

## Surface-Induced Dissociation of Protein Complexes Selected by Trapped Ion Mobility Spectrometry

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## Abstract

Native mass spectrometry, particularly in conjunction with gas-phase ion mobility spectrometry measurements, has proven useful as a structural biology tool for evaluating the stoichiometry, conformation, and topology of protein complexes. Here, we demonstrate the combination of trapped ion mobility spectrometry (TIMS) and surface-induced dissociation (SID) on a Bruker Solarix XR 15 T FT-ICR mass spectrometer for structural analysis of protein complexes. We successfully performed SID on mobility-selected protein complexes, including streptavidin tetramer and cholera toxin B with bound ligand. Additionally, TIMS-SID was employed on a mixture of peptides bradykinin desR1 and desR9 to mobility separate and identify the individual peptides. Importantly, results show that native-like conformations can be maintained throughout the TIMS analysis. The TIMS-SID spectra are analogous to SID spectra acquired using quadrupole mass selection, indicating little measurable, if any, structural rearrangement during mobility selection. Mobility parking was used on the ion of mobility of interest and 50 to 200 SID mass spectra were averaged. High quality TIMS-SID spectra were acquired over a period of 2-10 minutes, comparable to or slightly longer than SID coupled with ion mobility on various instrument platforms in our laboratory. The ultrahigh resolving power of the 15 T FT-ICR allowed for the identification and relative quantification of overlapping SID fragments with the same nominal  $m/z$  based on isotope patterns and shows promise as a platform to probe small mass differences, such as protein-ligand binding or post-translational modifications. These results represent the potential of TIMS-SID-MS for the analysis of both protein complexes and peptides.

## Introduction

Proteins often carry out their functions as complexes of multiple proteins. The understanding of their assembly is integral to determining how architecture affects function and possibly malfunction. Traditional biophysical methods, such as NMR, Cryo-EM, or x-ray crystallography, have long been useful tools for probing the structures and dynamics of biological molecules.<sup>4</sup> Native mass spectrometry (nMS), particularly in conjunction with gas-phase ion mobility measurements, offers complementary information for evaluating the stoichiometry, conformation, and topology of protein complexes. The information provided as well as its sensitivity and ease of use have made nMS a rapidly growing technique for analyzing proteins and protein complexes prepared in solutions that mimic the cellular environment, thus maintaining their native-like structure.<sup>1,2</sup> These biological systems are softly, but efficiently, ionized and transferred to the gas phase using nanoelectrospray ionization (nESI) in kinetically-trapped, native-like structures, in which structural disruption is limited and noncovalent interfaces are preserved.<sup>3</sup> Importantly, nMS is capable of probing multiple conformations and proteoforms of protein complexes with low sample amounts, less extensive sample preparation, and relatively uncomplicated data processing.<sup>5-7</sup> The combination of ion mobility spectrometry (IMS) and surface-induced dissociation (SID) in a single native mass spectrometry experiment has the potential to provide a wealth of structural information for protein complexes, including conformational landscape information and subunit stoichiometry, connectivity, and topology.<sup>8-11</sup>

IMS provides an extra dimension of structural characterization when coupled with mass spectrometry by separating ions not only by mass, but also by their rotationally averaged collision cross sections.<sup>9, 12</sup> There are several different ion mobility methods and instrument types available, and these have been compared in depth previously.<sup>12</sup> Both drift tube ion mobility spectrometry (DTIMS) and traveling wave ion mobility spectrometry (TWIMS) have been utilized regularly by the nMS community for the analysis of protein complexes. In linear cell drift tube IMS, a potential is applied along the ion mobility cell while ions are accelerated into a quasi-stationary bath gas, providing a drag force that opposes the ion motion along the potential gradient.<sup>13-15</sup> As a result, larger or more expanded ions undergo a greater number of collisions, therefore taking longer to traverse the IMS cell compared to more rigid or compact conformations. For ions of the same  $m/z$ , more compact ions, described as having higher mobility, will elute from the IMS cell sooner compared to expanded ions.

Trapped ion mobility spectrometry (TIMS), initially optimized and implemented for small molecules, has been gaining traction for the analysis of native proteins and protein complexes.<sup>16, 17</sup> TIMS differs from the previously mentioned techniques in that an electric field that opposes the ion motion is used to hold ions stationary against a bath gas which pushes ions along the TIMS analyzer (an ion funnel). Ions of different mobilities are trapped at different points (potentials) along the ion optical axis using the electric field gradient and are then eluted from the device over time, based on their mobility, as the TIMS potential gradient is reduced.<sup>18-21</sup> Because the bath gas flows in the same direction as the ions, larger or more expanded ions are trapped farther along the TIMS analyzer and elute sooner than smaller or compact ions. Because ions are trapped and separated in the TIMS funnel, resolving power and time of analysis can be user-defined and optimized for a given application. For example, if high resolving power is necessary to separate two ions of similar mobilities, the step size of the voltage profile (or slope) is decreased, and overall time of analysis increased. For species requiring less resolving power the voltage profile is stepped at larger increments and overall analysis time is much shorter, allowing for rapid analysis. Because the resolving power of TIMS is not limited to IMS analyzer length, as traditional drift tube or travelling wave ion mobility are, higher mobility resolution has been achieved with TIMS. Park and co-workers observed a resolving power up to ~300 for separation of ubiquitin conformations, allowing for the identification of

several additional conformations previously undetected using drift tube ion mobility spectrometry ( $R=20-50$ ).<sup>22-26</sup> The use of quadrupolar RF confinement in the TIMS funnel also decreases the radial spread and loss of ions compared to a traditional dc-only drift cell analyzer, therefore increasing total ion signal and allowing for ion activation of mobility-separated ions.<sup>27</sup> This is important in the results described here because having a confined and well defined ion beam after mobility analysis facilitates surface collision experiments.

SID is a gas-phase activation technique that can probe protein complex stoichiometry, connectivity, subunit interactions, and structural rearrangements.<sup>28-33</sup> To perform SID, ions are intentionally accelerated toward a rigid surface in the mass spectrometer. The result of the collision is a rapid increase in internal energy that often results in compact fragments indicative of the native topology of the complex, i.e., non-covalent bonds are cleaved to separate subcomplexes at the weaker of multiple interfaces. Most commercially available tandem mass spectrometers are equipped with gas-phase collision-induced dissociation (CID) capabilities, in which ions undergo numerous, low-energy collisions with neutral gas molecules.<sup>33</sup> CID can restructure protein complexes in a variety of ways that can include collapse or expansion/unfolding, often yielding highly charged monomer and the corresponding (n-1)mer, therefore limiting the amount of structural information gained from these experiments.<sup>34</sup> SID can access higher energy pathways with minimal restructuring, a pathway often unattainable by multi-step activation in CID, with SID often yielding lower-charge, compact subunits with symmetric charge partitioning. The implementation of SID capabilities on a Bruker Solarix XR 15T FT-ICR<sup>35-38</sup>, capable of high mass resolving power, has proven extremely beneficial for identifying overlapping oligomers produced from SID (i.e., dimer 4+ and monomer 2+) as well as probing small mass/charge differences, such as ligand binding or post translational modifications.<sup>29</sup>

TIMS combined with SID can be used to probe conformational dynamics, such as whether specific conformations can be identified as native-like, collapsed or expanded (including unfolding). With the typical symmetric charge partitioning of SID (and occasional/atypical symmetric charge partitioning of CID)<sup>39</sup>, product ions often overlap in  $m/z$  space, requiring another stage of separation for identification. The ultrahigh resolving power of the FT-ICR, however, allows for the identification of overlapping oligomers based on their isotope patterns. Here, we present the implementation and evaluation of TIMS combined with SID for the protein complexes streptavidin and cholera toxin B with GM1s ligand (sugar head group of GM1) and a peptide mixture containing bradykinin desArg1 and desArg9 on a 15 T FT-ICR mass spectrometer.

## Methods and Materials

### *Sample Preparation and Ionization*

Ammonium acetate, ethylenediamine diacetate (EDDA), cholera toxin B (CTB), the sodium salt of the pentasaccharide Gal $\beta$ 1-3GalNAc $\beta$ 1-4(Neu5Ac- $\alpha$ 2-3)-Gal $\beta$ 1-4Glc (GM1s), and bradykinin desArg1 (PPGFSPFR) and desArg9 (RPPGFSPF) were purchased from Sigma-Aldrich (St. Louis, MO). Streptavidin from *Streptomyces avidinii* was purchased from Thermo Scientific Pierce Biotechnology (Rockford, IL) and has the known issue that each protein monomer has 0 or 1 methionine at the N-terminus. Protein complexes were buffer exchanged twice into 200 mM ammonium acetate (pH near neutral) with size exclusion chromatography spin columns (Micro Bio-Spin 6, Bio-Rad, Hercules, CA). EDDA was used as a

charge-reducing agent and added to protein solutions at a final concentration of 40 mM EDDA and 160 mM ammonium acetate. Final protein concentrations ranged from 5-10  $\mu$ M. For the bradykinin desArg1 and desArg9 mixture, the peptides were mixed at equal concentrations in 1:1 water: acetonitrile, at a final concentration of 5  $\mu$ M each.

### *Instrumentation*

All experiments were performed on a Bruker Solarix XR 15 T FT-ICR mass spectrometer (depicted in **Figure 1**, Bruker Daltonics, Bremen, Germany). Ions for mass spectrometry experiments were generated using nanoelectrospray (nESI) ionization. Static nESI capillaries were prepared in-house using Sutter Instrument thin wall borosilicate capillaries with filament (outside diameter = 1.00 mm and inside diameter = 0.78 mm) and a Sutter Instrument P-97 Flaming/Brown type micropipette puller (Novato, CA). Capillary voltages of 0.6-1.1 kV were used for nESI via a 0.25mm diameter platinum wire (Sigma Aldrich, St. Louis, MO) inserted into the back of the capillary with 180 C, 5 L/min drying gas flow rate.

Once in the vacuum system ions may be analyzed and manipulated in a variety of ways. A prototype TIMS analyzer (**Figure 1A**) is incorporated in the first pumping region – serving the dual purpose of collecting the ions for transmission to downstream pumping stages and mobility analyzing the ions. The ions may also be mass filtered in an analytical quadrupole before arriving at the SID optic (**Figure 1B**). The ions may either be SID activated or allowed to pass unactivated into the collision cell. Typically, ions are collected in the collision cell before being transmitted to the ICR cell for mass analysis.

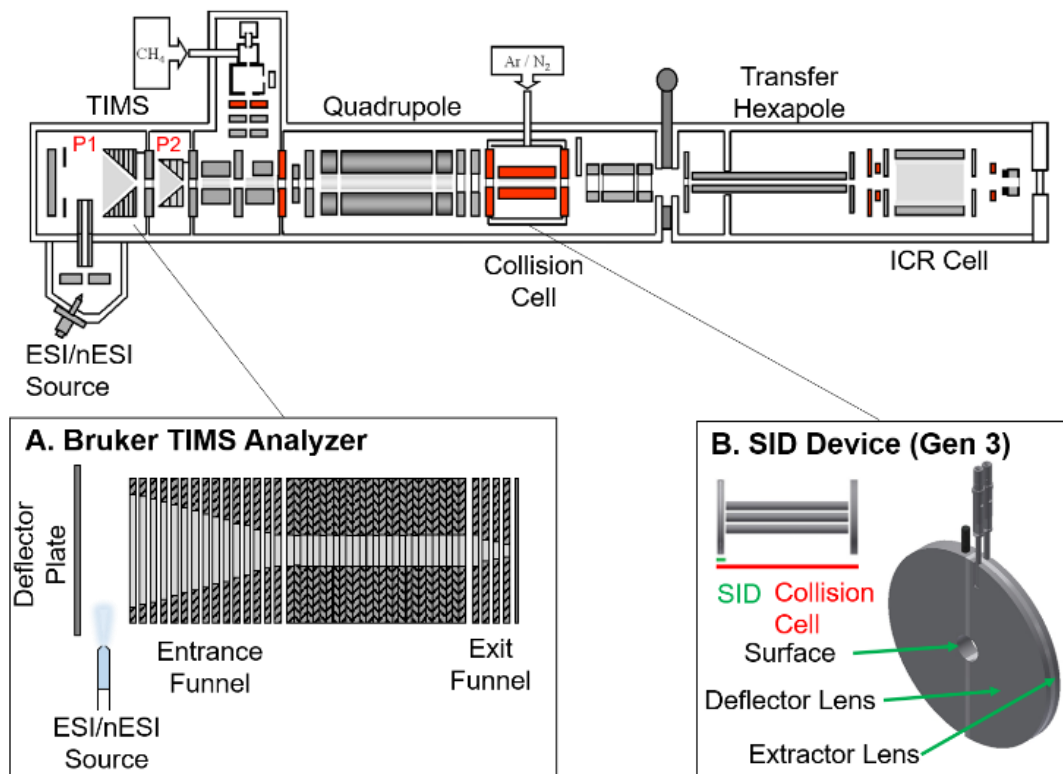
### *Surface-Induced Dissociation*

The front endcap of the standard Bruker collision cell was replaced with a split lens SID device, termed “Gen 3” and is described in a recent manuscript.<sup>35</sup> Gen 3 consists of three electrodes: a surface, deflector, and extractor lens (**Figure 1B**).<sup>35</sup> The device can be operated in both SID and transmission modes. The surface consists of a metal electrode (stainless steel, presumably coated with adventitious organics in the vacuum system). No significant neutralization/ion loss for large multiply-charged protein complexes was observed using a stainless steel surface compared to a gold surface coated with fluorocarbon self-assembled monolayers, as previously described by the Wysocki research group.<sup>35, 40</sup> In SID mode, positive ions are accelerated toward the surface by applying an attractive voltage (more negative than the exit of the quadrupole mass analyzer) to the surface, while applying a more positive voltage to the deflector (**Figure S13**). SID energy can be increased by applying an even more negative voltage to the surface electrode, thus increasing the ion acceleration into the surface. After surface collision, the activated ions are extracted directly into the collision cell by applying a voltage to the extractor lens that is equivalent to, or slightly more positive, than the surface voltage while simultaneously holding the collision cell rods at a DC bias lower than the surface. In transmission mode, the three lenses are set to equivalent potentials and ions simply traverse the front endcap without any surface collision. Transmission mode also allows for CID experiments in the collision cell without removal or modification of the Gen 3 SID device.

### *Implementation of TIMS with SID*

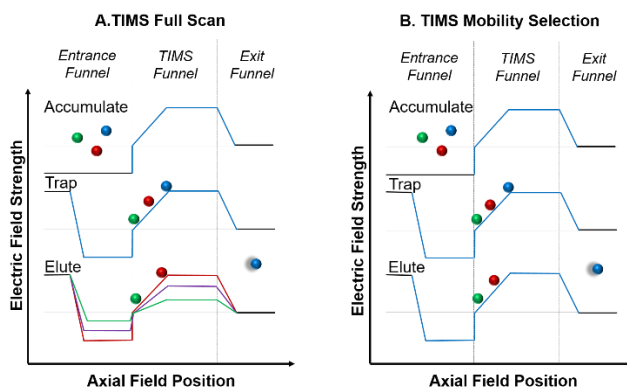
The ion funnel region was replaced with a custom TIMS analyzer (Bruker, Billerica, MA) equipped with an external power supply (**Figure 1A**).<sup>41, 21</sup> In-house software, written in C++ by Danielson Software Consulting (Richland, WA) utilizing National Instruments LabVIEW resources (Austin, TX), was used to control the

TIMS analyzer and was synchronized with the FT-ICR MS data acquisition program using a special pulse program. For TIMS analysis of large protein complex ions (greater than 3,000  $m/z$ ), the standard 900 kHz RF driver in the source (ion funnel) region was replaced with a 450 kHz RF driver capable of providing up to 300 V<sub>pp</sub>. Care was taken when setting the RF amplitude as high values can unintentionally activate protein and protein complex ions. Mobility separation was performed using nitrogen as a bath gas at approximately 300 K with P1 = 3.2 mbar and P2 = 1.4 mbar (**Figure 1**). The TIMS deflector plate (V<sub>def</sub>) voltage was set to 150 V for TIMS accumulation (open) and 0 V for the trapping and elution steps (closed) to prevent additional ions from entering the TIMS analyzer.



**Figure 1.** Instrument design and modifications, including a TIMS analyzer<sup>21</sup> (A) and SID device<sup>35</sup> (B). The TIMS analyzer replaced the entrance funnel and the SID device replaced the front-end cap of the commercial collision cell. The previously described SID device consists of three lenses: a semicircular surface onto which ions collide and two lenses with a non-conducting space in between: a deflector lens to push ions into the surface and an extractor lens element to extract and guide ions into the collision cell for accumulation prior to being pulsed into the ICR cell for detection.<sup>35</sup>

Additionally, the experimental setup includes an ion gate downstream of the TIMS funnel, as previously described elsewhere.<sup>21</sup> Briefly, the ion gate is used to select or eliminate ions of different mobilities, allowing for the coupling of TIMS with the slower FT-ICR mass analyzer.<sup>21</sup> The TIMS analyzer was operated using an accumulate/trap/elute (with gating)/quench sequence over 34 ms per mobility scan. Ions selected with the ion gate are accumulated in the Bruker collision cell before being pulsed into the ICR cell, with an average of 25 to 100 ion mobility scans per mass spectrum. For the filling step of one TIMS scan, ions were accumulated for 15 ms of the total 34 ms TIMS profile. The TIMS funnel was quenched at the conclusion of each scan before beginning the next. An example of the voltages and timings used in the TIMS ion funnel region can be found in the supplemental information (**Figures S11** and **S12**). For streptavidin tetramer, the starting and ending voltages of the TIMS ramp were set to -275 V and -50 V, respectively. After completion of a mobility of interest, voltage profile was lowered to the next mobility of interest.<sup>21</sup> A pictorial representation of the accumulate/trap/elute/quench sequence can be found in **Figure 2A**. When performing TIMS-SID experiments the trapping voltage was held constant at the elution voltage for the mobility corresponding to the precursor of interest, resulting in isolation of a selected mobility (referred to as “mobility parking”, **Figure 2B**). Approximately 50-200 mass spectra were averaged to yield the multidimensional IM-MS spectrum, resulting in an average analysis time ranging from 2-10 minutes (TIMS-SID required longer averaging time compared to performing SID without mobility selection due to the lower duty cycle of combining TIMS with SID, however the total experiment time is comparable to SID-IMS experiments performed on our Waters Synapt Q-SID-TWIMS-TOF mass spectrometers.<sup>34, 38</sup>



**Figure 2.** A visual representation of the TIMS voltage profiles used for full scans (**A**) or mobility selection for SID experiments (**B**). For the full scan, the voltage is lowered in a stepwise fashion to elute ions of different mobilities as a function of time. For mobility selection, the voltage is set for only the ion of interest to elute. The TIMS funnel is quenched after each mobility scan to remove any remaining ions before beginning the next accumulation step.

For each sample, an initial full TIMS scan was performed to determine the elution voltage for ions of particular interest, with the Gen 3 SID device held in transmission mode. To perform TIMS-SID experiments, the TIMS voltage ramp was held constant (or “parked”) at the elution voltage of interest and

the Gen 3 SID device was tuned for SID mode. The collision cell accumulation time was set to a value in the range of 0.6 to 3.0 seconds (depending on number of TIMS accumulations and need for increased ion throughput, similar to Bruker's typical CASI, continuous accumulation of selected ions, mode for CID). Post-accumulation, ion packets were pulsed into the ICR cell for detection. The process of pulsing an ion packet into the ICR was performed multiple times and the resulting mass spectra were averaged for a total analysis of time of 2-10 minutes. The time-of-flight setting, which controls the time ions have to travel between the collision cell into the ICR cell was optimized to ensure a balanced detection of low and high  $m/z$  ions (typically 2.2 to 3.0 milliseconds for protein complexes). A low time-of-flight setting results in a bias toward increased detection of low  $m/z$  ions, as high  $m/z$  ions require a longer time-of-flight. An example of the ICR tune settings used for nMS experiments are provided in **Figure S14**.

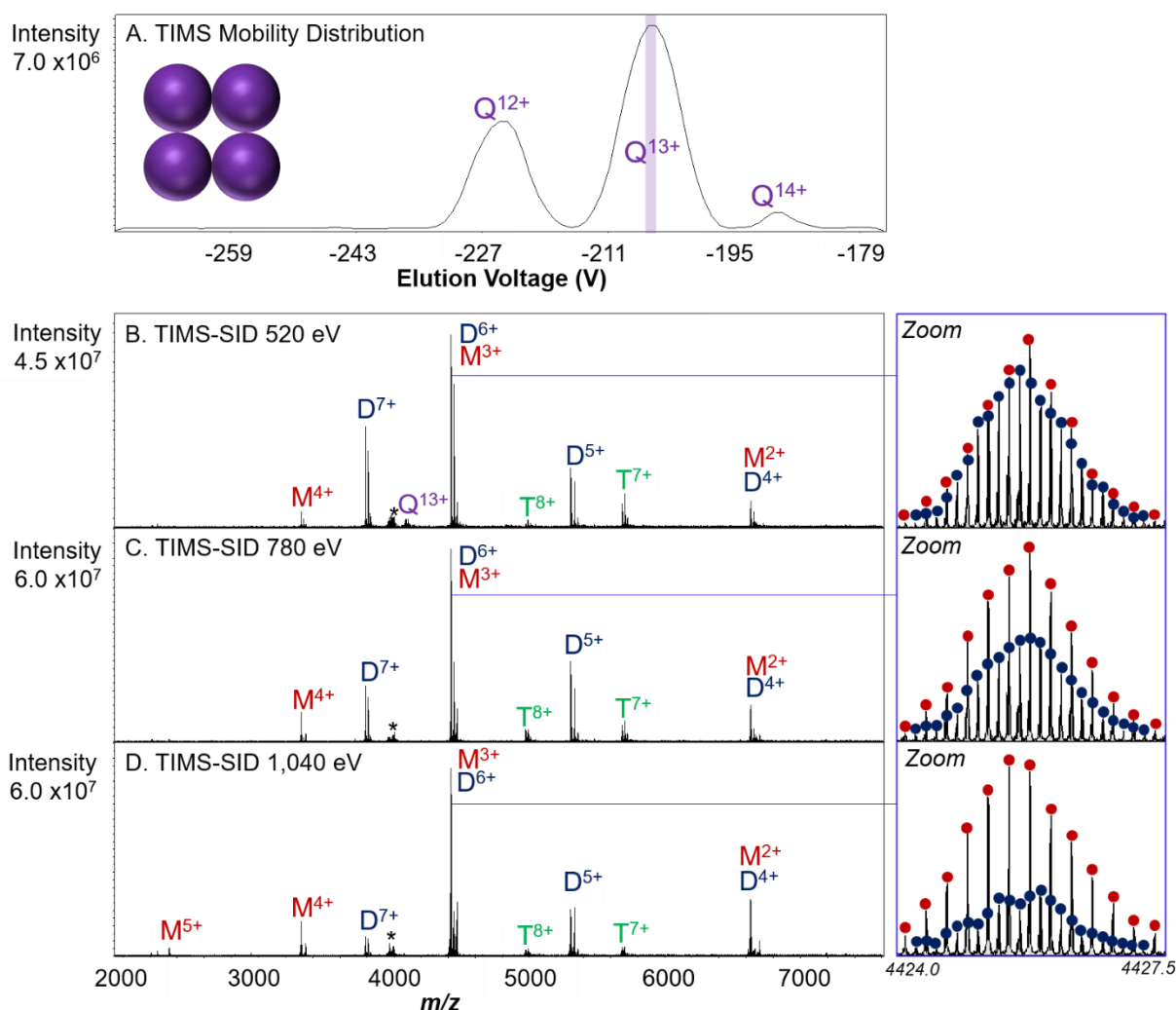
## Results

### *Application of TIMS-SID for Biological Samples*

#### *TIMS-SID of Streptavidin Tetramer*

Streptavidin, a 53 kDa homotetramer, is a dimer of dimers; therefore, SID yields predominately compact dimers at low energies. At higher SID laboratory collision energies, charge-symmetric, and therefore assumed to be, compact monomers and trimers form in competition with the formation of dimers.<sup>42</sup> As provided by Sigma, the streptavidin tetramer contains variable numbers of leading methionines, with zero or one on each monomer; these can be seen in the spectra for both the main series of peaks and for salt adducts. The results of the TIMS-SID-MS analysis of streptavidin, is shown in **Figure 3**. The full mass spectrum for charge-reduced streptavidin can be found in **Figure S4**. The full TIMS distribution for streptavidin yielded distinct mobility profiles for the 12+, 13+, and 14+ charge states (**Figure 3A**). In **Figures 3B-D**, TIMS was used to mobility select the 13+ charge state ( $V_{\text{elution}} = -203$  V) and then activate at three different SID energies.





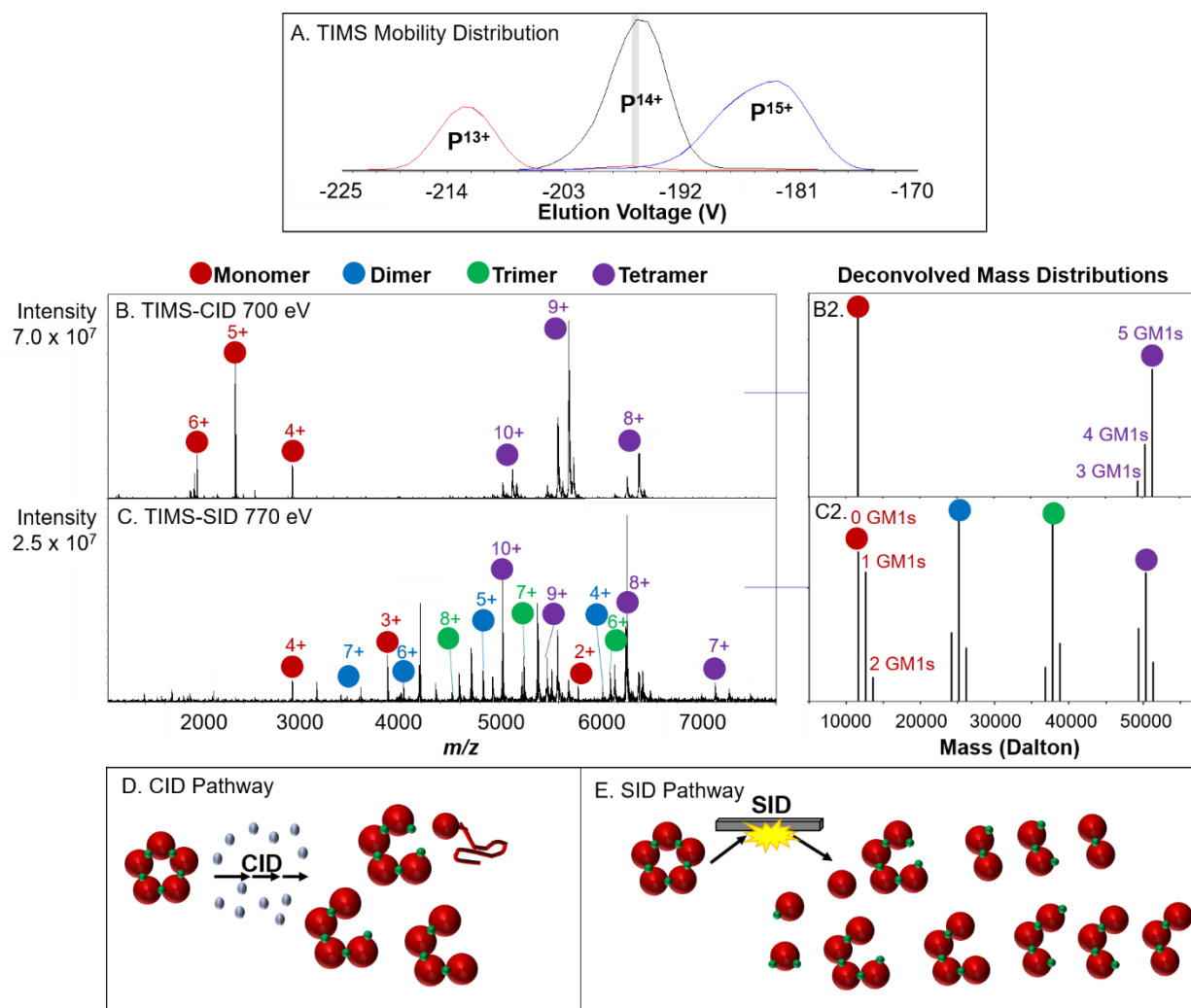
**Figure 3.** TMS mobility distribution of streptavidin tetramer with the mobility selection used for SID experiments highlighted by the purple slice (A). TMS-SID of mobility-selected 13+ streptavidin tetramer at different activation energies. At low SID energy (520 eV (B)), SID yields dimer and monomer products. As SID energy is increased to 780 eV (C) and 1,040 eV (D), trimers are also observed in the mass spectra. Even relatively low mass resolution ( $R_{\text{calculated}} = 138,000$ ) for the 15 T FT-ICR allows for identification of overlapping monomer and dimer based on isotopic distributions, as shown in the zoom-in panels. The ratio of total dimers to [monomer plus trimer] for each energy varies from 85:15 at 540 eV to 16:84 at 1040 eV. The asterisk denotes a noise peak.

TMS-SID of the mobility-selected Q<sup>13+</sup> tetramer yields mostly symmetrically charged dimer (on average, half the charge of the tetramer precursor) and symmetrically charged monomer, which can be indicative of compact subunit fragments.<sup>42</sup> As previously mentioned, streptavidin is considered a dimer of dimers, therefore dissociation based on its native interfaces should occur between the two dimers. As SID energy is increased, however, the complex dissociates into compact, low charge (5+, 4+, 3+, and 2+) monomers and their complementary trimers (Figure 3). Because of the high resolving power of the 15 T FT-ICR,

overlapping oligomers were also identified based on their isotopic distributions. For example, TIMS-SID of streptavidin shown in **Figure 3B** resulted in overlapping 3+ monomer and 6+ dimer. The differing amounts of monomer and dimer can be observed by zooming into the spectra at this region. An increase in SID energy resulted in the production of more monomer products, presumably from dimers that were observed to decrease at higher energies (trimers were of very low abundance at these energies). A comparison between TIMS-SID and TIMS-CID can be found in **Figure SI6**. Here, we obtained high signal-to-noise ratios ( $S/N_{m/z\ 4424} = 120.3$  for streptavidin) for protein complexes, evidenced by the accurate isotopic distributions of even species of low abundance.

#### *TIMS-CID and TIMS-SID of Cholera Toxin B Pentamer (with GM1 Ligand)*

TIMS ion mobility instruments operate by holding ions stationary in an electric field for user-defined periods of time, with typically longer trapping times for higher resolution applications. It has previously been suggested that the extended trapping times and applied electric fields may increase the internal energy of ions, resulting in restructuring or even fragmentation of smaller ions.<sup>41</sup> This raises concern when performing nMS experiments, in which maintaining the native-like structure and avoiding unintentional activation is of critical importance.<sup>21, 43, 44</sup> It has since been clarified that that ion heating likely occurs due to non-intrinsic properties of TIMS (space charge effects and RF power absorption) and that careful tuning prevents structural rearrangement, therefore making TIMS a suitable technology for nMS.<sup>44</sup> For the experiments performed in this manuscript, the amplitude of the RF-confining voltage in the TIMS funnel was set as low as possible, while still trapping a sufficient amount of ions, and trapping times in the TIMS analyzer were kept low to limit structural perturbation.<sup>21</sup> To confirm the results from Bleiholder and co-workers, TIMS-SID was performed using cholera toxin B (CTB), a 58 kDa pentamer, with bound GM1s ligands and compared to TIMS-CID of the same complex (TIMS parameters were identical for both TIMS-SID and TIMS-CID experiments). The CTB complex binds five GM1s molecules, located between two neighboring subunits. The results for CTB with GM1 are shown in **Figure 4**. The full mass spectrum for charge reduced CTB with GM1s is shown in **Figure SI7**.



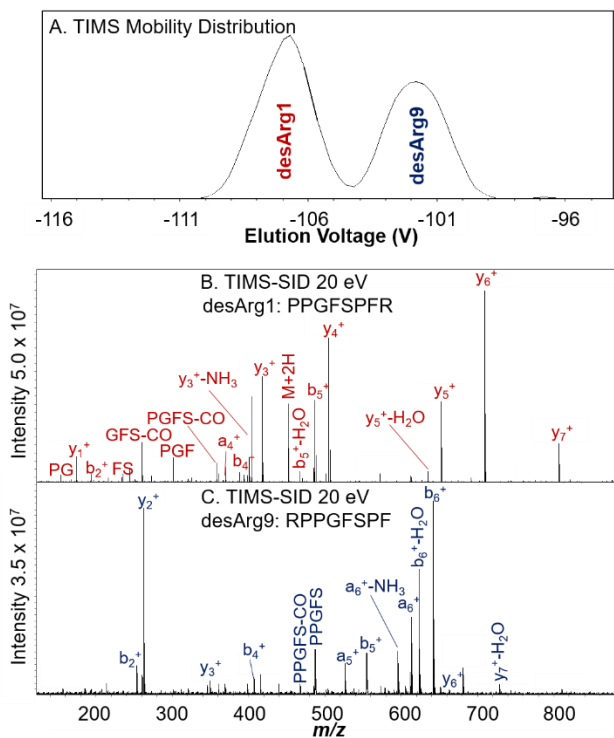
**Figure 4.** Tims mobility distribution of CTB pentamer with GM1s ligand bound with mobility selection for CID and SID experiments highlighted with gray slice (A). Tims-CID and Tims-SID of mobility-selected  $14^+$  (B, C) with corresponding deconvolution to show variations of GM1s retention on subunits (B2, C2). CID and SID pathways of CTB with GM1s and the observed products (D, E).

Pre-activation of a protein complex can often be probed by SID. Loss of native-like structure typically results in a SID spectrum that is more consistent with CID products, that is disproportionately high-charged monomer and the corresponding (n-1)mer. Additionally, pre-activation can lead to ligand loss from the complex. Tims-CID of the  $14^+$  CTB with GM1s complex yields the expected highly charged monomer and corresponding tetramer, which further undergoes ligand loss (Figure 4B, 4B2, 4D). Conversely, Tims-SID of CTB with GM1s yields monomer, dimer, trimer, and tetramer, indicative of the equal interfaces between each subunit.<sup>45</sup> For CTB with GM1s, the GM1s ligand binds between individual subunits, therefore SID results in monomer with 0, 1, or 2 GM1s bound and dimer with 1, 2, or 3 GM1s bound, and so forth (Figure 4C, 4C2, 4E).<sup>45</sup> The observation of charge-symmetric products, as well as retention of the ligand further suggests that carefully tuned Tims parameters do not result in significant,

measurable restructuring of protein complexes. Additionally, ICR instrument parameters downstream from the TIMS funnel were all tuned “softly” to prevent any inadvertent activation. We likely see less measurable rearrangement for protein complexes due to their higher barrier for intramolecular rearrangement to occur compared to thermometer ions or monomeric proteins, at least within a 15 ms time scale.<sup>46</sup>

#### *TIMS-SID of Bradykinin desArg1 and desArg9*

Bradykinin is a peptide comprised of nine amino acids (RPPGFSPFR). Removal of the C-terminal arginine (desArg9) or the N-terminal arginine (desArg1) results in two distinct peptides with the same exact mass. As a result, the two peptides cannot be separated using an analytical quadrupole. TIMS was performed on a 1:1 mixture of desArg1 and desArg9 bradykinin (**Figure 8A**) and two distinct distributions were observed for the doubly charged peptides. Each distribution was mobility-selected, and SID was performed (**Figure 5B and C**).



**Figure 5.** TIMS of a 1:1 mixture of desArg1 and desArg9 yielded baseline resolved distributions of the isobaric peptides (**A**). Each distribution was selected, and the corresponding peptide was assigned based on the SID fragmentation spectra (**B and C**).

Using TIMS, we observed baseline IMS resolution for desR1 and desR9 bradykinin, previously unachieved on a TWIMS platform.<sup>47</sup> Despite having the same nominal mass, desArg1 and desArg9 yield significantly different dissociation spectra. Most notable is the preferential formation of  $\gamma$ -type ions from desArg1 because the charge is sequestered on the C-terminal arginine. Comparatively, desArg9 preferentially forms more b-type fragment ions due to charge retention on the N-terminal arginine. Here, TIMS-SID is used for both the separation and identification of isobaric mixtures. The identification of the peptides was also confirmed by performing TIMS on the individual species (**Figure S18**). It was observed that the lower mass peptides were more sensitive to RF confining amplitude, as evidenced by intact peptide dissociating in the TIMS funnel at very high amplitudes. Low RF amplitude (70 V<sub>pp</sub> for bradykinin compared to 250 V<sub>pp</sub> for protein complexes) that did not yield any unintentional fragmentation was used for the results presented here. TIMS-SID shows promising potential of being applied for the analysis of complex mixtures, not just for nMS applications.

## Conclusions

Various protein complexes and a peptide mixture were successfully trapped and mobility-selected using trapped ion mobility spectrometry (TIMS) on an ultrahigh resolution 15T FT-ICR MS platform, further equipped with SID capabilities. The combination of TIMS and SID provided complementary structural information regarding the topology and conformation (relative drift times) of the protein complexes studied. Additionally, TIMS-SID allowed for the separation, fragmentation, and identification of peptide structural isomers present in a mixture. The SID spectra of streptavidin and CTB with GM1s ligand following IM parking were nearly identical to those from quadrupole isolation without any TIMS analysis, suggesting that little to no measurable restructuring or unfolding of protein complexes was observed during the TIMS analysis, supporting the conclusion that TIMS is applicable to nMS experiments in agreement with Bleiholder and co-workers. When analyzing peptides, however, low trapping RF V<sub>pp</sub> must be used to avoid unintentional ion activation (and possibly fragmentation). Additionally, the implementation of TIMS-SID on an ultrahigh resolution platform allows for identification and relative quantification of overlapping oligomeric SID products which would otherwise require a second stage of ion mobility for deconvolution.

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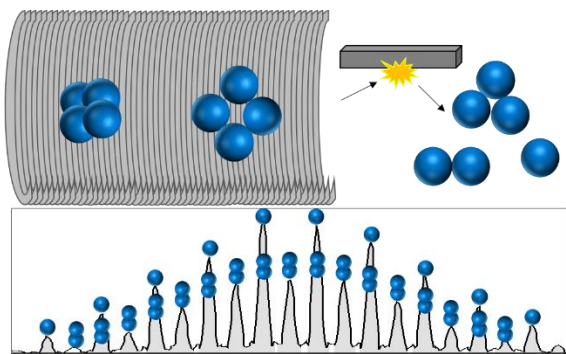
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**For TOC only**

## Surface-Induced Dissociation of Protein Complexes Selected by Trapped Ion Mobility Spectrometry

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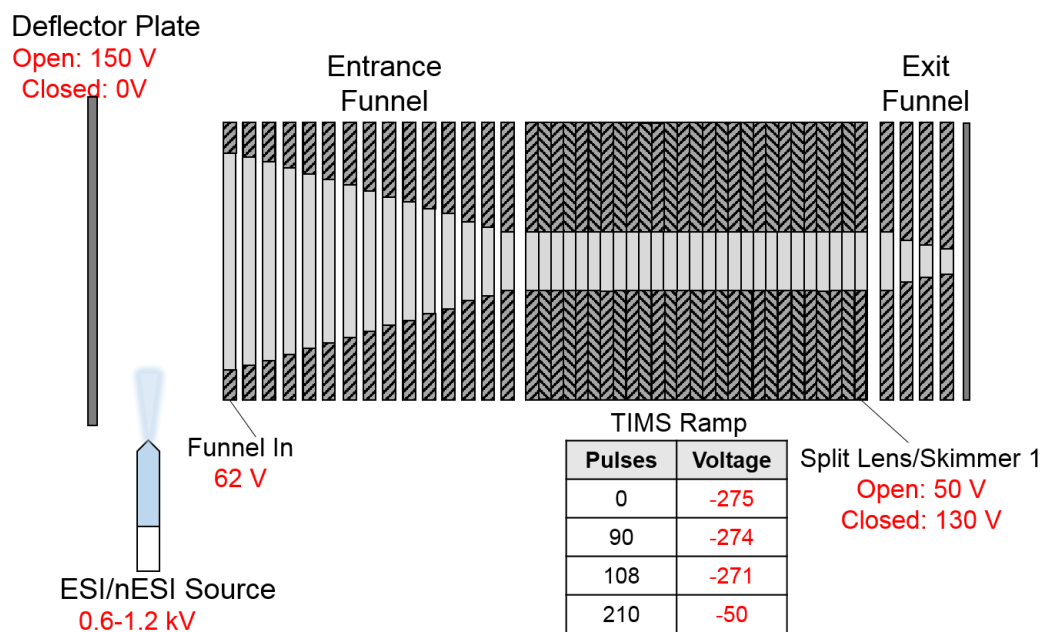
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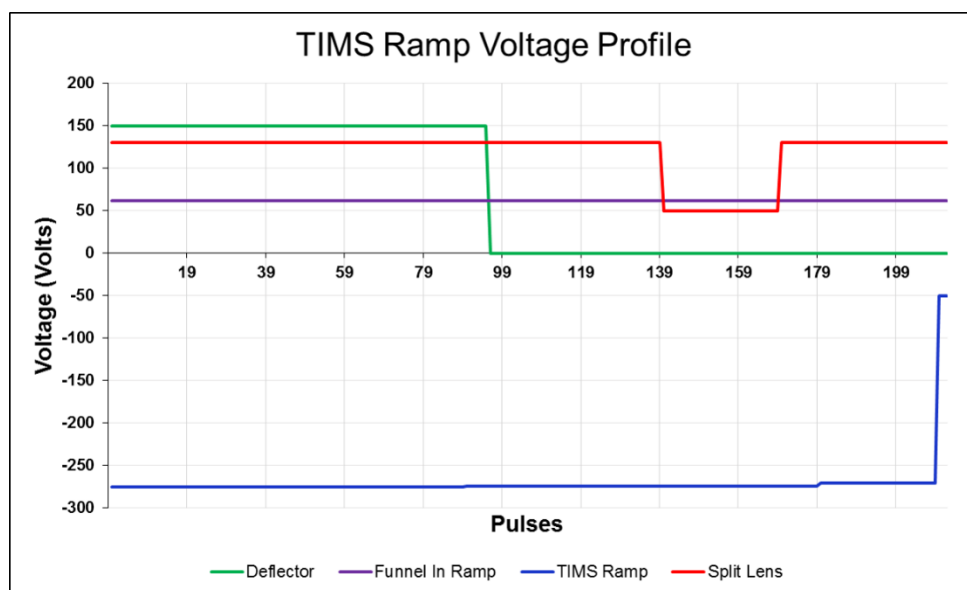
### Supplemental Information

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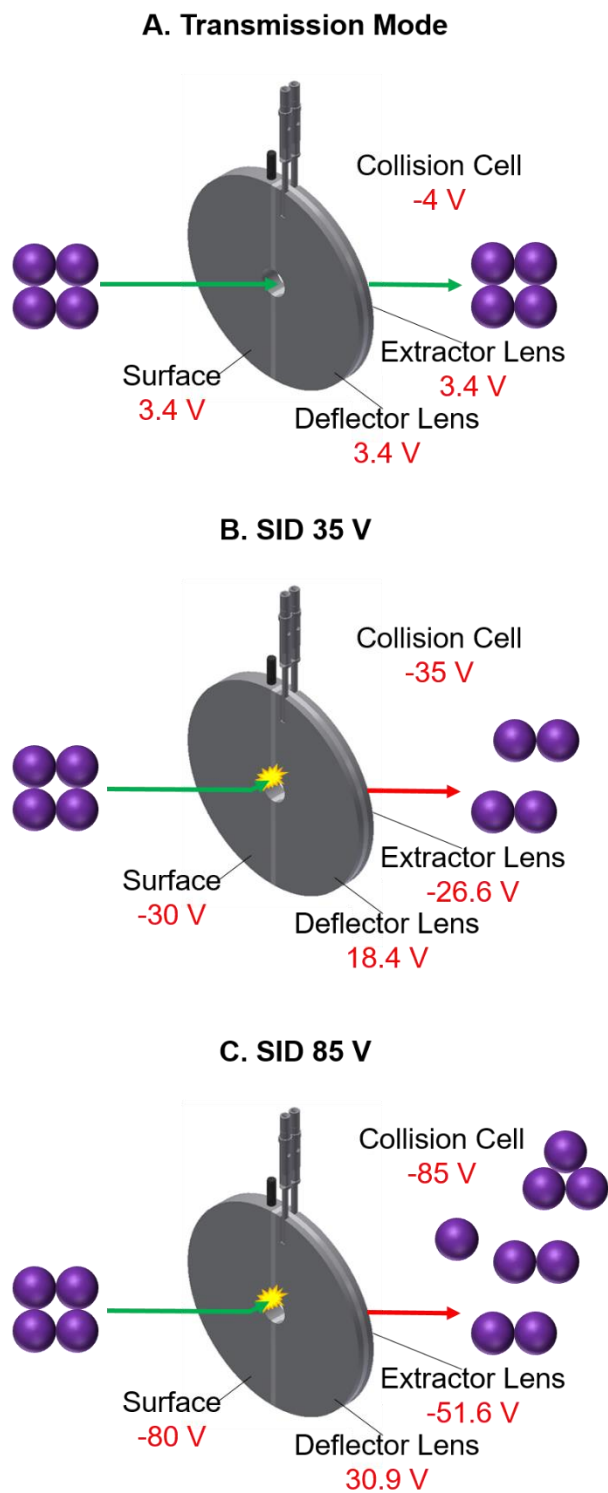
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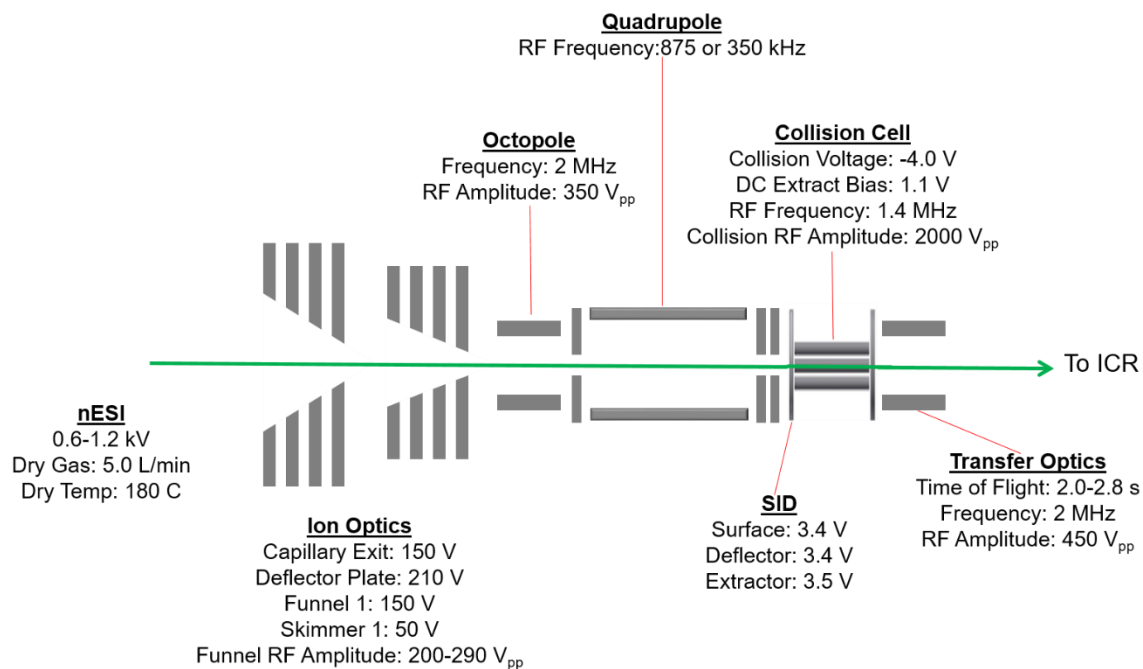
**Figure S1.** Example voltages used in the TIMS/entrance funnel region for a full mobility scan of streptavidin tetramer.



**Figure S2.** Voltage profiles used for the full mobility scan of streptavidin tetramer for a total profile of 212 pulses (34 ms). The deflector (green) was set to 150 V for ion accumulation (open) and switched to 0 V during mobility analysis. The funnel in ramp (purple) was set to 62 V for the entire profile. The TIMS ramp (blue) in the mobility analyzer spanned a range of -275 V to -50 V. The position of the split lens (red) can be altered along the TIMS ramp to only analyze ions of interest.

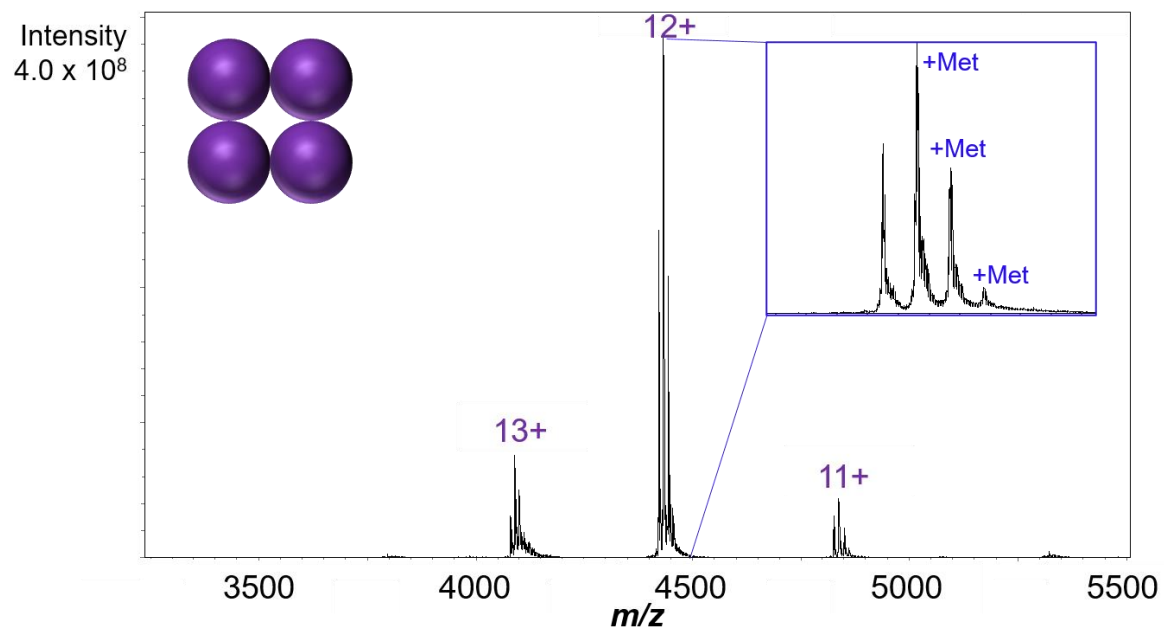


**Figure S3.** Example voltages applied to the SID device lenses and collision cell for flythrough mode (A), SID 35 V (B), and SID 85 V (C) for streptavidin tetramer. These voltages are typical for most protein complexes less than 120,000 Dalton.

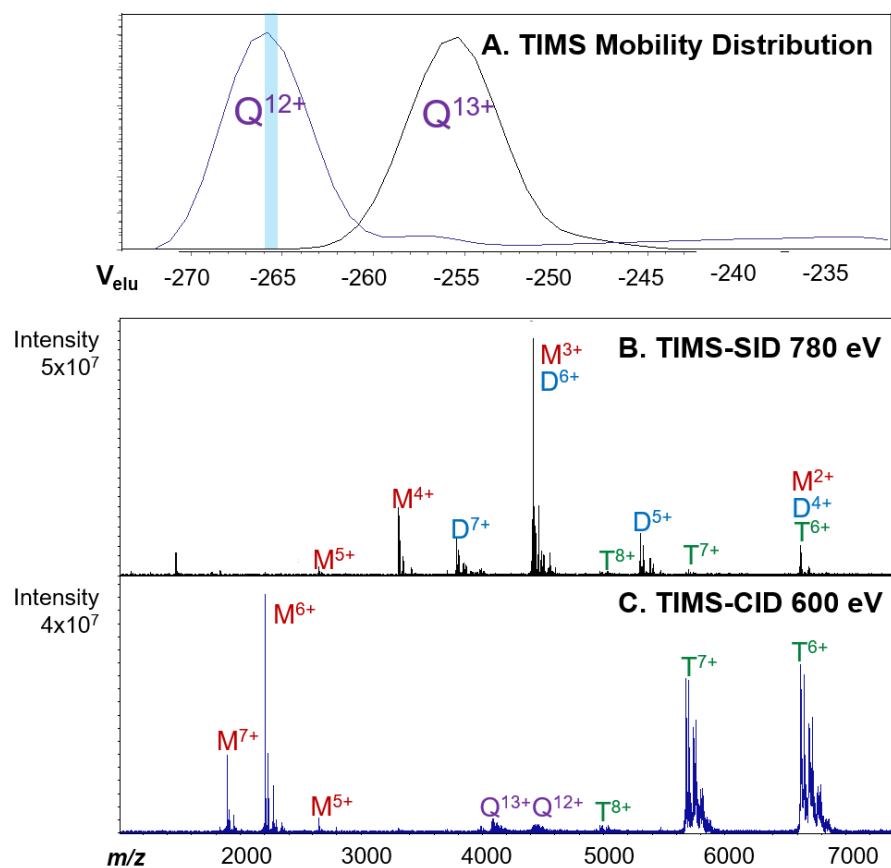


**Figure S4.** Example tune settings for the transmission and detection of streptavidin tetramer for the Bruker Solarix 15 T FT-ICR mass spectrometer. Tune settings were chosen to avoid any unintentional ion activation to retain native-like structures.

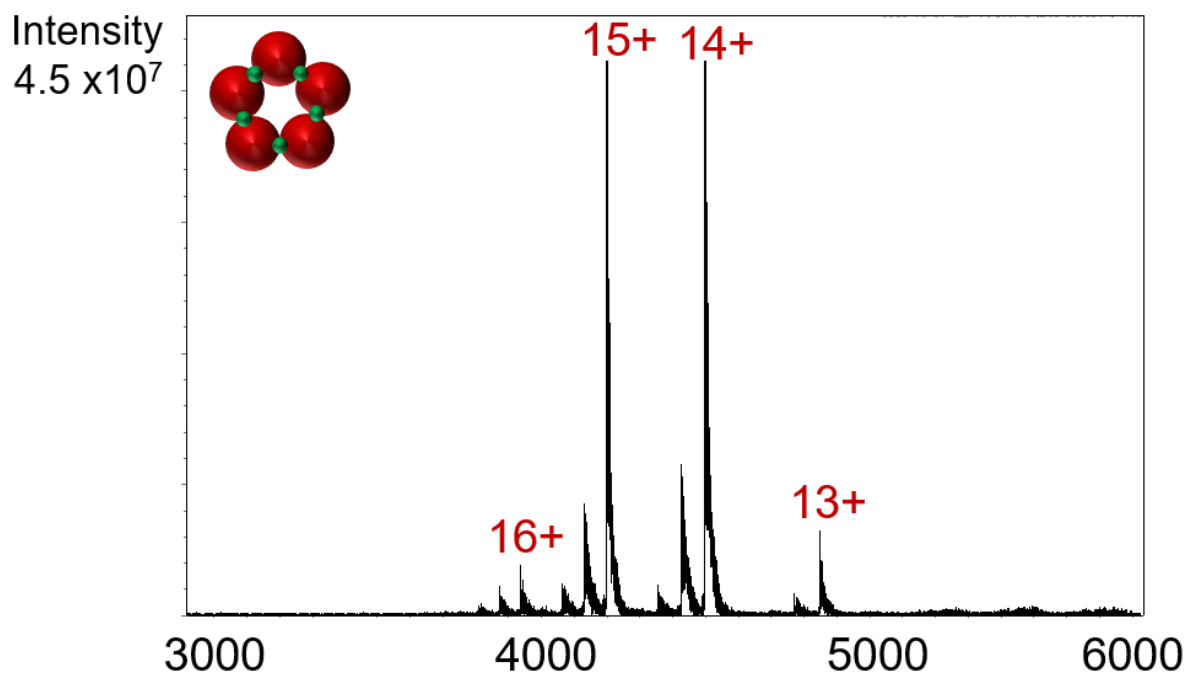




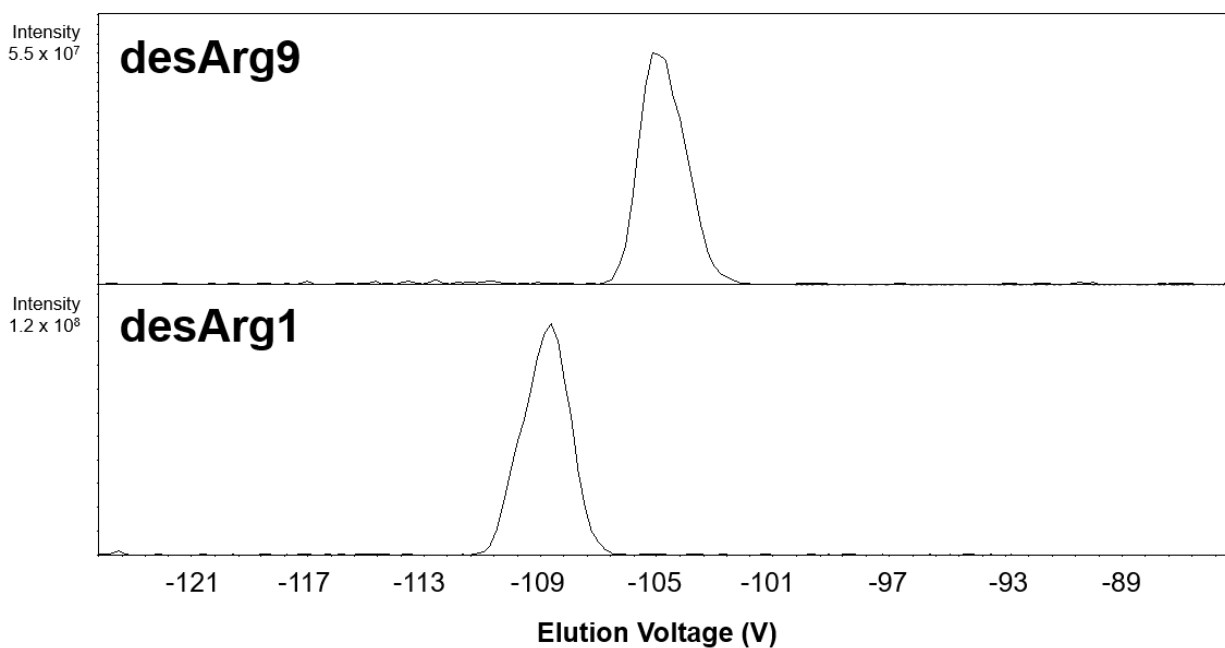
**Figure S5.** Full mass spectrum of streptavidin tetramer in 160 mM ammonium acetate and 40 mM EDDA charge-reducing agent. Individual methionine additions can be observed in the zoom-in of the  $12^+$  tetramer.



**Figure S6.** TMS mobility distribution of streptavidin tetramer in 160 mM ammonium acetate and 40 mM EDDA charge-reducing agent (A). SID (B) and CID (C) were performed using the selection of the 12+ tetramer (blue trace, A).



**Figure S7.** Full mass spectrum of cholera toxin B with GM1 in 160 mM ammonium acetate and 40 mM EDDA charge-reducing agent.



**Figure S8.** Individual mobility distributions for desArg9 and desArg1 peptides. The mobility profiles were compared to the mixture analysis shown in the main text (**Figure 5**) to confirm the correct identification of which peptide corresponded to either mobility distribution.