

Tuning Coordination Modes and Nucleation of Calcium Phosphate with Oligomeric Lactic Acid and pH Values: Theoretical and Experimental Study

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ABSTRACT: Mineralization of calcium phosphate (CaP) is ubiquitous in nature, which can facilitate biological organisms produce hierarchically structured minerals. The influences of the addition of oligomeric lactic acid (LAC_n, n=1, 8) and changing pH values on nucleation processes of CaP under the simulated body fluid (SBF) solution or aqueous solution were revealed by using ab initio molecular dynamics (AIMD) and molecular dynamics (MD) simulations. Through increasing pH value, the coordination between differently protonated phosphate species and Ca²⁺ ions could be tuned from the monodentate (η^1) to the coexisting monodentate and bidentate (η^2) modes. The carboxyl and hydroxyl groups of LAC molecules are capable of forming multiple interactions such as proton transfer, electrostatic interaction, and intermolecular hydrogen bonding with phosphate species and Ca²⁺ ions on both pre-nucleation clusters and hydroxyapatite (HA) surfaces. The high-throughput experimentations (HTE) with factors of the adding LAC, changing Ca/P ratios (1.25 ~ 2.50), using different solutions demonstrated that the UV-Vis absorbance values decreased with addition of LAC, indicating the inhibition of the nucleation process of CaP. At the physiological pH of 7.4, the CaP samples with different Ca/P ratios (1.50, 1.67 and 2.00) exhibited negative zeta potential values, which were correlated with the surface electrostatic potential distributions and potential biological applications. The precipitation was CaHPO₄ · 2H₂O (DCPD) in neutral condition at the early stage of nucleation process. Understanding the effects of different pH and Ca/P values on the nucleation process and interfacial interaction between LAC additive and the nanocluster is helpful to guide the rational design of biocompatible materials.

1. Introduction

Mineralization, which is an important biological process, facilitates biological organisms produce hierarchically structured minerals. Bones and teeth of vertebrates are known to come from CaP minerals,¹ whose nucleation process was proposed to have two possibilities, i.e., a classical aggregation or a two-step nucleation pathway.² In the classical viewpoint, the ions of solution could directly aggregate, nucleation, and finally crystallization. Another two-step nucleation pathway proposal indicated that the ions could self-assemble to form metastable amorphous clusters, and then aggregated together to fulfill final precipitation in solution. In fact, the real nucleation process of CaP in biological condition should be much more complicated because many factors such as adding additive, Ca/P ratio, carbonate ions and pH value could affect the occurrence of different phosphate species such as H_3PO_4 (called 3H for short), H_2PO_4^- (2H), HPO_4^{2-} (1H), and PO_4^{3-} (0H), shown in Figure 1a. It is known that in the CaP solution, starting from the ion pair CaHPO_4 , pre-nucleation cluster $\text{Ca}(\text{HPO}_4)_3^{4-}$ could be gradually formed at the early stages nucleation process.³⁻⁶ Such a CaP cluster $[\text{Ca}(\text{HPO}_4)_3]^{4-}$ was demonstrated to exist both η^1 and η^1 modes, i.e., $\text{Ca}(\eta^2\text{-HPO}_4^{2-})_2(\eta^1\text{-HPO}_4^{2-})(\text{H}_2\text{O})_2$ by using AIMD simulations with PBE functional.⁴ Subsequently, those small-sized clusters could aggregate into thermodynamically metastable amorphous CaP (ACP).⁷ The building blocks of ACP are further propagated with several nucleation mechanisms, such as post-nucleation cluster $\text{Ca}_2(\text{HPO}_4)_3^{2-}$ and Posner's cluster $\text{Ca}_9(\text{PO}_4)_6$.^{3, 7-9} With the increase in cluster size, the ACP phase has been formed gradually as the precursor, and then transformed into octocalcium phosphate (OCP, $\text{Ca}_8(\text{HPO}_4)_2(\text{PO}_4)_4 \cdot 5\text{H}_2\text{O}$) and crystalline phase.^{3, 7} CaP minerals in aqueous solution mainly include dicalcium phosphate dehydrate (DCPD, $\text{CaHPO}_4 \cdot 2\text{H}_2\text{O}$), monocalcium phosphate monohydrate (MCPM, $\text{Ca}(\text{H}_2\text{PO}_4)_2 \cdot \text{H}_2\text{O}$), OCP and hydroxyapatite (HA, $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$).¹⁰ Among them, HA is considered the most thermodynamically stable phase of CaP minerals. It has received much attention as a potential anti-tumor material due to the ability of inhibiting the proliferation of various tumor cells.¹¹⁻¹⁶

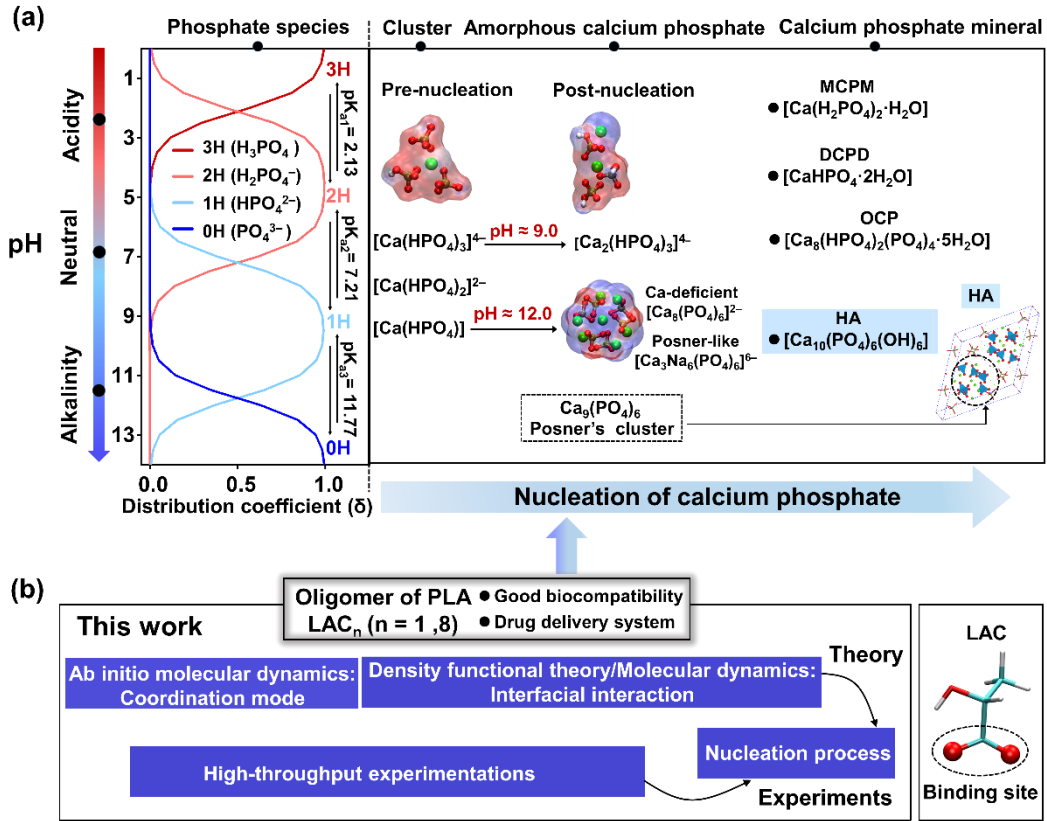


Figure 1. (a) The distribution coefficient δ of phosphate species, H_3PO_4 (3H), H_2PO_4^- (2H), HPO_4^{2-} (1H), and PO_4^{3-} (0H), as a function of pH values, and the schematic illustration of nucleation process of CaP, (b) The influence of LAC_n ($n=1, 8$) additives and pH values on the nucleation is investigated in this work through theoretical (AIMD, DFT, and MD) and experimental (UV-Vis, XRD, TEM, and Zeta potential) study.

The sequential nucleation and crystal growth processes in biomineralization have aroused intensive interest in both experimental and theoretical studies. Scanning electron microscopy (SEM) and XRD experiments indicated that the nucleation and growth of HA were promoted on some substrates, such as chitosan-coated polylactic acid (PLA) nanofiber, polylactide/gelatin composite nanofibers, lignin/polycaprolactone nanofibers and polydopamine,¹⁷⁻²⁵ due to chelation of the coated hydroxyl or carboxylate groups with Ca^{2+} ions in the simulated body fluid (SBF, whose ion concentrations and pH value are similar to the human blood plasma). It has also been found that the pH condition has great influence on the intrinsic

physicochemical characteristics of various CaP samples, including adsorption of organic acids²⁶, phase composition²⁷, morphology and crystal size²⁸. Moreover, zeta potentials were affected by pH values for many nanoparticles, such as iron oxide nanoparticles,²⁹ Ag@HA,³⁰ F-Cl@HA,³¹ and the complexes of HA with the additives of citric acid³², oxalic acid³³ and malic acid³⁴. Such negative zeta potentials of nanoparticles were proposed to favor osseointegration, apatite nucleation, and inhibition of tumor cell proliferation.^{14, 29, 31, 35, 36}

Some experimental phenomena have been rationalized by molecular dynamics simulations. For example, the addition of substrate such as cellulose nanofibrils³⁷ is less favorable for the formation of CaP clusters than those in solutions based on MD simulations with CSFF force field. The surface of protein is dominated by amino acid residues, which could provide adequate reactive sites to bind with Ca^{2+} or phosphate ions through electrostatic interaction or hydrogen bonding and facilitate the nucleation and growth of CaP. The regulation of nucleation process of CaP by the charged amino acids of collagen surface was demonstrated by CHARMM27 based MD study in solution.³⁸⁻⁴⁰ Neutral amino acids of collagen such as glycine and hydroxyproline residues, whose carboxylate groups were also found to be freely accessible to the Ca^{2+} ions based on generalized AMBER force field.⁴¹ The interactions between lysine and HA (100) surface was ascribed to the dominant electrostatic interaction in Ca-O ionic bonding, followed by hydrogen bonding between $\text{C}_\epsilon\text{-H}$ /amine-H and phosphate-O.⁴² Electrostatic interactions were also found to play an important role in binding of CaP clusters with proteins such as bone morphological protein-2⁴³ and clathrin-associated protein adaptin-2 (AP2)⁴⁴ especially with the charged residues. The important role of hydrogen bonding interaction in the nucleation of CaP was revealed by oxygen plasma treatment and MD simulations with GAFF force field, from which H_2PO_4^- ions were shown to be initially adsorbed at the -OH-modified parylene C surface accompanied by Ca^{2+} and HPO_4^{2-} ions.⁴⁵

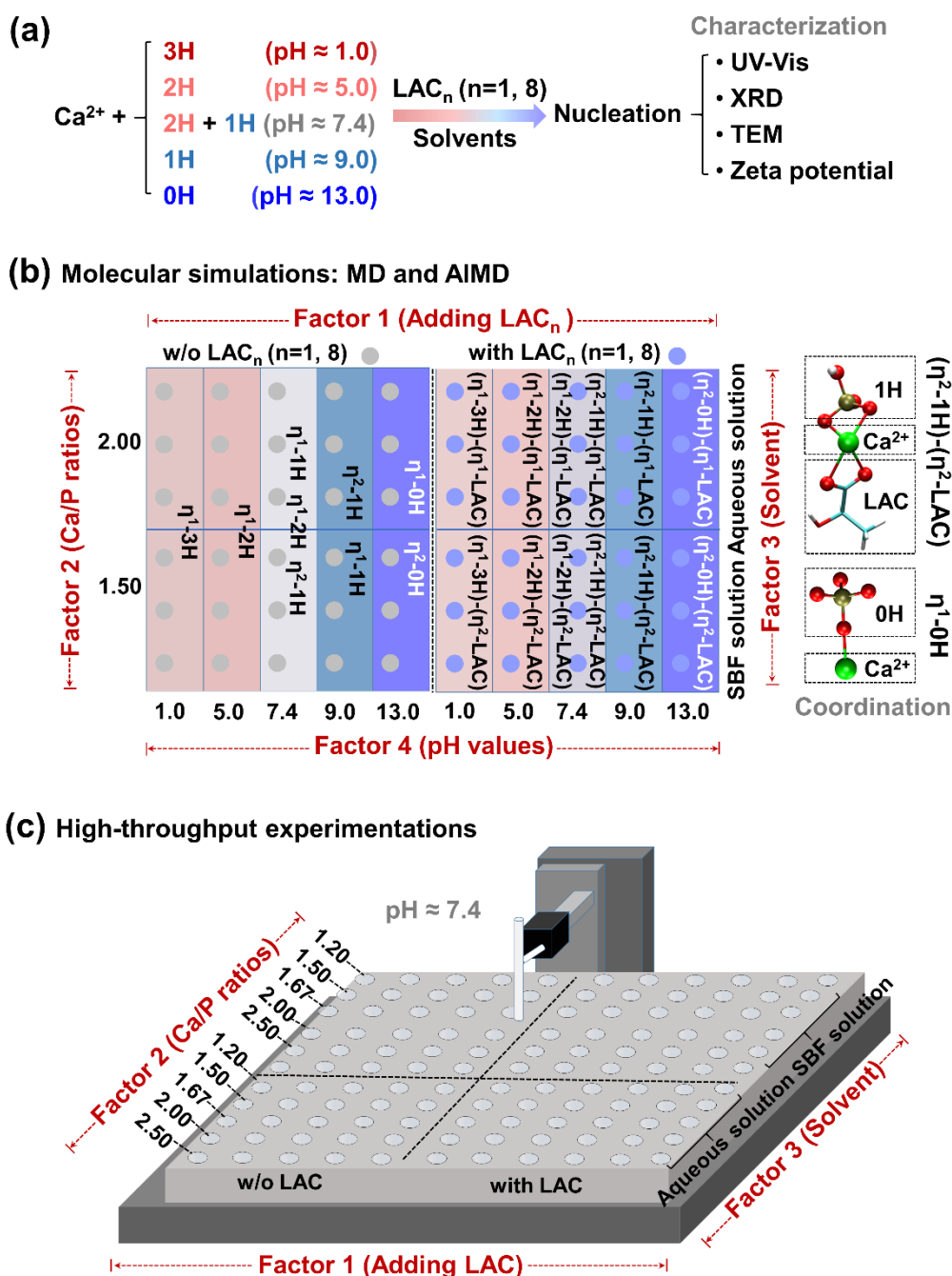


Figure 2. (a) The model build and experimental detection methods in studied systems, (b) the main factors affecting the nucleation of CaP, (c) schematic diagram of high-throughput experimentations equipment.

In the present work, we are interested in the interaction of the oxygen-containing molecules, such as LAC, with CaP systems, since LAC is the basic block of PLA, which is one of the widely used biodegradable polyesters in medical applications.⁴⁶ Lactic acid

is also a by-product of aerobic or anaerobic glycolysis, and its high amount is a characteristic feature of tumor cells.⁴⁷ It could function as a key signaling molecule in detecting cancer cell migration, angiogenesis and metastasis.⁴⁸ Taking into account the value of lactic acid dissociation constant of $pK_a = 3.86$,⁴⁹ the deprotonated lactic acid form (also called LAC for short) is the dominated species in the studied pH conditions except for the simulation with $pH = 1.0$ (Figure 1). The nucleation process of CaP based on the four factors (adding LAC_n ($n = 1, 8$), Ca/P ratio, pH value, and SBF/aqueous solution) (Figure 1b) was systematically investigated by using AIMD and MD simulations as well as high-throughput experimentations (HTE). Density functional theory (DFT) was employed to depict the coordination modes and their binding strength between Ca^{2+} ions and phosphate species with and without the addition of LAC oligomers. AIMD and MD simulations demonstrated the aggregation processes of CaP clusters in both SBF and aqueous solutions. HTE⁵⁰ allowed a large number of experiments about changing the Ca/P ratios, additives, and solvents to be executed in parallel (Figure 2). An in-depth atomistic insight into the nucleation process of CaP under different values of pH and Ca/P ratios with the addition of LAC is important to comprehend the influence of various factors on the binding modes and to guide the rational design of potential biomaterials.

2. Materials and Methods

2.1. Binding strength estimated by DFT calculations.

To study the influence of pH values and the addition of LAC on interfacial interactions of CaP with the surrounding species, the binding strength of Ca^{2+} (M) with the ligand (Y), including phosphate species, carbonate species (H_2CO_3 , HCO_3^- , CO_3^{2-}) in the simulated body fluid, LAC oligomers, and solvent molecules (water or CH_3COOH), was calculated by using density functional theory (DFT) with the implicit polarizable continuum model (PCM) at the level of DFT/M062X/6-311+g(d,p) within

the Gaussian 16 package⁵¹. The binding free energy, G_{bind} , of a CaP cluster in a certain pH condition was derived from the following equation,

$$G_{\text{bind}} = G_{\text{MY}} + 6 \times G_{\text{wt}} - G_{\text{Y}} - G_{\text{M(wt)}_6} \quad (1)$$

where $\text{M} = \text{Ca}^{2+}$, whose 6-coordinated complexation with water (wt), M(wt)_6^{2+} , is taken as a reference in binding Gibbs free energy (G) calculations. The optimized geometries of the studied systems are presented in Figure S1 and the calculated Gibbs free energies at 298 K are listed in Table S1. A negative value of G_{bind} of a complex indicates that this compound is thermodynamically more stable than the Ca(wt)_6^{2+} complex, causing the competitive interaction with Ca^{2+} ions in the presence of various additives.

With an increase in the cluster size, crystallization may take place after some treatments. The two-layer slab model with periodic boundary condition (PBC) was applied to simulate the binding of LAC on the HA surface using the DFT calculations with the plane wave-based Vienna ab initio simulation package (VASP)⁵². The details of slab models were given in supporting information. DFT calculations were performed within generalized gradient approximation (GGA) with Perdew-Burke-Ernzerhof (PBE)⁵³ functional. The van de Walls (vdW) correction with the Grimme approach (DFT-D3)⁵⁴ was performed to describe the interaction between the additive species and surfaces. The relative stabilities of two different surfaces, i.e., (001) versus (100) of HA were estimated by their surface energy, E_{surf} , with the computational details shown in supporting information. To estimate the interaction energy between LAC and HA slab with different Ca/P ratios (Figure S2 and Figure S3), the binding energies (E_{bind}) were calculated from the energy difference between the whole system and the individual components, as shown in Table S2. To correlate the binding strength with the extent of interfacial charge transfer, the atomic charges were evaluated by Bader charge analysis⁵⁵.

2.2. MD simulations of aggregation and nucleation of CaP. To study the nucleation process of CaP in SBF solution w/o or with oligomeric LAC_n ($n=1, 8$) additives, we carried out the MD simulations by using AMBER16 CUDA version⁵⁶. The octamer

LAC₈ was taken as an example to model the interfacial interaction of polylactic acid (PLA) polymer with the CaP clusters. The pK_a of LAC and oligomeric PLA are 3.86 and 3.10,^{49, 57} respectively. When the pH is greater than 5, they are both in deprotonated states (Figure S4). In addition, there are several kinds of anions in SBF solution, such as carbonate ions, whose protonation state distribution at different pH values depends on their pK_a values⁵⁸⁻⁶⁰, as shown in Figure S5. The GAFF2 force field was used for the phosphate species, with the force field parameters shown in Table S3. The 500 ns MD simulations were run at NPT ensemble (with the constant pressure of 1.0 bar and temperature of 300 K) for the model systems with different pH values and Ca/P ratios, as shown in Tables S4-S9. The temperature was controlled using Langevin dynamics⁶¹ with the collision frequency of 1 ps⁻¹. Integration step at 1 fs was set and the direct spatial nonbonded cutoff of 8.0 Å was used. The local binding modes were displayed from the radial distribution function (RDF) and spatial distribution function (SDF) using VMD⁸ and TRAVIS⁹ programs, respectively. Adaptive Poisson-Boltzmann Solver (APBS)⁶² was employed to study the electrostatic properties of the CaP clusters.

2.3. AIMD simulations for exploring the local coordination modes. In order to provide the more detailed information of the intermolecular interaction in the early aggregation stage, the AIMD simulations were carried out using CP2K with PBE functional and GTH-DZVP basis set^{63, 64} for all atoms. The Grimme algorithm (DFT-D3)⁵⁴ was employed to improve the description of van der Waals (vdW) interactions. The energy cutoff for plane wave basis set was 550 Ry. Several CaP systems with different protonation conditions (Table S10) with or without the addition of LAC were set by using the Packmol code⁶⁵ in a PBC cell of 14 × 14 × 14 Å³ to illustrate the coordination modes between different phosphate species and Ca²⁺ ions under different pH values. The counterions, Na⁺ or Cl⁻ ions, were added to neutralize the excess charge. The 20 ps AIMD trajectory was generated in an NPT ensemble at 300 K and 1 bar with a time step of 0.5 fs.

2.5. High-throughput experimentation (HTE) for tuning various factors. The high-throughput experimentations were carried out using the house-made synthesis

equipment (Figure 2c and Figure S6). Based on combinatoric chemistry method, we developed a high-through microarray synthesis equipment which is composed of a multi-channel sample collection platform, a multi-channel automatic liquid injection system and an automatic stirring system. The addition of reaction solution, the stirring speed and the stepping of XY-axis motion guide can be controlled by the computer with our self-compiled operating software. Therefore, the input ratio of different reactants, the pH value of solution, reaction temperature and stirring time can be auto-executed by preset system. Through this equipment, different samples can be automatically synthesized one by one.

Calcium chloride ($\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$) and diammonium hydrogen phosphate ($(\text{NH}_4)_2\text{HPO}_4$) were purchased from Sinopharm Chemical Reagent Co., Ltd. SBF solution was directly acquired from Shanghai Yuanye Bio-technology Co., Ltd. The species of lactic acid (LAC) used as factor 1 in the CaP nucleation experiments was provided by Shanghai Aladdin Bio-Chem Technology Co., Ltd. To obtain enough precipitate for XRD and TEM characterizations, LAC (purity $\geq 95\%$), $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$, $(\text{NH}_4)_2\text{HPO}_4$, and SBF solution were purchased from Shanghai Bide Chemical Reagent Co., Ltd., Shanghai Hushi Chemical Reagent Co., Ltd, and Phygene Life Sciences Co., Ltd., respectively. All chemicals were used without further purification.

For HTE, the concentration of $(\text{NH}_4)_2\text{HPO}_4$ was about 0.006700 and 0.003335 mol dm^{-3} , and remained unchanged. The concentration of CaCl_2 was determined by modifying the Ca/P ratios of 1.25, 150, 1.67, 2.00, 2.50 and 2.75 (factor 2), respectively. The solvents used in this HTE included aqueous solution and SBF solution (factor 3). Firstly, the prepared $(\text{NH}_4)_2\text{HPO}_4$, CaCl_2 and LAC solutions (factor 1) were adjusted to pH about 7.4 with $\text{NH}_3 \cdot \text{H}_2\text{O}$ or HCl . The above neutral solutions were mixed together in the order of LAC, $(\text{NH}_4)_2\text{HPO}_4$ and CaCl_2 (Figure S6, Tables S11-S18). Then, the reactions were automatically carried out through the high-throughput material synthesis equipment and stirred for three minutes for each experiment group. For each Ca/P ratio group, the experiments were repeated three times. Finally, the amounts of precipitation formed in the 120 samples were derived from UV-Vis absorbance at 450 nm, using a

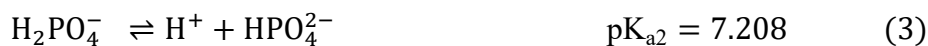
Shimadzu UV-2600 spectrophotometer instrument using SBF or the distilled water as the reference sample.

Zeta potential analysis was performed at room temperature on Zetasizer (Nano-Z, Malvern, UK) based on dynamic light scattering theory. Powder X-ray diffraction (XRD) analysis for the crystalline structures of the as-prepared products were conducted on a Shimadzu XRD-6000 instrument with Cu K α radiation ($\lambda=0.154$ nm) generated at 40 kV and 30 mA. Transmission electron microscopy (TEM) images were obtained on JEOL JEM-1011 instrument with an accelerating voltage of 120 kV to analyze the morphological and structural features of the as-made samples.

3. Results and Discussion

3.1. Protonation states of phosphate species under different pH values

Under different pH conditions the phosphate has different protonation states, called 3H, 2H, 1H, 0H, respectively, whose distribution coefficients could be derived from the following acid-base equations:



The pK_{a1} , pK_{a2} and pK_{a3} in equations (2)-(4) are the acid dissociation constants of phosphate⁵⁸. According to those pK_a values, one could predict the distribution coefficient, δ , of each protonation state at a certain pH value by using the following equations.

$$\delta(3\text{H}) = \frac{[\text{H}^+]^3}{[\text{H}^+]^3 + K_{a1}[\text{H}^+]^2 + K_{a1}K_{a2}[\text{H}^+] + K_{a1}K_{a2}K_{a3}} \quad (5)$$

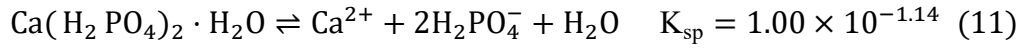
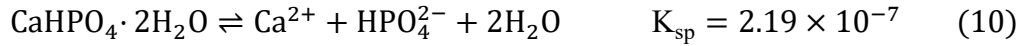
$$\delta(2\text{H}) = \frac{K_{a1}[\text{H}^+]^2}{[\text{H}^+]^3 + K_{a1}[\text{H}^+]^2 + K_{a1}K_{a2}[\text{H}^+] + K_{a1}K_{a2}K_{a3}} \quad (6)$$

$$\delta(1\text{H}) = \frac{K_{a1}K_{a2}[\text{H}^+]}{[\text{H}^+]^3 + K_{a1}[\text{H}^+]^2 + K_{a1}K_{a2}[\text{H}^+] + K_{a1}K_{a2}K_{a3}} \quad (7)$$

$$\delta(\text{OH}) = \frac{K_{a1}K_{a2}K_{a3}}{[\text{H}^+]^3 + K_{a1}[\text{H}^+]^2 + K_{a1}K_{a2}[\text{H}^+] + K_{a1}K_{a2}K_{a3}} \quad (8)$$

Here, the $[\text{H}^+]$ represents the concentration of the H^+ in phosphate aqueous solution, which could be derived from the experimentally detected pH values. According to the above equations, the pH dependent coefficients of different protonation states, 3H, 2H, 1H and 0H, were plotted in Figure 1a and as well as in literatures^{34, 66}. It is clear that phosphate solution is a mixture of different protonation states under the certain pH condition. For example, each of 1H and 2H accounted for approximately half with the $\text{pH} \approx 7.20$.

When those differently protonated phosphate species meet the Ca^{2+} ions, different kinds of minerals,¹⁰ such as TCP ($\text{Ca}_3(\text{PO}_4)_2$), MCPM ($\text{Ca}(\text{H}_2\text{PO}_4)_2 \cdot \text{H}_2\text{O}$), and DCPD ($\text{CaHPO}_4 \cdot 2\text{H}_2\text{O}$), would be formed (Figure 1b). The formation of those solid CaP precipitations is controlled by the following solid-liquid equilibrium equations.



The K_{sp} in equations (9)-(11) represents the solubility product^{67, 68}. It should be mentioned that the pH value also affects the above equations, since the concentration of ‘free’ dissociated phosphate species in equations (5)-(8) is also determined by the pK_a and the pH values shown in equations (2)-(4). Accordingly, the concentration of ‘free’ unbound Ca^{2+} ions could be derived from the K_{sp} values of phosphate species. Such a concentration is a useful parameter in designing an experiment: once the real concentration of Ca^{2+} ion exceeded this value, the CaP precipitation would immediately occur in solution. On the other hand, to obtain enough precipitate for X-ray diffraction (XRD) analysis, the concentration of Ca^{2+} ion should be much larger than this free Ca^{2+} concentration value.

3.2. Interfacial interactions of LAC with CaP clusters and HA surfaces

Coordination of calcium ions with phosphates. The different phosphate protonation states have different binding capacities with Ca^{2+} ions, as shown in Figure 3a and Table S1. The M062X/6-311+g(d,p) binding free energies, G_{bind} , of the complexes of phosphate species with calcium ions gradually increased as the pH value increases, indicating that the exposed oxygen atoms of the deprotonated phosphate state with negative partial charges are more affinitive to the positively charged Ca^{2+} ions than the protonated ones. It could be seen from Figure 3a that the $\text{Ca}(\eta^2\text{-OH})$ cluster had the largest binding free energy (-47.56 kcal/mol), demonstrating that the η^2 combination of Ca^{2+} ions with the fully deprotonated OH in alkaline condition was the most favorable, followed by $\text{Ca}(\eta^1\text{-OH})$ and $\text{Ca}(\eta^2\text{-1H})$. For OH and 1H species, the binding free energies with Ca^{2+} ions by η^2 mode were larger than those of the complexation in η^1 mode, which will be correlated with the charge transfer extent in the following subsection. In comparison with the complexation with 1H, the binding with 2H protonation state had the smaller binding free energy, implying that Ca^{2+} ions were relatively more easily chelated with 1H instead of 2H in neutral condition. The compound $\text{Ca}(\eta^1\text{-3H})$ in acidic condition had the smallest binding free energy (-6.15 kcal/mol). To sum up, the increase of pH value enhanced the electrostatic interaction between Ca^{2+} ions and the increased number of the exposed oxygen atoms of the deprotonated phosphate species. The increase of pH value was also found to enhance the interaction (solvation free energy) between phosphate species and water molecules by using the thermodynamic cycle at the COSMO/PBE/DNP level.⁶⁹

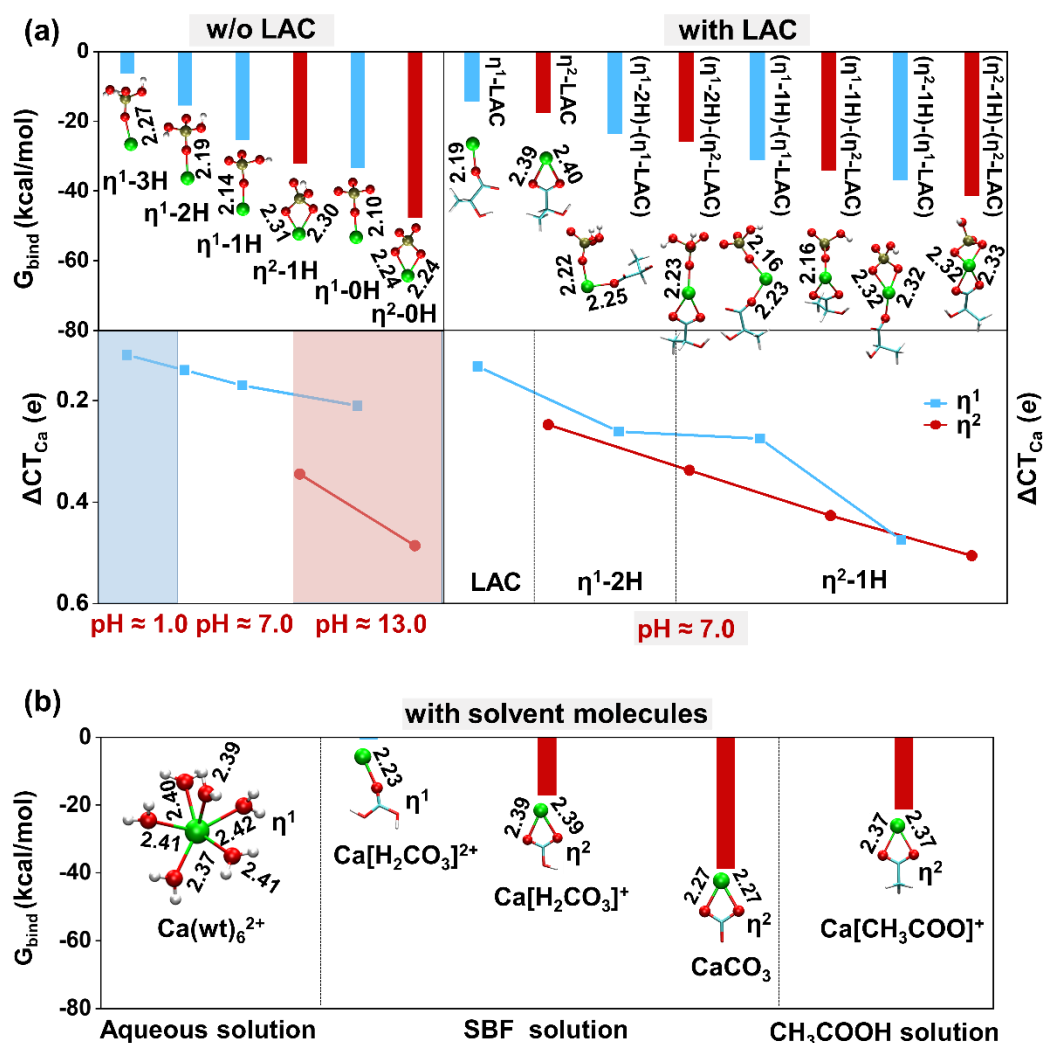


Figure 3. (a) The calculated binding free energies- G_{bind} (kcal/mol) of different compounds w/o and with LAC at the level of M062X/6-311+g(d,p); (b) the charge transfer extent described by charge difference between Ca^{2+} ion in Ca-O binding compound and an isolated Ca^{2+} ion ($\Delta\text{CT}_{\text{Ca}}$) of CaP complex without and with the attached LAC; (c) the relative binding free energies between Ca^{2+} ion and solvent molecules with the binding energy of the 6-coordinated water complex taken as a reference. The units of distance are Å. Green balls-Ca atoms, red balls-O atoms, brown balls-P atoms, white balls-H atoms.

Competitive interactions with LAC and solvent molecules. Besides phosphate species, Ca^{2+} ions could also bind with other oxygen-containing functional groups. When the pH is about 7, the LAC and the longer oligomers exist as the deprotonated states since the pK_a values of LAC and PLA are 3.86 and 3.10^{49,57}, respectively. As shown in Figure 3a, LAC could coordinate with Ca^{2+} ion both by η^1 and η^2 modes, indicating that LAC could compete with phosphate species to chelate Ca^{2+} ions in solution. Since the elongation of LAC oligomer from LAC monomer to LAC₈, does not change the binding strength too much (Table S1), we mainly analysis the binding modes of LAC with the CaP clusters. As demonstrated in Figure 3a, the stronger binding strength of η^2 coordination mode than η^1 is closely related to the extent of charge transfer between the interacting components, which was defined as $\Delta\text{CT}_{\text{Ca}}$ (charge variance of Ca caused by binding with phosphates without and with the presence of LAC). Thus, the electrostatics interaction is a dominating factor in tuning interfacial interactions with adding LAC or other additives and changing the pH conditions. With respect to the coordination of Ca^{2+} ion with water molecules in aqueous solution, the H_2CO_3 , CO_3^{2-} , and HCO_3^- ions in SBF solution and CH_3COO^- group in CH_3COOH solution may compete with phosphate species to chelate Ca^{2+} ions with non-negligible relative binding strengths (Figure 3b), whose effects were considered in the following MD simulations with the introduction of explicit solvent molecules. In addition, the formation of the new Ca-O bonds with adding LAC and other oxygen-containing solvent molecules would increase the Ca-O distances which are close to phosphorus atom (Figure 3a). For example, the Ca-O distance of 2.19 Å in η^1 -2H is elongated to 2.22 (in η^1) or 2.23 Å (in η^2) with the addition of LAC (Figures 3a). This indicates that the presence of LAC could weaken the binding ability of Ca^{2+} ions with phosphate species. It will be demonstrated through both MD simulations and experiments that the nucleation process of CaP would be slowed down by adding LAC.

It is also meaningful to survey the interfacial interactions of LAC with different HA surfaces, which represent the large sized CaP systems and crystallization product after nucleation. As displayed in Figure 4a, HA (Ca/P = 1.67) has a hexagonal crystal

structure with 44 atoms in each unit cell, in which the calcium ions are situated in two different sites, denoted as Ca1 (blue color) and Ca2 (green color), respectively. The difference between these two different kinds of calcium ions is told by the coordination number of Ca, called $N_{\text{Ca-contact}}$. The nine-coordinated Ca1 ($N_{\text{Ca-contact}} = 9$) is surrounded by nine oxygen atoms from the surrounding six phosphate groups, and Ca2 ($N_{\text{Ca-contact}} = 7$) is coordinated to seven oxygen atoms (where six coming from phosphate groups and one from the hydroxyl group). Both η^1 and η^2 coordination modes between PO_4^{3-} and Ca^{2+} ions coexist in HA crystal structure. The change of pH values could lead to the difference in Ca/P ratios,⁷⁰ and hence the different morphologies of the prepared HA.²⁸ Being stimulated by such pH-dependent phenomena, we attempt to tune the Ca/P ratios in our simulation models by changing the number of hydroxyl groups in HA unit cell (Figure 4b). In addition, the number of Ca^{2+} ions on surface per unit area, labelled as $N_{\text{Ca-surface}}/A$, was also changed with the different Ca/P ratios. Here, the surface area (A) is introduced to eliminate the influence of different surfaces. The parameter of $N_{\text{Ca-surface}}/A$ is one of the factors to affect the binding ability of different HA surface facets with different Ca/P ratios with the oxygen-containing additives.

Before we start to investigate the adsorption of LAC on HA slab, the relative stabilities of the different surfaces of HA were tested from their surface energy, E_{surf} , which is the energy difference per unit area between a slab model and the bulk crystal structure. As shown in Figure S8, among the studied HA surfaces, including (001), (100), (110), (210), (211), (112), the (001) surface was predicted to be the most stable one, in agreement with those reported DFT results using B3LYP or PBE functionals.⁷¹⁻⁷³ The thickness of four layers in the slab model is enough to give a converged value of surface energy. On the contrary, the (100) surface was expected to be most active toward adsorption of additives with the largest surface energy. In the following discussion, we would focus on (001) and (100) surfaces to study the adsorption of LAC on HA with different Ca/P ratios (Figure 4c).

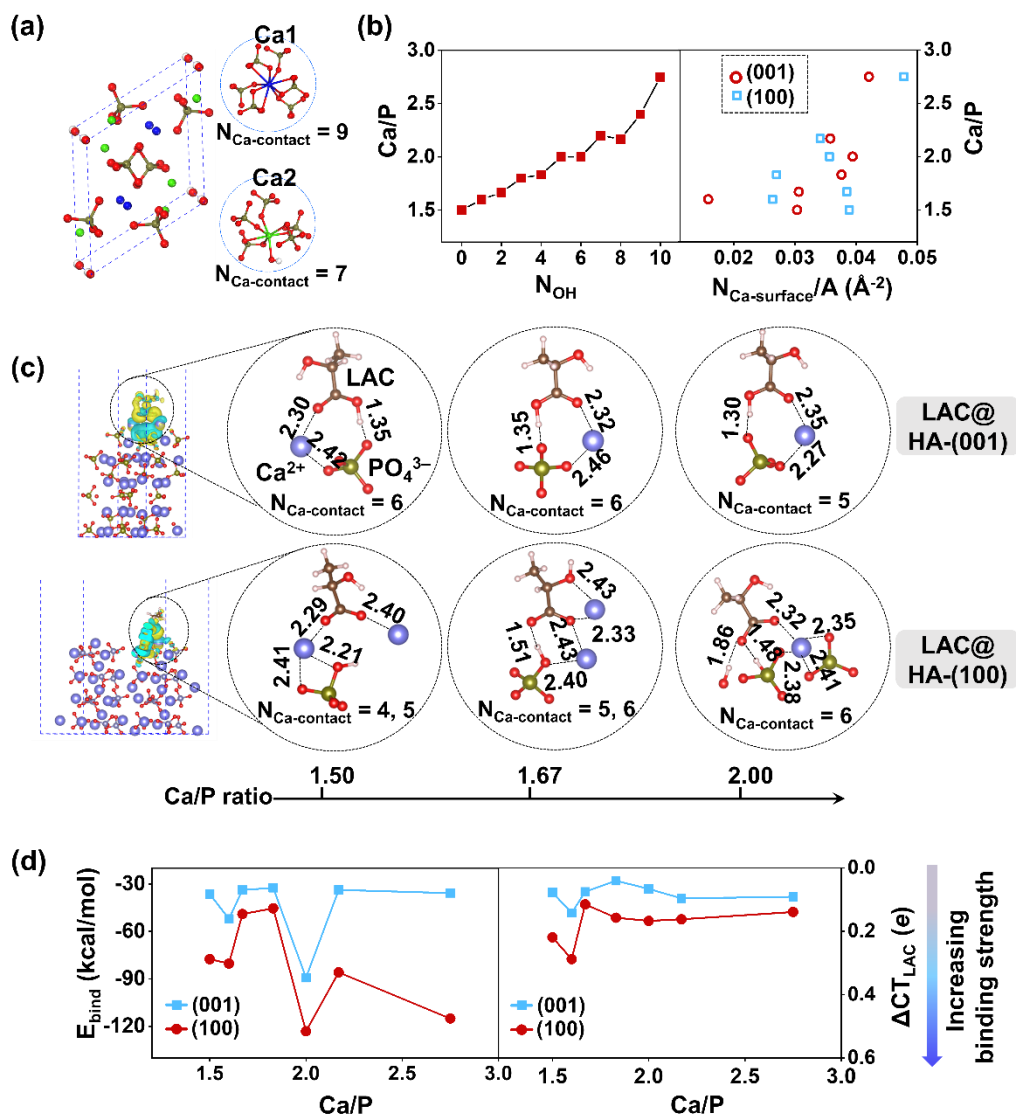


Figure 4. (a) Unit cell of HA crystal; (b) the variation of Ca/P ratios with the number of hydroxyl groups, N_{OH} , and the number of Ca^{2+} ions on surface per unit area, labelled as $N_{\text{Ca-surface}}/A$; (c) the interfacial binding modes and (d) the binding energies (E_{bind}) of LAC with the (001) and (100) surfaces with different Ca/P ratios. The charge transfer between the LAC and HA substrate is also displayed in (c) and (d), with the $\Delta\text{CT}_{\text{LAC}}$ estimated from the charge difference between LAC and the attached LAC on HA surface.

There are multiple interactions between LAC and HA surface including electrostatic interaction, proton transfer, and intermolecular hydrogen bond, as shown in Figure 4c. In addition to the above-mentioned Ca-O binding with LAC, the oxygen atoms of phosphates are favorable to form the interfacial hydrogen bonding interaction with the hydroxyl group of LAC molecule. These complicated interfacial interactions leading to the significant binding strength, E_{bind} , as shown in Figure 4d. Among the studied slab models with Ca/P ratios changing from 1.50 to 2.75, the Ca/P of 2.00 corresponds to the largest E_{bind} value (-123.13 kcal/mol on the (100) surface; -89.23 kcal/mol on the (001) surface) due to the stronger intermolecular hydrogen bonding than those of the other Ca/P ratios. The proton transfer from LAC to surface species is also be observed on (001) and (100) surfaces of HA with different Ca/P ratios (Figure 4c, Table S19). The positive values of $\Delta\text{CT}_{\text{LAC}}$ shown in Figure 4d meant that the surface acted as the electron donor and transferred charge to the LAC. This is consistent with the conclusion drawn from previous DFT study with PBE functional that the protons of adsorbates could be transferred to hydroxyl and phosphate groups on the surface.⁷² Compared to (001) surface, more active (100) surface owned the larger $\Delta\text{CT}_{\text{LAC}}$ value, because larger number of undercoordinated surface species on (100) surface facilitates the binding of LAC with dangling bonds.

3.3. The pH dependent coordination modes in CaP clusters

AIMD simulations of aqueous solution without presence of additives. AIMD simulations were further carried out to provide the more detailed information of the intermolecular interactions between phosphate species and Ca^{2+} ions in the early aggregation stage. Some simulation settings such as the number of water molecules (92), the size of box ($14 \times 14 \times 14 \text{ \AA}^3$) and functional (PBE) were similar to those used in AIMD study of the prenucleation $[\text{Ca}(\text{HPO}_4)_3]^{4-}$ cluster with PBE functional.⁴ In the present work, we aim to study the difference in coordination modes with Ca^{2+} ion between different phosphate species caused by the change of pH conditions. The selected snapshots from AIMD trajectories of aggregation of different phosphate

species around Ca^{2+} ions without (w/o) the presence of LAC are shown in Figure 5a. For 2H and 3H, which are dominant protonation states in acidity condition, the major coordination of Ca^{2+} is in η^1 modes, as displayed by a single sharp peak in the radial distribution function (RDF) of Ca-P distance. To the other end, the Ca^{2+} coordinated with the deprotonated species of 0H and 1H in basic conditions in both η^1 and η^2 modes, corresponding to the two overlapping peaks in RDF, respectively. The η^2 peak of Ca \cdots P distance is located around 3.1 Å, shorter than that (3.7 Å) in η^1 binding mode. This is also in line with our DFT optimizations of various CaP clusters in Figure 3a. For example, the Ca \cdots P distance in the strongly bound η^2 -0H cluster is 2.9 Å, shorter than that (3.9 Å) in η^1 coordination mode of η^1 -0H. As mentioned above, the simulation of pH \approx 7.4 involved the two coexisting 2H and 1H species, which have different binding modes toward Ca^{2+} ions, i.e., 2H is bound in η^1 but 1H is chelated in both η^1 and η^2 modes. The coexisting η^1 and η^2 modes between PO_4^{3-} and Ca^{2+} ion were also revealed by in situ XANES experiments that the most abundant clusters were detected to be $\text{Ca}(\eta^2\text{-PO}_4^{3-})_2\text{L}_2$ (L = H_2O or $\eta^1\text{-PO}_4^{3-}$) during ACP formation.⁷⁴ The prenucleation $[\text{Ca}(\text{HPO}_4)_3]^{4-}$ cluster was found to take the configuration of $(\text{Ca}(\eta^2\text{-HPO}_4^{2-})_2(\eta^1\text{-HPO}_4^{2-}))(\text{H}_2\text{O})_2$ from the AIMD study with PBE functional.⁴

It was found from the SDF (Figure 5a) that Ca^{2+} ions were mainly chelated with non-hydroxyl oxygen atoms of phosphate species. In the simulation of pH \approx 1.0, one hydrogen atom of the 3H was pulled by a nearby water molecule through the hydrogen bonding. Such a hydrated proton was stabilized by the formation of concerted hydrogen bonding chain in the surrounding water solvent molecules. As a result, the 3H in Figure 5a looks like to ‘lose’ a proton to form a deprotonated 2H species, which was demonstrated to have stronger binding free energies than the 3H by our DFT results (Figure 3a). When the pH value is increased to 5, the more compact CaP cluster is formed with the two 2H species bridged by two Ca^{2+} ions through non-hydroxyl oxygen atoms of the deprotonated phosphates. The number of non-hydroxyl oxygen atoms is increased with the further deprotonations in the increased pH conditions, leading to the formation of the cage-like CaP prenucleation cluster. Each Ca^{2+} ion sits at the cage

vertex to bridge 3 ~ 4 oxygen atoms of phosphates with significant binding energies. The water molecules were pushed outside the CaP cluster cages, exhibiting strong tendency of forming precipitate.

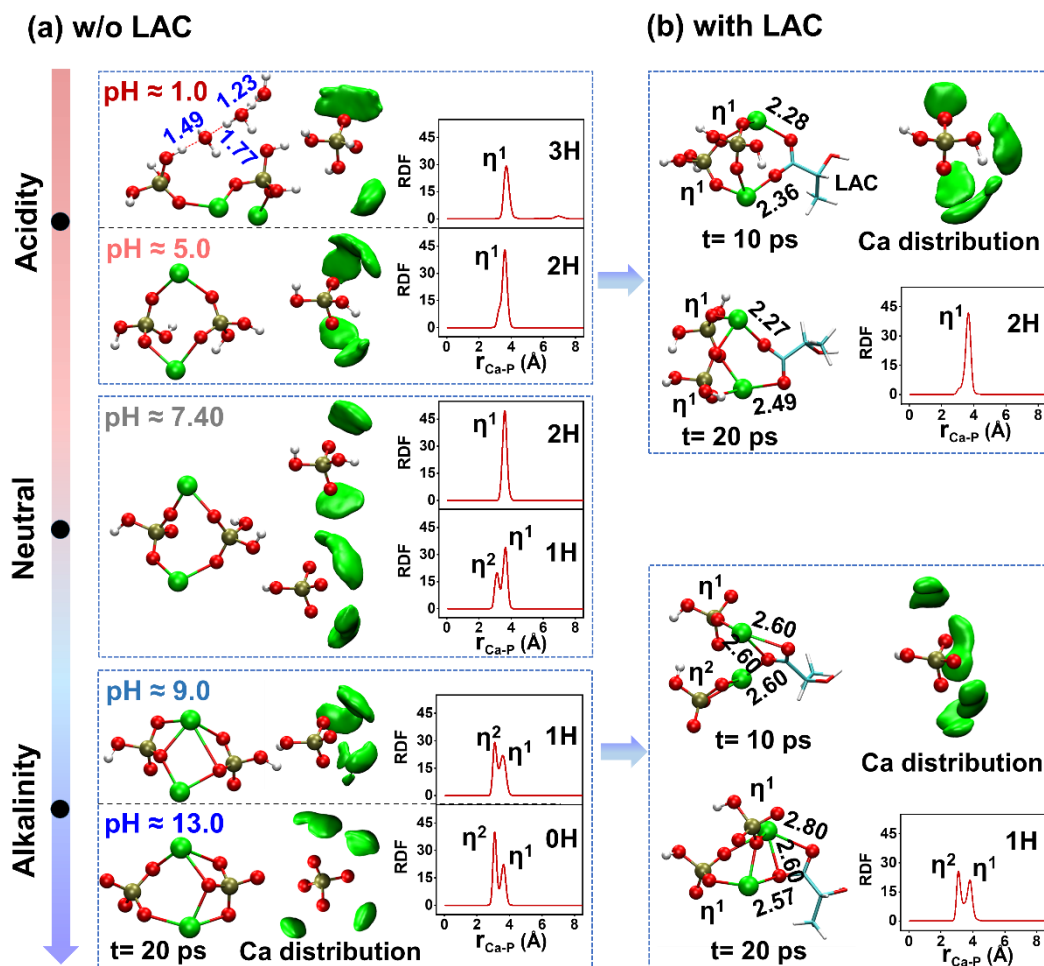


Figure 5. The selected AIMD snapshots, SDF (isosurfaces for Ca atoms are represented in green, and isovalues of Ca atoms are set to be 50 particles/nm³), and RDF of studied systems (a) w/o LAC and (b) with LAC, respectively. Only the adjacent water molecules with 2.0 Å are shown in the sampled snapshots at 20 ps for clarity.

Influence of LAC oligmers on the coorination number and hydrogen bonding.

The addition of LAC introduced the competitive coordination interactions with Ca²⁺ ions, destroying the cage structure of CaP cluster (Figure 5b). Consequently, the coordination number of Ca²⁺ ions, $N_{Ca-contact}$, is decreased from 3.0 to 1.5 in the

deprotonated 1H system in basic condition. As shown in Figure 5b, LAC would bind with Ca^{2+} ions in both η^2 and η^1 modes, decreasing the contact chance of phosphates with Ca^{2+} ions.

It is difficult to perform AIMD study on the intermolecular interactions of longer LAC_8 oligomer in CaP aqueous solutions with different pH conditions due to the huge computational costs with much larger simulation boxes to accommodate the LAC_8 chain (whose length was predicted to be 22.9 Å by DFT optimization). To study the influence of LAC oligomers in a much larger space and time scale, we carried out classical MD simulations based on GAFF2 on three model systems: w/o additives, with LAC and with LAC_8 , respectively, as shown in Figures 6. To gain insight of calcium phosphate precipitation and biological mineralization in the physiological environment, the SBF solutions with ion concentrations and pH value similar to the human blood plasma¹⁷⁻²⁵ were introduced in our MD simulations. As listed in Table S20, the SBF solutions are consisting of Na^+ , Cl^- , HCO_3^- , K^+ , Mg^{2+} , Ca^{2+} , HPO_4^{2-} and SO_4^{2-} ions.

It could be concluded that the coordination modes of phosphate species and Ca^{2+} ions obtained by GAFF2 based MD simulations (Figure 6, Figure S9) were similar to those revealed from AIMD simulations, validating the adopted force field parameters (see supporting information for the details). Compared with the control group (w/o additives), LAC molecules were mainly chelated with Ca^{2+} ions through its carboxyl group by η^2 and η^1 modes (Figure 6). Besides, one Ca^{2+} ion could chelate with two or more LAC molecules when the pH is larger than 5.0. In the strong acid solution of pH ≈ 1.0 , the LAC oligomers are not deprotonated and have little binding affinity to the CaP clusters, as indicated by the long distance between the LAC additives and the 3H species in Figure 6. Same as what has been found in AIMD simulations, the chelation of LAC and Ca^{2+} ions reduced the number of Ca^{2+} ions near the phosphate species.

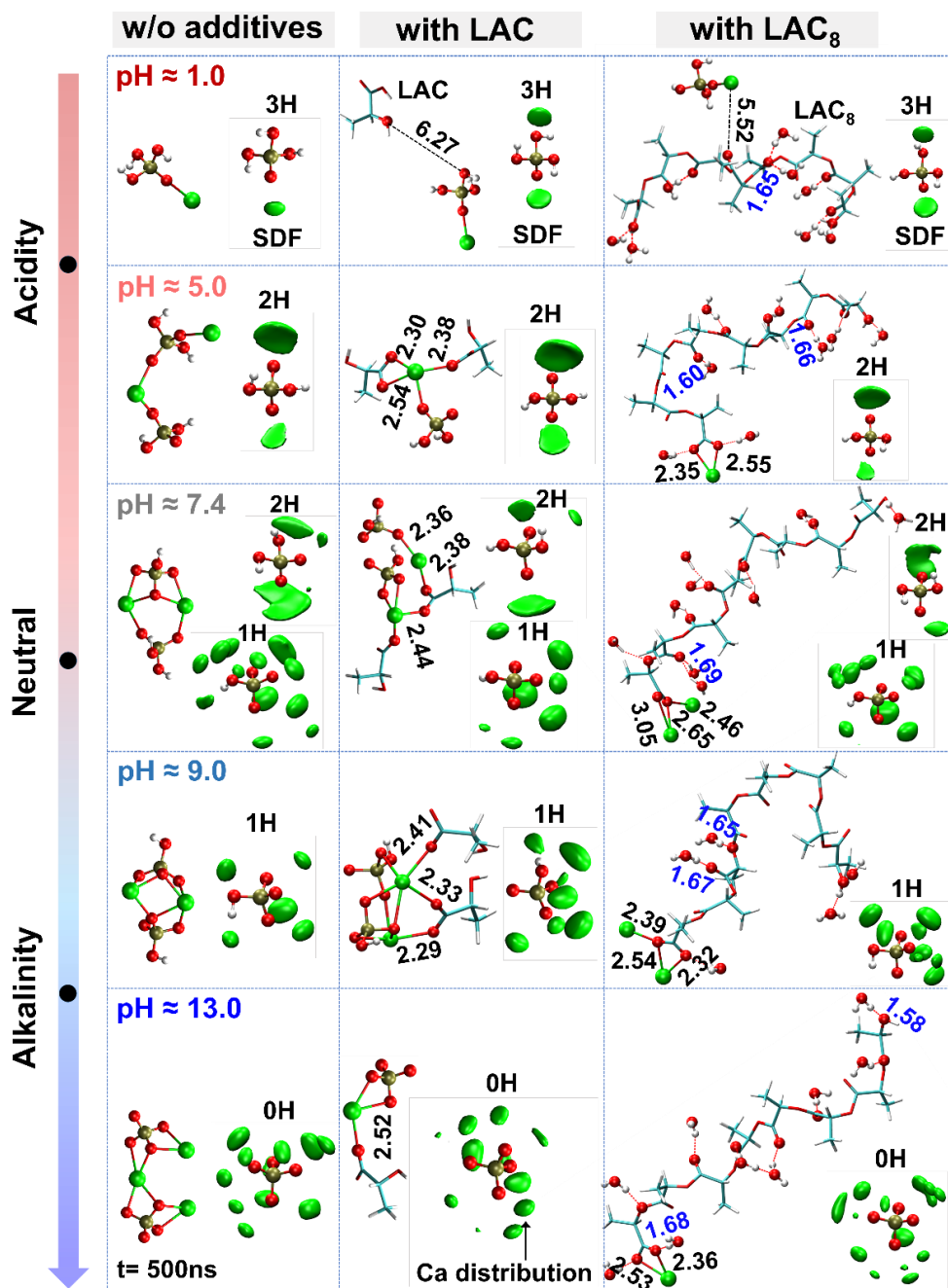


Figure 6. The selected GAFF2 based MD snapshots and SDF (isosurfaces for Ca atoms are represented in green, and isovalues of Ca atoms are set to be 50 particles/nm³ except the system with pH \approx 1.0 (20 particles/nm³)) of the CaP clusters in SBF solution w/o additives, with LAC, and with LAC₈, respectively. Only the adjacent water molecules with 2.0 Å are shown in the sampled snapshots at 20 ps for clarity. The ions of SBF were also hidden in snapshots.

The longer oligomer, LAC₈, is facvor to take a zigzag conformation in CaP aqueous solution. LAC₈ preferred to chelate with Ca²⁺ ions through its terminal carboxyl group, and the other oxygen-containing functional groups could form hydrogen bonds with nearby water molecules (Figure 6). The cage like CaP clstuter is diificult to form in the vicinity of LAC₈. To compare with the coordination modes of Ca²⁺ ions and phosphate species under SBF solution, the coordination modes of Ca²⁺ ions and phosphate species under aqueous solution with pH \approx 7.4 were also studied (Figure S9). The exsistence of various ion compositions in SBF solution brought little difference in coordination modes of CaP from those in aqueous solution. The present GAFF2 based MD simulations on aqueous solution of Ca²⁺ ions and OH species gave similar results to the previous MD simulations based on INTERFACE force field, which demonstrated the η^1 and η^2 coordination modes by the two peaks at 3.16 and 3.78 Å in RDF³⁹.

3.4. Aggregation of CaP clusters in aqueous and SBF solutions

In order to further trace the evolution process of CaP clusters at different simulation time durations, we presented the snapshots in Figure S10 at simulation time of 0 ns, 100 ns, 300 ns and 500 ns, respectively. We lay emphasis on the MD simulation models with pH \approx 7.4 and Ca/P = 2.00, in which 1H and 2H phosphates were randomly distributed in the SBF solutions, as shown in Figure 7a and Figure S10a. With the evolution of MD simulation, the initial small sized and loosely bound CaP clusters attracted each other and aggregated into the larger clusters consisting of about 10 Ca²⁺ ions at 500 ns. By the end of the MD simulation, about 58% of Ca²⁺ ions in those CaP clusters were coordinated by phosphate species, and those Ca²⁺ ions were found at the upmost surface of the formed clusters. It could be seen from Figure 7b and Figure S10b, LAC monomer could chelate with Ca²⁺ ions during the entire simulation time. However, the longer oligomer, LAC₈, was mainly keeping free because of its weaker binding ability to Ca²⁺ ions than LAC monomer. It is also shown that the carboxyl group of LAC molecules could compete with phosphate species (1H and 2H) to bind with Ca²⁺ ions in η^1 and η^2 modes (Figures 7b). We selected two typical clusters, cluster 1 and

cluster 2, to illustrate the binding modes of oxygen-containing groups, such as phosphates, LAC, and ions in SBF solution, with Ca^{2+} ions, as shown in the inset of Figure 7. In cluster 1, two bridging Ca^{2+} ions are in distorted tetrahedral and trihedral coordinations formed by the surrounding LAC molecules. The cluster 2 is bigger and more complicated than cluster 1. The two Ca^{2+} ions in cluster 2 binding with two LAC monomers and 2H phosphates through trihedral and bilateral coordinations, while all of the other three Ca^{2+} ions are tetrahedrally coordinated with the surrounding 1H species to form a compact CaP cage. There were still several free 2H species that did not coordinate with Ca^{2+} ions during 500 ns MD simulation (Figure S11) because of its relatively weaker chelate affinity to Ca^{2+} ions than 1H. The CSFF based MD study of interaction between CaP and cellulose nanofibrils gave a similar conclusion that the activity of hydroxyl groups of substrate was not high enough to cause adsorption of Ca^{2+} ions from the solution.³⁷

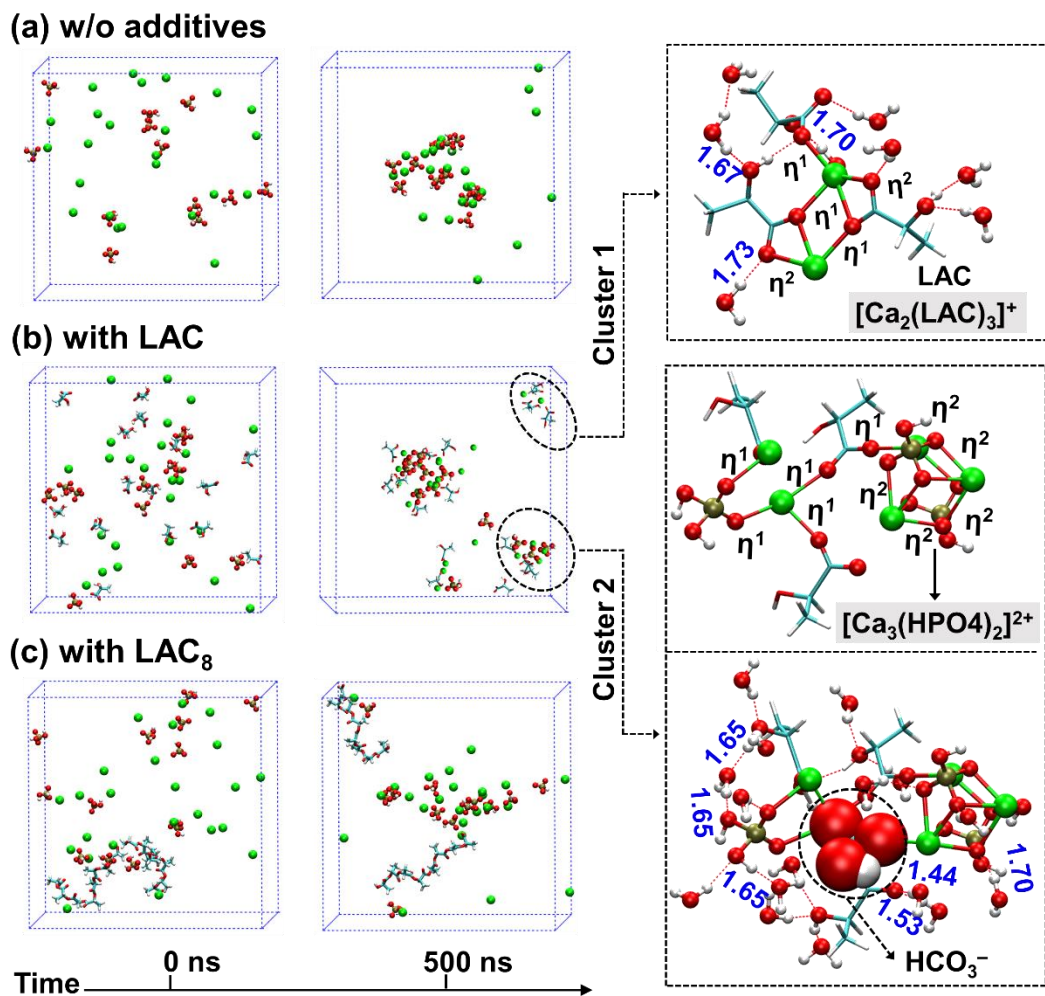


Figure 7. MD snapshots with the explicit illustration of Ca^{2+} ions, 1H, 2H ions, LAC and LAC_8 at 0 ns and 500 ns of CaP systems ($\text{Ca/P} = 2.00$, $\text{pH} \approx 7.4$) in SBF solutions (a) w/o additives, (b) with LAC and (c) with LAC_8 , respectively. Water molecules are omitted for clarity. Ca^{2+} ions are shown as green (VDW), 1H and 2H ions as CPK and VDW, LAC and LAC_8 as Licorice.

Some other oxygen-containing units such as HCO_3^- ions of SBF solution also competed with phosphate species to chelate Ca^{2+} ions through Ca-O ionic bonding. It is hence not surprising to find from our MD simulations that larger sized CaP clusters could be formed in a shorter simulation time under aqueous solution compared with that under SBF solution (Figure S12). For example, the snapshots at 100 ns for aqueous solution displayed a cluster that enclosing 11 Ca^{2+} ions, larger than that (6 Ca^{2+} ions)

appeared in SBF solution (Figures S12a, S10a). In other words, the lack of competitive interactions of HCO_3^- ions in aqueous solution could give more chance for phosphate species binding to Ca^{2+} ions, resulting in the formation of a larger CaP cluster with a short simulation time. In addition, LAC could form hydrogen bonds with nearby water molecules through its hydroxyl and carbonyl groups. Phosphate species could also function as hydrogen bonding donor or acceptor in binding with the water solvent molecules through its oxygen atoms and hydrogen atom of hydroxyl group.

MD simulations of basic conditions with $\text{pH} \approx 9.0$ and $\text{pH} \approx 13.0$ (Figures S13-S14) demonstrated similar aggregation behavior of CaP clusters to the case of $\text{pH} \approx 7.4$. But in the acid solution, 3H species owned the weak binding strength with Ca^{2+} ions and only small CaP clusters containing 2-3 Ca^{2+} ions were formed even at 500 ns trajectory (Figure S15). Nearly 92% Ca^{2+} ions and 83% 3H phosphates were staying in the free state with addition of LAC (Figure S15b).

In order to evaluate the whole evolution process of clusters aggregation with a qualitative descriptor, we counted the number of phosphate species and Ca^{2+} ions in each cluster. In the first step, we calculated the radial distribution functions (RDF) of Ca-P (phosphate species) (Figure S16), and then the cutoff distance (R) was set based on the local minimum of corresponding RDF. Accordingly, the number of phosphate species or Ca^{2+} ions in each cluster was counted within the cutoff radius R. In our code, we took one of the Ca^{2+} or phosphates as the center, and searched all the target species (Figure S17). Here, we selected the largest CaP cluster to estimate the value of $N_{\text{Ca-contact}}$, which represents the number of Ca^{2+} ions around phosphate species within the cutoff radius, with the results shown in Figures S18-S24. An increase in $N_{\text{Ca-contact}}$ was observed from Figures S18, S20, S23 as the simulation time increases. The chelation of LAC with Ca^{2+} in η^1 and η^2 modes prevented Ca^{2+} ions from binding with phosphate species, yielding small value of $N_{\text{Ca-contact}}$ in the neutral and alkaline conditions with different Ca/P ratios (Figures S18, S20, S23). In contrast, the presence of LAC_8 had little effect on the size of the largest CaP cluster and the number of $N_{\text{Ca-contact}}$ (as shown

in Figures S18, S20, S23) because of its weaker binding ability to Ca^{2+} ions in all pH conditions.

Electrostatic potential surfaces (EPS) of CaP clusters. The uneven charge distribution at the surface of the CaP cluster leads to the significant electrostatic interactions, which would play an important role in the binding affinities toward proteins and nucleation processes. In our recent work, the nano-sized CaP clusters had been found to have binding specificity toward AP2 protein through electrostatic interactions with the charged residues.⁴⁴ We are now curious about the change of electrostatic potential distribution of the CaP aggregates upon the complexation with LAC oligomers, LAC_n ($n = 1, 8$). As shown in Figure 8, without the presence of additives the CaP clusters had unevenly distributed EPS maps, in which the exposed groups are dominated by the positively charged calcium ions (blue color) on the cluster surfaces under both acid and basic conditions. Those exposed Ca^{2+} ions could be coordinated with nearby water molecules to stabilize the clusters. The addition of LAC led to the charge redistribution to allow negatively charged phosphate species (red color) being exposed at the outmost surface of CaP cluster. This was because LAC could compete with phosphate species to chelate with Ca^{2+} ions in both η^1 and η^2 modes, resulting in the exposed species to be phosphate species instead of Ca^{2+} ions. However, the relatively weaker electrostatic interaction between LAC_8 and Ca^{2+} ions did not result in such a large change in the EPS of CaP cluster as what was found above with the additive of LAC monomer.

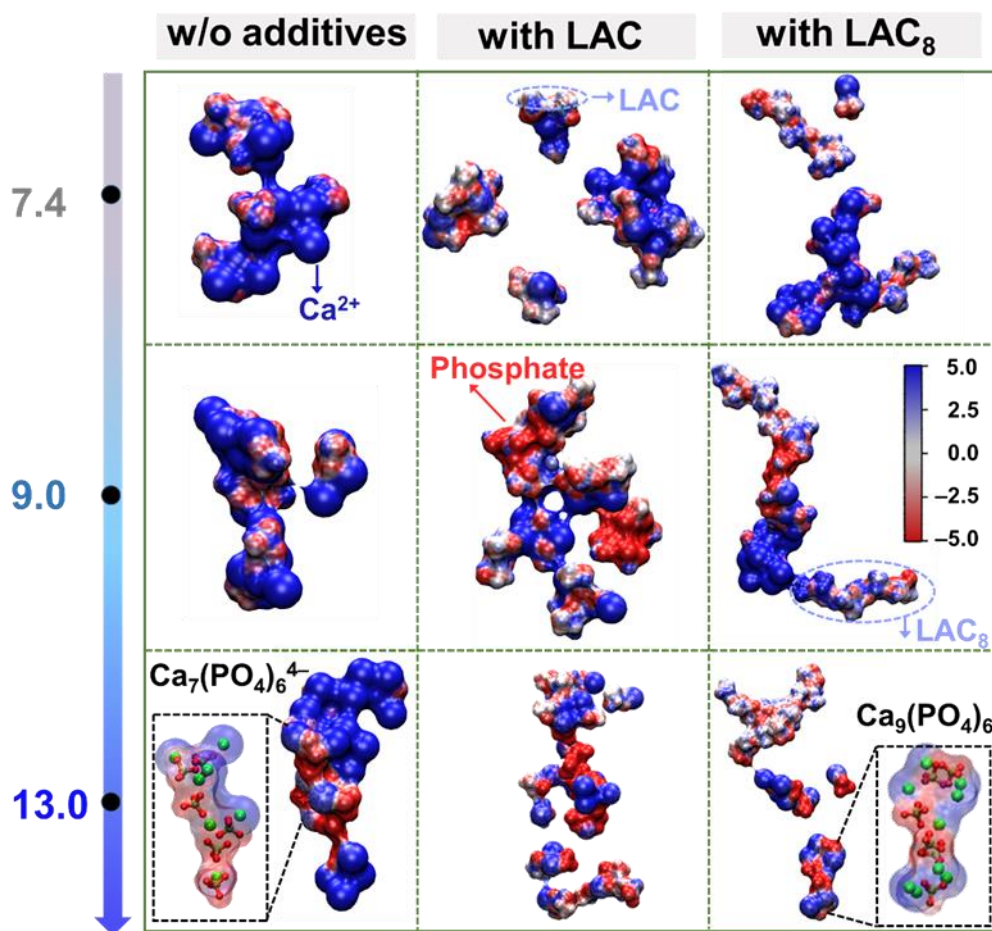


Figure 8. Electrostatic potential surface maps of CaP clusters without and with LAC_n oligomers ($n = 1, 8$) under different pH values ($\text{Ca/P} = 2.00$) sampled at 500 ns of MD simulation. The electrostatic potentials are calculated with APBS software. The unit for electrostatic potential is $k_B T/e$.

Moreover, our MD simulations demonstrated the formation of the Posner-like clusters, such as Ca-deficient complex $[\text{Ca}_7(\text{PO}_4)_6]^{4-}$ and Posner's cluster $\text{Ca}_9(\text{PO}_4)_6$ (Figure 6). The Posner's cluster⁷⁵ was usually considered to be crucial in the nucleation process of CaP in solution. It was also reported from classical molecular dynamics simulations that PO_4^{3-} and Ca^{2+} ions could aggregate into a $[\text{Ca}_8(\text{PO}_4)_6]^{2-}$ cluster, which is similar to Posner's cluster.⁹ Those small sized CaP clusters could also be assembled in the form of multiple Posner's clusters such as $\text{Ca}_{108}(\text{PO}_4)_{72}(\text{OH})_{10}$ (called Posner₁₂) on the collagen surface.³⁹ The electrostatic interaction between the positively charged

arginine residue and PO_4^{3-} ion could regulate the HA nucleation process by the arginine residue of collagen.⁴⁰ The distribution of positively and negatively charged groups at the CaP cluster surface could be correlated with the zeta potential experiments in the following subsection.

3.5. High-throughput experimentations (HTE) of CaP clustering processes

The above mentioned calculation results have shown that the nucleation process of CaP was controlled by comprehensive factors including the pH value, Ca/P ratio, additives, and solvent molecules. Thus, in order to further explore the influence of those factors on the nucleation process at macroscopic scales, the HTE technique was applied to synthesize various nanoparticles of CaP in the present work. The experimental details were introduced in the Supporting Information. Totally 120 experimental groups were prepared based on the three factors, i.e., w/o and with the addition of LAC (factor 1), changing Ca/P ratios from 1.20 to 2.50 (factor 2), and using different solvent media (factor 3), with the results shown in Figure 9. The UV-Vis absorbance values were used to detect the amount of the formed precipitation. It can be found from Figure 9a and 9b that the UV-Vis absorbance values decreased with addition of LAC in both SBF and aqueous solutions, indicating that the addition of LAC would inhibit the generation of precipitation. As the Ca/P ratio was greater than 2.0, the presence of LAC had small effect on the amount of precipitation with the concentration of Ca^{2+} ions, $[\text{Ca}^{2+}]$, of about 0.01 mol dm^{-3} . If the Ca^{2+} concentration was too low, $[\text{Ca}^{2+}] = 0.005 \text{ mol dm}^{-3}$, the UV-Vis absorbance of CaP aqueous solution vanished without and with the presence of LAC. It could be concluded from the above experiments that the addition of LAC inhibited the nucleation process of CaP, supporting the foregoing theoretical predictions.

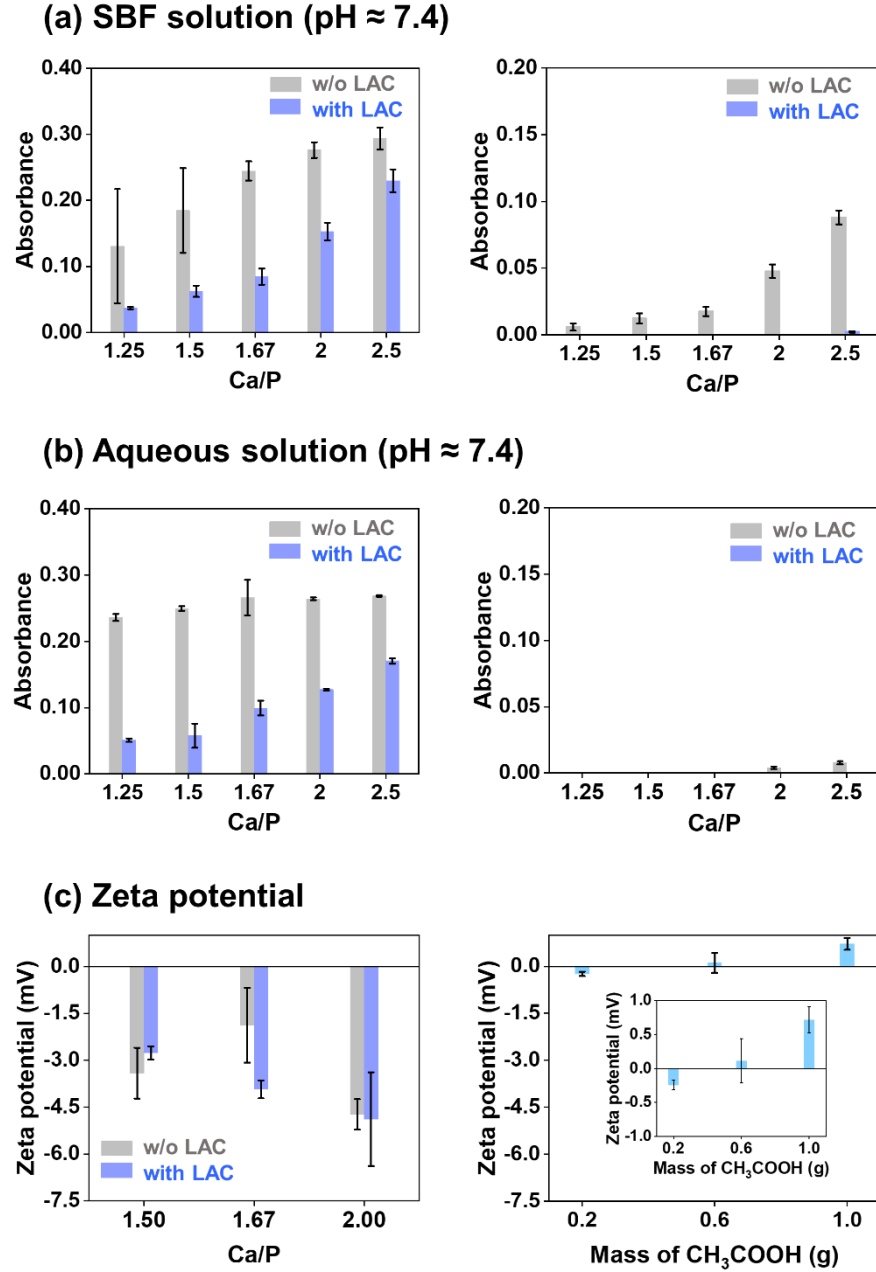


Figure 9. The absorbance values (450 nm) of the experimental groups with different Ca/P ratios under (a) SBF solution and (b) aqueous solution before and after adding LAC, where the concentrations of Ca^{2+} ions are 0.01 mol dm^{-3} (left) and $0.005 \text{ mol dm}^{-3}$ (right), respectively, (c) zeta potentials of CaP with different Ca/P ratios and as-prepared samples with different mass of CH_3COOH .

Zeta potential was used to estimate the average surface electronegativities. We selected three experimental groups with Ca/P of 1.50 ($\text{CaP}_{1.50}$), 1.67 ($\text{CaP}_{1.67}$) and 2.00

(CaP_{2.00}) , respectively, to measure their zeta potentials by increasing the concentration of Ca²⁺ ion to 0.15 mol dm⁻³. All these three samples had negative zeta potentials (Figure 9c, Table 1), indicating that they had net negative surface charges. The presence of LAC caused a slight increase in the zeta potentials of the samples CaP_{1.67} and CaP_{2.00} with different Ca/P ratios. Although the zeta potential values measured by different research groups may be different from each other, the experimental data of various nanoparticles collected in Table 1 could still give some general trends, especially for the potential of binding with the biological molecules. As shown in Table 1, the HA nanoparticles with different morphologies,¹⁴ Ag@HA,³⁰ F-Cl@HA,³¹ and Au/Ag@HA,³⁵ had negative zeta potentials, some of which were found to be efficiently taken up by A375 cells and MG-63 cells. The HA particles with several kinds of oxygen-containing additives, such as, citric acid@HA,³² oxalic acid@HA³³ and malic acid@HA³⁴ also exhibited negative zeta potentials, whose values were dependent on the pH values (Table 1). Similar to HA nanoparticles, some other nanoparticles including Au/Ag nanoparticles,³⁵ cerium oxide nanoparticles⁷⁶ and iron oxide nanoparticles²⁹ with negative zeta potentials were able to effectively absorbed on MG-63, A549, and HeLa cells. In addition to the aforementioned nanoparticles, two-dimensional materials such as arsenene could inhibit the proliferation of NB4 cancer cells (82% inhibition) with the negative zeta potential (-17.8 mV), small size, and the planar structure.³⁶ It was addressed that highly negative zeta potential contributed to accumulate on Ca²⁺ ions, which was conducive to bone regeneration, thus enhanced the bioactivity of the nanoparticle.³⁰ On the contrary, low zeta potential would lead to aggregation or coagulation of nanoparticles, favored by interparticle interaction. To summary, the surface electronegativities and zeta potential could be taken as one of the descriptors in understanding the aggregation and biological functions of CaP particles.

Table 1. The zeta potential values of the as-prepared CaP samples and some other nanoparticles with and without additives.

Samples	Zeta potential (mV)	Tumor cells	References
CaP with additives			
LAC@CaP _{1.50}	-2.77 ± 0.21 (pH ≈ 7.4)	/	This work
LAC@CaP _{1.67}	-3.92 ± 0.29 (pH ≈ 7.4)	/	This work
LAC@CaP _{2.00}	-4.89 ± 1.50 (pH ≈ 7.4)	/	This work
Citric acid@HA	0.00 ~ -50.00 (pH = 3~11)	/	32
Oxalic acid@HA	0.00 ~ -40.00 (pH = 4~11)	/	33
Malic acid@HA	0.00 ~ -20.00 (pH = 5~11)	/	34
CaP or HA nanoparticles without additives			
CaP _{1.50} (Ca/P=1.50)	-3.41 ± 0.82 (pH ≈ 7.4)	/	This work
CaP _{1.67} (Ca/P=1.67)	-1.88 ± 1.20 (pH ≈ 7.4)	/	This work
CaP _{2.00} (Ca/P=2.00)	-4.73 ± 0.49 (pH ≈ 7.4)	/	This work
HA-A (Ca/P=1.67)	-9.62 ± 0.65 (pH ≈ 10.0)	A375 cells	14
HA-B (Ca/P=1.67)	-10.90 ± 0.91 (pH ≈ 10.0)	A375 cells	14
HA-C (Ca/P=1.67)	-12.50 ± 0.78 (pH ≈ 10.0)	A375 cells	14
HA-D (Ca/P=1.67)	-16.60 ± 0.60 (pH ≈ 10.0)	A375 cells	14
HA-E(Ca/P=1.67)	-10.60 ± 0.69 (pH ≈ 10.0)	A375 cells	14
HA (Ca/P=1.21)	-2.0 (pH ≈ 7.2)	MG-63 cells	35
Au@HA	-15.0 (pH ≈ 7.2)	MG-63 cells	35
Ag@HA	-13.4 (pH ≈ 7.2)	MG-63 cells	35
Au-Ag@HA	-11.6 (pH ≈ 7.2)	MG-63 cells	35
Ag@HA	10.0 ~ -25.0 (pH = 4~11)	/	30
F-Cl@HA	-17.0 ~ -37.0 (pH = 5, 7.4, 9)	/	31
Other nanoparticles			
Au-NP	-31.5 (pH ≈ 7.2)	MG-63 cells	35
Ag-NP	-26.72 (pH ≈ 7.2)	MG-63 cells	35
Au-Ag-NP	-5.93 (pH ≈ 7.2)	MG-63 cells	35
Cerium oxide -1	-16.26 (pH ≈ 7.0)	A549 cells	76
Cerium oxide -2	-43.10 (pH ≈ 13.0)	A549 cells	76
Cerium oxide-5	-42.46 (pH ≈ 13.0)	A549 cells	76
Iron oxide	-9.0 ~ -45.0 (pH = 1~13)	HeLa cells	29

A CaP implant is partially dissolved and releases calcium and phosphate ions into the surrounding body fluid, resulting in an increase in the concentrations of calcium and phosphate ions in a local region under physiological environment. It was also interesting to study the role of LAC in nucleation process by increasing the concentrations of calcium and phosphate, so that the prepared samples were large

enough to get their XRD patterns (Figure 10a and Figure S25) The characteristic peaks of (020), (021), (041) without and with the addition of LAC were in good agreement with the standard XRD pattern for DCPD (JCPDS: 09-0077), which indicated that all of them were composed of DCPD phase. It could be seen that the intensity of characteristic peaks such as (020), (021), (041) fluctuated with the increase of the stirring time (Figure 10b), which indicated that the nucleation of CaP was a dynamic process in solution. It was reported that DCPD was the most kinetically favorable phase in the precipitation, when the concentrations of calcium and phosphate are 1 ~ 5 times higher than those in the normal SBF solution at pH values of 5, 7.4, and 10, respectively.⁷⁷ DCPD was generally regarded as a precursor phase for HA formation.^{78,}⁷⁹ As revealed in Figure S25c, the differences in the observed XRD patterns at different reaction times were insignificant, particularly when the stirring time was less than 24 h. The peaks at about 25.90 ° and 31.86 ° with the prepared samples at 24 h were consistent with the standard card (JCPDS: 09-0432)'s (002) and (211) reflection, suggesting that the prepared samples were HA. It means that DCPD could act as an intermediate phase during the formation of HA. The relative stability of DCPD could be rationalized by the relatively large binding free energy of the deprotonated 1H species with Ca²⁺ ions. The as-prepared samples showed the plate-like morphology with a dimension of 100-200 nm by transmission electron microscopy (TEM) without and with the addition of LAC (Figure 10c and Figure S26).

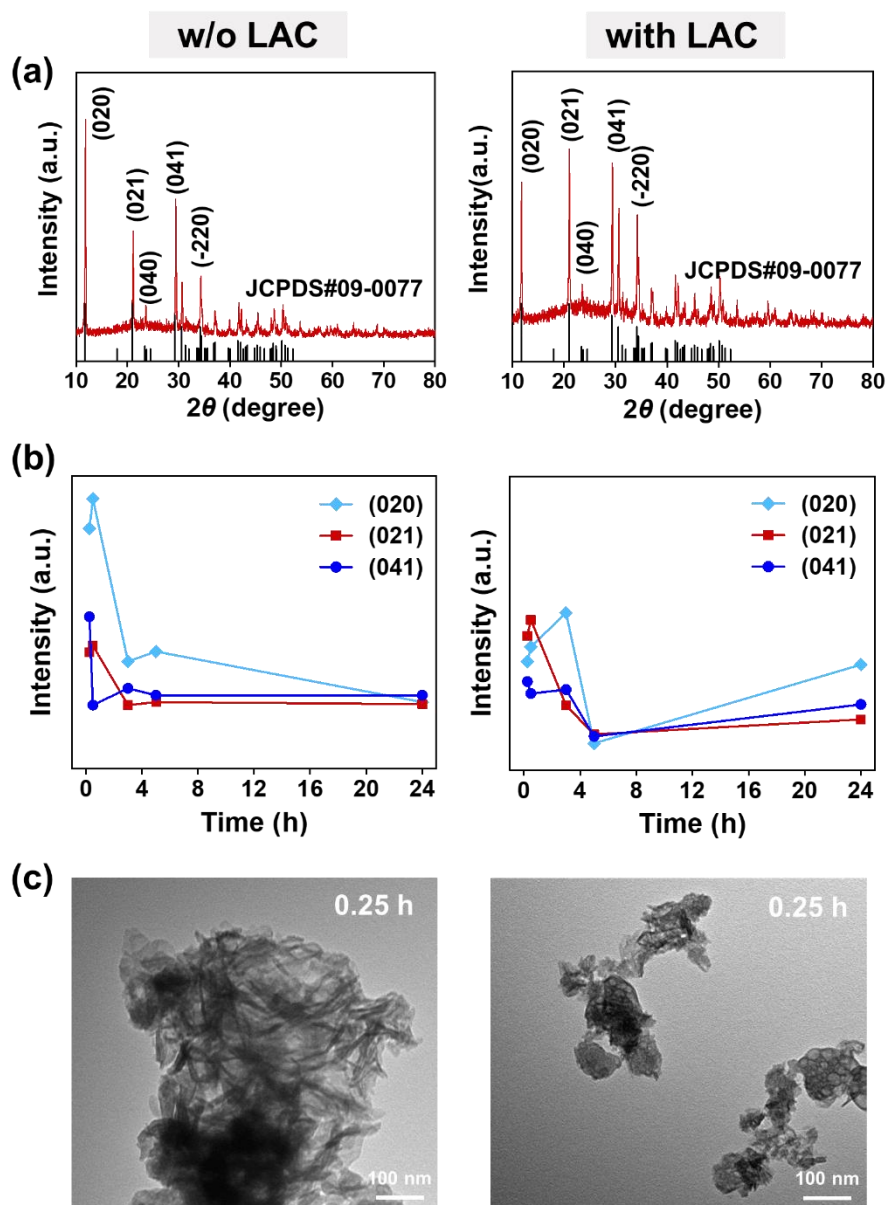


Figure 10. (a) XRD patterns, (b) intensity of characteristic peaks such as (020), (021), (041), (c) TEM images of the as-prepared samples, which were formed after 0.25 h of drying in a vacuum oven.

In addition, we used another method to prepare DCPD with the introduction of CH_3COOH in aqueous solution, with the preparation process referred to the literature⁸⁰, as shown in Figure 11a. With the increase of mass of CH_3COOH from 0.10 to 1.00 g (factor 1), the final precipitate was composed of DCPD and $\text{Ca}(\text{OH})_2$, judged by the characteristic XRD peaks (JCPDS: 09-0077 and 44-1481). The intensity of

characteristic peaks at about 11.68° (020) and 20.93° (021) of DCPD, 18.01° (001) and 34.10° (101) of $\text{Ca}(\text{OH})_2$ fluctuated greatly when the mass of CH_3COOH was increased from 0.40 to 0.70 g (Figure 11b). The zeta potentials of samples with mass of CH_3COOH of 0.2, 0.6 and 1.0 g were also analyzed, respectively. As shown in Figure 9c, the as-prepared samples had neutral surface net charges, favoring the formation of the larger sized precipitation.

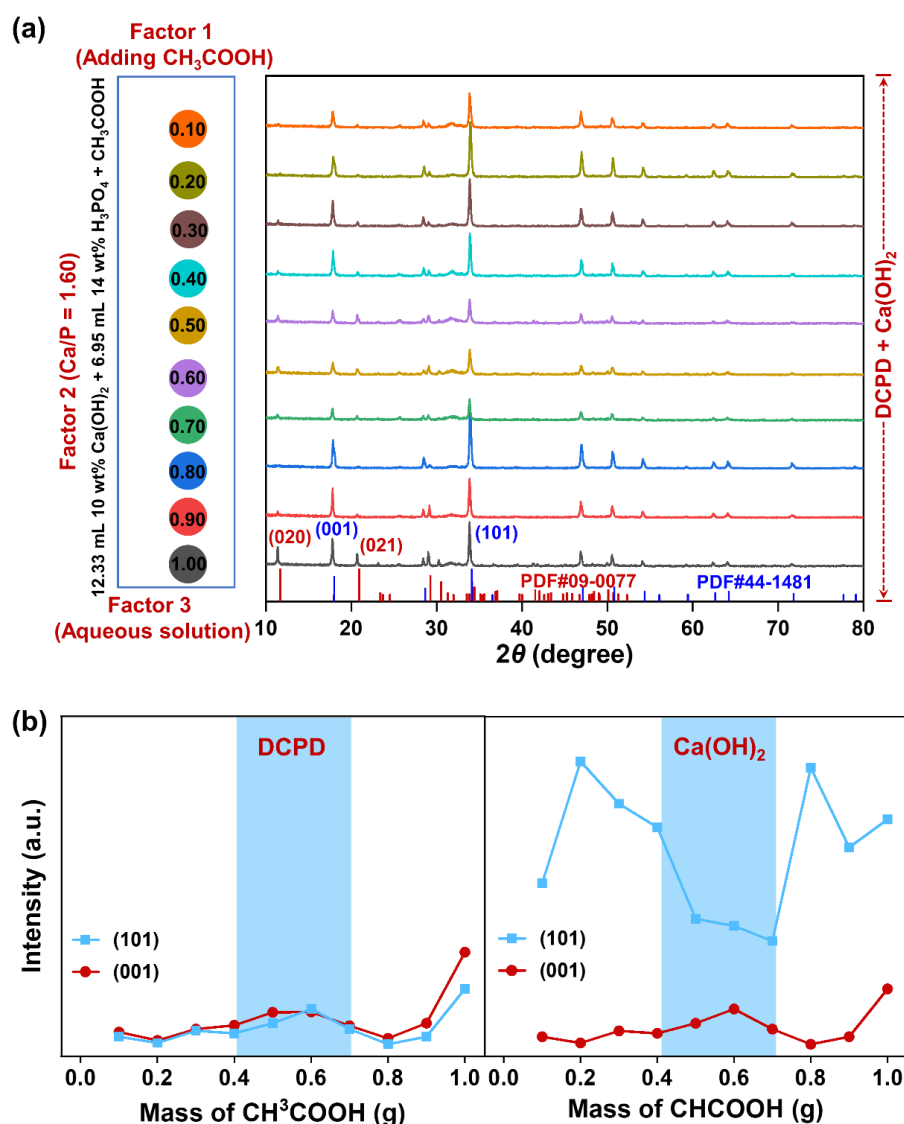


Figure 11. (a) The preparation process and XRD spectrum of as-prepared samples by high-throughput experimentations equipment, (b) intensity of characteristic peaks such as (020) and (021) surfaces of DCPD, (001) and (101) surfaces of $\text{Ca}(\text{OH})_2$, respectively.

4. Conclusion

In this work, we have systematically investigated the influence of adding LAC, changing the values of Ca/P ratio and pH, and introducing different solvent environments on the aggregation and nucleation of CaP. The pH dependent protonation states of phosphate species and their coordination modes of Ca^{2+} ions were analyzed through AIMD and MD simulations with different time and spatial scales. The competitive interaction of LAC with Ca^{2+} ions of CaP clusters or on the surfaces of HA was featured by electrostatic interaction, which was also affected by the intermolecular hydrogen bonding between LAC and phosphate. With the addition of LAC in neutral and alkaline solutions, those competitive interactions from the oxygen-containing groups led to a decrease of aggregation rate of forming CaP clusters. The presence of longer oligomer, LAC_8 , had little effect on the CaP clustering because of the relatively weak binding ability between Ca^{2+} ions and LAC_8 . The predicted inhibition of nucleation process of CaP was supported by our experiments which indicated that the UV-Vis absorbance values of CaP samples decreased with addition of LAC at the physiological pH value of 7.4. The charge redistribution at the outmost surface of CaP cluster was also observed in the presence of LAC, since the exposed groups at outsurface are negatively charged phosphate species instead of Ca^{2+} ions at the original CaP surface. The prepared CaP samples with different Ca/P ratios (1.50, 1.67 and 2.00) showed negative zeta potential values at $\text{pH} \approx 7.4$. The relationship between the zeta potentials of various nano clusters and their biological functions (such as osseointegration, apatite nucleation, and inhibition of tumor cell proliferation) was discussed. The final precipitation was found to be DCPD by XRD. Understand of the influence of interfacial interactions of LAC and other factors such as pH values and Ca/P ratios on the early stage of nucleation process of CaP is useful to guide the rational design of potential antitumor or tissue regeneration materials.

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Author Contributions

J.M. and H.C. designed all the computations. C.L., L.G., and X.G conceived the experiments and performed characterizations (XRD, TEM, UV-Vis and Zeta potential). M.M., Z.L., J.L., H.C., and X.Z. guided the high-throughput experimentations. J.H., H.D., Q.Z., Y.G., and Z.L provided the data analysis scripts. All authors contributed to discussing the results as well as the writing and revising of the manuscript.

Notes

The authors declare no competing financial interest.

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