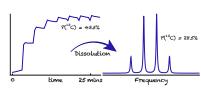
Boosting Dissolution-Dynamic Nuclear Polarization by Multiple-Step Dipolar Order Mediated ¹H→¹³C Cross-Polarization

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Abstract: Dissolution-dynamic nuclear polarization can be boosted by employing multiplecontact cross-polarization techniques to transfer polarization from ¹H to ¹³C spins. The method is efficient and significantly reduces polarization build-up times, however, it involves high-power radiofrequency pulses in a superfluid helium environment which limit its implementation and applicability and prevent a significant scaling-up of the sample size. We propose to overcome this limitation by a stepwise transfer of polarization using a low-



energy and low-peak power radiofrequency pulse sequence where the ${}^{1}H\rightarrow{}^{13}C$ polarization transfer is mediated by a dipolar spin order reservoir. An experimental demonstration is presented for $[1-{}^{13}C]$ sodium acetate. A solid-state ${}^{13}C$ polarization of ~43.5% was achieved using this method with a build-up time constant of ~5.1 minutes, leading to a ~28.5% ${}^{13}C$ polarization in the liquid-state after sample dissolution. The low-power multiple-step polarization transfer efficiency with respect to the most advanced and highest-power multiple-contact cross-polarization approach was found to be ~0.69.

Keywords: NMR, Hyperpolarization, DNP, dDNP, CP, dCP, dipolar order, dissolution

Ordinary magnetic resonance spectroscopy (MRS) and imaging (MRI) methods are often limited by the weak magnetic response from clusters of nuclear spins, even when placed within today's highest field superconducting magnets. The inherent insensitivity of a nuclear spin ensemble is engendered by the small differences in energy between nuclear spin states compared with the energy typically available at room temperature, which results in a rather flat Boltzmann distribution of nuclear spin populations.

To alleviate this issue, dissolution-dynamic nuclear polarization (*d*DNP) experiments are becoming increasingly employed. The hyperpolarization technique generates strong nuclear magnetic resonance (NMR) signals enhanced by factors approaching 10^4 [1] for a range of nuclear spins in various media, with applications in clinical research [2-4] and metabolomics studies [5], among others [6,7]. For dilute low- γ nuclear spins at low temperatures, the *d*DNP process [8] suffers from excessively long polarization build-up time constants τ_{DNP} sometimes exceeding an hour [9].

dDNP methods can be efficiently accelerated (by a factor of up to 40) by the implementation of radiofrequency (rf) pulse sequences such as cross-polarization (CP) [10-17], which indirectly transfer electron spin polarization to insensitive nuclear spins (such as ¹³C) via sensitive nuclear spins (such as ¹H). The acceleration of the DNP process, compared with direct polarization, is attributed to the generally quicker polarization build-up timescales of proton spins at low temperatures when polarized with nitroxide radicals [18]. Multiple applications of intense B_1 -matched (typ. > 15 kHz) simultaneous ¹H and ¹³C spin-locking rf-fields throughout an optimized contact period (typ. > 1 ms) allow the repeated indirect transfer of electron spin polarization, which is accumulated by the insensitive nuclear spins. This multiple-contact CP-DNP approach was implemented in the preparation and transportation of highly polarized metabolites [19].

Such a CP approach under *d*DNP conditions, *i.e.* at liquid helium temperatures, is significantly challenging, *e.g.* necessary high-energy and high-peak power *rf*-pulses can lead to detrimental arcing in the superfluid cryogenic bath [20]. As a result, CP is not widely implemented under *d*DNP conditions (typ. T = 1.0-1.6 K). Such difficulties not only restrict a broader implementation of CP, but also prevent the scaling-up of *d*DNP sample volumes required for human applications or for the parallelization of hyperpolarization [21].

We have recently demonstrated the use in a DNP context of an alternative *rf*-pulse sequence to CP which is of low-power, does not require synchronized B_1 -matched spin-locking *rf*-fields and can ultimately overcome all previous limitations [22]. In such cases, the transfer of spin polarization is mediated by an intermediary reservoir of nuclear dipolar spin order [23-33], and even though the exact mechanism underlying the polarization transfer is yet to be fully understood, the *rf*-pulse sequence has consequently been termed dipolar order mediated crosspolarization (*d*CP). It is expected that significant levels of ¹³C polarization can be accrued if an approach incorporating consecutive *d*CP transfers can be successfully implemented, and that a significant fraction of the resulting solid-state ¹³C polarization will be preserved upon hyperpolarization by dissolution to the liquid-state.

In the current Paper, we present an *rf*-pulse sequence containing multiple low-power polarization transfer elements in a way that is fully compatible with *d*DNP conditions. The *rf*-pulse sequence yielded a ¹³C polarization of ~43.5% with a build-up time constant $\tau_{dCP} = 5.1 \pm 0.2$ minutes for a sample of $[1^{-13}C]$ sodium acetate in the frozen solid-state, which was found to be ~0.69 of the efficiency realized by using a most advanced state-of-the-art and fully optimized multiple-contact

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CP experiment. We additionally investigated the dissolution of $[1-{}^{13}C]$ sodium acetate indirectly polarized via this technique, and achieved a liquid-state ${}^{13}C$ polarization level of ~28.5%.

Sample I (see experimental methods) was polarized by applying microwave irradiation at $f_{\mu\nu}$ = 198.128 GHz (negative lobe of the EPR line) with triangular frequency modulation of amplitude $\Delta f_{\mu w} = \pm 160 \text{ MHz} [34]$ and rate $f_{mod} = 0.5 \text{ kHz}$ at a power of ca. $P_{\mu w} = 125$ mW at the output of the microwave source and ca. $P_{\mu w}$ = 30 mW reaching the DNP cavity. Microwave gating was employed shortly before and during dDNP transfer elements to allow the electron spin ensemble to return to a highly polarized state, which happens on the timescale of the longitudinal electron relaxation time (typ. T_{1e} = 100 ms with $P_e = 99.93\%$ under our conditions) [35]. Microwave gating provides a way to strongly attenuate paramagnetic relaxation, therefore resulting in a significant increase in the nuclear spin relaxation times in the rf-field (or rotating) frame. This allows the application of longer dCP rfpulses, which significantly increases the efficiency of nuclear spin polarization transfer.

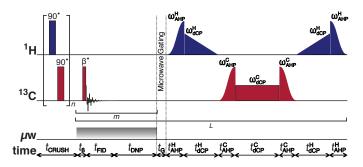


Figure 1: Schematic representation of the multiple-step dipolar order mediated cross-polarization (*multi-dCP*) *rf*-pulse sequence used for transferring ¹H Zeeman polarization to insensitive ¹³C nuclear spins in successive steps. The experiments used the following parameters, chosen to maximize ¹H \rightarrow ¹³C polarization transfer: n = 50; $\beta = 5^{\circ}$; $\tau_{\text{DNP}} = 30$ s; m = 7; $\tau_{\text{G}} = 0.5$ s; $\omega_{\text{AHP}}^{\text{H}}/2\pi = 27.8$ kHz; $t_{\text{AHP}}^{\text{H}} = 175$ μ s; $\omega_{\text{dCP}}^{\text{H}}/2\pi = 16.9$ kHz; $t_{\text{dCP}}^{\text{H}} = 450 \ \mu$ s; $\omega_{\text{CHP}}^{\text{C}}/2\pi = 14.6$ kHz; $t_{\text{dCP}}^{\text{C}} = 49$ ms; L = 8. AHP = Adiabatic Half-Passage. AHP sweep width = 100 kHz. All AHP and *dCP rf*-pulses have phase *x*. The $\pi/2$ crusher *rf*-pulses use a thirteen-step phase cycle to remove residual magnetization at the beginning of the experiment: {0, $\pi/18$, $5\pi/18$, $\pi/2$, $4\pi/9$, $5\pi/18$, $8\pi/9$, π , $10\pi/9$, $13\pi/9$, $\pi/18$, $5\pi/3$, $35\pi/18$ }. The resonance offset was placed at the centre of the ¹H and ¹³C NMR peaks.

To obtain the highest possible levels of 13 C polarization, it is of interest to perform multiple *d*CP *rf*-pulse sequence cycles to incrementally transfer ¹H polarization to dilute ¹³C nuclear spins embedded within the sample. Figure 1 shows such a suitable *rf*-pulse sequence capable of implementing numerous *d*CP *rf*-pulse sequence steps. Consequently, the *rf*-pulse sequence has been termed multiple-step dipolar order mediated cross-polarization (*multi-d*CP).

The *multi-d*CP *rf*-pulse sequence operates as follows:

(*i*) A crusher sequence of 90° *rf*-pulses with alternating phases separated by a short delay (typ. 11 ms) repeated *n* times (typ. n = 50) kills residual magnetization on both *rf*-channels;

(ii) The microwave source becomes active;

(*iii*) The ¹³C Zeeman magnetization trajectory is minimally perturbed by the application of a small flip-angle *rf*-pulse (typ. $\beta = 5^{\circ}$) with a short acquisition period (typ. $t_{FID} = 1$ ms) used for detection;

(*iv*) ¹H DNP builds-up during a time t_{DNP} (typ. $t_{DNP} = 30$ s);

(v) Stages *iii-iv* are cycled *m* times (typ. m = 7) in order to monitor the evolution of the ¹³C polarization between *d*CP steps;

(*vi*) The microwave source is gated and a delay of duration $t_G = 0.5$ s occurs before the next *d*CP step, thus permitting the electron spins to relax to their highly polarized thermal equilibrium state [35];

(*vii*) A ¹H adiabatic half-passage (AHP) *rf*-pulse followed by a linearly decreasing amplitude ramp ¹H *rf*-pulse of amplitude ω_{dCP}^{H} and length t_{dCP}^{H} presumably converts ¹H Zeeman polarization into ¹H-¹H dipolar order [36];

(*viii*) A ¹³C spin-locking *rf*-pulse of amplitude ω_{dCP}^{C} and length t_{dCP}^{C} sandwiched between two ¹³C AHP *rf*-pulses of opposite chronology generates ¹³C transverse magnetization;

(*ix*) Stages *ii-viii* are repeated in L units (typ. L = 8) to periodically transfer ¹H Zeeman polarization to ¹³C spins.

The DNP build-up time constant of ¹H polarization for sample I at 1.2 K was measured to be: $\tau_{\text{DNP}} = 225 \pm 1$ s (for the positive lobe of the DNP microwave spectrum). Consequently, the period between *d*CP and CP polarization transfer steps was chosen to be: $t_{\text{DNP}} = 210$ s, corresponding to m = 7, see Figure 1.

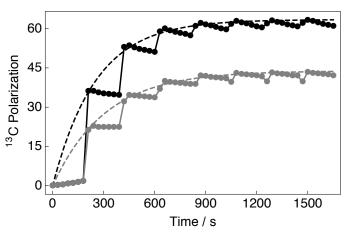


Figure 2: Experimental ¹³C polarization build-up curves for a sample of I acquired at 7.05 T (¹H nuclear Larmor frequency = 300.13 MHz, ¹³C nuclear Larmor frequency = 75.47 MHz) and 1.2 K with a single transient per data point. The build-up of ¹³C polarization was measured by using: Black curve: A state-of-the-art and high-power CP *rf*-pulse sequence described in the Supporting Information (SI); and Grey curve: The low-power *multi-dCP rf*-pulse sequence described in Figure 1. The traces have the same overall form, and plateau over a period of ~1500 s. The *d*CP and CP build-up curves were fitted with a mono-exponential build-up function A(1-exp{-t/ τ }) using the build-up time constants $\tau = \tau_{CP}$ and $\tau = \tau_{dCP}$, respectively. Build-up time constants: Black dashed curve (CP): $\tau_{CP} = 4.2 \pm 0.2$ minutes; Grey dashed curve (*d*CP): $\tau_{dCP} = 5.1 \pm 0.2$ minutes.

The ¹³C nuclear spin polarization level achieved by the *multi-d*CP *rf*-pulse sequence is ~32.2% after 7 minutes (2 transfer steps) and ultimately reaches ~43.5% with a build-up time constant $\tau_{dCP} = 5.1 \pm 0.2$ minutes, see Figure 2 (grey curve). The very first ¹H \rightarrow ¹³C nuclear polarization transfer step alone achieves a ¹³C polarization of ~21.2% after only 3.5 minutes. A multiple-step CP *rf*-pulse sequence (see the Supporting Information (SI) for more details) obtains a maximum ¹³C polarization level of ~63.3% under the same experimental conditions (black curve). The overall performance of the *multi-d*CP *rf*-pulse sequence compared with a sophisticated and high-power multiple-contact CP *rf*-pulse sequence is ~0.69, which is determined from integrals of the multiple-contact CP and *multi-d*CP ¹³C NMR signal maxima. The grey curve in Figure 2 also

shows a small decrease in ¹³C polarization following each transfer step. ¹³C NMR spectra acquired around these regions are shown in the SI.

Polarized samples of **I** were dissolved with 5 mL of D₂O solvent prepressurized at 6 bar, and subsequently heated to 180°C (and a pressure of 9 bar). The liquid sample was transferred in 10 s to a *Bruker Biospin* prototype *d*DNP injector placed in the bore of a 1.88 T (¹H nuclear Larmor frequency = 80.05 MHz, ¹³C nuclear Larmor frequency = 20.13 MHz) permanent magnet *Bruker Biospin Fourier 80* benchtop NMR system by pushing with helium gas at 6 bar through a PTFE tube (1.6 mm inner diameter) running inside a series of solenoid coils (2 A power source) producing a minimum magnetic field of 4 mT along the entire sample transfer path (~2.8 m length), see Figure 3a [37]. Experimental liquid-state ¹H and ¹³C NMR data were recorded and processed using *TopSpin* 4.0 software.

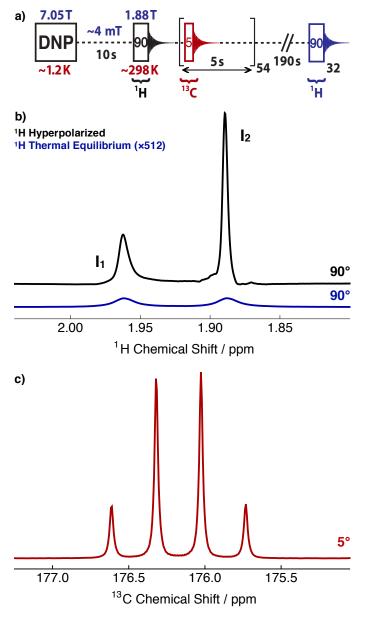


Figure 3: (a) Events and timings for DNP, sample dissolution and acquisition of liquid-state NMR spectra. Relevant portions of the experimental (b) ¹H and (c) ¹³C NMR spectra belonging to the methyl (CH₃) group and ¹³C-labelled carbonyl site, respectively, of I in approximately 0.6 mL of D₂O solution acquired at 1.88 T (¹H nuclear Larmor frequency = 80.05 MHz, ¹³C nuclear Larmor frequency = 20.13 MHz) and 298.15 K. The experimental NMR spectra were acquired in accordance with the events and timings depicted in (a). I₁ and I₂ refer to the ¹H NMR signal integrals of the multiplet lineshape components.

The first experimental 13 C NMR spectrum of the small flip-angle *rf*-pulse and acquire train in (a) is shown in (c).

The sequence of events given in Figure 3a details how the experimental NMR spectra were acquired. After DNP and sample dissolution and transfer, a 90° *rf*-pulse is used to record a hyperpolarized ¹H NMR spectrum (black). Subsequently, a train of 5° *rf*-pulses separated by 5 s is used to record the hyperpolarized ¹³C NMR spectra (red). The sample is allowed to rest in the 1.88 T magnet for an additional 190 s in order to achieve thermal equilibrium, and a proton NMR signal is acquired using a 90° *rf*-pulse (blue).

The black spectrum in Figure 3b shows the relevant region of the experimental ¹H NMR spectrum of **I** acquired immediately after sample dissolution and transfer. The sample was initially hyperpolarized in the solid-state by using the *multi-d*CP *rf*-pulse sequence. The methyl (CH₃) group resonance located at ~1.93ppm is split into two peaks due to a scalar coupling with the ¹³C-labelled carbonyl site (scalar coupling constant: $|^2J_{\text{HC}}| \simeq 5.8$ Hz).

The proton NMR lineshape in Figure 3b is highly asymmetric, and indicates that the scalar coupled ¹³C nuclear spins within the sample are significantly hyperpolarized. The degree of multiplet asymmetry can be used to infer the ¹³C polarization of the sample upon arrival inside the receiving magnet, a methodology known as SPY-MR [38]. The ¹³C polarization level $P(^{13}C)$ is given by the following expression:

$$P(^{13}C) = \frac{I_1 - I_2}{I_1 + I_2} \times 100\% (1)$$

where I₁ and I₂ are the ¹³C NMR signal integrals of the most and least intense multiplet peaks, respectively. In the case of the ¹H NMR spectrum presented in Figure 3b, by using Equation 1 it can be deduced that the ¹³C polarization $P(^{13}C)$ is ~28.5%. The hyperpolarized ¹³C NMR spectrum (Figure 3c) was also compared indirectly to the thermal equilibrium ¹H NMR spectrum of **I** (Figure 3b, blue spectrum), with a 0.2% agreement on the final ¹³C polarization.

The ¹³C NMR spectrum acquired immediately after sample dissolution and transfer is shown in Figure 3c. The ¹³C carbonyl resonance of **I** is positioned at ~176.2ppm and displays a 1:3:3:1 quartet structure due to the $|^2J_{\rm HC}|$ scalar coupling with the 3 methyl group protons. The signal-to-noise ratio (SNR) of the NMR peak belonging to the ¹³C-labelled site was determined to be ~1905.

The longitudinal relaxation time constant T_1 of the ¹³Clabelled carbonyl site in **I** was measured by applying a small flip-angle *rf*-pulse (5°) followed directly by ¹³C NMR signal detection (acquisition time = 3 s) every 5 s, see Figure 3a. The resulting curve was found to have a single exponential relaxation behaviour (data not shown), and is well fitted with a mono-exponential decay function using a sole relaxation time constant T_1 . Mono-exponential decay function: Aexp{-t/ T_1 }. The value of T_1 was determined to be: 99.0 ± 0.7 s, taking into account the influence of the small flip-angle *rf*-pulse train.

The relevant portion of the experimental ¹H NMR spectrum belonging to the methyl (CH₃) group protons of I acquired under thermal equilibrium conditions is also shown in Figure 3b (blue spectrum). This spectrum allows an estimate of the resulting liquid-state sample concentration for I after dissolution and transfer by comparing the ¹H NMR signal integral to that from a sample of I at a known concentration. The solution-state sample concentration of I was consequently found to be: ~ 14.7 mM.

In the solid-state, the multiple-contact CP *rf*-pulse sequence is clearly superior in terms of ${}^{1}\text{H} \rightarrow {}^{13}\text{C}$ polarization transfer and achieves a higher final ¹³C polarization before sample dissolution. A suspected reason for the lower efficiency of dCP polarization transfer is the quantity of ¹H polarization depleted to complete the dCP transfer step compared with the CP rf-pulse sequence. Proton NMR spectra showing this effect are given in the SI. It was found that a CP contact retains ~82.9% of the initial ¹H polarization, whereas the dCP rf-pulse sequence retains only ~19.8%. Consequently, there is less ¹H polarization available at subsequent polarization transfer steps. This result very likely inhibits the efficiency of the remaining polarization transfer stages in the dCP rf-pulse sequence. This effect should be investigated in the future, and the inter-dCP delays further optimized (and likely lengthened) to ultimately increase the overall dCP efficiency, since the current inter-dCP delays were intentionally matched to those of a fully optimized CP experiment. Liquid-state 13C polarizations on the order of ~40% have previously been demonstrated for [1-¹³C]sodium acetate by using a multiple-contact CP rf-pulse sequence prior to sample dissolution [39,40]. Nevertheless, a liquid-state ¹³C polarization of ~28.5% is encouraging given the initial solidstate ¹³C polarization of ~43.5%.

SPY-MR polarimetry [38] in liquid-state NMR works in the case that the nuclear spins involved are not participating in strong coupling (which cannot be the case for heteronuclear spins at sufficiently high magnetic fields). As a result; the SPY-MR approach was implemented to infer the level of ¹³C polarization from the hyperpolarized ¹H NMR spectrum in the liquid-state after dissolution. This is a feasible approach since the lower sensitivity of our benchtop magnet, with respect to higher magnetic field superconducting instruments, requires very long (on the order of days) accumulations of ¹³C thermal equilibrium spectra in order to obtain an NMR signal with a sufficient signal-to-noise ratio (SNR).

There is evidentially a discrepancy between solid-state and liquid-state ¹³C polarizations. Given the long T_1 of sample I at low magnetic field, it is unlikely that solely ¹³C nuclear spin relaxation is responsible for the difference in results. Another possibility is that ¹H-¹³C dipolar order is generated and survives the dissolution process and is not eradicated by changing magnetic field gradients during the sample transfer step to the detection magnet. The presence of such dipolar order would limit the applicability of the SPY-MR approach [38]. The excessive losses in ¹³C polarization are not accounted for at present and are largely thought to be related to the sample dissolution and transfer processes but also could be attributable to zero or double quantum coherences created in regions of low magnetic field and incoherent cross-relaxation phenomena [41].

An *rf*-pulse sequence which employs low-power *rf*-pulses for the stepwise transfer of ¹H polarization to ¹³C nuclear spins under *d*DNP conditions was demonstrated. The *multi-d*CP *rf*sequence achieves a ¹³C polarization level of ~43.5% in the solid-state after ~25 minutes (7 polarization transfer steps). The overall *d*CP polarization transfer efficiency of the approach was found to be ~0.69 with respect to a sophisticated and highpower multiple-contact CP experiment. After dissolution with a hot solvent, hyperpolarized liquid-state NMR signals were detected and a ¹³C polarization of ~28.5% was observed in a separate permanent magnet benchtop NMR system. These results are promising for future applications of indirect hyperpolarization techniques and dissolution of insensitive nuclear spins. The low-power nature of the *multi-d*CP approach may allow polarization transfer techniques to be implemented in larger sample volumes, paving the way to the use of indirect ${}^{1}\text{H} \rightarrow {}^{13}\text{C}$ polarization transfer schemes in (pre)clinical settings.

Experimental Methods

Sample Preparation. A solution of 3 M $[1^{-13}C]$ sodium acetate in the glass-forming mixture H₂O:D₂O:glycerol-*d*₈ (10%:30%:60% *v/v/v*) was doped with 30 mM TEMPOL radical (all compounds purchased from *Sigma Aldrich*) and sonicated for ~10 minutes. This sample is referred to as I in the main text. Paramagnetic TEMPOL radicals were chosen to most efficiently polarize ¹H spins under our *d*DNP conditions.

Sample Freezing. 100 μ L sample volumes were pipetted into a PEEK sample cup and inserted into a 7.05 T prototype Bruker Biospin polarizer equipped with a specialized *d*DNP probe and running *TopSpin 3.5* software. The sample temperature was reduced to 1.2 K by submerging the sample in liquid helium and reducing the pressure of the variable temperature insert (VTI) towards ~0.7 mbar.

Supporting Information

The Supporting Information is available free of charge. Multiple-contact CP *rf*-pulse sequence in Figure S1, ¹H detected NMR spectra after *d*CP and CP polarization transfer steps in Figure S2, and ¹³C detected NMR spectra around *d*CP polarization transfer steps in Figure S3.

Conflict of Interest

The authors declare no conflict of interest.

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References

J.-H. Ardenkjær-Larsen, B. Fridlund, A. Gram, G. Hansson, L. Hansson, M. H. Lerche, R. Servin, M. Thaning and K. Golman, Increase in signal-to-noise ratio of > 10,000 times in liquid-state NMR, *Proc. Natl. Acad. Sci. U.S.A.*, 2003, 100, 10158-10163.

[3] K. M. Brindle, S. E. Bohndiek, F. A. Gallagher and M. I. Kettunen, Tumor imaging using hyperpolarized ¹³C magnetic resonance spectroscopy, *Magn. Reson. Med.*, **2011**, 66, 505-519.

^[2] S. E. Day, M. I. Kettunen, F. A. Gallagher, D.-E. Hu, M. Lerche, J. Wolber, K. Golman, J. H. Ardenkjær-Larsen and K. M. Brindle, Detecting tumor response to treatment using hyperpolarized ¹³C magnetic resonance imaging and spectroscopy, *Nat. Med.*, **2007**, 13, 1382-1387.

[4] S. J. Nelson, J. Kurhanewicz, D. B. Vigneron, P. E. Z. Larson, A. L. Harzstark, M. Ferrone, M. van Criekinge, J. W. Chang, R. Bok, I. Park, G. Reed, L. Carvajal, E. J. Small, P. Munster, V. K. Weinberg, J. H. Ardenkjær-Larsen, A. P. Chen, R. E. Hurd, L.-I. Odegardstuen, F. J. Robb, J. Tropp and J. A. Murray, Metabolic imaging of patients with prostate cancer using hyperpolarized [1-¹³C]pyruvate, *Sci. Transl. Med.*, **2013**, 5, 198.

[5] A. Bornet, X. Ji, D. Mammoli, B. Vuichoud, J. Milani, G. Bodenhausen and S. Jannin, Long-Lived States of Magnetically Equivalent Spins Populated by Dissolution-DNP and Revealed by Enzymatic Reactions, *Chem.: Eur. J.*, **2014**, 20, 17113-17118.

[6] J.-N. Dumez, J. Milani, B. Vuichoud, A. Bornet, J. Lalande-Martin, I. Tea, M. Yon, M. Maucourt, C. Deborde, A. Moing, L. Frydman, G. Bodenhausen, S. Jannin and P. Giraudeau, Hyperpolarized NMR of plant and cancer cell extracts at natural abundance, *Analyst*, **2015**, 140, 5860-5863.

[7] A. Bornet, M. Maucourt, C. Deborde, D. Jacob, J. Milani, B. Vuichoud, X. Ji, J.-N. Dumez, A. Moing, G. Bodenhausen, S. Jannin and P. Giraudeau, Highly Repeatable Dissolution Dynamic Nuclear Polarization for Heteronuclear NMR Metabolomics, *Anal. Chem.*, **2016**, 88, 6179-6183.

[8] S. Jannin, J.-N. Dumez, P. Giraudeau and D. Kurzbach, Application and methodology of dissolution dynamic nuclear polarization in physical, chemical and biological contexts, *J. Magn. Reson.*, **2019**, 305, 41-50.

[9] S. Jannin, A. Bornet, S. Colombo and G. Bodenhausen, Low-temperature cross polarization in view of enhancing dissolution Dynamic Nuclear Polarization in NMR, *Chem. Phys. Lett.*, **2011**, 517, 234-236.

[10] S. R. Hartmann and E. L. Hahn, Nuclear Double Resonance in the Rotating Frame, *Phys. Rev.*, **1962**, 128, 204-205.

[11] A. Pines, M. Gibby and J. Waugh, Proton-enhanced nuclear induction spectroscopy ¹³C chemical shielding anisotropy in some organic solids, *Chem. Phys. Lett.*, **1972**, 15, 373-376.

[12] A. Bornet, R. Melzi, S. Jannin and G. Bodenhausen, Cross Polarization for Dissolution Dynamic Nuclear Polarization Experiments at Readily Accessible Temperatures 1.2 < T < 4.2 K, *Appl. Magn. Reson.*, **2012**, 43, 107-117.

[13] M. Batel, M. Krajewski, A. Däpp, A. Hunkeler, B. H. Meier, S. Kozerke and M. Ernst, Cross Polarization for Dissolution Dynamic Nuclear Polarization, *Chem. Phys. Lett.*, **2012**, 554, 72-76.

[14] A. Bornet, R. Melzi, A. J. Perez Linde, P. Hautle, B. van den Brandt, S. Jannin and G. Bodenhausen, Boosting Dissolution Dynamic Nuclear Polarization by Cross Polarization, *J. Chem. Phys. Lett.*, **2013**, 4, 111-114.

[15] B. Vuichoud, A. Bornet, F. de Nanteuil, J. Milani, E. Canet, X. Ji, P. Miéville, E. Weber, D. Kurzbach, A. Flamm, R. Konrat, A. D. Gossert, S. Jannin and G. Bodenhausen, Filterable Agents for Hyperpolarization of Water, Metabolites and Proteins, *Chem.: Eur. J.*, **2016**, 22, 14696-14700.

[16] M. Cavaillès, A. Bornet, X. Jaurand, B. Vuichoud, D. Baudouin, M. Baudin, L. Veyre, G. Bodenhausen, J.-N. Dumez, S. Jannin, C. Copéret and C. Thieuleux, Tailored Microstructured Hyperpolarizing Matrices for Optimal Magnetic Resonance Imaging, *Angew. Chem. Int. Ed.*, **2018**, 130, 7575-7579.
[17] A. J. Perez Linde, PhD thesis, University of Nottingham, UK, 2009.

[18] G. Hartmann, D. Hubert, S. Mango, C. C. Morehouse and K. Plog, Proton polarization in alcohols at 50 kG, 1 K, *Nucl. Instrum. Meth. A*, **1973**, 106, 9-12.
[19] X. Ji, A. Bornet, B. Vuichoud, J. Milani, D. Gajan, A. J. Rossini, L. Emsley, G. Bodenhausen and S. Jannin, Transportable Hyperpolarized Metabolites, *Nat. Commun.*, **2017**, 8, 13975.

[20] J. M. O. Vinther, V. Zhurbenko, M. M. Albannay and J.-H. Ardenkjær-Larsen, Design of a local quasi-distributed tuning and matching circuit for dissolution DNP cross polarization, *Solid State Nucl. Mag.*, **2019**, 102, 12-20.

[21] K. W. Lipsø, S. Bowen, O. Rybalko and J.-H. Ardenkjær-Larsen, Large dose hyperpolarized water with dissolution-DNP at high magnetic field, *J. Magn. Reson.*, **2017**, 274, 65-72.

[22] S. J. Elliott, S. F. Cousin, Q. Chappuis, O. Cala, M. Ceillier, A. Bornet and S. Jannin, Dipolar order mediated ${}^{1}H\rightarrow{}^{13}C$ cross-polarization for dissolution-dynamic nuclear polarization, *Magn. Reson.*, **2020**, 1, 89-96.

[23] J. Jeener and P. Broekaert, Nuclear Magnetic Resonance in Solids: Thermodynamic Effects of a Pair of rf Pulses, *Phys. Rev.*, **1967**, 157, 232-240.
[24] H.-M. Vieth and C. S. Yannoni, Cross polarization in solid state NMR spectroscopy. Efficient polarization transfer via the non-Zeeman spin reservoir, *Chem. Phys. Lett.*, **1993**, 205, 153-156.

[25] N. D. Kurur and G. Bodenhausen, Adiabatic Coherence Transfer in Magnetic Resonance of Homonuclear Scalar-Coupled Systems, *J. Magn. Reson. A*, **1995**, 114, 163-173.

[26] S. Emid, J. Konijnendijk, J. Smidt and A. Pines, On the short time behavior of the dipolar signal in relaxation measurements by the pulse method, *Physica* B+C, **1980**, 100, 215-218.

[27] S. Zhang, E. Stejskal, R. Fornes and X. Wu, Mismatching Cross Polarization in the Rotating Frame, *J. Magn. Reson., Ser A*, **1993**, 104, 177-179. [28] A. K. Khitrin, J. Xu and A. Ramamoorthy, Cross-correlations between low- γ nuclei in solids via a common dipolar bath, *J. Magn. Reson.*, **2011**, 212, 95-101.

[29] J. Jeener, R. Du Bois and P. Broekaert, "Zeeman" and "Dipolar" Spin Temperatures during a Strong rf Irradiation, *Phys. Rev.*, **1965**, 139, A1959-A1961.

[30] A. G. Redfield, Nuclear spin thermodynamics in the rotating frame, *Science*, **1969**, 164, 1015-1023.

[31] D. E. Demco, J. Tegenfeldt and J. S. Waugh, Dynamics of cross relaxation in nuclear magnetic double resonance, *Phys. Rev. B*, **1975**, 11, 4133-4151.

[32] M. Kunitomo, H. Hatanaka and T. Hashi, Adiabatic demagnetization in the rotating frame by non-resonant oscillating field, *Phys. Lett. A*, **1974**, 49, 135-136.

[33] J.-S. Lee and A. K. Khitrin, Thermodynamics of adiabatic cross polarization, *J. Chem. Phys.*, **2008**, 128, 114504.

[34] A. Bornet, J. Milani, B. Vuichoud, A. J. Perez Linde, G. Bodenhausen and S. Jannin, Microwave frequency modulation to enhance Dissolution Dynamic Nuclear Polarization, *Chem. Phys. Lett.*, **2014**, 602, 63-67.

[35] A. Bornet, A. Pinon, A. Jhajharia, M. Baudin, X. Ji, L. Emsley, G. Bodenhausen, J.-H. Ardenkjær-Larsen and S. Jannin, Microwave-gated dynamic nuclear polarization, *Phys. Chem. Chem. Phys.*, **2016**, 18, 30530-30535.

[36] S. J. Elliott, O. Cala, Q. Chappuis, S. F. Cousin, D. Eshchenko, R. Melzi, J. G. Kempf and S. Jannin, *Submitted*, **2021**.

[37] J. Milani, B. Vuichoud, A. Bornet, P. Miéville, R. Mottier, S. Jannin and G. Bodenhausen, A magnetic tunnel to shelter hyperpolarized fluids, *Rev. Sci. Instrum.*, **2015**, 80, 024101.

[38] B. Vuichoud, J. Milani, Q. Chappuis, A. Bornet, G. Bodenhausen and S. Jannin, Measuring absolute spin polarization in dissolution-DNP by Spin PolarimetrY Magnetic Resonance (SPY-MR), *J. Magn. Reson.*, **2015**, 260, 127-135.

[39] B. Vuichoud, J. Milani, A. Bornet, R. Melzi, S. Jannin and G. Bodenhausen, Hyperpolarization of Deuterated Metabolites via Remote Cross-Polarization and Dissolution Dynamic Nuclear Polarization, *J. Phys. Chem. B* **2014**, 118, 1411-1415.

[40] A. Bornet and S. Jannin, Optimizing dissolution dynamic nuclear polarization, *J. Magn. Reson.*, **2016**, 264, 13-21.

[41] J. E. Ollerenshaw, V. Tugarinov and L. W. Kay, *Magn. Reson. Chem.*, **2003**, 41, 843-852.