of the number of variables) of the variables
211 were fed to individual trees. The combination of out-of-bag cross-validation and leaf purity
212 was utilized to calculate the importance of individual variables on the final model. To select
213 the relevant variables, we divided variance explained by each variable by the largest one and
214 selected those that contributed more than 1% to the model, thus 230 out of 2036 variables.

215

216 To build the final model, the full acute fish toxicity dataset was used with the selected
217 variables. In this case all the selected variables were used for the final model building.
218 Additionally, this model was used to categorize the Norman dataset into the two types of
219 acute fish toxicity categories directly based on the curated descriptors.

220 **Applicability Domain**

221 To assess whether a chemical is well represented by the model training set, we performed
222 the applicability domain (AD) assessment. The AD assessment was done by calculating
223 the leverage of each chemical compared to the training set.³⁴ The leverage was calculated
224 using Eq.1, where X is the matrix of the training set (including the descriptors), x_i is the
225 vector of descriptors for an individual chemical, and the h_{ii} is the calculated leverage. The
226 leverage calculations are typically done only using the model variables, in other words only
227 the descriptors used for the optimized model. In this study we performed both the full
228 descriptor space (i.e. assuming the model using all the descriptors) and the model specific
229 descriptors (i.e. conventional approach). This strategy enabled us to systematically assess
230 which chemicals are well represented by the training set.

$$h_{ii} = x_i^T (X^T X)^{-1} x_i \quad (1)$$

231 **Calculations**

232 All calculations were performed using a personal computer (PC) with Intel Core i7 CPU
233 and 16 GB of RAM operating Ubuntu 20.04.2 LTS. All the data processing and statistical
234 analysis were performed using julia language 1.6.

235 **Results and discussion**

236 In this study, we developed a random forest-based direct classification model to convert the
237 molecular descriptors of chemicals to predefined acute fish toxicity categories. This model
238 was developed, validated, and tested via an experimentally defined dataset of 96h LC50
239 values for acute fish toxicity for 907 organic chemicals. The result of this strategy was directly
240 compared to the conventional two-step approach—first QSAR-based property prediction and
241 then toxicity categorization—both for the acute fish toxicity data and a dataset of ≈ 23000

242 chemicals from Norman SusDat.³³

243 Toxicity Categorization

244 The final k-means model resulted in a clustering accuracy of 97.5%. This model, then, was
245 fed the full acute fish toxicity dataset to define the toxicity category of each chemical in
246 that dataset. The final model was saved as a binary file to be used for prediction (Figure
247 3). The k-means and GHS categories were used as labels in two separate runs of the direct
248 classification model while the 96h LC50 values for acute fish toxicity predicted by the QSAR
249 regression model were converted into the two types of acute toxicity categories in a second
250 step.

251

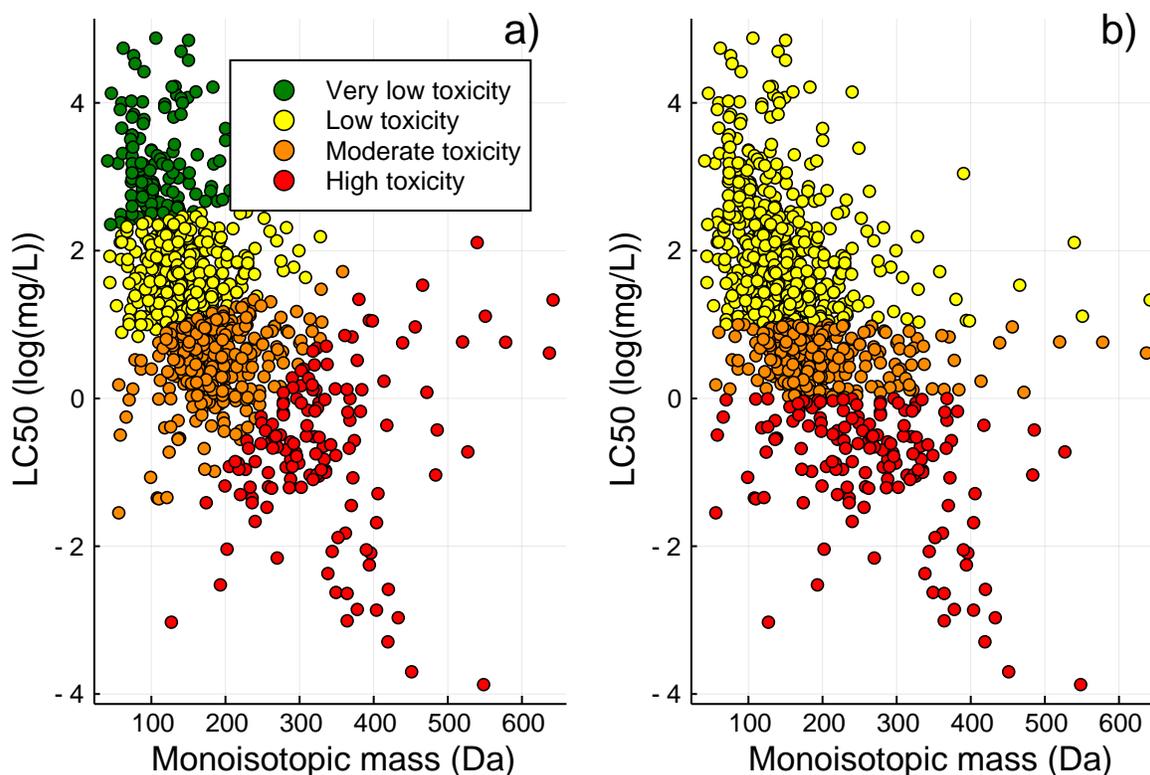


Figure 3: shows the distribution of the toxicity categories of the acute fish toxicity dataset via a) the best k-means clustering model and b) based on GHS categories.

252 When comparing the unsupervised k-means clustering-based categorization with the ex-
253 pert knowledge based categorization from the GHS, we see a high level of similarity in the
254 thresholds (Figure 3). In fact the main differences were observed for chemicals with a molec-
255 ular weight of ≥ 400 Da and LC50 values ≥ 1 mg/L ($0 \log(\text{mg/L})$). These chemicals in the
256 k-means categorization were considered part of the high toxicity category while based on the
257 GHS categories they were considered moderate to low toxicity. When calculating the similar-
258 ity scores between the descriptors of those chemicals and the two categories, we consistently
259 observed higher values for high toxicity category. This indicates that those chemicals may
260 be structurally more similar to the high toxicity category rather than the moderate and/or
261 low one. These similarities are better captured by the k-means model, given that it uses
262 two variables (96h LC50 and monoisotopic mass) and Euclidean distances for the cluster
263 creation.

264 **Performance of QSAR Regression Model**

265 The residuals of the final and optimized QSAR regression model were between -1 and 1 in
266 LC50 units for $\approx 95\%$ of the data (Figure S2). This model consisted of 600 trees and 8 vari-
267 ables, resulting in an R^2 of 0.86 for the training set and ≈ 0.7 for both median cross-validation
268 and test set. The observed levels of accuracy was comparable to previously reported linear
269 and non-linear QSAR models^{17,34} (Figure 4). We observed up to 2.1 $\log(\text{mg/L})$ overesti-
270 mation of the LC50 for values ≤ -1 while our model resulted in a slight underestimation of
271 toxicity for LC50 values ≥ 5 (Figures 4 and S2). Finally, we used the optimized model to
272 predict the 96h acute fish toxicity LC50 values for the Norman dataset. When comparing
273 the results of our predictions to the predictions by Aalizadeh et al,³⁴ a clear linear trend (i.e.
274 Pearson correlation coefficient of 0.68) between the two predictions was observed, further
275 indicating the validity of our model (Figure S3).

276

277 The optimized regression model included 8 variables from which two were related to the

278 logP of the chemicals in the training set (Figure S1). The most relevant variable was the
279 Crippen logP⁴⁶ value explaining around 35% variance of the final model. This logP was
280 calculated based on 68 atomic contributions. On the other hand, the second variable was
281 XLogP,⁴⁷ implemented within PubChem.^{38,48} This logP calculation also uses the atomic
282 contribution of 87 groups and additionally incorporates two correction factors, improving
283 its accuracy and expanding its applicability. Another relevant variable for our regression
284 model was the ZMIC1 descriptor which is a 2D descriptor indicating the level of symmetry
285 in the structure.³⁵ Finally, the remaining relevant descriptors (i.e. excluding logP, XlogP
286 and ZMIC1 descriptors) were related to molecular connectivity, polarizability, and hydrogen-
287 bond donation, which all have shown to be relevant in explaining physico-chemical properties
288 and toxicity of chemicals.^{15,17,34}

289

290 **Performance of Descriptor-Based Direct Classification Model**

291 The optimized direct classification model resulted in a classification accuracy of 92% for
292 the training set and around 80% for both the cross-validation and the test set, for the four
293 k-means categories. The final model used 230 variables out of a total of 2036 curated descrip-
294 tors. Similar to the regression model, most of the important variables were a combination
295 of 2D descriptors and fingerprints (i.e. 3D) (Figure S4). These descriptors included the
296 four logP calculations (e.g. CrippenlogP) as well as parameters related to polarizability and
297 charge distribution. These parameters are all highly relevant to the mobility of the chemicals
298 and their binding potential with the active sites.^{15,18} Differently from the regression model,
299 the most relevant variable only explained $\approx 1.5\%$ of variance (vs 35% for the regression
300 model) in the final model. Even though larger number of variables were included in the
301 model, the total number of variables were less than 30% of the number of measurements re-
302 sulting in a mathematically well-defined problem. Additionally, a larger number of variables
303 enables a better assessment of the model applicability domain.

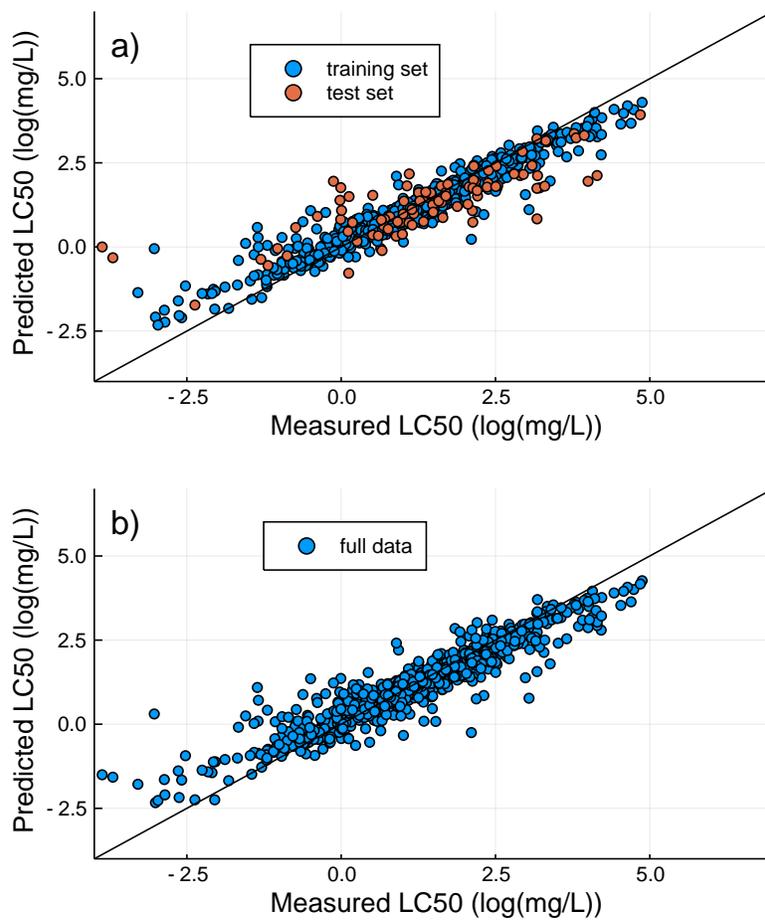


Figure 4: depicts the measured vs predicted 96h LC50 values (in log(mg/L)) for acute fish toxicity for a) the training and test set during the model optimization and b) the optimized model with the full acute fish toxicity dataset.

304

305 The direct classification model based on the three GHS categories, resulted in an accuracy
306 of 94% for the training set and around 85% for the cross-validation and test set. This model,
307 similar to the previous one, had 236 high importance variables that were included in the
308 final model. The high importance variables (e.g. top 20) for both models were exactly the
309 same as for the direct classification into the k-means categories with similar levels of variance
310 explained.

311 The reported statistics and the selected variables in our classification models further
312 indicated the applicability of our model for prediction of acute fish toxicity categories directly
313 from the molecular descriptors.

314 **Classification vs Regression**

315 The fish toxicity data were used for predicting the toxicity categories via both the conven-
316 tional QSAR regression model and the direct classification strategies. The QSAR regression
317 model resulted in predicted LC50 values that were converted to the two types of acute fish
318 toxicity categories in a subsequent step. On the other hand, the classification model directly
319 predicted the toxicity categories. The predicted acute fish toxicity categories based on both
320 methods were compared to the true categories coming from the measured 96h LC50 values
321 for fish toxicity to evaluate the accuracy of each approach.

322

323 The direct classification method, for both cases, resulted in around four times fewer
324 misclassifications when compared to the QSAR regression model. We observed 47 cases of
325 misclassification for the k-means based categories while for GHS categories the misclassified
326 cases were 41. This was in agreement with our expectations, given that the total number
327 of classes in GHS categories were smaller, thus a lower probability of wrong classification.
328 For the QSAR regression model, we observed 178 cases of wrong classifications for k-means
329 based categories whereas 163 incorrectly classified cases were observed for the GHS cate-

330 gories (Figure 5). The direct classification strategy showed a homogeneous distribution of
331 the miscategorized chemicals in the acute fish toxicity dataset, for both the k-means and
332 the GHS categories. For the k-means categorization the QSAR regression model resulted in
333 large and homogenous distribution of wrong categorization while for the GHS approach we
334 observed a high density of miscategorization for high and moderate toxicity groups, (Figure
335 5).

336

337 Around 85% of the miscategorized chemicals via direct classification overlapped with
338 those wrongly categorized via the QSAR regression model, independently of the type of cat-
339 egories. For example a chemical that was consistently wrongly categorized by all the methods
340 was 1-hydroxypyridine-2-thione (InChyKey:YBBJKCMMCRQZMA-UHFFFAOYSA-N) with measured
341 LC50 of 0.95 $\mu\text{g/L}$ (i.e. $-3.02 \log(\text{mg/L})$). This chemical was categorized as moderately toxic
342 by both models while it is actually a high toxicity chemical. When looking at the struc-
343 ture of this chemical, it is clear that this chemical is not very well covered by our training
344 set. In other words, there are not enough (at least 4) chemicals with a similar structure
345 to this one in our training set. This further indicates that the addition of more diverse
346 chemical structures to our training set will result in even more accurate prediction of the
347 toxicity categories. Additionally, the replacement of the molecular descriptors with the to-
348 pographical fingerprints,⁴⁹ given their stability, may further improve our prediction accuracy.

349

350 When comparing the distribution of the wrongly categorized chemicals, we observed a
351 higher levels of homogeneity in the k-means categories compared to the GHS ones. This was
352 consistent for both QSAR regression model and direct classification model. We also observed
353 that for the GHS categories, both the QSAR regression-based and the direct classification
354 model showed a high density of wrong categorization for chemicals at the border between
355 high toxicity and moderate toxicity region. We interpret that this is mainly caused by larger
356 number of categories and lower levels of rigidity in the k-means approach compared to hard

357 set thresholds (i.e. GHS approach).

358

359 The predicted LC50 values using our optimized QSAR regression model followed by the
360 k-means clustering categorization resulted in 81% consistent classification between the acute
361 fish toxicity categories generated by the direct classification method (Figure S5). On the
362 other hand, the predicted LC50 values using the model developed by Aalizadeh et al³⁴ re-
363 sulted in only 37% consistent toxicity categories. This may be due to the fact that our QSAR
364 regression and direct classification models both had the same training set as well as the fact
365 that our QSAR regression model uses 8 descriptors while the model by Aalizadeh et al uses
366 only 6 from which three are logP values.

367

368 Overall, our direct classification strategy showed a better performance in identifying the
369 acute fish toxicity categories of the chemicals directly from the molecular descriptors, rather
370 than passing via a QSAR regression model. We also observed a higher level of consistency
371 between the categories generated by our models compared to another prediction method (i.e.
372 Aalizadeh model). We interpret that the main reason behind the overall better performance
373 of the direct classification approaches is first and foremost the fact the uncertainties asso-
374 ciated with the QSAR regression models do not impact the categorization. Additionally,
375 the inclusion of a larger number of descriptors in such models implies that higher levels of
376 structural features are incorporated. In fact, the low level of variance explained by individ-
377 ual variables further confirms this hypothesis. Our direct classification model can be easily
378 adapted to different types of pre-defined (acute fish toxicity) categories, as demonstrated
379 here by classifying the chemicals following the categories for short-term (acute) aquatic haz-
380 ard of the GHS. Overall, these results indicate the viability of the classification strategy as
381 a means of chemical prioritization and management.

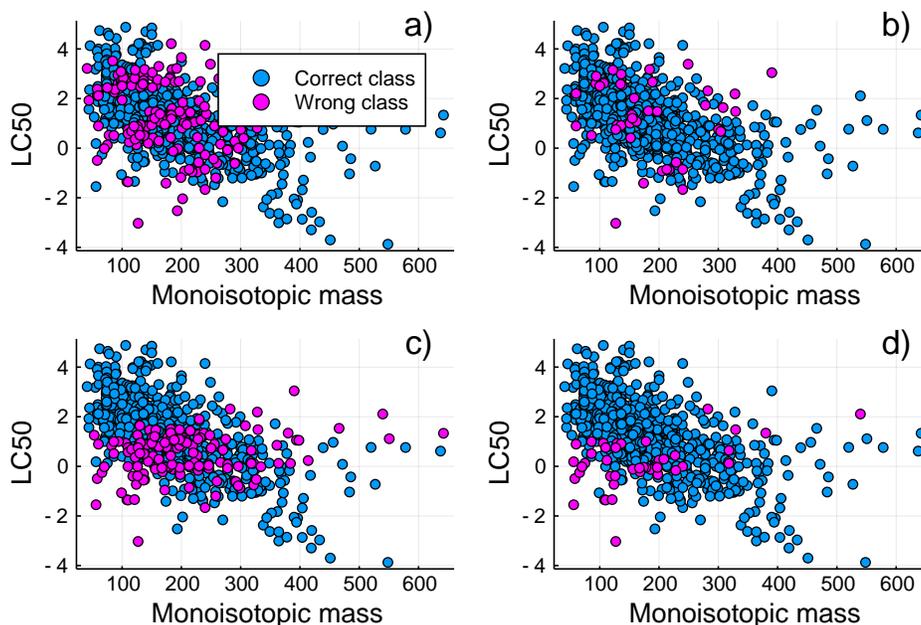


Figure 5: depicts the correctly vs wrongly predicted acute fish toxicity categories based on a) the QSAR regression model and k-means based categories; b) the direct classification strategy based on k-means categories; c) the QSAR regression model using the GHS categories categories; and d) the direct classification strategy with GHS categories.

382 Applicability Domain

383 We also evaluated the impact of the applicability domain AD selection for the assessment
 384 of the model coverage of the chemicals space. To perform such assessment, we calculated
 385 the leverage for full descriptor space, QSAR regression model descriptors, and the direct
 386 classification model descriptors. Figure 6 depicts the scores' plots for the training set and
 387 the Norman dataset and the associated applicability domains.

388

389 With the full descriptor space (i.e. the curated descriptors used for our model devel-
 390 opment), only 585 entries of Norman dataset were covered by the training set. Using the
 391 regression model descriptors (i.e. the 9 most relevant ones) resulted in around 31000 entries
 392 being covered by the training set. On the other hand, based on the descriptors of the direct
 393 classification model around 27000 entries were covered by the chemical space of the train-
 394 ing set. The observed trend is in agreement with our expectations, given that the larger

395 number of descriptors provides a better coverage of different structural characteristics of the
396 chemicals. When looking at the covered chemical space by the training set (i.e. 96h LC50
397 for acute fish toxicity) and the chemicals within the AD of the training set (i.e. the full de-
398 scriptor space) a good level of overlap is observed. This is not the case when looking at the
399 model specific ADs, implying an extrapolation with a much larger level of prediction error.
400 An example of such cases is carbonothioylbis(iminomethylene) bis(diethyldithiocarbamate)
401 (InChyKey: SPQBHESGHZSSMQ-UHFFFAOYSA-N), which was covered by the regression
402 model AD and was not covered by both the classification and the training set AD. In fact,
403 this chemical was one of the most different chemicals compared to the chemicals in the Nor-
404 man dataset (i.e. PC1 -11 and PC2 28). Therefore, it may be advisable to use the training
405 set AD (i.e. the full descriptor space) to assess the training set coverage of the chemical
406 space, rather than the individual model ADs.

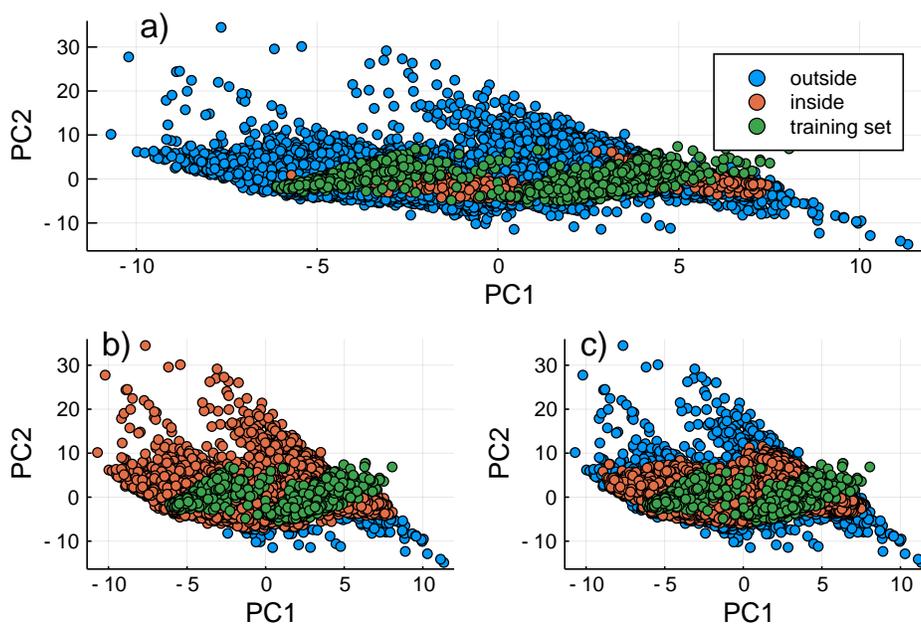


Figure 6: depicts the applicability domain (AD) assessment (i.e. the leverage calculation) of Norman dataset, based on a) the training set (i.e. the full molecular descriptor space), b) the QSAR regression model, and c) the direct classification model. The blue circles represent the chemicals that are outside of the AD while the orange circles are within the model applicability domain and the green circles are within the training set applicability domain.

407 **Implications for Chemical Assessment**

408 The results of our direct classification model showed its power in categorizing the chemicals
409 in terms their acute fish toxicity based on their specific molecular descriptors. Our strategy
410 can overcome the continuity assumption of QSAR models, which are conventionally used
411 to fill experimental data gaps in chemical assessment of structurally similar compounds, di-
412 rectly impacting the size of the training set. In other words, with our direct classification
413 approach the experimental datasets from different sources and for different chemical families
414 can be grouped to generate larger training sets resulting in higher accuracy predictions. As
415 demonstrated here with the direct classification of the chemicals in the Norman dataset into
416 hazard categories defined by the GHS (based on acute fish toxicity), our approach can be
417 adapted to different predefined categories as prescribed by various international regulations
418 and/or classification or labeling systems. The direct classification approach can be expanded
419 to other hazard categories (e.g. chronic toxicity) as well as to fate (e.g. mobility or persis-
420 tence) and shows great potential for improving in-silico tools for chemical hazard and risk
421 assessment.

422 **Code Availability**

423 The open access/source julia package for performing these calculations is available with
424 MIT license using the link here [https://bitbucket.org/SSamanipour/toxcatpred-jl/
425 src/main/](https://bitbucket.org/SSamanipour/toxcatpred-jl/src/main/). Additionally, all the scripts for the model building is available in the same Bit-
426 bucket repository. Finally, the predictions of both models, and all three ADs are available for
427 download and use via FigShare (Fish toxicity: <https://doi.org/10.21942/uva.20089751>,
428 Norman SusDat: <https://doi.org/10.21942/uva.20089787>, and model output: [https:
429 //doi.org/10.21942/uva.20089805](https://doi.org/10.21942/uva.20089805)).

430 padelpy: <https://github.com/ecrl/padelpy>

431 pubchempy API: <https://pubchempy.readthedocs.io/en/latest/>

432 ScikitLearn.jl: <https://scikitlearnjl.readthedocs.io/en/latest/>

433 **Acknowledgement**

434 SS is grateful to the UvA Data Science Center for financial support provided for this project.
435 Moreover, SS is thankful to the members of CAST for the fruitful discussions. JO is the
436 recipient of an NHMRC Emerging Leadership Fellowship (EL1 2009209). The Queensland
437 Alliance for Environmental Health Sciences (QAEHS), The University of Queensland, grate-
438 fully acknowledges the financial support of the Queensland Health and Australian Research
439 Council ARC Discovery Project (DP190102476). The authors thank Joanke van Dijk for
440 help with identifying the regulatory hazard categories.

441 **Supporting Information Available**

442 The Supporting Information containing the details related to the samples, parameter set-
443 tings, and the figures associated with the algorithms are available free of charge at ACS web
444 site.

445 **Author Information**

446 Corresponding Author:

447 Saer Samanipour

448 Van 't hoff institute for molecular sciences (HIMS),

449 University of Amsterdam,

450 the Netherlands

451 Email: s.samanipour@uva.nl

452 **ORCID**

453 Saer Samanipour: 0000-0001-8270-6979

454 Jake W. O'Brien: 0000-0001-9336-9656
455 Malcolm J. Reid: 0000-0002-9988-4867
456 Kevin V. Thomas: 0000-0002-2155-100X
457 Antonia Praetorius: 0000-0003-0197-0116

458 **References**

- 459 (1) Muir, D. C. G.; Howard, P. H. Are There Other Persistent Organic Pollutants? A
460 Challenge for Environmental Chemists. *Environ. Sci. Technol.* **2006**, *40*, 7157–7166.
- 461 (2) Wang, Z.; Walker, G. W.; Muir, D. C.; Nagatani-Yoshida, K. Toward a global under-
462 standing of chemical pollution: a first comprehensive analysis of national and regional
463 chemical inventories. *Environmental science & technology* **2020**, *54*, 2575–2584.
- 464 (3) Howard, P. H.; Muir, D. C. G. Identifying New Persistent and Bioaccumulative Organics
465 Among Chemicals in Commerce. *Environ. Sci. Technol.* **2010**, *44*, 2277–2285.
- 466 (4) Howard, P. H.; Muir, D. C. G. Identifying New Persistent and Bioaccumulative Or-
467 ganics Among Chemicals in Commerce II: Pharmaceuticals. *Environmental Science &*
468 *Technology* **2011**, *45*, 6938–6946.
- 469 (5) Howard, P. H.; Muir, D. C. G. Identifying New Persistent and Bioaccumulative Organics
470 Among Chemicals in Commerce II: Pharmaceuticals. *Environ. Sci. Technol.* **2011**, *45*,
471 6938–6946.
- 472 (6) Escher, B. I.; Stapleton, H. M.; Schymanski, E. L. Tracking complex mixtures of chem-
473 icals in our changing environment. *Science* **2020**, *367*, 388–392.
- 474 (7) Vermeulen, R.; Schymanski, E. L.; Barabási, A.-L.; Miller, G. W. The exposome and
475 health: Where chemistry meets biology. *Science* **2020**, *367*, 392–396.

- 476 (8) Williams, A. J.; Grulke, C. M.; Edwards, J.; McEachran, A. D.; Mansouri, K.;
477 Baker, N. C.; Patlewicz, G.; Shah, I.; Wambaugh, J. F.; Judson, R. S., et al. The
478 CompTox Chemistry Dashboard: a community data resource for environmental chem-
479 istry. *Journal of cheminformatics* **2017**, *9*, 1–27.
- 480 (9) Lai, A.; Clark, A. M.; Escher, B. I.; Fernandez, M.; McEwen, L. R.; Tian, Z.; Wang, Z.;
481 Schymanski, E. L. The Next Frontier of Environmental Unknowns: Substances of Un-
482 known or Variable Composition, Complex Reaction Products, or Biological Materials
483 (UVCBs). *Environmen. Sci. Technol.* **2022**, *56*, 7448.
- 484 (10) Liu, H.; Papa, E.; Gramatica, P. QSAR Prediction of Estrogen Activity for a Large
485 Set of Diverse Chemicals under the Guidance of OECD Principles. *Chem. Res. Toxicol.*
486 **2006**, *19*, 1540–1548.
- 487 (11) Wang, T.; Yuan, X.-s.; Wu, M.-B.; Lin, J.-P.; Yang, L.-R. The advancement of multi-
488 dimensional QSAR for novel drug discovery-where are we headed? *Expert opinion on*
489 *drug discovery* **2017**, *12*, 769–784.
- 490 (12) Sigurnjak Bureš, M.; Cvetnić, M.; Miloloža, M.; Kučić Grgić, D.; Markić, M.; Kušić, H.;
491 Bolanča, T.; Rogošić, M.; Ukić, Š. Modeling the toxicity of pollutants mixtures for risk
492 assessment: a review. *Environmental Chemistry Letters* **2021**, *19*, 1629–1655.
- 493 (13) Mao, J.; Akhtar, J.; Zhang, X.; Sun, L.; Guan, S.; Li, X.; Chen, G.; Liu, J.; Jeon, K.
494 M. S.; Hyeon-Nae; No, K. T.; Wang, G. Comprehensive strategies of machine-learning-
495 based quantitative structure-activity relationship models. *Iscience* **2021**, *24*, 103052.
- 496 (14) Aalizadeh, R.; Ohe, P. C. v. d.; Thomaidis, N. S. Prediction of acute toxicity of emerging
497 contaminants on the water flea *Daphnia magna* by Ant Colony Optimization–Support
498 Vector Machine QSTR models. *Environ. Sci.: Processes Impacts* **2017**, *19*, 438–448.
- 499 (15) Aalizadeh, R.; Nika, M.-C.; Thomaidis, N. S. Development and application of retention

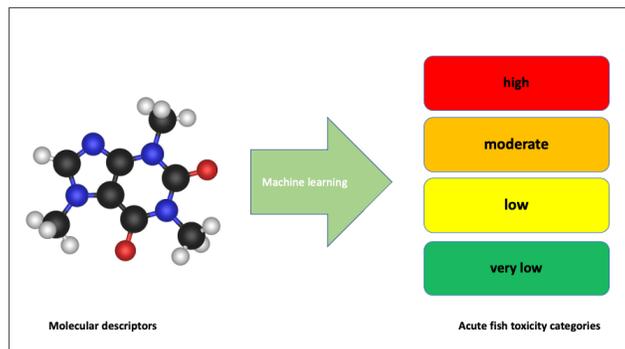
- 500 time prediction models in the suspect and non-target screening of emerging contami-
501 nants. *Journal of Hazardous Materials* **2019**, *363*, 277–285.
- 502 (16) Mansouri, K.; Grulke, C. M.; Judson, R. S.; Williams, A. J. OPERA models for pre-
503 dicting physicochemical properties and environmental fate endpoints. *J Cheminform*
504 **2018**, *10*, 10.
- 505 (17) Ballabio, D.; Consonni, V. Classification tools in chemistry. Part 1: linear models.
506 PLS-DA. *Analytical Methods* **2013**, *5*, 3790.
- 507 (18) Cassotti, M.; Ballabio, D.; Todeschini, R.; Consonni, V. A similarity-based QSAR
508 model for predicting acute toxicity towards the fathead minnow (*Pimephales promelas*).
509 *SAR and QSAR in Environmental Research* **2015**, *26*, 217–243.
- 510 (19) Svetnik, V.; Liaw, A.; Tong, C.; Culberson, J. C.; Sheridan, R. P.; Feuston, B. P.
511 Random forest: a classification and regression tool for compound classification and
512 QSAR modeling. *Journal of chemical information and computer sciences* **2003**, *43*,
513 1947–1958.
- 514 (20) Sheridan, R. P. Using random forest to model the domain applicability of another
515 random forest model. *Journal of chemical information and modeling* **2013**, *53*, 2837–
516 2850.
- 517 (21) Reppas-Chrysovitsinos, E.; Sobek, A.; MacLeod, M. Screening-level exposure-based
518 prioritization to identify potential POPs, vPvBs and planetary boundary threats among
519 Arctic contaminants. *Emerging Contaminants* **2017**, *3*, 85–94.
- 520 (22) Guo, J.; Sinclair, C. J.; Selby, K.; Boxall, A. B. Toxicological and ecotoxicological
521 risk-based prioritization of pharmaceuticals in the natural environment. *Environmental*
522 *toxicology and chemistry* **2016**, *35*, 1550–1559.

- 523 (23) Schulze, S.; Sättler, D.; Neumann, M.; Arp, H. P. H.; Reemtsma, T.; Berger, U. Using
524 REACH registration data to rank the environmental emission potential of persistent
525 and mobile organic chemicals. *Science of the Total Environment* **2018**, *625*, 1122–1128.
- 526 (24) Hale, S. E.; Arp, H. P. H.; Schliebner, I.; Neumann, M. Persistent, mobile and toxic
527 (PMT) and very persistent and very mobile (vPvM) substances pose an equivalent
528 level of concern to persistent, bioaccumulative and toxic (PBT) and very persistent
529 and very bioaccumulative (vPvB) substances under REACH. *Environmental Sciences
530 Europe* **2020**, *32*, 155.
- 531 (25) Williams, E. S.; Panko, J.; Paustenbach, D. J. The European Union’s REACH regula-
532 tion: a review of its history and requirements. *Critical reviews in toxicology* **2009**, *39*,
533 553–575.
- 534 (26) Kwiatkowski, C. F. et al. Scientific Basis for Managing PFAS as a Chemical Class.
535 *Environ. Sci. Technol. Lett.* **2020**, *7*, 532–543.
- 536 (27) Dulio, V. et al. The NORMAN Association and the European Partnership for Chemicals
537 Risk Assessment (PARC): let’s cooperate! *Environmental Sciences Europe* **2020**, *32*,
538 100.
- 539 (28) Rüdell, H.; Körner, W.; Letzel, T.; Neumann, M.; Nödler, K.; Reemtsma, T. Persistent,
540 mobile and toxic substances in the environment: a spotlight on current research and
541 regulatory activities. *Environmental Sciences Europe* **2020**, *32*, 5.
- 542 (29) Dulio, V.; van Bavel, B.; Brorström-Lundén, E.; Harmsen, J.; Hollender, J.;
543 Schlabach, M.; Slobodnik, J.; Thomas, K.; Koschorreck, J. Emerging pollutants in
544 the EU: 10 years of NORMAN in support of environmental policies and regulations.
545 *Environmental Sciences Europe* **2018**, *30*, 5.
- 546 (30) Moe, S. J.; Madsen, A. L.; Connors, K. A.; Rawlings, J. M.; Belanger, S. E.; Lan-
547 dis, W. G.; Wolf, R.; Lillicrap, A. D. Development of a hybrid Bayesian network model

- 548 for predicting acute fish toxicity using multiple lines of evidence. *Environmental Mod-*
549 *elling & Software* **2020**, *126*, 104655.
- 550 (31) Linkov, I.; Massey, O.; Keisler, J.; Rusyn, I.; Hartung, T. From” weight of evidence” to
551 quantitative data integration using multicriteria decision analysis and Bayesian meth-
552 ods. *Altex* **2015**, *32*, 3.
- 553 (32) Kjaerulff, U. B.; Madsen, A. L. Bayesian networks and influence diagrams. *Springer*
554 *Science+ Business Media* **2008**, *200*, 114.
- 555 (33) Schymanski, E. Update on NORMAN-SusDat NORMAN-SLE (Suspect List Ex-
556 change). **2021**,
- 557 (34) Aalizadeh, R.; Peter, C.; Thomaidis, N. S. Prediction of acute toxicity of emerging
558 contaminants on the water flea *Daphnia magna* by Ant Colony Optimization–Support
559 Vector Machine QSTR models. *Environmental Science: Processes & Impacts* **2017**, *19*,
560 438–448.
- 561 (35) Yap, C. W. PaDEL-descriptor: An open source software to calculate molecular descrip-
562 tors and fingerprints. *Journal of computational chemistry* **2011**, *32*, 1466–1474.
- 563 (36) Weininger, D. SMILES, a chemical language and information system. 1. Introduction
564 to methodology and encoding rules. *Journal of chemical information and computer*
565 *sciences* **1988**, *28*, 31–36.
- 566 (37) Heller, S. R.; McNaught, A. D. The IUPAC international chemical identifier (InChI).
567 *Chemistry International* **2009**, *31*, 7.
- 568 (38) Kim, S.; Thiessen, P. A.; Bolton, E. E.; Chen, J.; Fu, G.; Gindulyte, A.; Han, L.; He, J.;
569 He, S.; Shoemaker, B. A., et al. PubChem substance and compound databases. *Nucleic*
570 *acids research* **2016**, *44*, D1202–D1213.

- 571 (39) van den Berg, R. A.; Hoefsloot, H. C.; Westerhuis, J. A.; Smilde, A. K.; van der
572 Werf, M. J. Centering, scaling, and transformations: improving the biological informa-
573 tion content of metabolomics data. *BMC genomics* **2006**, *7*, 1–15.
- 574 (40) Hastie, T.; Tibshirani, R.; Friedman, J. H.; Friedman, J. H. *The elements of statistical*
575 *learning: data mining, inference, and prediction*; Springer, 2009; Vol. 2; p 587.
- 576 (41) Miyagawa, M. Globally harmonized system of classification and labelling of chemicals
577 (GHS) and its implementation in Japan. *Nihon Eiseigaku zasshi. Japanese Journal of*
578 *Hygiene* **2010**, *65*, 5–13.
- 579 (42) Globally Harmonized System of Classification and Labelling of Chemicals (GHS
580 Rev. 9, 2021) | UNECE. [https://unece.org/transport/standards/transport/](https://unece.org/transport/standards/transport/dangerous-goods/ghs-rev9-2021)
581 [dangerous-goods/ghs-rev9-2021](https://unece.org/transport/standards/transport/dangerous-goods/ghs-rev9-2021).
- 582 (43) Cassotti, M.; Ballabio, D.; Consonni, V.; Mauri, A.; Tetko, I. V.; Todeschini, R. Pre-
583 diction of acute aquatic toxicity toward daphnia magna by using the ga-k nn method.
584 *Alternatives to Laboratory Animals* **2014**, *42*, 31–41.
- 585 (44) Pedregosa, F.; Varoquaux, G.; Gramfort, A.; Michel, V.; Thirion, B.; Grisel, O.; Blon-
586 del, M.; Prettenhofer, P.; Weiss, R.; Dubourg, V., et al. Scikit-learn: Machine learning
587 in Python. *the Journal of machine Learning research* **2011**, *12*, 2825–2830.
- 588 (45) Cho, G.; Jung, K.; Hwang, H. Out-of-bag prediction error: A cross validation index
589 for generalized structured component analysis. *Multivariate Behavioral Research* **2019**,
590 *54*, 505–513.
- 591 (46) Wildman, S. A.; Crippen, G. M. Prediction of physicochemical parameters by atomic
592 contributions. *Journal of chemical information and computer sciences* **1999**, *39*, 868–
593 873.

- 594 (47) Cheng, T.; Zhao, Y.; Li, X.; Lin, F.; Xu, Y.; Zhang, X.; Li, Y.; Wang, R.; Lai, L.
595 Computation of octanol- water partition coefficients by guiding an additive model with
596 knowledge. *Journal of chemical information and modeling* **2007**, *47*, 2140–2148.
- 597 (48) Kim, S.; Chen, J.; Cheng, T.; Gindulyte, A.; He, J.; He, S.; Li, Q.; Shoemaker, B. A.;
598 Thiessen, P. A.; Yu, B.; Zaslavsky, L.; Zhang, J.; Bolton, E. E. PubChem 2019 update:
599 improved access to chemical data. *Nucleic Acids Research* **2019**, *47*, D1102–D1109.
- 600 (49) Rogers, D.; Hahn, M. Extended-connectivity fingerprints. *J. Chem. Inf. Model.* **2010**,
601 *50*, 742–754.



Review only.