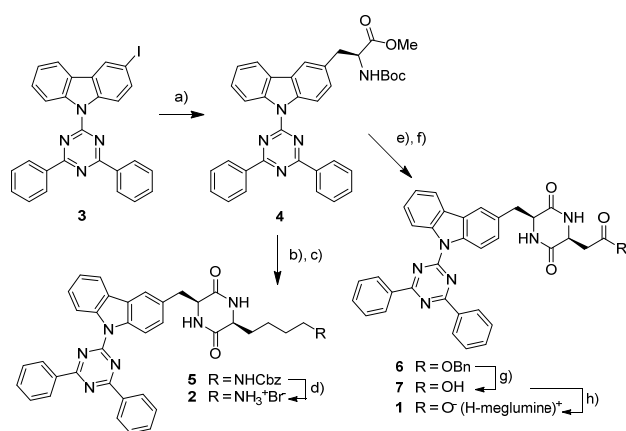


hydrophilicity and solubility of the gelators.^[17,30–33] The *N*-methyl-D-glucamine (meglumine) containing salt **1** was prepared since meglumine has been shown to drastically improve the solubility of an otherwise hydrophobic carboxylic acid salt in water compared to its sodium salt.^[34]

Results and Discussion

Synthesis

Gelators **1** and **2** were synthesized from the protected amino acid **4**. The synthesis of the key intermediate **4** was accomplished by a palladium-catalyzed NEGISHI cross-coupling reaction of the iodinated heterocycle **3** with an iodoalanine derivative in 51% yield (Scheme 2).^[35]



Scheme 2. Synthesis of diketopiperazines **1** and **2**. Conditions: a) Pd₂dba₃, SPhos, (R)-BocNHCH(CH₂ZnI)CO₂Me, DMF/THF, 30 °C, 51%. b) i. TFA, DCM, rt, 18 h; ii. NEt₃, Boc-Lys(Z)-OH, DCM/DMF, HBTU, 0 °C - rt, 2.5 d, 90%. c) i. TFA, DCM, rt, 18 h; ii. NEt₃, AcOH, MeCN, *iso*-BuOH, 100 °C, 3 h, 90%. d) HBr/AcOH, TFA, rt, 75 min, 86%. e) i. TFA, DCM, rt, 18 h; ii. NEt₃, Boc-Asp(OBn)-OH, DCM/DMF, HBTU, 0 °C - rt, 2.5 d, 87%. f) i. TFA, DCM, rt, 18 h; ii. NEt₃, AcOH, MeCN, *iso*-BuOH, 100 °C, 4 h, 81%. g) BBr₃, DCM, -15 °C - rt, 20 h, 84%. h) *N*-methyl-D-glucamine, EtOH, 50 °C, 3 h, 85%. For detailed synthetic procedures and the full synthesis plan see ESI.

Building block **4** was then converted into lysine- and aspartic acid-derived, protected diketopiperazines **5** and **6** in high yields via Boc-deprotection, amide coupling, second Boc-deprotection and acid-catalyzed cyclization (**5**: 81% yield over four steps, **6**: 70% yield over four steps; see ESI for a full scheme).^[25,36] The deprotection of **5** proceeded smoothly in 86% yield using hydrobromic acid in acetic acid.^[37] The deprotection of **6** proved to be difficult at first (see ESI chapter 11). However, we were delighted to find that the deprotection proceeded in high yield of 84% using an excess of boron tribromide in dry dichloromethane.^[38] Finally, the free acid **7** was converted to its meglumine salt **1** in 85% yield by simple heating in ethanol.^[34] Notably, all steps proceeding from **4** were performed without the use of column chromatography, which underlines the robustness of the synthesis of the chosen target structures.

General luminescence properties of the synthesized DPhCzT derivatives

Compound **4** and all derivatives showed blue fluorescence in the solid state (ESI Fig. S3). Compounds **1**, **2** and **7** showed almost the same fluorescence emission spectra in the solid state, with quantum yields ranging between 3% - 4% (ESI Fig. S28-30). DPhCzT has been described to show ultra-long phosphorescence at room temperature in the solid state,^[29] but none was found for **3** or any of its derivatives. In a recent publication by CHEN *et al.*, the long-lived phosphorescence of DPhCzT was attributed to an isomer present as a trace impurity, in which the carbazole unit is replaced by a benzo[*f*]indole unit. The compound stemmed from a 1*H*-benzo[*f*]indole impurity present in commercial carbazole, which reacted in the same way as carbazole, forming the isomer.^[39] In this synthesis, the commercial carbazole was iodinated and the product of the reaction recrystallized prior to the synthesis of **3** (see ESI for the full synthesis), which has most likely removed the impurity and would explain why no phosphorescence was found for the presently prepared samples of **3** or any of its derivatives.

Diketopiperazines **1** and **2** showed aggregation-induced blue-shifted emission (AIBSE) in DMSO/water mixtures (Fig. 1, ESI Fig. S4-5, S7-8). With increasing water content, fluorescence emission changed from a yellow/slightly greenish color to a blue color (ESI Fig. S6, S9). In hexafluoroisopropanol (HFIP)/water mixtures, both compounds showed AIE behavior with increasing water content, as indicated by an increase in intensity of the fluorescence emission (ESI Fig. S10, S13) and a strong blue-shift thereof (Fig. 1, ESI Fig. S11-12, S14-15).

Gelation experiments

After compound **2** was synthesized, its gelation abilities were investigated. **2** was not completely soluble at a concentration of 10 mg/mL in any common organic solvent, not even upon heating. Upon the addition of small amounts of trifluoroacetic acid, **2** readily dissolved in DCM, CHCl₃ and HFIP, resulting in a bright yellow solution that showed increased UV light absorption (ESI Fig. S55). We also found the same reaction towards TFA with **3** in DCM, as well as any derivative of it, except for **1**. We hypothesize that this as a solubilizing, de-aggregating hydrogen bonding interaction between TFA and the heterocyclic nitrogens. The addition of a co-solvent to the HFIP/TFA solution of **2** finally induced gelation accompanied by a decolorization (Table 1, ESI Fig. S25). Translucent gels were formed with water, DMF and DMSO as co-solvents. Opaque gels formed with methanol, isopropyl alcohol, tetrahydrofuran, 1,4-dioxane and ethyl acetate as co-solvents. Notably, mixtures with acetonitrile, acetone and ethyl acetate only formed gels after prolonged standing while simultaneously becoming more translucent. Gels with DMF and DMSO as co-solvents became transparent after three days (ESI Fig. S26). **2** formed gels with all tested co-solvents except for toluene and dichloromethane. When methanol was added to both of these solutions, gelation occurred within 24 h. In all cases, the gelation was not reversible by heating.

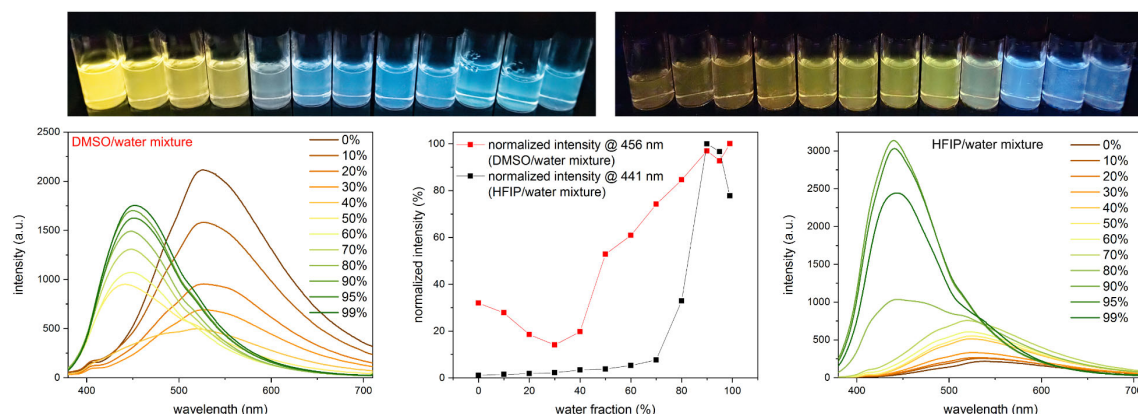


Figure 1. Aggregation behavior of **2** in DMSO and HFIP with increasing water content ($c = 100 \mu\text{M}$, excitation at 365 nm). (Left) fluorescence emission spectra showing AIBSE behavior in DMSO/water mixtures. (Middle) comparison of normalized fluorescence emission intensities at the respective maximum caused by aggregation. (Right) fluorescence emission spectra showing AIE behavior in HFIP/water mixtures..

The critical gelator concentration (CGC) for the HFIP/water (1:1) gel was determined to be 0.4 wt% (7.4 mM) and gels showed blue fluorescence (ESI Fig. S39).

Table 1. Gelation properties of **2** in HFIP/co-solvent mixtures.^[a]

| Co-solvent | 2 ^[b] | Co-solvent | 2 ^[b] |
|--------------------|-------------------------|--------------------|-------------------------|
| water | TG | dimethyl sulfoxide | TG/G ^[c] |
| methanol | OG | tetrahydrofuran | OG |
| isopropyl alcohol | OG | 1,4-dioxane | OG |
| acetonitrile | Sol/TG ^[c] | ethyl acetate | Sol/OG ^[c] |
| acetone | Sol/TG ^[c] | toluene | S/OG ^[d] |
| dimethyl formamide | TG/G ^[c] | dichloromethane | S/OG ^[d] |

[a] A co-solvent (500 μL) was added to a solution of **2** (10 mg) and TFA (15 μL) in HFIP (500 μL). [b] 1 h after addition of the co-solvent. G: Transparent gel; OG: Opaque gel; S: Solution; TG: Translucent gel; [c] after 3 d. [d] 24 h after addition of MeOH (150 μL).

Further experiments were performed to find the smallest fraction of water that causes a HFIP/TFA/water mixture of **1** to form a stable gel (Fig. 2, ESI Table S3). At 0.5 wt% of **1**, we found that a water fraction of 10% did not cause gelation, while a water fraction of 20% or higher caused gelation. The gel obtained from a water fraction of 20% had an intense yellow color that did not fade over time, showing the presence of TFA-*N*-heterocycle interactions within the gel.

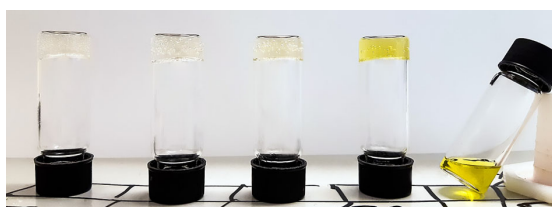


Figure 2. Gelation behavior of **2** in HFIP/water/TFA mixtures at 0.5 wt% with different water fractions. From left to right: (50, 40, 30, 20, 10) vol% H_2O . See ESI Table S3 for more information.

At higher water fractions, the gels were almost colorless (30% and 40% water) or colorless (50% water). (Protic) co-solvents could influence TFA interactions with the gelator in a twofold way: On the one hand, solubilizing hydrogen bonding between TFA and the heterocycle might be broken up, leading to aggregation through aggregation of the heterocycles. On the other hand, co-solvents might break up solubilizing hydrogen bonding between TFA, HFIP and the DKP residue since HFIP has been shown to be significantly potent at solubilizing peptides,^[40] leading to gelation through aggregation by hydrogen bonding of the DKP moieties.

Next, we investigated the hydrogelation properties of the more hydrophilic **1** (Scheme 2). To our delight, **1** was able to form pure hydrogels through a sol-gel process. Heating finely suspended **1** in water at 80 $^\circ\text{C}$ for a few minutes caused a seeming dissolution of the solid. A clear sol instead of a solution formed, that showed blue fluorescence indicating aggregation (ESI Fig. S17, S33). Filtering the sol through a syringe filter (0.3 μm) yielded a filtrate showing no visible fluorescence, confirming the presence of large aggregates in the unfiltered sol. Similarly, DRAPER *et al.* recently showed the effect of temperature on the solution of a peptide-based hydrogelator and its gelation properties.^[41] We found the sol to be stable for months at room temperature without any gelation or precipitation occurring over time. The addition of different gelation triggers to the sol, such as metal ions, acids or other ions, which have been shown to have dramatic effects on gel properties,^[42,43] then led to hydrogelation and to the formation of translucent or transparent hydrogels (Fig. 3).



Figure 3. Hydrogels from **1** (1 wt% in H_2O) and different gelation triggers (3.0 equiv each), 24 h after addition of the trigger. 1) $\text{Zn}(\text{OAc})_2 \cdot 2\text{H}_2\text{O}$; 2)

Ni(OAc)₂·3H₂O; 3) MgSO₄·7H₂O; 4) CuSO₄·H₂O; 5) FeSO₄·7H₂O; 6) AcOH; 7) TFA; 8) Glucono-δ-lactone; 9) *n*Bu₄NBF₄; 10) Ph₂I⁺(OTf)⁻.

Diaryliodonium salts are known photo acids.^[44] However, the addition of the salt to the sol already led to gelation. When the hydrogel prepared from it was irradiated with UV light, fast decomposition of the gel occurred. Very homogenous gels were obtained by gelation with glucono-δ-lactone (GdL).^[45] Using GdL, hydrogels of **1** could be obtained with a CGC as low as 0.3 wt% (3.9 mM). The gelation was in all cases not reversible by heating. This indicates a cation exchange or protonation of **1** taking place prior to gelation, forming the much less water-soluble free acid **7** or different metal salts thereof, which then form the gel network. The indication is supported by the fact that no gels were formed when 0.25 equivalents or less of GdL were added to a 1 wt% sol of **1** in water, indicating that solely the cation-exchanged species are responsible for the gel formation. All hydrogels (except for those from copper and iron salts) showed blue fluorescence (example in ESI Fig. S17, S35). The gels remained stable for as long as they were observed (>6 months) when kept from light and tightly closed. Only the Ph₂I⁺(OTf)⁻ containing gel showed significant sensitivity towards light.

1 was insoluble at concentrations of 10 mg/mL in pure organic solvents, even when heated. Therefore, we further explored the gelation properties of **1** in mixed water/organic solvent systems without additives (Table 2, ESI Table S1). At room temperature, **1** only dissolved completely in a water/THF mixture. This was indicated by a change in fluorescence emission to a yellow/slightly greenish color and confirmed by filtering the solution, which did not change its fluorescence emission intensity (ESI Fig. S31). In contrast, gels formed from mixtures with co-solvents HFIP, acetone, acetonitrile and 1,4-dioxane within 17 h of addition of the co-solvent (ESI Fig. S18). Clear sols formed from methanol, DMSO and DMF as co-solvents. No gels formed in ethanol and *iso*-propanol mixtures, instead slight precipitation occurred. At 80 °C, solutions were formed from mixtures with methanol, ethanol, *iso*-propanol and acetone as co-solvents, as indicated by a change in fluorescence emission from a blue to a yellow/slightly greenish color. The acetonitrile gel and DMF sol both turned to a sol with a yellowish fluorescence containing solid particles showing blue fluorescence, while the HFIP and 1,4-dioxane gels and the DMSO sol remained largely unchanged at 80 °C (ESI Fig. S19). After cooling down, all mixtures formed gels except for the mixtures containing methanol, DMSO, THF and 1,4-dioxane (Fig. 4, ESI Fig. S20).

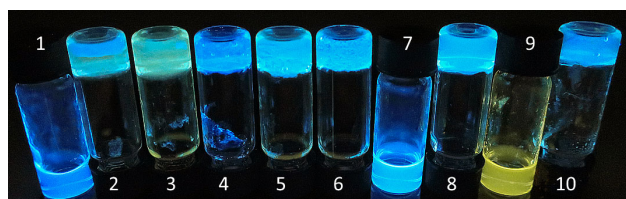


Figure 4. Fluorescence at 366 nm UV-light irradiation of **1** in 1:1 water/organic solvent mixtures (10 mg/mL), after heating to 80 °C and

standing for 1 d. Co-solvents from left to right: 1) MeOH, 2) EtOH, 3) *i*-PrOH, 4) HFIP, 5) acetone, 6) CH₃CN, 7) DMSO, 8) DMF, 9) THF, 10) 1,4-dioxane.

The gels from mixtures containing ethanol, *iso*-propanol, acetone, acetonitrile and DMF showed reversible gelation by heating to 80 °C again and persisted at room temperature for up to 17 h (ESI Table S1).

Full gelation of the mixtures did not necessarily afford the same blue fluorescence indicating aggregation as seen in solid **1**, its sol or its GdL gel, as exemplified by the water/*i*-PrOH gel, which showed a much greener fluorescence. This could either be a solvent effect or might indicate partly disaggregated (solvated) DPhCzT moieties within the gel.

When experimenting with additives for these mixtures, we found that addition of more meglumine (10 mg/mL) to the sol of **1** increased its gelation abilities as well as the transparency of the obtained gels in all mixtures except for the water/HFIP mixture (ESI Table S2, Fig. S23). Initially, there were only minor differences between the mixtures with and without meglumine 17 h after addition of the co-solvents. While the same co-solvents caused gelation and sol formation, the main differences were slight precipitate formation with the co-solvent HFIP and a weaker gel resulting from the co-solvent acetonitrile (ESI Fig. S21). However, when these mixtures were heated to 80 °C, the same yellow/slightly greenish fluorescence emission was observed indicating solution. The only exception was the HFIP mixture which contained insoluble blue fluorescent particles. (ESI Fig. S22). 17 h after cooling down, all mixtures except for the water/THF mixture had formed gels. While **1** alone could not form gels in a water/methanol or water/DMSO mixture at 10 mg/mL, transparent gels were formed with additional meglumine (Table 2).

Table 2. Gelation properties of **1** and **1** + meglumine in H₂O/organic solvent mixtures.^[a]

| Co-solvent | 1 [b] | 1 + meglumine [b][c] | Co-solvent | 1 ^[b] | 1 + meglumine [b][c] |
|----------------|-----------------|-----------------------------------|--------------|-------------------------|-----------------------------------|
| methanol | Sol | G | acetonitrile | TG | G |
| ethanol | TG | G | DMSO | Sol | G |
| <i>i</i> -PrOH | TG | G | DMF | G | G |
| HFIP | TG | PG | THF | S | S |
| acetone | G | G | 1,4-dioxane | TG | TG |

[a] A co-solvent (500 µL) was added to a solution of **1** (10 mg) in H₂O (500 µL). The mixture was left standing for 17 h, then heated to 80 °C for 10 min, then left standing for 24 h. [b] G: Transparent gel; TG: Translucent gel; S: Solution; PG: Partly gel. [c] With 10 mg/mL meglumine. For more detailed gelation properties see ESI Tables S1-2.

Moreover, samples of **1** with additional meglumine formed transparent gels with ethanol, isopropyl alcohol and acetonitrile, whereas **1** alone formed translucent gels (Table 2). Notably, the HFIP containing mixture gelled only partly, dividing into gel and a gel-like precipitate. Gelation was reversible for all gels with added meglumine under heating.

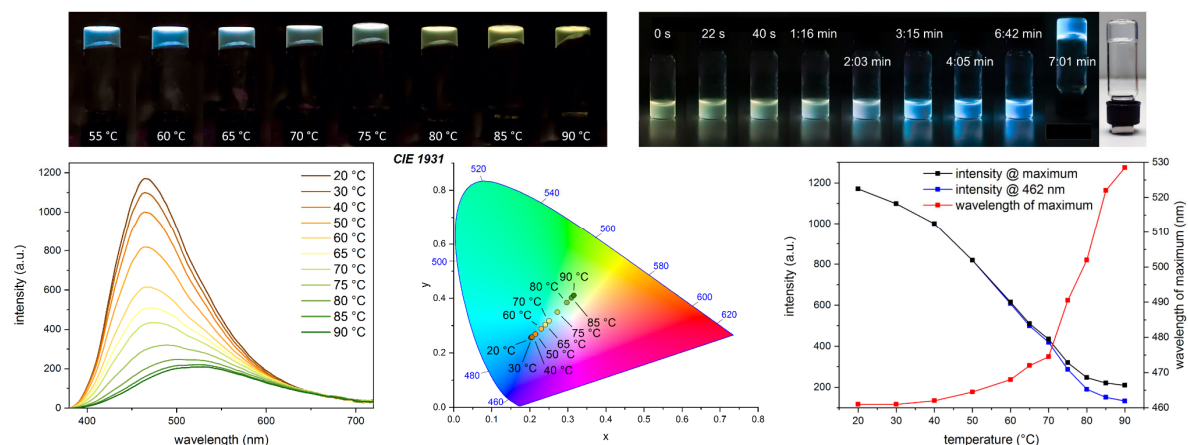


Figure 5. (Top left) temperature dependent fluorescence of the gel of **1** (10 mg/mL) from water:acetonitrile (1:1, with 10 mg/mL meglumine) while heating, and while cooling down (top right) (excitation at 365 nm). (Left) temperature dependent fluorescence emission spectra. (Middle) CIE 1931 chromaticity plot of the temperature dependent fluorescence emission spectra. For the table of emission color coordinates, see ESI Fig. S42. (Right) temperature dependence of the maximum, the AIE-maximum (462 nm) and the wavelength of the maximum of the gel above.

Fluorescence measurements

To quantify these observations, we initially measured the temperature dependent fluorescence emission spectra of the water/acetonitrile gel of **1** (10 mg/mL) with added meglumine (Fig. 5). With increasing temperature, the fluorescence emission intensity decreased and its maximum became more redshifted. Above 70 °C, the red-shifting of the maximum accelerated noticeably with higher temperatures and the fluorescence emission spectrum converged to almost the same spectrum as that of **1** dissolved in water/THF, indicating solution. The temperature dependent behavior of the gel was also studied (Fig. 5, ESI Fig. S46). It melted completely after 10 min at 90 °C, beginning very slowly at 80 - 85 °C, although a noticeable shift in fluorescence emission was already visible at 70 °C (ESI Fig. S44). Upon cooling down, the solution quickly formed a gel after the fluorescence emission had changed back to blue, which indicated aggregation (ESI Fig. S45). This shows that the gel structure remains stable for a short time even after almost complete disaggregation of the **DPhCzT** moieties, as shown by the fluorescence spectra. It further implies that the gel network is mainly stabilized by the hydrogen-bonding of the DKP ring systems. To rule out that the change in fluorescence emission is only an effect of

temperature, the experiment was repeated with the GdL gel of **1** (1 wt% in H₂O). For this gel, gelation was not reversible by heating and it showed a decrease of fluorescence intensity with increasing temperature, but no shift of the fluorescence emission (ESI Fig. S47-48). In this case, the gel is possibly mainly stabilized by the hydrophobic interactions of the **DPhCzT** moieties, which are driven to aggregate by the aqueous environment. Both results are in good alignment with findings by MAKAREVIĆ *et al.*, who found that gels of bis(amino acid) oxalyl amides were stabilized mainly by lipophilic interactions in aqueous environments and by intermolecular hydrogen bonding in organic solvents.^[46]

The effect of aggregation on the fluorescence emission could also be seen for water/THF solutions of **1**. Adding GdL to a filtered solution of **1** (6.7 mg/mL) in 33% THF and 67% water or **1** (6.0 mg/mL) in a 40% THF and 60% water led to a change in fluorescence emission from slightly greenish/yellow to blue within seconds and rose well until after a gel was formed. Through this effect, the gel formation could be tracked *via* monitoring the fluorescence emission intensity of the aggregation-induced maximum over time, directly after the addition of GdL (Fig. 6).

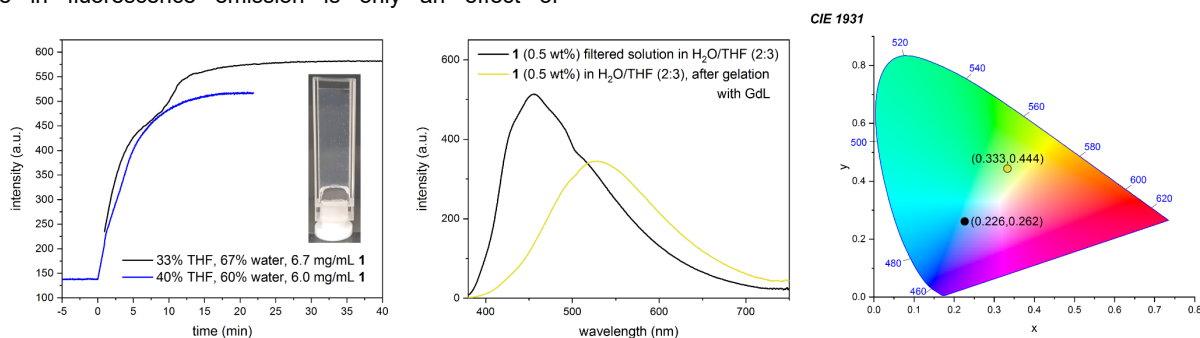


Figure 6. (Left) evolution of the fluorescence emission intensity of the aggregation maximum of filtered water/THF solutions of **1** after addition of GdL (12 equiv, at $t = 0$), and picture showing the resulting gel 1 h after addition of GdL (40/60% mixture, ambient light). (Middle) fluorescence emission spectrum of the 40/60% mixture before and 1 hour after the addition of GdL, showing AIBSE. (Right) CIE 1931 chromaticity plot of the spectra. Excitation at 365 nm.

The form of the observed curve for the 33/67% mixture is in excellent agreement with results from CHEN *et al.*, who investigated the self-assembly mechanism for a naphthalene-dipeptide hydrogelator. The mixture shows the same two plateaus that were found during the hydrogelation of a naphthalene-dipeptide hydrogelator with GdL, whereas the first plateau corresponded to a drop of the pH of the mixture to that of the pK_a of the carboxylic acid group of the peptide.^[47] In their work, the authors used thioflavin T (ThT) to track the aggregation of the peptides and the gelation process *via* the fluorescence emission of ThT, which was incorporated into the gel during gelation. By incorporation into the gel, it showed an increase in fluorescence intensity due to restricted intramolecular rotation (RIR).^[48] Furthermore, the AIBSE phenomenon has been shown to be caused by RIR in a theoretical study.^[49] This implies that the observed AIBSE in our case is caused by RIR of the **DPhCzT** moieties, giving direct experimental evidence for the theoretical study.

The results also show that even a very slight change in solvent composition can cause huge differences in the self-assembly process, as the 40/60% mixture aggregates too fast for the plateau to really show despite a slightly lower concentration of **1**. The resulting gels were only stable for two days, after which they both degraded to a solution containing a voluminous precipitate.

When the ratio of THF to water was further raised to 50:50, no gel formed. Instead, addition of GdL to this solution led to precipitate formation shortly after (Fig. 7). Addition of 12 equivalents of GdL resulted in a very fast increase of fluorescence emission intensity at 456 nm within 5 min, followed by a sharp decline to a lower intensity than before the addition due to the scattering of light by the precipitate.

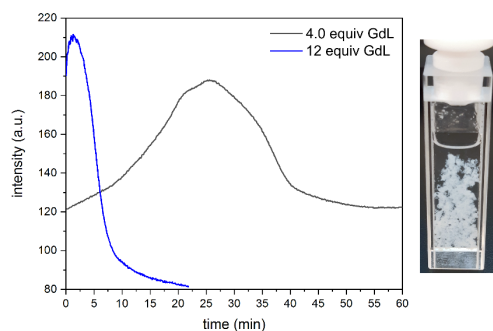


Figure 7. Evolution of the fluorescence emission intensity of the aggregation maximum (456 nm, excitation at 365 nm) of a filtered water/THF (1:1) solution of **1** (5 mg/mL) after addition of GdL, and picture (ambient light) showing the resulting suspension 1 h after addition of 12 equivalents GdL.

The addition of 4.0 equivalents of GdL instead resulted in a much slower increase in fluorescence emission intensity, but led to the same result. The at first accelerating increase was significantly slowed down 20 min after the addition and declined significantly between 25 min and 40 min to a level similar to that before the addition, which was also due to the formation of light-scattering precipitate. The explanation for the drastic effect of the THF content on the gelation process can

be rationalized by the better solvation of the **DPhCzT** moieties with a higher THF content. If the content is too high, the gel state, which presents a kinetic trap for aggregation, can be overcome and the thermodynamic minimum (precipitation) reached.^[50]

To further investigate the aggregation of the **DPhCzT** moieties, fluorescence lifetimes of the solids and some of the gels were measured. **1** showed an average fluorescence lifetime of $\tau_{avg} = (6.30 \pm 0.16)$ ns as a solid, a slightly shorter lifetime of $\tau_{avg} = (5.93 \pm 0.13)$ ns as a sol in water and a significantly shorter lifetime of $\tau_{avg} = (4.37 \pm 0.14)$ ns as 17 h old GdL gel (Fig. 8). For **2**, the same reduction in fluorescence lifetime was observed after gelation as for **1** (Table 3, ESI Fig. S53).

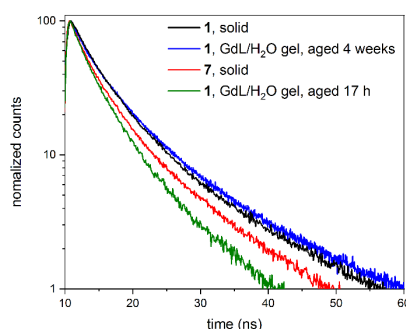


Figure 8. Comparison of fluorescence lifetime decay profiles (excitation at 373 nm). Comparison of GdL gels of **1** to the solids **1** and **7**. For an enlarged picture see ESI Fig. S51.

As found before by hydrogelation experiments, the GdL gel of **1** should consist mainly of the free acid **7** since substoichiometric amounts of GdL did not cause gelation. However, after aging for four weeks, the lifetime of the GdL gel increased to $\tau_{avg} = (6.61 \pm 0.08)$ ns, which is longer than that of **7** ($\tau_{avg} = (5.46 \pm 0.17)$ ns) and very similar to solid **1**, indicating a similar microenvironment. The at first significantly decreased lifetime after gelation can be rationalized by a competing aggregation of the **DPhCzT** moieties on the one hand and the DKP ring systems on the other hand. First, the gel structure forms through hydrogen bonding of the DKP ring systems, leaving aggregates of the **DPhCzT** moieties out of equilibrium compared to the sol. Over time, the gel structure reorganizes to a local kinetic minimum at which aggregation interactions are maximized, leading to conformationally stabilized excited states, explaining the longer lifetime of the aged gel. The water/acetonitrile gel of **1** with added meglumine, in contrast, showed a slightly increased fluorescence lifetime 17 h after gelation compared to the sol of **1** in water. A recent study by DEBNATH *et al.* has shown that π -stacking dominates hydrogen bonding at higher temperatures for the gelation of Fmoc-tyrosyl-leucine.^[51] In our case, this could be a possible explanation for the increased lifetime, as dominating π -stacking would then lead to initially conformationally stabilized excited states. A second explanation for the increased lifetimes of the aged GdL gel and the water/acetonitrile + meglumine gel of **1** might be a partial displacement of oxygen from the gel structure. The effect of oxygen on the fluorescence lifetime of

1 was shown by deaerating the aqueous sol of **1**, which led to an almost doubling of the lifetime (Table 3). A partial displacement of oxygen could take place through the formation of hydrophobic domains within the gel after gelation, and the effect increased by further rearrangement of the **DPhCzT** moieties after gelation as described above.

Table 3: Overview of average fluorescence lifetimes of select compounds and formulations (excitation at 373 nm). For detailed formulations, see ESI chapter 10.1.

| Compound | Entry | $\lambda_{\text{max, emission}}$ | $\tau_{\text{avg}}/\text{ns}$ |
|----------|---|----------------------------------|-------------------------------|
| 1 | Solid | 465 nm | 6.30 ± 0.16 |
| 2 | Solid | 466 nm | 5.54 ± 0.13 |
| 7 | Solid | 463 nm | 5.46 ± 0.17 |
| 1 | Solution in THF/H ₂ O (2/3) (6.0 mg/mL) | 527 nm | 4.67 ± 0.35 |
| 1 | H ₂ O sol (1 wt%) | 466 nm | 5.93 ± 0.13 |
| 1 | H ₂ O sol, deaerated (1 wt%) | n.d. | 10.3 ± 0.1 |
| 1 | GdL/H ₂ O gel 17 h old (1 wt%) | 462 nm | 4.37 ± 0.14 |
| 1 | GdL/H ₂ O gel 4 weeks old (1 wt%) | n.d. | 6.61 ± 0.08 |
| 1 | H ₂ O/MeCN + meglumin gel (10 mg/mL) | 462 nm | 6.45 ± 0.24 |
| 2 | H ₂ O/HFIP + TFA gel 17 h old (10 mg/mL) | 445 nm | 4.37 ± 0.11 |

Material properties

Selected gels and the sol of **1** in water were further analyzed for their microstructures by performing scanning electron microscopy (SEM) after freeze-drying (Fig. 9). The freeze-dried sol of **1** showed a highly porous network of thin ribbons, as an effect of its aggregation and possible preorganization in water before freeze-drying (Fig. 9A) The ribbons were approx. 60 nm - 200 nm thick (with most around ~ 100 nm). The *n*Bu₄NBF₄ gel of the same sol in comparison shows significantly thicker ribbons (approx. 100 nm - 400 nm) with visible fibers, cross-linked to a gel-network and demonstrating the difference between the hydrogel and the viscous sol (Fig. 9B). In stark contrast, the GdL/water gel of **1** is characterized by very long lamellar sheets that are approx. 0.1 μm - 1 μm thick (average around 0.4 μm , Fig. 9C). The water/acetonitrile + meglumine gel of **1** and the water/HFIP + TFA gel of **2** both consisted of a network of much thicker fibers, with a thickness above 1 μm and up to 4 μm for the gel of **2**, and up to 10 μm for the gel of **1** (Figures 9D and 9E).

Rheological experiments were performed on the GdL/water gel of **1**, the water/acetonitrile + meglumine gel of **1** and the water/HFIP + TFA gel of **2** (Fig. 10).

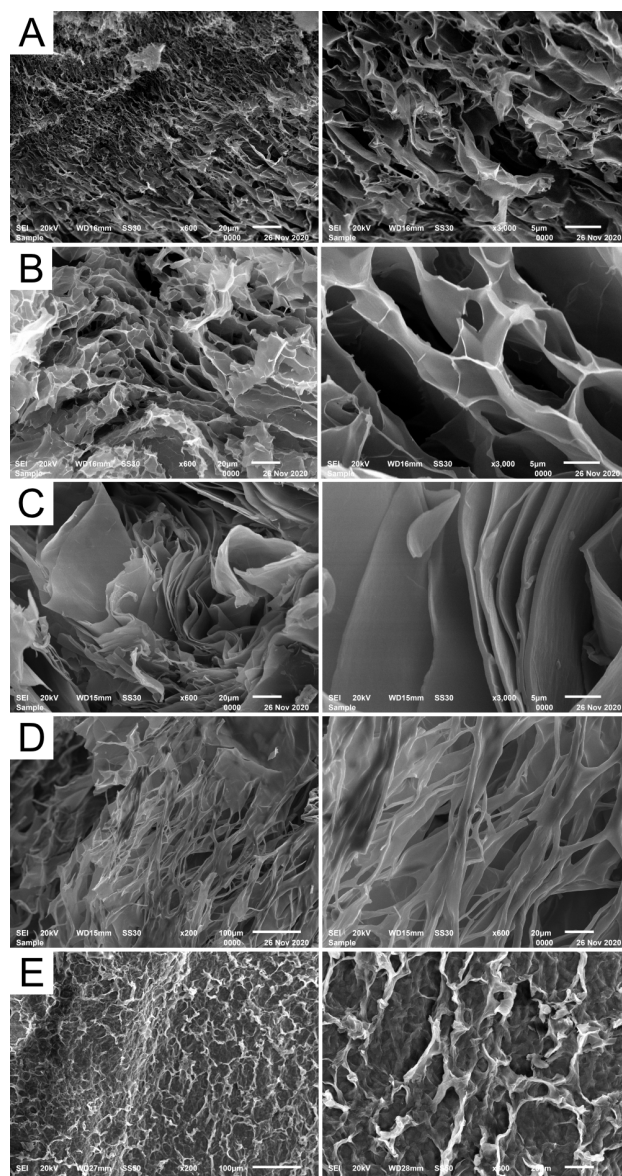


Figure 9. SEM images of select xerogels of **1** and **2**. A) Freeze dried sol of **1** in water (1 wt%); B) *n*Bu₄NBF₄ (3.0 equiv) gel of **1** (1 wt%) in water, aged four weeks; C) GdL gel of **1** (1 wt%, 12 equiv. GdL), aged 7 d; D) water/acetonitrile (1:1) gel of **1** (10 mg/mL) with additional meglumine (10 mg/mL), aged 7 d; E) water/HFIP (1:1) + TFA (1%) gel of **2** (10 mg/mL), aged 7 d.

Despite showing gel formation by tube-inversion already 30 min after addition of GdL, the gel was too unstable to be transferred to the rheometer even after 17 h. We see this as an indication of a direct link between the gel strength and the relative fluorescence lifetime of the gel over time. Amplitude sweep experiments showed a mechanical stability for all gels, aged one week, up to 0.1% shear strain. The experiments confirmed that all gels were physical hydrogels and not viscoelastic fluids, since the storage modulus G' was in all cases about one order of magnitude higher than the loss modulus G'' .

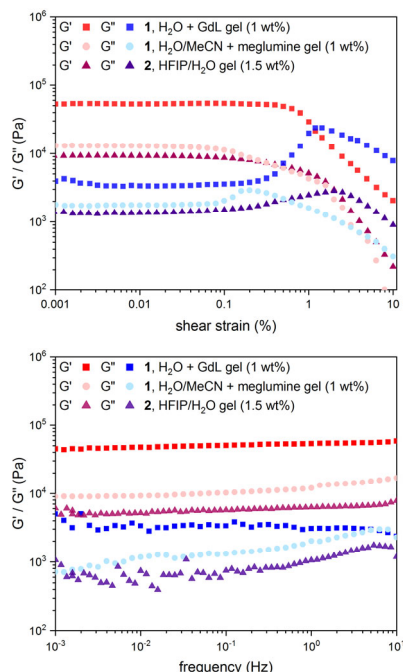


Figure 10. Comparison of frequency sweeps of the GdL (12 equiv) gel of **1** (1.5 wt%), the water/acetonitrile (1:1) + meglumine (10 mg/mL) gel of **1** (10 mg/mL) and of the water/HFIP (1:1) + TFA (1%) gel of **2** (15 mg/mL). All gels aged 7 days prior to the measurement. Single pictures are also in the ESI, Fig. S62-64.

Frequency sweep experiments further showed only a weak dependency of G' and G'' on frequency, which are both in a range common for DKPs with aromatic residues.^[17,21,30,31]

Conclusion

In conclusion, our work shows that efficient hydrogelation is possible for minimalistic, DKP based hydrogelators even if they contain a challengingly insoluble AIEgen. Although the heterocyclic compound **DPhCzT** made up 51.1% of the mass of hydrogelator **1** and 57.8% of the mass of gelator **2**, efficient hydrogelation by many triggers was possible for **1** with a CGC down to 0.3 wt% (3.9 mM), while a wide variety of organogels could be formed by **2** at a concentration of 1 wt% or below. Aggregation properties were investigated *via* different fluorescence techniques as well as measurements of basic material properties. The results show the importance of solvent choice, additives and temperature for aggregation properties and therefore gel properties. Moreover, the results illustrate the delicate balance needed between hydrophilic and hydrophobic aggregation interactions to avoid competing interactions, which can result in lower gel stability or precipitation. We believe our work demonstrates the usefulness of the minimalistic, modular system consisting of a DKP and an AIEgen for the synthesis and study of efficient hydrogelators containing challenging hydrophobic structure motifs. The system allows for quick and manifold variations in structure for fine-tuning gel properties through the choice of readily available amino acid side-chains or counter-ions. Further

modifications of the structure are possible through functionalization of the peptide side chains.

Keywords

fluorescence • gels • peptides • sol-gel-process • supramolecular chemistry

Experimental Section

For full experimental procedures as well as spectroscopic and analytical data for all new compounds including copies of NMR spectra, see the ESI.

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