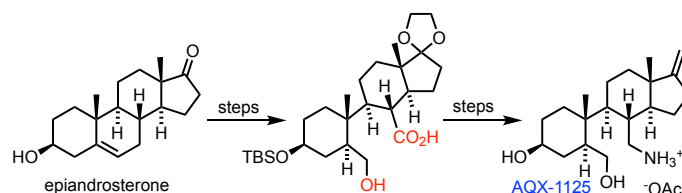


Synthetic Studies on the Indane SHIP1 Agonist AQX-1125.

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Supporting Information Placeholder



ABSTRACT: A new synthetic route to the SHIP1 agonist AQX-1125 has been developed. This sequence utilizes a hydroxy-acid intermediate which allows for ready differentiation of the C6 and C7 positions. The role of the C17 alkene in the biological activity of the system is also investigated, and this functional group is not required for SHIP1 agonist activity.

The PI3K signaling axis is a major cell signaling pathway, trafficking information about the extracellular environment from outside the plasma membrane through the cytoplasm to the nucleus.¹ This signaling influences cell metabolism, effector functions, proliferation, and survival.² Membrane receptors mediate these effects, initiating signaling cascades through a network of enzymes and second messengers inside the cell, with phosphoinositides acting as participants.³ The pattern of phosphorylation present on the phosphoinositide ring are detected by PH and C2-like domains present in protein kinases and other signaling adapters leading to the transmission of information to the nucleus. Phosphatidylinositol-3,4,5-trisphosphate (PI(3,4,5)P₃) (Figure 1) is regarded as a key secondary messenger in the PI3K pathway. Normally PI(3,4,5)P₃ is maintained at a low concentration, but activation of PI3K leads to the local synthesis of PI(3,4,5)P₃ from PI(4,5)P₂, rapidly increasing the intracellular concentration of this phosphoinositide and activation of downstream signaling elements (e.g., Akt, mTOR).⁴ Inositol phosphorylation is therefore closely regulated by inositol kinases and phosphatases.⁵ Modulation of inositol kinases and phosphatases has become an active research area, as aberrant activation or loss of function in these enzymes is implicated in many disorders.⁶ Inhibition of PI3K has been shown to have widespread influence on cellular physiology,¹ and several PI3K inhibitors are now being utilized as cancer treatments.⁷ Resistance has already been documented,^{6a,8} however, leading to the investigation of alternative approaches to control signaling including the modulation of inositol phosphatases. The primary inositol phosphatases involved in processing PI(3,4,5)P₃ are PTEN (phosphatase and tensin homolog protein) and SHIP (src homology 2 (SH2) – containing inositol 5'-phosphatase).⁹ Genetic studies have indicated that modulation of inositol phosphatase activity

may influence the development and progression of disorders involving inflammation and cell division.¹⁰ Although PTEN and SHIP both negatively regulate the PI3K pathway, they do so in different ways, with PTEN converting PI(3,4,5)P₃ to PI(4,5)P₂ while SHIP converts PI(3,4,5)P₃ to PI(3,4)P₂.^{9a} By decreasing the cellular concentration of PI(3,4,5)P₃, activation of SHIP provides an alternative means to influence signaling.¹¹

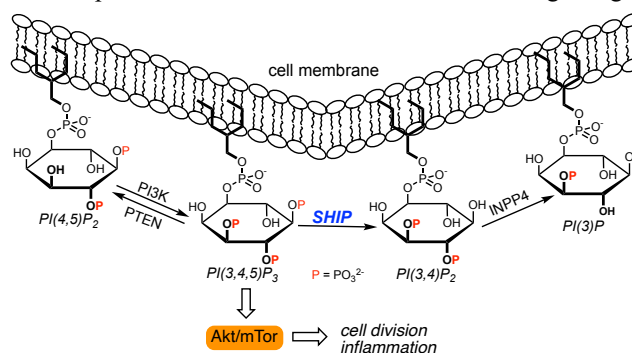


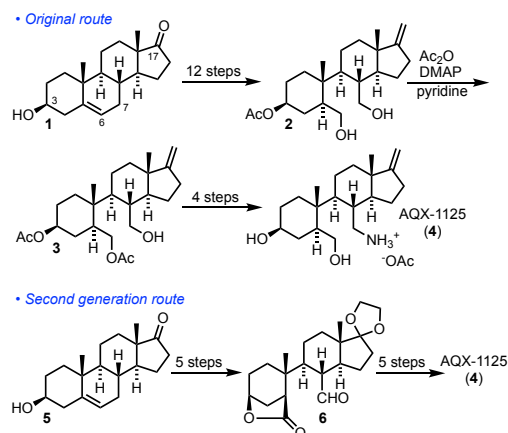
Figure 1. Modification of phosphatidylinositol second messengers by kinases and phosphatases in the PI3K pathway.

Both SHIP1 agonists (as antitumor¹² and anti-inflammatory agents¹³) and SHIP1 inhibitors (as antitumor agents,¹⁴ immunotherapeutics,¹⁵ and Alzheimer's therapeutics¹⁶) have been investigated. The best known SHIP1 agonist is the indane AQX-1125 which was evaluated in clinical trials even though the molecule showed only moderate activation of the SHIP1 enzyme (~20% increase in SHIP1 activity at 300 μ M in the Malachite Green assay¹³). AQX-1125 was evaluated in the clinic for the treatment of bladder pain syndrome/interstitial cystitis (BPS/IC),¹⁷ but development was halted due to lack of

efficacy. Given our recent results indicating that SHP1 deficiency in the intestine is associated with IBD and Crohn's disease,¹⁸ we began a program to access known SHP1 agonists to evaluate their effects in models of IBD, and therefore undertook synthetic studies on AQX-1125.

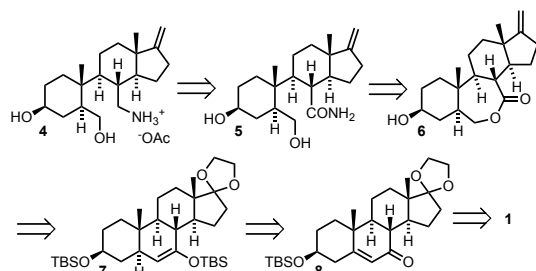
The initial synthesis¹⁹ of AQX-1125 begins with dehydroepiandrosterone **1** (Scheme 1). The route utilizes numerous functional group interconversions until intermediate **2** is reached. Diol **2** is then acylated selectively at the C6 hydroxyl (steroid numbering), as this alcohol is evidently less hindered. The acetate **3** is then taken on to the desired AQX-1125 **4**, with the entire route requiring 17 synthetic steps. After initiation of this work,²⁰ a number of different synthetic routes to AQX-1125 were disclosed in the patent literature.²¹ The route presented with the most detail utilizes the lactone **6** as an intermediate, which differentiates C6 and C7 while also protecting the C3 alcohol.

Scheme 1. Reported Syntheses of AQX-1125



In order to evaluate AQX-1125 in our assays we developed a new synthetic route to the molecule. This new route utilized a different lactone intermediate to differentiate C6 and C7 positions. Beginning with dehydroepiandrosterone **1**, the synthesis would proceed through ketone **8**, which would be converted into the silyl enol ether **7** (Scheme 2). Oxidative cleavage of the silyl enol ether followed by cyclization would provide lactone **6**, which can provide access to amide **5** and eventually AQX-1125 **4**. This new route avoids the diol intermediate **2** and provides access to new analogs that can be evaluated as SHP1 agonists.

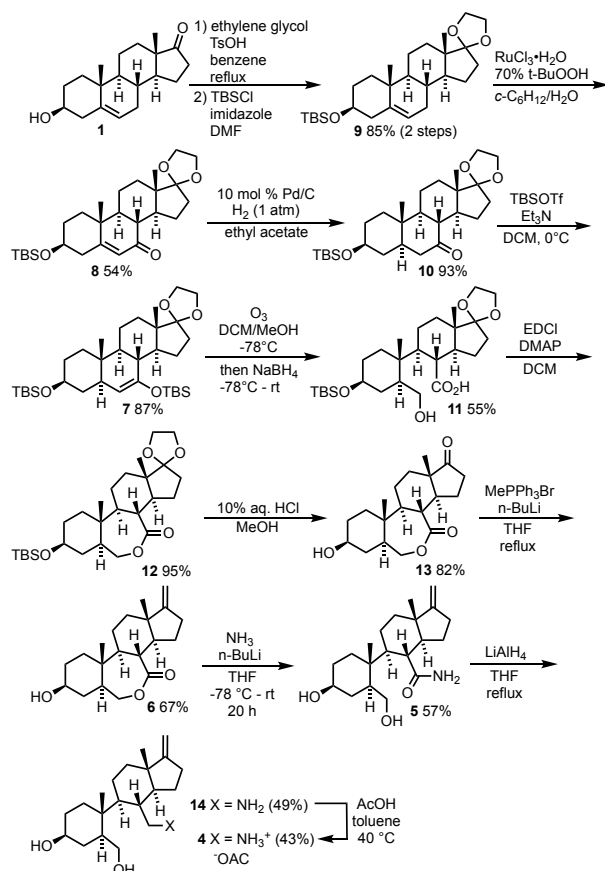
Scheme 2. Retrosynthetic Analysis of AQX-1125



Following this plan, the C17 ketone of epiandrosterone **1** was protected as the ketal followed by formation of the silyl ether **4** (Scheme 3). Ruthenium-catalyzed allylic oxidation²² of the C7 position then provided ketone **5**. Hydrogenation of the C5-6 alkene then gave ketone **10**. The stereochemistry of this transformation is well precedented,²³ leading to the addition of an α -hydrogen at C5. This outcome is rationalized by the cata-

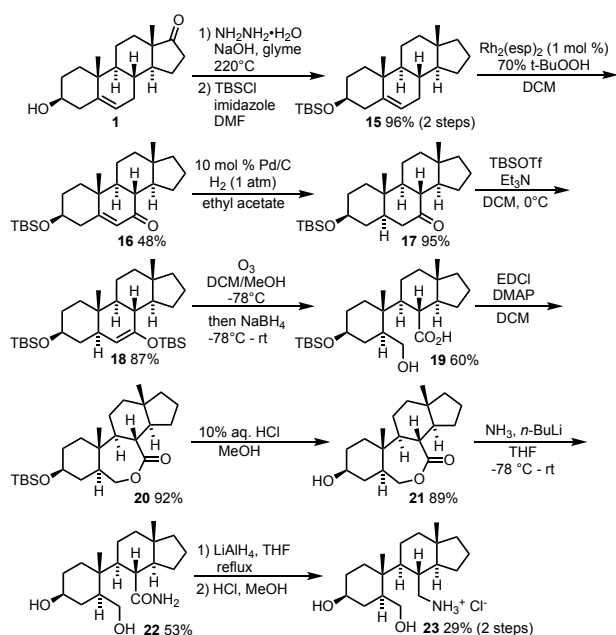
lyst avoiding the axial C10 methyl group. Formation of the silyl enol ether was then accomplished following a procedure from Deslongchamps²⁴ in 95% yield. Oxidative cleavage of the resulting silyl enol ether was then investigated, with ozonolysis being the most successful (two step protocols involving Rubottom oxidation and periodate cleavage gave lower yields). Direct reduction of the ozonide with sodium borohydride provided the hydroxy-acid **11** in 55% yield. The carboxylic acid **11** was then cyclized to the seven-membered lactone with EDCI. Simultaneous deprotection of the ketal and silyl ether was accomplished utilizing aqueous HCl in methanol leading to ketone **13**. A Wittig reaction then installed the C17 methylene providing alkene **6**. Opening of the lactone with lithium amide (formed *in situ* from anhydrous ammonia and *n*-butyllithium) provided the desired amide **5**, which could be reduced with LiAlH₄ to access amine **14**. Formation of the acetate salt then provided AQX-1125 (**4**).

Scheme 3. New Synthetic Route to AQX-1125



In addition to providing material for our biological assays, the synthetic studies on AQX-1125 provided an opportunity to study some structure activity relationships. In particular, the role the C17 alkene played in the SHP1 agonist activity was of interest, as this functional group was hypothesized to be uninvolved in interactions with the enzyme and lengthened the synthetic route. Should the C17 alkene not be required, the analog could also be utilized in our studies, providing a more expedient route to SHP1 agonists. To explore this possibility, a synthesis of the analog **23** was undertaken (Scheme 4). The C17 ketone of epiandrosterone **1** was reduced to the alkane utilizing Wolff-Kishner conditions,²⁵ and the C3 alcohol protected as a TBS ether. Oxidation of the C7 position utilizing Ru-catalyzed conditions resulted in an unexpectedly low 25% yield of enone **16** along with numerous side products. This

Scheme 4. Analog Synthesis



Both SHIP1 agonists and antagonists can induce cell death by induction of cell-intrinsic²⁸ or -extrinsic apoptosis,^{18b} but the lack of activity of AQX-1125 is difficult to rationalize. Recently Mui and co-workers demonstrated that AQX-1125 is ineffective in treating inflammation in IL-10 knockout mice, while more potent SHIP1 agonists are effective in this model.³¹ This report also showed that AQX-1125 only binds to SHIP1 weakly, with SHIP1 perhaps not being the primary cellular target of the molecule. These results may explain the lack of activity of AQX-1125 on OPM2 cells.

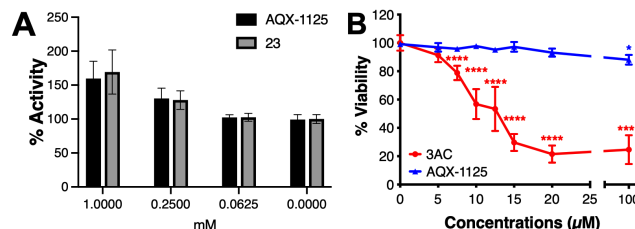


Figure 2. Bioactivity of AQX-1125 (4) and 23. (A) SHP1 agonist activity of AQX-1125 (4) and 23 in the Malachite Green Phosphatase Release Assay. (B) Effects of AQX-1125 and 3 α -amincholestanol on cell viability using OPM2 multiple myeloma cells as determined by Dojindo CCK-8 Cell Viability Assay.

In summary we have developed a new synthetic route to the AQX-1125 and synthesized a simplified analog that showed similar bioactivity. Preliminary evaluation of AQX-1125 showed that while the molecule appears to be a SHIP1 agonist in *in vitro* assays, it does not display cytotoxic effects against OPM2 cells like other SHIP1 agonists. This is similar to the lack of activity in anti-inflammatory assays that was recently reported by Mui.³¹ The poor results in cell based assays indicate a need for more potent SHIP1 agonists with better performance in cell based assays and *in vivo*. We have therefore turned our attention to alternative SHIP1 agonist scaffolds for evaluation in cancer, IBD, and colitis models, as the AQX-1125 system appears to have little utility in these areas.

ASSOCIATED CONTENT

Supporting Information

Experimental procedures, new compound characterization data, and NMR spectra. (PDF)

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Notes

S.D., S. F-D., W.G.K. and J.D.C. have patents on small molecules targeting of SHIP1 and SHIP2 in disease. The other authors have no conflicts to disclose.

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REFERENCES

- (1) Cantley, L. C. The phosphoinositide-3-kinase pathway. *Science* **2002**, *296*, 1655.

- (2) (a) Vanhaesebroeck, B.; Stephens, L.; Hawkins, P. PI3K signaling: The path to discovery and understanding. *Nat. Rev. Mol. Cell Biol.* **2012**, *13*, 195. (b) Wymann, M. P.; Schultz, C. The chemical biology of phosphoinositide 3-kinases. *ChemBioChem* **2012**, *13*, 2022.
- (3) Catimel, B.; Yin, M.-X.; Schieber, C.; Condron, M.; Patsiouras, H.; Catimel, J.; Robinson, D. E. J. E.; Wong, L. S.-M.; Nice, E. C.; Holmes, A. B.; Burgess, A. W. PI(3,4,5)P₃ Interactome. *J. Proteome Res.* **2009**, *8*, 3712.
- (4) Marion, F.; Williams, D. E.; Patrick, B. O.; Hollander, I.; Mallon, R.; Kim, S. C.; Roll, D. M.; Feldberg, L.; Van Soest, R.; Andersen, R. J. Liphagal, a Selective Inhibitor of PI3 Kinase Isolated from the Sponge Aka coralliphaga: Structure Elucidation and Biomimetic Synthesis. *Org. Lett.* **2006**, *8*, 321.
- (5) Balla, T. Phosphoinositides: tiny lipids with giant impact on cell regulation. *Physiol. Rev.* **2013**, *93*, 1019.
- (6) (a) Fruman, D. A.; Chiu, H.; Hopkins, B. D.; Bagrodia, S.; Cantley, L. C.; Abraham, R. T. The PI3K pathway in Human Disease. *Cell* **2017**, *170*, 605. (b) Xu, F.; Na, L.; Li, Y.; Chen, L. Roles of the PI3K/AKT/mTOR signalling pathways in neurodegenerative diseases and tumours. *Cell Biosci.* **2020**, *10*, 54.
- (7) (a) Garces, A. E.; Stocks, M. J. Class 1 PI3K Clinical Candidates and Recent Inhibitor Design Strategies: A Medicinal Chemistry Perspective. *J. Med. Chem.* **2019**, *62*, 4815. (b) Elmenier, F. M.; Lasheen, D. S.; Abouzid, K. A. M. Phosphatidylinositol 3 kinase (PI3K) inhibitors as new weapon to combat cancer. *Eur. J. Med. Chem.* **2019**, *183*, 111718.
- (8) (a) Park, S.; Kim, Y. S.; Kim, D. Y.; So, I.; Jeon, J.-H. PI3K pathway in prostate cancer: All resistant roads lead to PI3K. *Biochim. Biophys. Acta, Rev. Cancer* **2018**, *1870*, 198. (b) Brown, K. K.; Toker, A. The phosphoinositide 3-kinase pathway and therapy resistance in cancer. *F1000Prime Rep.* **2015**, *7*, 1. (c) Yap, T. A.; Bjerke, L.; Clarke, P. A.; Workman, P. Drugging PI3K in cancer: refining targets and therapeutic strategies. *Curr. Opin. Pharmacol.* **2015**, *23*, 98.
- (9) (a) Leslie, N. R.; Biondi, R. M.; Alessi, D. R. Phosphoinositide-regulated kinases and phosphoinositide phosphatases. *Chem. Rev.* **2001**, *101*, 2365. (b) Krystal, G. Lipid phosphatases in the immune system. *Semin. Immunol.* **2000**, *12*, 397.
- (10) (a) Eramo, M. J.; Mitchell, C. A. Regulation of PtdIns(3,4,5)P₃/Akt signaling by inositol polyphosphate 5-phosphatases. *Biochem. Soc. Trans.* **2016**, *44*, 240. (b) Billcliff, P. G.; Lowe, M. Inositol lipid phosphatases in membrane trafficking and human disease. *Biochem. J.* **2014**, *461*, 159.
- (11) (a) Viernes, D. R.; Choi, L. B.; Kerr, W. G.; Chisholm, J. D. Discovery and Development of Small Molecule SHIP Phosphatase Modulators. *Med. Res. Rev.* **2014**, *34*, 795. (b) McLoughlin, N. M.; Mueller, C.; Grossmann, T. N. The Therapeutic Potential of PTEN Modulation: Targeting Strategies from Gene to Protein. *Cell Chem. Biol.* **2018**, *25*, 19.
- (12) Kennah, M.; Yau, T. Y.; Nodwell, M.; Krystal, G.; Anderson, R. J.; Ong, C. J.; Mui, A. L. Activation of SHIP via a small molecule agonist kills multiple myeloma cells. *Exp. Hematol.* **2009**, *37*, 1274.
- (13) (a) Stenton, G. R.; MacKenzie, L. F.; Tam, P.; Cross, J. L.; Harwig, C.; Raymond, J.; Toews, J.; Wu, J.; Ogden, N.; MacRury, T.; Szabo, C. Characterization of AQX-1125, a small-molecule SHIP1 activator. Part 1. Effects on inflammatory cell activation and chemotaxis in vitro and pharmacokinetic characterization in vivo. *Br. J. Pharmacol.* **2013**, *168*, 1506. (b) Stenton, G. R.; Mackenzie, L. F.; Tam, P.; Cross, J. L.; Harwig, C.; Raymond, J.; Toews, J.; Chernoff, D.; MacRury, T.; Szabo, C. Characterization of AQX-1125, a small-molecule SHIP1 activator. Part 2. Efficacy studies in allergic and pulmonary inflammation models in vivo. *Br. J. Pharmacol.* **2013**, *168*, 1519.
- (14) (a) Fuhler, G. M.; Brooks, R.; Toms, B.; Iyer, S.; Gengo, E. A.; Park, M.-Y.; Gumbleton, M.; Viernes, D. R.; Chisholm, J. D.; Kerr, W. G. Therapeutic potential of SH2 domain-containing inositol-5'-phosphatase 1 (SHIP1) and SHIP2 inhibition in cancer. *Mol. Med.* **2012**, *18*, 65. (b) Chen, Z.; Shojaaee, S.; Buchner, M.; Geng, H.; Lee, J. W.; Klemm, L.; Titz, B.; Graeber, T. G.; Park, E.; Tan, Y. X.; Satterthwaite, A.; Paietta, E.; Hunger, S. P.; Willman, C. L.; Melnick, A.; Loh, M. L.; Jung, J. U.; Coligan, J. E.; Bolland, S.; Mak, T. W.; Limnander, A.; Jumaa, H.; Reth, M.; Weiss, A.; Lowell, C. A.; Muschen, M. Signalling thresholds and negative B-cell selection in acute lymphoblastic leukaemia. *Nature* **2015**, *521*, 357.
- (15) Gumbleton, M.; Sudan, R.; Fernandes, S.; Engelman, R. W.; Russo, C. M.; Chisholm, J. D.; Kerr, W. G. Dual enhancement of T and NK cell function by pulsatile inhibition of SHIP1 improves anti-tumor immunity and survival. *Sci Signal* **2017**, *10*, eaam5353.
- (16) Pedicone, C.; Fernandes, S.; Dungan, O. M.; Dormann, S. M.; Viernes, D. R.; Adhikari, A. A.; Choi, L. B.; De Jong, E. P.; Chisholm, J. D.; Kerr, W. G. Pan-SHIP1/2 inhibitors promote microglia effector functions essential for CNS homeostasis. *J. Cell Sci.* **2020**, *133*, jcs238030.
- (17) Nickel, J. C.; Egerdie, B.; Davis, E.; Evans, R.; Mackenzie, L.; Shrewsbury, S. B. A Phase II Study of the Efficacy and Safety of the Novel Oral SHIP1 Activator AQX-1125 in Subjects with Moderate to Severe Interstitial Cystitis/Bladder Pain Syndrome. *J. Urol.* **2016**, *196*, 747.
- (18) (a) Fernandes, S.; Srivastava, N.; Sudan, R.; Middleton, F. A.; Shergill, A. K.; Ryan, J. C.; Kerr, W. G. SHIP1 deficiency in inflammatory bowel disease is associated with severe Crohn's disease and peripheral T cell reduction. *Front. Immunol.* **2018**, *9*, 1100/1. (b) Park, M. Y.; Srivastava, N.; Sudan, R.; Viernes, D. R.; Chisholm, J. D.; Engelman, R. W.; Kerr, W. G. Impaired T-cell survival promotes mucosal inflammatory disease in SHIP1-deficient mice. *Mucosal Immunol.* **2014**, *7*, 1429.
- (19) Raymond, J. R.; Han, K.; Zhou, Y.; He, Y.; Noren, B.; Yee, J. G. K. Indene derivatives as pharmaceutical agents. US20140371252A1, 2014.
- (20) Duffy, B. C. Thioetherification and Etherification Utilizing Trichloroacetimidates Under Thermal Conditions & Progress Towards an Efficient Synthesis of AQX-1125, PhD Thesis, Dissertation Abstracts:10157481. Syracuse University, Syracuse, NY, 2016.
- (21) Harwig, C.; Seenisamy, J.; Keregadde, M. N.; Chetia, L. Synthesis of a substituted indene derivative. US10053415B2, 2018.
- (22) Miller, R. A.; Li, W.; Humphrey, G. R. A ruthenium-catalyzed oxidation of steroidal alkenes to enones. *Tetrahedron Lett.* **1996**, *37*, 3429.
- (23) (a) Krishnan, K.; Manion, B. D.; Taylor, A.; Bracamontes, J.; Steinbach, J. H.; Reichert, D. E.; Evers, A. S.; Zorumski, C. F.; Mennerick, S.; Covey, D. F. Neurosteroid Analogues. 17. Inverted Binding Orientations of Androsterone Enantiomers at the Steroid Potentiation Site on γ -Aminobutyric Acid Type A Receptors. *J. Med. Chem.* **2012**, *55*, 1334. (b) Zhao, C.; Ye, Z.; Ma, Z.-x.; Wildman, S. A.; Blaszczyk, S. A.; Hu, L.; Guizei, I. A.; Tang, W. A general strategy for diversifying complex natural products to polycyclic scaffolds with medium-sized rings. *Nat. Commun.* **2019**, *10*, 1.
- (24) Reddy, M. S.; Zhang, H.; Phoenix, S.; Deslongchamps, P. Total synthesis of ouabagenin and ouabain. *Chem. - Asian J.* **2009**, *4*, 725.
- (25) Bazin, M.-A.; Travert, C.; Carreau, S.; Rault, S.; El Kihel, L. First synthesis of 7 α - and 7 β -amino-DHEA, dehydroepiandrosterone (DHEA) analogues and preliminary evaluation of their cytotoxicity on Leydig cells and TM4 Sertoli cells. *Bioorg. Med. Chem.* **2007**, *15*, 3152.
- (26) Wang, Y.; Kuang, Y.; Zhang, H.; Ma, R.; Wang, Y. Recyclable Dirhodium(II) Catalyst Rh2(esp)2 for the Allylic Oxidation of Δ^5 -Steroids. *J. Org. Chem.* **2017**, *82*, 4729.
- (27) Cogan, E. B.; Birrell, G. B.; Griffith, O. H. A Robotics-Based Automated Assay for Inorganic and Organic Phosphates. *Anal. Biochem.* **1999**, *271*, 29.
- (28) Brooks, R.; Fuhler, G. M.; Iyer, S.; Smith, M. J.; Park, M.-Y.; Paraiso, K. H. T.; Engelman, R. W.; Kerr, W. G. SHIP1 Inhibition Increases Immunoregulatory Capacity and Triggers Apoptosis of Hematopoietic Cancer Cells. *J. Immunol.* **2010**, *184*, 3582.
- (29) Ong, C. J.; Ming-Lum, A.; Nodwell, M.; Ghanipour, A.; Yang, L.; Williams, D. E.; Kim, J.; Demirjian, L.; Qasimi, P.; Ruschmann, J.; Cao, L. P.; Ma, K.; Chung, S. W.; Duronio, V.; Anderson, R. J.; Krystal, G.; Mui, A. L. Small-molecule agonists of SHIP1 inhibit the phosphoinositide 3-kinase pathway in hematopoietic cells. *Blood* **2007**, *110*, 1942.
- (30) Kerr, W. G. Inhibitor and Activator: Dual Functions for SHIP in Immunity and Cancer. *Ann. N.Y. Acad. Sci.* **2011**, *1217*, 1.

(31) Chamberlain, T. C.; Cheung, S. T.; Yoon, J. S. J.; Ming-Lum, A.; Gardill, B. R.; Shakibakho, S.; Dzananovic, E.; Ban, F.; Samiea, A.; Jawanda, K.; Priatel, J.; Krystal, G.; Ong, C. J.; Cherkasov, A.; Andersen, R. J.; McKenna, S. A.; Van Petegem, F.; Mui, A. L. F.

Interleukin-10 and Small Molecule SHIP1 Allosteric Regulators Trigger Anti-inflammatory Effects through SHIP1/STAT3 Complexes. *iScience* **2020**, 23, 101433.
