A Sulfamate-Tethered *Aza*-Wacker Cyclization Strategy for the Syntheses of 2-Amino-2-Deoxyhexoses: Preparation of Orthogonally Protected D-Galactosamines

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A New Strategy for Orthogonally Protected D-Galactosamines

Abstract

We present a new strategy for the assembly of protected D-galactosamine synthons. Our route uses a sulfamate-tethered *aza*-Wacker cyclization as a key step and commences from D-erythrono-1,4-lactone. This stands in contrast to most literature syntheses of 2-amino-2-deoxyhexose derivatives, as these generally employ glycals or hexoses as starting materials. This strategy may serve as a template for the assembly of many other 2-amino-2-deoxyhexoses with protection patterns difficult to access by conventional methods.



Figure 1. D-galactosamine and N-acetyl-D-galactosamine are available from hydrolysis of chondroitin sulfates, but orthogonally protected derivatives require *de novo* syntheses.

D-galactosamine (**Figure 1**) is one of the 2-amino-2-deoxyhexose monosaccharides, which are important components of polysaccharides and glycoproteins.¹ D-galactosamine itself is biologically active and is a known hepatotoxin; for this reason, D-galactosamine is used as a reagent to induce hepatitis in animal models.² N-acetyl-D-galactosamine is an essential component of chondroitin sulfate, a glycosaminoglycan which is abundant in cartilage.³ D-galactosamine derivatives are also found in blood group antigens, antifreeze glycoproteins, and glycosphingolipids.^{1,4}

D-galactosamine and N-acetyl-D-galactosamine are obtainable from hydrolysis of chondroitin sulfates,⁵ but differentially protected derivatives require total syntheses. To date, almost all orthogonally protected D-galactosamines have been prepared using "heterocycle → heterocycle" approaches (Scheme 1A) with either glycals⁶⁻⁸ or other hexoses⁹⁻¹⁴ as starting materials.¹⁵ Diverse alkene functionalization reactions have been employed for the transformation of glycals into D-galactosamine derivatives.¹⁶⁻²³ A variety of nucleophilic substitution protocols have been developed for the conversion of D-glucosamines into D-galactosamines.²⁴⁻³⁰ Kulkarni and Emmadi have employed D-mannose as a starting material for D-galactosamine thioglycosides.^{31, 32}

We envisioned an alternate strategy for the preparation of an orthogonally protected D-galactosamine (**Scheme 1B**). Our laboratory has a programmatic focus on the development of tethered *aza*-

Wacker technology for complex molecule synthesis.³³⁻³⁶ In general, both classical³⁷⁻³⁹ and tethered aza-Wacker cyclization reactions^{40, 41} have been underemployed as key steps in total syntheses. We thus imagined preparing new, orthogonally-protected D-galactosamines using our laboratory's sulfamate-tethered *aza*-Wacker cyclization as a key step. Success with such a synthesis would allow access to interesting D-galactosamine synthons, but, more importantly, could represent a unique and potentially general strategy for the assembly of a variety of 2-amino-2-deoxyhexose monosaccharides.

<u>A. Prior art</u>: Heterocycle \rightarrow Heterocycle Approach

OH

"Glycal → Aza-Pyranose" Strategy Lemieux and Ratcliffe, 1978

OAc AcO AcO

"Hexose → Hexose" Strategy



OBn Scheme 1. (A) Historically, syntheses of D-galactosamine and protected variants have commenced from glycals or other pyranoses (select examples shown). (B) Our approach uses a different chiron and a sulfamate-tethered *aza*-Wacker cyclization strategy.

O

HN

OН



Scheme 2. Retrosynthetic analysis incorporates a sulfamate-tethered *aza*-Wacker cyclization as a key step.

Our retrosynthetic analysis of an orthogonally protected D-galactosamine is shown in **Scheme 2**. Late-stage hemiacetalization of **B** would form target **A**. Aldehyde **B** would be synthesized from intermediate **C** by oxidative alkene cleavage and oxathiazinane ring-opening. Sulfamate **D**, obtained from sulfamoylation of **E**, would be converted into oxathiazinane **C** using our laboratory's sulfamate-tethered *aza*-Wacker cyclization reaction. We envisioned that **E** would be accessible from intermediate **F** using a Grignard addition/ketone reduction sequence.



Scheme 3. Opening Sequence of Reactions.

Our synthesis commenced with ring-opening of D-erythrono-1,4-lactone (1) with morpholine followed by ketalization with 2,2-dimethoxypropane and 10-CSA (Scheme 3).⁴² Protection of the secondary alcohol into its corresponding TBS ether proceeded smoothly with TBSOTf/2,6-lutidine to furnish 3. Subsequent addition of propenyl magnesium bromide (commercially available as a *cis/trans* mixture or freshly prepared from 1-bromo-1-propene and magnesium) formed enone 4. While the reaction did not go to completion even with 3 equivalents of the Grignard reagent, the overall mass balance was satisfactory, and starting material 3 could be recovered and recycled. Luche reduction with NaBH₄/CeCl₃•7H₂O formed allylic alcohol 5 with excellent chemoselectivity and diastereocontrol. The allylic alcohol was transformed into its corresponding benzoate ester 6 using benzoyl chloride/TMEDA. The TBS group was cleaved with TBAF/THF, and the resulting alcohol was sulfamoylated with CISO₂NH₂ (prepared *in situ* by addition of HCO₂H to CISO₂NCO⁴³) in a mixture of CH₃CN/DMA.



 $\label{eq:scheme 4.} Scheme 4. Attempted sulfamate-tethered aza-Wacker cyclization with an allylic benzoate present fails, presumably because of a Pd(II)-catalyzed [3,3]-sigmatropic rearrangement.$

Sulfamate **8** was then subjected to one of the protocols that our laboratory has previously developed for *aza*-Wacker cyclizations³⁵ (**Scheme 4**). When **8** was heated to 55 °C with a mixture of $Pd(OAc)_2$ and $Cu(OAc)_2$ under 1 atm of O_2 in CH₃CN, the mass balance of the reaction was poor, and no desired product was observed. Several investigators have established that Pd(II) salts catalyze efficient [3,3]-sigmatropic rearrangements of allylic esters.⁴⁴⁻⁴⁸ We know from our own experience that allylic sulfamates are generally very unstable; thus, we hypothesized that a Pd(II)-catalyzed [3,3]-sigmatropic rearrangement was a likely pathway leading to unproductive consumption of **8**.



Scheme 5. Optimization of the sulfamate-tethered aza-Wacker cyclization.

The benzoate ester was hydrolyzed using K₂CO₃/MeOH to form **9** (Scheme **5**). We were pleased to see that when **9** was heated with Pd(OAc)₂ (15 mol%) and 1 equivalent of Cu(OAc)₂ under 1 atm of O₂ in CH₃CN, cyclized product **10** formed in a 32% yield (Scheme **5**, Entry **1**). A crystal structure of **10** allowed us to assign product identity and stereochemistry unambiguously (CCDC **2204976**). The reaction yield was similar with 15 mol% of Pd₂(dba)₃ (Scheme **5**, Entry **2**), but we observed a much better result upon switching to PdCl₂(nbd) and Pd(PhCN)₂Cl₂ (Scheme **5**, Entries **3**-**4**). Switching solvents from CH₃CN to dioxane or to DMA (Scheme **5**, Entries **5**-**6**) was deleterious, but reaction performance was reasonable in DMSO (Scheme **5**, Entry **7**). The Yu laboratory has shown that mono-protected amino acid (MPAA) ligands are excellent in promoting Pd(II) – Pd(0) catalytic cycles in C–H functionalization reactions.⁴⁹⁻⁵¹ *Aza*-Wacker cyclization reactions also proceed *via* a Pd(II) – Pd(0) redox manifold, and we thus hypothesized that an MPAA ligand may boost reaction performance. In line with this idea, we were pleased to see a 15% boost in yield when 1 equivalent of Fmoc-Gly-OH and 4 Å molecular sieves were added to the reaction mixture (Scheme **5**, Entry **9**).



Scheme 6. Completion of orthogonally protected D-galactosamines.

Moving forward, the secondary alcohol was transformed into the corresponding TBS ether using TBSOTf/2,6-lutidine (**Scheme 6**). We³⁴ and others^{43, 52, 53} have established that activated oxathiazinane heterocycles are excellent synthons for ring-opening with a variety of nucleophiles. Accordingly, a Cbz group was appended to oxathiazinane **11** using benzyl chloroformate and TMEDA, and ring opening was effected by heating **12** with KOAc in DMSO to form linear intermediate **13**. Many biologically active natural products contain carbamoylated hexose components.⁵⁴ Thus, in our D-galactosamine synthons, we planned that one of the OH protecting groups would be a carbamate. Accordingly, the acetate of **13** was hydrolyzed using K₂CO₃ in MeOH, and alcohol **14** was converted into benzyl carbamate **15** by heating with benzyl isocyanate in toluene. Dihydroxylation of **15** proceeded smoothly with 10 mol% K₂OsO₄•2H₂O and stoichiometric NMO. Cleavage with NaIO₄ formed an aldehyde which was unstable to purification and thus was immediately subjected to hemiacetalization with TsOH. Based on the coupling constant of the anomeric proton (~2 Hz), we conclude that cyclization selectively furnished the alpha anomer. The free alcohols of pyranose **17** were converted into their benzoate esters using benzoyl chloride and TMEDA. The anomeric

benzoate could be selectively deprotected using MeNH₂ in THF.⁵⁵ Overall, **17**, **18**, and **19** represent new D-galactosamine synthons.

In summary, we present a new strategy for the assembly of protected D-galactosamine molecules, which we envision may be employed as synthetic intermediates for various applications. Our route uses a sulfamate-tethered *aza*-Wacker cyclization as a key step and commences from D-erythrono-1,4-lactone. This stands in contrast to most literature syntheses of 2-amino-2-deoxyhexose derivatives, which generally employ glycals or hexoses as starting materials. While we have focused specifically on D-galactosamine syntheses, we hope that this strategy may be employed in the syntheses of many other 2-amino-2-deoxyhexoses.

Experimental Section

General Considerations: All reagents were obtained commercially unless otherwise noted. Solvents were purified by passage under 10 psi N₂ through activated alumina columns. Infrared (IR) spectra were recorded on a Thermo ScientificTM NicoletTM iS^{TM5} FT-IR Spectrometer; data are reported in frequency of absorption (cm⁻¹). ¹H NMR spectra were recorded at 400, 500, or 600 MHz. Data are recorded as: chemical shift in ppm referenced internally using residual solvent peaks, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet or overlap of nonequivalent resonances), integration, coupling constant (Hz). ¹³C NMR spectra were recorded at 101 MHz or at 126 MHz. Exact mass spectra were recorded using an electrospray ion source (ESI) either in positive mode or negative mode and with a time-of-flight (TOF) analyzer on a Waters LCT PremierTM mass spectrometer and are given in m/z. TLC was performed on pre-coated glass plates (Merck) and visualized either with a UV lamp (254 nm) or by dipping into a solution of KMnO₄–K₂CO₃ in water followed by heating. Flash chromatography was performed on silica gel (230-400 mesh) or Florisil (60-100 mesh).



A 250 mL round-bottom flask was charged with a stir bar, **1** (10 g, 84.7 mmol, 1 equiv.), and MeOH (100 mL). Subsequently, morpholine (8.0 mL, 8.08 g, 92.8 mmol, 1.1 equiv.) was added at room temperature, and the reaction mixture was stirred for 14 hours. Following this time, the reaction was concentrated under reduced pressure using toluene as an azeotrope (50 mL x 3) to remove excess morpholine. Then, CH_2Cl_2 (100 mL) and acetone (25 mL) were added to the crude product, and the mixture was cooled to 0 °C using an ice-water bath. 2,2-dimethoxypropane (20.2 mL, 17.2 g, 165 mmol, 2 equiv.) and camphorsulfonic acid (6.6 g, 28 mmol, 0.33 equiv.) were added sequentially. The reaction mixture was warmed to room temperature over a period of 3 hours. Following this time, the reaction was quenched by addition of Et₃N (8.0 mL) and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 50% EtOAc/hexanes on silica gel to yield **2** (white solid, 18.2 g, 74.2 mmol, 88% yield).



(R)-2-((R)-2,2-dimethyl-1,3-dioxolan-4-yl)-2-hydroxy-1-morpholinoethan-1-one

Compound 2

¹H NMR (400 MHz, CDCl₃) δ 4.31 (d, J = 7.9 Hz, 1H), 4.19 (dd, J = 8.5, 6.1 Hz, 1H), 4.06 – 3.95 (m, 2H), 3.95 – 3.88 (m, 1H), 3.80 – 3.61 (m, 5H), 3.53 – 3.41 (m, 2H), 1.41 (s, 3H), 1.32 (s, 3H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ 171.1, 110.0, 77.5, 69.0, 67.8, 66.8, 66.7, 46.2, 43.2, 26.3, 24.9.

IR (v_{max}) 3500, 2986, 1644, 1373, 1270, 1067 cm⁻¹.

 $[\alpha]_{D}^{23.7} = -36.42$ (*c* 0.98, CH₃Cl).

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{11}H_{19}NO_5Na^+$ 268.1155 Found 268.1137 (6.7 ppm error).



A 250 mL round-bottom flask was charged with a stir bar, **2** (11.4 g, 46.5 mmol, 1 equiv.) and CH₂Cl₂ (80 mL). The reaction flask was cooled to 0 °C using an ice-water bath. 2,6-lutidine (16.3 mL, 15.1 g, 140 mmol, 3 equiv.) and TBSOTf (21.4 mL, 24.6 g, 93.0 mmol, 2 equiv.) were sequentially added dropwise. The reaction mixture was stirred at 0 °C for 30 minutes. The reaction was then quenched by addition of a saturated aqueous solution of NaHCO₃ (100 mL), and the mixture was transferred to a separatory funnel. The aqueous layer was extracted with CH₂Cl₂ (2 x 80 mL). The organic layers were collected, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 20% EtOAc/hexanes on silica gel to yield **3** (colorless oil, 15.7 g, 43.7 mmol, 94% yield).



(R)-2-((tert-butyldimethylsilyl)oxy)-2-((R)-2,2-dimethyl-1,3-dioxolan-4-yl)-1-morpholinoethan-1-one

Compound 3

¹H NMR (400 MHz, CDCl₃) δ 4.38 (d, *J* = 7.0 Hz, 1H), 4.21 (dt, *J* = 7.0, 6.0 Hz, 1H), 4.06 (dd, *J* = 8.6, 6.2 Hz, 1H), 3.90 (dd, *J* = 8.6, 5.9 Hz, 1H), 3.80 – 3.50 (m, 8H), 1.39 (s, 3H), 1.30 (s, 3H), 0.86 (s, 9H), 0.06 (s, 3H), 0.05 (s, 3H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ 169.4, 109.9, 77.3, 73.6, 67.1, 67.0, 66.8, 46.1, 42.7, 26.6, 25.7, 25.2, 18.2, -4.4, -5.1.

IR (v_{max}) 2929, 1658, 1253, 1116 cm⁻¹.

 $[\alpha]_{D}^{24.7} = +7.75$ (*c* 0.92, CH₃Cl).

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{17}H_{33}NO_5SiNa^+$ 382.2020 Found 382.1993 (7.1 ppm error).



Note: Commercially available 1-propenylmagnesium bromide solution (sold as a *cis/trans* mixture) can also be used for this reaction and gives a similar yield. Subsequent steps are also not affected by using products which are mixtures of geometric isomers.

Preparation of *trans*-1-propenyl-magnesium bromide solution (This procedure can also be used for preparation of *cis*-1-propenyl-magnesium bromide solution): A 500 mL two-neck oven dried round-bottom flask was charged with a stir bar, Mg turnings (2.88 g, 118 mmol, 1.2 equiv.), I_2 (0.254 g, 1 mmol, 0.01 equiv.), and THF (200 mL). A reflux condenser was attached to one neck of the flask. *Trans*-1-bromo-1-propene (8.5 mL, 12.0 g, 99 mmol, 1 equiv.) was added to the flask dropwise at room temperature, and the heterogeneous mixture was heated to 60 °C for 3 hours using an oil bath. Following this time, the reaction was cooled to room temperature and stirred for an additional 12 hours.

Note: The following reaction was performed in two separate batches of 6 g each. The yield is reported for both batches combined.

A 250 mL round-bottom flask was charged with a stir bar, compound **3** (6 g, 16.7 mmol, 1 equiv.), THF (40 mL), and cooled to 0 °C using an ice-water bath. Subsequently, the freshly prepared Grignard solution (~0.5 M concentration, 100 mL, ~50 mmol, ~3 equiv.) was added dropwise. The reaction mixture was warmed to room temperature over a period of 7 hours. Following this time, the reaction was cooled to 0 °C using an ice-water bath and quenched by slow addition of a saturated aqueous solution of NH₄Cl (100 mL). The mixture was transferred to a separatory funnel and further diluted with water (100 mL). The aqueous layer was extracted with EtOAc (2 x 100 mL). The organic layers were collected, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 10% EtOAc/hexanes on silica gel to yield **4**. Over two batches: (colorless oil, 5.7 g, 18.1 mmol, 54% yield) and recovered starting material: 3 g (25%).



(R,E)-1-((tert-butyldimethylsilyl)oxy)-1-((R)-2,2-dimethyl-1,3-dioxolan-4-yl)pent-3-en-2-one

Compound 4

¹H NMR (400 MHz, CDCl₃) δ 7.03 (dq, *J* = 15.6, 6.9 Hz, 1H), 6.53 (dq, *J* = 15.6, 1.7 Hz, 1H), 4.24 – 4.15 (m, 2H), 3.94 (qdd, *J* = 8.3, 4.3, 1.5 Hz, 2H), 1.92 (dd, *J* = 6.9, 1.7 Hz, 3H), 1.42 (s, 3H), 1.32 (s, 3H), 0.91 (s, 9H), 0.08 (s, 3H), 0.03 (s, 3H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ 198.7, 144.8, 126.9, 109.8, 78.3, 77.3, 65.8, 26.6, 25.8, 25.5, 18.6, 18.3, -4.6, -4.7.

IR (v_{max}) 2931, 1692, 1632, 1253, 1076 cm⁻¹.

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{16}H_{30}O_4SiNa^+$ 337.1806 Found 337.1811 (1.5 ppm error).



A 250 mL round-bottom flask was charged with a stir bar, **4** (5.6 g, 17.8 mmol, 1 equiv.), and MeOH (40 mL). The reaction flask was cooled to -10 °C using a NaCl/ice-water bath. CeCl₃•7H₂O (7.3 g, 19.6 mmol, 1.1 equiv.) was added and, after 15 minutes, NaBH₄ (0.732 g, 19.3 mmol, 1.1 equiv.) was added portionwise. The reaction mixture was stirred at -10 °C for 30 minutes. Following this time, the reaction was quenched with a saturated aqueous solution of NH₄Cl (50 mL), and the MeOH was evaporated under reduced pressure. The aqueous mixture was then transferred to a separatory funnel and extracted with EtOAc (2 x 50 mL). The organic layers were combined, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 20% EtOAc/hexanes on silica gel to yield **5** (colorless oil, 4.4 g, 13.9 mmol, 78% yield).



(1S,2R)-1-((tert-butyldimethylsilyl)oxy)-1-((R)-2,2-dimethyl-1,3-dioxolan-4-yl)pent-3-en-2-ol

Compound 5

¹H NMR (400 MHz, CDCl₃) δ 5.81 – 5.69 (m, 1H), 5.53 (ddq, *J* = 15.3, 5.4, 1.6 Hz, 1H), 4.17 – 4.04 (m, 2H), 4.01 (dd, *J* = 8.1, 6.2 Hz, 1H), 3.81 (dd, *J* = 8.1, 6.9 Hz, 1H), 3.75 (dd, *J* = 6.2, 3.3 Hz, 1H), 1.73 (dt, *J* = 6.5, 1.4 Hz, 3H), 1.42 (s, 3H), 1.34 (s, 3H), 0.89 (s, 9H), 0.11 (s, 3H), 0.08 (s, 3H).

¹³C{¹H} NMR (126 MHz, CDCl₃) δ 130.9, 127.5, 109.1, 76.8, 75.0, 73.7, 66.8, 26.9, 26.1, 25.6, 18.4, 18.1, -3.9, -4.0.

IR (v_{max}) 3500, 2931, 1473, 1253 cm⁻¹.

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{16}H_{32}O_4SiNa^+$ 339.1962 Found 339.1985 (6.8 ppm error).



A 100 mL round-bottom flask was charged with a stir bar, **5** (4.3 g, 13.6 mmol, 1 equiv.), and CH₂Cl₂ (30 mL). The reaction flask was cooled to 0 °C using an ice-water bath. TMEDA (2.0 mL, 1.55 g, 13.3 mmol, 1 equiv.) was added followed by dropwise addition of benzoyl chloride (2.6 mL, 3.15 g, 22.4 mmol, 1.6 equiv.). The reaction mixture was stirred at 0 °C for 0.5 hours. Following this time, the reaction was quenched with a saturated aqueous solution of NH₄Cl (50 mL), and the mixture was transferred to a separatory funnel. The aqueous layer was extracted with CH₂Cl₂ (2 x 30 mL). The organic layers were collected, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 5% EtOAc/hexanes on silica gel to yield **6** (colorless oil, 5.4 g, 12.8 mmol, 94% yield).



(1S,2R)-1-((tert-butyldimethylsilyl)oxy)-1-((R)-2,2-dimethyl-1,3-dioxolan-4-yl)pent-3-en-2-yl benzoate

Compound 6

¹H NMR (400 MHz, CDCl₃) δ 8.09 – 8.01 (m, 2H), 7.56 (ddt, *J* = 7.9, 6.9, 1.4 Hz, 1H), 7.49 – 7.40 (m, 2H), 5.82 (dqd, *J* = 15.2, 6.5, 0.9 Hz, 1H), 5.54 (ddq, *J* = 15.0, 6.6, 1.5 Hz, 1H), 5.47 (ddt, *J* = 6.5, 4.1, 1.0 Hz, 1H), 4.22 – 4.14 (m, 1H), 4.14 – 4.08 (m, 1H), 3.94 (t, *J* = 7.6 Hz, 1H), 3.87 (dd, *J* = 7.8, 6.4 Hz, 1H), 1.72 (ddd, *J* = 6.5, 1.6, 1.0 Hz, 3H), 1.39 (s, 3H), 1.31 (s, 3H), 0.91 (s, 9H), 0.13 (s, 3H), 0.12 (s, 3H).

¹³C{¹H} NMR (126 MHz, CDCl₃) δ 165.5, 133.1, 130.4, 129.9, 129.8, 128.5, 126.0, 108.3, 76.3, 75.7, 73.2, 65.0, 26.6, 25.9, 25.3, 18.2, 18.0, -4.0, -4.1.

IR (v_{max}) 2931, 1726, 1267 cm⁻¹.

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{23}H_{36}O_5SiNa^+$ 443.2224 Found 443.2214 (2.3 ppm error).



A 100 mL round-bottom flask was charged with a stir bar, **6** (5.3 g, 12.6 mmol, 1 equiv.), and THF (25 mL). The reaction flask was cooled to 0 °C using an ice-water bath, and TBAF (1M in THF, 15.2 mL, 15.2 mmol, 1.2 equiv.) was added dropwise. The reaction mixture was warmed to room temperature over a period of 3 hours. Following this time, the reaction was quenched by addition of a saturated aqueous solution of NH₄Cl (30 mL) and water (20 mL). The mixture was transferred to a separatory funnel. The

aqueous layer was extracted with EtOAc ($2 \times 80 \text{ mL}$). The organic layers were collected, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 5% EtOAc/hexanes on silica gel to yield **7** (colorless oil, 3.7 g, 12.1 mmol, 96% yield).



(1S,2R)-1-((R)-2,2-dimethyl-1,3-dioxolan-4-yl)-1-hydroxypent-3-en-2-yl benzoate

Compound 7

¹H NMR (500 MHz, CDCl₃) δ 8.08 – 8.01 (m, 2H), 7.57 (ddt, *J* = 7.9, 7.0, 1.3 Hz, 1H), 7.51 – 7.41 (m, 2H), 5.91 (dqd, *J* = 15.3, 6.5, 0.8 Hz, 1H), 5.65 (ddq, *J* = 15.1, 7.1, 1.6 Hz, 1H), 5.59 (ddt, *J* = 7.1, 3.9, 0.8 Hz, 1H), 4.12 (q, *J* = 6.1 Hz, 1H), 4.06 (dd, *J* = 8.3, 6.0 Hz, 1H), 3.99 (dd, *J* = 8.3, 6.3 Hz, 1H), 3.90 (dd, *J* = 5.8, 3.8 Hz, 1H), 1.78 – 1.71 (m, 3H), 1.43 (s, 3H), 1.33 (s, 3H).

¹³C{¹H} NMR (126 MHz, CDCl₃) δ 165.6, 133.3, 131.5, 130.2, 129.8, 128.6, 125.9, 109.2, 75.9, 74.9, 73.5, 65.6, 26.8, 25.4, 18.0.

IR (v_{max}) 3471, 2929, 1715, 1270 cm⁻¹.

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{17}H_{22}O_5Na^+$ 329.1359 Found 329.1368 (2.7 ppm error).



Preparation of ClSO₂NH₂: A 50 mL oven-dried round-bottom flask was fitted with a balloon of N₂ gas and charged with a stir bar. ClSO₂NCO (2.0 mL, 3.26 g, 23.0 mmol, 2 equiv.) was added, and the flask was cooled to 0 $^{\circ}$ C using an ice-water bath. HCO₂H (0.9 mL, 1.10 g, 23.9 mmol, 2 equiv.) was added dropwise

(Caution: vigorous gas evolution upon addition). The mixture solidified into a white solid within five minutes of addition. CH_3CN (12 mL) was added, and the reaction mixture was warmed to room temperature over a period of 5 hours.

A separate 100 mL oven-dried round-bottom flask was fitted with a nitrogen balloon and charged with a stir bar, **7** (3.6 g, 11.75 mmol, 1 equiv.), and DMA (25 mL). The reaction flask was cooled to 0 °C using an ice-water bath, and freshly prepared $ClSO_2NH_2$ (in 12 mL of CH_3CN) was added dropwise. The reaction mixture was warmed to room temperature over a period of 2 hours. Following this time, the reaction was quenched by addition of saturated aqueous NaHCO₃ solution (20 mL). The mixture was transferred to a separatory funnel and further diluted with water (20 mL). The aqueous layer was extracted with EtOAc (3 x 30 mL). The organic layers were combined, dried over MgSO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 25% EtOAc/hexanes on silica gel to yield **8** (colorless oil, 3.4 g, 8.82 mmol, 75% yield).



(1S,2R)-1-((R)-2,2-dimethyl-1,3-dioxolan-4-yl)-1-(sulfamoyloxy)pent-3-en-2-yl benzoate

Compound 8

¹H NMR (400 MHz, CDCl₃) δ 8.10 – 8.01 (m, 2H), 7.62 – 7.54 (m, 1H), 7.49 – 7.41 (m, 2H), 5.98 (dq, *J* = 14.4, 6.6 Hz, 1H), 5.67 – 5.55 (m, 2H), 5.01 (dd, *J* = 4.7, 3.9 Hz, 1H), 4.97 (broad s, 2H), 4.31 (td, *J* = 6.8, 4.0 Hz, 1H), 4.06 (dd, *J* = 8.5, 7.0 Hz, 1H), 3.98 (dd, *J* = 8.5, 6.5 Hz, 1H), 1.76 (dd, *J* = 6.5, 1.4 Hz, 3H), 1.42 (s, 3H), 1.36 – 1.33 (m, 3H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ 165.4, 133.6, 133.3, 129.9, 129.6, 128.7, 124.4, 109.4, 81.9, 74.6, 73.5, 64.8, 26.5, 25.2, 18.0.

IR (v_{max}) 3450, 2988, 1710, 1373, 1270 cm⁻¹.

HRMS (ESI) m/z: $[M+Na^+]$ calculated mass for $C_{17}H_{23}NO_7SNa^+$ 408.1087 Found 408.1092 (1.2 ppm error).



A 100 mL round-bottom flask was charged with a stir bar, **8** (3.4 g, 8.8 mmol, 1 equiv.), and MeOH (20 mL). The reaction flask was cooled to 0 °C using an ice-water bath, and K_2CO_3 (1.5 g, 10.9 mmol, 1.2 equiv.) was added in one bolus. The reaction mixture was warmed to room temperature over a period of 5 hours. Then, the reaction was quenched with a saturated aqueous solution of NH₄Cl (20 mL), and the methanol was removed under reduced pressure. The mixture was transferred to a separatory funnel, further diluted with water (20 mL), and the aqueous layer was extracted with EtOAc (2 x 20 mL). The organic layers were collected, dried over MgSO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 35% EtOAc/hexanes on silica gel to yield **9** (colorless oil, 1.9 g, 6.8 mmol, 77% yield).



(1S,2R)-1-((R)-2,2-dimethyl-1,3-dioxolan-4-yl)-2-hydroxypent-3-en-1-yl sulfamate

Compound 9

¹H NMR (400 MHz, CDCl₃) δ 5.84 (dqd, J = 15.3, 6.5, 1.1 Hz, 1H), 5.57 (ddq, J = 15.3, 7.0, 1.6 Hz, 1H), 5.19 (s, 2H), 4.70 (dd, J = 5.3, 4.5 Hz, 1H), 4.29 (qd, J = 6.7, 2.8 Hz, 2H), 4.08 – 3.90 (m, 2H), 2.57 (broad s, 1H), 1.76 (ddd, J = 6.5, 1.7, 0.8 Hz, 3H), 1.48 – 1.42 (s, 3H), 1.40 – 1.34 (s, 3H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ 130.8, 128.3, 109.7, 83.8, 74.6, 72.4, 65.1, 26.4, 25.3, 17.9.

IR (v_{max}) 3500, 2930, 1373, 1216 cm⁻¹.

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{10}H_{19}NO_6SNa^+$ 304.0825 Found 304.0845 (6.6 ppm error).



A 250 mL round-bottom flask was charged with a stir bar, **9** (1.4 g, 4.98 mmol, 1 equiv.), Pd(PhCN)₂Cl₂ (0.282 g, 0.73 mmol, 0.15 equiv.), Cu(OAc)₂ (0.890 g, 4.9 mmol, 1 equiv.), Fmoc-Gly-OH (1.4 g, 4.7 mmol, 1 equiv.), molecular sieves (4Å, 725 mg (5 mg/mL of solvent)), and DMSO (145 mL, final concentration: 0.034 M). The reaction vessel was evacuated and backfilled with O₂ gas three times. Then, it was submerged in an oil bath preheated to 60 °C and kept at this temperature under a balloon of O₂ (~1 atm) for 17 hours. Following this time, the reaction mixture was filtered through a pad of silica gel. The filter cake was further washed with EtOAc. The filtrate was diluted with H₂O (500 mL), and the mixture was transferred to a separatory funnel. The aqueous layer was extracted with EtOAc (3 x 200 mL). The organic layers were collected, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 35% EtOAc/hexanes on silica gel to yield **10** (crystalline solid, 1.0 g, 3.58 mmol, 72% yield).



(4S,5R,6S)-6-((R)-2,2-dimethyl-1,3-dioxolan-4-yl)-5-hydroxy-4-vinyl-1,2,3-oxathiazinane 2,2-dioxide

Compound 10

¹H NMR (400 MHz, CDCl₃) δ 5.89 (ddd, J = 17.3, 10.7, 4.7 Hz, 1H), 5.46 (dd, J = 5.8, 1.8 Hz, 1H), 5.43 (d, J = 1.8 Hz, 1H), 4.80 (d, J = 11.3 Hz, 1H), 4.54 (dd, J = 7.9, 1.2 Hz, 1H), 4.46 – 4.38 (m, 2H), 4.18 (dd, J = 9.2, 6.2 Hz, 1H), 4.05 (dd, J = 9.2, 4.3 Hz, 1H), 3.93 (dt, J = 4.6, 1.4 Hz, 1H), 2.71 (d, J = 4.6 Hz, 1H), 1.46 (s, 3H), 1.39 (s, 3H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ 132.2, 119.1, 110.5, 84.8, 72.9, 66.7, 62.7, 60.9, 26.9, 25.0.

IR (v_{max}) 3548, 2937, 1370, 1190 cm⁻¹.

 $[\alpha]_{D}^{25.5} = -20.35$ (*c* 0.48, CH₃Cl).

HRMS (ESI) m/z = [M - H] calculated mass for $C_{10}H_{16}NO_6S^2$ 278.0704 Found 278.0685 (6.8 ppm error).



A 50 mL round-bottom flask was charged with a stir bar, **10** (0.824 g, 2.95 mmol, 1 equiv.), and CH_2Cl_2 (15 mL). The reaction flask was cooled to 0 °C using an ice-water bath. 2,6-lutidine (1.72 mL, 1.59 g, 14.8 mmol, 5 equiv.) and TBSOTf (2.8 mL, 3.22 g, 12.2 mmol, 4 equiv.) were sequentially added dropwise. The reaction mixture was stirred at 0 °C for 30 minutes. The reaction was then quenched by addition of a saturated aqueous solution of NaHCO₃ (15 mL), and the mixture was transferred to a separatory funnel. The aqueous layer was extracted with CH_2Cl_2 (3 x 15 mL). The organic layer was collected, dried over

Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 15% EtOAc/hexanes on silica gel to yield compound **11** (colorless oil, 0.937 g, 2.38 mmol, 81% yield).



(4*S*,5*R*,6*R*)-5-((*tert*-butyldimethylsilyl)oxy)-6-((*R*)-2,2-dimethyl-1,3-dioxolan-4-yl)-4-vinyl-1,2,3-oxathiazinane 2,2-dioxide

Compound 11

¹H NMR (400 MHz, CDCl₃) δ 5.76 (ddd, *J* = 17.2, 10.7, 4.1 Hz, 1H), 5.41 – 5.29 (m, 2H), 4.44 – 4.31 (m, 3H), 4.22 – 4.14 (m, 2H), 4.01 – 3.96 (m, 1H), 3.90 (t, *J* = 1.0 Hz, 1H), 1.42 (s, 3H), 1.35 (s, 3H), 0.90 (s, 9H), 0.18 (s, 3H), 0.09 (s, 3H).

¹³C{¹H} NMR (126 MHz, CDCl₃) δ 133.5, 118.2, 110.4, 86.5, 71.2, 67.3, 64.5, 61.9, 26.9, 26.3, 25.1, 18.6, -3.6, -4.2.

IR (v_{max}) 3271, 2931, 1373, 1196 cm⁻¹.

 $[\alpha]_{D}^{26.0} = -20.55 \ (c \ 0.85, \text{CH}_3\text{Cl}).$

HRMS (ESI) m/z = [M - H] calculated mass for $C_{16}H_{30}NO_6SSi^-$ 392.1569 Found 392.1561 (2.0 ppm error).



A 100 mL round-bottom flask was charged with a stir bar, **11** (920 mg, 2.33 mmol, 1 equiv.), and CH_2Cl_2 (25 mL). The reaction flask was cooled to 0 °C using an ice-water bath. Sequentially, TMEDA (0.7 mL,

0.55 g, 4.7 mmol, 2 equiv.) and CbzCl (1.0 mL, 1.2 g, 7.03 mmol, 3 equiv.) were added dropwise. The reaction mixture was warmed to room temperature over a period of 40 hours. Subsequently, the reaction was quenched with a saturated aqueous solution of NH₄Cl (20 mL), and the mixture was transferred to a separatory funnel. The aqueous layer was extracted with CH_2Cl_2 (2 x 30 mL). The organic layers were collected, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 20% EtOAc/hexanes on silica gel to yield **12** (colorless oil, 860 mg, 1.63 mmol, 70% yield).



benzyl (4*S*,5*R*,6*R*)-5-((*tert*-butyldimethylsilyl)oxy)-6-((*R*)-2,2-dimethyl-1,3-dioxolan-4-yl)-4-vinyl-1,2,3-oxathiazinane-3-carboxylate 2,2-dioxide

Compound 12

¹H NMR (400 MHz, CDCl₃) δ 7.47 – 7.29 (m, 5H), 6.15 (ddd, *J* = 17.1, 10.5, 7.8 Hz, 1H), 5.41 – 5.27 (m, 4H), 5.19 (tt, *J* = 7.6, 1.2 Hz, 1H), 4.80 (dd, *J* = 6.7, 5.8 Hz, 1H), 4.60 (dd, *J* = 7.4, 5.8 Hz, 1H), 4.43 (td, *J* = 6.6, 5.6 Hz, 1H), 4.13 (dd, *J* = 9.1, 6.4 Hz, 1H), 4.02 (dd, *J* = 9.0, 5.7 Hz, 1H), 1.43 – 1.38 (m, 3H), 1.37 – 1.31 (m, 3H), 0.90 (s, 9H), 0.11 (s, 3H), 0.08 (s, 3H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ 151.7, 134.6, 131.8, 128.7, 128.6, 127.8, 121.2, 110.1, 85.3, 72.1,

69.8, 67.7, 66.1, 63.6, 26.6, 25.8, 25.3, 18.1, -4.4, -4.9.

IR (v_{max}) 2971, 1704, 1284, 1190 cm⁻¹.

 $[\alpha]_{D}^{26.0} = -10.8 (c \ 1.30, CH_{3}Cl)$

HRMS (ESI) m/z = [M - H] calculated mass for $C_{24}H_{36}NO_8SSi^-526.1936$ Found 526.1925 (2.1 ppm error).



A 50 mL round-bottom flask was charged with a stir bar, **12** (0.820 g, 1.55 mmol, 1 equiv.), DMSO (20 mL), and KOAc (0.610 g, 6.21 mmol, 4 equiv.). The reaction vessel was then submerged in an oil bath preheated to 80 °C and kept at this temperature under a balloon of N_2 (~1 atm) for 4 hours. Following this time, the reaction was cooled to room temperature, diluted with water (80 mL), and transferred to a separatory funnel. The aqueous layer was extracted with EtOAc (2 x 50 mL). The organic layers were collected, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 20% EtOAc/hexanes on silica gel to yield **13** (colorless oil, 0.661 g, 1.30 mmol, 84% yield).



(1S,2R,3S)-3-(((benzyloxy)carbonyl)amino)-2-((*tert*-butyldimethylsilyl)oxy)-1-((*R*)-2,2-dimethyl-1,3-dioxolan-4-yl)pent-4-en-1-yl acetate

Compound 13

¹H NMR (400 MHz, CDCl₃) δ 7.43 – 7.29 (m, 5H), 5.81 (ddd, J = 17.2, 10.4, 4.5 Hz, 1H), 5.28 – 5.17 (m, 2H), 5.08 (dd, J = 11.4, 7.1 Hz, 3H), 4.87 (dd, J = 8.6, 1.9 Hz, 1H), 4.44 (ddt, J = 10.8, 3.9, 1.8 Hz, 1H), 4.35 (td, J = 6.6, 1.9 Hz, 1H), 4.13 (dd, J = 8.6, 1.7 Hz, 1H), 3.98 (dd, J = 8.7, 6.8 Hz, 1H), 3.67 (dd, J = 8.7, 6.4 Hz, 1H), 2.10 (s, 3H), 1.41 (s, 3H), 1.34 (s, 3H), 0.87 (s, 9H), 0.14 (s, 3H), 0.08 (s, 3H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ 170.6, 156.0, 136.7, 136.6, 128.6, 128.4, 128.2, 116.2, 109.4, 73.6, 72.9, 71.5, 67.0, 65.7, 54.2, 26.2, 25.6, 21.1, 18.3, -3.3, -4.4.

IR (v_{max}) 3448, 2954, 1747, 1730, 1504, 1224 cm⁻¹.

 $[\alpha]_{D}^{26.6} = -24.89 (c \ 1.30, CH_{3}Cl).$

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for C₂₆H₄₁NO₇SiNa⁺ 530.2545 Found 530.2546 (0.2 ppm error).



A 100 mL round-bottom flask was charged with a stir bar, **13** (0.65 g, 1.28 mmol, 1 equiv.), and MeOH (6 mL). The reaction flask was cooled to 0 °C using an ice-water bath, and K_2CO_3 (0.177 g, 1.28 mmol, 1.0 equiv.) was added in one portion. The reaction mixture was warmed to room temperature over a period of 3 hours. Following this time, the reaction was quenched with a saturated aqueous solution of NH₄Cl (15 mL), further diluted with water (20 mL), and the mixture was transferred to a separatory funnel. The aqueous layer was extracted with EtOAc (3 x 30 mL). The organic layers were collected, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 20% EtOAc/hexanes on silica gel to yield **14** (colorless oil, 0.381 g, 0.818 mmol, 64% yield).



benzyl ((3*S*,4*R*,5*S*)-4-((*tert*-butyldimethylsilyl)oxy)-5-((*R*)-2,2-dimethyl-1,3-dioxolan-4-yl)-5-hydroxypent-1-en-3-yl)carbamate

Compound 14

¹H NMR (400 MHz, CDCl₃) δ 7.44 – 7.28 (m, 5H), 5.94 (ddd, *J* = 17.3, 10.5, 4.4 Hz, 1H), 5.60 (d, *J* = 9.8 Hz, 1H), 5.31 – 5.20 (m, 2H), 5.19 – 5.05 (m, 2H), 4.61 (dtd, *J* = 10.6, 6.2, 5.3, 3.1 Hz, 1H), 4.30 (td, J = 10.6, 6.2, 5.3, 5.3, 5.1, 5.2)

7.0, 2.2 Hz, 1H), 4.00 (dd, *J* = 8.0, 6.7 Hz, 1H), 3.87 (t, *J* = 7.6 Hz, 1H), 3.72 (dd, *J* = 9.1, 2.5 Hz, 1H), 3.38 (t, *J* = 7.6 Hz, 1H), 2.49 (d, *J* = 8.1 Hz, 1H), 1.44 (s, 3H), 1.35 (s, 3H), 0.90 (s, 9H), 0.15 (s, 3H), 0.12 (s, 3H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ 156.2, 136.6, 135.2, 128.6, 128.3, 128.2, 116.0, 109.4, 74.3, 74.1, 70.9, 67.0, 66.2, 55.8, 26.5, 26.0, 25.2, 18.1, -3.8, -4.6.

IR (v_{max}) 3434, 2931, 1724, 1504, 1258 cm⁻¹.

 $[\alpha]_{D}^{26.3} = -58.15$ (*c* 1.00, CH₃Cl).

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{24}H_{39}NO_6SiNa^+ 488.2439$ Found 488.2438 (0.2 ppm error).



A 50 mL round-bottom flask was charged with a stir bar, **14** (0.370 g, 0.795 mmol, 1 equiv.), toluene (6 mL), and benzyl isocyanate (0.2 mL, 0.216 g, 1.62 mmol, 2 equiv.). The reaction vessel was then submerged into an oil bath preheated to 115 °C and kept at this temperature under a balloon of N₂ (~1 atm) for 24 hours. Subsequently, the reaction was cooled to room temperature, and the solvent was removed under reduced pressure. The resulting residue was purified using a gradient of 0 to 20% EtOAc/hexanes on silica gel to yield **15** (colorless oil, 0.302 g, 0.504 mmol, 64% yield).



(1S,2R,3S)-3-(((benzyloxy)carbonyl)amino)-2-((*tert*-butyldimethylsilyl)oxy)-1-((R)-2,2-dimethyl-1,3-dioxolan-4-yl)pent-4-en-1-yl benzylcarbamate

Compound 15

¹H NMR (400 MHz, Chloroform-*d*) δ 7.47 – 7.27 (m, 10H), 5.85 (ddd, *J* = 17.2, 10.3, 4.6 Hz, 1H), 5.27 – 5.15 (m, 3H), 5.10 (s, 2H), 5.01 (t, *J* = 5.9 Hz, 1H), 4.89 – 4.79 (m, 1H), 4.50 (d, *J* = 8.1 Hz, 1H), 4.43 – 4.24 (m, 3H), 4.14 – 3.94 (m, 2H), 3.79 (dd, *J* = 8.6, 6.8 Hz, 1H), 1.39 (s, 3H), 1.33 (s, 3H), 0.88 (s, 9H), 0.14 (s, 3H), 0.10 (s, 3H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ 156.0, 155.9, 138.3, 136.8, 136.3, 128.7, 128.6, 128.3, 128.2, 127.6, 127.5, 116.2, 109.3, 74.1, 73.0, 72.5, 66.9, 65.7, 54.7, 45.4, 26.3, 26.1, 25.6, 18.3, -3.5, -4.5.

IR (v_{max}) 3340, 2931, 1738, 1654, 1415 cm⁻¹.

 $[\alpha]_{D}^{26.7} = -8.02 (c \ 1.40, CH_{3}Cl).$

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{32}H_{46}N_2O_7SiNa^+$ 621.2966 Found 621.2987 (3.4 ppm error).



A 50 mL round-bottom flask was charged with a stir bar, **15** (0.25 g, 0.417 mmol, 1 equiv.), and 'BuOH/H₂O (2:1 mixture, 4 mL total volume). The reaction flask was cooled to 0 °C using an ice-water bath. $K_2OsO_4 \cdot 2H_2O$ (16 mg, 0.043 mmol, 0.1 equiv.) and NMO \cdot H₂O (195 mg, 1.44 mmol, 3.5 equiv.) were

added sequentially. The reaction mixture was warmed to room temperature over a period of 14 hours. Following this time, it was quenched with a saturated, aqueous solution of Na_2SO_3 (20 mL), and the mixture was transferred to a separatory funnel. The aqueous layer was extracted with EtOAc (2 x 15 mL). The organic layers were collected, dried over Na_2SO_4 , and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 60% EtOAc/hexanes on silica gel to yield **16** (colorless oil, 0.181 g, 0.286 mmol, 69% yield).



(1*S*,2*R*,3*S*)-3-(((benzyloxy)carbonyl)amino)-2-((*tert*-butyldimethylsilyl)oxy)-1-((*R*)-2,2-dimethyl-1,3-dioxolan-4-yl)-4,5-dihydroxypentyl benzylcarbamate

Compound 16

¹H NMR (500 MHz, CD₃OD) δ 7.41 – 7.18 (m, 10H), 5.12 – 5.00 (m, 2H), 4.78 (ddd, *J* = 10.3, 7.4, 2.8 Hz, 1H), 4.51 (dd, *J* = 7.5, 1.2 Hz, 1H), 4.41 (td, *J* = 6.7, 2.7 Hz, 1H), 4.31 – 4.21 (m, 2H), 4.07 – 3.98 (m, 1H), 3.81 – 3.72 (m, 2H), 3.62 (dtd, *J* = 9.4, 4.6, 2.3 Hz, 1H), 3.57 – 3.49 (m, 2H), 1.38 (s, 3H), 1.34 (s, 3H), 0.92 (s, 9H), 0.22 (s, 3H), 0.20 (s, 3H).

¹³C{¹H} NMR (126 MHz, CD₃OD) δ 158.7, 158.3, 140.4, 138.0, 129.5, 129.4, 129.2, 129.1, 128.2, 128.0, 110.6, 75.5, 74.1, 71.9, 70.3, 68.1, 66.5, 65.2, 53.7, 45.6, 26.7, 26.4, 25.9, 19.2, -4.0, -4.3.

IR (v_{max}) 3446, 2929, 1701, 1427, 1253 cm⁻¹.

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{32}H_{48}N_2O_9SiNa^+ 655.3021$ Found 655.3051 (4.6 ppm error).



A 50 mL round-bottom flask was charged with a stir bar, **16** (0.170 g, 0.27 mmol, 1 equiv.), and CH_3CN/H_2O (2:1 mixture; 6 mL total volume). The reaction flask was cooled to 0 °C using an ice-water bath. NaIO₄ (0.115 g, 0.54 mmol, 2 equiv.) and NaHCO₃ (0.068 g, 0.81 mmol, 3 equiv.) were added sequentially. The reaction mixture was warmed to room temperature over a period of 2 hours. Following this time, the reaction mixture was filtered through a sintered glass funnel, and the filtrate was transferred to a separatory funnel with 20 mL of H₂O. The aqueous layer was extracted with EtOAc (3 x 15 mL). The organic layers were collected, dried over Na₂SO₄, and concentrated under reduced pressure. The crude product was used in the next step without further purification.

A 50 mL round-bottom flask was charged with a stir bar, aldehyde starting material, and CH_3CN/H_2O (50:1 mixture, 4 mL total volume). The reaction flask was cooled to 0 °C using an ice-water bath, and p-TsOH•H₂O (10 mg, 0.053 mmol, 0.2 equiv.) was added. The reaction mixture was warmed to room temperature over a period of 4 hours. Following this time, the reaction was quenched with saturated aqueous NaHCO₃ solution (10 mL), and the mixture was transferred to a separatory funnel. The aqueous layer was extracted with EtOAc (2 x 10 mL). The organic layers were collected, dried over Na₂SO₄, and concentrated under reduced pressure. The resulting residue was purified using a gradient of 50 to 100% EtOAc/hexanes on silica gel to yield **17** (colorless oil, 0.084 g, 0.150 mmol, 56% yield over two steps).



(2R,3S,4R,5R,6S)-5-(((benzyloxy)carbonyl)amino)-4-((*tert*-butyldimethylsilyl)oxy)-6-hydroxy-2-(hydroxymethyl)tetrahydro-2*H*-pyran-3-yl benzylcarbamate

Compound 17

¹H NMR (500 MHz, CD₃OD) δ 7.46 – 7.17 (m, 10H), 5.10 – 5.09 (m, 1H), 5.06 (broad s, 3H), 4.33 – 4.21 (m, 2H), 4.15 (t, *J* = 6.4 Hz, 1H), 4.04 – 3.98 (m, 2H), 3.59 – 3.50 (m, 2H), 0.80 (s, 9H), 0.07 (s, 3H), 0.03 (s, 3H).

¹³C{¹H} NMR (126 MHz, CD₃OD) δ 159.1, 158.6, 140.4, 138.1, 129.46, 129.45, 129.1, 129.0, 128.4, 128.1, 93.7, 73.2, 71.1, 69.7, 67.6, 62.2, 54.1, 45.5, 26.2, 18.7, -4.5, -4.8.

IR (v_{max}) 3412, 2951, 1715, 1521, 1253 cm⁻¹.

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{28}H_{40}N_2O_8SiNa^+$ 583.2446 Found 583.2472 (4.8 ppm error).



A 25 mL round-bottom flask was charged with a stir bar, **17** (0.028 g, 0.050 mmol, 1 equiv.) and CH₂Cl₂ (2 mL). The reaction flask was cooled to 0 °C using an ice-water bath. Subsequently, TMEDA (7 μ L, 6 mg, 0.049 mmol, 1 equiv.) followed by BzCl (13 μ L, 15 mg, 0.107 mmol, 2.1 equiv.) were added dropwise. The reaction mixture was stirred at 0 °C for 4 hours. Following this time, the reaction was quenched with a saturated aqueous solution of NH₄Cl (10 mL), and the mixture was transferred to a separatory funnel. The

aqueous layer was extracted with CH_2Cl_2 (3 x 5 mL). The organic layers were collected, dried over Na_2SO_4 , and concentrated under reduced pressure. The resulting residue was purified using a gradient of 0 to 5% EtOAc/hexanes on silica gel to yield **18** (colorless oil, 0.037 g, 0.048 mmol, 96% yield).



((2R,3S,4R,5R,6R)-6-(benzoyloxy)-3-((benzylcarbamoyl)oxy)-5-(((benzyloxy)carbonyl)amino)-4-((*tert*-butyldimethylsilyl)oxy)tetrahydro-2*H*-pyran-2-yl)methyl benzoate

Compound 18

¹H NMR (500 MHz, CD₃OD) δ 8.14 – 8.07 (m, 2H), 7.91 – 7.83 (m, 2H), 7.68 – 7.58 (m, 1H), 7.58 – 7.46 (m, 3H), 7.44 – 7.18 (m, 12H), 6.41 (d, *J* = 3.8 Hz, 1H), 5.35 (d, *J* = 3.5 Hz, 1H), 5.11 – 5.01 (m, 2H), 4.52 (dd, *J* = 7.4, 4.5 Hz, 1H), 4.43 – 4.32 (m, 3H), 4.26 (td, *J* = 5.7, 3.4 Hz, 3H), 0.82 (s, 9H), 0.16 (s, 3H), 0.10 (s, 3H).

¹³C{¹H} NMR (126 MHz, CD₃OD) δ 164.6, 163.7, 156.0, 155.4, 137.3, 135.1, 131.8, 131.3, 128.1, 127.6, 126.7, 126.6, 126.5, 126.4, 126.1, 125.5, 125.4, 125.1, 91.1, 69.4, 68.5, 66.7, 64.9, 61.8, 50.1, 42.6, 23.2, 15.8, -7.3, -7.6.

IR (v_{max}) 2929, 1724, 1453, 1260 cm⁻¹.

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{42}H_{48}N_2O_{10}SiNa^+$ 791.2970 Found 791.2997 (3.4 ppm error).



A 25 mL round-bottom flask was charged with a stir bar, **18** (0.03 g, 0.039 mmol, 1 equiv.), and THF (6 mL). MeNH₂ (2.0 M solution in THF, 0.039 mL, 0.078 mmol, 2 equiv.) was added at room temperature. The reaction mixture was stirred for 48 hours. Following this time, the solvent was removed under reduced pressure. The resulting residue was purified using a gradient of 0 to 20% EtOAc/hexanes on silica gel to yield **19** (colorless oil, 0.022 g, 0.033 mmol, 85% yield).



((2*R*,3*S*,4*R*,5*R*,6*S*)-3-((benzylcarbamoyl)oxy)-5-(((benzyloxy)carbonyl)amino)-4-((*tert*-butyldimethylsilyl)oxy)-6hydroxytetrahydro-2*H*-pyran-2-yl)methyl benzoate

Compound 19

¹H NMR (500 MHz, CD₃OD) δ 8.04 (dt, *J* = 8.1, 1.2 Hz, 2H), 7.59 (tq, *J* = 7.0, 1.3 Hz, 1H), 7.49 – 7.42 (m, 2H), 7.39 – 7.17 (m, 10H), 5.25 – 5.21 (m, 1H), 5.15 – 5.13 (m, 1H), 5.06 (s, 2H), 4.54 – 4.46 (m, 1H), 4.42 – 4.33 (m, 2H), 4.30 – 4.20 (m, 2H), 4.09 – 4.03 (m, 2H), 0.81 (s, 9H), 0.10 (s, 3H), 0.05 (s, 3H).

¹³C{¹H} NMR (126 MHz, CD₃OD) δ 167.7, 158.6, 140.3, 138.1, 134.2, 131.2, 130.7, 129.5, 129.45, 129.40, 129.1, 129.0, 128.3, 128.0, 93.8, 73.1, 69.5, 68.4, 67.6, 64.9, 54.0, 45.5, 26.2, 18.6, -4.5, -4.7.

IR (v_{max}) 3348, 2929, 1730, 1267 cm⁻¹.

HRMS (ESI) $m/z = [M + Na^+]$ calculated mass for $C_{35}H_{44}N_2O_9SiNa^+$ 687.2708 Found 687.2711 (0.4 ppm error).

Acknowledgements

This work was supported by a National Institutes of Health grant R35GM142499 awarded to Shyam Sathyamoorthi. Justin Douglas and Sarah Neuenswander (KU NMR Lab) are acknowledged for help with structural elucidation. Lawrence Seib and Anita Saraf (KU Mass Spectrometry Facility) are acknowledged for help acquiring HRMS data. Joel T. Mague thanks Tulane University for support of the Tulane Crystallography Laboratory. We gratefully acknowledge Professor Mark Farrell and Professor Robert A. Pascal, Jr. for many discussions regarding this manuscript and the larger area of carbohydrate chemistry.

References

 Banoub, J.; Boullanger, P.; Lafont, D., Synthesis of oligosaccharides of 2-amino-2-deoxy sugars. *Chem. Rev.* 1992, 92, 1167-1195.

Stachlewitz, R. F.; Seabra, V.; Bradford, B.; Bradham, C. A.; Rusyn, I.; Germolec, D.; Thurman,
 R. G., Glycine and uridine prevent d-galactosamine hepatotoxicity in the rat: Role of kupffer cells.
 Hepatology 1999, 29, 737-745.

3. Sugahara, K.; Mikami, T.; Uyama, T.; Mizuguchi, S.; Nomura, K.; Kitagawa, H., Recent advances in the structural biology of chondroitin sulfate and dermatan sulfate. *Curr. Opin. Struct. Biol.* **2003**, *13*, 612-620.

4. Herzner, H.; Reipen, T.; Schultz, M.; Kunz, H., Synthesis of Glycopeptides Containing Carbohydrate and Peptide Recognition Motifs. *Chem. Rev.* **2000**, *100*, 4495-4538.

5. Wolfrom, M. L.; Onodera, K., Dithioacetals of D-Glucuronic Acid and 2-Amino-2-deoxy-D-galactose. *J. Am. Chem. Soc.* **1957**, *79*, 4737-4740.

6. Danishefsky, S. J.; Bilodeau, M. T., Glycals in Organic Synthesis: The Evolution of Comprehensive Strategies for the Assembly of Oligosaccharides and Glycoconjugates of Biological Consequence. *Angew. Chem. Int. Ed.* **1996**, *35*, 1380-1419.

7. Kinfe, H. H., Versatility of glycals in synthetic organic chemistry: coupling reactions, diversity oriented synthesis and natural product synthesis. *Org. Biomol. Chem.* **2019**, *17*, 4153-4182.

8. Mirabella, S.; Cardona, F.; Goti, A., From glycals to aminosugars: a challenging test for new stereoselective aminohydroxylation and related methodologies. *Org. Biomol. Chem.* **2016**, *14*, 5186-5204.

James, S. P.; Smith, F.; Stacey, M.; Wiggins, L. F., 130. The action of alkaline reagents on 2 : 3-1
: 6- and 3 : 4-1 : 6-dianhydro β-talose. A constitutional synthesis of chondrosamine and other amino-sugar derivatives. *J. Chem. Soc. (Resumed)* **1946**, 625-628.

10. Dhurandhare, V. M.; Wen, Y.-S.; Gawande, S. D.; Liao, P.-H.; Wang, C.-C., Synthesis of d-Galactosamine and d-Allosamine Derivatives via a Microwave-Assisted Preparation of 1,6-Anhydroglucosamine. *J. Org. Chem.* **2016**, *81*, 11521-11528.

11. Inoue, K.; Nishimoto, M.; Kitaoka, M., One-pot enzymatic production of 2-acetamido-2-deoxy-d-galactose (GalNAc) from 2-acetamido-2-deoxy-d-glucose (GlcNAc). *Carbohydr. Res.* **2011**, *346*, 2432-2436.

12. Černý, M.; Staněk, J., 1,6-Anhydro Derivatives of Aldohexoses. In *Advances in Carbohydrate Chemistry and Biochemistry*, Stuart Tipson, R.; Horton, D., Eds. Academic Press1977; Vol. 34, pp 23-177.

13. Wrodnigg, T. M.; Lundt, I.; Stütz, A. E., Synthesis of N-Protected Galactosamine Building Blocks from d-Tagatose via the Heyns Rearrangement. *J. Carbohydr. Chem.* **2006**, *25*, 33-41.

14. Guazzelli, L.; Catelani, G.; D'Andrea, F.; Giannarelli, A., Stereoselective entry into the d-GalNAc series starting from the d-Gal one: a new access to N-acetyl-d-galactosamine and derivatives thereof. *Carbohydr. Res.* **2009**, *344*, 298-303.

Kulkarni, S. S.; Wang, C.-C.; Sabbavarapu, N. M.; Podilapu, A. R.; Liao, P.-H.; Hung, S.-C., "One-Pot" Protection, Glycosylation, and Protection–Glycosylation Strategies of Carbohydrates. *Chem. Rev.* 2018, *118*, 8025-8104.

Lemieux, R. U.; Nagabhushan, T. L., The synthesis of 2-amino-2-deoxyhexoses: D-glucosamine,
 D-mannosamine, D-galactosamine, and D-talosamine. *Can. J. Chem.* **1968**, *46*, 401-403.

 Lemieux, R. U.; Ratcliffe, R. M., The azidonitration of tri-O-acetyl-D-galactal. *Can. J. Chem.* 1979, 57, 1244-1251.

Czernecki, S.; Ayadi, E.; Randriamandimby, D., New and efficient synthesis of protected 2-azido 2-deoxy-glycopyranoses from the corresponding glycal. *J. Chem. Soc., Chem. Commun.* 1994, 35-36.

19. Plattner, C.; Höfener, M.; Sewald, N., One-Pot Azidochlorination of Glycals. *Org. Lett.* **2011**, *13*, 545-547.

20. Das, J.; Schmidt, R. R., Convenient Glycoside Synthesis of Amino Sugars: Michael-Type Addition to 2-Nitro-D-galactal. *Eur. J. Org. Chem.* **1998**, *1998*, 1609-1613.

21. Santoyo-Gonzalez, F.; Calvo-Flores, F. G.; Garcia-Mendoza, P.; Hernandez-Mateo, F.; Isac-Garcia, J.; Robles-Diaz, R., Synthesis of phenyl 2-azido-2-deoxy-1-selenoglycosides from glycals. *J. Org. Chem.* **1993**, *58*, 6122-6125.

22. Liu, J.; Gin, D. Y., C2-Amidoglycosylation. Scope and Mechanism of Nitrogen Transfer. *J. Am. Chem. Soc.* **2002**, *124*, 9789-9797.

 Guberman, M.; Pieber, B.; Seeberger, P. H., Safe and Scalable Continuous Flow Azidophenylselenylation of Galactal to Prepare Galactosamine Building Blocks. *Org. Process Res. Dev.* 2019, 23, 2764-2770.

24. Cai, Y.; Ling, C.-C.; Bundle, D. R., Concise and Efficient Synthesis of 2-Acetamido-2-deoxy-β-dhexopyranosides of Diverse Aminosugars from 2-Acetamido-2-deoxy-β-d-glucose. *J. Org. Chem.* **2009**, *74*, 580-589.

 McGeary, R. P.; Wright, K.; Toth, I., Conversion of Glucosamine to Galactosamine and Allosamine Derivatives: Control of Inversions of Stereochemistry at C-3 and C-4. *J. Org. Chem.* 2001, *66*, 5102-5105.
 Feng, J.; Ling, C.-C., An efficient conversion of N-acetyl-d-glucosamine to N-acetyl-d-glucosamine and derivatives. *Carbohydr. Res.* 2010, *345*, 2450-2457. 27. Glibstrup, E.; Pedersen, C. M., Scalable Synthesis of Anomerically Pure Orthogonal-Protected GlcN3 and GalN3 from d-Glucosamine. *Org. Lett.* **2016**, *18*, 4424-4427.

28. Hederos, M.; Konradsson, P., Efficient Routes to Ethyl-2-Deoxy-2-phthalimido-1-β-D-thiogalactosamine Derivatives via Epimerization of the Corresponding Glucosamine Compounds. *J. Carbohydr. Chem.* **2005**, *24*, 297-320.

29. Ochiai, H.; Niwa, T.; Hosoya, T., Stereoinversion of Stereocongested Carbocyclic Alcohols via Triflylation and Subsequent Treatment with Aqueous N,N-Dimethylformamide. *Org. Lett.* **2016**, *18*, 5982-5985.

30. Chatterjee, D.; Nayak, S.; Paul, A.; Yadav, S., Syntheses of Orthogonally Protected d-Galactosamine, d-Allosamine and d-Gulosamine Thioglycoside Building Blocks with N-phthalimido Groups. *Asian J. Org. Chem.* **2019**, *8*, 2065-2072.

31. Emmadi, M.; Kulkarni, S. S., Rapid Transformation of d-Mannose into Orthogonally Protected d-Glucosamine and d-Galactosamine Thioglycosides. *J. Org. Chem.* **2011**, *76*, 4703-4709.

32. Emmadi, M.; Kulkarni, S. S., Synthesis of orthogonally protected bacterial, rare-sugar and D-glycosamine building blocks. *Nat. Protoc.* **2013**, *8*, 1870-1889.

33. Shinde, A. H.; Sathyamoorthi, S., Oxidative Cyclization of Sulfamates onto Pendant Alkenes. *Org. Lett.* **2020**, *22*, 896-901.

34. Shinde, A. H.; Nagamalla, S.; Sathyamoorthi, S., N-arylated oxathiazinane heterocycles are convenient synthons for 1,3-amino ethers and 1,3-amino thioethers. *Med. Chem. Res.* **2020**, *29*, 1223-1229.

35. Nagamalla, S.; Johnson, D. K.; Sathyamoorthi, S., Sulfamate-tethered aza-Wacker approach towards analogs of Bactobolin A. *Med. Chem. Res.* **2021**, *30*, 1348-1357.

36. Shinde, A. H.; Thomas, A. A.; Mague, J. T.; Sathyamoorthi, S., Highly Regio- and Diastereoselective Tethered Aza-Wacker Cyclizations of Alkenyl Phosphoramidates. *J. Org. Chem.* **2021**, *86*, 14732-14758.

37. Kotov, V.; Scarborough, C. C.; Stahl, S. S., Palladium-Catalyzed Aerobic Oxidative Amination of Alkenes: Development of Intra- and Intermolecular Aza-Wacker Reactions. *Inorg. Chem.* **2007**, *46*, 1910-1923.

38. Minatti, A.; Muñiz, K., Intramolecular aminopalladation of alkenes as a key step to pyrrolidines and related heterocycles. *Chem. Soc. Rev.* **2007**, *36*, 1142-1152.

39. Zeni, G.; Larock, R. C., Synthesis of Heterocycles via Palladium π -Olefin and π -Alkyne Chemistry. *Chem. Rev.* **2004**, *104*, 2285-2310.

40. Weinstein, A. B.; Schuman, D. P.; Tan, Z. X.; Stahl, S. S., Synthesis of Vicinal Aminoalcohols by Stereoselective Aza-Wacker Cyclizations: Access to (–)-Acosamine by Redox Relay. *Angew. Chem. Int. Ed.* **2013**, *52*, 11867-11870.

41. Thomas, A. A.; Nagamalla, S.; Sathyamoorthi, S., Salient features of the aza-Wacker cyclization reaction. *Chem. Sci.* **2020**, *11*, 8073-8088.

42. Borkar, S. R.; Bokolia, N.; Aidhen, I. S.; Khan, I. A., Synthesis of threo- and erythro-configured trihydroxy open chain lipophilic ketones as possible anti-mycobacterial agents. *Tetrahedron: Asymmetry* **2017**, *28*, 186-195.

43. Espino, C. G.; Wehn, P. M.; Chow, J.; Du Bois, J., Synthesis of 1,3-Difunctionalized Amine Derivatives through Selective C–H Bond Oxidation. *J. Am. Chem. Soc.* **2001**, *123*, 6935-6936.

44. Henry, P. M., Palladium(II)-catalyzed exchange and isomerization reactions. III. Allylic esters isomerization in acetic acid catalyzed by palladium(II) chloride. *J. Am. Chem. Soc.***1972**, *94*, 5200-5206.

45. Overman, L. E.; Knoll, F. M., Palladium (II) - catalyzed rearrangement of allylic acetates. *Tetrahedron Lett.* **1979**, *20*, 321-324.

46. Ariza, X.; Fernández, N.; Garcia, J.; López, M.; Montserrat, L.; Ortiz, J., [3,3]-Sigmatropic Rearrangements in the Enantioselective Synthesis of (-)-Methylenolactocin. *Synthesis* **2004**, *2004*, 128-134.

47. Trost, B. M.; Lee, C. B., Geminal Dicarboxylates as Carbonyl Surrogates for Asymmetric Synthesis. Part II. Scope and Applications. *J. Am. Chem. Soc.* **2001**, *123*, 3687-3696.

48. Saito, S.; Kuroda, A.; Matsunaga, H.; Ikeda, S., Synthesis of chiral 3E,5E-octadiene-1,2R,7R,8tetraol frameworks by means of palladium(II)-promoted hetero-Claisen rearrangement: Mechanistic aspect. *Tetrahedron* **1996**, *52*, 13919-13932.

49. Shao, Q.; Wu, K.; Zhuang, Z.; Qian, S.; Yu, J.-Q., From Pd(OAc)2 to Chiral Catalysts: The Discovery and Development of Bifunctional Mono-N-Protected Amino Acid Ligands for Diverse C–H Functionalization Reactions. *Acc. Chem. Res.* **2020**, *53*, 833-851.

50. Engle, K. M.; Wang, D.-H.; Yu, J.-Q., Ligand-Accelerated C–H Activation Reactions: Evidence for a Switch of Mechanism. *J. Am. Chem. Soc.* **2010**, *132*, 14137-14151.

51. Shi, B.-F.; Maugel, N.; Zhang, Y.-H.; Yu, J.-Q., PdII-Catalyzed Enantioselective Activation of C(sp2)-H and C(sp3)-H Bonds Using Monoprotected Amino Acids as Chiral Ligands. *Angew. Chem. Int. Ed.* **2008**, *47*, 4882-4886.

Paradine, S. M.; Griffin, J. R.; Zhao, J.; Petronico, A. L.; Miller, S. M.; Christina White, M., A manganese catalyst for highly reactive yet chemoselective intramolecular C(sp3)–H amination. *Nat. Chem.* 2015, 7, 987-994.

53. Fleming, J. J.; McReynolds, M. D.; Du Bois, J., (+)-Saxitoxin: A First and Second Generation Stereoselective Synthesis. *J. Am. Chem. Soc.* **2007**, *129*, 9964-9975.

54. Elshahawi, S. I.; Shaaban, K. A.; Kharel, M. K.; Thorson, J. S., A comprehensive review of glycosylated bacterial natural products. *Chem. Soc. Rev.* **2015**, *44*, 7591-7697.

55. Montel, E.; Hrmova, M.; Fincher, G. B.; Driguez, H.; Cottaz, S., A Chemoenzymatic Route to Conjugatable (13)-Glucan Oligosaccharides*. *Aust. J. Chem.* **2009**, *62*, 575-58