¹ On-Board Reagent Storage and Release

² by Solvent-Selective, Rotationally Opened

³ Membranes – A Digital Twin Approach

4 Jens Ducrée

5 School of Physical Sciences, Dublin City University, Glasnevin, Dublin 9, Ireland, email:

6 jens.ducree@dcu.ie

7 Abstract

8 Decentralized bioanalytical testing in resource-poor settings ranges among the prime applications of 9 microfluidic systems. The high operational autonomy in such point-of-care / point-of-use scenarios 10 requires on-board stored liquid reagents, which need to be safely contained during long-term storage, 11 transport and handling, and reliably released prior to activation. Over the recent decades, centrifugal 12 microfluidic technologies have demonstrated the capability of integrated, automated and parallelized 13 sample preparation and detection of bioanalytical protocols. This paper introduces a novel concept 14 for onboard storage of liquid reagents which can be delivered in a well-defined manner by a rotational 15 stimulus of the system-innate spindle motor, while still aligning with the conceptual simplicity of such 16 "Lab-on-a-Disc" (LoaD) concepts. The reagent storage technology is captured by a digital twin which 17 allows making complex performance analysis and algorithmic design optimization according to given

18 objectives as expressed by target metrics.

19 Introduction

20 The automation of bioanalytical assay panels has been a paramount objective of microfluidic technologies since their debut in the early 1990s [1-5]. In the meantime, these Lab-on-a-Chip devices 21 22 have pervaded manifold application spaces, primarily in biomedical point-of-care and global 23 diagnostics, liquid handling automation for the life sciences, process analytical techniques and cell line 24 development for biopharma, as well as monitoring the environment, infrastructure, industrial 25 processes and agrifood [6-11]. Their compliance with the relevant workflows, infrastructure, operator 26 skill and competitive cost of ownership / per test are vital for deployment in locations outside 27 sophisticated medical infrastructure.

Various miniaturized liquid handling platforms have been introduced, which may be distinguished by their pumping scheme; among them are pressure sources, capillary force, electrokinetics, electrowetting on dielectric, bulk and surface-acoustic waves. Throughout the last three decades, centrifugal microfluidic platforms have been at the center of commercial and academic endeavors. Driven by a rugged spindle motor, these conceptually simple "Lab-on-a-Disc" (LoaD) systems [12-35] excel through their high-performance centrifugal sample preparation solely actuated by a conventional spindle motor

Mostly operating in batch-mode, biosamples are preconditioned through a series of centrifugally implemented Laboratory Unit Operations (LUOs), such as metering [36, 37], mixing [38-41], incubation, purification / concentration / extraction [42, 43], homogenization [44, 45], particle filtering [46-51], and droplet generation [52-54], while transiently held back in each step by a downstream valve. Note that some assay steps may also be realized by transferring functionalized magneticparticles between reagent chambers [55, 56].

41 Akin to the pick-up heads familiar from digital data storage technologies like CD, DVD or Blu-ray, most 42 detection schemes for LoaD systems are based on optical detection [22-24, 57-74]. While the radially 43 directed, outwards pointing centrifugal field is independent of the outer contours of the microfluidic 44 chip, a disc shape complies with the rotational symmetry and supports mechanical balance, layout, 45 and mold flow in common mass manufacturing such as (compression-)injection molding; yet, 46 deviations from the common 12-cm diameter and 1.2-mm thick CD format have been implemented, 47 e.g., smaller "mini-discs", tubes, or rectangular microscope slides. Furthermore, the alignment of the 48 inlet ports, outlets and detection chambers may also be important for seamless interfacing with 49 standard well plate-formats, liquid handling robotics, and associated equipment like readers. For 50 better readability, we refer to all these LoaD variants as "discs" in the following.

51 Liquid volumes concurrently residing on a rotor experience the same spin rate, and are thus 52 simultaneously exposed to the rotationally induced centrifugal field, which further depend on their 53 individual radial coordinates. Hence, and other than for most conventional Lab-on-a-Chip systems, 54 valving represents a key ingredient for automating sequential liquid handling protocols of the LoaD. 55 In principle, the disc could be halted for valve opening, for instance, by a manual or instrument-based, 56 e.g., mechanical, thermal or radiation-based actuator [75-83]. However, it is usually preferrable to 57 keep the disc-based liquid volumes at bay by at least moderate centrifugation, which requires a co-58 rotating power source, e.g., through pneumatic pumps [84, 85] or electro-thermal or radiative units 59 for melting sacrificial barriers film [83, 86, 87].

60 However, this work focusses on rotationally controlled valving concepts, which have been chosen by 61 many researchers due to their smooth alignment with the low complexity of the LoaD platform. In 62 these passive valves, the centrifugal pressure head driving liquid segments towards the disc perimeter 63 is combined with other pressure contributions which are independent of external power. In their high-64 pass variants, the centrifugal driving force is opposed by a capillary barrier, while low-pass siphons 65 typically feature hydrophilic coatings in inbound sections or pneumatic effects, so that valving is 66 ushered by reducing the spin rate below a critical threshold. Also, various centrifugo- or thermo-67 pneumatic flow control mechanisms [88-97] have been elaborated to create forward or reverse 68 pressure differentials.

To provide full-fledged sample-to-answer automation in compliance with point-of-care applications, the disc has to be pre-loaded with liquid or dry reagents, to avoid managing the logistics and loading from separate stocks; the user then only needs to introduce the sample into the disc. As opposed to many concepts conceived for temporarily retaining liquid volumes while carrying out LUOs along liquid handling sequences, reagent valves need to impede diffusion or exposure to ambient humidity over a shelf lives of months to years, and possibly rough handling during transport.
Various concepts based on physical barriers, like pouches, cartridges or wax plugs, have been

developed; similar to the previously discussed active valves, these containers may be opened by stimuli such as mechanical piercing, illumination by a high-power laser, or by local heating. Essential performance criteria are their compatibility with requirements of manufacture and assembly, evaporation, absorption into the bulk material, and possible leaching of chemicals during extended contact. Furthermore, the reliability of the release mechanism and the accuracy, precision and recovery ratio of the stored liquid volume need to be accounted for.

Dovetailing the centrifugal microfluidics covered in this work, also rotationally induced opening has 82 83 been demonstrated. For these concepts, it needs to be considered that, e.g., for limited motor power 84 and safety, there is an upper limit for practically achievable spin rates, and the reservoir might have 85 to be placed centrally, which would severely restrict obtainable pressure heads to fractions of 86 common atmospheric pressures. Thus, the reservoir valve has to yield at rather small pressure heads. 87 Furthermore, the mechanical strength of a designated weak point is hard to define, thus smearing out the associated spin rate threshold for release. Operational reliability hence demands setting a high 88 89 spin frequency for release, which tends to counteract the options for fluidic multiplexing of 90 concurrently loaded liquids [85].

This paper focuses on a novel type of rotationally actuated valve; during storage of an aqueous reagent, a (water) dissolvable film (DF) presents a diffusion barrier which is initially protected by an oleophilic, ancillary liquid having a certain, specific density [50, 75, 96, 98-100]. Upon spinning, a centrifugo-hydrostatic equilibrium is reached in which the interface between the two immiscible liquids contacts and thus dissolves the DF. In an idealized model, the reagent will be released at any finite spin rate.

97 In addition to the general prerequisites for on-board storage of liquid (and potentially also for 98 protecting dry / lyophilized) reagents, such valves need to meet further specifications. During logistics 99 and manual handling, acceleration due to shaking and terrestrial gravity act on the liquid volumes, 100 while the meniscus between the immiscible fluids needs to stay near its default rest position to avoid 101 premature opening. Furthermore, manufacturing and dispensing precision, evaporation rates, and 102 structural fidelity affect the release mechanism, and potentially enclosed or emerging gas bubbles 103 need to be tolerated.

This work first elaborates the operational principles for these rotationally controlled on-board storage and release valving. Then strategic features of the basic layout are introduced, and first motivated in a mostly qualitative manner. Next, key performance indicators (KPIs) are defined as design objectives, which are individually or collectively optimized along the "digital twin" derived from the underlying

108 functional model.

109 Working Principle

- 110 Figure 1 illustrates the fundamental mechanism underpinning storage and release of liquid reagents.
- 111 In the portrayed, exemplary fluidic structure, which is referred in the following to as Γ , two reservoirs
- of upper and lower cross sections A and a, and heights H and h on the left, and A' and a', and H' and
- 113 h' on the right, hold the aqueous reagent and the immiscible ancillary liquid of densities ρ and ρ' ,
- 114 respectively. These containers are interconnected by an isoradial channel at an inner radial position *R*
- of axial length \mathcal{L} , radial height \mathcal{H} , and of cross section \mathcal{A} , which accommodates a water-dissolvable
- film of axial extension δZ at the (mean) position Z on the *z*-axis.
- 117 For storage and transport ($\omega = 0$), the phase interface between the immiscible liquids is to reside
- within a (coaxial) segment of a cross section a of axial length δz , at a target position z = z, or at least
- 119 $|z z| \le 0.5 \cdot \delta z$. For activation and release in hydrostatic equilibrium at an angular spin rate at $\omega = 2\pi \cdot v > 0$, the meniscus needs to move beyond the DF at z = Z.





122 Figure 1 Basic storage and release mechanism (linearized, 2-dimensional display of cylindrical coordinates, dimensions 123 not to scale). The two-pronged structure, referred to as Γ , is divided into upper and lower radial sections of cross 124 sections A and A', and a and a', and heights of H, H', h and h', respectively. An isoradial channel at an inner radial 125 position *R* possesses an axial length \mathcal{L} , a radial extension \mathcal{H} and cross section \mathcal{A} ; a section of radial height \hbar and cross 126 section a and length $\delta z < \mathcal{L}$ is centered at z to provide a further degree of freedom. (Left) In a first step at rest ($\omega =$ 127 0), the aqueous reagent and an immiscible ancillary liquid of volumes U and U', densities ρ and ρ' , and viscosities η and 128 η' , are loaded to the left and right reservoirs with meniscus positions r_0 and r'_0 of their liquid distributions $\Lambda(\omega)$ and 129 $\Lambda'(\omega)$, respectively; their phase interface z is targeted to be located within the center of the z-segment centered at z =130 z in the isoradial channel, or at least $|\tilde{z} - z| \le 0.5 \cdot \delta z$. These compartments are then isolated at pneumatic pressures 131 $p_V(\omega = 0) = p'_V(\omega = 0) = p_0$ (9) from the ambient pressure to $p_0 \approx p_{std} = 1013.25$ hPa. (Right) Before its (default) 132 on-site usage, both seals are removed from the disc. During rotation at (theoretically any) $\omega > 0$, the initial difference 133 $\Delta p_{\omega}(\varrho, \varrho', U, U', R, \Gamma, \omega, z) = p_{\omega}(\varrho, U, R, \Gamma, \omega, z) - p_{\omega}(\varrho', U', R, \Gamma, \omega, z) > 0$ resulting from the pressure heads p_{ω} (4) drives the liquid distributions $\Lambda(\omega)$ and $\Lambda'(\omega)$ towards hydrostatic equilibrium $\Delta p_{\omega}(\varrho, \varrho', U, U', R, \Gamma, \omega, z) = 0$ (4), and 134 135 consequently new inner meniscus positions $r > r_0$ and $r' < r'_0$. For triggering the reagent release through an outlet, 136 e.g., located in a lower disc layer connected by a vertical via, the liquid-liquid interface at $z = z(\varrho, \varrho', U, U', R, \Gamma, \omega)$ in 137 hydrostatic equilibrium needs to cover the DF at z = Z having an axial extension δZ that was so far protected by the ancillary liquid, i.e., $z \ge Z$. Note that the geometry Γ is composed of cuboid segments to facilitate calculations. The 138 139 default values for the parameters indicated are compiled in Table A1. Evidently, by suitably adjusting the dissolution 140 characteristics of the DF, also non-aqueous liquids may be stored and released with the same mechanism.

141 The conservation of liquid volumes is generally expressed as

$$U(\check{r},\hat{r}) = \int_{\check{r}}^{\hat{r}} A(r) dr$$
(1)

for general geometries Γ by the integral over the function A(r) representing the dependence of the (partially compartmentalized) cross section A on the radial coordinate r between the inner and outer radial confinements of the liquid distributions Λ or Λ' ; this liquid volume U (1) spans between its inner and other radial confinements \check{r} and \hat{r} , respectively.

146 To simplify calculations without compromising the outcomes of this work, we consider a structure Γ 147 composed of cuboidal segments in Figure 1, with $\check{r} = r_0$ or r, and $\hat{r} = R + \hbar$. Instead of having to 148 (numerically) solve the possibly complex integral $U(\check{r}, \hat{r})$ in (1), the liquid volumes can then be 149 calculated (assuming $R - h - H \le r \le R - h$) by the algebraic formulas

$$U = A \cdot (R - h - r) + a \cdot h + \mathcal{A} \cdot z \Rightarrow r = r(U, R, \Gamma, z)$$
⁽²⁾

150 for the aqueous reagent and (assuming $R - h' - H' \le r' \le R - h'$)

$$U' = A' \cdot (R - h' - r') + a' \cdot h' + \mathcal{A} \cdot (L - z) \Rightarrow r'(U', R, \Gamma, z)$$
(3)

for the ancillary liquid. Equations (2) and (3) link the loaded liquid volumes U and U' to the radial position R of Γ , its structural parameters $A, A', a, a', h, h', \mathcal{A}, a, z, \delta z$ and \mathcal{L} as represented by Γ , and the interface at $0 < z < \mathcal{L}$; the liquid levels at rest can therefore be expressed as $r = r(U, R, \Gamma)$ in (2) and $r'(U', R, \Gamma)$ in (3).

155 Loading, Storage and Transport

Following a suitable, well-reproducible, possibly closed-loop controlled experimental loading 156 procedure at $\omega = 0$, the two liquids are introduced at ambient pressure p_0 (typically $p_0 \approx p_{std}$ with 157 the standard atmospheric pressure $p_{std} = 1013.25$ hPa) to place the phase interface at z within the 158 159 center of the isoradial segment at z. For a starting position z = z of the initial interface after properly loading U and U' at $\omega = 0$, the conservation of mass (1) trivially yields the initial filling levels $r_0 =$ 160 $r_0(U, \Gamma, z)$ and $r'_0 = r'_0(U', \Gamma, z)$. To suppress evaporation and contamination during subsequent 161 storage, transport and handling, the two reservoirs are then isolated from ambient by a membrane 162 163 exhibiting good barrier properties.

164 Liquid Release

165 In response to spinning at a finite spin speed $\omega > 0$, the centrifugal pressure heads

$$p_{\omega} = \varrho \cdot \bar{r} \Delta r \cdot \omega^2$$
 and $p'_{\omega} = \varrho' \cdot \bar{r}' \Delta r' \cdot \omega^2$ (4)

are induced to reshape the liquid distributions Λ and Λ' , which are confined by their menisci at the radially inner r, r' and common outer positions R, respectively. (Note that $\mathcal{H}/R \ll 1$ and $\hbar/R \ll 1$ are assumed throughout.) The products $\bar{r}\Delta r$ and $\bar{r}'\Delta r'$ in (4) are composed of the mean radial positions $\bar{r} = 0.5 \cdot (R + r)$ and $\bar{r}' = 0.5 \cdot (R + r')$, and the liquid level differences $\Delta r = R - r$ and $\Lambda r' = R - r'$.

- 171 In default actuation mode, the pneumatic seals are removed from the disc prior to launching the 172 centrifugal assay protocol $\omega(t)$. While rotating at sufficiently high ω , so that $p_{\omega} \propto \omega^2$ (4) overcomes 173 adverse stiction and capillary effects a hydrostatic equilibrium
- adverse stiction and capillary effects, a hydrostatic equilibrium

$$\Delta p_{\omega} = p_{\omega} - p_{\omega}^{\prime} \tag{5}$$

174 establishes which can be rewritten

$$\varrho \cdot (R+r) \cdot (R-r) = \varrho' \cdot (R+r') \cdot (R-r') \tag{6}$$

and therefore only relates the two liquid levels $r = r(\varrho, \varrho', U, U', R, \Gamma)$ and $r' = r'(\varrho, \varrho', U, U', R, \Gamma)$ to the radial position R of Γ , the loaded liquid volumes U and U', and their densities ϱ and ϱ' , but not to ω . The new, centrifugally stabilized position of the phase interface is defined by $\Delta p_{\omega}(\varrho, \varrho', U, U', R, \Gamma, z) = 0$ (5), and hence directly obtained from r and Γ , i.e., $z = z(\varrho, \varrho', U, U', R, \Gamma)$.

180 Consequently, centrifugally triggered release through the DF at \mathcal{Z} comes down to $z \ge \mathcal{Z}$, which 181 requires $\Delta p_{\omega}(z) > 0$ (5) during the transition of the meniscus at z all along the way from the position 182 during storage at z to the DF at $\mathcal{Z} > z$. This condition entails that a minimum reagent volume

$$U_{\Delta}(z) = \int_{z}^{z} A(\mathfrak{z}) d\mathfrak{z} = 0.5 \cdot a \cdot \delta z + \mathcal{A} \cdot (z - z - 0.5 \cdot \delta z)$$
(7)

183 (for $z + 0.5 \cdot \delta z \le z \le Z$) needs to be displaced for valve opening, and the shift of the left liquid level

- 184 from r_0 to $r > r_0$, while the meniscus of the ancillary liquid moves radially inbound from r'_0 to $r' < r'_0$.
- 185 If all meniscus position r_0 , r, r'_0 and r' remain in their respective inner compartments of cross sections

186 *A* and *A'* during this reconfiguration towards centrifugo-hydrostatic equilibrium from z = z to Z, we 187 obtain $r = r_0 + U_{\Delta}(Z)/A$ and $r' = r'_0 - U_{\Delta}(Z)/A'$.

188 Reliability

- 189 In practical applications, tolerances, mainly in the geometrical dimensions, as quantified by the
- 190 standard deviations referred to as $\Delta\Gamma$, and in the liquid volumes ΔU and $\Delta U'$ after pipetting, impact
- 191 the spread Δz of the interface z from their target positions z and Z at $\omega = 0$ and $\omega > 0$, respectively.
- Using (5), (2) and (3) for the meniscus position $z = z(\varrho, \varrho', U, U', R, \Gamma)$, its standard deviation

$$\Delta z \left(\left\{ \frac{\partial z}{\partial \gamma_k} \right\}, \left\{ \Delta \gamma_k \right\} \right) = \sqrt{\left(\sum_k \frac{\partial z(\varrho, \varrho', U, U', R, \Gamma)}{\partial \gamma_k} \cdot \Delta \gamma_k \right)^2}$$
(8)

 $\gamma_k \in \{\varrho, \varrho', U, U', R, \Gamma\},$ their deviations 193 depends on the values standard $\Delta \gamma_k \in$ $\{\Delta \varrho, \Delta \varrho', \Delta U, \Delta U', \Delta R, \Delta \Gamma\}$, and the partial derivatives $\partial z / \partial \gamma_k$ evaluated at the critical positions z and 194 Z, respectively. Note that strictly speaking, equation (8) only holds for small deviations $\{\Delta \gamma_k\}$. 195 196 Alternatively, as used for robustness analysis further below, Monte-Carlo methods can be employed 197 to compute Δz at the two target positions z = z and Z.

According to this fluidic model, operational robustness during storage, transport and rotation caused by statistical variations { $\Delta \gamma_k$ } is tightly linked to squeezing the interval of the actual interface positions $z \pm 0.5 \cdot M \cdot \Delta z$ (8) within the narrow segment after loading at rest, i.e., $M \cdot \Delta z(\omega = 0) < 0.5 \cdot \delta z$, and for reliable release at $\omega > 0$, $M \cdot \Delta z(\omega > 0) < 0.5 \cdot \delta z$. The factor M quantifies the targeted degree of operational robustness, with 68%, 95%, 99.7%, 99.99%, ... for M = 1,2,3,4,...

203 Design Characterization and Optimization

204 The digital twin [101-103] developed here allows to configure the free experimental parameters $\{\gamma_k\}$ 205 to achieve key performance goals, while staying commensurate with design-for-manufacture and scale-up of fabrication [104]. Underlying design optimization is facilitated by the multi-segmented 206 207 structure Γ (Figure 1) featuring cross sections A, a, A', a', A and a with respect to the axial direction 208 (and their respective axial heights / lengths $H, h, H', h', \delta z$ and \mathcal{L}). The core motivation of this draft 209 layout is now briefly outlined on a qualitative, heuristic manner; note that the due to the huge variety 210 of possible application cases in the multi-dimensional parameter space $\{\varrho, \varrho', U, U', R, \Gamma\}$, only computational optimization towards well-defined target metrics will eventually provide the proper 211 212 geometry.

The liquid volumes U and U' are chosen to settle the menisci r and r' in the inner (wider) region of 213 214 their reservoirs with cross sections A and A' so that effects of volume deviations, whether related to 215 systematic loss by evaporation or dispenser precision ΔU and $\Delta U'$, on the initial liquid levels r_0 , and 216 r'_0 , and thus the centrifugal equilibrium (5) determining z at $\omega > 0$, are mitigated. For given volumes U and U', the lower radial segments of the reservoirs display smaller cross sections a and a'; for 217 instance, a < A amplifies $\Delta r = r' - r$ entering the net pressure $p_{\omega} \propto \Delta r$ (4) for pumping the 218 minimum volume fraction U_{Δ} (7) of U to reach z = Z, as required for prompting disintegration of the 219 220 DF.

The additional segment centered at z in the isoradial channel featuring a profile a over an axial extension δz has been introduced for supporting the definition of the liquid-liquid interface, and to suppress the shift of the meniscus by a high flow resistance scaling with $\delta z/a^2$ during storage and transport. Evidently, it is critical that the actual position of the meniscus at \tilde{z} , when factoring in experimental tolerances in { $\varrho, \varrho', U, U', R, \Gamma$, }, remains between the edges, i.e., $|\tilde{z} - z| < 0.5 \cdot \delta z$

- 226 (while avoiding enclosure of gas between the liquid phases). A large extension δz is desirable for
- 227 improving the tolerance to discrepancies in the loading procedure of the two liquids.

228 Loading

The reservoirs are filled at the factory (at rest, i.e., $\omega = 0$) with the two immiscible liquids, in the main

- 230 case considered here water and FC-72 (3M[™] Fluorinert[™] Electronic Liquid FC-72), of target volumes
- 231 U and U' and densities ρ and ρ' through their designated inlet ports. The loading procedure should
- be highly reproducible, and ideally be monitored until the actual meniscus location \tilde{z} matches z, or at
- least settles sufficiently central in their designated channel section around z, i.e., $|\tilde{z} z| \ll 0.5 \cdot \delta z$.

234 Transport

- 235 When taken out of its storage, a LoaD may experience various accelerations β , e.g., repetitively during
- 236 manual handling, walking or in a moving vehicle, or punctually and more forcefully when its full weight
- hits solid ground after falling from a height. The directions of the resulting, usually unintended forces
- tend to be random, but may possess components parallel to the designated radial axis during spinning.
- The unknown number, magnitude, orientation and duration of such arbitrary inertial effects make it
- impossible to exactly quantify their impact on the deviation of the actual meniscus position \tilde{z} from its
- target value z. As successive accelerations might point in opposite directions, and thus neutralize their
- effect on the meniscus position, it is mostly likely that a single hard impact aligned in \mathscr{V} -direction will compromise the liquid distributions Λ and Λ' . We consider here two main mechanisms for pinning the
- 243 compromise the inquite distributions M and M. We consider here two matrix
- 244 phase interface during transport.

245 Pneumatic Stabilization

- Air-tight membranes seal the inlet ports of the reservoirs after loading the reagent and ancillary liquid
- 247 to prevent evaporation. These barriers also pneumatically stabilize Λ and Λ' by virtue of Boyle's law

$$p_V = p_0 \cdot \frac{V_0}{V} \tag{9}$$

stating that a change of an originally confined gas volume V_0 at p_0 to V alters the pressure to $p_V \neq p_0$ (9). So, for instance, according to the conservation of liquid volume dU/dt = 0 (1), transport-related disruption may induce a shift of the liquid level r on the (left) aqueous side towards the center of rotation, which ensues a peripheral displacement of the liquid level r' of the ancillary fluid (right) towards R (Figure 1).

The resultant changes in the gas volumes V and V' lead to an increase in p_V and a reduction of p'_V , thus seeking to restore $\tilde{z} \mapsto z$. The driving pressure (difference)

$$\Delta p_V(z) = p_V'(z) - p_V(z) = p_0 \cdot \left[\frac{V_0'}{V'(z)} - \frac{V_0}{V(z)} \right] = p_0 \cdot \left[\frac{V_0'}{V_0' - U_\Delta(z)} - \frac{V_0}{V_0 + U_\Delta(z)} \right]$$
(10)

should thus be maximized through adjusting the *z*-dependent displaced liquid volume $U_{\Delta}(z)$ in (7), and the initial gas volumes V_0 and V'_0 underneath the seal for stabilizing the interface position near z = z during transport. Figure 2(left) illustrates the dependency of the effective counter pressure Δp_V (10) in response to (left) a shift *z* from the default z = 10 mm. Figure 2(right) reveals that the restoring pressure $\Delta p_V < 0$ (10), evaluated at the downstream boundary of the *z*-segment z = z + $0.5 \cdot \delta z$, approaches 0 towards scaling the initially enclosed gas volumes V_0 and V'_0 by the (same) factor ξ .



Figure 2 Net pneumatic (counter-)pressure Δp_V (10) as a function of (left) the axial coordinate *z* with the equilibrium $\Delta p_V = 0$ at z = z, and (right) scaling the initial volumes V_0 and V'_0 by a by applying factor ξ to their heights while pinning the meniscus position at the end of the *z*-section $z = z + 0.5 \cdot \delta z$. Reducing the initial gas volumes V_0 and V'_0 thus stabilizes the actual meniscus position \tilde{z} in the vicinity of *z* during transport. Default values (Table A1) are used in this example.

267 Geometrical Factors

It would be operationally disastrous if the full volume $U_{\Delta}(\mathcal{Z})$ in (7) was displaced so that the aqueous reagent already reaches $z \ge \mathcal{Z}$ to open the DF during storage, transport and handling. This fatal event would happen when the liquid distributions Λ and Λ' have experienced an acceleration β with a strong component parallel to the radial r-direction of the centrifugal field (Figure 1) for a sufficiently long duty cycle τ . Note that even not discussed here for the sake of clarity, the interface may also leave the designated isoradial region, i.e., $z > z + 0.5 \cdot \delta z$, under the impact of β to disrupt the phase interface.

- 275 In response to an acceleration β with a major component parallel to the γ -axis, the liquid distributions
- 276 Λ and Λ' seek hydrostatic equilibrium (5). The resulting flow is driven by the pressure differential

$$p_{\beta} \approx \beta \cdot \left[\varrho \cdot \Delta r - \varrho' \cdot \Delta r' \right] \tag{11}$$

(initially) applying along the axial *z*-axis towards \mathcal{Z} , and throttled by the aggregate hydrodynamic resistance

$$\mathcal{R} = \sum_{q} \mathcal{R}_{q} = \sum_{q} c_{q} \cdot \frac{\eta_{q} \cdot l_{q}}{A_{q}^{2}}$$
(12)

of the structural segments indexed by q, possessing a cross section A_q , and filled over an axial length l_q with the reagent and ancillary liquid with viscosities η or η' , respectively. The numerical coefficients c_q amount to 8π for a round cross section. This law of Hagen-Poiseuille delivers a volume flow rate

$$\dot{U}_{V} = \frac{dU}{dt} = \frac{p_{\beta}}{\mathcal{R}} \approx \frac{\beta \cdot [\varrho \cdot \Delta r - \varrho' \cdot \Delta r']}{\sum_{q} c_{q} \cdot \eta_{q} \cdot l_{q} / A_{q}^{2}}$$
(13)

which is governed by p_{β} (11) and $\{\mathcal{R}_q\}$ (12). As the outlet opens for $\dot{U}_V \cdot \tau \ge U_{\Delta}(z)$ in (7), the duty cycle $\tau = U_{\Delta}/\dot{U}_V$ ought to be maximized to best suppress operationally disastrous premature release of reagent during transport. However, since β in unknown, we define a resilience (evaluated for z = 285 Z)

$$\tau_{\beta} = \frac{\tau}{\beta} = \frac{U_{\Delta}}{\beta \cdot \dot{U}_{V}} \approx \frac{\sum_{q} c_{q} \cdot \eta_{q} \cdot l_{q} / A_{q}^{2}}{[\varrho \cdot \Delta r - \varrho' \cdot \Delta r']} \cdot [0.5 \cdot a \cdot \delta z + \mathcal{A} \cdot (\mathcal{Z} - z - 0.5 \cdot \delta z)]$$
(14)

expressed in units of $s^3 \cdot m^{-1}$, to be maximized by adjusting to the structure Γ (Figure 1) for mitigating adverse effects owing to transport conditions.



Figure 3 Resilience quantified by τ_{β} (14) of the liquid distributions Λ and Λ' contained in the structure Γ as a function of (left) the length $l_q = \delta z$ of the isoradial segmented centered at z, and (right) its cross section $a_q = a$. Is inferred from its definition (14), the resilience steeply increases towards high flow resistance \mathcal{R} (12), i.e., with l_q/a_q^2 . The gridlines mark the values when using default parameters (Table A1).

292 This design goal of maximizing τ_{β} (14) translates into maximizing the dead volumes $U_{\Delta}(Z)$ (7) of the 293 isoradial channel extending between z = z on the left side, and Z on the right, mostly determined via 294 the part having the (larger) cross section \mathcal{A} . Furthermore, the flow rate \dot{U}_V (13) should be minimized; this implies that the (initial) liquid level difference r' - r should be minimized, e.g., by adjusting the 295 296 cross sections or the reservoirs A, a, A' and a', while the dominant flow resistance \mathcal{R} (12), as imposed by the isoradial segment of length $l_q = \delta z$ and cross section $A_q = a$, and scaling with the geometrical 297 ratio $\delta z/a^2$, should be high. As the reagent is usually given, an ancillary liquid possessing a high 298 299 viscosity η' would also be beneficial to increase τ_{β} (14). Figure 3 shows the dependency of the 300 resilience τ_{β} (14) on (left) the length δz , and (right) the cross section a of the z-segment, which 301 accounts for the highest impact on the flow resistance \mathcal{R} (12) through $\mathcal{R}_q \propto \delta z/a^2$.

302 Design Optimized for Transport

- By maximizing the quantities Δp_V (10) and τ_β (14) within the practical ranges of their input parameters
- 304 $\{\gamma_k\}$ and their tolerances $\{\Delta \gamma_k\}$, the parametrized structure Γ can be algorithmically optimized to
- achieve these design goals. Figure 4 shows (left) the layouts for highest restoring pressure Δp_V (10),
- 306 (center) the resilience τ_{β} (14), and (right) their product $\Delta p_V \cdot \tau_{\beta}$.



Figure 4 Algorithmically optimized layouts Γ according to given design metrics for stabilizing the liquid-liquid interface at z = z during transport. (Left) Pneumatic Δp_V (10): Both ports closed, (Center), Resilience τ_β (14): Both ports open, and (Right) Combination of stabilization of pneumatics and resilience $\Delta p_V \cdot \tau_\beta$ with both ports closed, which finds the right balance between partially contradictory design guidelines.

311 Actuation

312 Pneumatic Modes

- 313 With the default scenario portrayed in Figure 1, the inlets of the reservoirs for the aqueous reagent
- and the ancillary liquid are vented during rotation ($\omega > 0$), which can be interpreted as $V_0 \mapsto \infty$ and
- 315 $V'_0 \mapsto \infty$ in (10), and thus $\Delta p_V \mapsto 0$ (10). So as long as the centrifugally induced pressure head remains

positive, i.e., $\Delta p_{\omega} > 0$ (5) for z < Z, the meniscus will reach $z \ge Z$, and thus open DF at the outlet. However, especially for multiplexed flow control [103, 105], it is advantageous to create a critical spin rate

$$\Omega = \sqrt{\frac{p_V' - p_V}{\varrho \bar{r} \Delta r - \varrho' \bar{r}' \Delta r'}}$$
(15)

which first needs to be surpassed, i.e., $\omega > \Omega$, before trigging reagent release. In such centrifugopneumatic actuation, the ambient pressure p_0 applies to open, and p_V and p'_V (9) to sealed ports.



Figure 5 Burst frequency $\Omega/2\pi$ required to establish z = Z when both inlet ports remain sealed during rotation when reducing the ancillary volume U' by a factor of χ . Towards $\chi \mapsto 1$, Ω reaches values that are way out of reach for typical LoaD instruments, thus effectively preventing release. Yet, spin rates below the practical upper limit $\omega/2\pi \approx 100$ Hz can be achieved below $\chi \approx 0.55$. Default parameters (Table A1) are used, except that the cross sections A and a of the isoradial channel have been reduced by a factor of 3 to still assure its complete filling.

As the default experimental parameters ρ , ρ' , U, U', R and Γ (Table A1) are geared to result in Δp_{ω} = 327 328 0 (5) at z = Z, and thus $\Omega \mapsto \infty$, centrifugo-pneumatic actuation requires adjusting some of them. In general, the digital twin allows to determine the changes required to within the rather complex 329 330 correlation between $\varrho, \varrho', U, U', R, \Gamma$ and Ω in order realize certain design targets. The example 331 portrayed in Figure 5 shows the scaling of the critical spin rate $\Omega/2\pi$ (15) with the ancillary volume 332 $\chi \cdot U'$. Spindle speeds $\Omega/2\pi$ (15) within the experimentally achievable range not surpassing about 333 100 Hz only emerge below $\chi \approx 0.55$. Note that in order to make sure that the isoradial channel is 334 always filled with the ancillary liquid, the cross sections \mathcal{A} and α are both reduced by a factor of 3 335 with respect to the values in Table A1.

Note that equation (10) also discloses $\Delta p_V \propto p_0$. This scaling with the atmospheric pressure p_0 only affects the overall magnitude of Δp_V , but not the direction and ratio of the pneumatic pressures. We therefore refer to previous work on centrifugo-pneumatic valving where the impact and possible compensation of variation in the atmospheric pressure by weather, and particularly local altitude, have been examined in more detail [27, 98, 103, 105].

341 The critical spin rate Ω (15) also determines the maximum (density) of the centrifugal field $f_{\omega} = \rho_{\text{part}}$.

342 $R_{LUO} \cdot \Omega^2$ that can be sustained by the reagent valve, e.g., while an LUO such as plasma extraction is

- simultaneously processed to separate blood cells of (relative) density ρ_{part} at $\omega < \Omega \pm M \cdot \Delta \Omega$.
- 344 Systematic Volume Losses
- 345 Due to related to evaporation or absorption at rates \dot{U} and \dot{U}' , liquid volumes may appreciably decline
- during storage over time periods T, typically lasting months to a few years, by $\delta U = \dot{U} \cdot T$ and $\delta U' = \dot{U} \cdot T$
- 347 $\dot{U}' \cdot T$. Even for open inlet ports during rotation, the reduced volumes $U \delta U$ and $U' \delta U'$ may lead

to $\Delta p_{\omega}(\varrho, \varrho', U - \delta U, U' - \delta U', R, \Gamma, Z) < 0$ (5), and thus valve malfunction by failing to dissolve the DF at z = Z. (In addition, these losses may also affect the outcome of quantitative assays.)



Figure 6 Systematic reductions $\delta U = \dot{U} \cdot T$ and $\delta U' = \dot{U}' \cdot T$ of the originally loaded liquid volumes U and U' at rates \dot{U} and \dot{U}' change the centrifugal pressure balance p_{ω}/ω^2 (6) over a time T. In this example, annual loss rates of \dot{U} and \dot{U}' of 5% and 1% are assumed, and $\dot{U} > \dot{U}'$ is compensated by loading 10% more reagent volume U. The curve shows that the opening condition $p_{\omega}/\omega^2 > 0$ is assured beyond a typical minimum storage period of 24 months.

355 The main impact of the liquid losses δU and $\delta U'$ on the density-weighted radial products $\bar{r}\Delta r$ and 356 $\bar{r}'\Delta r'$ in the centrifugal equilibrium (6) is through $\Delta r = R - r$ and $\Delta r' = R - r'$ via $r(U, \delta U, A) =$ $(U - \dot{U} \cdot T)/A$ and $r'(U', \delta U', A') = (U' - \dot{U}' \cdot T)/A'$. Enlarged cross sections A and A' thus mitigate 357 358 the effect of evaporation or other losses of the reagent and ancillary phases at \dot{U} and \dot{U}' . Figure 6 359 shows that the prerequisite $\Delta p_{\omega} > 0$ for z = Z is provided for nearly 2.5 years at exemplary annual evaporation losses of 5% and 1% for the reagent and the ancillary liquid, respectively. This condition 360 of a positive centrifugal pressure differential Δp_{ω} (10) also implies that sufficient liquid volumes U and 361 U' are at available for all interface locations $z \le z \le Z$ to displace U_{Δ} (7) from the reagent side 362 through the isoradial segment into the ancillary reservoir. 363

364 Statistical Tolerances

350

Consistent reagent release hinges upon $|\tilde{z}(\varrho, \varrho', U, U', R, \Gamma, \Delta \varrho, \Delta \varrho', \Delta U, \Delta U', \Delta R, \Delta \Gamma) - \mathcal{Z}| \leq M \cdot$ 365 366 $\Delta z(z=Z)$ in hydrostatic equilibrium (5) at $\omega > 0$. The standard deviation Δz (8) is governed by the unavoidable spreads $\{\Delta \gamma_k\}$ of the experimental input parameters $\{\gamma_k\}$, mainly in the geometrical 367 368 dimensions $\Delta\Gamma$ and ΔU defining the structure Γ and the loaded liquid volumes U and U', respectively. The histogram in Figure 7 displays the distribution of \tilde{z} with a mean $\bar{z} = 19.97$ mm close to Z =369 370 20 mm and a standard deviation $\Delta z = 4.02$ mm (8) obtained from a Monte-Carlo simulation with 371 1000 runs using the default values and realistic tolerances Δd and Δw in vertical and lateral machining, and pipetting the liquid volumes ΔU and $\Delta U'$ as listed in Table A1. 372



Figure 7 Monte-Carlo simulation of the distribution of actual meniscus position \tilde{z} when targeting z = Z = 20 mm a centrifugal equilibrium $\Delta p_{\omega} = 0$ obtained with the default parameters $\varrho, \varrho', U, U', R$ and Γ while factoring in their respective tolerances $\Delta U, \Delta U'$ and $\Delta \Gamma$ (Table A1). After 1000 (time consuming) runs, the histogram features a mean position $\bar{z} = 19.99$ mm with a standard deviation $\Delta z = 0.387$ mm. The vertical lines indicate (magenta) the default 378 center and limits of the DF at $z = Z, Z \pm 0.5 \cdot \delta Z$, respectively, and (red shades) $z = Z \pm M \cdot \Delta z$ with $M = \{1,2,3\}$ with 379 the standard deviation Δz (8) of the \tilde{z} -distribution.

380 Gas Enclosure

- 381 During priming or storage, bubbles may emerge within the liquids. For developing a semi-quantitative
- 382 understanding of their influence on valving, we consider the case of a gas of volume $V_{g,0}$ entrapped at
- 383 ambient pressure p_0 in the center of the -isoradial z-segment after loading. Upon reaching equilibrium
- 384 during spinning at $\omega > 0$, the original volume $V_{g,0}$ is compressed to

$$V_{\rm g}(\omega) = V_{\rm g,0} \cdot \frac{p_0}{p_0 + p_\omega} = V_{\rm g,0} \cdot \frac{p_0}{p_0 + \varrho \cdot \bar{r} \Delta r \cdot \omega^2}$$
(16)

- 385 while now residing at z = Z. To still open the DF in presence of the entrapped gas, the liquid volumes U and U' need to be sized so that the meniscus at z would have to be shifted by a further $0.5 \cdot$ 386 387 $V_{g}(\omega)/\mathcal{A}$ (16) in the isoradial z-axis compared to the absence of a bubble. (Alternatively, the position 388
- Z can be appropriately adjusted to provide bubble tolerance.)

389 Assuming symmetrical displacement into each lateral reservoir (which is, strictly speaking, only the 390 case for $\rho = \rho'$), the liquid levels r and r' in hydrostatic equilibrium (5) are lifted by about $0.5 \cdot$ 391 $V_{\rm g}(\omega)/A$ and $0.5 \cdot V_{\rm g}(\omega)/A'$ towards the center of rotation, respectively. The impact of such a gas enclosure is assessed in a back-ot-the-envelope calculation; for this, we assume $V_0 = 1 \mu l$, $p_0 = p_{std}$, 392 393 $v = \omega/2\pi = 25$ Hz and default values for the other parameters (Table A1), to arrive at typical values of $V \approx 0.9 V_{g,0}$ and $\delta r \approx \delta r' \approx 1$ mm. 394

395 Refinements

- 396 In order to illustrate the concept and potential of a digital twin for optimizing the long-term storage 397 and release mechanism towards strategic design goals, we introduced a simple structure Γ (Figure 1); 398 this way, engineering objectives could be quantified and expressed by algebraic equations, which can 399 be solved on reasonable time scales by commonly available computing power.
- 400 Pegging the forward meniscus of the first introduced liquid by a small capillary barrier, sometimes also 401 referred to as phase guide, at z = z, is amongst many possible improvements. A possible trapping of 402 an interstitial bubble after filling the second liquid may be prevented by a gas permeable membrane, 403 located near z = z, or a local outlet to be sealed after priming has completed. Stiction, e.g., caused 404 by capillary pinning to manufacturing-related artefacts or dust, may be overcome by choosing a 405 sufficiently high spin rate $\omega \gg 0$ for reaching hydrostatic equilibrium (5) at z = Z.
- 406 To avoid possible interference of with the assay protocol, the ancillary liquid might be cleanly removed 407 under prevalent laminar flow conditions through an additional side pocket and centrifugal 408 stratification. The permanently gas filled parts of reservoirs may be placed at distal locations as long 409 as they are still pneumatically connected through conduits, e.g., to make efficient use of precious disc 410 real estate required for multiplexed assay panels. The pneumatic seals may also be removed through 411 secondary mechanism, e.g., akin to venting procedures implemented for centrifugo-pneumatic valves 412 based on mechanical [76], laser- [82, 106] or pneumatic [37, 66, 86, 88-91, 105-112] principles.

Summary and Outlook 413

Summary 414

- 415 A novel technology has been introduced for Lab-on-a-Disc systems which offers a physical evaporation
- 416 barrier and stabilization of liquid distributions during long-term storage, transport, and handling.
- 417 Reagent release proceeds through a dissolvable film disintegrating upon centrifugally induced contact
- 418 with the aqueous reagent. The complex interdependencies governing the operational principle over

its multiparameter space have been modelled to characterize system robustness and behavior *in silico*.
The resulting digital twin further enables computational design optimization towards given
performance objectives within practically achievable ranges regimes of experimental input
parameters. In addition, systematic volume losses, e.g., through evaporation during storage, artefacts
like enclosed gas bubbles, and statistical deviations in experimental input parameters can be factored
in.

425 Outlook

426 Evidently, the work presented represents the blueprint for setting up digital twins to efficiently 427 characterize and improve other functional elements of (centrifugal) microfluidic Lab-on-a-Disc 428 systems. Its simplified representation of the valving structure by cuboidal elements can be significantly 429 refined to optimize flow, e.g., by curved contours of the compartments, their inclination with respect 430 to the radial orientation, and fins to guide the relocation of liquids and gases. Similar to the entrapped 431 bubble, further parasitic effects observed during experimental testing, and additional elements of the 432 layout can be included in the digital twin. Such enhancements will require a more complex 433 computational fluid dynamic (CFD) simulation, which should also include inertia of the liquid and 434 elastic components such as the sealing membranes, which may bend or even yield under pressure.

435 It is well known that tests with real systems will show effects that are not included in the digital-twin 436 modelling. So, it is well expected that experimental validation will remain a substantial tool for arriving 437 at a product. However, in particular during early stage of development where manufacturing and 438 testing is mostly manual, only very limited numbers of fluidic chips are available, which usually prevent 439 collecting sufficient statistics for proper device performance and reliability analysis. The digital twin 440 presented in this work can then efficiently expedite design iteration of microfluidic systems by 441 providing virtual prototyping and testing. Such a tool hence empowers failure mode and effects 442 analysis (FMEA) regarding unavoidable tolerances, and in silico optimization of the layout for given 443 design targets. Adding similar programs for simulating manufacturing and biochemical processes 444 would also be desirable to combine with the digital twin for fluidics presented here.

On the bigger picture, the digital twin concept can boost microfluidic industries by standardization [113-115], interpreted in a way that validated boundary conditions issued by foundries can incorporated in computational design software to guarantee manufacturability and acceptable performance within given cost limits. Moreover, the digital twin modelling published here lends itself for open platform concepts in an increasingly digitized world, which can leverage crowdsourcing of brains, hands, infrastructure and equipment, e.g., coordinated by the rapidly emerging, decentralized blockchain technology [116-119].

452 Appendix

 $\begin{array}{ll} \textbf{453} & \text{Table A1 Default dimensions and boundary conditions for experimental parameters of the valving structure } \Gamma \mbox{ (Figure } \textbf{454} & 1). \end{array}$

Isoradial Channel	R = 30 mm	$\mathcal{L} = 30 \text{ mm}$	$\mathcal{H} = 3 \text{ mm}$
Isoradial <i>z</i> -Segment	z = 10 mm	$\delta z = 5 \text{ mm}$	h = 2 mm
DF Region	$\mathcal{Z} = 15 \text{ mm}$	$\delta Z = 5 \text{ mm}$	
Cross Sections	$A = 1 \text{ mm} \times 10 \text{ mm}$	$A' = 1 \text{ mm} \times 10 \text{ mm}$	$\mathcal{A} = 1 \text{ mm} \times 3 \text{ mm}$
(depth $ imes$ width)	$a = 1 \text{ mm} \times 5 \text{ mm}$	$a' = 1 \text{ mm} \times 4.5 \text{ mm}$	$a = 1 \text{ mm} \times 2 \text{ mm}$
Depths	D = d = 1 mm	D' = d' = 1 mm	$\mathcal{D} = d = 1 \text{ mm}$
Reservoir Heights	H = 15 mm	H' = 10 mm	
	h = 5 mm	h' = 2.5 mm	
Minimum Dimensions	Vertical $\geq 300 \ \mu m$	Lateral $\geq 200 \ \mu m$	Wall Thickness $\geq 1 \text{ mm}$

Structurable area:	$R_{\min} = 7.5 \text{ mm}$	$R_{\rm max} = 55 \ \rm mm$	
Geom. Tolerances	Vertical: 30 µm	Lateral: 20 µm	
Liquid Volumes	$U = 160 \mu l$	$U' \approx 60.68 \mu l$	$\Delta U = \Delta U' = 100 \text{ nl}$
Minimum Filling Gap		$\delta H = \delta H' = 1 \text{ mm}$	
Liquid Densities	$\varrho = 997 \text{ kg} \cdot \text{m}^{-3}$	$\varrho' = 1680 \text{ kg} \cdot \text{m}^{-3}$	
Liquid Viscosities	$\eta = 1.0016 \text{ mPa} \cdot \text{s}$	$\eta' = 0.64 \text{ mPa} \cdot \text{s}$	
Ambient pressure	$p_0 = p_{\rm std} = 1013.25 \rm hPa$		

455 The default parameters of the basic structure Γ in Figure 1 are listed in Table A1. The default tolerances 456 in vertical and lateral dimensions are $\Delta d = 30 \ \mu m$ and $\Delta w = 20 \ \mu m$, respectively. The minimum wall 457 thickness between fluidic cavities is set to 1 mm to account for scale-up of production by injection molding, and sufficient surface bonding. The properties of water and "FC-72" (3M™ Fluorinert™ 458 459 Electronic Liquid FC-72) representing an aqueous reagent and an immiscible ancillary liquid (at 25° C) are used. The volume of the ancillary fluid U' is chosen to settle z = Z at $\omega > 0$ (for both reservoirs 460 open). For the definition of liquid volumes, $\Delta U = \Delta U' = 100$ nl are assumed, and minimum gap δH 461 between the filling level and the seal is implemented for facilitating loading. Fluctuations in p_0 with 462 463 respect to the standard atmospheric pressure p_{std} are limited to the range following whether 464 conditions, i.e., about 4%; the impact of the local altitude on p_0 is more pronounced when operating the LoaD in mountainous regions. The impact of variance in $\varrho, \varrho', z, \delta z, Z, \delta Z, \mathcal{L}$ and R on the 465 466 meniscus position z is assumed to be neglectable.

467 References

- Manz, A., N. Graber, and H.M. Widmer *Miniaturized total chemical analysis systems: A novel concept for chemical sensing*. Sensors and Actuators B: Chemical, 1990. 1, 244-248 DOI:
 10.1016/0925-4005(90)80209-I.
- 471 2. Auroux, P.-A., D. Iossifidis, D.R. Reyes, and A. Manz *Micro Total Analysis Systems. 2. Analytical*472 *Standard Operations and Applications*. Analytical Chemistry, 2002. **74**, 2637-2652 DOI:
 473 10.1021/ac020239t.
- 474 3. Reyes, D.R., D. lossifidis, P.-A. Auroux, and A. Manz *Micro Total Analysis Systems*. 1.
 475 *Introduction, Theory, and Technology*. Analytical Chemistry, 2002. 74, 2623-2636 DOI: 10.1021/ac0202435.
- 477 4. Whitesides, G.M. *The origins and the future of microfluidics*. Nature, 2006. 442, 368-373 DOI:
 478 10.1038/nature05058.
- Janasek, D., J. Franzke, and A. Manz *Scaling and the design of miniaturized chemical-analysis systems*. Nature, 2006. **442**, 374-380 DOI: 10.1038/nature05059.
- 481 6. Gijs, M.A.M., F. Lacharme, and U. Lehmann *Microfluidic Applications of Magnetic Particles for*482 *Biological Analysis and Catalysis*. Chemical Reviews, 2010. **110**, 1518-1563 DOI:
 483 10.1021/cr9001929.
- 484 7. Nge, P.N., C.I. Rogers, and A.T. Woolley Advances in Microfluidic Materials, Functions,
 485 Integration, and Applications. Chemical Reviews, 2013. 113, 2550-2583 DOI:
 486 10.1021/cr300337x.
- 487 8. Liu, Q., C. Wu, H. Cai, N. Hu, J. Zhou, and P. Wang *Cell-Based Biosensors and Their Application*488 *in Biomedicine*. Chemical Reviews, 2014. **114**, 6423-6461 DOI: 10.1021/cr2003129.
- Mauk, M., J. Song, H.H. Bau, R. Gross, F.D. Bushman, R.G. Collman, and C. Liu *Miniaturized devices for point of care molecular detection of HIV*. Lab on a Chip, 2017. **17**, 382-394 DOI: 10.1039/c6lc01239f.
- 492 10. Yuan, X. and R.D. Oleschuk *Advances in Microchip Liquid Chromatography*. Analytical
 493 Chemistry, 2018. **90**, 283-301 DOI: 10.1021/acs.analchem.7b04329.
- 494 11. Olanrewaju, A., M. Beaugrand, M. Yafia, and D. Juncker *Capillary microfluidics in microchannels: from microfluidic networks to capillaric circuits*. Lab on a Chip, 2018. 18, 2323496 2347 DOI: 10.1039/c8lc00458g.

- 497 12. Schembri, C.T., V. Ostoich, P.J. Lingane, T.L. Burd, and S.N. Buhl *Portable Simultaneous*498 *Multiple Analyte Whole-Blood Analyzer for Point-of-Care Testing*. Clinical Chemistry, 1992. 38,
 499 1665-1670 DOI: 10.1093/clinchem/38.9.1665.
- Schembri, C.T., T.L. Burd, A.R. Kopfsill, L.R. Shea, and B. Braynin *Centrifugation and Capillarity Integrated into a Multiple Analyte Whole-Blood Analyzer*. Journal of Automatic Chemistry,
 1995. 17, 99-104 DOI: 10.1155/S1463924695000174.
- 503 14. *Abaxis (Piccolo Express)*. Accessed: 14/06/2021; Available on: <u>https://www.abaxis.com/</u>.
- 50415.Andersson, P., G. Jesson, G. Kylberg, G. Ekstrand, and G. Thorsen Parallel nanoliter microfluidic505analysis system. Analytical Chemistry, 2007. **79**, 4022-4030 DOI: 10.1021/ac061692y.
- Inganas, M., H. Derand, A. Eckersten, G. Ekstrand, A.K. Honerud, G. Jesson, G. Thorsen, T.
 Soderman, and P. Andersson Integrated microfluidic compact disc device with potential use in both centralized and point-of-care laboratory settings. Clinical Chemistry, 2005. 51, 1985-7
 DOI: 10.1373/clinchem.2005.053181.
- 51017.GyrosProteinTechnologies.Accessed:14/06/2021;Availableon:511https://www.gyrosproteintechnologies.com/.
- 18. Madou, M.J. and G.J. Kellogg *The LabCD (TM): A centrifuge-based microfluidic platform for diagnostics*. Systems and Technologies for Clinical Diagnostics and Drug Discovery,
 Proceedings Of, 1998. **3259**, 80-93 DOI: 10.1117/12.307314.
- 51519.Shea, M. ADMET Assays on Tecan's LabCD-ADMET System. Journal of the Association for516Laboratory Automation, 2003. 8, 74-77 DOI: 10.1016/s1535-5535(04)00260-6.
- Smith, S., D. Mager, A. Perebikovsky, E. Shamloo, D. Kinahan, R. Mishra, S.M.T. Delgado, H.
 Kido, S. Saha, J. Ducrée, M. Madou, K. Land, and J.G. Korvink *CD-Based Microfluidics for Primary Care in Extreme Point-of-Care Settings*. Micromachines, 2016. 7, DOI:
 10.3390/mi7020022.
- Kong, L.X., A. Perebikovsky, J. Moebius, L. Kulinsky, and M. Madou Lab-on-a-CD: A Fully
 Integrated Molecular Diagnostic System. Journal of the Association for Laboratory
 Automation, 2016. 21, 323-355 DOI: 10.1177/2211068215588456.
- Maguire, I., R. O'Kennedy, J. Ducrée, and F. Regan A review of centrifugal microfluidics in environmental monitoring. Analytical Methods, 2018. 10, 1497-1515 DOI: 10.1039/c8ay00361k.
- Sorkin, R., J. Park, J. Siegrist, M. Amasia, B.S. Lee, J.M. Park, J. Kim, H. Kim, M. Madou, and Y.K.
 Cho *Centrifugal microfluidics for biomedical applications*. Lab on a Chip, 2010. 10, 1758-1773
 DOI: 10.1039/b924109d.
- 530 24. Burger, R., L. Amato, and A. Boisen *Detection methods for centrifugal microfluidic platforms*.
 531 Biosensors and Bioelectronics, 2016. **76**, 54-67 DOI: 10.1016/j.bios.2015.06.075.
- Aeinehvand, M.M., F. Ibrahim, W. Al-Faqheri, K. Joseph, and M.J. Madou *Recent advances in the development of micropumps, microvalves and micromixers and the integration of carbon electrodes on centrifugal microfluidic platforms*. International Journal of Nanotechnology, 2018. 15, 53-68 DOI: 10.1504/IJNT.2018.089559.
- 53626.Sciuto, E.L., S. Petralia, G. Calabrese, and S. Conoci An integrated biosensor platform for537extraction and detection of nucleic acids. Biotechnol Bioeng, 2020. DOI: 10.1002/bit.27290.
- 53827.Ducrée, J. Design optimization of centrifugal microfluidic "Lab-on-a-Disc" systems towards539fluidic larger-scale integration. Appled Sciences, 2021. **11**, 5839 DOI: 10.3390/app11135839.
- Ramachandraiah, H., M. Amasia, J. Cole, P. Sheard, S. Pickhaver, C. Walker, V. Wirta, P. Lexow,
 R. Lione, and A. Russom, *Lab-on-DVD: standard DVD drives as a novel laser scanning microscope for image based point of care diagnostics.* Lab Chip, 2013. 13(8): p. 1578-85.
- 54329.Thompson, B.L., C. Birch, D.A. Nelson, J. Li, J.A. DuVall, D. Le Roux, A.C. Tsuei, D.L. Mills, B.E.544Root, and J.P. Landers A centrifugal microfluidic device with integrated gold leaf electrodes for545the electrophoretic separation of DNA. Lab on a Chip, 2016.54610.1039/c6lc00953k.

- 54730.Krauss, S.T., M.S. Woolf, K.C. Hadley, N.M. Collins, A.Q. Nauman, and J.P. Landers Centrifugal548microfluidic devices using low-volume reagent storage and inward fluid displacement for549presumptive drug detection. Sensors and Actuators B: Chemical, 2019. 284, 704-710 DOI:55010.1016/j.snb.2018.12.113.
- 55131.Abi-Samra, K., L. Clime, L. Kong, R. Gorkin, T.H. Kim, Y.K. Cho, and M. Madou Thermo-552pneumatic pumping in centrifugal microfluidic platforms. Microfluidics and Nanofluidics,5532011. **11**, 643-652 DOI: 10.1007/s10404-011-0830-5.
- Thompson, B.L., R.J. Gilbert, M. Mejia, N. Shukla, D.M. Haverstick, G.T. Garner, and J.P.
 Landers *Hematocrit analysis through the use of an inexpensive centrifugal polyester-toner device with finger-to-chip blood loading capability*. Analytica Chimica Acta, 2016. **924**, 1-8 DOI:
 10.1016/j.aca.2016.04.028.
- Watts, A.S., A.A. Urbas, E. Moschou, V.G. Gavalas, J.V. Zoval, M. Madou, and L.G. Bachas *Centrifugal microfluidics with integrated sensing microdome optodes for multiion detection*.
 Analytical Chemistry, 2007. **79**, 8046-8054 DOI: 10.1021/ac0709100.
- 561 34. Kim, T.H., K. Abi-Samra, V. Sunkara, D.K. Park, M. Amasia, N. Kim, J. Kim, H. Kim, M. Madou,
 562 and Y.K. Cho, *Flow-enhanced electrochemical immunosensors on centrifugal microfluidic*563 *platforms.* Lab on a Chip, 2013. **13**(18): p. 3747-3754.
- Moschou, E.A., A.D. Nicholson, G.Y. Jia, J.V. Zoval, M.J. Madou, L.G. Bachas, and S. Daunert, *Integration of microcolumns and microfluidic fractionators on multitasking centrifugal microfluidic platforms for the analysis of biomolecules.* Analytical and Bioanalytical Chemistry,
 2006. 385(3): p. 596-605.
- 56836.Mark, D., S. Haeberle, T. Metz, S. Lutz, J. Ducrée, R. Zengerle, and F. von Stetten Aliquoting569structure for centrifugal microfluidics based on a new pneumatic valve. MEMS 2008: 21st leee570International Conference on Micro Electro Mechanical Systems, Technical Digest, 2008. 611-571+.
- 572 37. Keller, M., S. Wadle, N. Paust, L. Dreesen, C. Nuese, O. Strohmeier, R. Zengerle, and F. von
 573 Stetten *Centrifugo-thermopneumatic fluid control for valving and aliquoting applied to*574 *multiplex real-time PCR on off-the-shelf centrifugal thermocycler*. RSC Advances, 2015. 5,
 575 89603-89611 DOI: 10.1039/c5ra16095b.
- 57638.Grumann, M., A. Geipel, L. Riegger, R. Zengerle, and J. Ducrée Batch-mode mixing on577centrifugal microfluidic platforms. Lab on a Chip, 2005. 5, 560-5 DOI: 10.1039/b418253g.
- 578 39. Ducrée, J., T. Brenner, S. Haeberle, T. Glatzel, and R. Zengerle *Multilamination of flows in planar networks of rotating microchannels*. Microfluidics and Nanofluidics, 2006. 2, 78-84 DOI: 10.1007/s10404-005-0056-5.
- 40. Burger, R., D. Kinahan, H. Cayron, N. Reis, J. Garcia da Fonseca, and J. Ducrée *Siphon-induced droplet break-off for enhanced mixing on a centrifugal platform*. Inventions, 2020. 5, DOI:
 10.3390/inventions5010001.
- 584 41. Ducrée, J., S. Haeberle, T. Brenner, T. Glatzel, and R. Zengerle *Patterning of flow and mixing in rotating radial microchannels*. Microfluidics and Nanofluidics, 2006. 2, 97-105 DOI: 10.1007/s10404-005-0049-4.
- 587 42. Strohmeier, O., S. Keil, B. Kanat, P. Patel, M. Niedrig, M. Weidmann, F. Hufert, J. Drexler, R.
 588 Zengerle, and F. von Stetten Automated nucleic acid extraction from whole blood, B. subtilis,
 589 E. coli, and Rift Valley fever virus on a centrifugal microfluidic LabDisk. RSC Advances, 2015. 5,
 590 32144-32150 DOI: 10.1039/c5ra03399c.
- 43. Brassard, D., M. Geissler, M. Descarreaux, D. Tremblay, J. Daoud, L. Clime, M. Mounier, D.
 592 Charlebois, and T. Veres *Extraction of nucleic acids from blood: unveiling the potential of active*593 *pneumatic pumping in centrifugal microfluidics for integration and automation of sample*594 *preparation processes.* Lab on a Chip, 2019. **19**, 1941-1952 DOI: 10.1039/c9lc00276f.
- 59544.Karle, M., J. Miwa, G. Roth, R. Zengerle, and F. von Stetten A Novel Microfluidic Platform for596Continuous DNA Extraction and Purification Using Laminar Flow Magnetophoresis. IEEE 22nd

- 597International Conference on Micro Electro Mechanical Systems (MEMS 2009), 2009. 276-279598DOI: 10.1109/Memsys.2009.4805372.
- Kido, H., M. Micic, D. Smith, J. Zoval, J. Norton, and M. Madou *A novel, compact disk-like centrifugal microfluidics system for cell lysis and sample homogenization*. Colloids and
 Surfaces B-Biointerfaces, 2007. 58, 44-51 DOI: 10.1016/j.colsurfb.2007.03.015.
- 46. Haeberle, S., T. Brenner, R. Zengerle, and J. Ducrée *Centrifugal extraction of plasma from whole blood on a rotating disk*. Lab on a Chip, 2006. 6, 776-781 DOI: 10.1039/b604145k.
- 47. Steigert, J., T. Brenner, M. Grumann, L. Riegger, S. Lutz, R. Zengerle, and J. Ducrée *Integrated*siphon-based metering and sedimentation of whole blood on a hydrophilic lab-on-a-disk.
 Biomedical Microdevices, 2007. 9, 675-679 DOI: 10.1007/s10544-007-9076-0.
- Kinahan, D.J., S.M. Kearney, N.A. Kilcawley, P.L. Early, M.T. Glynn, and J. Ducrée *Density-Gradient Mediated Band Extraction of Leukocytes from Whole Blood Using Centrifugo-Pneumatic Siphon Valving on Centrifugal Microfluidic Discs*. PLOS ONE, 2016. **11**, e0155545
 DOI: 10.1371/journal.pone.0155545.
- 49. Dimov, N., J. Gaughran, D. Mc Auley, D. Boyle, D.J. Kinahan, and J. Ducrée *Centrifugally Automated Solid-Phase Purification of RNA*. 2014 IEEE 27th International Conference on Micro
 Electro Mechanical Systems (MEMS), 2014. 260-263 DOI: 10.1109/MEMSYS.2014.6765625.
- 614 50. Gaughran, J., D. Boyle, J. Murphy, R. Kelly, and J. Ducrée *Phase-selective graphene oxide*615 *membranes for advanced microfluidic flow control*. Microsystems and Nanoengineering, 2016.
 616 2, 16008 DOI: 10.1038/micronano.2016.8.
- 51. Zehnle, S., M. Rombach, R. Zengerle, F. von Stetten, and N. Paust *Network simulation-based*optimization of centrifugopneumatic blood plasma separation. Biomicrofluidics, 2017. 11,
 DOI: 10.1063/1.4979044.
- Haeberle, S., R. Zengerle, and J. Ducrée *Centrifugal generation and manipulation of droplet emulsions*. Microfluidics and Nanofluidics, 2007. 3, 65-75 DOI: 10.1007/s10404-006-0106-7.
- 53. Schuler, F., F. Schwemmer, M. Trotter, S. Wadle, R. Zengerle, F. von Stetten, and N. Paust *Centrifugal step emulsification applied for absolute quantification of nucleic acids by digital droplet RPA*. Lab on a Chip, 2015. **15**, 2759-2766 DOI: 10.1039/c5lc00291e.
- 54. Schuler, F., M. Trotter, M. Geltman, F. Schwemmer, S. Wadle, E. Dominguez-Garrido, M.
 Lopez, C. Cervera-Acedo, P. Santibanez, F. von Stetten, R. Zengerle, and N. Paust *Digital droplet PCR on disk*. Lab on a Chip, 2016. 16, 208-216 DOI: 10.1039/c5lc01068c.
- 55. Czilwik, G., S.K. Vashist, V. Klein, A. Buderer, G. Roth, F. von Stetten, R. Zengerle, and D. Mark,
 Magnetic chemiluminescent immunoassay for human C-reactive protein on the centrifugal
 microfluidics platform. Rsc Advances, 2015. 5(76): p. 61906-61912.
- 631 56. Grumann, M., A. Geipel, L. Riegger, R. Zengerle, and J. Ducrée, *Magneto-hydrodynamic*632 *micromixing for centrifugal lab-on-a-disk platforms*, in *Micro Total Analysis Systems 2004, Vol*633 1. 2005. p. 593-595
- 57. Ducrée, J., S. Haeberle, S. Lutz, S. Pausch, F. von Stetten, and R. Zengerle *The centrifugal microfluidic Bio-Disk platform*. Journal of Micromechanics and Microengineering, 2007. **17**,
 5103-S115 DOI: 10.1088/0960-1317/17/7/S07.
- 58. Lutz, S., D. Mark, G. Roth, R. Zengerle, and F. von Stetten *Centrifugal Microfluidic Platforms for Molecular Diagnostics*. Clinical Chemistry and Laboratory Medicine, 2011. 49, S608-S608.
- 59. Tang, M., G. Wang, S.-K. Kong, and H.-P. Ho *A Review of Biomedical Centrifugal Microfluidic Platforms*. Micromachines, 2016. 7, DOI: 10.3390/mi7020026.
- 60. Duffy, D.C., H.L. Gillis, J. Lin, N.F. Sheppard, and G.J. Kellogg *Microfabricated Centrifugal Microfluidic Systems: Characterization and Multiple Enzymatic Assays*. Analytical Chemistry,
 1999. **71**, 4669-4678 DOI: 10.1021/ac990682c.
- 644 61. Azimi-Boulali, J., M. Madadelahi, M.J. Madou, and S.O. Martinez-Chapa *Droplet and Particle*645 *Generation on Centrifugal Microfluidic Platforms: A Review*. Micromachines, 2020. 11, DOI:
 646 10.3390/mi11060603.

- 647 62. Strohmeier, O., M. Keller, F. Schwemmer, S. Zehnle, D. Mark, F. von Stetten, R. Zengerle, and
 648 N. Paust *Centrifugal microfluidic platforms: Advanced unit operations and applications*.
 649 Chemical Society Reviews, 2015. 44, 6187-229 DOI: 10.1039/c4cs00371c.
- 63. Kong, L.X., A. Perebikovsky, J. Moebius, L. Kulinsky, and M. Madou *Lab-on-a-CD*. Journal of
 Laboratory Automation, 2016. 21, 323-355 DOI: 10.1177/2211068215588456.
- 64. Aeinehvand, M.M., P. Magaña, M.S. Aeinehvand, O. Aguilar, M.J. Madou, and S.O. Martinez653 Chapa Ultra-rapid and low-cost fabrication of centrifugal microfluidic platforms with active
 654 mechanical valves. RSC Advances, 2017. 7, 55400-55407 DOI: 10.1039/c7ra11532f.
- 655 65. Aeinehvand, M.M., L. Weber, M. Jiménez, A. Palermo, M. Bauer, F.F. Loeffler, F. Ibrahim, F.
 656 Breitling, J. Korvink, M. Madou, D. Mager, and S.O. Martínez-Chapa *Elastic reversible valves*657 *on centrifugal microfluidic platforms*. Lab on a Chip, 2019. **19**, 1090-1100 DOI:
 658 10.1039/C8LC00849C.
- 659 66. Hess, J.F., S. Zehnle, P. Juelg, T. Hutzenlaub, R. Zengerle, and N. Paust *Review on pneumatic*660 *operations in centrifugal microfluidics*. Lab on a Chip, 2019. **19**, 3745-3770 DOI:
 661 10.1039/C9LC00441F.
- 662 67. Nguyen, H.V., V.D. Nguyen, H.Q. Nguyen, T.H.T. Chau, E.Y. Lee, and T.S. Seo *Nucleic acid diagnostics on the total integrated lab-on-a-disc for point-of-care testing*. Biosensors and Bioelectronics, 2019. **141**, 111466 DOI: 10.1016/j.bios.2019.111466.
- 665 68. Rombach, M., S. Hin, M. Specht, B. Johannsen, J. Lüddecke, N. Paust, R. Zengerle, L. Roux, T.
 666 Sutcliffe, J.R. Peham, C. Herz, M. Panning, O. Donoso Mantke, and K. Mitsakakis *RespiDisk: A*667 *point-of-care platform for fully automated detection of respiratory tract infection pathogens*668 *in clinical samples*. The Analyst, 2020. **145**, 7040-7047 DOI: 10.1039/d0an01226b.
- 669 69. Homann, A.R., L. Niebling, S. Zehnle, M. Beutler, L. Delamotte, M.-C. Rothmund, D. Czurratis,
 670 K.-D. Beller, R. Zengerle, H. Hoffmann, and N. Paust A microfluidic cartridge for fast and
 671 accurate diagnosis of Mycobacterium tuberculosis infections on standard laboratory
 672 equipment. Lab on a Chip, 2021. DOI: 10.1039/d1lc00035g.
- 673 70. Madadelahi, M., L.F. Acosta-Soto, S. Hosseini, S.O. Martinez-Chapa, and M.J. Madou
 674 *Mathematical modeling and computational analysis of centrifugal microfluidic platforms: A*675 *review*. Lab on a Chip, 2020. 20, 1318-1357 DOI: 10.1039/c9lc00775j.
- 676 71. Miyazaki, C.M., E. Carthy, and D.J. Kinahan *Biosensing on the Centrifugal Microfluidic Lab-on-* 677 *a-Disc Platform*. Processes, 2020. **8**, 1360 DOI: 10.3390/pr8111360.
- Rombach, M., S. Hin, M. Specht, B. Johannsen, J. Lüddecke, N. Paust, R. Zengerle, L. Roux, T.
 Sutcliffe, J.R. Peham, C. Herz, M. Panning, O. Donoso Mantke, and K. Mitsakakis, *RespiDisk: a point-of-care platform for fully automated detection of respiratory tract infection pathogens in clinical samples.* The Analyst, 2020. **145**(21): p. 7040-7047.
- Brennan, D., H. Coughlan, E. Clancy, N. Dimov, T. Barry, D. Kinahan, J. Ducrée, T.J. Smith, and
 P. Galvin *Development of an on-disc isothermal in vitro amplification and detection of bacterial RNA*. Sensors and Actuators, B: Chemical, 2017. 239, 235-242 DOI: 10.1016/j.snb.2016.08.018.
- Delgado, S.M.T., D.J. Kinahan, F.S. Sandoval, L.A.N. Julius, N.A. Kilcawley, J. Ducrée, and D.
 Mager Fully automated chemiluminescence detection using an electrified-Lab-on-a-Disc
 (eLoaD) platform. Lab on a Chip, 2016. 16, 4002-4011 DOI: 10.1039/c6lc00973e.
- Mishra, R., J. Gaughran, D. Kinahan, and J. Ducrée *Functional Membranes for Enhanced Rotational Flow Control on Centrifugal Microfluidic Platforms*. Reference Module in Materials
 Science and Materials Engineering, 2017. DOI: 10.1016/b978-0-12-803581-8.04041-8.
- Kinahan, D.J., P.L. Early, A. Vembadi, E. MacNamara, N.A. Kilcawley, T. Glennon, D. Diamond,
 D. Brabazon, and J. Ducrée *Xurography actuated valving for centrifugal flow control*. Lab on a
 Chip, 2016. 16, 3454-3459 DOI: 10.1039/c6lc00568c.
- 69477.SpinXTechnologies.Accessed:15/06/2021;Availableon:695https://web.archive.org/web/20040414090409/http://www.spinx-technologies.com/.

- Abi-Samra, K., R. Hanson, M. Madou, and R.A. Gorkin *Infrared controlled waxes for liquid handling and storage on a CD-microfluidic platform*. Lab on a Chip, 2011. **11**, 723-726 DOI:
 10.1039/c0lc00160k.
- Kong, L.X., K. Parate, K. Abi-Samra, and M. Madou *Multifunctional wax valves for liquid handling and incubation on a microfluidic CD*. Microfluidics and Nanofluidics, 2015. 18, 1031-1037 DOI: 10.1007/s10404-014-1492-x.
- Al-Faqheri, W., F. Ibrahim, T.H.G. Thio, J. Moebius, K. Joseph, H. Arof, and M. Madou *Vacuum/Compression Valving (VCV) Using Parrafin-Wax on a Centrifugal Microfluidic CD Platform.* PLOS ONE, 2013. 8, DOI: 10.1371/journal.pone.0058523.
- 81. García-Cordero, J.L., F. Benito-Lopez, D. Diamond, J. Ducrée, and A.J. Ricco *Low-Cost*Microfluidic Single-Use Valves and on-Board Reagent Storage Using Laser-Printer Technology.
 IEEE 22nd International Conference on Micro Electro Mechanical Systems (MEMS 2009), 2009.
 439-442 DOI: 10.1109/Memsys.2009.4805413.
- 709 82. García-Cordero, J.L., D. Kurzbuch, F. Benito-Lopez, D. Diamond, L.P. Lee, and A.J. Ricco
 710 *Optically addressable single-use microfluidic valves by laser printer lithography*. Lab on a Chip,
 711 2010. 10, 2680-7 DOI: 10.1039/c004980h.
- 712 83. Torres Delgado, S.M., D.J. Kinahan, L.A. Nirupa Julius, A. Mallette, D.S. Ardila, R. Mishra, C.M.
 713 Miyazaki, J.G. Korvink, J. Ducrée, and D. Mager *Wirelessly powered and remotely controlled*714 *valve-array for highly multiplexed analytical assay automation on a centrifugal microfluidic*715 *platform*. Biosensors and Bioelectronics, 2018. **109**, 214-223 DOI: 10.1016/j.bios.2018.03.012.
- 716 84. Clime, L., D. Brassard, M. Geissler, and T. Veres Active pneumatic control of centrifugal 717 microfluidic flows for lab-on-a-chip applications. Lab on a Chip, 2015. 15, 2400-2411 DOI: 718 10.1039/c4lc01490a.
- Clime, L., J. Daoud, D. Brassard, L. Malic, M. Geissler, and T. Veres *Active pumping and control of flows in centrifugal microfluidics*. Microfluidics and Nanofluidics, 2019. 23, DOI: 10.1007/s10404-019-2198-x.
- Kinahan, D.J., S.M. Delgado, L.A.N. Julius, A. Mallette, D. Saenz-Ardila, R. Mishra, C.M.
 Miyazaki, J. Korvink, D. Mager, and J. Ducrée, *Wireless Closed-Loop Control of Centrifugo- Pneumatic Valving Towards Large-Scale Microfluidic Process Integration*, in 2018 IEEE Micro
 Electro Mechanical Systems (MEMS). 2018: Belfast, Northern Ireland. p. 1213-1216 DOI:
 10.1109/MEMSYS.2018.8346781.
- P27 87. Delgado, S.M.T., D.J. Kinahan, L.A.N. Julius, A. Mallette, D.S. Ardila, R. Mishra, C.M. Miyazaki,
 J.G. Korvink, J. Ducrée, and D. Mager *Wirelessly powered and remotely controlled valve-array for highly multiplexed analytical assay automation on a centrifugal microfluidic platform*.
 Biosensors & Bioelectronics, 2018. **109**, 214-223 DOI: 10.1016/j.bios.2018.03.012.
- 88. Godino, N., R. Gorkin, 3rd, A.V. Linares, R. Burger, and J. Ducrée *Comprehensive integration of homogeneous bioassays via centrifugo-pneumatic cascading*. Lab on a Chip, 2013. 13, 685-94
 DOI: 10.1039/c2lc40722a.
- Schwemmer, F., T. Hutzenlaub, D. Buselmeier, N. Paust, F. von Stetten, D. Mark, R. Zengerle,
 and D. Kosse *Centrifugo-pneumatic multi-liquid aliquoting-parallel aliquoting and combination of multiple liquids in centrifugal microfluidics*. Lab on a Chip, 2015. 15, 3250-3258
 DOI: 10.1039/c5lc00513b.
- 738 90. Zhao, Y., F. Schwemmer, S. Zehnle, F. von Stetten, R. Zengerle, and N. Paust *Centrifugo-*739 *pneumatic sedimentation, re-suspension and transport of microparticles*. Lab on a Chip, 2015.
 740 **15**, 4133-4137 DOI: 10.1039/c5lc00508f.
- 741 91. Zehnle, S., F. Schwemmer, R. Bergmann, F. von Stetten, R. Zengerle, and N. Paust *Pneumatic*742 *siphon valving and switching in centrifugal microfluidics controlled by rotational frequency or*743 *rotational acceleration*. Microfluidics and Nanofluidics, 2015. **19**, 1259-1269 DOI:
 744 10.1007/s10404-015-1634-9.

- Henderson, B.D., D.J. Kinahan, J. Rio, R. Mishra, D. King, S.M. Torres-Delgado, D. Mager, J.G.
 Korvink, and J. Ducrée *Siphon-Controlled Automation on a Lab-on-a-Disc Using Event- Triggered Dissolvable Film Valves*. Biosensors, 2021. 11, DOI: 10.3390/1108103.
- 93. Gorkin, R., 3rd, C.E. Nwankire, J. Gaughran, X. Zhang, G.G. Donohoe, M. Rook, R. O'Kennedy,
 and J. Ducrée *Centrifugo-pneumatic valving utilizing dissolvable films*. Lab on a Chip, 2012. 12,
 2894-902 DOI: 10.1039/c2lc20973j.
- Kinahan, D.J., S.M. Kearney, N. Dimov, M.T. Glynn, and J. Ducrée *Event-triggered logical flow control for comprehensive process integration of multi-step assays on centrifugal microfluidic platforms*. Lab on a Chip, 2014. 14, 2249-58 DOI: 10.1039/c4lc00380b.
- 754 95. Kinahan, D.J., S.M. Kearney, O.P. Faneuil, M.T. Glynn, N. Dimov, and J. Ducrée *Paper imbibition*755 *for timing of multi-step liquid handling protocols on event-triggered centrifugal microfluidic*756 *lab-on-a-disc platforms*. RSC Advances, 2015. 5, 1818-1826 DOI: 10.1039/c4ra14887h.
- Mishra, R., R. Alam, D.J. Kinahan, K. Anderson, and J. Ducrée, *Lipophilic-Membrane Based Routing for Centrifugal Automation of Heterogeneous Immunoassays*, in 2015 28th IEEE *International Conference on Micro Electro Mechanical Systems (MEMS 2015)*. 2015: Estoril,
 Portugal. p. 523-526 DOI: 10.1109/MEMSYS.2015.7051007.
- 97. Schwemmer, F., S. Zehnle, D. Mark, F. von Stetten, R. Zengerle, and N. Paust A microfluidic timer for timed valving and pumping in centrifugal microfluidics. Lab on a Chip, 2015. 15, 1545-1553 DOI: 10.1039/C4LC01269K.
- 98. Ducrée, J. Anti-counterfeit technologies for microfluidic "Lab-on-a-Disc" systems. 2021. DOI:
 10.20944/preprints202107.0443.v1.
- P9. Lu, Y., R. Mishra, D. McAuley, D. Boyle, and J. Ducrée, *Reliable liquid reagent storage and* rotational release for centrifugal sample-to-answer automation, in Proceedings of the 24th
 International Conference on Miniaturized Systems for Chemistry and Life Sciences (μTAS 2020),
 October 04–08, S.L.G.a.H. Lu, Editor. 2020, The Chemical and Biological Microsystems Society
 (CBMS): Virtual. p. 134-135
- Mishra, R., D. McAuley, N. Rolinska, D. Boyle, and J. Ducrée, *Barrier-film based reagent storage*and release on microfluidic platforms for sample-to-answer automation of bioassays, in
 Proceedings of the 24th International Conference on Miniaturized Systems for Chemistry and
 Life Sciences (μTAS 2020), S.L.G.a.H. Lu, Editor. 2020, The Chemical and Biological
 Microsystems Society (CBMS): Virtual. p. 382-383
- 776101.DigitalTwin.2021;Accessed:25/05/2021;Availableon:777https://en.wikipedia.org/wiki/Digital_twin.
- 778102.Marr, B. What Is Digital Twin Technology And Why Is It So Important? 2017 Published:77906/03/2017; Accessed: 25/05/2021; Available on:780https://www.forbes.com/sites/bernardmarr/2017/03/06/what-is-digital-twin-technology-781and-why-is-it-so-important/.
- 103. Ducrée, J. Systematic review of centrifugal valving based on digital twin modelling towards
 highly integrated Lab-on-a-Disc systems. Nature Microsystems & Nanoengineering, 2021.
 DOI: 10.20944/preprints202105.0683.v2.
- 785104.Ducrée, J. Efficient development of integrated Lab-On-A-Chip systems featuring operational786robustness and nanufacturability. Micromachines, 2019. 10, 12 DOI: 10.3390/mi10120886.
- 787105.Ducrée, J. Secure air traffic control at the hub of multiplexing on the centrifugo-pneumatic Lab-788on-a-Disc platform. Micromachines, 2021. 12, 700 DOI: 10.3390/mi12060700.
- 106. Mishra, R., G. Reilly, M. Agnew, A. Garvey, C. Rogers, E. Andrade, H. Ma, S. Fitzgerald, J.
 Zapatero, R. O'Kennedy, and J. Ducrée, *Laser-Actuated Centrifugo-Pneumatic Flow Control Towards 'Sample-to-Answer' Integrated Detection of Multi-Marker Panels at the Point-of- Care*, in *2018 IEEE Micro Electro Mechanical Systems (MEMS)*. 2018: Belfast, Northern Ireland.
 p. 1185-1188 DOI: 10.1109/MEMSYS.2018.8346774.
- 794107.Godino, N., R. Gorkin, A.V. Linares, R. Burger, and J. Ducrée, A Centrifugo-Pneumatic Cascade795for Fully Integrated and Multiplexed Biological Analysis, in 2012 IEEE 25th International

- 796 *Conference on Micro Electro Mechanical Systems (MEMS).* 2012: Paris, France DOI: 10.1109/MEMSYS.2012.6170352.
- Gorkin, R., L. Clime, M. Madou, and H. Kido, *Pneumatic pumping in centrifugal microfluidic platforms*. Microfluidics and Nanofluidics, 2010. 9(2-3): p. 541-549.
- 800 109. Gorkin, R., C. Nwankire, J. Siegrist, R. Burger, J. Gaughran, and J. Ducree, *Rotationally* 801 *controlled centrifugo-pneumatic valving utilizing dissolvable films*, in 2011 16th International
 802 *Solid-State Sensors, Actuators and Microsystems Conference*. 2011. p. 1276-1279 DOI:
 803 10.1109/transducers.2011.5969448.
- Kinahan, D.J., M. Renou, D. Kurzbuch, N.A. Kilcawley, E. Bailey, M.T. Glynn, C. McDonagh, and
 J. Ducrée *Baking Powder Actuated Centrifugo-Pneumatic Valving for Automation of Multi-Step Bioassays*. Micromachines, 2016. 7, DOI: 10.3390/mi7100175.
- Mark, D., S. Haeberle, T. Metz, S. Lutz, J. Ducrée, R. Zengerle, and F. von Stetten Aliquoting
 structure for centrifugal microfluidics based on a new pneumatic valve. MEMS 2008: 21st IEEE
 International Conference on Micro Electro Mechanical Systems, Technical Digest, 2008. 611 614 DOI: 10.1109/MEMSYS.2008.4443730.
- 811 112. Mark, D., T. Metz, S. Haeberle, S. Lutz, J. Ducrée, R. Zengerle, and F. von Stetten *Centrifugo-*812 *pneumatic valve for metering of highly wetting liquids on centrifugal microfluidic platforms*.
 813 Lab on a Chip, 2009. **9**, 3599-3603 DOI: 10.1039/b914415c.
- van Heeren, H. Standards for connecting microfluidic devices? Lab on a Chip, 2012. 12, 10221025 DOI: 10.1039/C2LC20937C.
- 816 114. Stavis, S.M. *A glowing future for lab on a chip testing standards*. Lab on a Chip, 2012. 12, 3008 817 11 DOI: 10.1039/c2lc40511c.
- 818 115. Reyes, D.R., H.v. Heeren, S. Guha, L.H. Herbertson, A.P. Tzannis, J. Ducrée, H. Bissig, and H.
 819 Becker Accelerating Innovation and Commercialization Through Standardization of 820 Microfluidic-Based Medical Devices. Lab on a Chip, 2021. 21, 9-21 DOI: 10.1039/D0LC00963F.
- 116. Ducrée, J., M. Gravitt, R. Walshe, S. Bartling, M. Etzrodt, and T. Harrington *Open Platform Concept for Blockchain-Enabled Crowdsourcing of Technology Development and Supply Chains.* Frontiers in Blockchain, 2020. 3, 386525 DOI: 10.3389/fbloc.2020.586525.
- 117. Ducrée, J., M. Etzrodt, B. Gordijn, M. Gravitt, S. Bartling, R. Walshe, and T. Harrington
 Blockchain for Organising Effective Grass-Roots Actions on a Global Commons: Saving The
 Planet. Frontiers in Blockchain, 2020. 3, 33 DOI: 10.3389/fbloc.2020.00033.
- 118. Ducrée, J., M. Etzrodt, S. Bartling, R. Walshe, T. Harrington, N. Wittek, S. Posth, K.W.A. Ionita,
 W. Prinz, D. Kogias, T. Paixão, I. Peterfi, and J. Lawton Unchaining collective intelligence for
 science, research and technology development by blockchain-boosted community
 participation. Frontiers in Blockchain, 2021. DOI: 10.3389/fbloc.2021.631648.
- 119. Ducrée, J. *Research A blockchain of knowledge*? Blockchain Research and Applications,
 2020. 1, 100005 DOI: 10.1016/j.bcra.2020.100005.
- 833