Facile surface modification with hexamethyldisilazane to generate atomically flat and hydrophobic substrates for filament gliding assays with molecular motors

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In vitro gliding assay is a well-established assay for determining the activity of protein motors, such as actin-associated myosins and microtubule-associated kinesins and dyneins. In one of the conventional methods, protein motors are immobilized onto a nitrocellulose-coated coverslip and it propels actin filaments in the presence of ATP. Gliding assays also serve as the foundation for protein-motor-based nanotechnological devices such as biosensing and sorting. However, the preparation of nitrocellulose-coated coverslips produces rough surfaces and in some protocols, requires up to an overnight incubation for the nitrocellulose film to completely dry. Furthermore, the nitrocellulose matrix exhibits high background autofluorescence, which reduces the signal-to-noise ratio of fluorescence images. Here, we investigated the use of hexamethyldisilazane (HMDS) to study actomyosin function and characterized its physical properties on glass coverslips and glass capillary tubes. We showed that the total preparation time to coat a coverslip with HMDS is <30 minutes and generates an atomically-flat surface with $\ll 0.1\times$ of autofluorescence than HMDS. In addition, HMDS does not interfere with myosin function, which is indicated by the similar actin gliding speed when compared with nitrocellulose. Our results show that HMDS vapor deposition is a more favorable surface treatment to nitrocellulose for *in vitro* gliding assay and potentially for a broad class of biophysics experiments that require protein immobilization on hydrophobic surfaces.

Introduction

Molecular motors are involved in many cellular functions including mediating active cargo transport, muscle contraction, and cell division.¹ Motor proteins such as myosins form a group and they walk on actin filaments to perform those processes. *In vitro* gliding assays are often used^{2–7} to elucidate the underlying mechanism and principles of the ATP-driven actin-myosin interaction. In this assay, isolated myosin or its fragments are adsorbed onto a treated glass coverslip where they propel the gliding of fluorescent-labeled actin filaments in the presence of ATP.⁸ In addition to providing considerable insight into the fundamentals of actomyosin biophysics, *in vitro* gliding assays have been proposed to serve as an active transport system in artificial environments for biomedical devices and molecular cybernetics.^{9–13}

In conventional *in vitro* gliding assays, glass coverslips and slides are functionalized with nitrocellulose to bind myosin motors for *in vitro* gliding assay experiments. Nitrocellulose (**Fig. 1A**) is a polymer that is derived by nitrating cellulose via chemical a reaction involving a mixture of nitric acid and sulfuric acid. However, the preparation of coverslips with nitrocellulose is requires an overnight drying step and produces coated glass surfaces with a high background autofluorescence. Furthermore, gliding assays with nitrocellulose tend to give a lower fraction of motile filaments.^{14–16}

Fig. 1 Nitrocellulose and HMDS surface coatings for gliding assays. Chemical structures of nitrocellulose (A) and HMDS (B). Workflow schematics of coverslip modification with nitrocellulose (C) and HMDS (D). (E) Schematic of gliding assay with HMDS-coated coverslip. For schematic clarity, the relative orientation of myosin VI motors on HMDS was oversimplified. Mode of myosin VI absorption is unknown, but is expected to be stochastic.

Using silanes to coat coverslips has been shown to increase the gliding speed and gliding duration of actin filaments.^{6,14,16,17} Among these studies, it is shown that trimethylchlorosilane is superior in improving the fidelity of gliding assay and gives the most reproducible result compared to other silanes such as octadecylchlorosilane, dimethyldichlorosilane¹⁴, triethylchlorosilane, tripropylchlorosilane, tributylchlorosilane, phenyldimethylchlorosilane, and diphenylmethylchlorosilane¹⁶. The silanization process using these chemicals involves a lot of steps, including but not limited to piranha cleaning, then dipping the coverslips into silane/chloroform mixtures.

Hexamethyldisilazane (HMDS; Fig. 1B) is one of the silane reagents that derivatizes surfaces through the formation of trimethylsilyl groups.¹⁸ In contrast to nitrocellulose that forms a disordered matrix¹⁹ (Fig. 1C), HMDS chemically bonds as a monolayer on the surfaces (Fig. 1D). HMDS reacts with oxidized surfaces, releasing ammonia, and forming a monolayer that consists of non-polar trimethylsilyl groups away from the substrate. This reaction forms a hydrophobic surface, which is a preferable condition to adsorb proteins with hydrophobic patches. Interestingly, HMDS vapor deposition has been used to prevent highly-hydrophilic DNA origami nanostructures from binding to glass surfaces.^{20,21} However, the use of HMDS for *in vitro* gliding assays of protein motors has not been explored.

Derivatization of surfaces with HMDS can be easily done through chemical vapor deposition of a negatively-charged surface for less than 30 min, which is significantly faster than nitrocellulose coating that requires up to an overnight drying before it is ready for functional assays (Fig. 1E and F). Silanization with HMDS does not require glass coverslip to undergo an extensive cleaning procedure with piranha solution

and plasma cleaning is sufficient for this modification.²⁰ In addition, the versatility of vapor deposition has the potential to modify less-exposed surfaces such as narrow capillary tubes that might be challenging to modify with viscous nitrocellulose.

Here, we evaluate the use of HMDS as a substrate to immobilize myosin by characterizing the physical properties such as hydrophobicity, surface height distribution, and autofluorescence of the modified coverslips. Then, we have compared the motility of actin on glass surfaces treated with HMDS and nitrocellulose. We tested our method to modify capillary tubes for gliding assays. We used myosin VI dimer constructs as a model system for motor proteins. We show that HMDS vapor deposition can modify exposed and less-exposed glass surfaces to allow immobilization of motor proteins. The immobilized myosins were functional for *in vitro* gliding assays.

Results

HMDS vapor deposition forms a uniform layer on glass coverslips. We deposited monolayer HMDS on Corning D263 glass coverslips using the vapor deposition method at room temperature and atmospheric pressure (Materials and Methods; **Fig. 1**). We also deposited nitrocellulose on the same model of coverslips. The overall process of surface modification with HMDS takes less than 30 min, while the nitrocellulose deposition process takes 16 hours until the coverslip is ready to use (**Fig. 1E**). Representative AFM images of both surface coating techniques are presented in **Fig. 2A** and B with the same color scale to allow a direct comparison of the samples. We increased the range of the color scale to reduce the appearance of noise for a better visual analysis. AFM images with real-time scaling mode show a periodic noise (**Fig. S1** and **S2**). HMDS-coated surfaces are cleaner than nitrocellulose-coated surfaces as indicated by the presence of blobs (bright spots) on nitrocellulose-coated surfaces (**Fig. 2B**). There is a clear difference of the surface height distribution between HMDS-coated surface (**Fig. 2C**) and nitrocellulose-coated surfaces (**Fig. 2D**). There is an asymmetry observed in the histogram of nitrocellulose-coated surface heights and they are more variable compared to the heights of HMDS-coated surfaces.

TIRF assays of partially modified glass coverslips with nitrocellulose show that the mean height of the deposited nitrocellulose is 456 ± 125 nm thick, which indicates that it has multilayer thickness. In contrast, AFM characterization of partially modified glass coverslips (data not shown) shows that the deposited HMDS layer is 1 monolayer thick (<1 nm), which is three orders of magnitude thinner than nitrocellulose films (**Fig. 2E**). The monolayer formation of HMDS is consistent with previous reports that HMDS interacts with activated SiO_2 and not with trimethylsilyl groups.

HMDS modifies the hydrophilic plasma-cleaned surface to a hydrophobic surface. Plasma-cleaning (PC) reduces the contact angle of untreated (non-plasma-cleaned; NPC) glass surfaces from $40.4^\circ \pm 1.9^\circ$ ($N = 3$) into completely hydrophilic (0° contact angle; $N = 3$; **Fig. 3F and G**). After vapor deposition of HMDS, PC glass surface becomes hydrophobic which is indicated by the increase of contact angle to $64.1^\circ \pm 3.2^\circ$ ($N = 3$). Surprisingly, HMDS vapor deposition of inactive NPC glass also renders the surface hydrophobic ($70.7^\circ \pm 1.8^\circ$ ($N = 3$; **Fig. 2H**), which is significantly higher ($p = 0.037$) than HMDS-coated PC glass (**Fig. 2I**). The similar contact angle of nitrocellulose-coated NPC glass ($59.5^\circ \pm 0.1^\circ$) and PC glass ($61.1^\circ \pm 2.0^\circ$) shows that plasma cleaning of glass does not affect the hydrophobicity of nitrocellulose coating ($p = 0.236$; **Fig. 2J-L**). The contact angles of HMDS-coated PC surfaces are on par with the contact angle of both nitrocellulose-coated NPC and PC surfaces.

HMDS have lower autofluorescence than nitrocellulose. We quantified the autofluorescence intensity and photo-bleaching of nitrocellulose- and HMDS-coated coverslips in a 2-min period (**Movies S1–S3**). Nitrocellulose-coated surfaces exhibit higher background intensity than HMDS-coated surfaces and bare glass (**Fig. 2M**). In addition to that, HMDS-coated surfaces have similar intensities as the standard bare glass. The intensity of nitrocellulose-coated surfaces is reduced over time. However, even after 2 min, the intensity of nitrocellulose background is still higher than the glass surface.

Fig. 2 Physical characterization of HMDS-coated and nitrocellulose-coated glass coverslips. (A–B) Representative AFM images of HMDS-coated (A) and nitrocellulose-coated (B) surfaces. Scale bar represents 5 μm . (C–D) Height distribution of HMDS-coated (C) and nitrocellulose-coated (D) surfaces. (E) Film thickness of treated surfaces (N = 3). (F–K) Representative images of water droplets on non-plasma-cleaned (NPC; F), plasma-cleaned (NPC; G), NPC glass treated with HMDS (H), PC glass treated with HMDS (I), NPC glass coated with nitrocellulose (J), and PC glass coated with nitrocellulose (K). (L) Contact angle measurements of different treatments of glass coverslip (N = 3 samples). Scale bar represents 1 mm. (M) Mean fluorescence/pixel intensity of glass, HMDS-coated, and nitrocellulose-coated surfaces (N = 3 field of views). Error bars are $\pm$ SD.

Myosin-actin gliding assay is compatible with HMDS. Fluorescent time-lapse images of gliding assays in the absence of ATP revealed that myosin VI motors bind to HMDS (**Fig. 3A**). This is indicated by the binding of actin filaments to HMDS surfaces deposited with myosin VI motors. However, at 0 mM ATP actin filaments do not glide along the myosin surface (0 $\mu\text{m/s}$ gliding speed). As seen in **Fig. 3B**, actin filaments are not adsorbed onto HMDS-coated surfaces in the absence of myosin motors. This result is similar to when common passivating agent bovine serum albumin (BSA) was used to coat the glass surfaces and HMDS-coated surfaces (**Fig. 3C and D**). As expected, actin filaments are adsorbed to bare plasma-cleaned glass surfaces (**Fig. 3E**). This result suggests that HMDS is a compatible substrate to use for the *in vitro* gliding assay because it does not interact with actin filaments on its own.

Gliding assay (**Fig. 2F**) shows that at 2 mM ATP, there is gliding of actin filaments which indicates that the myosin are immobilized and functional when immobilized on HMDS- and nitrocellulose-coated surfaces (**Fig. 2G, H, Movies S4 and S5**). The fraction of motile filaments on HMDS-coated surfaces does not have any significant difference (p value = 0.15) with that on nitrocellulose-coated surfaces (**Fig. 3I**). Accordingly, the average gliding speed on HMDS-coated surfaces is similar (63 ± 14 nm/s) to that on nitrocellulose-coated surfaces (64 ± 14 nm/s). In addition, the actin-length-dependent gliding velocities of myosin VI on HMDS- and nitrocellulose-coated surfaces are qualitatively similar (**Fig. 3J**).

HMDS functionalization is compatible with commonly used passivating agents such as BSA and Tween-20 (**Table 1**). Here, the coverslips were plasma-cleaned before any treatment. HMDS-coated surfaces were pretreated with passivating agents before myosin VI deposition. After myosin deposition on the treated surfaces, we measured the actin gliding speed. Pre-treatment with BSA does not outcompete myosin VI binding to HMDS, indicated by the statistically identical gliding speed between experiments using BSA (139 ± 25 nm/s ($N = 50$ filaments); **Movie S6**) and without BSA (63 ± 14 nm/s ($N = 50$ filaments); **Movie S4**). In control experiment without the HMDS coating, gliding speed when only BSA was used to treat the surface is 43 ± 7 nm/s ($N = 20$ filaments; **Movie S7**), which is 32% lower than the actin gliding speed on HMDS-BSA-coated surfaces. Tween-20 inhibits myosin VI binding to HMDS-coated surfaces and plasma cleaned glass as there were no actin binding and gliding observed on both surfaces (**Movies S8 and S9**). Within our experiments, the observed actin gliding speed is the slowest when the glass surface was only plasma-cleaned without further surface treatment (40 ± 12 nm/s; $N = 20$ filaments; **Movie S10**), presumably because of the strong interactions between the unpassivated plasma-cleaned surface and myosin VI.

Sample preparation				Measurement
Step 1	Step 2	Step 3	Step 4	Gliding speed (nm/s)
Plasma-cleaning	HMDS	Passivating agent	myosin VI	
+	+	–	+	63 ± 14
+	+	BSA	+	139 ± 25
+	+	Tween-20	+	Not observed
+	–	BSA	+	43 ± 7
+	–	Tween-20	+	Not observed
+	–	–	+	40 ± 12

Table 1 Gliding speed of indicated experiment using different surface passivating strategies ($N \geq 20$ filaments). The addition of Tween-20 detergent after HMDS vapor deposition block myosin VI from binding to the passivated surfaces.

HMDS vapor deposition modifies the surface hydrophobicity of capillary tubes. Vapor deposition of HMDS can modify geometrically less exposed surfaces, namely, the inner surface capillary tubes. Here, instead of using capillary tubes with circular cross sections, we evaluated the HMDS surface chemistry of the inner surface of capillary tubes with a uniform rectangular cross section. The rectangular cross

F Gliding assay protocol

Fig. 3 Actomyosin function on different coverslip surface modifications. (A) Fluorescence image of rhodamine-labelled F-actin on HMDS-coated myosin VI surface with 0 mM ATP. Fluorescence images of rhodamine-labelled F-actin adsorption on HMDS- (B), BSA- (C), HMDS + BSA-coated (D), and plasma-cleaned glass (E) surfaces without myosin VI at 2 mM ATP. (F) Gliding assay protocol to measure actin gliding speeds and fraction of motile filaments. Fluorescence micrograph time series at 20 s intervals displaying motile and non-motile actin filaments on HMDS-coated (G) and nitrocellulose-coated (H) surfaces (Movies S4–S5). Scale bar represents 5 μm . (I) Fraction of motile filaments (motile filaments divided by total number of filaments on a 20 μm $\times$ 20 μm area, $N = 3$ fields of views) on HMDS-coated and nitrocellulose-coated surfaces. (J) Gliding speed of actin filaments on HMDS surface (blue) and nitrocellulose (orange). Error bars are $\pm\text{SD}$ ($N = 50$ filaments).

section enables microscopy assays for measuring the speed of gliding actin filaments. The inner dimensions of the capillary tubes are 2.0 mm $\times$ 0.20 mm $\times$ 50 mm. Fig. 4A compares HMDS and nitrocellulose

modifications of rectangular capillary tubes. The height of the dyed water was recorded by measuring the distance from the bottom of the capillary tube to the meniscus of the dyed water. The more hydrophilic the interior walls of the capillary tube are, the higher the water rises in the capillary tubes: Plasma cleaning of the untreated capillary surface increased the capillary rise to the other end of the tube (**Fig. 4A**). **Fig. 2** shows that hydrophobicity manifests in a larger contact angle. The plasma cleaned capillary tubes were treated with HMDS or nitrocellulose and both of these treatments made the surfaces hydrophobic (**Fig. 4A and B**). In the context of vertical capillary tubes (**Fig. 4**), Jurin’s Law predicts the height h of the water given the contact angle θ , surface tension γ , acceleration due to gravity g , density ρ , and tube radius r . More importantly, it establishes a relation between height and contact angle that is used to convert the height data to contact angles.

$$h = \frac{2\gamma \cos \theta}{\rho g r} \quad [1]$$

The caveat of our model is that Jurin’s law applies for cylinders, which is different than the rectangular cross section of our capillary tubes. Consequently, the rectangular tubes used in this experiment do not have a radius r that can be found directly. We used 2 physical models which provide upper and lower bound estimates. The first assumption is a cylindrical cross section and equates the areas such that width $\times$ length $= \pi r^2$. Using the first model, the effective r is estimated to be 0.357 mm. The second assumption is where we model the capillary tube as two parallel plates spaced closely together. The capillary rise for two parallel plates is given by a similar formula where r is replaced with the spacing between the two plates ($d = 0.2$ mm). Note that Jurin’s law only works for heights that produce values within the range accepted by the cosine inverse function. In addition, the water height is limited by the height of the capillary tube, so calculations with heights that exceed the max height (*i.e.* all plasma-cleaned tubes) have been excluded from the data (undefined; **Fig. 4B**).

The final contact angles of both glass coverslips and capillary tubes after HMDS treatment are similar ($53.0^\circ - 69.9^\circ$). This indicates that our HMDS vapor deposition is able to modify unexposed glass or unconventional 3D chamber. Nitrocellulose-coated capillary tubes have a slightly higher contact angle ($72.2^\circ - 79.9^\circ$) than HMDS-coated capillary tubes (. However, nitrocellulose coating had to be done with extra care to avoid clogging of the tubes.

HMDS surface modification is compatible with preparing the internal surface of rectangular capillary tubes for filament gliding assays with myosin VI. Fluorescent time-lapse images of gliding assay with HMDS-coated capillary tubes show that actin filaments glide along the surface (**Fig. 4C**, **Movie S11**). The gliding speed of the actin filaments on the capillary tube setup was measured to be 107 ± 35 nm/sec ($N = 29$ filaments), which is within 23% of the gliding speed observed from the HMDS-coated coverslip setup (139 ± 25 nm/sec; **Table 1**(second row)). We observed a lack of actin filaments gliding in unmodified or plasma-cleaned tubes (**Movies S12 and S13**), similar to the low numbers of gliding in filaments in plasma-cleaned only coverslips (Table 1). These results demonstrated that the myosin VI binds to HMDS-treated rectangular capillary tubes as well. We were unable to get gliding assay data on nitrocellulose-coated capillary tubes because the nitrocellulose and gliding assay mix (Go mix; Supporting Information) was not able to flow through the capillary tubes.

Discussion

Here, we demonstrated a simple approach using HMDS vapor deposition to immobilize myosin motors. Our results show that HMDS can provide an atomically flat surface with lower autofluorescence and a faster preparation time that is suitable for *in vitro* gliding assay. Compared to nitrocellulose-coated surfaces, HMDS-coated surfaces have improved physical properties for *in vitro* gliding assays. According to previous studies¹⁴, silanized surfaces are smoother than nitrocellulose-coated surfaces. The height of the nitrocellulose film was significantly thicker and more variable than the HMDS monolayer. One explanation is that, unlike

Fig. 4 Physical properties of modified capillary tubes and actomyosin function on HMDS-treated capillary tubes. (A) Capillary rise recorded for each step of the HMDS and nitrocellulose preparation (N = 3). The image was adjusted to remove perspective distortion for accurate height comparison. (B) Inferred contact angles calculated from the capillary rise data. Results from cylindrical and parallel plate assumptions are compared to values obtained for coverslips. (C) Fluorescence micrograph time series of a gliding actin filament (highlighted in blue) relative to the non-motile actin filaments (Movies S11). Scale bar is 5 μm. Error bars are ±SEM.

nitrocellulose, HMDS cannot form a polymer on the negatively charged coverslip. Furthermore, in line with the idea that HMDS is the superior option for coverslip-coating, HMDS-coated surfaces have a significantly lower autofluorescence, which solves the problem of interference of background autofluorescence that persists in nitrocellulose coatings. HMDS-coated surfaces have similar hydrophobicity to nitrocellulose, which is appropriate for binding functional myosin VI molecules. Our silane coating with HMDS has a lower contact angle than that of TMCS ($76.4 \pm 1.6^\circ$) reported in the earlier study.¹⁴ However, in comparison to the results in that study, our control nitrocellulose-coated surfaces also have a lower contact angle. This lower contact angle can possibly be attributed to the different type of coverslip and the treatment methods we used in our study.

We performed gliding assays on HMDS-coated surfaces to identify actin and myosin VI functions. The measured gliding speeds of actin filaments on myosin VI immobilized on HMDS-coated surfaces are within 98% with the gliding speeds on the standard nitrocellulose-coated surfaces (Fig. 3). These results are 21% slower than the previously reported gliding speed measured using myosin VI-labeled DNA patterned on DNA nanotubes²² and two-dimensional DNA origami scaffolds^{23,24}. We found that the average actin gliding speed increased when HMDS was preincubated with BSA before myosin VI immobilization. This indicated that BSA has a marked effect on gliding speeds, which might be attributed to the lower density or number of myosin VI immobilized on the surface. With a large number of myosins, the actin gliding speeds is effectively independent of the number of proteins²². These results are 57% slower than the speeds of single-myosin VI movement on actin filaments.²⁵ The speed decrease is similar to that previously reported for one to six myosin VI motors on two-dimensional actin networks.²⁶ The selective adsorption of protein to HMDS can be explained by the different hydrophobicity and charge of myosin VI and actin filaments. We do not know the mode of myosin VI adsorption to the silanized surface. However, because the silanized surface is hydrophobic, the hydrophobic part of myosin VI (the tail domain²⁷) is possibly bound to it

through hydrophobic-hydrophobic interactions. Some studies suggest that hydrophobicity is among the underlying mechanisms of myosin molecules adsorption.^{28,29}

In conclusion, HMDS can provide what is needed of nitrocellulose for *in vitro* gliding assay but with faster preparation time, no autofluorescence, and a uniformly nanostructured layer without being restricted by the geometry of the surface. Our tests with capillary tubes indicate that HMDS is a versatile surface modification that can be used for treating glass surfaces with unique geometries. Contact angles and successful gliding attempts confirm that plasma-cleaned glass is compatible with HMDS, even in tight chambers where flow is restricted. Since we use vapor deposition in a vacuum chamber, no flow is required for HMDS modification. The contact angle measurements that match the values obtained with coverslips and the observation of gliding confirms the consistency of this method. However, future research should investigate the mechanism of myosin VI binding to the silanized surface. It would be fruitful to explore the application of HMDS as a substrate for other motor proteins, such as kinesin and dynein.

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Author contributions

I.F.T. and R.F.H. contributed to the experimental design, planning, and execution. I.F.T. and N.K. performed the experiments involving capillary tubes. D.K. collected AFM images. S.D. measured the contact angle of modified glass surfaces. I.F.T., N.K., and R.F.H. wrote the manuscript and prepared figures.

Conflict of interest

The author declares there is no conflict of interest.

Data and material availability. The images, raw data files, and the Mathematica code for performing the data analysis are on a [GitHub](#) repository.

<https://github.com/bionicslab/HMDS-manuscript>

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Materials and Methods

Buffers. 1× assay buffer (AB): 25 mM imidazole (pH 7.5), 4 mM Magnesium Chloride, 1 mM Ethylene glycol-bis(β -aminoethyl ether)-N,N,N',N'-tetraacetic acid tetrasodium salt, 25 mM Potassium Chloride, 10 mM Dithiothreitol. 1× AB.BSA buffer: AB + 1 mg/mL BSA. 1× AB.BSA.CAM: 1× AB.BSA + 9 μ M calmodulin. 1× G-buffer, 4 mM Tris-Cl (pH 8.0), and 0.2 mM Calcium Chloride. 15× HKEM: 150 mM Hepes, 750 mM KCl, 75 mM MgCl₂, 15 mM EGTA.Na. Go mix: 1/25× F-actin, 0.05 mg/mL glucose oxidase, 0.01 mg/mL catalase, 1.5 mg/mL glucose, and 2 mM of ATP in 1× AB.BSA.CAM.

myosin VI expression in Sf9 cells. Recombinant myosin VI was cloned into pBiEX-1 plasmid (Novagen). The construct contains *Homo sapiens* myosin VI, an N-terminal FLAG tag for purification by anti-FLAG resin, 6xHis tag for alternative purification using Ni-NTA resin, and a SNAP tag for optional attachment of benzylguanine (BG)-oligo. This construct was transfected into sf9 insect cells using transient transfection system (Escort IV, Sigma). The protein was expressed for 48–72 hours at 27°C with constant shaking at 150 rpm. The expressed myosin VI is affinity purified using Anti-FLAG resin. The BG-oligo sequence is 5'-AAAAAAAAAAAAAAAAAAAAA-3' with Cy5 modification at the 5'. The optional BG-oligo attachment does not change the gliding speed of actin (data not shown).

Actin Filament (F-actin) Polymerization. Rabbit skeletal muscle actin (Cytoskeleton, Inc;) and rhodamine-labelled actin (Cytoskeleton, Inc; AR05) monomers were mixed at the ratio of 4 to 1 at a final concentration of 30 μ M. Then, 4 μ L of the monomer mix was slowly pipetted into actin polymerization buffer which was prepared by mixing 30 μ L of Premix (3× HKEM, 30 mM DTT, and 8 mM ATP), 6 μ L of 1× HKEM, and 51 μ L 1× G-buffer in a microcentrifuge tube. The tube was incubated on ice for 1 h, then 1 μ L of 10 mM phalloidin was added to the mix. The F-actin was stored at 4°C for up to 2 months.

Coverslip Preparation. Corning D263 glass coverslips (22 mm × 50 mm) were washed with isopropanol and then rinsed with deionized water before drying with nitrogen gas. After washing and drying, the coverslips were plasma-cleaned for 2 min at high setting (Harrick Plasma; PDC-32G) to remove impurities and organic residues. For nitrocellulose coating: coverslips were dipped in 1% nitrocellulose in amyl acetate for 2 sec. Nitrocellulose solution was prepared by diluting 2% nitrocellulose (Electron Microscopy Sciences; 12620-10) with amyl acetate (Electron Microscopy Sciences; 10815). Excess nitrocellulose was blotted from the corner of the coverslip with delicate task wipes before drying overnight in a vertical position. For HMDS coating: immediately after plasma cleaning, the coverslips were placed inside a vacuum desiccator together with a vial containing 300 μ L of HMDS solution at room temperature. Then, the desiccator was connected to a vacuum pump in a fume hood to deposit the HMDS vapor for 20 min.

Coverslip Contact Angle Measurements. The hydrophobicity of treated and nontreated glass surfaces were characterized by contact angle measurements using a Krüss advance goniometer. De-ionized water droplets (1 μ L) were dispensed by an EasyDrop syringe pump system. After the droplet dispensation, an ellipse

fit was used to obtain the contact angle of the water droplet at 20°C. Droplet profiles were captured and their angles were analyzed by the in-built Krüss advance program version 1.11.0.15801. The angle of each replicate was the mean value of the left-most and right-most angle of each droplet at random locations on the coverslip.

Autofluorescence measurements. The epifluorescence images were obtained with an Oxford NanoImaging (ONi) microscope at 10% laser power (0.7–0.8 mW), with 532 nm excitation, and 100 ms exposure. 2-min movies were taken. Autofluorescence of the samples are measured as mean pixel intensity (AU) of 400×400 pixel² ($46.68 \times 46.68 \mu\text{m}^2$) areas in the center of the image using ImageJ tools.³⁰

Gliding Assay and Data Collection. Flow cell was made by sandwiching double-sided tape (Kapton) between the coverslips. A rectangle is cut on the double-sided tape with Graphtech cutting plotter. We estimate the volume of the flow cell to be 10 μL .

Gliding assay was performed by flowing 10 μL of myosin VI into the flow cell followed by 10 min incubation to allow binding of myosin VI to the treated surface. Excess myosin was washed 3 times with $1 \times$ AB.BSA.CAM. Go mix containing 1/25X F-actin, 0.05 mg/mL glucose oxidase, 0.01 mg/mL catalase, 1.5 mg/mL glucose, and 2 mM of ATP in $1 \times$ AB.BSA.CAM was introduced to the flow chamber. Then, imaging was immediately performed with ONi microscope (5% power, excitation of a 1 W 532 nm laser, and a illumination angle of 54°). Movies of gliding actin filaments were obtained at >10 frames/sec.

Actin Gliding Speed Analysis. The gliding assay movies were analyzed with ImageJ.³⁰ The movements of actin filaments were tracked with MTrackJ³¹ plugin based on the movement of their plus (pointed) end between consecutive frames (1 sec intervals) for at least 15 frames. Gliding speed of each filament was measured by dividing the total distance traveled by the elapsed time ($\Delta t = 15\text{--}25$ sec) to cover the measured distance.

Atomic Force Microscopy. All AFM images were acquired using a Dimension FastScan Bio (Bruker) using the “short and fat”, or “long and thin” ScanAsyst-IN AIR or ScanAsyst-FLUID+ cantilever of an SNL probe (“sharp nitride lever”, Bruker) in ScanAsyst Air or Fluid mode. NanoScope Analysis 2.0 and Mathematical (Wolfram Research) were used to measure the surface height distribution, HMDS film thickness, and generate the AFM image with appropriate color scales.

Capillary Tube Preparation. Rectangle Boro Capillary Tubes ($0.2 \text{ mm} \times 2 \text{ mm} \times 50 \text{ mm}$; Vitrotubes (product number: 3520) were washed with isopropanol and then rinsed with deionized water before drying with a nitrogen gas. For nitrocellulose and HMDS treatments, the capillary tubes were plasma cleaned for 5 min at high RF setting (Harrick Plasma; Plasma Cleaner PDC-32G). For nitrocellulose coating: one end of the capillary tube was dipped into a microcentrifuge tube containing 30 μL 1% nitrocellulose in amyl acetate until the liquid reached the other end. The excess nitrocellulose was drained by blotting one end with an absorbent task wipe, then hung vertically to dry overnight. For HMDS coating: The capillary tubes were placed vertically inside a vacuum desiccator chamber together with a vial containing 300 μL of HMDS solution at room temperature. Then, the chamber was connected to a vacuum pump in a fume hood to deposit the HMDS vapor for 20 min.

Capillary Tube Contact Angle Measurements. The capillary tubes were arranged in a clear flat bottom tray with multiple wells and stood upright. A Canon EOS 77D camera body with an EF-S 35mm f/2.8 Macro IS STM Canon lens was arranged reasonably far away to minimize camera distortion effects but close enough to capture the 4 tubes such that the meniscus and bottom of the capillary tube were clearly visible. 70 μL of a solution of 1 mL MilliQ water and $\sim 5 \mu\text{L}$ of red food coloring was loaded into each well that contained a capillary tube. The capillary tubes were left undisturbed for 20 min after which a still photo was taken. The photo was analyzed using Logger Pro 3 by Vernier. The height of each capillary tube was measured to be 50.9 mm and was used to set the scale for each height measurement. The height

from the base of the capillary tube to the lower meniscus was recorded as the height. 3 repeats were taken with freshly prepared tubes each time. Jurin’s Law was used to calculate the contact angles, first using the assumption that the “radius” of the capillary tube was circular

$$r \sim \sqrt{\frac{l \times w}{\pi}} = \sqrt{\frac{0.2 \text{ mm} \times 2 \text{ mm}}{\pi}}, \quad [2]$$

then using the assumption that the capillary tube was a pair of parallel plates ($r = 0.2 \text{ mm}$). Jurin’s law is unable to calculate the contact angle when the water height is the full length of the capillary tube which happens consistently when the capillary tube is plasma cleaned.

Gliding Assay with Capillary Tubes. The volume of a capillary tube is 200 μL according to the specifications given by the manufacturer. 200 μL of myosin VI is flowed into the capillary tube by dipping one end into the solution and tipping the tube such that it is horizontal. This allows the solution to be taken up via capillary effect. The myosin VI is allowed to incubate for 15 min. In order to flow the next few solutions, a different approach must be used. The capillary tube is placed in the solution and a dry wipe is used to absorb the solution from the other end causing the capillary tube to take in the solution it is placed in. Using this method, we washed the capillary tube with approximately 600 μL of $1\times$ AB.BSA.CAM followed by 200 μL of Go-mixture containing $1/25\times$ F-actin, 0.05 mg/mL glucose oxidase, 0.01 mg/mL catalase, 1.5 mg/mL glucose, and 2 mM of ATP in 1X AB.BSA.CAM. Then, imaging was immediately performed with ONi microscope (8% power, excitation of 532 nm or green laser, and a TIRF excitation angle of $\sim 50^\circ$). In most experiments, 2-min movies of gliding actin filaments were obtained at 10 fps.

Statistical analysis. The p -values were computed by two-tail unpaired Student’s t test using Prism 9 (GraphPad). We calculated the error bars as $\text{mean} \pm \text{SD}$ throughout this work, with the exception of Fig. 4B ($\text{mean} \pm \text{SEM}$).

Supporting Information

Facile surface modification with hexamethyldisilazane to generate atomically flat and hydrophobic substrates for filament gliding assays with molecular motors

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S1. Supplementary Figures

Fig. S1 AFM image of HMDS-coated glass surface. Vapor deposition of HMDS maintains the surface flatness.

Fig. S2 AFM image of nitrocellulose-coated glass surface shows that nitrocellulose coating does not produce a flat surface.

Fig. S3 Gliding speed of actin filaments in capillary tubes ($N=29$ filaments). One data point (296 nm/sec) was excluded from the plot and from calculating the mean $\pm$ SD. This designated outlier is $>5\sigma$ from the estimated mean $\pm$ SD (107 $\pm$ 35 nm/sec). The mean was calculated after excluding the outlier.

S2. Supplementary Movies

Movies S1–S13 are available at <https://github.com/bionicslab/HMDS-manuscript>.

Movie S1 Photobleaching of glass coverslip.

Movie S2 Photobleaching of HMDS-coated coverslip.

Movie S3 Photobleaching of nitrocellulose-coated coverslip.

Movie S4 Gliding assay of actin filaments on myosin VI immobilized on HMDS-coated surface (PC glass + HMDS).

Movie S5 Gliding assay of actin filaments on myosin VI immobilized on nitrocellulose-coated surface (PC glass + nitrocellulose).

Movie S6 Gliding assay of actin filaments on myosin VI immobilized on HMDS-coated surface preincubated with BSA (PC glass + HMDS + BSA).

Movie S7 Gliding assay of actin filaments on myosin VI immobilized on plasma-cleaned glass preincubated with BSA (PC glass + BSA).

- Movie S8** Gliding assay of actin filaments on myosin VI immobilized on HMDS-coated surface preincubated with Tween-20 (PC glass + HMDS + Tween-20).
- Movie S9** Gliding assay of actin filaments on myosin VI immobilized on plasma-cleaned glass preincubated with Tween-20 (PC glass + Tween-20).
- Movie S10** Gliding assay of actin filaments on myosin VI immobilized on plasma-cleaned glass (PC glass).
- Movie S11** Gliding assay of actin filaments on myosin VI immobilized on HMDS-coated capillary tube (PC glass + HMDS).
- Movie S12** Gliding assay of actin filaments on myosin VI immobilized on untreated capillary tube.
- Movie S13** Gliding assay of actin filaments on myosin VI immobilized on plasma-cleaned capillary tube (PC glass).