

# Biosynthesis of mycotoxin fusaric acid and application of a PLP-dependent enzyme for chemoenzymatic synthesis of substituted L-pipecolic acids

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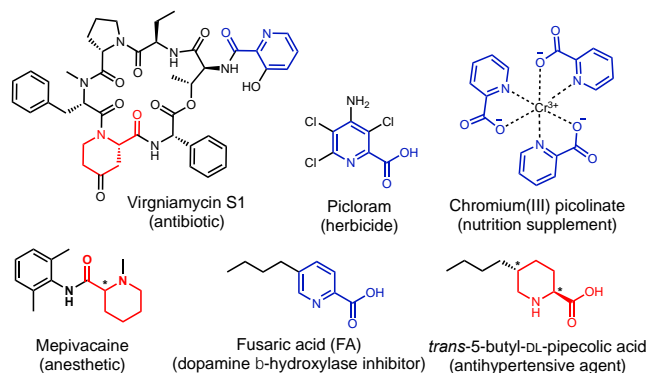
## Supporting Information Placeholder

**ABSTRACT:** Fusaric acid (FA) is a well-known mycotoxin that plays an important role in plant pathology. The biosynthetic gene cluster for FA has been identified but the biosynthetic pathway remains unclarified. Here, we elucidated the biosynthesis of FA, which features a two-enzyme catalytic cascade, a pyridoxal 5'-phosphate (PLP)-dependent enzyme (Fub7) and a flavin mononucleotide (FMN)-dependent oxidase (Fub9) in synthesizing the picolinic acid scaffold. FA biosynthesis also involves an off-line collaboration between a highly reducing polyketide synthase (HRPKS, Fub1) and a nonribosomal peptide synthetase (NRPS)-like carboxylic acid reductase (Fub8) in making an aliphatic  $\alpha,\beta$ -unsaturated aldehyde. By harnessing the stereoselective C-C bond forming activity of Fub7, we established a chemoenzymatic route for stereoconvergent synthesis of a series of 5-alkyl, 5,5-dialkyl, and 5,5,6-trialkyl-L-pipecolic acids of high diastereomeric ratio.

## INTRODUCTION

Picolinic acids and their derivatives (including the ring hydrogenated derivatives, pipecolic acids) are privileged scaffolds frequently used in pharmaceuticals and agrochemicals, and also common moieties found in many natural products (**Figure 1**).<sup>1,2</sup> As a bidentate chelator, picolinic acids are also important metal-binding pharmacophores (MBPs) that show activity against several metalloenzymes.<sup>2c</sup> Fusaric acid (FA), a picolinic acid-based fungal natural product, is a classic mycotoxin isolated from numerous *Fusarium* species (**Figure 1**).<sup>3</sup> It exhibits strong phytotoxicity and causes wilt symptoms in a variety of plants, and has long been regarded as a virulence factor of pathogenic *Fusarium* species.<sup>4</sup> FA also shows mild toxicity to animals and is found to be an uncompetitive inhibitor of mammalian dopamine  $\beta$ -hydroxylase.<sup>5a</sup> Consequently, the amidated derivative of FA, bupicomide (5-butyl-2-pyridine-carboxamide), was clinically used as a vasodilator and

experimentally as a  $\beta$ -blocker.<sup>5b</sup> Furthermore, the hydrogenated derivative of FA, *trans*-5-butyl-DL-pipecolic acid mixtures (**Figure 1**) was also shown to exhibit antihypertensive activity.<sup>6</sup>



**Figure 1.** Representative picolinic acids and pipecolic acids-containing natural products, pharmaceuticals, and agrochemicals. The racemate stereocenters are indicated with asterisks.

Despite the importance of FA in plant pathology, fungal ecology and food safety,<sup>7</sup> the biochemical basis for FA biosynthesis has not been established. Early isotope-labeled precursor feeding experiments revealed that FA was derived from aspartate (or oxaloacetate) and three acetate units (**Figure 2A**).<sup>8</sup> Recently, the well-conserved biosynthetic gene cluster for FA in multiple *Fusarium* species has been identified via transcriptomics and comparative genomics approaches (**Figure 2B**).<sup>9</sup> Gene inactivation studies showed that five genes (*fub1*, *fub4*, and *fub6-8*) were essential for FA biosynthesis, which encode a highly-reducing polyketide synthase (HRPKS, *fub1*), an  $\alpha/\beta$  hydrolase (*fub4*), a medium-chain dehydrogenase/reductase (MDR, *fub6*), a PLP-dependent enzyme (*fub7*), and a nonribosomal peptide synthetase (NRPS)-like carboxylic acid reductase (*fub8*). Addition-



**Figure 2.** Biosynthesis of fusaric acid. (A) Feeding experiment summary.<sup>8</sup> (B) Reported gene cluster *fub* in *Fusarium fujikuroi*.<sup>9b</sup> (C) Previously proposed biosynthetic pathway of FA by Studt et al.<sup>9d</sup> Domain abbreviations: KS, keto-synthase; MAT, malonyl acyltransferase; DH, dehydratase; MT<sup>0</sup>, degenerate methyltransferase domain; ER, enoyl-reductase; KR, keto-reductase; ACP, acyl-carrier protein; A, adenylation; T, thiolation; R, thioester reductase. (D) Biosynthetic pathway established in this study.

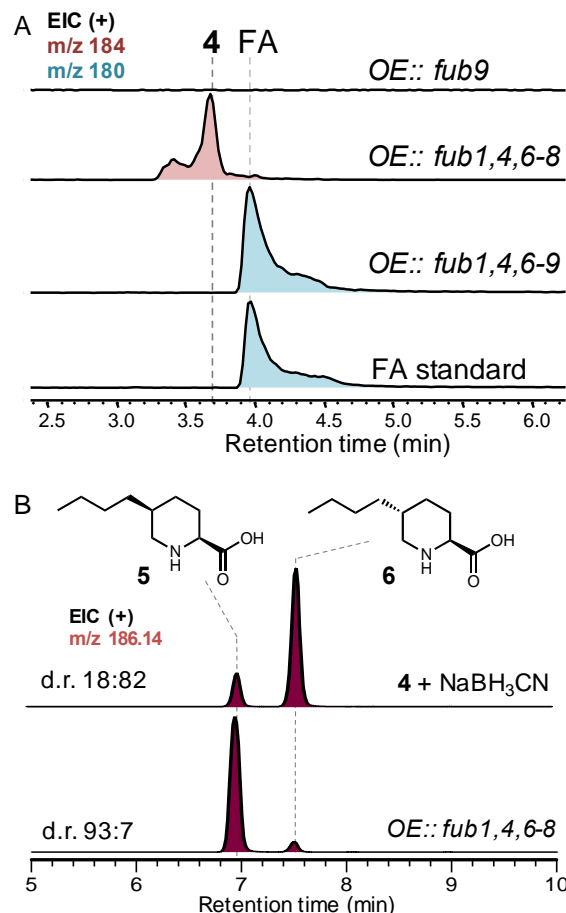
## RESULTS AND DISCUSSIONS

### Reconstitution of FA biosynthesis

To elucidate FA biosynthetic pathway, we first attempted to reconstitute FA production using the *Aspergillus nidulans*  $\Delta$ EM strain as a heterologous host.<sup>12</sup> We initially expressed only the essential genes (*fub1*, 4, 6-8) as suggested by the knockout studies. (ref. 9d) and relied on the endogenous metabolic pathway in *A. nidulans* to supply the putative amino acid precursor O-acetyl-L-homoserine (OAH).<sup>13</sup> However, we did not observe production of FA. Instead, a new compound **4** ( $m/z$  of 184 for  $[M+H]^+$ ) emerged, of which the molecular weight is in accord with the proposed tetrahydro-fusaric acid intermediate (**Figure 3A**). Derivatizing this compound with 2-aminobenzaldehyde (2AB) gave the expected dihydroquinazolinium adduct (2AB-**4**,  $m/z$  of 287  $[M+H]^+$  and  $\lambda_{max}$  at 460 nm) suggesting that this compound indeed bears an imino functional group (**Fig. S1**). Chemical reduction of **4** by  $NaBH_3CN$  afforded **5** and **6** with a diastereomeric ratio (d.r.) of 18:82 (**Figure 3B**), which were isolated and determined to be *cis*-5-butyl-L-pipecolic acid and *trans*-5-butyl-L-pipecolic acid, respectively (**Table S1, S2**). Characterization of **5** and **6** as the reduction product of **4** not only confirmed that **4** is tetrahydrofusaric acid, but also implied that **4** is epimerized at C<sub>5</sub> ( $pK_a \approx 8$ , ref. 14) presumably through facile tautomerization between the imine tautomer and the enamine tautomer **4'** (**Figure 2D**). Interestingly, we also observed **5** and **6** as shunt products from the same *A. nidulans* transformant that produced **4** when it was cultured on solid medium. The diastereomeric ratio (93:7) between **5** and **6** formed in vivo is completely reversed as compared to the ones formed by chemical reduction, which suggests that a yet-unknown endogenous imine reductase (IREd) from *A. nidulans* may preferentially catalyze reduction on the thermodynamically less favored *cis*-diastereomer of **4** (*cis*-**4**) leading to **5** as the dominant shunt product.

Nevertheless, observation of **4** instead of FA in our initial reconstitution indicates the enzyme responsible for the last 4-electron oxidation step converting **4** to FA was missing in our heterologous expression experiment. We reasoned that one of the remaining genes, *fub9*, which encodes an FMN-dependent oxidase, could be a likely candidate responsible for this oxidation event. Indeed, including *fub9* in our reconstitution successfully yielded FA (**Figure 3A**). The discrepancy between our result that Fub9 is involved in FA biosynthesis and the previous gene-inactivation study (ref. 9d), which showed *fub9* was unnecessary for FA formation in *Fusarium verticillioides*, suggests there must be a redundant pathway from **4** to FA in *F. verticillioides*. We have hence now defined the minimal set of

genes required to synthesize FA in a heterologous host and set the stage for detailed biochemical investigation of these enzymes.



**Figure 3.** Heterologous production of fusaric acid and 5-butyl-L-pipecolic acids. (A) Reconstitution of FA biosynthesis in *A. nidulans*. (B) Reduction of **4** led to pipecolic acid diastereomer **5** and **6**. Traces shown here are extracted ion chromatograms (EIC).

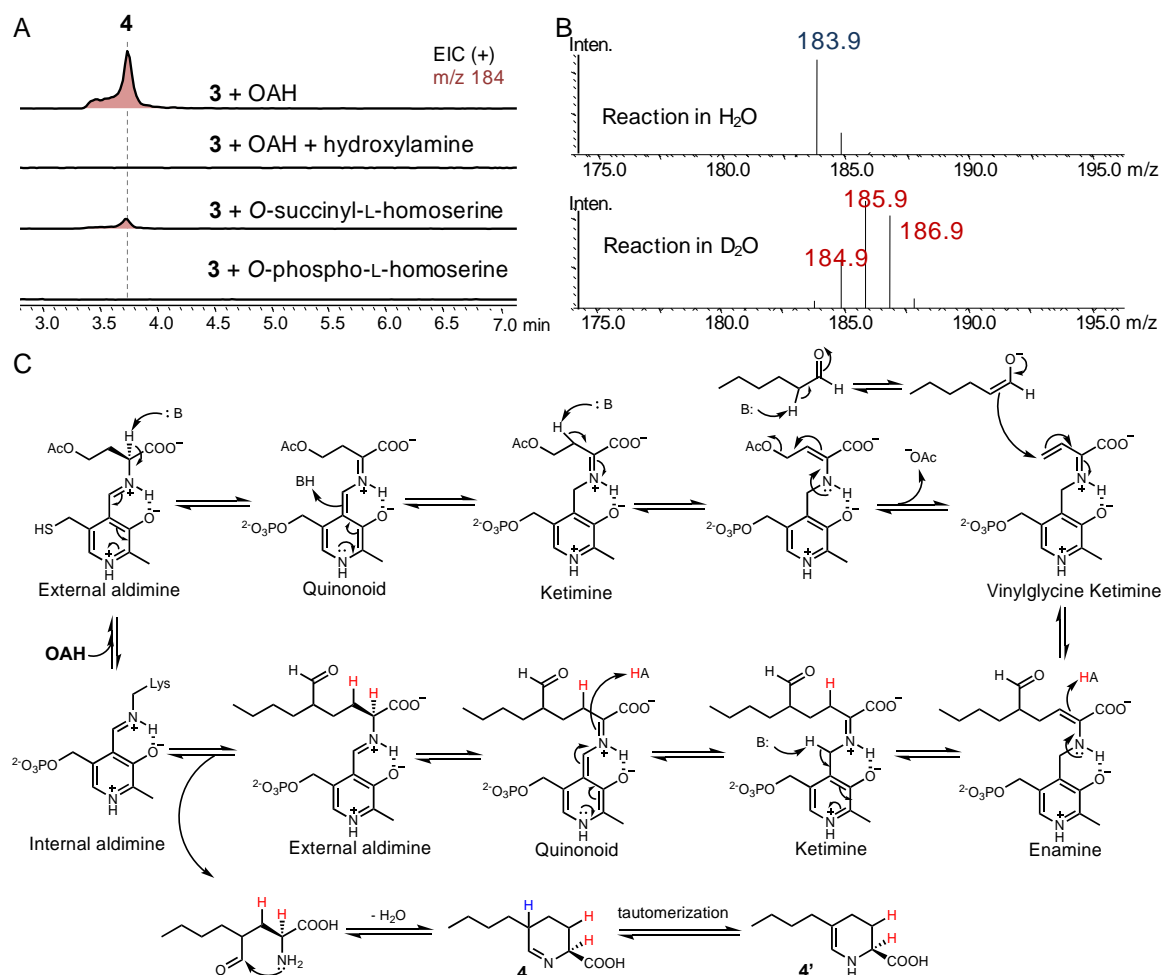
### Fub7 is a PLP-dependent enzyme catalyzing C-C bond formation

The cyclic Schiff Base **4** is in equilibrium with the (2S)-2-amino-5-formylnonanoic acid (**Figure 2D**). Retrobiosynthetic dissection of the  $\gamma$ - $\delta$  C-C bond in (2S)-2-amino-5-formylnonanoic acid revealed this compound could be synthesized from OAH and *n*-hexanal (**3**) through an apparent  $\gamma$ -substitution reaction. Such reactions can be catalyzed by PLP-dependent enzymes,<sup>11</sup> of which Fub7 is predicted to be (**Figure 2D**). To test this hypothesis, we purified recombinant Fub7 from *E. coli* and characterized its function in vitro (**Figure S3**). As-isolated Fub7 was yellow and exhibited maximal absorbance at  $\sim 425$  nm (**Figure S4a**), indicating the presence of PLP cofactor that is covalently

bound as an internal aldimine and exists in the form of ketoenamine.<sup>15</sup> Fub7 (1  $\mu$ M) readily afforded **4** in the presence of OAH (1 mM) and **3** (1 mM) as substrates (**Figure 4a**). Fub7 activity was abolished and no OAH consumption was observed when the enzyme was pre-treated with hydroxylamine (1 mM), a common inhibitor for PLP-dependent enzymes. O-succinyl-L-homoserine could also serve as the amino acid donor, albeit with much lower efficiency. Other O-substituted homoserines or S-substituted homocysteines/methionines are not substrates for Fub7 (**Figure S4b**). By using a Fub9-peroxidase-coupled assay (vide infra), we performed steady-state kinetic analysis of Fub7-catalyzed C-C bond formation between **3** and OAH (**Figure S4c**). The reaction has an apparent  $K_M$  of 1.8 mM for **3** and an apparent  $K_M$  of 15  $\mu$ M for OAH, with an overall  $k_{cat}$  of 0.14 s<sup>-1</sup>.

To test whether OAH and **3** undergo a direct displacement reaction ( $S_N2$ ), or the reaction follow an elimination-addition mechanism, we repeated the enzymatic assay in D<sub>2</sub>O. If the reaction proceeds via a  $S_N2$  pathway, only one deuterium atom (blue H in **Fig-**

**ure 4C**) should be “washed in” as a result of the non-enzymatic epimerization at C<sub>5</sub> (+1 Da mass shift of **4**). Alternatively, two additional deuterium atoms (red H in **Figure 4C**) would be incorporated if the reaction proceeds through an elimination-addition mechanism.<sup>16</sup> As shown in **Figure 4B**, a +3 Da mass shift of **4** was observed when the reaction was performed in D<sub>2</sub>O, which clearly rules out the  $S_N2$  mechanism and supports a  $\gamma$ -elimination-Michael addition pathway analogous to that of cystathionine  $\gamma$ -synthase (**Figure 4C**).<sup>16</sup> Briefly, Fub7 first generates a vinylglycine intermediate (ketimine form) from OAH, which then undergoes conjugate addition by attack from the enolate tautomer of **3** to give an enamine adduct. Protonation at C <sub>$\beta$</sub>  (where deuterium is incorporated) followed by quinonoid formation, and protonation at C <sub>$\alpha$</sub>  (where the second deuterium is incorporated) yields the final external aldimine adduct. Offloading the product and regeneration of the internal aldimine set the enzyme ready for another catalytic cycle. Released (2S)-2-amino-5-formylnonanoic acid can undergo intramolecular cyclization and dehydration to afford **4**.



**Figure 4.** Characterization of Fub7 function. (A) In vitro biochemical assay of Fub7. (B) MS analysis of Fub7 catalyzed reaction in D<sub>2</sub>O. A +3 Da mass shift of product **4** is shown. (C) Proposed catalytic mechanism of Fub7. The proton exchanged with solvent due to spontaneous epimerization is colored in blue, whereas the protons incorporated through enzyme catalyzed protonation are colored in red. All three protons can be exchanged with deuterium when the reaction is performed in D<sub>2</sub>O.



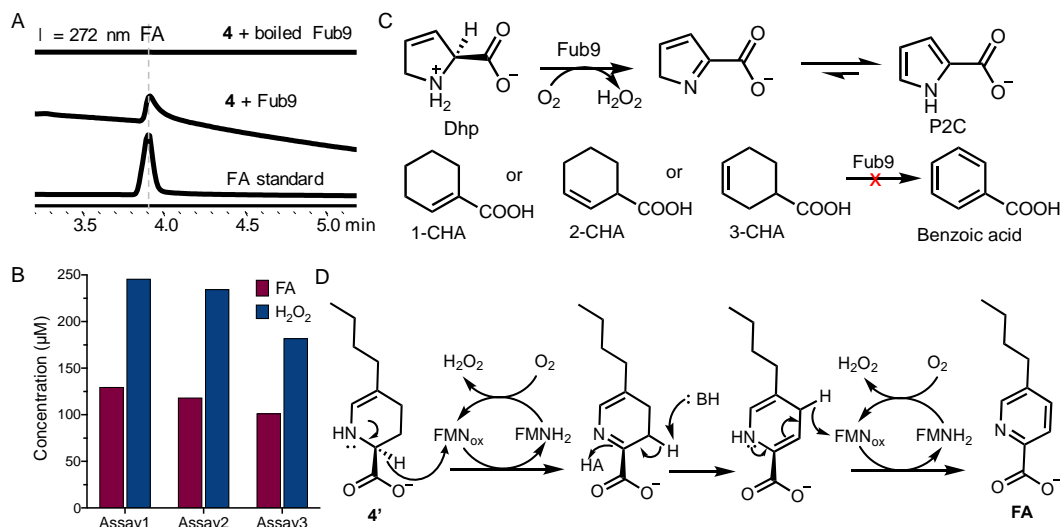
PLP-dependent aldolases generally use aldehyde substrates as electrophiles, such as threonine aldolase and serine hydroxymethyltransferase.<sup>17</sup> To our knowledge, Fub7 is the first PLP-dependent enzyme shown to use aldehyde substrates as nucleophiles. We next examined whether Fub7 can accept other nucleophiles (S, O, or N) to form carbon-heteroatom bond.<sup>18</sup> In the presence of hydrogen sulfide and OAH, Fub7 can synthesize L-homocysteine (**Figure S5**), which demonstrates its sulfhydrylase activity. Furthermore, with aliphatic alcohols and thiols as nucleophiles, Fub7 could afford an array of O-alkyl-L-homoserines and S-alkyl-L-homocysteines (**Figure S5, S6**). With aliphatic carboxylic acids as nucleophiles, Fub7 formally acts as a transacylase to convert OAH into corresponding O-acyl-L-homoserines (**Figure S7**). In contrast, Fub7 did not accept aliphatic amines (e.g. *n*-butylamine) as nucleophiles. These results together suggest Fub7 has relaxed substrate specificity towards different nucleophiles. In addition, the “unnatural” O-acyl-L-homoserines synthesized by the transacylase activity could also undergo  $\gamma$ -elimination and act as amino acid donor in synthesizing **4** (**Figure S8**). This result not only suggest above reactions all go through the elimination/addition mechanism via the same key vinylglycine ketimine intermediate, but also indicate the transacylation reaction is fully reversible and thermodynamically controlled (i.e. in equilibrium).

#### Fub9 is an FMN-dependent oxidase catalyzing dehydrogenative aromatization.

Preparation of **4** by Fub7 in vitro allowed us to further verify the function of Fub9. Purified recombinant Fub9 appeared yellow, which is attributed to the enzyme-bound cofactor FMN<sub>ox</sub> (**Figure S7**). When we added Fub9 into Fub7 reaction mixture using OAH and **3** as substrates, oxidation of **4** into FA was observed

(**Figure 5A**). By employing a horseradish peroxidase-based fluorometric assay, we also detected formation of H<sub>2</sub>O<sub>2</sub>. The stoichiometry between product FA and H<sub>2</sub>O<sub>2</sub> was determined to be 1:2 (**Figure 5B**), which clearly indicates that two equivalents of molecular oxygen are reduced to hydrogen peroxide accompanying each 4-electron oxidation of **4**. This results also demonstrates that Fub9 is a bona fide oxidase and **4** is an on-pathway intermediate en route to FA.

To gain more insight into Fub9 function, we tested several substrate analogues (**Figure 5C**). The five-membered ring analogue 3,4-dehydro-L-proline (Dhp), which mimics **4'** instead of **4**, can be oxidized to pyrrole-2-carboxylate (P2C) by Fub9. The steady-state kinetics was determined based on the rate of H<sub>2</sub>O<sub>2</sub> formation. The initial catalytic rate was not saturated even up to 10 mM Dhp (app.  $K_m > 50$  mM) and an apparent  $k_{cat}/K_m$  of 421 M<sup>-1</sup>s<sup>-1</sup> was obtained (**Figure S8**). The poor affinity to Dhp is perhaps due to the lack of alkyl substitution group on the substrate. In contrast, neither 2-cyclohexene-1-carboxylate (2-CHA) nor 3-cyclohexene-1-carboxylate (3-CHA) was a substrate for Fub9, which mimic imine **4** and enamine **4'** respectively, (**Figure S8**). These results together suggest that Fub9 specifically oxidizes C-N bond instead of C-C bond, which is reminiscent of the well-characterized flavin adenine dinucleotide (FAD)-dependent oxidase NikD involved in nikkomycin biosynthesis.<sup>19</sup> Similar to NikD, we propose the reactive substrate for Fub9 is the enamine **4'** instead of imine **4**. Upon the first hydride transfer from C<sub>2</sub> to FMN<sub>ox</sub>, Fub9 catalyzes tautomerization to yield another enamine, dihydrofusaric acid, which then transfers hydride from C<sub>4</sub> to a regenerated FMN<sub>ox</sub> to complete the dehydrogenative aromatization (**Figure 5D**).



**Figure 5.** Characterization of Fub9. (A) In vitro biochemical assay of Fub9. (B) Determining the stoichiometry of FA and H<sub>2</sub>O<sub>2</sub> by quantifying the product concentration. Three independent assays were performed. (C) Substrate analogues tested for Fub9. (D) Proposed catalytic mechanism of Fub9.

### Biosynthesis of *n*-hexanal (**3**) involves a collaboration between an HRPKS and an NRPS-like enzyme

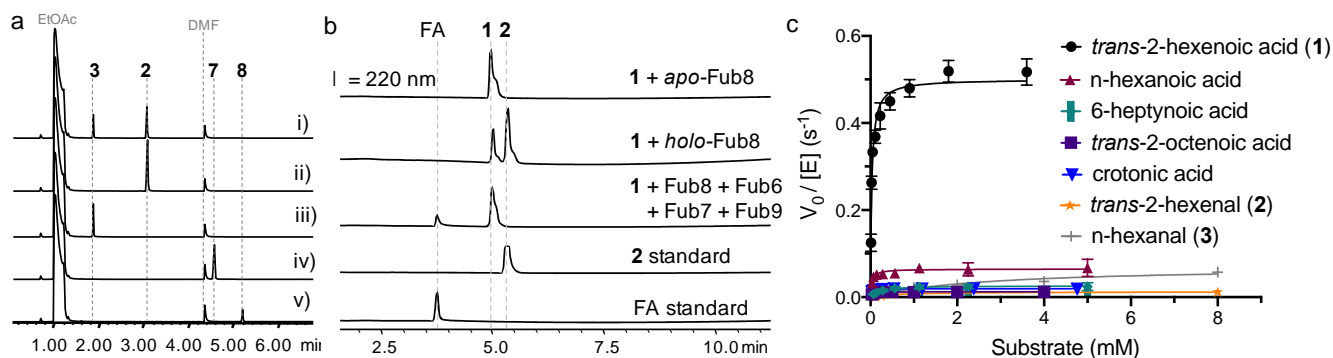
Having elucidated the biosynthetic mechanism for picolinic acid formation, we then focused on the biogenesis of the alkyl building block *n*-hexanal (**3**). Since *trans*-2-hexenal (**2**) accumulated in both  $\Delta fub6$  and  $\Delta fub7$  mutants was confirmed to be an on-pathway intermediate (ref. 9d), we reasoned that the MDR Fub6 may catalyze ene-reduction of **2** to **3**. Indeed, addition of purified recombinant Fub6 into solution containing **2** and NADPH (but not NADH) led to rapid decrease of absorbance at 340 nm, indicating oxidation of NADPH (Figure S9). Analyzing the reaction mixture by GC-MS revealed that **2** was reduced to **3** exclusively, without formation of *n*-hexanol (**7**) or *trans*-2-hexenol (**8**). Therefore, Fub6 is a strict ene-reductase and does not possess any aldehyde reductase activity (Figure 6A). Fub6 does not show any metal ion dependence and the reaction has a  $K_M$ (app) of 60  $\mu$ M for **2** and a  $k_{cat}$  of 3.1 s<sup>-1</sup> (Figure S9). Low apparent  $K_M$  implies that the intracellular concentration of the good Michael-acceptor **2** is maintained at a low level to avoid undesired nonenzymatic reactions. Other  $\alpha/\beta$ -unsaturated aldehydes, such as crotonaldehyde and cinnamaldehyde, are also substrates for Fub6, but the catalytic efficiencies are more than 10-fold lower than that for **2** (Table S3). Like other ene-reductases,<sup>20</sup> Fub6 can only reduce activated C=C double bonds, as the non-activated *trans*-2-hexenol (**8**) and the weakly-activated *trans*-2-hexenoic acid (**1**) are not substrates for Fub6 (Figure S9).

The remaining essential enzymes for FA biosynthesis, which include Fub1 (HRPKS), Fub4 ( $\alpha,\beta$ -hydro-lase), and Fub8 (NRPS-like carboxylic acid reductase), must be responsible for synthesizing *trans*-2-hexenal (**2**). We propose that Fub1 and Fub4 synthesize and hydrolytically release the triketide *trans*-2-hexenoic acid (**1**), which is then activated through adenylation and reduced by Fub8 to give **2** (Figure 2D). To test this proposal, we purified recombinant Fub8 from *E. coli* (Figure S10). Apo-Fub8 was enzymatically converted to the *holo*-form by using phosphopantetheinyl transferase NpgA as described previously.<sup>21</sup> In the presence of ATP, NADPH, and MgCl<sub>2</sub>, **1** can be reduced to **2** by *holo*-Fub8, but not *apo*-Fub8 (Figure 6B). Although prolonged incubation of Fub8 with **1** in the presence of excess amount of NADPH led to formation of over-re-

duced alcohol product **8** (Figure S10), the catalytic efficiency of Fub8 aldehyde reductase activity (**8** as substrate, app.  $k_{cat}/K_M = 19 \text{ M}^{-1}\text{s}^{-1}$ ) is three orders of magnitude lower than that of its carboxylic acid reductase activity (**1** as substrate, app.  $k_{cat}/K_M = 17,000 \text{ M}^{-1}\text{s}^{-1}$ , Table S3). Thus, like many other NRPS-like carboxylic acid reductases,<sup>22</sup> Fub8 does not have processive 2+2 electron reduction activities under physiological conditions. Successfully reconstituting Fub8 activity also allowed us to synthesize FA from **1** and OAH in one-pot using a four-enzyme cocktail (Figure 6B).

Furthermore, Fub8 is shown to be highly specific to **1** in terms of both chain length and functional groups (Figure 6C). Reduction of the saturated substrate analogue *n*-hexanoic acid is 10-fold slower than that of **1**, and neither the 2-carbon shorter crotonic acid ( $k_{cat} = 0.02 \text{ s}^{-1}$ ) or the 2-carbon longer *trans*-2-octenoic acid ( $k_{cat} = 0.01 \text{ s}^{-1}$ ) is a good substrate for Fub8. Such stringent substrate specificity of Fub8 suggests a dedicated role of Fub8 in reducing the triketide precursor **1**, proposed to be synthesized by Fub1 and Fub4. Although we are not able to reconstitute the activity of the HRPKS Fub1 and Fub4 in vitro, such combination of HRPKS and hydrolase in producing polyketides is well-documented.<sup>23</sup> Therefore, biosynthesis of **2** requires the collaborative effort between two multi-domain enzymes: an HRPKS (Fub1) and an NRPS-like carboxylic acid reductase (Fub8).

The revised biosynthetic pathway to FA is shown in Figure 2D. Unexpectedly, it takes a village (i.e. four enzymes) to biosynthesize **3**, a seemingly simple, saturated C6 aldehyde. This biosynthetic logic is intriguing, since one would not design such an atomically non-economical pathway. For instance, Fub6 could be bypassed if the HRPKS system can afford the fully saturated *n*-hexanoic acid, which is well within the biosynthetic capability of HRPKSs. Moreover, the HRPKS Fub1 could directly release the polyketide chain to yield **3** using a reductase (R) release domain as seen in the biosynthesis of betaenone B.<sup>24</sup> These strategies are all energetically more efficient as compared to the current pathway. One possible explanation why *Fusarium* species adopt the current pathway is that the free intermediates **1** and **2** may serve other purposes, such as participating in other yet-unknown metabolic pathways, or act as signaling or regulatory molecules. Future studies are required to understand this logic.



**Figure 6.** Characterization of Fub6 and Fub8. (A) GC-MS analysis of Fub6-catalyzed ene-reduction of **2** in vitro: i) reaction extract (**2**); ii) **2** standard; iii) *n*-hexanal (**3**) standard, iv) *n*-hexanol (**7**) standard, v) *trans*-2-hexenol (**8**) standard. (B) HPLC analysis of Fub8-catalyzed ATP-dependent reduction of **1**. (C) Substrate specificity of Fub8 determined through steady-state kinetics analysis.

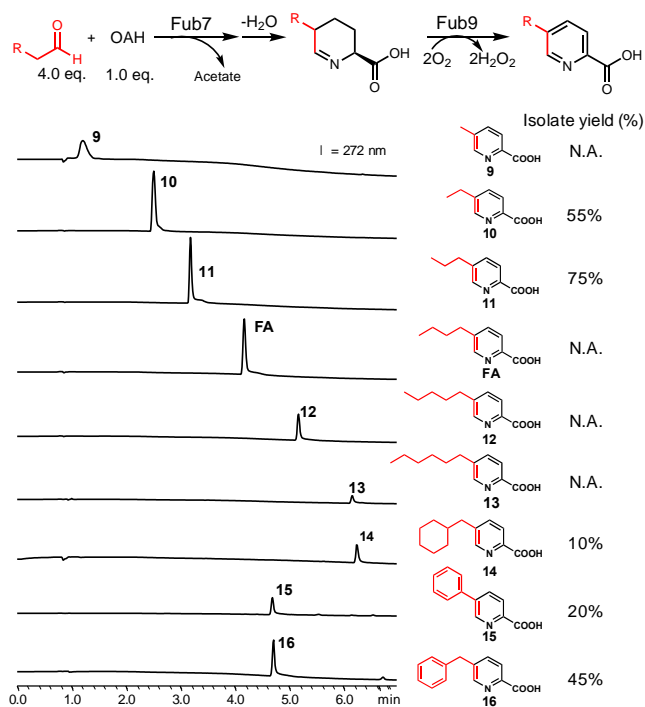
### Biocatalytic utility of Fub7

Our biochemical characterization established the biosynthetic pathway of FA, which features a four-enzyme combination to make the short-chain fully saturated aliphatic aldehyde (**3**), a PLP-dependent enzyme (Fub7) to connect **3** with an amino acid primary metabolite L-homoserine to generate the nitrogen-containing heterocycle **4**, and an oxidase to furnish the aromatic pyridine ring at last. Many of these enzymes characterized here can be useful reagents in biocatalysis and synthetic biology.<sup>20</sup> Among them, the C-C bond forming activity of Fub7 intrigued us to further explore its biocatalytic potential.

We first tested Fub7 substrate specificity using different  $\alpha$ -unsubstituted aldehyde substrates with chain length varying from C3 to C8. OAH was used as the limiting reagent in each reaction. To facilitate product detection and quantification, we coupled this assay to Fub9 enabling oxidation of any cyclic imine products into chromophoric picolinic acids. As shown in **Figure 7**, HPLC analysis indicates a series of 5-alkyl picolinic acids (**9-16**) can be synthesized using this coupled enzymatic system starting from OAH and aldehydes. Five compounds (**10**, **11**, **14-16**) were isolated from scaled-up, over-night reactions (volume varies between 10 to 30 mL depending on the yield) for structural characterization. Analysis of the reactions by LC-MS did not reveal any accumulated tetrahydropicolinic acid intermediates, therefore the yield of each product is likely limited by the substrate specificity of Fub7. Aldehydes bearing bulky groups at C $\beta$  position are not favored by Fub7 as 2-phenylacetaldehyde gave low yield (product **15**) and 2-cyclohexylacetaldehyde was completely inactive (vide infra), presumably due to increased steric hinderance impeding Fub7-catalyzed enolization of aldehydes. Overall, Fub7 exhibits somewhat relaxed substrate specificity towards  $\alpha$ -unsubstituted aliphatic aldehydes and prefers substrate chain length C4-C7.

This relaxed substrate specificity of Fub7 encouraged us to repurpose its activity for the synthesis of 5-alkyl- and 5-dialkyl-L-pipecolic acids by intercepting

Fub7-synthesized cyclic imine products through chemical reduction using NaBH<sub>3</sub>CN. Compared to the reported method in which 5-alkyl-pipecolic acids were prepared through hydrogenation of 5-alkyl-picolinic acids,<sup>6</sup> our chemoenzymatic route is advantageous in that it preserves the stereocenter at C<sub>2</sub> position (L-configuration) originated from OAH. Furthermore, we reasoned that C-C bond formation catalyzed by Fub7 should be stereoselective, which can be leveraged to make L-pipecolic acids bearing a quaternary stereocenter at C<sub>5</sub> position by starting from asymmetric  $\alpha$ -substituted aldehydes.



**Figure 7.** Biocatalytic synthesis of 5-alkyl picolinic acid derivatives. Analytical HPLC chromatograms of each reactions at small scale (200  $\mu$ L). Assay conditions: 2 mM OAH, 10  $\mu$ M Fub7, 5  $\mu$ M Fub9, 0.1  $\mu$ M catalase, and 8 mM aldehydes mixed in phosphate buffer [50 mM K<sub>2</sub>HPO<sub>4</sub> (pH 7.5), 100 mM NaCl, 3% (v/v) DMSO]

and incubated at 20 °C. Five compounds (**10**, **11**, **14-16**) were isolated for structural characterization. N.D., not determined.

As shown in **Figure 8**, a library of 5-substituted-L-pipecolic acids can be prepared chemoenzymatically. When the yield is sufficiently high, the final products were purified and structurally characterized by NMR as indicated in the figure. Diastereomeric ratios were determined by either NMR or by LCMS when the compounds can be separated. As expected, when  $\alpha$ -unsubstituted aliphatic aldehydes were tested, a pair of 2,5-diastereomers (*cis* and *trans*) were formed with the *trans*-isomer being the major product (**Figure 8**, **Figure S12**). For  $\alpha$ -branched aldehyde substrates (entry 5-12, **Figure 8**), Fub7 could accept a series of  $\alpha$ -methyl aldehydes with carbon chain length C4-C7 preferred (products **20-24**), but did not tolerate aldehydes with ethyl or any other larger substitution groups at C $_{\alpha}$ , which is also likely due to steric clash with enzyme active site residues. It is noteworthy to mention that one of the products, 5,5-dimethyl-L-pipecolic acid (**20**), is a natural metabolite involved in flavunoidine biosynthesis.<sup>25</sup> It was proposed that **20** could be synthesized from OAH and  $\alpha$ -keto-isovalerate by the PLP-dependent enzyme FlvA and imine reductase FlvB. Our result shown here shed more light on the biosynthetic mechanism of **20** that the carbon nucleophile for FlvA could be isobutylaldehyde instead of  $\alpha$ -keto-isovalerate.

Moreover, reactions with asymmetric racemate  $\alpha$ -methyl aldehyde substrates (entry 6-8, 14, **Figure 8**) all convergently yielded *trans*-5-alkyl-5-methyl-L-pipecolic acids. Because this C<sub>5</sub> quaternary stereocenter can no longer epimerize, this result strongly supports our hypothesis that Fub7 stereoselectively catalyzes C-C bond formation between aldehyde-derived enolate and PLP-bound vinylglycine ketimine. Notably, product **24** is a 5,5,6-trialkyl-L-pipecolic acid with three contiguous stereocenters, which are set through a cascade reaction triggered by Fub7: stereoselective C-C bond formation between 2,6-dimethyl-5-heptenal and OAH, followed by cyclization and dehydration; the newly-formed imine intermediate then undergoes a Pictet-

Spengler-like reaction to give the final bicyclic scaffold. Synthesis of **24** also demonstrates that the reactive Fub7-synthesized imine products can be leveraged to generate new scaffolds which could open up new opportunities in constructing more complex L-pipecolic acids.

Last but not least, Fub7 can also accept various cyclic aldehydes (entry 15-20, **Figure 8**) leading to pipecolic acids derived spirocycles (product **26-31**). The low diastereomeric ratios of product **29** and **30** indicates that significant substrate desymmetrization (appending substitution groups) will be required for Fub7 to achieve diastereoselectivity, as Fub7 does not distinguish the *re*-face and *si*-face of the asymmetric enolate substrates (entry 18-19) very well. However, additional methyl substitution could not be accommodated (entry 21-22), which further suggests that the enzyme active site is naturally tailored to best suit linear aliphatic aldehyde, such as its natural substrate **3**. Enlarging the active site by protein engineering on Fub7 may be required to recognize more bulky substrates.

## Summary

In this work, we elucidated the biosynthetic pathway for fusaric acid and characterized the enzymatic activity of four key enzymes, Fub7-9. Among these, we demonstrated the synthetic potential of Fub7 in synthesizing either 5-alkyl-picolinic acids when coupled with Fub9 or substituted pipecolic acids via a chemoenzymatic route. In particular, the stereoselective C-C bond formation catalyzed by Fub7 can be harnessed to stereoconvergent access 5-alkyl-5-methyl-L-pipecolic acids with high diastereomeric ratio. Fub7-enabled cascade reaction synthesizing **24** indicates the Fub7-synthesized asymmetric imine products can be borrowed by different downstream pathways leading to more complex structural scaffolds. Our study also demonstrated how biosynthesis study could facilitate biocatalytic innovations. Given the vast genetic potential of microbial genomes, more PLP-dependent biosynthetic enzyme will be uncovered and hold promise in biocatalysis and synthetic biology.



Entry	Aldehyde Substrates	Products	Diastereomeric Ratio <sup>a</sup>	Entry	Aldehyde Substrates	Products	Diastereomeric Ratio
1		 20% isolated yield	d.r. <sup>a</sup> = 80:20	12		Inactive	
2		 30% isolated yield	d.r. <sup>a</sup> = 85:15	13		Inactive	
3		 9% isolated yield (mixtures of diastereomers)	d.r. = 70:30	14 <sup>c</sup>		 37% isolated yield	d.r. > 95:5
4		Inactive		14		 26% isolated yield	N.A.
5		 Not isolated, low yield, identity confirmed by coelution with standard	N.A.	15		 88% isolated yield	N.A.
6		 36% isolated yield	d.r. > 95:5	16		 40% isolated yield	N.A.
7		 32% isolated yield	d.r. = 90:10	17		 15% isolated yield	N.A.
8		 15% isolated yield	d.r. > 95:5	18		 37% isolated yield (mixtures of diastereomers)	d.r. = 75:25
9		Not isolated (low yield) <sup>b</sup>		19		 39% isolated yield (mixtures of diastereomers)	d.r. = 50:50
10		Inactive		20		Inactive	
11		Inactive		21		Inactive	

**Figure 8.** Substrate scope of Fub7 in making 5-alkyl-L-pipecolic acids and 5,5-dialkyl-L-pipecolic acids. Reaction conditions: 2 mM OAH, 10  $\mu$ M Fub7, and 8 mM aldehydes mixed in phosphate buffer [50 mM  $\text{K}_2\text{HPO}_4$  (pH 7.5), 100 mM NaCl, 3% (v/v) DMSO] and incubated at 20 °C. Reaction volume varies between 20 to 40 mL depending on the yield. N.A., not applicable. <sup>a</sup>The d.r. values were analyzed by NMR or LC-MS when the diastereomers can be separated by HPLC. <sup>b</sup>Attempts to purify the target product from a 40 mL reaction did not yield enough materials for NMR characterization (<0.5 mg). <sup>c</sup> $\text{NaBH}_3\text{CN}$  was not used in this reaction.

## ASSOCIATED CONTENT

## Supporting Information.

This material is available free of charge via the Internet at <http://pubs.acs.org>.  
Experimental procedures, and spectroscopic data.

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## Notes

No competing financial interests have been declared.

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