Fluorinated Aliphatic Diazirines: Preparation, Characterization, and Photolabeling Studies

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The previously unknown difluoromethyl- and the previously neglected trifluoromethyl-aliphatic diazirines were synthesized and characterized. Model photolabeling experiments and biological studies revealed that the stable trifluoromethyl-aliphatic diazirines could indeed be safely used as photoaffinity labels, while the labile difluoromethylated analogues - could not.

N. H Ņ. H \dot{N} H [secondary amines] [¹⁹F-aliphatic diazirines] [synthesis, logD, solubility, X-Ray, pK_a, UV] N H N $N \rightarrow$ F F and analogues $CF₃$ N N[:] $\dot{\texttt{N}}$ H [bioactivity, photolabeling experiments] >100 drugs

Introduction and Aim. Aliphatic diazirines (type A, Figure 1) were discovered first in the 1960s by *Abendroth* with Henrich;^[1] *Paulsen*;^[2] and *Schmitz* with *Ohme*.^[3] Their thermal decomposition was originally studied as an approach to carbenes.^[4] Photochemical irradiation of diazirines also leads to the elimination of nitrogen and the formation of carbenes.[5] The latter can insert at the C-H and N-H, C-H, S-H bonds to cross-link with proximal species. Therefore, aliphatic diazirines have become common labels for photoaffinity studies.[6-11] It is worth noting that aliphatic diazirines tend to undergo the undesired 1,2-hydrogen rearrangement into alkenes.[12]

In 1973, Knowles studied the utility of aromatic diazirines for photolabeling.^[13] Under irradiation, however, they extensively isomerized into the more stable diazo compounds that caused the nonspecific labeling.[14] In 1980, Brunner demonstrated that the trifluoromethyl-substituted aromatic diazirines (type B, Figure 1) were more useful as photoaffinity labels, because the undesired isomerization products - CF_3 -diazo compounds $CF_3C(N_2)Ar$, were inert and did not cause the non-selective labeling.^[15] Since that time, chemists have been commonly using trifluoromethylsubstituted aromatic diazirines as labels to replace aromatic fragments in bioactive compounds for photoaffinity studies.^[16-18]

Both types of diazirines - dialkyl and trifluoromethyl-aryl (types A and B, Figure 1) - are popular today. $[9,19-23]$

In 1984, Meese showed that in strict contrast to diazomethane or alkyl diazomethanes $(CH_2N_2, \text{MeCHN}_2)$, trifluorodiazoethane $(CF₃CHN₂)$ did not react with carboxylic acids.^[24,25] Therefore, we wondered if aliphatic fluoroalkyl-substituted diazirines (type C, Figure 1) could be used as photoaffinity labels. The side photoisomerization products - fluoroalkyl diazo compounds, should not cause the undesired site-selective labeling, due to lower activity.[26]

Unexpectedly, chemists almost did not use aliphatic trifluoromethyl diazirines before. Moreover, these compounds were mostly unknown from the synthetic standpoint. We could find only three single molecules in the literature with a linear aliphatic CF_3 -diazirine motif,^[27-29] while cyclic molecules were not reported. Aliphatic difluoromethyl-substituted diazirines were completely unknown. Presumably, the lack of synthetic efforts to fluoroalkyl aliphatic diazirines led to the inability of biochemists to systematically study them, and use them as photoaffinity labels. No one made them, and no one could use them.

In this work, we addressed this gap in chemistry: we elaborated a general practical method for trifluoromethyl and difluoromethyl aliphatic diazirines, characterized them and studied them in model photolabeling and biological experiments (Figure 2). Moreover, we have shown that the undeservedly neglected before aliphatic trifluoromethyl diazirines could be safely used as photoaffinity labels, while the difluoromethylating analogues – cannot, due to chemical instability.

Results and Discussion. Azetidine, pyrrolidine, and piperidine are among the most popular secondary amines in drug discovery.^[30,31] Scientists use these amines in almost every medicinal chemistry project. In addition, they are a part of more than one hundred drugs.^[32] Therefore, it is even hard to believe that in the $21st$ century, the corresponding CF₃-/CHF₂-diazirines for replacing azetidine, pyrrolidine, and piperidine in bioactive compounds for subsequent photoaffinity studies, remain unknown. During the past years, we often received requests from pharmaceutical companies on those structures.[33] Therefore, we needed to elaborate first on a general practical synthetic route to access them.

Figure 1. Diazirines: state of the art. ^aSubstructure search at Reaxys db (1.02.2022). ^bManuscripts and patents.

Figure 2. Motivation and aim of this work.

1) Synthesis. We started our synthetic efforts by trying to reach the simplest representative of this type – the derivative of azetidine (Scheme 1). Swern oxidation of the alcohol group in the commercially available compound 1a, followed by fluoridemediated nucleophilic addition of $CF₃TMS^[34]$ to the intermediate aldehyde gave the substituted trifluoroethanol 1b in 79% yield (Scheme 1). Oxidation of the latter gave ketone 1c that existed in the equilibrium with the corresponding hydrate. The obtained mixture of products was directly treated with hydroxylamine under heating to obtain oxime 1a in a 40% yield. O-Tosylation smoothly afforded product 1e in 85% yield. Reaction with liquid ammonia and oxidation of the crude intermediate with molecular iodine afforded the needed N-Boc substituted CF_3 -diazirine 1f in 70% yield. Finally, acidic cleavage of the N-Boc protecting group

Scheme 1. Synthesis of CF₃-diazirines 1g·HCl-11g·HCl. ^aTotal yield.

gave the desired amine hydrochloride 1g as a white crystalline gave the desired amine hydrochloride 1g as a white crystalline solid. All synthetic steps were optimized, and we could subsequently synthesize amine $1g$ in a five-gram scale in a single run. The structure of 1g was confirmed by crystallographic analysis (Figure 3).^[35]

Following the elaborated strategy, a library of CF_3 -diazirine containing isomeric/homologous pyrrolidines 2g-5g and piperidines 6g-11g was rapidly synthesized from the corresponding piperidines $6g-11g$ was rapidly synthesized from the corresponding commercially available alcohols (please, see SI for details). O -Mesylation (3e-5e, 7e, 9e-11e) was shown to work equally well as O-tosylation (1e, 2e, 6e, 8e) and we used both methods allowing some flexibility. Structures of amines 4g and 10g were confirmed by crystallographic analysis (Figure 3).^[35]

In addition, we performed the synthesis of linear derivatives of CF_3 -diazirine-substituted ethylamine and propylamine (Scheme 2). From the commercially available epoxide 12, the needed amine 13 was obtained in nine steps. Alternatively, from fluoral 14, amine 15 was synthesized in ten steps.

2) Scale. Original syntheses of all compounds were performed on a milligram scale. However, after validation and optimization of on a milligram scale. However, after validation and optimization of the protocol, all amines 1g-11g, 13, and 15 were synthesized in 5-30 gram amounts in a single run (Scheme 1, Scheme 2).

 70% 1e
-------------- Scheme 2. Synthesis of CF₃-diazirines 13 HCl and 15 HCl.

 N and 10b⋅HCl.³⁵ Hydrogen and chlorine atoms are omitted for N^2 clarity H claimy. N right *J. A-Kay* crystal subcure of diaziriles 10 fK N Figure 3. X-Ray crystal structure of diazirines 1b HCl , 4b HCl , clarity.

 $N = N$ $N = N$ CF_3 crystalline white solids. They were air-stable and moisture-stable. CF_3 $\downarrow \downarrow \downarrow$ All compounds 1g·HCl-11g·HCl, 13·HCl, and 15·HCl were [piperidine es] decomposition into nitrogen and carbenes. Therefore, we next $N-N$ checked the stability of the synthesized trifluoromethyl diazirines. $\frac{\%^{a}}{\%}$ (12 g; 27%^a) 3) Chemical stability. Aliphatic diazirines undergo thermal (¹³⁾ For $\begin{pmatrix} 18 \\ 16 \\ 19 \\ 19 \\ 109 \\ 108 \\ 109 \\ 109 \\ 100 \\ 100 \\ 101 \\ 102 \\ 103 \\ 104 \\ 105 \\ 106 \\ 107 \\ 108 \\ 109 \\ 101 \\ 101 \\ 102 \\ 103 \\ 104 \\ 105 \\ 106 \\ 107 \\ 108 \\ 109 \\ 101 \\ 101 \\ 102 \\ 103 \\ 104 \\ 105 \\ 106 \\ 107 \\ 108 \\ 109 \\ 101 \\ 101 \\ 1$) three months and observed no detectable decomposition according We stored them at room temperature in closed vials on the shelf for to ${}^{1}H$ NMR. Upon heating below 100 °C, all compounds also remained stable. Visual decomposition was observed at temperatures >120 °C with the elimination of nitrogen. Treatment of representative amines 2g, 6g, and 8g with aq. 1M hydrochloric acid, or aq. 1M sodium hydroxide at room temperature for one hour also did not lead to any decomposition.

CF₃

The CF₃ (Fig. 10 (Fig. 4) Basicity of amines. Having synthesized the needed products, we next studied the influence of the trifluoromethyl-diazirine fragment on the basicity of the nitrogen atom. For that, we experimentally measured pK_a values of hydrochlorides 1g-11g by standard titration method (Figure 4). Incorporation of the trifluoromethyl-diazirine unit at the β-position of the azetidine ring decreased its basicity by more than two orders of magnitude: pK_a $(16 \text{ HCl}) = 11.3 \text{ vs } pK_a$ $(1g \text{ HCl}) = 8.8$. A similar effect was $(16 \text{ HCl}) = 11.3 \text{ vs } pK_a$ $(1g \text{ HCl}) = 8.8$. A similar effect was observed during the incorporation of the trifluoromethyl-diazirine unit at the β -position of the pyrrolidine ring – basicity of the nitrogen atom decreased by more than two orders of magnitude: pK_a (17·HCl) = 11.3 *vs* pK_a (2g·HCl) = 9.1. Introduction of the pK_a (17 HCl) = 11.3 *vs* pK_a (2g HCl) = 9.1. Introduction of the trifluoromethyl-diazirine fragment at the α-position of the pyrrolidine ring led to an even more dramatic increase of basicity by more than four orders of magnitude: pK_a (17*HCl) = 11.3 *vs* pK_a (4g·HCl) = 7.1. A similar trend was observed with piperidine pK_a (**4g**:HCl) = 7.1. A similar trend was observed with piperidine (**18**): incorporation of the trifluoromethyl-diazirine unit at the most distal γ-position (6g) led to a decrease in basicity by ca. one magnitude of order, at the β -position $(8g)$ – by two magnitudes, at the α-position (10g) – by more than four magnitudes of order: pK_a $(19 \text{-} HCl) = 11.2 \text{ vs } pK_a (6g \text{-} HCl) = 9.7 \text{ vs } pK_a (8g \text{-} HCl) = 9.1 \text{ vs } pK_a$ $(10g·HCl) = 6.9.$

Figure 4. ${}^{\text{a}}$ Experimental p K_{a} values of amine hydrochlorides.

In a brief summary, two conclusions can be made:

1. Incorporation of the trifluoromethyl-diazirine fragment at the structure of azetidine, pyrrolidine, and piperidine leads to a significant decrease in the basicity of the nitrogen atom with ΔpK_a = -(1.5-4.3). These experiments indicate that trifluoromethyldiazirine substituent is strongly electron-withdrawing, similar to other fluoroalkyl groups.[36,37]

2. Even though the basicity of the nitrogen atom in 1g-11g is reduced compared to the original secondary amines, it is still sufficient for the standard amide coupling – the most popular reaction in medicinal chemistry (see Table 1).^[38]

5) Physicochemical properties. We also studied the effect of the trifluoromethyl-diazirine fragment on solubility and lipophilicity of azetidine, pyrrolidine, and piperidine. Therefore, we first synthesized model compounds 19-27 (Table 1) by standard acylation with para-phenyl benzoic acid. Indeed, despite reduced basicity, all amines 1g-11g were compatible with the standard protocol of amide coupling under standard conditions (please, see SI). Incorporation of the trifluoromethyl-diazirine unit into model compounds 19, 21 and 24 significantly decreased their water solubility: 78-331 μM (19, 21, 24) *vs* <10 μM (20, 22, 23, 25, 26, 27) (Table 1). Analogously, lipophilicity (logD) of the synthesized trifluoromethyl-diazirine containing derivatives was higher: 3.0-3.3 (19, 21, 24) vs 3.9-5.0 µM (20, 22, 23, 25, 26, 27).

As a short summary, incorporation of the trifluoromethyldiazirine fragment into the structure of azetidine, pyrrolidine, and piperidine decreased their solubility and increased lipophilicity.

6) Aliphatic difluoromethyl diazirines. Next, we wondered if the strategy for aliphatic trifluoromethyl diazirines could also be extended to the difluoromethylated analogues. Importantly, aliphatic CHF₂-diazirines were unknown in the literature before. As a representative example, we first synthesized the azetidine derivative. Commercially available amino acid 28a was converted first into the difluoromethyl ketone 28c following literature protocol (Scheme 3).^[39] Subsequent reaction of the latter with hydroxylamine gave oxime 28d in 69% yield. O-mesylation (28e), reaction with liquid ammonia, and oxidation provided N-Boc diazirine 28f in 50% yield. Final acidic deprotection of N-Boc gave the needed CHF₂-azetidine $28g$ HCl in 22% yield.

Table 1. Experimental lipophilicity (logD) and water solubility of model compounds 19-27.

 a^2 Experimental *n*-octanol/water distribution coefficient (log) at pH 7.4; ^bKinetic aqueous solubility (μ M) in 50 mM phosphate buffer (pH 7.4).

Synthesis of the difluoromethyl piperidine 29g·HCl was also analogously performed following this strategy from carboxylic acid 29a in 19% overall yield and 20 g scale (Scheme 3).

Scheme 3. Synthesis of CHF_2 -diazirines **28g** HCl and **29g** HCl.

7) Model photolabeling experiments. To check if fluoroalkyl aliphatic diazirines could be used as photoaffinity labels, we performed next several standard photolabelling experiments. For that, we synthesized a model compound 30 by N-tosyl protection of trifluoromethyl piperidine 6g (please, see SI; Scheme 4). N-Ts group was specifically chosen so that we could quantitatively analyze the reaction mixtures by LC-MS with UV detection at 254 nm (absorbance of the phenyl ring in the tosyl group).

Irradiation of diazirine 30 in methanol at 350 nm for 15 minutes led to the predominant formation of alkene 31 with the only minor formation of the needed O-H insertion products 32 and 33 (Scheme 4). At 310 nm, the formation of 13% of the needed insertion product 31 was already observed. Both compounds 31 and 32 were isolated and fully characterized to additionally confirm their structure that was originally proposed based on LC-MS data. Irradiation of diazirine 30 in the acetonitrile-water mixture (9:1) at 310 nm already gave a significant amount of the needed O-H insertion product 33. Moreover, irradiation at 350 nm provided >40% of insertion products 33 and 34 (via Ritter-type reaction). The latter experiment clearly demonstrated the full compatibility of the aliphatic trifluoromethyl diazirine motif with photoaffinity experiments.

Additionally, another model compound 35 with improved water solubility^[40] was synthesized (Scheme 4). Its irradiation in methanol under different wavelengths gave mainly alkene 36. However, irradiation in acetonitrile-water mixtures already gave >30% of the needed insertion products 38, 39. Importantly, during experiments with aliphatic trifluoromethyl diazirines, the trifluoromethyl group was stable, and we did not observe its cleavage.

Scheme 4. Photolabelling experiments of CF_3 -diazirines 30, 35 in different solvents. LC-MS data is shown. $c = 0.005$ M.

To check the compatibility of aliphatic difluoromethyl diazirines with photoaffinity studies, we synthesized model compound 40 (please, see SI; Scheme 5). Its irradiation in methanol unexpectedly gave a complex mixture, where we could identify five compounds 41-45 based on the LC-MS data. In three of them (41, 44, 45), the C-F bond was cleaved due to the solvolysis. The formation of undesired fluoroalkenes 42 and 43 was also observed. The needed O-H insertion products with the difluoromethyl group were not detected. A similar situation was observed after irradiation of diazirine 40 in acetonitrile-water mixtures. Extensive cleavage of the C-F bond was seen (41, 46) with the predominant formation of the undesired alkenes (41-43). Again, we did not observe the needed O-H insertion products with the difluoromethyl group.

Scheme 5. Photolabeling experiments of CHF₂-diazirine 40 in different solvents. LC-MS data is shown. Isomeric alkenes 42 and 43 have identical m/z , and their structures were arbitrarily assigned.

In a short summary, experiments with model compounds 30, 35 revealed full compatibility of aliphatic trifluoromethyl diazirines with photoaffinity studies. Under non-optimized conditions, >40% of the O-H insertion products were achieved under irradiation at 350 nm. During the irradiation, the trifluoromethyl group was stable. At the same time, under identical conditions, the difluoromethyl diazirine motif was chemically labile due to extensive C-F bond solvolysis.

8) Incorporation into the bioactive compound. Finally, we needed to show the utility of aliphatic CF₃-diazirines for medicinal chemistry projects. Therefore, we incorporated representative CF₃diazirine into a bioactive compound. Previously, sulfonamide 47 was identified as a promising broad-spectrum inhibitor of cancer cell lines (Table 2).^[41] Here, we replaced the fragment of piperidine with the corresponding CF_3 -diazirine $6g$ to obtain compound 48 (please, see SI for details). Both compounds were tested for their in vitro antitumor activity against a panel of sixty cancer cell lines (please, see SI).^[42] Results against five selected tumor cell lines are summarized in Figure 4. Original compound 47 inhibited the growth of all five tumor cells K-562, NCI-H522, HCT-15, SW-620, OVCAR-3 (Table 2). It's CF_3 -diazirine derivative 48 was also active against three cell lines NCI-H522, HCT-15, SW-620. These results demonstrated that the trifluoromethyl-diazirine fragment was compatible with the bioactive compound, did not significantly affect its bioactivity, and hence could be used in medicinal chemistry projects.

Table 2. In vitro inhibition of growth of cell cancer lines (%) with compounds 47 and 48 ($c = 10 \mu M$). Single-dose experiment. (green color) - inhibition of tumor growth.

Summary. In the 1960s, pioneering studies of *Abendroth* with Henrich;^[1] Paulsen;^[2] and Schmitz with Ohme^[3] showed that diazirines could form reactive carbenes upon heating or exposure to light. Since then, diazirines became useful labels in photoaffinity studies of bioactive compounds.^[9,10,18-23] Especially popular today are dialkyl diazirines and trifluoromethyl aromatic diazirines (types A, B in Figure 1). Herein, we prepared the previously unknown difluoromethyl- and the previously neglected trifluoromethylaliphatic diazirines (types C, D in Figure 1), and comprehensively characterized them. Model photolabeling experiments and biological studies indicated that trifluoromethyl aliphatic diazirines could indeed be used as photoaffinity labels. At the same time, the difluoromethyl aliphatic diazirines were chemically labile due to C-F bond solvolysis.

We expect that with these results, the previously neglected CF_3 aliphatic diazirines will soon be commonly used by chemists and biochemists in photoaffinity labeling studies, in the very same way as dialkyl diazirines and trifluoromethyl aromatic diazirines are already being used.

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Keywords: diazirine • photoaffinity • labeling • trifluoromethyl • difluoromethyl

References

- [1] Abendroth, H. J.; Henrich, G. Angew. Chem. 1959, 71, 283−283.
- [2] Paulsen, S. R. Angew. Chem. 1960, 72, 781-782.
- [3] Schmitz, E.; Ohme, R. Chem. Ber. 1961, 94, 2166-2173.
- [4] Schmitz, E.; Habisch, D.; Stark, A. Angew. Chem. Int. Ed. 1963, 2, 548-548.
- [5] Liu, M. T. H. Chem. Soc. Rev. 1982, 11, 127−140.
- [6] Vila-Perello, M.; Pratt, M. R.; Tulin, F.; Muir, T. W. J. Am. Chem. Soc. 2007, 129, 8068−8069.
- [7] Shi, H.; Zhang, C. J.; Chen, G. Y.; Yao, S. Q. J. Am. Chem. Soc. 2012, 134, 3001−3014.
- [8] West, A. V.; Muncipinto, G.; Wu, H.-Y.; Huang, A. C.; Labenski, M. T.; Jones, L. H.; Woo, C. M. J. Am. Chem. Soc. 2021, 143, 6691–6700.
- [9] Das, J. Chem. Rev. 2011, 111, 4405-4417.
- [10] Zhang, T.; Ondrus, A. E. Synlett 2021, 32, 1053-1059.
- [11] Ziemianowicz, D. S.; Bomgarden, R.; Etienne, C.; Schriemer, D. C. J. Am. Soc. Mass Spectrom. 2017, 28, 2011– 2021.
- [12] Modarelli, D. A.; Morgan, S.; Platz, M. S. J. Am. Chem. Soc. 1992, 114, 7034−7041.
- [13] Smith, R. A. G.; Knowles, J. R. J. Am. Chem. Soc. 1973, 95, 5072−5073.
- [14] Smith, R. A. G.; Knowles, J. R. J. Chem. Soc., Perkin Trans. 2 1975, 7, 686−694.
- [15] Brunner, J.; Senn, H.; Richards, F. M. J. Biol. Chem. 1980, 255, 3313−3318.
- [16] Nassal, M. J. Am. Chem. Soc. 1984, 106, 7540-7545.
- [17] Šimon, P.; Knedlík, T.; Blažková, K.; Dvořáková, P.; Březinová, A.; Kostka, L.; Šubr, V.; Konvalinka, J.; Šácha, P. ACS Chem. Biol. 2018, 13, 12, 3333–3342.
- [18] (a) Geri, J. B.; Oakley, J. V.; Reyes-Robles, T.; Wang, T.; McCarver, S. J.; White, C. H.; Rodriguez-Rivera, F. P.; Parker, D. L.; Hett, E. C.; Fadeyi, O. O.; Oslund, R. C.; MacMillan, D. W. C. Science 2020, 367, 1091-1097. (b) Musolino, S. F.; Pei, Z.; Bi, L.; DiLabio, G. A.; Wulff, J. E. Chem. Sci. 2021, 12, 12138-12148.
- [19] Blencowe, A.; Hayes, W. Soft Matter 2005, 1, 178–205.
- [20] Dubinsky, L.; Krom, B. P.; Meijler, M. M. Bioorg. Med. Chem. 2012, 20, 554−570.
- [21] Halloran, M. W.; Lumb, J.-P. Chem. Eur. J. 2019, 25, 4885- 4898.
- [22] Ge, S.-S.; Chen, B.; Wu, Y.-Y.; Long, O.-S.; Zhao, Y.-L.; Wang, P.-Y.; Yang, S. RSC Adv. 2018, 8, 29428–29454.
- [23] Hill, J. R.; Robertson, A. A. B. J. Med. Chem. 2018, 61, 6945–6963.
- [24] Meese, C. O. Synthesis 1984, 1041-1042.
- [25] Mykhailiuk, P. K. Chem. Rev. 2020, 120, 12718-12755.
- [26] Mertens, L.; Koenigs, R. M. Org. Biomol. Chem. 2016, 14, 10547-10556.
- [27] Yip, G. M. S.; Chen, Z.-W.; Edge, C. J.; Smith, E. H.; Dickinson, R.; Hohenester, E.; Townsend, R. R.; Fuchs, K.; Sieghart, W.; Evers, A. S.; Franks, N. P. Nature Chem. Biol. 2013, 9, 715–720.
- [28] Masuda, Y.; Aoyama, K.; Yoshida, M.; Kobayashi, K.; Ohshiro, T.; Tomoda, H.; Doi, T. Chem. Pharm. Bull. 2016, 64, 754−765.
- [29] The use of aliphatic CF_3 -diazirines is also briefly mentioned in two patents: (a) patent US2016307747A1; (b) patent CN112358414A.
- [30] Taylor, R. D.; MacCoss, M.; Lawson, A. D. G. J. Med. Chem. 2014, 57, 5845−5859.
- [31] Vitaku, E.; Smith, D. T.; Njardarson, J. T. J. Med. Chem. 2014, 57, 10257−10274.
- [32] The search was performed at www.drugbank.ca.
- [33] Authors are employees of Enamine Ltd that is a supplier of building blocks for pharmaceutical and agrochemical companies.
- [34] Prakash, G. K. S.; Krishnamurti, R.; Olah, G. A. J. Am. Chem. Soc. 1989, 111, 393–395.
- [35] CCDC numbers: 2085823 (1g·HCl); 2085822 (4g·HCl); 2093700 (10g·HCl).
- [36] Huheey, J. E. J. Phys. Chem. 1965, 69, 3284-3291.
- [37] M. Morgenthaler, E. Schweizer, A. Hoffmann-Roeder, F. Benini, R. E. Martin, G. Jaeschke, B. Wagner, H. Fischer, S. Bendels, D. Zimmerli, J. Schneider, F. Diederich, M. Kansy, K. Muller. ChemMedChem 2007, 2, 1100-1115.
- [38] J. Boström, D. G. Brown, R. J. Young, G. M. Keserü Nat. Rev. Drug Discov. 2018, 17, 709–727.
- [39] Miele, M.; Citarella, A.; Micale, N.; Holzer, W.; Pace, V. Org. Lett. 2019, 21, 8261-8265.
- [40] Water solubility of organic molecules could be significantly inhanced by introduction of the P(O)Me₂-group: Stambirskyi, M. V.; Kostiuk, T.; Sirobaba, S. I.; Rudnichenko, A.; Titikaiev, D. L.; Dmytriv, Y. V.; Kuznietsova, H.; Pishel, I.; Borysko, P.; Mykhailiuk, P. K. J. Org. Chem. 2021, 86, 12783–12801.
- [41] Shablykin, O. V.; Kornii, Y. E.; Dyakonenko, V. V.; Shablykina, O. V.; Brovarets, V. S. Curr. Chem. Lett. 2019, 8, 199-210.
- [42] Biological experiments were performed at the National Cancer Institute, USA, within the framework of Developmental Therapeutic Program (http://dtp.cancer.gov).