Krein Support Vector Machine Classification of Antimicrobial Peptides

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Antimicrobial peptides (AMPs) represent a potential solution to the growing problem of antimicrobial resistance, yet their identification through wet-lab experiments is a costly and time-consuming process. Accurate computational predictions would allow rapid in silico screening of candidate AMPs, thereby accelerating the discovery process. Kernel methods are a class of machine learning algorithms that utilise a kernel function to transform input data into a new representation. When appropriately normalised, the kernel function can be regarded as a notion of similarity between instances. However, many expressive notions of similarity are not valid kernel functions, meaning they cannot be used with standard kernel methods such as the support-vector machine (SVM). The Kreĭn-SVM represents generalisation of the standard SVM that admits a much larger class of similarity functions. In this study, we propose and develop Kreĭn-SVM models for AMP classification and prediction by employing the Levenshtein distance and local alignment score as sequence similarity functions. Utilising two datasets from the literature, each containing more than 3000 peptides, we train models to predict general antimicrobial activity. Our best models achieve an AUC of 0.967 and 0.863 on the test sets of each respective dataset, outperforming the in-house and literature baselines in both cases. We also curate a dataset of experimentally validated peptides, measured against Staphylococcus aureus and Pseudomonas aeruginosa, in order to evaluate the applicability of our methodology in predicting microbe-specific activity. In this case, our best models achieve an AUC of 0.982 and 0.891, respectively. Models to predict both general and microbe-specific activities are made available as web applications.

1 Introduction

Kernel methods are a class of machine learning algorithms that incorporate a kernel function in order to model non-linear relationships. Standard kernel methods assume that a given kernel function is positive-definite. Those kernel functions which do not satisfy this assumption are known as indefinite kernels. The assumption of positive-definiteness is restrictive, as it limits the number of applicable functions. Recent developments in the theory of learning with indefinite kernels have now removed this requirement, allowing a much broader class of functions to be incorporated into kernel methods ^{1–3}. Leveraging these developments, we study the effectiveness of learning with established sequence-similarity functions for the classification of antimicrobial peptides (AMPs) based on their amino acid sequences. We evaluate the ability of the proposed methodology to predict both general and species-specific antimicrobial activity.

AMPs, also known as host defense peptides, are a class of evolutionary conserved molecules that form an important component in the innate immune system^{4–6}. These molecules are usually made of 12 to 50 amino acid residues, and they typically possess certain properties, including cationicity, 30-50% hydropho-

bicity, and amphiphilicity. They exhibit good antimicrobial activity against a broad range of bacteria, viruses, fungi, and parasites. In addition, they have an inherent low risk of developing antimicrobial resistance (AMR), largely attributed to their underlying rapid membrane permeabilising activity^{4,7,8}. Such broad-spectrum and rapid antimicrobial activity has prompted researchers to consider AMPs as a potential remedy to the growing problem of AMR, which is a major global health threat ^{9,10}. Nonetheless, there has so far been a lack of success in translating AMP-based therapy to clinical use, due to challenges such as complex structure-activity relationship (SAR), drug toxicity, instability in host and infective environment, and low financial incentives 11,12. Owing to the complex SAR and the costly and timeconsuming process of wet-lab experiments associated with AMP investigations, many researchers have proposed computational approaches, including molecular dynamics (MD) simulations and machine learning (ML) algorithms, to accelerate the discovery and development of potential AMPs for clinical use ^{13–19}.

Several studies have highlighted the promise of ML algorithms in predicting the antimicrobial activity, dissecting the complex SAR, and informing the drug design of AMPs ^{13–15}. A wide range of ML algorithms have been utilised, including random forests ²⁰, support vector machines (SVMs) ^{20–24} and artificial neural networks ^{20–22,25,26}. Many of these algorithms are used in combination with a carefully selected set of peptide features, which can be divided into two categories: compositional and physicochemical. The amino acid composition is the simplest example of a compositional feature, which is a vector containing counts of each amino acid in a given peptide. There are various extensions, such as the

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reduced amino acid composition ²⁷ and the pseudo amino acid composition ²⁸. When computing the reduced amino acid composition, a peptide is represented in a reduced alphabet in which similar amino acids are grouped together. The pseudo amino acid composition accounts for composition as well as sequence-order information, as this is not considered in the standard amino acid composition. The set of physicochemical features include peptide properties such as the charge, hydrophobicity and isoelectric point ^{20,24,29}. These features are typically average values of the respective properties calculated over the length of the peptide.

Classical sequence alignment algorithms, such as the Smith-Waterman³⁰ and Needleman-Wunsch³¹ algorithms, are computationally intensive and do not scale well to large problems. Many papers have advocated the use of alignment-free methods to determine sequence similarity ^{19,32–35}. The success of these endeavours notwithstanding, sequence alignment functions are effective notions of biological-sequence similarity that can reflect ancestral, structural or functional similarity and therefore should not be overlooked. Several studies have utilised sequence alignment functions for AMP prediction. For example, Wang et al. 36 and Ng et al. 37 utilised the BLAST algorithm 38 in a classification model. Whilst these approaches led to accurate models, the BLAST algorithm is a heuristic method that finds only approximate optimal alignments. Hence, it is interesting to consider whether the same approaches using the optimal alignment score would improve the models.

The SVM is a well-known ML algorithm for classification and can incorporate a kernel function in order to learn non-linear classification boundaries. The kernel function greatly influences the performance of the resulting classification model. When appropriately normalised, a kernel function can be regarded as a similarity function. A useful kernel function should produce similarities that are relevant to the problem. Many expressive notions of similarity are not valid kernel functions ^{39–42}, in that they are indefinite, meaning they cannot be used with an SVM. Recent developments have now alleviated this problem, facilitating a much larger class of similarity functions to be used in conjunction with an SVM. Loosli et al. 1 present an algorithm for learning an SVM with indefinite kernels. Their approach relies on a method of stabilisation, meaning there is no guarantee of global optimality. On the other hand, the Kreĭn-SVM³ is an algorithm for learning an SVM with indefinite kernels that is guaranteed to find a globally optimal solution.

In this work, we utilised the Kreĭn-SVM algorithm to assess the effectiveness of sequence alignment functions for AMP classification. We performed an empirical comparison of both the local alignment score and Levenshtein distance 43,44 on two AMP datasets from the literature. Furthermore, we tested the ability of our approach to detect species-specific activity on a dataset of experimentally validated peptides measured against both Staphylococcus aureus ATCC 29213 and Pseudomonas aeruginosa ATCC 27853, henceforth referred to as S. aureus ATCC 29213 and P. aeruginosa ATCC 27853. Our trained models are made available as web applications at http://comp.chem.nottingham.ac.uk/KreinAMP, for the prediction of both general and species-specific activities.

2 Methods

AMP prediction models were developed using the Kreĭn-SVM algorithm in conjunction with sequence alignment functions, which we formally define in this section. We initially describe the more familiar SVM, before moving on to the Kreĭn-SVM. We then define the Levenshtein distance ⁴⁴ and the local alignment score. Finally, we describe our computational and microbiological methodology.

2.1 SVM and Krein-SVM

2.1.1 SVM.

The SVM is a ML algorithm used for classification. The result of training an SVM is a hyperplane whose distance to the closest training instance, in either class, is maximal. Furthermore, instances from each class are required to reside on separate sides of the hyperplane. New instances are classified based solely on which side of the hyperplane they are located. The distance from the hyperplane to the closest training instance is known as the margin. The hyperplane that maximises the margin is the maximum-margin hyperplane and this is what is produced when training an SVM.

The decision surface associated to a hyperplane is inherently linear, which can be restrictive when the two classes are not linearly separable. This issue is mitigated through the use of a kernel function, which implicitly maps the instances into a new space. The space in which the instances are mapped to is known as a reproducing kernel Hilbert Space (RKHS), and every kernel function is uniquely associated to a RKHS. When incorporating a kernel function, the SVM finds the maximum-margin hyperplane in the associated RKHS and this can correspond to a non-linear decision surface in the original space. Equation (1) presents the optimisation problem that is solved when training an SVM with L2 loss, commonly known as an L2-SVM. We opted for an L2-SVM as greater penalisation is placed on instances which violate the constraints.

$$\underset{f \in \mathcal{H}, \xi \in \mathbb{R}^n}{\arg \min} \sum_{i=1}^n \xi_i^2 + \lambda \|f\|_{\mathcal{H}}^2, \tag{1}$$

subject to $y_i f(x_i) \ge 1 - \xi_i$, i = 1, ..., n.

We denote by x_i the i-th training instance and by y_i its corresponding label. Furthermore, \mathcal{H} denotes the RKHS from which a solution is found. The solution is a function $f \in \mathcal{H}$ and a vector of slack variables ξ which minimise the objective function, subject to the constraints. The constraint $y_i f(x_i) \geq 1 - \xi_i$ imposes that the i-th instance lies on the correct side of the hyperplane and that its distance to the hyperplane is greater than or equal to the margin. An instance x_j which does not satisfy this constraint contributes a value of ξ_j^2 to the objective function, where ξ_j is the distance between x_j and the margin. The $\sum_{i=1}^n \xi_i^2$ term in the objective function is the total contribution of all instances which do not satisfy the constraints. The $\|f\|_{\mathcal{H}}^2$ term is the norm of the considered function and acts as a regulariser. The hyperparameter λ can be tuned to provide a balance between the number of instances that violate the constraints and the regulariser.

2.1.2 Kreĭn-SVM.

The Kreĭn-SVM is a classification algorithm that is defined to incorporate a much broader class of kernel functions, known as indefinite kernel functions. Similarly to a standard kernel function, an indefinite kernel function implicitly maps instances into a new space. However, the space associated to an indefinite kernel function is known as a reproducing kernel Kreĭn Space (RKKS). Whilst operating in different spaces, the SVM and Kreĭn-SVM are conceptually similar. Both algorithms incorporate a kernel function, find the maximum-margin hyperplane in the associated space and are capable of learning non-linear decision surfaces.

$$\underset{f \in \mathcal{K}, \xi \in \mathbb{R}^n}{\operatorname{arg\,min}} \sum_{i=1}^n \xi_i^2 + \lambda_+ \|f_+\|_{\mathcal{H}_+}^2 + \lambda_- \|f_-\|_{\mathcal{H}_-}^2.$$
(2)

subject to
$$y_i f(x_i) \ge 1 - \xi_i$$
, $i = 1, ..., n$.

Equation (2) presents the optimisation problem that is solved when training the Kreı̆n-SVM with L2 loss. We denote by $\mathcal K$ the associated RKKS. Similarly to the SVM, the solution is a function $f \in \mathcal K$ and a vector of slack variables ξ which minimise the objective function. The first term in the objective function, as well as the constraints, have the same interpretation as in eq. (1). The only notable difference to the SVM is the method of regularisation. Any RKKS $\mathcal K$ can be expressed as a direct sum of the form

$$\mathcal{K} = \mathcal{H}_+ \oplus \mathcal{H}_-,$$

where \mathcal{H}_{\pm} are RKHSs. This means that any function $f \in \mathcal{K}$ can be decomposed as $f = f_+ \oplus f_-$, where $f_{\pm} \in \mathcal{H}_{\pm}$. Hence, regularisation in eq. (2) is performed by separately regularising each component of the decomposition. The hyperparameters λ_{\pm} can be tuned to provide a balance between the number of instances that violate the constraints and the regularisation of each decomposition component.

2.2 Sequence Similarities and Distances

We now proceed to define the sequence similarities used throughout this work. This section closely follows the works of Setubal and Meidanis 45 and Yujian and Bo 43 . First, we clarify our terminology.

2.2.1 Notation and Terminology.

Let Σ be a finite alphabet, $\Sigma^n \subseteq \Sigma$ be the set of all strings of length n from Σ and Σ^* the set of all strings from that alphabet. A string $s \in \Sigma^*$ of length n is a sequence of characters that can be indexed as $s = s_1 \dots s_n$. Given a string $s \in \Sigma^n$, we say that $u \in \Sigma^m$ is a subsequence of s if there exists a set of indices $I = \{i_1, \dots, i_m\}$ with $1 \le i_1 \le \dots \le i_m \le n$, such that $u_j = s_{i_j}$ for $j = 1, \dots, m$. We write u = s[I] for short. We say that $v \in \Sigma^I$ is a substring of s if v is a subsequence of s with index set $J = \{j_1, \dots, j_I\}$ such that $j_{r+1} = j_r + 1$ for $r = 1, \dots, l-1$. That is, v is a subsequence consisting of consecutive characters of s.

2.2.2 Global Alignments.

The goal of a sequence alignment is to establish a correspondence between the characters in two sequences. In the context of bioinformatics, a pairwise alignment can indicate ancestral, structural or functional similarities between the pair of sequences. In this section, we provide a formal review of global sequence alignment.

Definition 2.1 (Global Alignment). Let Σ be an alphabet and let $s \in \Sigma^n$ and $t \in \Sigma^m$ be two strings over Σ . Define $\Sigma_g = \Sigma \cup \{\text{``-''}\}$ as the extension of Σ with the gap character "-". The tuple $\alpha(s,t) = (s',t')$ is a global alignment of sequences s and t if and only if

- 1. $s',t' \in \Sigma_g^*$
- 2. |s'| = |t'| = l, such that $\max(n, m) \le l \le m + n$,
- 3. The subsequence of *s'* obtained by removing all gap characters is equal to *s*,
- 4. The subsequence of t' obtained by removing all gap characters is equal to t,

5.
$$\{i|s_i' = "-"\} \cap \{i|t_i' = "-"\} = \varnothing$$
.

Definition 2.1 provides a formal definition of global alignment. Whilst many possible alignments exist for two strings, the goal of sequence alignment is to find an alignment that optimises some criterion. A scoring function, presented in definition 2.2, can be used to quantify the "appropriateness" of an alignment. An optimal global alignment is then one which is optimal with respect to a given scoring function, as shown in definition 2.3.

Definition 2.2 (Scoring Functions). Let Σ be an alphabet, $\Sigma_g = \Sigma \cup \{\text{``-''}\}$ be the extension of Σ with the gap character "-" and $p: \Sigma_g \times \Sigma_g \to \mathbb{R}$ be a function defined over the elements of Σ_g . We say p is a *similarity* scoring function over Σ_g if, for all $x,y \in \Sigma$, we have

- 1. p(x,x) > 0,
- 2. p(x,x) > p(x,y),
- 3. p(x,y) = p(y,x),
- 4. $p(x, "-") \le 0$.

Similarly, we say p is a *distance* scoring function over Σ_g if, for all $x, y, z \in \Sigma$, we have

- 1. p(x,x) = 0,
- 2. p(x,y) > 0,
- 3. p(x,y) = p(y,x),
- 4. p(x, "-") > 0
- 5. $p(x,z) \le p(x,y) + p(y,z)$

Definition 2.3 (Optimal Global Alignment). Let Σ be an alphabet, $\Sigma_g = \Sigma \cup \{\text{``-''}\}$ be the extension of Σ with the gap character "-" and consider two strings $s \in \Sigma^n$, $t \in \Sigma^m$. Let $\alpha(s,t) = (s',t')$ be a valid global alignment of s and t (valid in the sense that it satisfies the conditions of definition 2.1), $p: \Sigma_g \times \Sigma_g \to \mathbb{R}$ be a scoring function over Σ_g and $\mathcal{A}(s,t)$ be the space of all valid alignments of

s and *t*. The score $S_p(\alpha(s,t))$ of $\alpha(s,t)$ with respect to the scoring function *p* is defined as

$$S_p(\alpha(s,t)) = \sum_{i=1}^l p(s_i',t_i').$$

If p is a similarity scoring function, then the optimal global alignment $\alpha^*(s,t)$ with respect to p is defined as

$$\alpha^*(s,t) = \underset{\alpha(s,t) \in \mathcal{A}(s,t)}{\arg \max} \mathcal{S}_p(\alpha(s,t)).$$

Similarly, if p is a distance scoring function then the optimal global alignment $\alpha^*(s,t)$ with respect to p is defined as

$$\alpha^*(s,t) = \underset{\alpha(s,t) \in \mathcal{A}(s,t)}{\arg \min} \mathcal{S}_p(\alpha(s,t)).$$

Since an alignment is optimal with respect to a given scoring function, it is natural to consider which scoring function to use in order to obtain the most meaningful alignments. In the context of biological sequences, researchers have been considering this problem for many years. A number of families of scoring matrices have been designed to encode useful notions of similarity. In this work, we only considered the BLOSUM62 scoring matrix ⁴⁶, as it is a standard choice when performing sequence alignment.

2.2.3 Levenshtein Distance.

The string edit distance defines a useful notion of distance between a pair of strings. It is informally defined as the minimum number of edit operations required to transform one string into another. The Levenshtein distance is a variant of the string edit distance that allows the operations of substitution, deletion and insertion of characters, and these are defined in definition 2.4.

Definition 2.4 (Elementary Edit Operations). Let Σ be an alphabet. For two characters $a,b\in\Sigma$, we denote by $a\to b$ the elementary edit operation that substitutes a with b. Denoting by ε the null character (the empty string), we can define the elementary edit operations of insertion and deletion as $\varepsilon\to b$ and $a\to\varepsilon$, respectively.

In order to define more complex transformations, one can consider the consecutive application of a sequence of elementary edit operations. Of special interest to the Levenshtein distance are those sequences of operations that transform one string into another. Such a sequence is known as an edit path and its length is defined as the number of operations in the sequence. The Levenshtein distance between two strings is defined as the length of the minimum length edit path, as seen in definition 2.5.

Definition 2.5 (Levenshtein Distance). Let Σ be an alphabet and consider two strings $s \in \Sigma^n$ and $t \in \Sigma^m$ from Σ . An edit path from s to t is denoted by $P_{s,t}$ and represents a sequence of elementary edit operations that transforms s into t. Denote by $|P_{s,t}|$ the number of operations contained in $P_{s,t}$ and by $\mathcal{P}_{s,t}$ the space of all edit paths from s to t. The Levenshtein distance $d_L(s,t)$ between s and t is defined as

$$d_L(s,t) = \min_{P_{s,t} \in \mathcal{P}_{s,t}} |P_{s,t}|.$$

Definition 2.5 shares some interesting similarities with definition 2.3. Both problems solve a combinatorial optimisation problem and, indeed, the Levenshtein distance can be realised as a special case of global alignment. More specifically, consider the distance scoring function $p: \Sigma_g \times \Sigma_g \to \{0,1\}$ defined as

$$p(x,y) = \begin{cases} 0, & \text{if } x = y \\ 1, & \text{otherwise.} \end{cases}$$

For two strings $s \in \Sigma^n$ and $t \in \Sigma^m$, let their optimal global alignment with respect to p be equal to $\alpha^*(s,t) = \{s',t'\}$. Define the set U as

$$U = \{i \mid s_i' \neq t_i'\}.$$

Then the score $S_p(\alpha^*(s,t))$ of $\alpha^*(s,t)$ with respect to p is equal to the cardinality of U. This is exactly equal to the Levenshtein distance between s and t.

2.2.4 Local Alignments.

A global alignment produces an alignment which spans the whole length of a pair of strings. It is based on the assumption that the strings are related in their entirety. This assumption can be restrictive, since it is often the case that certain substrings exhibit high similarity whilst others do not. A local alignment produces an alignment that finds those high similarity substrings. That is, it finds the highest scoring global alignment from all possible substrings of the pair of strings. We formalise this notion in definition 2.6.

Definition 2.6 (Optimal Local Alignment). Let Σ be an alphabet, $\Sigma_g = \Sigma \cup \{\text{``-''}\}$ be the extension of Σ with the gap character ``-'' and $p: \Sigma_g \times \Sigma_g \to \mathbb{R}$ be a similarity scoring function. For a string $s \in \Sigma^n$, let \mathcal{I}_s be the space of all index sets such that for any $I_s \in \mathcal{I}_s$, $s[I_s]$ is a valid substring of s. Similarly, for a string $t \in \Sigma^m$, let \mathcal{I}_t be the space of all index sets such that for any $I_t \in \mathcal{I}_t$, $t[I_t]$ is a valid substring of t. For any $I_s \in \mathcal{I}_s$ and $I_t \in \mathcal{I}_t$, denote by $\alpha^*(s[I_s], s[I_t])$ the optimal global alignment of $s[I_s]$ and $t[I_t]$ with respect to p. The optimal local alignment $\alpha_L^*(s,t)$ of s and t with respect to p is defined as

$$\alpha_L^*(s,t) = \underset{I_s \in \mathcal{I}_s, I_t \in \mathcal{I}_t}{\arg\max} S_p(\alpha^*(s[I_s],t[I_t])).$$

2.3 Computational Methodology

This section discusses the setup of our computational evaluation, as well as the datasets used.

2.3.1 Computational Setup.

To assess the usage of learning with sequence alignment functions, we performed a series of computational experiments on a number of AMP datasets. In each of our evaluations, we tested both the local alignment score (LA) and the Levenshtein distance (LEV) in conjunction with the Kreı̆n-SVM algorithm. We compared against a baseline SVM using the gapped k-mer kernel. This is a positive-definite kernel function that has produced accurate models in a number of biological-sequence classification tasks $^{47-49}$ and hence makes for a useful baseline. When applicable, we also compared our models with AMP identification tools

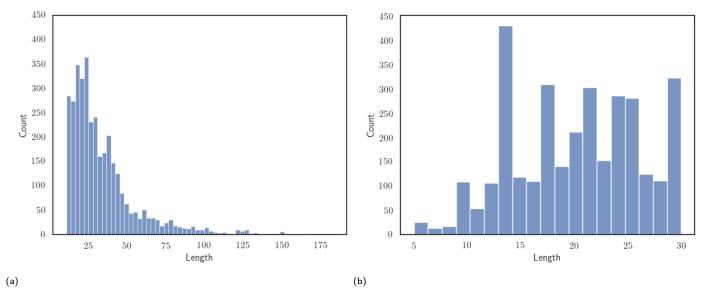


Fig. 1 The distribution of peptide lengths for (a) the AMPScan and (b) DeepAMP datasets.

from the literature. The parasail package 50 was used to compute local alignment scores. We only considered normalised variants of the local alignment score and Levenshtein distance, with the normalisation performed according to Schölkopf et al. 51 and Yujian and Bo 43, respectively. We report the accuracy and the area under the receiver operating characteristic curves (AUC) to compare models. The accuracy is defined as the proportion of correctly classified instances and the AUC as the probability that a classifier will score a randomly chosen positive instance higher than a randomly chosen negative instance 52. In order to allow for a fair comparison, all models used the same training and test splits. The optimal hyperparameters were selected by performing an exhaustive grid search over the training set, using 10-fold cross validation. The λ hyperparameter of the SVM algorithm, as well as the λ_+ and λ_- hyperparameters of the Kreĭn-SVM algorithm, were selected from {0.01,0.1,1,10,100}. The Levenshtein distance has no hyperparameters to control and we used the default values for the hyperparameters of the local alignment score. The gapped k-mer kernel has two hyperparameters g and m and is quite susceptible to their values. The optimal value of g was selected from $\{1,2,3,4,5\}$ and the optimal value of m was selected from $\{1, 2, 3, \dots, 10\}$. It is required that g > m, so only valid combinations of the two were considered. In our nested crossvalidation experiments, we used 10 inner and 10 outer folds and the reported results are averaged over the outer fold test sets.

2.3.2 General Antimicrobial Datasets.

We selected two AMP classification datasets from the literature, which we refer to as AMPScan²⁶ and DeepAMP²⁵, in order to test the ability of approach to predict general antimicrobial activity. Detailed discussions on the creation of these datasets can be found in the original studies. The AMPScan and DeepAMP datasets contain 3556 and 3246 instances, respectively. Each dataset also contains a 50:50 ratio of AMPs to non-AMPs, allowing

us to avoid issues that result from class imbalance. Associated to each dataset is a specific test set, and reporting results on this set allows comparison with the authors' proposed models. Despite being of similar size, one major differentiating factor between the two datasets is the length of peptides. Figure 1 displays the empirical distribution of peptide lengths for both datasets. DeepAMP contains generally shorter peptides than AMPScan. Indeed, the DeepAMP dataset was curated since short-length AMPs have been shown to exhibit enhanced activity, lower toxicity and higher stability as opposed to their longer counterparts ^{53,54}. More importantly, synthesis is cheaper for the short AMPs than the full-length AMPs, which increases the potential for clinical translation and commercialisation ¹².

2.3.3 Species-Specific Datasets.

To test the ability of our methodology to identify species-specific activity, we have utilised an external dataset of 16 peptides with minimum inhibitory concentration (MIC) measured against *S. aureus* ATCC 29213 and *P. aeruginosa* ATCC 27853. To make this dataset suitable for classification, we label a peptide as active if its MIC $< 100 \mu g/mL$ and inactive otherwise. Further details of the microbiological experimentation used to construct this dataset can be found in section 2.4.

Vishnepolsky *et al.* ⁵⁵ have shown that, given an appropriate training dataset, predictive models of peptide activity against specific species can be constructed. This involves training a separate model for each species of interest. Their model predicting activity against *E. coli* ATCC 25922 achieved a balanced accuracy of 0.79, which was greater than a number of common AMP prediction tools ⁵⁵. Furthermore, models to predict activity against *S. aureus* ATCC 25923 and *P. aeruginosa* ATCC 27853 were made publicly available as web-tools.

We follow the methodology of Vishnepolsky *et al.* to construct useful training datasets for our problem. We utilised the Database

Table 1 Descriptive statistics of the $\mathsf{SA}_{29213},\ \mathsf{SA}_{25923}$ and PA_{27853} datasets

| Dataset | Size | Class Ratio |
|---------------------|------|-------------|
| SA ₂₉₂₁₃ | 463 | 0.644 |
| SA ₂₅₉₂₃ | 808 | 0.646 |
| PA_{27853} | 686 | 0.547 |

of Antimicrobial Activity and Structure of Peptides (DBAASP) as a source of data. DBAASP contains peptide activity measurements against a wide-range of species 56, including those of interest to us. We extracted from DBAASP all peptides with activity measured against S. aureus ATCC 29213, S. aureus ATCC 25923 or *P. aeruginosa* ATCC 27853 subject to the following conditions: *i*) peptide length in the range [6, 18], ii) without intrachain bonds, iii) without non-standard amino acids and iv) MIC measured in μ g/mL or μ M. Condition i) was imposed as that is the range of peptide lengths in our external test set. Conditions ii) and iii) were imposed following the recommendation of the Vishnepolsky et al. and condition iv) was imposed as conversion from μM to μ g/mL is possible by estimating the molecular weight of a given sequence. Since no web-tool to predict activity against S. aureus ATCC 29213 was available, we collected data for peptides active against S. aureus ATCC 25923. This allowed us to compare our models with the state of the art provided by Vishnepolsky et al.

Three seperate datasets of peptides with activity measured against *S. aureus* ATCC 29213, *S. aureus* ATCC 25923 and *P. aeruginosa* ATCC 27853 were created using the data collected from DBAASP. We refer to these datasets as SA_{29213} , SA_{25923} , and PA_{27853} , respectively. Each dataset is constructed from highly active peptides (MIC $\leq 25\mu g/mL$) and inactive peptides (MIC $\geq 100\mu g/mL$). A peptide with $25\mu g/mL < MIC < 100\mu g/mL$ would not be included in our training dataset. This large interval allows us to account for experimental errors, which in turn increases the confidence in our class labels. In the case that a peptide was associated to multiple activity measurements, the median value was taken to represent its activity. As shown in table 1, the three training datasets are all relatively small and contain slightly more active peptides than inactive peptides.

2.4 Microbiological experiments

A previously established dataset of 16 peptides was used to test the ability of the developed ML algorithms in predicting species-specific antimicrobial activity⁷. A brief summary of the microbiological experiments in relation to this dataset is described in this section.

2.4.1 Synthesis of peptides.

All 16 peptides were commercially synthesised by Mimotopes (Mulgrave Victoria, Australia) via solid phase Fmoc synthesis method⁷. All the synthetic peptides were purified by reverse-phase high performance liquid chromatography (RP-HPLC) to >95% purity and characterised by mass spectrometry. The design of the peptides was based on the native templates of human beta-defensins (HBD)-1 to -3 and human cathelicidin (LL-37), which

are important components of AMPs.

2.4.2 Range of organisms used.

Two laboratory strains of bacteria were obtained from the American Type Culture Collection (ATCC; Virginia, U.S.A.), namely *S. aureus* ATCC SA29213 and *P. aeruginosa* ATCC PA27853, to examine the antimicrobial efficacy of the 16 synthetic peptides. The notation following species name indicates the specific strain of that species. For instance, *S. aureus* ATCC SA29213 and *S. aureus* ATCC SA25923 are two different strains of *S. aureus*.

S. aureus (a Gram-positive bacteria) and *P. aeruginosa* (a Gramnegative bacteria) were chosen for this study as they represent two of the important ESKAPE pathogens, which consist of *Enterococcus faecium*, *S. aureus, Klebsiella pneumoniae, Acinetobacteria baumannii, P. aeruginosa*, and *Enterobacter spp*. This group of pathogens have been recognised to be largely responsible for life-threatening nosocomial infections and are the key culprits in AMR-related systemic and non-systemic infections, including infectious keratitis ^{9,57–60}.

2.4.3 Minimum inhibitory concentration (MIC) assay.

The antimicrobial activity of the 16 peptides was examined using the Clinical and Laboratory Standards Institute (CLSI)-approved MIC assay with broth microdilution method. Briefly, the microorganisms were cultured on Tryptone Soya Agar (TSA) and incubated overnight for 18-21 hours at 37 °C. Three to five bacterial colonies were obtained from the agar plate and inoculated into an Eppendorf tube containing 1 mL of cation-adjusted Muller-Hinton broth (caMHB). The bacterial suspension was adjusted to achieve a turbidity equivalent to 0.1 OD600 or 0.5 MacFarland, containing 1.5×10^8 colony-forming unit (CFU)/mL, which was then further diluted in 1:150 in caMHB to reach a final bacterial concentration of 1×10^6 CFU/mL. The peptide was prepared in 1:2 serial dilution in a 96-well polypropylene microplate, with a starting concentration of 200 µg/mL and decreasing to 0.78 $\mu \text{g/mL}.$ 50 μl of 1×10^6 CFU/mL bacteria was added into each well containing of equal volume of peptide. The MIC values, defined as the lowest concentration of the antimicrobial agent that inhibits any visible bacterial growth, were determined after 24 hours of incubation with treatment.

3 Results

In section 3.1, we discuss the ability of our models to identify general antimicrobial activity. We observe that our proposed models consistently outperform the baseline and, in some cases, the models proposed in the literature. One shortcoming of any computational model that identifies general antimicrobial activity is that it cannot be used to identify activity against specific species. We address this shortcoming in section 3.2, by training our models to identify species-specific activity against S. *aureus* ATCC 29213 and P. *aeruginosa* ATCC 27853 on an experimentally validated dataset of 16 peptides, for which the proposed approach produces accurate models.

Table 2 Quality of the predictions on the AMPScan and DeepAMP datasets. The average accuracy and AUC are reported (standard deviation in parentheses), computed over the outer fold test sets of the nested cross-validation procedure. Results are presented for the Kreĭn-SVM with local alignment score (LA-KSVM), the Kreĭn-SVM with Levenshtein distance (LEV-KSVM) and the SVM with Gapped *k*-mer kernel (GKM-SVM)

| | AMPScan | | DeepAMP | |
|----------|---------------|---------------|---------------|---------------|
| Model | Accuracy | AUC | Accuracy | AUC |
| LA-KSVM | 0.920 (0.017) | 0.969 (0.006) | 0.760 (0.025) | 0.821 (0.028) |
| LEV-KSVM | 0.910 (0.021) | 0.966 (0.010) | 0.756 (0.032) | 0.819 (0.029) |
| GKM-SVM | 0.899 (0.015) | 0.957 (0.007) | 0.751 (0.032) | 0.817 (0.029) |

3.1 Identifying General Antimicrobial Activity

3.1.1 Nested Cross-Validation

The performance of the AMP classifiers on the considered datasets is reported in table 2. The results are averaged over the multiple test sets generated by nested-cross validation. On both datasets, the proposed models achieve a greater average accuracy and AUC than the baseline, with the local alignment score achieving the best values in all cases. The Welch t-test 61 , with p=0.05, is used to compare the test set AUC of our proposed models against the baseline across the outer folds of nested cross-validation. Adjusting for the testing of multiple hypotheses with the Bonferroni correction, we observe a significant difference between the mean AUC of the local alignment score and that of the baseline on the AMPScan dataset. All other comparisons, including those on the DeepAMP dataset, are not significant. The performance of all models is greater on the AMPScan dataset than on the DeepAMP dataset.

3.1.2 Predefined Test Set.

Table 3 reports the results of all models on the predefined test set associated with each dataset. For the sake of completeness, we also include the performance of the neural network-based classifiers proposed by the authors of each dataset. As for the nested cross-validation results, we observe that all models perform better on the AMPScan dataset than the DeepAMP dataset. Considering the former, the local alignment score achieves the largest accuracy and AUC, and is followed closely by the literature model. On the DeepAMP dataset, the performance is similar among all methods. The local alignment score achieves the best AUC but the Levenshtein distance achieves the best accuracy. However, the Levenshtein distance outperforms the literature model both in terms of accuracy and AUC. On both datasets, the baseline is the least predictive model. It is encouraging to observe that the sequence alignment functions can produce classifiers that match, and also outperform, the neural network-based classifiers.

3.2 Identifying Species-Specific Activity

In this section, we highlight the ability of our models to identify AMPs that are active against specific species, particularly S. *aureus* ATCC 29213 and P. *aeruginosa* ATCC 27853. Table 4 displays the accuracy on the external test set for models trained on the AMP-Scan and DeepAMP datasets. Once again, we also include the performance of the neural-network-based classifiers proposed by the authors of the AMPScan and DeepAMP datasets. We observe that the performance of all models is very poor. We noticed in our

Table 3 Quality of the predictions on the AMPScan and DeepAMP datasets. The accuracy and AUC are reported, computed on the predefined test sets. Results are presented for the Kreĭn-SVM with local alignment score (LA-KSVM), the Kreĭn-SVM with Levenshtein distance (LEV-KSVM) and the SVM with Gapped *k*-mer kernel (GKM-SVM). Results from the respective neural network-based classifiers ^{25,26} proposed by the authors of each dataset are also presented, denoted by Literature

| | AMPS | AMPScan | | DeepAMP | |
|------------|----------|---------|----------|---------|--|
| Model | Accuracy | AUC | Accuracy | AUC | |
| LA-KSVM | 0.911 | 0.967 | 0.761 | 0.863 | |
| LEV-KSVM | 0.904 | 0.960 | 0.798 | 0.860 | |
| GKM-SVM | 0.900 | 0.954 | 0.782 | 0.838 | |
| Literature | 0.910 | 0.965 | 0.771 | 0.853 | |

investigations that the majority of models predicted *active* for a large proportion of the peptides. This general poor performance is to be expected. We attribute it to the fact that these models have been trained to recognise if a peptide exhibits antimicrobial activity against any type of species. It is therefore unreasonable to assume that they are able to discriminate activity against specific species.

Table 4 Predictive accuracy of the Kreĭn-SVM with local alignment score (LA-KSVM), the Kreĭn-SVM with Levenshtein distance (LEV-KSVM) and the SVM with Gapped *k*-mer kernel (GKM-SVM) on the species-specific test sets of 16 peptides. Results from the respective neural network-based classifiers ^{25,26} proposed by the authors of each dataset are also presented, denoted by Literature. The dataset column indicates which dataset a model was trained on. The heading S. *aureus* indicates the model was predicting activity against S. *aureus* ATCC 29213 and the heading P. *aeruginosa* indicates the model was predicting activity against P. *aeruginosa* ATCC 27853

| Model | Dataset | S. aureus | P. aeruginosa |
|------------|---------|-----------|---------------|
| LA-KSVM | AMPScan | 0.312 | 0.312 |
| | DeepAMP | 0.250 | 0.250 |
| LEV-KSVM | AMPScan | 0.312 | 0.312 |
| | DeepAMP | 0.312 | 0.312 |
| GKM-SVM | AMPScan | 0.312 | 0.312 |
| | DeepAMP | 0.375 | 0.375 |
| Literature | AMPScan | 0.250 | 0.250 |
| | DeepAMP | 0.438 | 0.438 |

Table 5 displays the accuracy on the external test set for models trained on the species-specific datasets. We also present the performance of the web-tools provided by Vishnepolsky et al. 55 As mentioned in section 2.3.3, at the time of publication, there is no web-tool to predict activity against S. aureus ATCC 29213. Hence, we also provide results for models trained on SA₂₅₉₂₃ and tasked with predicting activity against S. aureus ATCC 29213. There is clearly a general improvement over the models trained on the AMPScan and DeepAMP datasets, indicating that the models have much greater discriminative power. On the SA₂₉₂₁₃ dataset, all models achieve the same accuracy. The baseline achieves the greatest AUC, but is closely followed by the Levenshtein distance. Considering the SA₂₅₉₂₃ dataset, the Levenshtein distance produces a model with the same accuracy as the web-tool but with a larger AUC. It is interesting to note that the models trained on SA₂₅₉₂₃ can still make accurate predictions on S. aureus ATCC 29213. Whilst these are two different strains, the findings suggest that the antimicrobial susceptibility to the AMPs is similar for both strains, implying similar mechanisms work in the same species. Considering the PA₂₇₈₅₃ dataset, we find that the local alignment score, Levenshtein distance and baseline produce equally accurate models, all of which are more accurate than the web-tool. However, the AUC of the local alignment score and Levenshtein distance are considerably higher than that of the baseline. Whilst it is difficult to make any strong conclusions on such a small dataset, it is still encouraging to observe that our models achieve similar, and sometimes greater, accuracy than both the baseline models and web-tools.

4 Conclusions

We have assessed the capabilities of sequence alignment functions coupled with the Krein-SVM as AMP classification models. Our investigations indicate that the proposed methodology produces accurate classifiers of both general and species-specific antimicrobial activity of AMPs. The utility of our methodology is twofold. Firstly, since sequence alignment algorithms operate directly on amino acid sequences, these methods do not explicitly require the use of peptide features. This removes the need for the practitioner to decide which features to use, which is often a detailed and time-consuming process. Secondly, in all of our experiments, we used the local alignment score with its default hyperparameters. Having achieved such promising results, it prompts the question of whether more accurate models could be attained by also tuning the various hyperparameters of the local alignment score, such as the choice of scoring function, which we will explore in future work.

As the chemical space of natural peptides is extremely large, the development of highly accurate classifiers will help accelerate the discovery and development of novel *de novo* AMPs. In addition, the promising results generated from this study open a number of possible avenues for further work. Our identification of species-specific activity could be improved using a larger external test set, allowing us to draw stronger conclusions. Furthermore, the methodology we have presented is not specific to the classification of AMPs. We suspect that the Kreĭn-SVM coupled with sequence alignment functions could be applied to other biological-

sequence classification tasks. Our computational findings demonstrate not only the feasibility of the proposed approach but more generally the utility of the Kreĭn-SVM as a classification algorithm. Its use of indefinite kernel functions provides a means for practitioners to learn from domain-specific similarity functions without the concern of verifying the positive-definite assumption. This is beneficial since it is often well beyond the expertise of the practitioner to verify this assumption. Whilst we have explored its use when combined with sequence alignment functions, there exist many more indefinite kernel functions with which the Krein-SVM could be combined. Furthermore, the theoretical insight of separately regularising the decomposition components of a function in a Kreın space could be applied to develop other indefinite kernelbased learning algorithms. A notable example that relates to the current study would be to extend the One-Class SVM 62 to incorporate indefinite kernel functions. The One-Class SVM is a kernelbased learning algorithm that performs anomaly detection ⁶³. It has previously been applied to identify the domain of applicability of virtual screening models ⁶⁴. An indefinite kernel extension of the One-Class SVM could be directly applied to estimate the domain of applicability of our models.

Author Contributions

Joseph Redshaw: conceptualisation, data curation, formal analysis, investigation, software, methodology, visualisation, writing – original draft. Darren S. J. Ting: conceptualisation, methodology, resources, writing - original draft. Alex Brown: validation, software, writing - review & editing. Jonathan D. Hirst: conceptualisation, methodology, project administration, resources, supervision, writing - review & editing. Thomas Gärtner: conceptualisation, methodology, supervision, writing - review & editing.

Conflicts of interest

A.B. is an employee and shareholder in GSK.

Data availability

The source code and data used to reproduce the computational experiments is available at https://github.com/Mrjoeybux/KreinAMP.

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Table 5 Quality of the predictions of the Kreĭn-SVM with local alignment score (LA-KSVM), the Kreĭn-SVM with Levenshtein distance (LEV-KSVM) and the SVM with Gapped k-mer kernel (GKM-SVM) on the test sets of 16 peptides. Results from the DBAASP Web-tools are also presented 55,56 . The headings in the third to fifth columns indicate which dataset the models were trained on. The models trained on both SA₂₉₂₁₃ and SA₂₅₉₂₃ were tasked with predicting activity against S. aureus ATCC 29213. The models trained on PA₂₇₈₅₃ were tasked with predicting activity against P. aeruginosa ATCC 27853

| Model | Evaluation Metric | Training Dataset | | |
|-----------------|-------------------|---------------------|---------------------|---------------------|
| | | SA ₂₉₂₁₃ | SA ₂₅₉₂₃ | PA ₂₇₈₅₃ |
| LA-KSVM | Accuracy | 0.688 | 0.688 | 0.750 |
| | AUC | 0.873 | 0.909 | 0.891 |
| LEV-KSVM | Accuracy | 0.688 | 0.875 | 0.750 |
| | AUC | 0.927 | 0.982 | 0.855 |
| GKM-SVM | Accuracy | 0.688 | 0.688 | 0.750 |
| | AUC | 0.945 | 0.891 | 0.655 |
| DBAASP Web-tool | Accuracy | - | 0.875 | 0.688 |
| | AUC | - | 0.945 | 0.718 |

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