A Photochemical Strategy for Carbon Isotope Exchange with CO₂

Victor Babin,[†] Alex Talbot,[†] Alexandre Labiche,[†] Gianluca Destro,[†] Antonio Del Vecchio,[†] Charles S. Elmore,[‡] Frédéric Taran,[†] Antoine Sallustrau,[†] Davide Audisio[†]*

[†] Service de Chimie Bio-organique et Marquage (SCBM), CEA/DRF/JOLIOT, Université Paris Saclay, F-91191, Gif-sur-Yvette, France. [‡] Isotope Chemistry, Pharmaceutical Science, R&D, AstraZeneca, Gothenburg, Sweden

* davide.audisio@cea.fr

ABSTRACT: A novel photocatalytic approach for carbon isotope exchange is reported. Utilizing [¹³C]CO₂ as primary C1 sources, this protocol allows the insertion of the desired carbon isotope into phenyl acetic acids without the need of structural modifications or pre-functionalization, in one single step. The exceptionally mild conditions required for this traceless transformation are in stark contrast with previous methods requiring the use of harsh thermal conditions.

Developments in visible light photoredox catalysis have led to the invention of an ample array of chemical transformations, which would be either challenging or even impossible to perform under thermal conditions. Carbon-carbon bond formation represents a major challenge in photocatalysis and, in particular, the valorization of carbon dioxide (CO₂) is drawing much attention. Indeed, the functionalization of this one-carbon (C1) building block has implications that go far behind the scientific community, and affect the environment and our society as a whole.^{1, 2} Recently, successful examples of photocatalytic CO_2 functionalization have appeared.^{3,4,5}

Besides valorizing this abundant greenhouse gas, these transformations are relevant in the field of carbon isotope labeling and particularly for carbon-14 (¹⁴C), where [¹⁴C]CO₂ represents the primary source of radioisotope.⁶ ¹⁴C (β ⁻ emitter, half-life 5730 years) is the gold standard for the preparation of radiotracers utilized in human and veterinary absorption, distribution, metabolism, and excretion (ADME) determination, agrochemical and environmental fate studies.⁷ While ¹⁴C has traditionally been introduced into biologically relevant target compounds in multi-step fashion (Figure 1A), drawbacks related to the limited available raw materials, their prohibitive costs ([¹⁴C]CO₂: 1600 \in per mmol) and the generation of long-lasting waste are standing challenges.⁸

In the last couple of years, late-stage ¹⁴C-labeling has undergone sudden growth.⁹ In particular, carbon isotope exchange (CIE), which allows for ¹²C-¹²C bond cleavage and ¹²C-¹⁴C bond formation in one single step, emerged as privileged strategy (Figure 1B).¹⁰ Mostly focused on [¹²C]/[¹⁴C]CO₂ exchange,^{11,12,13} these methodologies are based on the use of transition metals, which are in some cases even required in stoichiometric amounts.¹⁴ In 2020, our group and Lundgren's independently reported the transition metal-free thermal CIE of phenyl acetic acids (PAA).¹⁵ By heating the corresponding cesium or potassium carboxylates in presence of labeled CO₂, reversible decarboxylation/carboxylation takes place and the desired acids were obtained with good isotope incorporation. While appealing, the requirement of harsh thermal heating is compulsory for non-activated PAAs. Prolonged heating at 160 °C for 48 hrs¹⁵ was required to label a series of well-known Non-Steroidal Anti-Inflammatory Drugs (NSAIDs), such as Ibuprofen, Fenoprofen, Ketoprofen, Naproxen and Diclofenac.¹⁶ Drastic conditions for an extended period of time are generally unsuitable, and especially when handling of radioactive materials is involved.

A Previous multi-step methods:



Figure 1: State of the art. (A) Multi-step carbon labeling technologies for the insertion of the isotope into a pharmaceutically relevant PAA. (B) Reported CIE procedures. (C) Photocatalytic CIE. Blue circles represent the positions of isotopically labeled carbon. PC= photocatalyst; SET= single electron transfer.

The invention of a reversible carboxylation process under mild conditions still constitutes a fundamental challenge. Herein, we report the first photocatalytic approach for carbon isotope exchange (Figure 1C). This protocol allows the insertion of the desired isotope into phenyl acetic acids, including non-natural phenyl glycine amino acids, at 42 °C only and without the need of structural modifications or pre-functionalization. These exceptionally mild conditions stand in stark contrast with previous methods requiring the use of brutal thermal force.¹⁷

The optimized reaction conditions for the photocatalytic CIE of carboxylic acids are shown in Table 1. Model substrate 2-phenylacetic acid **1** was labeled in 72% isolated yield and 47% isotopic enrichment (IE), in presence of the photocatalyst (PC) 4CzIPN (6 mol%), K_3PO_4 and $[^{13}C]CO_2$ as a convenient surrogate for $[^{14}C]CO_2$ in dry DMF within 6 hours. The reactions were performed employing 0.1 mmol of substrate and 0.3 mmol of $[^{13}C]CO_2$ were precisely added using the RC Tritec manifold.¹⁸ A higher IE of 62% was obtained by increasing the catalyst load to 12 mol% (entry 2), but the isolated yield diminished to 43%, while proto-decarboxylation side-product **1b** was formed in 47%. Under the reaction conditions, we noticed that PC **2** was entirely converted into **3** (4CzBnBN), which is likely to

be the active PC in the reaction. This observation is in agreement with previous reports by König and Tunge.¹⁹ Notably, the use of other photocatalysts resulted in low isotope incorporation (SI, Table S4), while only **3** provided comparable results (entry 3). To exclude PC degradation over the reaction condition, **3** was isolated from the crude mixture and successfully reengaged in photocatalytic CIE (see Table S4 SI for details). Other bases such as K_2CO_3 and CsOAc were also compatible but gave lower IE (Table 1, entries 4, 5 and Table S3), while the absence of the base resulted in no reaction (Table 1, entry 6). The use of carbonate bases was excluded to avoid isotope dilution, as potential source of unlabeled [^{12}C]CO₂.^{5a,5d,20} When the reaction was performed starting from the corresponding potassium carboxylate [^{13}C]1 was obtained in 50% IE and 94% yield (see SI). The use of other polar aprotic solvent such as DMSO provided [^{13}C]1 in 66 % IE, but drastically eroded the yield (entry 6 and Table S2). When [^{13}C]CO₂ was replaced by nitrogen complete proto-decarboxylation was observed (Table 1, entry 8). Finally, removing 4CzIPN from the reaction or the absence of light resulted in no isotope incorporation, showcasing that no background reaction occurs on non-activated substrate (Table 1, entries 9 and 10).



Table 1: Optimization of the reaction. ^a Yields were determined from the crude ¹H NMR spectra using 1,3,5- trimethoxybenzene as an internal standard. ^b Isotopic enrichments (I.E.) were determined by mass spectrometry. ^c Isolated yield. The temperature of the reaction was to 42 ± 2 °C.

With these optimized conditions in hand, we directed our studies toward the scope (Figure 2). Regardless to the position, in presence of electron donating groups on the aromatic ring, isotopic enrichment was observed in substituted phenyl acetic acids bearing alkyl [¹³C]4-5 or methoxy [¹³C]6-9 moieties. It is worth to note that higher incorporation was achieved with *ortho* substitution but a lower isolated yield was obtained ([¹³C]8, IE = 71%, 29% yield). Halogen were also tolerated and [¹³C]10-16 were labeled in 29 to 70% IE and good yields. Only substrate 13 could not be labeled and de-iodination occurred without insertion of ¹³C in the acid. Investigation with stronger electro-withdrawing groups led to successful labeling of various substrates such as trifluoromethyl [¹³C]17-19 (IE = 33-70%), *m*-nitrile [¹³C]20 (IE = 64%) and ester derivatives [¹³C]21 (IE = 63%). Importantly, on this electron poor substrates no background reaction is observed in the absence of PC.^{15b} Labeling of dicarboxylic acids 22-23 required using DMSO in place of DMF, for solubility reasons, and 2 equiv. of base. Interestingly, [¹³C]22 was selectively labeled in 30% IE, while for [¹³C]23 only a single label was recovered (IE 15%).



Figure 2: Photocatalyzed CIE labeling of phenyl acetic acids and pharmaceutical compounds. Green colored circles and numbers denote the positions of the carbon atoms labeled and the percent incorporation of the carbon isotope. DMF, *N*,*N*-dimethylformamide. [a] Using 2 equiv. of K_3PO_4 and DMSO instead of DMF. [b] ¹H NMR yield determined using DMF-d7 instead of DMF and 1,3,5-trimethoxybenzene as internal standard. [c] Reaction time: 3 h instead of 6 h. The temperature of the reaction was to 42 ± 2 °C.

It is worth noting that isotope incorporation was also possible in the presence of labile protons such as amide or alcohol [¹³C]27-29 (IE = 40-70% and 21-72% yield). Pleasingly, functionalization in benzylic position was not detrimental for the reaction and applying the procedure to such substrates could

afford the expected labeled phenyl acetic acids. The presence of alkyl substituents α to the carboxylic acid was tolerated [¹³C]30-33. While the *gem*-dimethyl **31** was effectively labeled, the presence of a cyclopropyl ring allowed only minimal exchange ([¹³C]32 IE < 7 %). CIE was performed on particularly challenging non-natural protected amino acids [¹³C]35-36 and successfully enable the exchange in 39 to 52% IE.

Next, we turned our attention to the labeling of pharmaceutically relevant derivatives, Felbinac [¹³C]37 was labeled with an isotopic enrichment of 63% and 51% yield. Diclofenac **38** and Fenclofenac **39** were obtained respectively in 49% and 57% of recovered products with 10% and 35% of isotope incorporation. The structural similarity of these drugs shows that the presence of an aniline in *ortho* position to the acid have a deleterious effect on both IE and yield. Fenoprofen **40** was labeled with 68% of IE and a correct isolated yield of 57%. To obtain labeled **41** and **42** in useful recovered amounts and avoid extensive proto-decarboxylation, a slight modification of the conditions was required. Reducing the time of the reaction from 6 to 3 hours allowed to isolate Flurbiprofen [¹³C]**41** and Naproxen [¹³C]**42** with 46% and 68% of isotopic incorporations and 55% to 30% yields, respectively.

An optimization of the reaction was performed to effectively label the most notorious NSAID on the market Ibuprofen **43**. It was found that the use 12 mol% of PC allowed an enhancement of the isotopic incorporation with a minor modification of the final isolated yield (IE = 56%, 46%). The utilization of other polar non protic solvents (DMSO and DMA) drastically reduced the isolated yield.

In conclusion, we developed the first photocatalytic carbon isotope procedure for the carbon labeling of phenyl acetic acids. This reaction proceeds under exceptionally mild reaction conditions compared to previous CIE technologies and provides a complementary approach to the challenging carbon labeling of pharmaceuticals. Implementation of this novel transformation towards radioactive ¹⁴C radiolabeling is currently ongoing in our laboratory.

Acknowledgments

This work was supported by CEA and by the European Union's Horizon 2020 research and innovation program under the Marie Sklodowska-Curie grant agreement N°675071, the European Research Council (ERC-2019-COG – 864576) and FET-OPEN No 862179. The authors thank A. Goudet, S. Lebrequier and D.-A. Buisson (DRF-JOLIOT-SCBM, CEA) for the excellent analytical support.

References

[1] (a) Aresta, M.; Dibenedetto, A.; Angelini, A., Catalysis for the Valorization of Exhaust Carbon: from CO₂ to Chemicals, Materials, and Fuels. Technological Use of CO₂. *Chem. Rev.* 2014, *114*, 1709-1742.
(b) Civiš S., Ferus M., Knížek A. (2019) Carbon Dioxide and the Effects on Climate. In: The Chemistry of CO₂ and TiO2. SpringerBriefs in Molecular Science. Springer, Cham. https://doi.org/10.1007/978-3-030-24032-5_1.

[2] For a non-comprehensive series of reviews in the field, see: (a) Börjesson, M.; Moragas, T.; Gallego, D.; Martin, R., Metal-Catalyzed Carboxylation of Organic (Pseudo)halides with CO₂. *ACS Catal.* **2016**, *6*, 6739-6749. (b) Wang, S.; Xi, C., Recent advances in nucleophile-triggered CO₂-incorporated cyclization leading to heterocycles. *Chem. Soc. Rev.* **2019**, *48*, 382-404. (c) Liu, Q.; Wu, L.; Jackstell, R.; Beller, M., Using carbon dioxide as a building block in organic synthesis. *Nat. Commun.* **2015**, *6*, 5933. (d) Huang, K.; Sun, C.-L.; Shi, Z.-J., Transition-metal-catalyzed C–C bond formation through the fixation of carbon

dioxide. *Chem. Soc. Rev.* **2011**, *40*, 2435-2452. (e) Yang, Y.; Lee, J.-W., Toward ideal carbon dioxide functionalization. *Chem. Sci.* **2019**, *10*, 3905-3926. (f) Juliá-Hernández, F.; Gaydou, M.; Serrano, E.; van Gemmeren, M.; Martin, R., Ni- and Fe-catalyzed Carboxylation of Unsaturated Hydrocarbons with CO₂. *Top. Cur. Chem.* **2016**, *374*, 45.

[3] For recent reviews, see: (a) Yeung, C. S., Photoredox Catalysis as a Strategy for CO₂ Incorporation: Direct Access to Carboxylic Acids from a Renewable Feedstock. *Angew. Chem. Int. Ed.* **2019**, *58*, 5492-5502. (b) He, X.; Qiu, L.-Q.; Wang, W.-J.; Chen, K.-H.; He, L.-N., Photocarboxylation with CO₂: an appealing and sustainable strategy for CO₂ fixation. *Green Chem.* **2020**, DOI: 10.1039/D0GC02743J.

[4] For photocatalytic carboxylations of styrenes, see : (a) Yatham, V. R.; Shen, Y.; Martin, R., Catalytic Intermolecular Dicarbofunctionalization of Styrenes with CO₂ and Radical Precursors. *Angew. Chem. Int. Ed.* **2017,** *56*, 10915-10919. (b) Ju, T.; Fu, Q.; Ye, J.-H.; Zhang, Z.; Liao, L.-L.; Yan, S.-S.; Tian, X.-Y.; Luo, S.-P.; Li, J.; Yu, D.-G., Selective and Catalytic Hydrocarboxylation of Enamides and Imines with CO₂ to Generate α,α -Disubstituted α -Amino Acids. *Angew. Chem. Int. Ed.* **2018,** *57*, 13897-13901. (c) Hou, J.; Ee, A.; Cao, H.; Ong, H.-W.; Xu, J.-H.; Wu, J., Visible-Light-Mediated Metal-Free Difunctionalization of Alkenes with CO₂ and Silanes or C(sp3)–H Alkanes. *Angew. Chem. Int. Ed.* **2018,** *57*, 17220-17224. (d) Seo, H.; Liu, A.; Jamison, T. F., Direct β -Selective Hydrocarboxylation of Styrenes with CO₂ Enabled by Continuous Flow Photoredox Catalysis. *J. Am. Chem. Soc.* **2017,** *139*, 13969-13972. (e) Liao, L.-L.; Cao, G.-M.; Ye, J.-H.; Sun, G.-Q.; Zhou, W.-J.; Gui, Y.-Y.; Yan, S.-S.; Shen, G.; Yu, D.-G., Visible-Light-Driven External-Reductant-Free Cross-Electrophile Couplings of Tetraalkyl Ammonium Salts. *J. Am. Chem. Soc.* **2018,** *140*, 17338-17342. (f) Meng, Q.-Y.; Schirmer, T. E.; Berger, A. L.; Donabauer, K.; König, B., Photocarboxylation of Benzylic C–H Bonds. *J. Am. Chem. Soc.* **2019,** *141*, 11393-11397. (g) Wang, H.; Gao, Y.; Zhou, C.; Li, G., Visible-Light-Driven Reductive Carboarylation of Styrenes with CO₂ and Aryl Halides. *J. Am. Chem. Soc.* **2019,** *141*, 11393-11397. (g) Wang,

[5] (a) Meng, Q.-Y.; Wang, S.; König, B., Carboxylation of Aromatic and Aliphatic Bromides and Triflates with CO₂ by Dual Visible-Light–Nickel Catalysis. *Angew. Chem. Int. Ed.* **2017**, *56*, 13426-13430. (b) Ye, J.-H.; Miao, M.; Huang, H.; Yan, S.-S.; Yin, Z.-B.; Zhou, W.-J.; Yu, D.-G., Visible-Light-Driven Iron-Promoted Thiocarboxylation of Styrenes and Acrylates with CO₂. *Angew. Chem. Int. Ed.* **2017**, *56*, 15416-15420. (c) Ishida, N.; Masuda, Y.; Imamura, Y.; Yamazaki, K.; Murakami, M., Carboxylation of Benzylic and Aliphatic C–H Bonds with CO₂ Induced by Light/Ketone/Nickel. *J. Am. Chem. Soc.* **2019**, *141*, 19611-19615. (d) Song, L.; Fu, D.-M.; Chen, L.; Jiang, Y.-X.; Ye, J.-H.; Zhu, L.; Lan, Y.; Fu, Q.; Yu, D.-G., Visible-Light Photoredox-Catalyzed Remote Difunctionalizing Carboxylation of Unactivated Alkenes with CO₂. *Angew. Chem. Int. Ed.* 10.1002/anie.202008630.

[6] Kamen, M. D. Early History of Carbon-14: Discovery of this Supremely Important Tracer Was Expected in the Physical Sense but Not in the Chemical Sense. *Science*, **1963**, *140*, 584-590.

[7] (a) Isin, E. M.; Elmore, C. S.; Nilsson, G. N.; Thompson, R. A.; Weidolf, L. Use of Radiolabeled compounds in drug metabolism and pharmacokinetic studies. *Chem. Res. Toxicol.* 2009, *25*, 532–542.
(b) Elmore, C. S.; Bragg, R. A. Isotope chemistry; a useful tool in the drug discovery Arsenal. *Bioorg. Med. Chem. Lett.* 2015, *25*, 167–171. (c) Marathe, P. H.; Shyu, W. C.; Humphreys, W. G. The Use of radiolabeled compounds for ADME studies in discovery and exploratory development. *Curr. Pharm. Des.* 2004, *10*, 2991-3008. (d) Maxwell, B. D.; Elmore, C. S. Eds. Radiosynthesis for ADME studies; 461–471 (John Wiley & Sons, Inc: Hoboken, 2012). (e) Elmore, C. S., The use of isotopically labeled compounds in drug discovery. *Ed. Annu. Rep. Med. Chem.* 2009, *44*, 515–534.

[8] Voges, R.; Heys, J. R.; Moenius, T. Preparation of Compounds Labeled with Tritium and Carbon-14 (John Wiley & Sons, 2009).

[9] For examples of late-stage ¹⁴C labeling, see : (a) Song, F.; Salter, R.; Chen, L. Development of decarboxylative cyanation reactions for C-13/C-14 carboxylic acid labeling using an electrophilic cyanating reagent. *J. Org. Chem.* **2017**, *82*, 3530-3537. (b) Del Vecchio, A.; Caillé, F.; Chevalier, A.; Loreau, O.; Horkka, K.; Halldin, C.; Schou, M.; Camus, N.; Kessler, P.; Kuhnast, B.; Taran, F.; Audisio, D. Late-Stage Isotopic Carbon Labeling of Pharmaceutically Relevant Cyclic Ureas Directly from CO₂. *Angew. Chem. Int. Ed.*, **2018**, *57*, 9744-9748. (c) Donslund, A. S.; Pedersen, S. S.; Gaardbo, C.; Neumann, K. T.; Kingston, L.; Elmore, C. S.; Skrydstrup, T., Direct Access to Isotopically Labeled Aliphatic Ketones Mediated by Nickel(I) Activation. *Angew. Chem. Int. Ed.* **2020**, *59*, 8099-8103. (d) Del Vecchio, A.; Talbot, A.; Caillé, F.; Chevalier, A.; Sallustrau, A.; Loreau, O.; Destro, G.; Taran, F.; Audisio, D., Carbon Isotope Labeling of Carbamates by Late-Stage [¹¹C], [¹³C] and [¹⁴C] Carbon Dioxide Incorporation. *Chem. Commun.* **2020**, 10.1039/D0CC05031H.

[10] Hinsinger, K.; Pieters, G. The emergence of carbon isotope exchange. *Angew. Chem. Int. Ed.* **2019**, *58*, 9678-9680.

[11] (a) D. Audisio, T. Cantat, G. Destro, EP18305407 (**2018**); WO 2019/193068 A1. (b) Destro, G.; Loreau, O.; Marcon, E.; Taran, F.; Cantat, T.; Audisio. D.; Dynamic carbon isotope exchange of pharmaceuticals with labeled CO₂. *J. Am. Chem. Soc.* **2019**, *141*, 780-784. (c) Tortajada, A.; Duan, Y.; Sahoo, B.; Cong, F.; Toupalas,G.; Sallustrau, A.; Loreau, O.; Audisio, D.; Martin, R. Catalytic decarboxylation/carboxylation platform for accessing isotopically labeled carboxylic acids. *ACS Catal.* **2019**, *9*, 5897–5901.

[12] For CIE using ¹⁴C-carbon monoxide starting from acid chlorides, see : Gauthier, Jr. D. R.; Rivera, N. L.; Yang, Y.; Schultz, D. M.; Shultz, C. S. Palladium-catalyzed carbon isotope exchange on aliphatic and benzoic acid chlorides. *J. Am. Chem. Soc.* **2018**, *140*, 15596–15600.

[13] For an early example of CIE procedure using strong basic conditions (LDA) in presence of HMPA, see : a) Parnes, H. A method for the preparation of ¹⁴C-labeled carboxylic acids. Synthesis of 6,11dihydro[b,e]thiepin-11-one-3-yl acetic ¹⁴C-acid. *J. Label. Compd. Radiopharm.* **1979**, *16*, 771-775.

[14] For an example of CIE using stoichiometric amounts of nickel, see : Kingston, C.; Wallace, M. A.; Allentoff, A. J.; deGruyter, J. N.; Chen, J. S.; Gong, S. X.; Bonacorsi, Jr., S.; Baran., P., S.; Direct carbon isotope exchange through decarboxylative carboxylation. *J. Am. Chem. Soc.* **2019**, *141*, 774-779.

[15] (a) Destro, G.; Horkka, K.; Loreau, O.; Buisson, D.-A.; Kingston, L.; Del Vecchio, A.; Schou, M.; Elmore, C. S.; Taran, F.; Cantat, T.; Audisio, D., Transition-Metal-Free Carbon Isotope Exchange of Phenyl Acetic Acids. *Angew. Chem. Int. Ed.* **2020**, *59* (32), 13490-13495. (b) Kong, D.; Moon, P. J.; Lui, E. K. J.; Bsharat, O.; Lundgren, R. J., Direct reversible decarboxylation from stable organic acids in dimethylformamide solution. *Science* **2020**, *369*, 557-561.

[16] For multi-step labeling of PAAs, see: (a) Horio, Y.; Torisawa, Y.; Ikegami. S. A synthesis of ¹⁴C-labeled sodium 2-[o-[2,6-dichlorophenyl)-amino]phenyl]acetate ([¹⁴C]diclofenac sodium). *Chem. Pharm. Bull.* **1985**, *33*, 5562-5564. (b) Hayball, P.J.; Nation, R.L.; Bochner, F.; Newton, J.L.; Massy-Westropp, R.A.; Hamon, D.P.G. Plasma protein binding of ketoprofen enantiomers in man: method development and its application. *Chirality*, **1991**, *3*, 460-466. (c) Corrie, J. E. T.; Munasinghe, V. R. N. Preparation of [carboxy-¹³C]-4-nitrophenylacetic acid. *J. Label. Compd. Radiopharm.* **2005**; *48*, 231-233. (d) Shackleford, D. M.; Hayball, P. J.; Reynolds, G. D.; Hamon, D. P.; Evans, A. M.; Milne, R. W.; Nation, R. L. A small-scale synthesis and enantiomeric resolution of (R,S)-[1-¹⁴C]-2-phenylpropionic acid and biosynthesis of its diastereomeric acyl glucuronides. *J. Label. Compd. Radiopharm.* **2001**, *44*, 225-234. (e) Turnbull, LB.; Johnson III, CJ.; Chen, YH.; Sancilio, LF.; Bruce, RB.; Disposition and metabolism of 4-methyl-2-(4-phenylbenzyl)-2-oxazoline-4-methanol in the rat and dog. *J. Med. Chem*, **1974**, *17*,

45-48. (f) Marsh, M.V.; Caldwell, J.; Sloan, T.P.; Smith, R.L.; The metabolism of fenclofenac in the horse. *Xenobiotica*, **1983**, *13*, 233-240.

[17] For exchange of carboxylate groups in simple aliphatic acids with labeled CO₂ under extremely harsh, pyrolytic conditions (290-440 °C), see: (a) Szabolcs, A.; Szammer, J.; Noszkó, L. A new method for the preparation of carboxyl-labelled aliphatic carboxylic acids. *Tetrahedron* **1974**, *30*, 3647-3648.
(b) Nakai, R.; Sugii, M.; Nakao, H., Isotopic Tracer Studies of the Ketonic Pyrolysis of Sodium Carboxylates. *J. Am. Chem. Soc.* **1959**, *81*, 1003-1006.

[18] Isotopic Enrichment was determined by mass spectrometry. In this study, the maximum theoretical IE is: 3 equiv. $CO_2/(3 \text{ equiv. } CO_2 + 1 \text{ equiv. } ^{12}CO_2 \text{ substrate})$ 100 = 75%.

[19] (a) Donabauer, K.; Maity, M.; Berger, A. L.; Huff, G. S.; Crespi, S.; König, B., Photocatalytic carbanion generation – benzylation of aliphatic aldehydes to secondary alcohols. *Chem. Sci.* **2019**, *10*, 5162-5166. (b) Cartwright, K. C.; Tunge, J. A., Organophotoredox/palladium dual catalytic decarboxylative Csp3–Csp3 coupling of carboxylic acids and π -electrophiles. *Chem. Sci.* **2020**, *11*, 8167-8175.

[20] See references 5a and 5d for exemples where carbonate bases, K_2CO_3 and Cs_2CO_3 , have been shown to be souce of CO_2 in photocatalytic carboxylations.