

Biomimetic Total Syntheses of (+)-Chloropupukeananin, (–)-Chloropupukeanolide D, and Chloropestolides

Takahiro Suzuki,^{*,†} Soichiro Watanabe,[‡] Wataru Ikeda,[‡] Susumu Kobayashi,[§] and Keiji Tanino[†]

[†]Department of Chemistry, Faculty of Science, Hokkaido University, Sapporo, 060-0810, Hokkaido, Japan

[‡]Graduate School of Chemical Sciences and Engineering, Hokkaido University, Sapporo, 060-0810, Hokkaido, Japan

[§]Faculty of Pharmaceutical Sciences, Tokyo University of Science, 2641 Yamazaki, Noda, Chiba, 278-8510, Japan

ABSTRACT: Chloropupukeananin, chloropupukeanolides, and chloropestolides are a family of structurally complex bioactive natural products that possess highly functionalized tricyclo[4.3.1.0^{3,7}]decane or bicyclo[2.2.2]octane skeletons. Biosynthesis of the chloropupukeananin family is triggered by the intermolecular heterodimeric Diels-Alder reaction between maldoxin and iso-A82775C; however, the enzymes involved have not yet been identified. We herein report the one-pot biomimetic synthesis of chloropupukeananin and chloropupukeanolide D. Moreover, the effect of solvent on the intermolecular Diels-Alder reaction of siccayne and maldoxin suggested that the biosynthesis of the chloropupukeananin family involves a Diels-Alderase catalyzed heterodimeric Diels-Alder reaction.

INTRODUCTION

Natural products owe their structural diversity to enzymes. Enzymes, such as prenyl transferases¹ and terpene cyclases,² which catalyse the carbon-carbon bond formation and cyclisation reactions, are responsible for the diversity in the carbon skeleton of natural products. The Diels-Alder reaction is one of the most powerful tools to introduce molecular complexity through biosynthetic,³ and chemical synthesis approaches.⁴ Several natural products are synthesized through the Diels-Alder reaction in nature. However, most of these are only artifacts or products of spontaneous cycloaddition.⁵ Since the discovery of a Diels-Alderase in the biosynthesis of solanapyrone,^{6a} only a few classes of this enzyme, that can catalyze intramolecular reactions in polyketide synthesis,^{6b-e} have been identified. Very recently, enzymes promoting the intermolecular Diels-Alder reactions to produce pseudo-dimeric resveratrols have been identified.⁷ Artificial Diels-Alderases, such as antibody catalysts,⁸ artificial enzymes,⁹ and super molecules,¹⁰ have been developed to realize intermolecular Diels-Alder reactions owing to the high synthetic convergence and broad diversity of products of this process. If an intermolecular heterodimeric Diels-Alderase is identified, its genetic modifications should pave the way for the versatile production of bioactive natural product-like compounds.¹¹

Chloropupukeananin (**1**), an inhibitor of HIV-1 replication, was isolated from the plant endophytic fungus *Pestalotiopsis fici* by Che et al. as the first chlorinated pupukeanane derivative

(Figure 1).¹² Simultaneously, iso-A82775C (**2**) and pestheic acid (**3**) were also isolated and proposed as the biosynthetic precursors of **1**. One of the prominent structural features of **1** is its highly functionalized tricyclo[4.3.1.0^{3,7}]decane moiety that bears the vinyl allene moiety of **2** and the methyl benzoate moiety of **3**. Thus, we proposed that (*R*)-maldoxin (**4**), which is generated by the asymmetric oxidative dearomatization of **3**, is the actual biosynthetic precursor instead of **3**.¹³ Subsequently, an intermolecular reverse-electron-demand Diels-Alder reaction between **2** and **4** and an intramolecular carbonyl-ene reaction leads to the construction of the tricyclo[4.3.1.0^{3,7}]decane skeleton. Finally, migration of the *p*-orsellinate moiety results in **1**. Continuous efforts by Che et al. to unveil the biosynthetic pathway of **1** led to the identification of many natural heterodimeric congeners.¹⁴ These included (i) chloropestolides A–F (**5–10**),^{14a,b} the isomers of the Diels-Alder reactions, (ii) chloropupukeanolide D and C (**11**, **12**),^{14c} a carbonyl-ene products of **7** and **8**, and (iii) other degradation products.¹⁵ Finally, optically active (*R*)-**4**,^{14d} which was the missing link in the biosynthesis of **1**, was isolated from the related fungus, *P. theae*. These results support our hypothesis on the biosynthesis of **1**. Chloropestolides H–K (**13–16**)^{14e} were also isolated during the identification of the biosynthetic gene cluster of **2** using a prenyl transferase gene-disrupted strain. By inhibiting the biosynthesis of **2**, intermediate **17** was found to undergo intermolecular Diels-Alder reactions with **4** to afford **13–16**. Isolation of the chloropestolides (**5–8**, **13–16**) indicates poor facial selectivity of the Diels-Alder reaction during biosynthesis (Figure 2). Briefly, during the formation of these bicyclo[2.2.2]octane-containing cycloadducts, the dienophiles approach the *Re*- or *Si*-face of C4' of diene **4**, with *syn*- or *anti*-orientation between the R group of the dienophiles and the C2 acetal moiety of **4**. The intermolecular Diels-Alder reaction produces all the possible cycloadducts, with some *Si-syn* selectivity; these adducts are artifacts owing to the poor selectivity. Thus, the chemical synthesis of a chloropupukeananin family through the Diels-Alder reaction between (+)-**2** and (–)-**4** can elucidate the biosynthetic pathway, especially clarifying whether a non-enzymatic Diels-Alder reaction occurs or not. If an enzymatic reaction occurs, this enzyme might be the first example of an intermolecular heterodimeric Diels-Alderase that could construct a highly functionalized bicyclo[2.2.2]octane skeleton. Thus, we herein report the biomimetic total synthesis of (+)-**1** from (+)-**2** and (–)-**4** using a one-pot Diels-Alder/carbonyl-ene cascade reaction and migration of the *p*-orsellinate group.

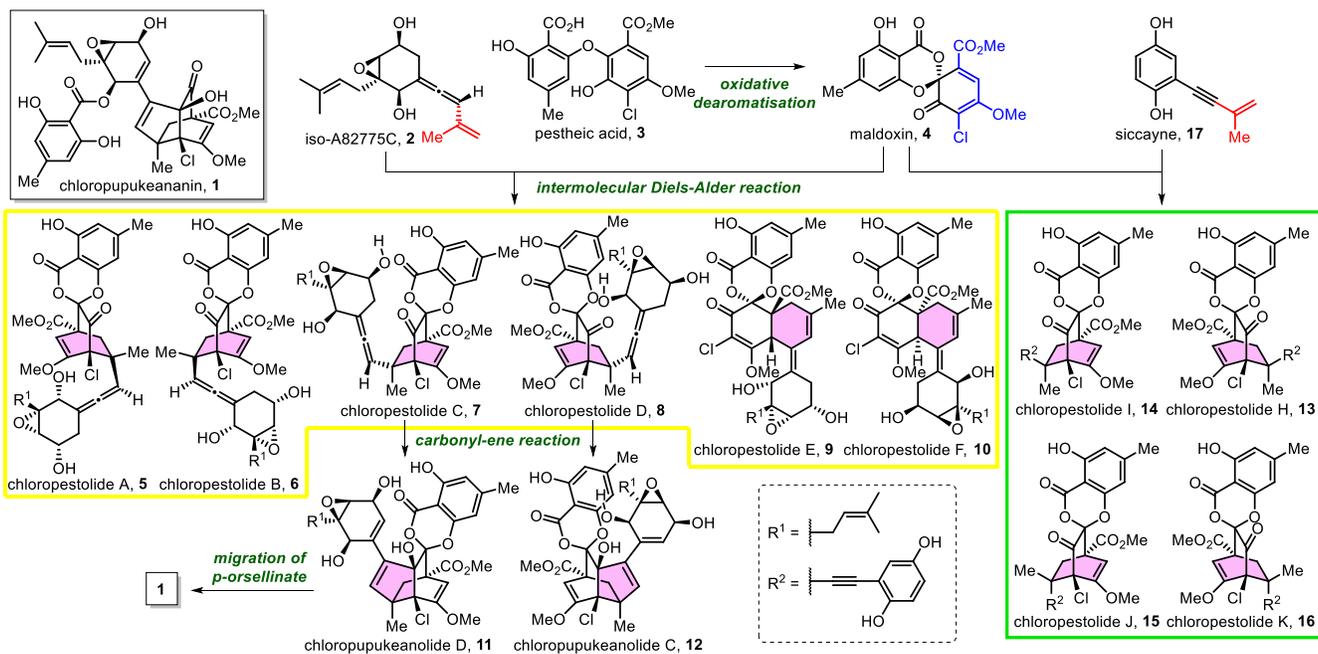


Figure 1. Proposed biosynthetic pathway of the chloropupukeananin family (the Diels-Alder adducts from **2** and **4** are framed in yellow box and those from **4** and **17** are framed in green box)

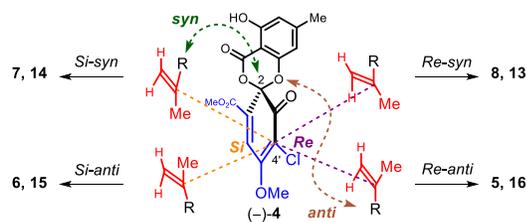
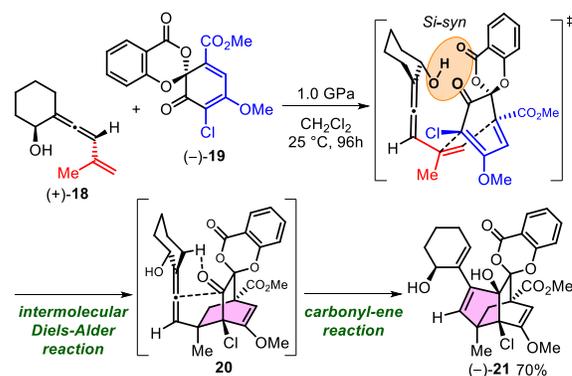


Figure 2. Four orientations of the diene and dienophile moieties in the intramolecular Diels-Alder reaction

RESULTS AND DISCUSSION

Our previous synthetic study on chloropupukeananin¹⁶ demonstrated the selective construction of a tricyclo[4.3.1.0^{3,7}]decane skeleton under high pressure conditions (Scheme 1). The Diels-Alder/carbonyl-ene cascade reaction of model compounds (+)-**18** and (-)-**19** results in tricyclic compound (-)-**21** in 70% yield via *Si-syn* cycloadduct **20**. The *Si-syn* transition state is stabilized by hydrogen bonding networks. Therefore, the chemical Diels-Alder reactions between (+)-**2** and (-)-**4** should selectively produce *Si-syn* cycloadduct **7**, whereas that in nature exhibits poor selectivity, forming cycloadducts **5–10**. This is contradictory to the usual biosynthetic enzyme reactions. However, it is also possible that *Si-syn* **7** is biosynthetically obtained as a major product, without the involvement of enzymes. The subsequent reactions, such as the carbonyl-ene reaction or other degradation reactions, consumed most of **7** in the presence or absence of enzymes. Consequently, all the cycloaddition products, **5–10**, are isolated in a non-selective manner. Hence, a simple system to directly compare the cycloadducts of the chemical synthesis with the natural cycloadducts is required.

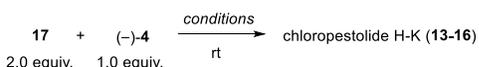
Scheme 1. Diels-Alder/carbonyl-ene cascade reaction in our previous study



Accordingly, we investigated the biosynthetic Diels-Alder reaction of **17**¹⁷ and (-)-**4** to produce **13–16** (Table 1). The reaction was initially conducted under neat conditions (entry 1), resulting in a mixture of products **13–16** (**13**:**14**:**15**:**16** yield ratio = 9:32:51:8) produced in quantitative yield. Owing to the nature of (-)-**4**,¹⁸ the *Si*-face was preferred. Performing the reaction in MeOH/H₂O (entry 2) resulted in a mixture containing predominantly *Si-anti* **15** (**13**:**14**:**15**:**16** = 4:11:74:11), which was incompatible with the *Si-syn* selectivity of isolation.^{14e} Surprisingly, the product ratio obtained in the aqueous solvent differed from that of the isolated sample; generally, hydrophilic solvents can simulate the biosynthetic reaction conditions. Next, we considered the relationship between the product ratio and solvent to reproduce the isolated ratio of **13–16**. After investigating the Diels-Alder reaction in common organic solvents (entries 3–10; for the complete solvent screening data, see the Supporting Information), we established that the product ratio, especially for **14**:**15**, depended on the solvent basicity (SB)¹⁹ and not on the dielectric constant, dipole moment, or solvent polarity parameter.²⁰ In low-basicity solvents (SB < 0.2), **14** and **15** were obtained in a ratio of nearly 1:1. The best result

for the *Si-syn* selectivity was obtained in CH₂Cl₂ (entry 4), giving **14** and **15** in isolated yields of 39% and 34%, respectively.²¹ In contrast, in high-basicity solvents (SB > 0.2), the Diels-Alder reaction resulted in predominantly **15**. The same trends were observed under high-pressure conditions (entries 11 and 12). Comparing the ratio of **13-16** isolated from nature^{14e} revealed that the preference for **14** was not reproduced in any of organic solvents. Comparatively similar results were obtained in lipophilic solvents (entries 3 and 4) rather than in hydrophilic solvents (entries 2, 7, and 9). Thus, there was no possibility that cycloadducts **13-16** are artifacts during cell cultivation and isolation. These results suggest that the biosynthetic production of the chloropupekeananin family involves the heterodimeric Diels-Alderase catalyzed the intermolecular Diels-Alder reaction of (-)-**4**.

Table 1. Intermolecular Diels-Alder reaction between (-)-4** and **17****



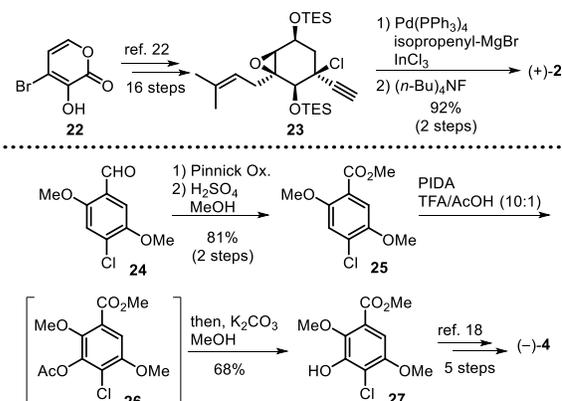
entry	conditions	SB	time (h)	ratio ^a			
				13	14	15	16
	isolation (ref.14e)			17	52	18	13
1	neat		24	9	32	51	8
2	MeOH/H ₂ O (4/1)		48	4	11	74	11
3	benzene	0.124	48	9	42	44	5
4 ^b	CH ₂ Cl ₂	0.178	48	13	44 (39)	37 (34)	6
5	CH ₃ CN	0.286	48	10	28	53	9
6	acetone	0.475	48	6	20	64	10
7	MeOH	0.545	48	4	11	75	10
8	Et ₂ O	0.562	48	6	17	69	7
9	EtOH	0.658	48	3	7	80	10
10	<i>i</i> -PrOH	0.762	48	3	5	81	10
11 ^c	1.0 GPa, MOH/CH ₂ Cl ₂ (10/1)		1	3	11	75	11
12 ^c	1.0 GPa, CH ₂ Cl ₂		1	15	46	33	6

^a Ratio was determined by ¹H-NMR. ^b Isolated yield in parentheses. ^c 1.1 Equiv. of **17** was used.

Subsequently, the next stage of the biomimetic total synthesis of chloropupekeananin, based on our proposed biosynthetic pathway, was considered. Scaling up of the previously reported syntheses of (+)-**2**²² and (-)-**4**¹⁸ is associated with limitations, such as low conversion in the *anti*-S_N2' reaction of **23** using a vinyl copper reagent in the total synthesis of (+)-**2** (Scheme 2). This problem was addressed by using an organoindium reagent and Pd catalyst,²³ and subsequently deprotecting the triethylsilyl (TES) groups of the resulting allene, resulted in (+)-**2** in 92% yield on a sub-gram scale.²⁴ In the original synthesis of (-)-**4**, phenol **27** was prepared from 5-methoxysalicylic acid in five steps, one of which was a non-selective chlorination reaction.¹³ The alternative synthesis was initiated with **24**, which was prepared by the formylation of commercially available 2-chloro-1,4-dimethoxybenzene.²⁵ The Pinnick oxidation and esterification of **24** gave salicylate **25** in 81% yield. Under Kita's conditions,²⁶ site-selective oxidation

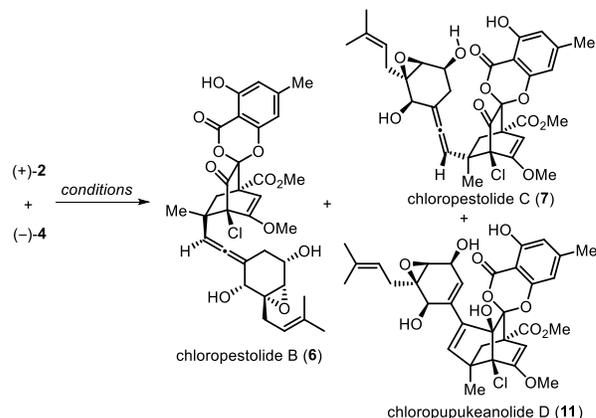
using (diacetoxyiodo)benzene (PIDA) in a trifluoroacetic acid (TFA)/acetic acid mixed solvent²⁴ gave acetate **26**, which underwent a one-pot hydrolysis to form **27** in 68% yield. Phenol **27** was converted to (-)-**4** according to our previously reported synthetic protocol.¹⁸

Scheme 2. Improved synthesis of (+)-2** and (-)-**4**.**



Next, we investigated the Diels-Alder/carbonyl-ene cascade reaction using the biosynthetic precursors (+)-**2** and (-)-**4** (Table 2). A mixture of (+)-**2** and (-)-**4** was first subjected to a high pressure in CH₂Cl₂ resulting in the formation of the desired *Si-syn* **7** and one diastereomer, *Si-anti* **6**, after 1 h (entry 1). Notably, no other cycloadducts were detected except for a small

Table 2. Intermolecular Diels-Alder/carbonyl-ene cascade reactions (+)-2** and (-)-**4** under high pressure and atmospheric pressure conditions**



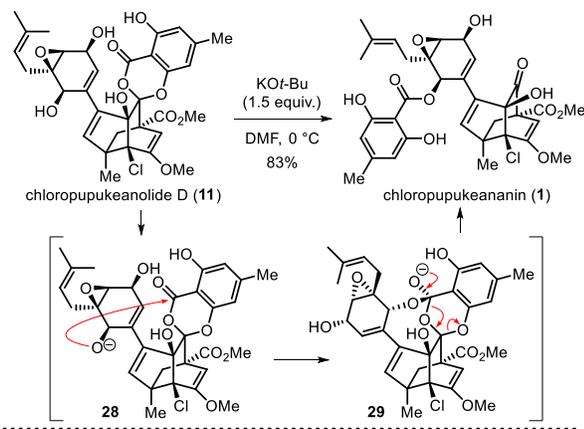
entry	ratio (2/4)	conditions	temp. (°C)	time (h)	yield (%) ^a		
					6	7	11
1	1.3/1.0	1.0 GPa, CH ₂ Cl ₂	25	1	17	67	3
2	1.3/1.0	1.0 GPa, CH ₂ Cl ₂	25	4	17	62	13
3	1.3/1.0	1.0 GPa, CH ₂ Cl ₂	25	16	17	42	40
4	1.3/1.0	1.0 GPa, CH ₂ Cl ₂	25	64	17	5	71
5 ^b	1.3/1.0	(i) 1.0 GPa, CH ₂ Cl ₂ (ii) 0.1 MPa, neat	25 60	64 9	21	-	69
6	1.0/1.5	0.1 MPa, (CH ₂ Cl) ₂	70	30	21	13	19
7 ^b	1.0/1.0	(i) 0.1 MPa, neat (ii) 0.1 MPa, neat	25 60	120 68	25	-	57

^a NMR yield otherwise noted. ^b Isolated yield.

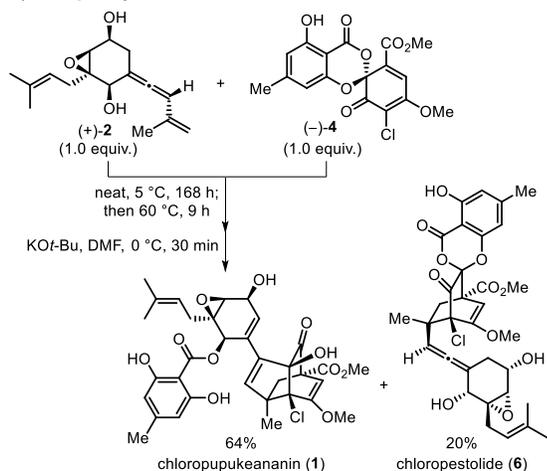
amount of **11**. The *Si-syn* selectivity would be attributed to the hydrogen bonding networks, as shown in Scheme 1. Prolonging the reaction time facilitated the intramolecular carbonyl-ene reaction; when the reaction was conducted for 64 h (entry 4), **11** was obtained in 71% yield. Since the mixture of **7** and **11** was inseparable, it was heated under atmospheric pressure for the carbonyl-ene reaction to reach completion. After the high-pressure reaction (entry 5), the reaction mixture was concentrated and heated to 60 °C to afford **11** and **6** in isolated yields of 69% and 21%, respectively. We also investigated the Diels-Alder reaction under atmospheric pressure. The thermal reaction (entry 6) produced a mixture of **6**, **7**, and **11** in low yields owing to the decomposition of the starting materials. Next, the reaction was conducted under neat conditions (entry 7), followed by heating to 60 °C to produce the desired compound **11** in 57% yield, along with **6** in 25% yield. These results indicated that a high pressure significantly accelerated the Diels-Alder reactions between (+)-**2** and (-)-**4** but did not affect the facial selectivity. This synthetic protocol demonstrates the intriguing *Si*-face selectivity of the intermolecular Diels-Alder reaction between (+)-**2** and (-)-**4**; i.e., only two cycloadducts, **6** and **7**, were obtained in the chemical synthesis, while six types of cycloadducts **5–10** were isolated as natural products.^{14a,b} This suggests that the biosynthetic intermolecular Diels-Alder reactions of **5–10** and **13–16** involve enzymes that activate the substrates to form diverse Diels-Alder products.

Scheme 3. Completion of biomimetic total synthesis of chloropupukeananin

a) Migration of the *p*-orsellinate group of **11**



b) One-pot synthesis of **1**



Finally, the migration of the *p*-orsellinate group of **11** was examined (Scheme 3a). We postulated that the secondary alkoxide group of **28** generated from **11** in basic conditions attacks the carbonyl carbon of *p*-orsellinate group to form intermediate **29**. Then, elimination reaction completes the migration to give **1**. Treatment of **11** with KOt-Bu in DMF afforded **1** in 83% yield. Because the Diels-Alder/carbonyl-ene cascade reaction did not require any reagents, we attempted the one-pot biomimetic transformation from (+)-**2** and (-)-**4** (Scheme 3b). The atmospheric pressure cascade reaction (neat, 5 °C, 7 days; then 60 °C, 9 h) and the following migration reaction provided **1** in 64% yield and **6** in 20% yield. Thus, the first asymmetric total synthesis of **1** was accomplished. The spectral data of **1**, **6**, and **11**²⁷ were identical to those of the corresponding natural compounds.^{12,14b,c}

CONCLUSION

The biomimetic total synthesis of (+)-chloropupukeananin and (-)-chloropupukeanolide D through an intermolecular Diels-Alder reaction using (-)-maldoxin and (+)-iso-A82775C and a carbonyl-ene reaction was achieved. The biomimetic synthesis of chloropestolides H–K using (-)-maldoxin and siccayne unveiled the effect of solvent on the intermolecular Diels-Alder reaction. The origin of the intermolecular Diels-Alder reaction stereoselectivity is still elusive; however, biosynthesis of the chloropupukeananin family possibly involves an enzymatically catalyzed intermolecular heterodimeric Diels-Alder reaction. We expected that our chemical synthesis of the family of chloropupukeananin advance the identification of the first intermolecular heterodimeric Diels-Alderase, which can produce diverse highly functionalized bicyclo[2.2.2]octanes.

ASSOCIATED CONTENT

Supporting Information

This material is available free of charge via the Internet at <http://pubs.acs.org>.
Experimental procedures and compound characterization (PDF)

AUTHOR INFORMATION

Corresponding Author

* Takahiro Suzuki
ORCID: 0000-0002-3842-1025
E-mail: takahiro-suzuki@sci.hokudai.ac.jp

Notes

The authors declare no competing financial interests.

ACKNOWLEDGMENT

This research was supported in part by JSPS KAKENHI grant number JP15H05842 in the Middle Molecular Strategy, JP18H01970 in Grant-in-Aid for Scientific Research(B), JP20K05485 in Grant-in-Aid for Scientific Research(C), and the Photo-excitonix Project of Hokkaido University. We thank Dr. M. Sodeoka and Dr. S. Kawamura (RIKEN) for assistance with the operation of the high-pressure apparatus. We also thank the Naito Foundation, the Uehara Memorial Foundation, the

Kurata Memorial Hitachi Science and Technology Foundation, and the Research Foundation for Pharmaceutical Sciences for their financial support.

REFERENCES

- (1) (a) Liang, P. H.; Ko, T. P.; Wang, A. H., Structure, mechanism and function of prenyltransferases. *Eur. J. Biochem.* **2002**, *269*, 3339–3354. (b) Takahashi, S.; Koyama, T., Structure and function of cis-prenyl chain elongating enzymes. *Chem. Rec.* **2006**, *6*, 194–205.
- (2) Christianson, D. W., Structural and Chemical Biology of Terpenoid Cyclases. *Chem. Rev.* **2017**, *117*, 11570–11648.
- (3) (a) Klas, K.; Tsukamoto, S.; Sherman, D. H.; Williams, R. M., Natural Diels-Alderase: Elusive and Irresistible. *J. Org. Chem.* **2015**, *80*, 11672–85. (b) Minami, A.; Oikawa, H., Recent advances of Diels-Alderase involved in natural product biosynthesis. *J. Antibiot.* **2016**, *69*, 500–506. (c) Jeon, B. S.; Wang, S. A.; Ruszczycy, M. W.; Liu, H. W., Natural [4+2]-Cyclases. *Chem. Rev.* **2017**, *117*, 5367–5388. (d) Jamieson, C. S.; Ohashi, M.; Liu, F.; Tang, Y.; Houk, K. N., The expanding world of biosynthetic pericyclases: cooperation of experiment and theory for discovery. *Nat. Prod. Rep.* **2019**, *36*, 698–713.
- (4) Nicolaou, K. C.; Snyder, S. A.; Montagnon, T.; Vassilikogiannakis, G., The Diels-Alder Reaction in Total Synthesis. *Angew. Chem. Int. Ed.* **2002**, *41*, 1668–1698.
- (5) (a) Gravel, E.; Poupon, E., Biogenesis and Biomimetic Chemistry: Can Complex Natural Products Be Assembled Spontaneously? *Eur. J. Org. Chem.* **2007**, *2008*, 27–42. (b) Wang, T.; Hoye, T. R., Diels-Alderase-free, bis-pericyclic, [4+2] dimerization in the biosynthesis of (+/-)-paracaseolide A. *Nat. Chem.* **2015**, *7*, 641–645. (c) Godfrey, R. C.; Green, N. J.; Nichol, G. S.; Lawrence, A. L., Total synthesis of brevianamide A. *Nat. Chem.* **2020**, *12*, 615–619.
- (6) (a) Oikawa, H.; Suzuki, Y.; Naya, A.; Katayama, K.; Ichihara, A., First Direct Evidence in Biological Diels-Alder Reaction of Incorporation of Diene-Dienophile Precursors in the Biosynthesis of Solanapyrones. *J. Am. Chem. Soc.* **1994**, *116*, 3605–3606. (b) Auclair, K.; Sutherland, A.; Kennedy, J.; Witter, D. J.; Van den Heever, J. P.; Hutchinson, C. R.; Vederas, J. C., Lovastatin Nonaketide Synthase Catalyzes an Intramolecular Diels-Alder Reaction of a Substrate Analogue. *J. Am. Chem. Soc.* **2000**, *122*, 11519–11520. (c) Hashimoto, T.; Hashimoto, J.; Teruya, K.; Hirano, T.; Shin-ya, K.; Ikeda, H.; Liu, H. W.; Nishiyama, M.; Kuzuyama, T., Biosynthesis of versipelostatatin: identification of an enzyme-catalyzed [4+2]-cycloaddition required for macrocyclization of spirotetrone-containing polyketides. *J. Am. Chem. Soc.* **2015**, *137*, 572–575. (d) Byrne, M. J.; Lees, N. R.; Han, L. C.; van der Kamp, M. W.; Mulholland, A. J.; Stach, J. E.; Willis, C. L.; Race, P. R., The Catalytic Mechanism of a Natural Diels-Alderase Revealed in Molecular Detail. *J. Am. Chem. Soc.* **2016**, *138*, 6095–6098. (e) Kim, H. J.; Ruszczycy, M. W.; Choi, S. H.; Liu, Y. N.; Liu, H. W., Enzyme-catalyzed [4+2] cycloaddition is a key step in the biosynthesis of spinosyn A. *Nature* **2011**, *473*, 109–112.
- (7) Gao, L.; Su, C.; Du, X.; Wang, R.; Chen, S.; Zhou, Y.; Liu, C.; Liu, X.; Tian, R.; Zhang, L.; Xie, K.; Chen, S.; Guo, Q.; Guo, L.; Hano, Y.; Shimazaki, M.; Minami, A.; Oikawa, H.; Huang, N.; Houk, K. N.; Huang, L.; Dai, J.; Lei, X., FAD-dependent enzyme-catalyzed intermolecular [4+2] cycloaddition in natural product biosynthesis. *Nat. Chem.* **2020**, *12*, 620–628.
- (8) Gouverneur, V. E.; Houk, K. N.; de Pascual-Teresa, B.; Beno, B.; Janda, K. D.; Lerner, R. A., Control of the exo and endo pathways of the Diels-Alder reaction by antibody catalysis. *Science* **1993**, *262*, 204–208.
- (9) Siegel, J. B.; Zanghellini, A.; Lovick, H. M.; Kiss, G.; Lambert, A. R.; St Clair, J. L.; Gallaher, J. L.; Hilvert, D.; Gelb, M. H.; Stoddard, B. L.; Houk, K. N.; Michael, F. E.; Baker, D., Computational design of an enzyme catalyst for a stereoselective bimolecular Diels-Alder reaction. *Science* **2010**, *329*, 309–313.
- (10) Palma, A.; Artelsmair, M.; Wu, G.; Lu, X.; Barrow, S. J.; Uddin, N.; Rosta, E.; Masson, E.; Scherman, O. A., Cucurbit[7]uril as a Supramolecular Artificial Enzyme for Diels-Alder Reactions. *Angew. Chem. Int. Ed.* **2017**, *56*, 15688–15692.
- (11) Basler, S.; Studer, S.; Zou, Y.; Mori, T.; Ota, Y.; Camus, A.; Bunzel, H. A.; Helgeson, R. C.; Houk, K. N.; Jimenez-Oses, G.; Hilvert, D., Efficient Lewis acid catalysis of an abiological reaction in a de novo protein scaffold. *Nat. Chem.* **2021**, *13*, 231–235.
- (12) Liu, L.; Liu, S.; Jiang, L.; Chen, X.; Guo, L.; Che, Y., Chloropupukeananin, the first chlorinated pupukeananine derivative, and its precursors from *Pestalotiopsis fici*. *Org. Lett.* **2008**, *10*, 1397–1400.
- (13) Suzuki, T.; Kobayashi, S., Concise approach to pupukeananine skeleton: synthetic study of chloropupukeananin. *Org. Lett.* **2010**, *12*, 2920–2923.
- (14) (a) Liu, L.; Li, Y.; Liu, S.; Zheng, Z.; Chen, X.; Zhang, H.; Guo, L.; Che, Y., Chloropestolide A, an antitumor metabolite with an unprecedented spiroketal skeleton from *Pestalotiopsis fici*. *Org. Lett.* **2009**, *11*, 2836–2839. (b) Liu, L.; Li, Y.; Li, L.; Cao, Y.; Guo, L.; Liu, G.; Che, Y., Spiroketal of *Pestalotiopsis fici* provide evidence for a biosynthetic hypothesis involving diversified Diels-Alder reaction cascades. *J. Org. Chem.* **2013**, *78*, 2992–3000. (c) Liu, L.; Bruhn, T.; Guo, L.; Gotz, D. C.; Brun, R.; Stich, A.; Che, Y.; Bringmann, G., Chloropupukeanolides C-E: cytotoxic pupukeananine chlorides with a spiroketal skeleton from *Pestalotiopsis fici*. *Chem. Eur. J.* **2011**, *17*, 2604–2613. (d) Liu, L.; Han, Y.; Xiao, J.; Li, L.; Guo, L.; Jiang, X.; Kong, L.; Che, Y., Chlorotheolides A and B, Spiroketal Generated via Diels-Alder Reactions in the Endophytic Fungus *Pestalotiopsis theae*. *J. Nat. Prod.* **2016**, *79*, 2616–2623. (e) Pan, Y.; Liu, L.; Guan, F.; Li, E.; Jin, J.; Li, J.; Che, Y.; Liu, G., Characterization of a Prenyltransferase for Iso-A82775C Biosynthesis and Generation of New Congeners of Chloropestolides. *ACS Chem. Biol.* **2018**, *13*, 703–711.
- (15) See the Supporting Information.
- (16) Suzuki, T.; Miyajima, Y.; Suzuki, K.; Iwakiri, K.; Koshimizu, M.; Hirai, G.; Sodeoka, M.; Kobayashi, S., Unexpected Diels-Alder/carbonylene cascade toward the biomimetic synthesis of chloropupukeananin. *Org. Lett.* **2013**, *15*, 1748–51.
- (17) Pinault, M.; Frangin, Y.; Genet, J. P.; Zamarlik, H., Total Synthesis of Siccayne. *Synthesis* **1990**, 935–937.
- (18) Suzuki, T.; Watanabe, S.; Uyanik, M.; Ishihara, K.; Kobayashi, S.; Tanino, K., Asymmetric Total Synthesis of (-)-Maldoxin, a Common Biosynthetic Ancestor of the Chloropupukeananin Family. *Org. Lett.* **2018**, *20*, 3919–3922.
- (19) (a) Catalán, J.; Palomar, J.; Díaz, C.; de Paz, J. L. G., On Solvent Basicity: Analysis of the SB Scale. *J. Phys. Chem. A* **1997**, *101*, 5183–5189. (b) Catalán, J.; Díaz, C.; López, V.; Pérez, P.; De Paz, J.-L. G.; Rodríguez, J. G., A Generalized Solvent Basicity Scale: The Solvatochromism of 5-Nitroindoline and Its Homomorph 1-Methyl-5-nitroindoline. *Liebigs Ann. Chem.* **1996**, *1996*, 1785–1794.
- (20) Reichardt, C.; Welton, T., *Solvents and Solvent Effects in Organic Chemistry Fourth, Updated and Enlarged Edition*, Wiley, **2011**, pp 549–553.
- (21) The spectral data of **14** and **15** were identical to those reported in ref 14e. The remaining **13** and **16** was obtained as an inseparable mixture in 17% yield.
- (22) Suzuki, T.; Watanabe, S.; Kobayashi, S.; Tanino, K., Enantioselective Total Synthesis of (+)-Iso-A82775C, a Proposed Biosynthetic Precursor of Chloropupukeananin. *Org. Lett.* **2017**, *19*, 922–925.
- (23) Riveiros, R.; Rodriguez, D.; Perez Sestelo, J.; Sarandeses, L. A., Palladium-catalyzed cross-coupling reaction of triorganoindium reagents with propargylic esters. *Org. Lett.* **2006**, *8*, 1403–1406.
- (24) See the Supporting Information for the details of the reaction optimization.
- (25) Bloomer, J. L.; Stagliano, K. W.; Gazzillo, J. A., Preparation of functionalized juglone acetates and juglones via 1,4-dimethoxynaphthalene derivatives: synthesis of anthraquinones related to rhein and aloemodin. *J. Org. Chem.*, **1993**, *58*, 7906–7912.
- (26) Kita, Y.; Tohma, H.; Hatanaka, K.; Takada, T.; Fujita, S.; Mitoh, S.; Sakurai, H.; Oka, S., Hypervalent Iodine-Induced Nucleophilic Substitution of para-Substituted Phenol Ethers. Generation of Cation Radicals as Reactive Intermediates. *J. Am. Chem. Soc.* **1994**, *116*, 3684–3691.
- (27) The optical rotation value of our synthetic **11** is -3.8 (c 0.12, MeOH), whereas that of natural **11** reported in ref. 14c is $+13.2$ (c 0.1, MeOH). Because natural **11** was obtained as an inseparable mixture with (+)-**1**, the reported optical rotation must turn to dextrorotatory. Therefore, the actual optical rotation of **11** is levorotatory, in a manner similar to model compound (-)-**21**.

