Unexpected isomerization of oxetane-carboxylic acids

Bohdan Chalyk,^{1,2} Anastasiia Grynyova,¹ Kateryna Filimonova,¹ Tymofii V. Rudenko,^{1,2} Dmitry Dibchak,¹ Pavel K. Mykhailiuk¹*

Correspondence: Pavel.Mykhailiuk@gmail.com

¹Enamine Ltd.; Chervonotkatska 78, 02094 Kyiv (Ukraine), www.enamine.net; www.mykhailiukchem.org

² Institute of Organic Chemistry, National Academy of Sciences of Ukraine, Murmanska St. 5, 02094Kyiv (Ukraine).



Dedicated to people of Ukraine

ABSTRACT: We unexpectedly discovered that popular oxetane-carboxylic acids are intrinsically unstable. They easily isomerize into lactones under storage at rt, or under slight heating. Chemists should keep in mind the high instability of these common molecules, as this could dramatically affect the reaction yields and lead to negative results (especially in those reactions that require heating).

Introduction. More than a decade ago, oxetanes received a second breath, when they were shown to mimic a carbonyl group in bioactive compounds (Scheme 1). Moreover, the oxetane fragment was demonstrated to increase water solubility, improve metabolic stability, and lower the lipophilicity of organic molecules.¹ Since that time oxetanes have been gaining popularity in different areas of chemistry including organic synthesis, chemical biology, and medicinal chemistry.²⁻⁴ In particular, oxetane-carboxylic acids have been mentioned in >200 peer-reviewed manuscripts and patents as both bioactive compounds⁵ and starting materials in synthesis (Scheme 1).⁶ Not surprising that during the past years, we received many requests for their preparation. Some oxetanecarboxylic acids were known in the literature, some of them we needed to synthesize for the first time. Over time, however, we realized that many oxetane-carboxylic acids were intrinsically unstable - they easily isomerized into lactones under storage at room temperature or under slight heating. Here, we want to disclose this unknown in the literature phenomenon, as chemists keep on using these molecules (Scheme 1), without realizing that many of them are unstable, as that could dramatically affect reaction yields and lead to negative results (especially in those reactions that require heating).

Intended intramolecular isomerizations of oxetanes by nucleophiles were reported in the literature (Scheme 1).^{7,8} In most cases these reactions required additional activation of the oxetane ring by Lewis-acid catalysts (In, Sc, Fe, BF₃, Co, Pd, phosphoric acids, *etc*). In our case, many oxetane-carboxylic acids easily isomerized under simple storage or slight heating and required no external catalysis.

Results and Discussion. Unexpected discovery. Previously, we developed an approach to spirocyclic pyrrolidines via [3+2]-cycloaddition.⁹ As a part of this project, we synthesized several oxetane-carboxylic acids. In particular, alkali saponification of ester 1 followed by acidification with NaHSO₄ gave crude acid 1a. At that time, we obtained pure product 1a by simple washing the crude product with MeOtBu to remove soluble impurities. Inspection of this product three months later revealed the presence of ca. 25% of the individual impurity according to ¹H NMR. After one year of storage at room temperature, ca. 50% of this impurity was present. Simple heating of this mixture in isopropanol led to the exclusive formation of the "impurity" that was isolated and identified as lactone 1b. At that point, we did not pay much attention to that observation. Several months later, however, we received a request from a pharmaceutical company on the synthesis of oxetane-carboxylic acid 2a (Scheme 2). This acid was known in the literature^{6e,10} and scientists often used it in amide coupling.^{6e,11} Synthesis of **2a** was described in a patent,^{10a} and we followed the procedure: hydrogenation of alkene 2 (obtained in one step from 3-oxetanone) using palladium on charcoal in methanol smoothly gave the desired product 2a. However, an inspection of this product by ¹H NMR after one week revealed the presence of ca. 7% of an impurity. After one month of storage at room



Scheme 1. Oxetane-carboxylic acids in organic synthesis and medicinal chemistry. Aim of this work.



Scheme 2. Unexpected isomerization of oxetane-carboxylic acids 1a, 2a under storage at room temperature.

temperature, already 16% of this impurity was present. After one year of storage, the compound completely isomerized into the "impurity." We isolated and identified it as lactone **2b**. Moreover, even under slight heating at 50 °C in dioxane/water mixture, acid **2a** cleanly isomerized into lactone **2b**. At this point, it became clear that a tendency of oxetane-carboxylic acids to isomerize into lactones has a general character. We were very much surprised because while chemists have been actively using oxetane-carboxylic acids (including **2a**) in the research (Scheme 1), we could not find any systematic studies of that phenomenon. Only one example of such transformation was found in a patent with no detailed experimental data.¹²

Scope and stability. We decided next to inspect all oxetanecarboxylic acids that we had on stock. Most of these compounds were synthesized by simple saponification of ethyl/methyl esters with NaOH and acidification with NaHSO4. These results are summarized in Scheme 3. Acids 3a-10a were obtained from esters 3-10. They were stable upon storage at room temperature, and after one year we did not observe any decomposition according to ¹H NMR. Presumably, bulky (hetero)aromatic substituents (3a-7a), zwitterionic structures (8a, 9a), or polycyclic conformationally rigid core (10a) stabilized these molecules. Nevertheless, compounds 3a-7a, 9a, 10a isomerized into lactones 3b-7b, 9b, 10b under heating in dioxane/water at 100 °C. Zwitterionic acid 8a remained stable. It seems that the high basicity of the imidazole group prevented an intramolecular protonation of the oxetane-ring by the carboxylic group (intermediate A in Scheme 1), stabilizing thereby the compound. The strained structure of core 10a led to additional hydrolytic cleavage of the tetrahydrofuran ring. The structure of product 10b was proven by crystallographic analysis (Figure 1).13 Importantly, all isomerizations took place under simple heating with no external activation of the oxetane ring by HCl, HBr, or other Lewis acids.

Next, we examined reaction mixtures after hydrolysis of ethyl esters 1, 11-19 with NaOH at room temperature and acidification. We were surprised to find out that in all cases,

>20% of lactone was already present. Moreover, with each compound that percentage significantly varied from 20% to 70% depending on a synthesis run. Finally, we understood that isomerization must have been taking place during evaporation of the solvent (EtOAc) on a rotary evaporator (extraction of the product after acidification). Even though we typically heated external water batch at ca. 40 °C, it was enough for the cyclization to happen. Indeed, heating of all reaction mixtures after saponification in dioxane-water at 50 °C, smoothly completed the isomerization, and the corresponding pure lactones 1b, 2b, 11b-19b were obtained (Scheme 3). The structure of product 18b was proven by crystallographic analysis (Figure 1).13 Interestingly, in contrast to unstable nonfluorinated analogs, fluorine-containing acids 20a-22a were stable under storage. After one year at room temperature on the shelf, we did not observe their decomposition according to ¹H NMR. However, all three acids easily isomerized into the corresponding lactones 20b-22b under heating at 50 °C in dioxane-water.

Next, we studied the stability of higher homologs of oxetanecarboxylic acids (Scheme 4). Acids 23a, 24a (obtained by standard saponification of the corresponding ethyl esters 23, 24) with bulky (hetero)aromatic substituents were stable under storage at room temperature at least for one year. Isomerization of 23, 24 into valerolactones 23b, 24b took place only under prolonged heating in dioxane-water at 100 °C. The structure of product 23b was proven by crystallographic analysis (Figure 1).¹³ However, hydrolysis of alkyl ester **25** already gave a mixture of lactone and acid. The same trend was observed with alkyl esters 26-29. Content of lactones varied dramatically in each case depending on the synthesis run, indicating again that the cyclization must have been taking place under heating during the evaporation of the solvent. Additional heating of those mixtures in dioxane-water gave valerolactones 25b, 26b, and dioxanones 27b-29b.

Acids **30a-34a** with bulky substituents were stable under storage (Scheme 4). Moreover, heating of acids **33a** and **34a** in dioxane-water did not lead to isomerization – both compounds remained intact. Acids **31a**, **32a** under slight heating, however, gave the corresponding dioxanones **31b**, **32b**.

It was interesting to discover a dramatically different stability of structurally similar acids **35a** and **36a**. Pure compound **36a** was obtained by saponification of ethyl ester **36** under standard conditions. It was stable under storage, and under heating at 100 °C overnight did not isomerize at all. Presumably, an intramolecular hydrogen bonding between the N-H and the carbonyl group (**36a** is a derivative of 2-aminobenzoic acid) "freezes" the conformation and stabilizes hereby the molecule. Ester **35**, however, after hydrolysis with NaOH provided ca. 1/1 mixture of acid **35a** and lactone **35b**. Additional heating of this mixture at 50 °C led to complete isomerization into product **35b**.



Scheme 3. *Reaction conditions*. Group A: dioxane/water (10/1), 100 °C, 12h; Group B: dioxane/water (10/1), 50 °C, 12h. ^areaction time 48h; ^bWater, 100 °C, 12h; ^c*i*PrOH, 82 °C, 10h; ^dDioxane/water (10/1), 50 °C, 12h. ^e**21a** was obtained from **20a** with *m*CPBA.



Scheme 4. *Reaction conditions*. Group A: dioxane/water (10/1), 100 °C, 12h; Group B: dioxane/water (10/1), 50 °C, 12h. ^aMeOH, reflux, 4h; ^bdioxane/water (10/1), 100 °C, 12h; ^cMeOH, 50 °C, 12 days; ^ddioxane/water (10/1), 100 °C, 48h. ^ereaction time 10 days; ^freaction time 16 days; ^greaction time 7 days. ^hdioxane/water (10/1), 50 °C, 48h.

In addition, we also observed similar isomerization from another project. When we treated bromide **37** with *t*BuLi in THF at -78 °C followed by the addition of dry ice, the crude lithium salt of the corresponding carboxylic acid was obtained (Scheme 4). However, careful acidification of the salt with NaHSO₄ led to the immediate formation of the bicyclic product **37b**, which was isolated in 53% yield. Formation of the eliminated alkene was also observed, but the expected acid **37a** was not seen.



Figure 1. X-Ray structure of compounds 10b, 18b, 22b, 23b.¹³

Mechanism and kinetics. Ring-opening of oxetanes usually requires activation of the four-membered ring by Brønsted acids.^{7,8} In our case, isomerization required no catalysis. Presumably, the internal carboxylic group activated the oxetane ring via the formation of intermediate *Int-1* (Scheme 5). This intermediate is dually activated towards isomerization: it has a protonated oxetane ring and an already deprotonated (more nucleophilic) carboxylate group. Indeed, in isopropanol under heating, isomerization of compound **1a** showed the 1st order kinetics with k=0.0064 min⁻¹ (please, see SI). The proposed mechanism and obtained kinetics were in full accordance with the known data on the intermolecular aqueous hydrolysis of the oxetane ring.¹⁴ In addition, ethyl ester **1**that cannot form analogous intermediate *Int-1* remained stable under these conditions (Scheme 5).

Proposed mechanism



Scheme 5. Proposed mechanism for the isomerization of oxetanecarboxylic acid 1a into lactone 1b.

Application in synthesis. The innate tendency of oxetanecarboxylic acids to isomerization can be also beneficially used in organic synthesis to prepare novel molecules (Schemes 3, 4), or to significantly simplify the synthesis of known ones. For example, dioxanone 28b was previously synthesized in four steps (18% total yield) from glycerine (39).¹⁵ Our approach allowed the three-step synthesis (29% total yield) of this product from the commercially available 3-oxetanol (39) (Scheme 6). Lactone 12b was previously synthesized in seven steps (10% total yield) from diester 40.16 In this work, we could obtain this molecule in just four steps from commercially available 3-oxetanone (41). Unsaturated lactone 42 was prepared before in three steps from triol 43.¹⁷ Here, we could also employ the isomerization strategy to obtain this compound in just two steps from 3-oxetanone (41). In that case, the final hydrolysis-isomerization was performed in one step in aqueous hydrochloric acid.



Scheme 6. Synthesis of compounds 28b, 12b, 42: literature approaches *vs* our approach.

Summary. During the past decade, oxetanes have been playing an important role in chemistry as bioactive compounds and valuable starting materials in synthesis. Oxetane-carboxylic acids have been used in more than 200 manuscripts and patents (Scheme 1). Here, we unexpectedly discovered that many of these molecules are intrinsically unstable. Some of them isomerized into lactones under simple storage at room temperature, others - under slight heating. For these acids, we recommend storage of the corresponding stable esters or Li- or Na-salts (for most of the unstable acids we could obtain their stable salts – please see SI).

Of course, the innate tendency of oxetane-carboxylic acids to isomerization can be beneficially used in the synthesis to make new molecules (Schemes 3, 4), and to simplify the preparation of the known ones (Scheme 6). However, the key message of this work is to inform *chemists on the innate instability of oxetane-carboxylic acids, as this could dramatically lower reaction yields and even lead to negative results (especially in those reactions that require heating).*

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors are grateful to Enamine Ltd for financial support. PM is grateful to Mrs. I. Sadkova for the help with the preparation of the manuscript, to Dr. E. Rusanov (IOC) for Xray analysis of compounds **10b**, **23b**; and Dr. S. Shishkina for X-ray analysis of compounds **18b** and **22b**. Mr. I. Pervak is acknowledged for the synthesis of the starting material **2a**. Authors are also grateful to Mr. I. Logvinenko, Mr. Y. Galuschak, Dr. D. Inshin, Mr. M. Leonenko for the support of this project. PM is very grateful to Mrs. Y. Fil for the experimental help in this project – the synthesis of compound **37b** (YF). T. Rudenko is grateful to Dr. O Stepaniuk for helpful suggestions.

Keywords: isomerization • medicinal chemistry • carboxylic acids • oxetanes • lactones

References

- (a) Burkhard, J. A.; Wuitschik, G.; Rogers-Evans, M.; Müller, K.; Carreira, E. M. Angew. Chem. Int. Ed. 2010, 49, 9052–9067. (b) Wuitschik, G.; Carreira, E. M.; Wagner, B.; Fischer, H.; Parrilla, I.; Schuler, F.; Rogers-Evans, M.; Müller, K. J. Med. Chem. 2010, 53, 3227– 3246. (c) Wuitschik, G.; Rogers-Evans, M.; Müller, K.; Fischer, H.; Wagner, B.; Schuler, F.; Polonchuk, L.; Carreira, E. M. Angew. Chem. Int. Ed. 2006, 45, 7736– 7739. (d) J. A. Burkhard, G. Wuitschik, J.-M. Plancher, M. Rogers-Evans, E. M. Carreira Org. Lett. 2013, 15, 4312-4315.
- For use of oxetanes in medchem campaigns, see for example: (a) Collier, P. N.; Twin, H. C.; Knegtel, R. M. A.; Boyall, D.; Brenchley, G.; Davis, C. J.; Keily, S.; Mak, C.; Miller, A.; Pierard, F.; Settimo, L.; Bolton, C. M.;

Chiu, P.; Curnock, A.; Doyle, E.; Tanner, A. J.; Jimenez, J. M. ACS Med. Chem. Lett. 2019, 10, 1134-1139. (b) Dowling, J. E.; Alimzhanov, M.; Bao, L.; Block, M. H.; Chuaqui, C.; Cooke, E. L.; Denz, C. R.; Hird, A.; Huang, S.; Larsen, N. A.; Peng, B.; Pontz, T. W.; Rivard-Costa, C.; Saeh, J. C.; Thakur, K.; Ye, Q.; Zhang, T.; Lyne, P. D. ACS Med. Chem. Lett. 2013, 4, 800-805. (c) Dubois, M. A. J.; Croft, R. A.; Ding, Y.; Choi, C.; Owen, D. R.; Bull, J. A.; Mousseau, J. J. RSC Med. Chem. 2021, 12, 2045-2052. (d) Kung, P. P.; Bingham, P.; Brooun, A.; Collins, M.; Deng, Y. L.; Dinh, D.; Fan, C.; Gajiwala, K. S.; Grantner, R.; Gukasyan, H. J.; Hu, W.; Huang, B.; Kania, R.; Kephart, S. E.; Krivacic, C.; Kumpf, R. A.; Khamphavong, P.; Kraus, M.; Liu, W.; Maegley, K. A.; Nguyen, L.; Ren, S.; Richter, D.; Rollins, R. A.; Sach, N.; Sharma, S.; Sherrill, J.; Spangler, J.; Stewart, A. E.; Sutton, S.; Uryu, S.; Verhelle, D.; Wang, H.; Wang, S.; Wythes, M.; Xin, S.; Yamazaki, S.; Zhu, H.; Zhu, J.; Zehnder, L.; Edwards, M. J. Med. Chem. 2018, 61, 650-665. (e) Lassalas, P.; Oukoloff, K.; Makani, V.; James, M.; Tran, V.; Yao, Y.; Huang, L.; Vijayendran, K.; Monti, L.; Trojanowski, J. Q.; Lee, V. M. Y.; Kozlowski, M. C.; Smith, A. B.; Brunden, K. R.; Ballatore, C. ACS Med. Chem. Lett. 2017, 8, 864-868. (f) Stepan, A. F.; Karki, K.; McDonald, W. S.; Dorff, P. H.; Dutra, J. K.; Dirico, K. J.; Won, A.; Subramanyam, C.; Efremov, I. V.; O'Donnell, C. J.; Nolan, C. E.; Becker, S. L.; Pustilnik, L. R.; Sneed, B.; Sun, H.; Lu, Y.; Robshaw, A. E.; Riddell, D.; O'Sullivan, T. J.; Sibley, E.; Capetta, S.; Atchison, K.; Hallgren, A. J.; Miller, E.; Wood, A.; Obach, R. S. J. Med. Chem. 2011, 54, 7772-Stepan, A. F.; Kauffman, G. W.; Keefer, C. 7783. (g) E.; Verhoest, P. R.; Edwards, M. J. Med. Chem. 2013, 56, 6985–6990. (h) Burkhard, J. A.; Wuitschik, G.; Plancher, J. M.; Rogers-Evans, M.; Carreira, E. M. Org. Lett. 2013, 15, 4312-4315. (i) Toselli, F.; Fredenwall, M.; Svensson, P.; Li, X. Q.; Johansson, A.; Weidolf, L.; Hayes, M. A. J. Med. Chem. 2019, 62, 7383-7399. (j) J. J. Rojas, R. A. Croft, A. J. Sterling, E. L. Briggs, D. Antermite, D. C. Schmitt, L. Blagojevic, P. Haycock, A. J. P. White, F. Duarte, C. Choi, J. J. Mousseau, J. A. Bull Nat. Chem. 2022, 14, 160-169.

For use of oxetanes in peptide studies, see: (a) Beadle, J. 3. D.; Knuhtsen, A.; Hoose, A.; Raubo, P.; Jamieson, A. G.; Shipman, M. Org. Lett. 2017, 19, 3303–3306. (b) McDougall, L.; Draper, E. R.; Beadle, J. D.; Shipman, M.; Raubo, P.; Jamieson, A. G.; Adams, D. J. Chem. Commun. **2018**, *54*, 1793–1796. (c) Möller, G. P.; Müller, S.; Wolfstädter, B. T.; Wolfrum, S.; Schepmann, D.; Wünsch, B.; Carreira, E. M. Org. Lett. 2017, 19, 2510-2513. (d) Powell, N. H.; Clarkson, G. J.; Notman, R.; Raubo, P.; Martin, N. G.; Shipman, M. Chem. Commun. 2014, 50, 8797-8800. (e) Roesner, S.; Beadle, J. D.; Tam, L. K. B.; Wilkening, I.; Clarkson, G. J.; Raubo, P.; Shipman, M. Org. Biomol. Chem. 2020, 18, 5400-5405. (f) Roesner, S.; Saunders, G. J.; Wilkening, I.; Jayawant, E.; Geden, J. V.; Kerby, P.; Dixon, A. M.; Notman, R.; Shipman, M. *Chem. Sci.* **2019**, *10*, 2465–2472.

- Bull, J. A.; Croft, R. A.; Davis, O. A.; Doran, R.; Morgan, K. F. *Chem. Rev.* 2016, *116*, 12150–12233.
- 5. For bioactive compounds with fragments of oxetanecarboxylic acids, see for example: (a) Del Carmen Terán Moldes, M.; Costantino, G.; Marinozzi, M.; Pellicciari, R. Farmaco 2001, 56, 609-613. (b) Ellis, B. D.; Milligan, J. C.; White, A. R.; Duong, V.; Altman, P. X.; Mohammed, L. Y.; Crump, M. P.; Crosby, J.; Luo, R.; Vanderwal, C. D.; Tsai, S. C. J. Am. Chem. Soc. 2018, 140, 4961-4964. (c) Furukawa, H.; Miyamoto, Y.; Hirata, Y.; Watanabe, K.; Hitomi, Y.; Yoshitomi, Y.; Aida, J.; Noguchi, N.; Takakura, N.; Takami, K.; Miwatashi, S.; Hirozane, Y.; Hamada, T.; Ito, R.; Ookawara, M.; Moritoh, Y.; Watanabe, M.; Maekawa, T. J. Med. Chem. 2020, 63, 10352-10379. (d) Kong, D.; Guo, S.; Yang, Y.; Guo, B.; Xie, X.; Hu, W. Bioorg. Med. Chem. Lett. 2019, 29, 848-852.
- 6. Use of oxetane-carboxylic acids as starting materials for further modifications: (a) Michiyuki, T.; Osaka, I.; Komeyama, K. Chem. Commun. 2020, 56, 1247-1250. (b) Adrian Meredith, J.; Wallberg, H.; Vrang, L.; Oscarson, S.; Parkes, K.; Hallberg, A.; Samuelsson, B. Eur. J. Med. Chem. 2010, 45, 160-170. (c) Collier, P. N.; Twin, H. C.; Knegtel, R. M. A.; Boyall, D.; Brenchley, G.; Davis, C. J.; Keily, S.; Mak, C.; Miller, A.; Pierard, F.; Settimo, L.; Bolton, C. M.; Chiu, P.; Curnock, A.; Doyle, E.; Tanner, A. J.; Jimenez, J. M. ACS Med. Chem. Lett. 2019, 10, 1134-1139. (d) Tran, T. D.; Wakenhut, F.; Pickford, C.; Shaw, S.; Westby, M.; Smith-Burchnell, C.; Watson, L.; Paradowski, M.; Milbank, J.; Brimage, R. A.; Halstead, R.; Glen, R.; Wilson, C. P.; Adam, F.; Hay, D.; Chiva, J. Y.; Nichols, C.; Blakemore, D. C.; Gardner, I.; Dayal, S.; Pike, A.; Webster, R.; Pryde, D. C. ChemMedChem 2014, 9, 1378-1386. (e) Tse, E. G.; Houston, S. D.; Williams, C. M.; Savage, G. P.; Rendina, L. M.; Hallyburton, I.; Anderson, M.; Sharma, R.; Walker, G. S.; Obach, R. S.; Todd, M. H. J. Med. Chem. 2020, 63, 11585-11601. (f) Zak, M.; Yuen, P. W.; Liu, X.; Patel, S.; Sampath, D.; Oeh, J.; Liederer, B. M.; Wang, W.; O'Brien, T.; Xiao, Y.; Skelton, N.; Hua, R.; Sodhi, J.; Wang, Y.; Zhang, L.; Zhao, G.; Zheng, X.; Ho, Y. C.; Bair, K. W.; Dragovich, P. S. J. Med. Chem. 2016, 59, 8345-8368.
- (a) Huang, H.; Yang, W.; Chen, Z.; Lai, Z.; Sun, J. Chem. Sci. 2019, 10, 9586–9590. (b) Huang, H.; Zhang, T.; Sun, J. Angew. Chem. Int. Ed. 2021, 60, 2668–2673. (c) R. N. Loy, E. N. Jacobsen J. Am. Chem. Soc. 2009, 131, 2786– 2787. (d) Bagal, S. K.; Bodnarchuk, M. S.; King, T. A.; McKerrecher, D.; Luo, X.; Wang, P.; Steward, O. R. Synlett 2020, 31, 502–506. (e) Brady, P. B.; Carreira, E. M. Org. Lett. 2015, 17, 3350–3353. (f) Deratt, L. G.; Lawson, E. C.; Kumar, K.; Hwang, S. S.; Desjarlais, R. L.; Kuduk, S. D. Org. Lett. 2020, 22, 5828–5832. (g) Deratt, L. G.; Lawson, E. C.; Wang, C. Y.; Kuduk, S. D. Org. Lett. 2019, 21, 9642–9645. (h) Zhang, R.; Guo, W.; Duan, M.; Houk,

K. N.; Sun, J. Angew. Chem. Int. Ed. 2019, 58, 18055-18060. (i) Zou, X.; Sun, G.; Huang, H.; Wang, J.; Yang, W.; Sun, J. Org. Lett. 2020, 22, 249-252. (j) Ruider, S. A.; Müller, S.; Carreira, E. M. Angew. Chem. Int. Ed. 2013, 52, 11908–11911. (k) Yang, W.; Sun, J. Angew. Chem. Int. Ed. 2016, 55, 1868-1871. (1) Wang, G.; Huang, H.; Guo, W.; Qian, C.; Sun, J. Angew. Chem. Int. Ed. 2020, 59, 11245-11249. (m) White, A. R.; Kozlowski, R. A.; Tsai, S. C.; Vanderwal, C. D. A. Angew. Chem. Int. Ed. 2017, 56, 10525-10529. (n) L. G. DeRatt, C.-Y. Wang, S. D. Kuduk J. Org. Chem. 2021, 86, 17482-17486. (o) J. J. Rojas, E. Torrisi, M. A. J. Dubois, R. Hossain, A. J. P. White, G. Zappia, J. J. Mousseau, C. Choi, J. A. Bull Org. Lett. 2022, 24, 2365–2370. (p) R. Zhang, M. Sun, Q. Yan, X. Lin, X. Li, X. Fang, H. H. Y. Sung, I. D. Williams, J. Sun, Org. Lett. 2022, 24, 2359-2364.

- Reviews on isomerizations of oxetanes: (a) Ahmad, S.; Yousaf, M.; Mansha, A.; Rasool, N.; Zahoor, A. F.; Hafeez, F.; Rizvi, S. M. A. *Synth. Commun.* **2016**, *46*, 1397–1416; (b) Malapit, C. A.; Howell, A. R. J. Org. Chem. **2015**, *80*, 8489–8495; (c) Sandvoß, A.; Wiest, J. M. Chem. Eur. J. **2021**, *27*, 5871–5879.
- (a) B. A. Chalyk, M. V. Butko, O. O. Yanshyna, K. S. Gavrilenko, T. V. Druzhenko, P. K. Mykhailiuk *Chem. Eur. J.* 2017, 23, 16782-16786. (b) B. Chalyk, A. Isakov, M. Butko, K. Hrebeniuk, O. Savych, O. Kucher, K. Gavrilenko, T. Druzhenko, V. Yarmolchuk, S. Zozulya, P. Mykhailiuk. *Eur. J. Org. Chem.* 2017, 2017, 4530-4542.
- 10. (a) Patent CN106565637A in Chinese language; (b) Patent WO2018195075A1.
- Compound 2a as a starting material in amide synthesis: (a) Ref. 6e; (b) Reddy, S.; Prasad, K. R. S. Asian J. Chem. 2021, 33, 577-582; (c) patent WO2015180612; (d) patent WO2018195075; (e) patent WO2018195075.
- 12. Patent CN106478558A in Chinese language.
- 13. CCDC numbers: 2090821 (10b); 1470391 (18b); 1470392 (22b); 2096431 (23b).
- J. G. Pritchard, F. A. Long J. Am. Chem. Soc. 1958, 80, 4162–4165.
- (a) Broggini, G.; Zecchi, G. Org. Prep. Proced. Int. 1991, 23 (6), 762–764. (b) Stacey, M.; Brimacombe, J. S.; Foster, A. B.; Stacey, M.; Whiffen, D. H. J. Chem. Soc. 1960, 520, 2574–2581.
- (a) Ishida, A.; Saijo, S.; Himizu, J.-I. *Chem. Pharm. Bull.* **1980**, 28, 783-788. (b) Ishida, A.; Noguchi, K.; Saijo, S.; Himizu, J.-I.; Wada, M. *Chem. Pharm. Bull.* **1979**, 27, 2281-2285.
- (a) Hoppe, D.; Schmincke, H.; Kleemann, H. W. *Tetrahedron* 1989, 45, 687–694. (b) Reimann, E.; Renz, M.; Unger, H. *Monats. Chem.* 2002, 133, 1285–1290.