# A Theoretical and Experimental Study of Electrochemical pH Control at Gold Interdigitated Microband Arrays

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### Abstract

In electroanalysis, solution pH is a critical parameter that often needs to be adjusted and controlled for the detection of particular analytes. This is most commonly performed by the addition of chemicals, such as strong acids or bases. Electrochemical in-situ pH control offers the possibility for the local adjustment of pH at the point of detection, without additional reagents. FEA simulations have been performed to guide experimental design for both electroanalysis and in-situ control of solution pH. No previous model exists that describes the generation of protons at an interdigitated electrode array in buffered solution with one comb acting as a protonator, and the other as the sensor. In this work, FEA models are developed to provide insight into the optimum conditions necessary for electrochemical pH control. The magnitude of applied galvanostatic current has a direct relation to the flux of protons generated and subsequent change in pH. Increasing the separation between the electrodes increases the time taken for protons to diffuse across the gap. The final pH achieved at both, protonators and sensor electrodes, after 1 second, was shown to be largely uninfluenced by the initial pH of the solution. The impact of buffer concentration was modelled and investigated. In practice, the pH at the electrode surface was probed by means of cyclic voltammetry, i.e., by cycling a gold electrode in solution and identifying the potential of the gold oxide reduction peak. A pH indicator, methyl red, was used to visualise the solution pH change at the electrodes, comparing well with the model's prediction.

## Keywords

Finite element analysis simulation, in-situ pH control, solid state sensors, Electroanalysis, Microband interdigitated electrode array(6 max)

### 1. Introduction

The pH of a solution is an important parameter in many biological and chemical reactions, either as a catalyst or where the protons are active as reactant or product. The detection of numerous analytes, including porphyrins[1], nitrates/nitrites[2, 3], chlorine[4] and human haemoglobin[5], requires the adjustment of pH from neutral to acidic conditions. The typical method of changing solution pH is the rudimentary addition of buffers or strong acids and bases. A number of issues can arise from this method including the purchase of additional chemicals, the requirement for trained personnel to handle them, the added safety risk and finally the potential to act as a source of contamination. Controlling the pH by in-situ electrochemical methods allows the user to adjust the pH both temporally and spatially and perform electroanalysis with the same device; thereby eliminating the extra steps. Potentially important electrochemical pH control methods include electrochemical redox reactions[6] and water electrolysis[7]. The latter is the splitting of water with an electrical current and can be used to reduce the solution pH by applying a positive potential to consumes hydroxyl ions in a basic solution or produce protons in acidic conditions.

Finite element analysis (FEA) simulations have been used to provide valuable insight into a variety of electrochemical processes. Mass transport at electrode arrays has been modelled to determine inter-electrode spacing needed to achieve diffusional independence [8, 9]. The simulations directly informed the subsequent sensor design and fabrication. Similarly, the theoretical study of ionic transfer at liquid-liquid interfaces has guided the development of micro and nano-ITIES (interface between two immiscible electrolyte solutions) systems[10]. Adsorption processes at electrode surfaces have also been simulated, to assess reactions taking place at the electrode as the electrode itself is modified[11]. Wahl et al. demonstrated the use of simulations for iron detection at interdigitated microband electrodes(IDE) and the expected collection efficiency in generator-collector(GC) mode[12]. The spacing between the electrode combs is inversely proportional to the diffusional overlap and thus, at smaller inter-comb distances a greater degree of overlap is present.

The modelling of solution pH change and its effects has also been used to inform on experimental design and comparison to results. Read et al[13] modelled the change in pH at concentric boron doped diamond ring electrodes. Protons were generated at the outer electrode and the decreased local pH was monitored by use of an iridium oxide electrode. While elegant, this model did not account for a solution that contained buffering agents. Critelli et al[14] have shown the effect of buffer reacting with generated protons and the impact on the rate of solution pH change. Proton flux, buffering effects and competing simultaneous chemical reactions were all investigated. While this model focused on the effects of probing the pH near the electrode surface, similar equations are applicable to models focused on directly changing the local pH. The combination of both models would allow the simulation of proton flux generated by a fixed current and the subsequent buffering effects to be accounted for.

In this work, FEA simulations are performed for proton generation by water electrolysis. This approach builds upon the works of Read and Critelli mentioned above by modelling the generation of protons at one comb of interdigitated electrodes and their buffered diffusion towards the other comb and towards the bulk solution. A galvanostatic current is applied at one array, with the flux of protons proportional to the applied current. The importance of inter-electrode distance is investigated by modelling the proton generation at 3 different electrode separations. In response to these simulations, gold interdigitated microband arrays are designed and fabricated for the monitoring and control of solution pH. Solution buffer capacity was also investigated as it can impact on the change in pH at the surface of the sensing electrodes. In our approach, the gold oxide reduction peak is used as a probe for local solution pH at the working electrode array is visualized optically by use of a pH indicator, methyl red. The region of colour change is compared to the simulated proton concentration profiles.

## 2. Materials and Methods

## 2.1. Materials and Electrode Characterization

Nitric acid (35%), sodium chloride, ferrocene carboxylic acid (FcCOOH), phosphate buffer solution (PBS), copper chloride, disodium phosphate and citric acid were purchased from Sigma Aldrich, Ireland and used as received. All solutions were prepared with DI water with resistivity of 18 M $\Omega$  cm<sup>-1</sup>. A CHI 920 bi-potentiostat was used for the analytical measurements with the images captured on a Zeiss microscope equipped with a charge-coupled detector camera(CCD). All experiments were undertaken in a four electrode cell comprising of the on chip gold counter electrode and either an Ag/AgCl as external reference electrode or the on chip platinum pseudo reference electrode. The interdigitated working electrodes were electrochemically characterized by cyclic voltammetry using 1 mM FcCOOH in 10 mM PBS pH 7.4 solution. The potential was cycled between -0.15 V and 0.45 V vs. an on chip platinum pseudo-reference electrode with a scan rate of 100 mV/s. Sensing electrodes of 1  $\mu$ m width with a 2  $\mu$ m gap to the protonators typically exhibited a plateau current of 10 nA in non-GC mode, and 28 nA in GC mode analysis.

# 2.2. In-situ pH control simulation study

Figure S1A shows a schematic of the in-situ pH control concept. GC electrode array devices composed of two combs of interdigitated electrodes were employed for in-situ pH control in this work. A comb, in this regard, refers to one half of an interdigitated electrode array. The

interdigitated electrodes are spaced 2  $\mu$ m apart. By imposing an appropriate potential at one ("protonator") comb of electrodes, a pH change occurs in the local environment that tailors the pH at the other ("sensor") comb. The sensor comb may then be used to perform sensing in conditions that differ from the bulk solution. In our approach, hydrolysis of water is employed to produce protons at the protonator electrode, which then diffuse to and lower the pH in the region of the sensor electrode; compared to the bulk solution.[13]The pH may be tailored to the required value by control of the applied current density.[15] In acidic conditions, the pH can be changed through the water splitting reactions of equations 1-2:[16]

Cathode: 
$$2H^+ + 2e^- \rightarrow H_2$$
 (1)  
Anode:  $2H_2O \rightarrow O_2 + 4H^+ + 4e^-$  (2)

In basic conditions, the mechanism becomes:

Cathode: 
$$2H_2O + 2e^- \rightarrow 2OH^- + H_2$$
 (3)  
Anode:  $4OH^- \rightarrow O_2 + 2H_2O + 4e^-$  (4)

An acidic medium can be made more basic by consumption of protons in a reduction reaction, or alternatively more acidic by production of protons in an oxidation reaction (depending on the potential imposed at an electrode), and vice versa. It is important to note that the maximum pH changes occur close to the electrode and diminishes with increasing distance from the electrode.

A simulation model was designed to explore generation and diffusion of protons at the microband electrodes, using the FEA software COMSOL Multiphysics 5.3, in line with the galvanostatic model shown by Read et al[13]. The geometry of the model consisted of a 5 mm square box as the experimental domain, and two sets of interdigitated 1  $\mu$ m wide microband electrodes of 50 nm height (14 protonator electrodes and 13 sensing electrodes), separated by 2  $\mu$ m; see Figure S1B and C. In the model, a flux of protons was applied to the surface of the protonators, by applying a fixed anodic current. The flux was assumed to be proportional to a galvanostatic current applied at the electrodes;

$$Flux = \frac{i}{F.A} \quad (5)$$

where "i" is the current applied, F is Faraday's constant and A is the surface area of the electrodes. The initial pH was set to pH 7. The proton diffusion coefficient used for the simulation was 9.31 x  $10^{-5}$  cm<sup>2</sup> s<sup>-1</sup>.[17] Diffusion of protons was modelled according to Fick's second law. The possible effects of alkalinity, electrode separation and buffering capacity on the diffusion profiles were

investigated. The bulk pH modelled was from modelled pH 6 to 10. Interdigitated electrode array separation of 1 and 10  $\mu$ m were also modelled.

The effects of a buffer were modelled by introduction of chemical equilibriums, as shown by Critelli et al [14]reflecting the reactions between the protons, the buffer molecule and water dissociation

 $H^{+} + 0H^{-} \rightarrow H_2 0 \qquad k_w = -14 \quad (6)$  $H^{+} + A^{-} \rightarrow HA \qquad k_a = -6.8 \quad (7)$ 

Ions OH<sup>-</sup>, HA and A<sup>-</sup> were defined, reflecting the hydroxyl ion, protonated and unprotonated buffer molecules, respectively.  $k_w$  is the water dissociation constant, while  $k_a$  is the acid dissociation constant. Initial concentration of the buffer was set at 1 mM to reflect the concentration of sodium bicarbonate in the experimental buffer. The diffusion coefficients utilised were previously reported: proton 9.31 x 10<sup>-5</sup> cm<sup>2</sup> s <sup>-1</sup>; hydroxyl 5.31 x 10<sup>-5</sup> cm<sup>2</sup> s <sup>-1</sup>; protonated and unprotonated buffer 1 x 10<sup>-5</sup> cm<sup>2</sup> s <sup>-1</sup>.[14, 17]

Mesh quality is an important consideration when using FEA. As the model domain is divided into smaller sections or "elements", the collection of lines of division are known as a mesh. A key balance to meshing is the relationship between mesh quality and its computational time or cost. It is necessary for mesh quality to be high enough to obtain an accurate result, but a higher mesh quality results in more elements and thus a longer time to obtain a solution. Extra refinement is needed around the smaller features of the domain (the microbands) in comparison to the remainder of the domain. In these experiments, an error of 1% was targeted. The mesh refinement was acceptable once increasing the refinement no longer produced a discernible change on the concentration profiles.

#### 2.3. Electrode Fabrication

Silicon chip sensor devices, consisting of interdigitated gold microband electrode arrays, a gold counter electrode and a platinum pseudo reference, were fabricated as described previously[12, 18]; see Figure S2A. In brief, interdigitated gold microbands were patterned in resist by photolithography followed by metal evaporation (Ti/Au 5/50 nm) and standard lift-off. Similarly, photolithography, metal evaporation (Ti/Au 10/90), and lift-off procedures were then employed to overlay electrical interconnection tracks including peripheral probe pads. Macroscale gold and platinum counter and pseudo-reference electrodes, respectively, was also deposited during this process. Finally, a silicon nitride passivation layer (500 nm thick) was deposited to passivate the

entire chip and windows selectively opened with a dry etch to allow exclusive contact between the working, reference and counter electrodes with the solution of interest.

# 2.4. Electrochemical approaches

# Gold oxide reduction peak

The effect of the solution pH on gold oxide reduction was studied. Gold oxide was formed and then reduced by cyclic voltammetry over a potential range of -0.15 V to 1.2 V on a gold working electrode array at 100 mV/s. The solution pH was varied chemically over a range of pH 3 to 8 by preparing a series of buffer solutions of citric acid and disodium phosphate from pH 3 to 8. For electrochemical pH control, a series of potentials high enough to induce water electrolysis (1.3 V to 1.55 V) was applied on the protonator electrode array during the CV.

# 2.5. pH Change visualisation

An artificial drinking water solution (ADW) containing the pH indicator, methyl red, was used to visualize the change in pH experimentally. ADW was made up by dissolving 1 g of sodium bicarbonate, 0.0654 g of magnesium sulfate (Sigma Aldrich, 99.5% anhydrous), 0.3414 g calcium sulfate dehydrate (Honeywell, 99%), 0.007 g potassium phosphate dibasic (Fluka, 98%), potassium phosphate monobasic (Sigma Aldrich, 99%) and 0.01 g sodium nitrate (Sigma Aldrich, 99%) in 10 litres of deionized water. 50 mg of methyl red was dissolved in 25 ml of ethanol. This solution was then made up to 250 mL with 25 mM ADW solution. A portable CHI 1222c potentiostat was used for visualization experiments. A current of 500 nA was applied at the protonator. The images were captured on a Zeiss microscope equipped with a charge-coupled detector camera.

# 3. Results and Discussion

# 3.1. Simulation of pH change vs Increasing Current

FEA simulations were performed to study the generation and diffusion of protons at an interdigitated array of electrodes. The array studied consisted of interdigitated combs of 1  $\mu$ m wide electrodes (13 sensors, 14 protonators) of 50 nm height separated by 2  $\mu$ m in a 5 mm square domain. A flux of protons is generated at the protonator array (surface area of 15.4  $\mu$ m<sup>2</sup>) by applying a galvanostatic current. Figures 1A-D show proton concentration profiles as pH after 10 seconds of flux in a solution with starting pH of 7 (blue) over a range of applied currents. Figure

1A, with a current of 1 nA, presents a semicircular change in colour (pH) around the electrodes, with the largest colour change (red) closest to the electrodes. This change in colour represents the diffusion zone of the protons generated at the protonator array, with a greater pH change near the electrodes and gradually returning to bulk conditions at greater distances from the electrodes. Figures 1B, C and D are the concentration profiles for applied currents of 10, 100 and 1000 nA, respectively, and each possess a diffusion zone with an increased diameter and a greater proportion of red (pH 4 and below) than the lower currents. From equation 5, a larger current generates a larger flux of protons and the resulting lower pH. Figure 1E is a plot of solution pH at the electrode top surface (50 nm) across the width of the domain. A lower pH is reached with a larger current. At each applied current, there is a very sharp decrease in pH observed at the center of the domain (shown in Figure 1E at 2,500  $\mu$ m on the x-axis), with a more gradual change in pH moving from the center to the edges until bulk conditions again towards the edges. This enhanced drop in pH in the central regions (Figure 1E inset) occurs directly at the electrodes since the protonators are generating protons at a higher rate than they can diffuse away thereby creating a region of lower pH above the electrodes.



Figure 1Simulated pH profile after 10 seconds with applied current of 1 (A), 10 (B), 100 (C) and 1000 (D) nA at protonators (Surface Area 15.4  $\mu$  m<sup>2</sup>); (E) Plot of pH at height over the electrodes vs width of model domain. Inset: Magnified image of region directly surrounding electrode array.

## **3.2.** Electrode Separations

The effect of the changing the electrode separations in the interdigitated arrays viz 1  $\mu$ m, 2  $\mu$ m and 10  $\mu$ m between the protonator and sensing combs was investigated after 100 microseconds and 1 second of applied current of 100 nA since increasing the duration of applied current should impact distance over which protons diffuse. We note that the 80  $\mu$ m passivation window, in which the array is located, limited the number of 1  $\mu$ m width microband electrodes that can be present as the spacing between combs increases. A 1  $\mu$ m separation array contained 20 protonators and 20

sensors, a 2 µm array contains 14 protonators and 13 sensors, a 10 µm array contained 4 protonators and 4 sensors. Figure 2A top panel shows the proton concentration profile after 100 microseconds of the 10 µm gap array. There are semicircular diffusion zones around each of the 4 protonator electrodes representing the change in pH. The clear separation between the diffusion zones is demonstrated in Figure 2A bottom panel (black), where bulk solution pH is present at the sensing electrodes (the midpoint between protonators). Figure 2A bottom panel (red) also displays the pH after 1 second which has decreased significantly. After this time, the zones of decreased pH around each of the 4 protonator electrodes have overlapped significantly, though we note there is a small deviation of ~0.1 pH units midpoint between protonators (over the sensing electrodes). Figure 2B top panel shows the proton concentration profile after 100 microseconds of the 2 µm gap array. There are semicircular diffusion zones around each of the 14 protonator electrodes representing the change in pH. The overlap of these individual diffusion zones is demonstrated in Figure 2B bottom panel (black), where bulk solution pH is not present at the sensing electrodes (the midpoint between protonators). Figure 5B bottom panel (red) shows that the pH variation midpoint between protonators (i.e., at the sensors), has decreased to ~0.02 pH units after 1 second. Figure 2C top panel, shows the proton concentration profile after 100 microseconds of the 1 µm gap array. There are semicircular diffusion zones around each of the 20 protonator electrodes representing the change in pH. The overlap of these individual diffusion zones is demonstrated in Figure 2C bottom panel (black), where bulk solution pH is not present at the sensing electrodes (the midpoint between protonators). With decreasing separations between the electrodes it can be observed that the generated pH change is becoming more uniform across the array even at this short time interval. The overlap of these individual diffusion zones after 1 second is demonstrated in Figure 5C bottom panel (red), where the variation in pH, midpoint between protonators, is negligible. Consequently, the simulation study suggests that the pH has been sufficiently adjusted after only 1 second of pH control at these different gaps.



Figure 2 Top Panel: Simulated pH profile after 100  $\mu$ s with applied current of 100 nA at working electrode arrays with gaps of 10 (A), 2(B) and 1 (C)  $\mu$ m. Bottom Panel (A, B, C) Plots of pH at electrode surfaces across array after 100  $\mu$ s and 1 s

#### **3.3.** Effect of initial solution pH

The effect of varying the initial starting pH of the bulk solution, over the range of pH 6 to pH 10, on the final pH was investigated. To this end, only the 2 µm spacing interdigitated electrode arrays were presented as these were the electrodes that were challenged in the subsequent experimental study below. A galvanostatic current of 100 nA was again applied to the protonators within the interdigitated electrode array. Figure 3A shows the proton concentration profile after 100 microseconds of applied current with the starting bulk pH set at pH 6 (yellow). Individual diffusion zones observed around each of the protonators indicate the decrease in pH due to the flux of protons and their subsequent diffusion, highlighted by the red regions. The inset, a magnified image, shows a small degree of overlap between the diffusion zones, indicating that the local pH at the sensing electrodes remains relatively unchanged. Figure 3B shows the equivalent simulation when the starting pH was set at 10. Again a decrease in pH was observed following proton production at the protonators and their subsequent diffusion. The inset shows that there is relatively more overlap of the diffusion profiles and pH still appears to reach pH  $\sim 6.0$  above the sensors. To confirm this, the final pH in the plane parallel to and 50 nm above the chip surface (i.e. at the sensor surface) is plotted in Figure 3C where a saw tooth pH profile was observed as before across the array. The peak maxima were again located over the sensors while the peak minima were over the protonators, as expected. It was observed that regardless of the initial solution pH, a final pH of ~6.0 was achieved (pH 5.7 for starting pH 6 was reached while a final pH of 6 was reached for starting pH 7, 9 and 10, respectively). The pH at the protonator surface was always observed to

reach pH 4.2 for different starting pH values. Clearly, after 100  $\mu$ s there is a change of ~1.7 pH units of the solution between the protonators and the sensors. Figure 3D shows the equivalent simulation conditions after 1 s of applied current. Here it was observed that a final pH, with little spatial variation, of ~2.8 was observed across the entire array. The inset shows a less magnified view to highlight initial starting pH of the simulations. Consequently, these results indicate that the starting pH does not have a significant effect on achieving the final desired pH at the sensors within one second of applied protonator current.



Figure 3 Simulated pH profile after 100 microseconds with applied current of 100 nA at 2  $\mu$ m gap working electrode arrays with starting bulk pH of 6 (A) and 10 (B). (C) and (D) show plots of the pH at the sensor electrode surface after 100 microseconds and 1 seconds

## **3.4.** Solution Buffer capacity

The model employed in the previous sections was reported by Read et al[13] and based on a purely diffusion dependant model employing a galvanostatic current. To factor in the effects of proton buffering reactions, the chemical equations from the Critelli et al[14] analysis were added (equations 5 and 6) to provide a more realistic model. Figure 4 features concentration profiles focused on a 1 mm x 1mm square by the electrodes in the modelled 5 x 5 mm domain.

Figure 4A top panel is the proton concentration profile for a solution of 1 mM bicarbonate buffer after 1 second. The diameter of the region of lower pH is smaller than that of the non-buffered

model. There is a sharper transition in pH from protonator electrodes to the bulk pH. The 1 mM buffer is sufficient to narrow the diameter the area of decreased pH (340  $\mu$ m vs. 1,150  $\mu$ m unbuffered), however the flux at the electrodes still reaches pH 4 and below. Figure 4A bottom panel is the proton concentration profile for a solution with a 1 mM buffer after 10 seconds, where the diffusion zone of changed pH has increased in diameter with time (1,050  $\mu$ m compared to 3,600  $\mu$ m for unbuffered).

Figure 4B top panel is the proton concentration profile for a solution with a 10 mM buffer after 1 second. The diameter of the region of lower pH is again reduced (263 um). The pH at the electrodes has not reached pH 4 after 1 second in a 10 mM buffer. Figure 4B bottom panel is the proton concentration profile for a solution with a 10 mM buffer after 10 seconds. The diffusion zone diameter has increased (740 um) and local pH has now decreased below pH 4. It was observed that the increased volume of protons over the 9 second interval has surpassed the buffering capacity of the buffer.

Figure 4C top panel is the proton concentration profile for a solution with a 100 mM buffer after 1 second. The colour change indicating a lower pH is barely evident (184  $\mu$ m with a lowest pH of 6.57). 100 mM buffer significantly inhibits the electrochemical pH control for this applied current and duration. Figure 4C bottom panel is the proton concentration profile for a solution with a 100 mM buffer after 10 seconds. There is a negligible decrease in pH after the 9 seconds interval, due to the buffering capacity of the 100 mM solution being greater than the flux of protons.

Figure 4D top panel is a plot of pH at the electrode surfaces after 1 second. For 1 mM, the pH at the protonators and sensors is as homogenous as an unbuffered solution. There is a large change in pH between the 2 combs for 10 mM. This buffer is effective after 1 second at scavenging protons over the 2  $\mu$ m gap between the combs. There is a slight pH difference between the combs at 100 mM. This difference being smaller than that of the 10 mM is due to the smaller relative change in pH. Figure 4D bottom panel is a plot of pH at the electrode surfaces after 10 seconds. Both the 1 and 10 mM buffer solutions now have the homogeneity of local pH at both electrode combs, with less of a decrease for the 10 mM buffer. For the 100 mM, buffer the same scenario exists after 10 seconds albeit at a slightly lower pH (6.40). These results would suggest that to decrease the pH in high buffer concentrations would require a much higher current, which will be the subject of future work.



Figure 4 Top Panel: Simulated pH profile after 1 second with applied current of 100 nA with buffer concentration of 1 (A), 10 (B) and 100 (C) mM with (D) showing a plot of pH across the electrode surfaces; Bottom Panel: Simulated pH profile after 10 seconds with

### 3.5. Electrode Characterisation

Experiments were undertaken, using silicon chip substrates, to confirm the simulation study presented above. Interdigitated microband electrodes were fabricated as described in the experimental section. Figure S3A shows a typical voltammogram in PBS of pH 7.4 over the potential range of -0.15 V to 1.2 V. An increase in current beginning at 0.6 V indicates the onset of gold oxidation. The plateau and subsequent increase at 0.75 V is indicative of the formation of a complex gold oxide morphology/ multilayers. The sharp increase in current beginning at 1.1 V is attributed to the onset of oxygen evolution described in equations 2 or 4. The electrochemical characterization was undertaken by cyclic voltammetry in FcCOOH from -0.15 V to 0.45 V at 100 mV/s in both three and four electrode configurations using the on-chip gold counter and platinum reference electrodes. Figure S3B presents typical voltammograms obtained. In the three electrode configuration, each IDE sensing electrode array exhibited a current magnitude of  $\sim 10$  nA, typical of such an array. The magnitude of this current strongly suggests that oxidation of FcCOOH only occurs at the interdigitated electrodes and that the interconnection metallization is completely passivated by the silicon nitride passivation layer. The diffusion limited peak shape arose from the electrodes of the array not being diffusionally independent due to their close proximity, as expected [12]. By contrast, in four electrode GC mode, a constant potential of -0.15 V was applied at the protonator electrode while the sensor electrode was scanned. A current of 28 nA was obtained at the sensing array electrodes with steady-state sigmoidal CV profiles observed. The increase in current arose from redox cycling of the FcCOOH between the protonator and sensor electrodes enabling the sensors to operate as diffusionally independent electrodes. The steadystate behavior again supports the thesis that the interconnection tracks were fully passivated and that redox reactions were only occurring at the IDEs.

# 3.6. Gold Oxide Reduction and pH Control

Gold oxide was formed on the sensor electrodes by sweeping the potential positively above 1.0 V (vs. Ag/AgCl). The potential of the subsequent gold oxide reduction was used to monitor solution pH at the microband electrodes as this potential is known to be pH sensitive[4]. Figure 5A shows an overlay of the gold oxide reduction peaks obtained at a sensor IDE immersed in different buffer solutions over the pH range 3 to 8. The sensor IDE was scanned from -0.2 to 1.2 V. The reduction potential of the gold oxide was observed to increase as the pH decreased. The change in reduction potential was then plotted versus pH and presented in Figure 5B. The plot exhibited excellent linearity with an  $R^2 = 0.994$  and a super Nernstian response of -72.4 mV/ pH., clearly indicating that gold oxide potential may be used as a method of pH monitoring over the time scale of these experiments. A possible reason for the super-Nernstian response is the presence of different oxidation states of gold, as shown with other metals oxide such as ruthenium and iridium when used as pH probes.[19] We note for longer timescales, dissolution of gold from the electrode could limit this approach.

Figure 5C shows an overlay of the gold oxide reduction peak obtained at a sensor electrode using artificial drinking water solution. The sensor IDE was again scanned from -0.2 to 1.2 V with the protonator electrode array biased at a constant potential. A variety of different potentials ranging from 1.3 to 1.55 V were applied to the protonator and the reduction potential measured. An increase in the reduction potential was observed as the applied potential is increased suggesting that the pH is decreasing as expected. This reduction peak potential was again plotted against pH, presented in figure 5D and again exhibited very good linearity with  $R^2 = 0.993$  over the timescale of the experiment. It may be concluded from these results confirm the simulation study; that the application of a suitable potential to the protonator can cause a change in solution pH (measured at the sensor electrodes), and that applying a greater potential leads to a larger change in pH, as a greater flux of protons are produced.



Figure 5 Reduction peak of gold oxide in chemically controlled pH buffers (A) with calibration plot of reduction potential vs pH(B); Reduction peak of gold oxide in electrochemically pH controlled buffer (C) with calibration plot of reduction potential vs applied potential(D).

### 3.7. Visualization of Electrochemical pH control

To further confirm the simulation study, electrochemically induced pH change was visualized experimentally by addition of methyl red, a pH indicator with a pH range of 4.2 (red) to 6.2 (yellow), to a 1 mM solution of ADW. The change of solution pH was monitored by use of a portable potentiostat and a microscope equipped with a CCD, as described in the experimental section. The simulated concentration profiles of a 1 mM buffered solution were compared to the results obtained experimentally. Figure 6A top panel shows the simulated concentration profile at t = 0 seconds, while Figure 6A bottom panel is an optical image of the electrodes t = 0. In both images, there is no change in colour as no current has been applied and consequently no change in pH has taken place. Figure 6B top panel shows the simulated concentration profile at t = 1s, while Figure 6B bottom panel is the corresponding optical image of the electrodes. A colour change is visible around the electrodes in both the simulated profile and the optical image at this time. The optical colour change arises from the generated protons interacting with the methyl red in the solution, with the colour change from yellow to red indicating that the pH has decreased to pH 4.2 or below. The simulated region below pH has a diameter of ~180 um, while the red area in the optical image has a diameter of ~175 um. Figure 6C top panel shows the simulated concentration

profile at t = 10 s, while Figure 6C bottom panel is the corresponding experimental optical image of the electrodes. The diameter of the colour change has increased in both the simulated and experimental results to 530 µm and ~450 µm, respectively. There is a slight discrepancy between the simulation's diffusion zone and the colour change visualized experimentally. There are a few potential sources of this: The ethanol added to dissolve the methyl red will change the diffusion coefficient of protons in a water based solution. The visualization technique requires a certain colour intensity to be visible. However, it is possible that protons have diffused beyond this visible region, but the colour is not intense enough to be yet observed. The addition of methyl red may also affect the diffusion coefficient of protons in solution. It has been reported by Xu et al.[20] that proton diffusion can be suppressed in solutions containing tetra methyl urea, an amphiphile with functional group similarity to methyl red.



Figure 6 Top panel: Simulated pH profile of proton generation at protonator electrodes at 0 (A), 1 (B) and 10 (C) seconds in 1 mM buffer; Bottom panel: Optical images of working electrode array as current applied at protonators at 0 (A), 1 (B) and 10 (C) seconds

#### 4. Conclusion

We have shown a simulation and experimental study of electrochemical pH control at interdigitated gold microband arrays. The application of a galvanostatic current at the protonator array generates protons by water electrolysis, which diffuse away and reduce the local solution

pH. This method allows the pH at a sensor array to be adjusted while eliminating the traditional need for the skilled handling of dangerous chemicals or the inherent expense and complexity thereof. The model has highlighted the impact of the spacing between the interdigitated electrodes, where the desired pH is reached at the sensing electrodes more rapidly at smaller distances. The initial bulk pH was shown by the simulation to be unimportant in achieving the final pH at the electrode surfaces. Buffering reactions were factored into the model, with the buffer concentration having a clear impact on the diffusion of protons from the protonator electrodes to the sensors. Microband electrodes were fabricated in line with the geometries modelled and the of the gold oxide reduction peak was used as a measurement probe for the local solution pH at the surface of the electrodes. This work shows the feasibility of using a pH indicator to corroborate the region of pH change around the protonator and sensing electrodes predicted by FEA. This in turn supports the use of electrochemical pH control in electroanalysis, where the analytes of interest are pH dependent.

# Acknowledgements

The authors would like to acknowledge funding from the Irish Research Council (Project GOIPG/2015/2925). This work has been supported in part by a research grant for the VistaMilk Centre Science Foundation Ireland (SFI); Department of Agriculture, Food and the Marine (DAFM) under Grant Number 16/RC/3835

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