Synthesis and Biological Evaluation of Iodinated Fidaxomicin Antibiotics

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Fidaxomicin (**1**, tiacumicin B, lipiarmycin A3) is a marketed antibiotic that is used in the treatment of *C. difficile* infections. Based on the analysis of a cryo-EM structure of fidaxomicin binding to its target enzyme (RNA-polymerase), a cation-π interaction of the aromatic moiety with an arginine residue was identified. Therefore, the variation of the substituents and concurrently changing the electronic properties of the aryl moiety represents an interesting strategy in search for new fidaxomicin analogs. Herein, we report the first semisynthetic access to new fidaxomicin analogs with varying halogen substitutents *via* a Pd-catalyzed hydrodechlorination reaction. Subsequent iodination gave access to the first iodo-fidaxomicin derivatives, which matched or improved antibacterial properties compared to fidaxomicin against *Mycobacterium tuberculosis* and *Staphylococcus aureus* ATCC 29213.

Keywords: fidaxomicin • antibiotics • iodination • semisynthesis • natural products

Introduction

Fidaxomicin (**1**, tiacumicin B, lipiarmycin A3) constitutes a glycosylated macrocyclic lactone first discovered in 1972 in a culture of *A. deccanensis*[1–3] and later re-isolated several times over the years.[4–11] This complex natural product has been introduced in the clinic and was reported to display superior properties compared to vancomycin and metronidazole.[12] Fidaxomicin shows a narrower spectrum of activity and it is particularly active against *C. difficile*, allowing the normal gut flora to recover.[13] Furthermore, fidaxomicin is poorly absorbed when orally administered, which reduces its side effects.[14]

Over the last decades, the number of strains exhibiting reduced susceptibility and even resistance to antibiotics is increasing steadily in the clinic,[15] which led various institutions such as governmental agencies and the World Health Organization to develop plans for action.[16][17] In the past, natural products were successfully used as lead structures for the development of new drugs with improved activities and properties.[18–20] Therefore, the synthesis of novel fidaxomicin derivatives can provide avenues to successfully combat antibiotic resistance[21] and consequently access to various fidaxomicin derivatives will be needed in future.

Fidaxomicin is an RNA-Polymerase (RNAP) inhibitor with a unique mode of action.[22–28] The recent disclosure of cryo-EM structures[29,30] of fidaxomicin binding to RNAP offers a tool for structural design of new derivatives. Notably, these cryo-EM structures reveal a cation-π interaction between the aromatic ring of fidaxomicin's homodichloroorsellinic acid moiety and the RNAP residue β'R84 (Figure 1). Determination of fidaxomicin's pK_a-value (pK_a=5.6)[31] confirms an acidic character of the phenol that will be mainly deprotonated in physiological environment and therefore result in a negative charge. Variations on the substituents of this aromatic ring will therefore influence its electronic properties and thereby alters the binding properties to the enzyme.

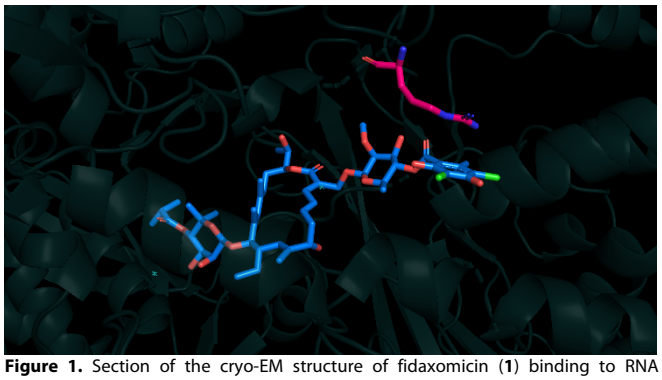
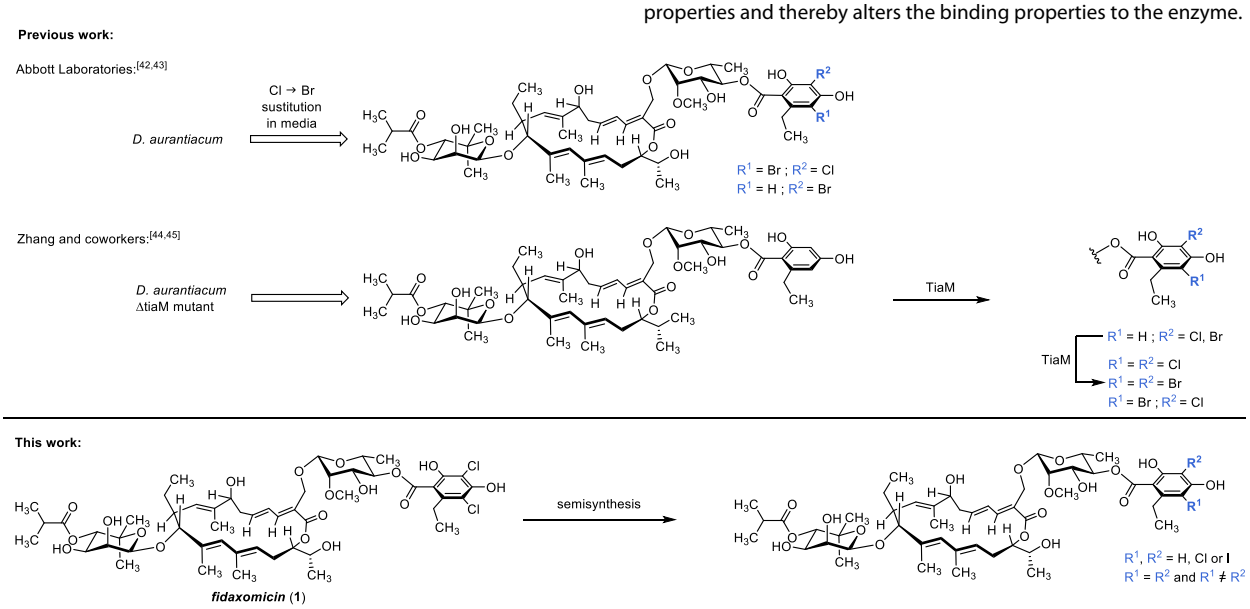


Figure 1. Section of the cryo-EM structure of fidaxomicin (**1**) binding to RNA polymerase (PDB-ID: 6FBV).[29] The cation-π interaction between β'R84 (magenta) and the dichloroorsellinate moiety of **1** (blue) is highlighted.



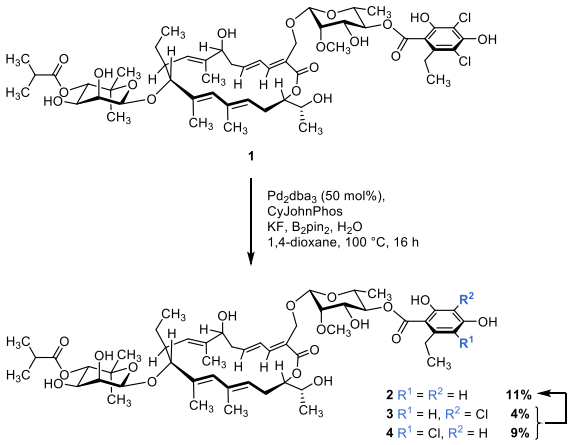
Scheme 1. Comparison of previous work and this work. tiaM = halogenase, ΔtiaM = tiaM inactivated mutant.

Besides total synthesis studies,[32–39] only a few examples of synthetic modifications on fidaxomicin are known to date.[29,40,41] Researchers from Abbott Laboratories investigated the replacement of chloride by implementation of bromide into the natural product, by supplying the fermentation broth of *D. aurantiacum* subsp. *hamdenensis* with bromide (Scheme 1).[42,43] Later, Zhang and coworkers performed gene-knockout

studies to elucidate the biosynthetic pathway.[44,45] According to their study, the halogenase TiaM is not only able to insert chloride to the homoorsellinic acid moiety, but also bromide can be introduced. However, the introduction of other halogen atoms was not achieved in this study.
In this context, we were interested in replacing the Cl-substituents from the natural product by semisynthetic approaches to study the influence of these substituents on the antibacterial properties of fidaxomicin.

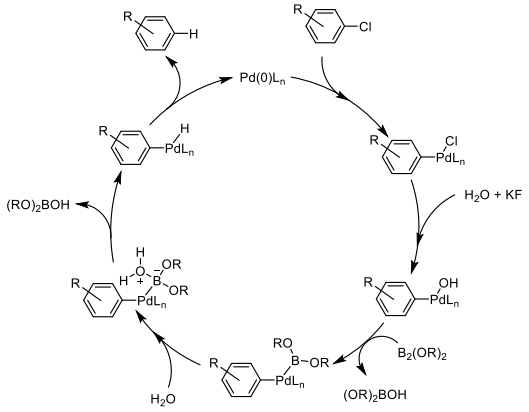
Results and Discussion

More recent results of Zhang and coworkers also showed improved activity for dechlorinated fidaxomicin derivatives.[44] Attracted by these results, we investigated a method for the preparation of such derivatives by using a semisynthetic approach from the natural product. Nevertheless, the direct transformation of aryl chlorides to aryl iodides is considered rather difficult. Indeed, exchange of a chlorine to a heavier halogen atom lead to the formation of a less stable compound (bond dissociation energies: Ph-Cl (407 kJ mol^{-1}) vs. Ph-I (280 kJ mol^{-1})).[46] Therefore, usually Cu– and Ni–mediated Finkelstein-type reactions are performed under harsh conditions (>100 °C) even for comparatively simple substrates,[46,47] potentially limiting their synthetic application with sensitive functional groups and/or scaffolds. Nevertheless, we decided to experimentally evaluate some of these Finkelstein reactions and found that either no conversion or upon increasing the temperature to above 150 °C decomposition of **1** was observed, rendering this classical approach not suitable in our case. Introduction of electron-withdrawing substituents like acetyl-protection of the phenolic hydroxy groups was not successful neither. We therefore turned our attention to the use of milder two-step procedures involving Pd-catalyzed silylations or borylations followed by an iodination step to access the desired iodinated derivatives.[48,49] To our delight, Buchwald's Pd-catalyzed silylation of aryl chlorides[49] led to formation of traces of dehalogenated fidaxomicins **2**, **3** and **4**, which indicated that the oxidative addition step into the hindered and unreactive C-Cl bonds of the homoorsellinic acid moiety took place.
Unfortunately, efforts to promote the reductive elimination using bulkier phosphine ligands such as *t*Bu-Davephos did not lead to any improvement. Nevertheless, encouraged by this result, we thus decided to study the reactivity of B_2pin_2 towards this reaction and we were pleased to obtain complete conversion to a mixture of mono- and didechlorinated compounds **2**, **3** and **4** (Scheme 2).

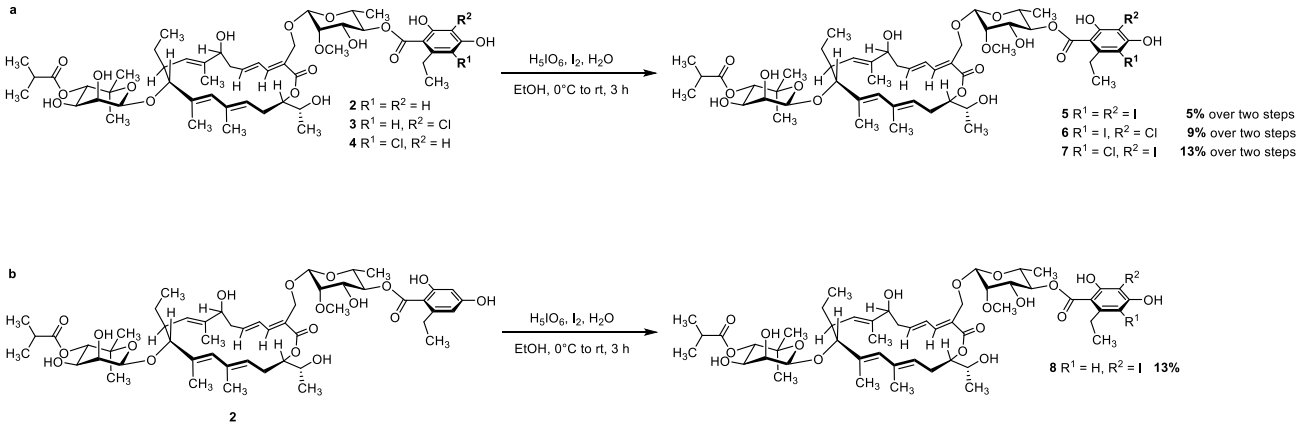


Scheme 2. Optimized conditions for the preparation of dechloro-fidaxomicin derivatives **2**–**4**.

Intrigued by the mechanism and the difference of reactivity between Me_6Si_2 and B_2pin_2, we performed control experiments in order to clarify the role of each reagent in these reaction conditions. Treatment of fidaxomicin under these conditions without water, base or the diboron source, respectively, led to the formation of only traces of the desired product, indicating their pivotal role.
Further mechanistic investigations using D_2O led to deuterium incorporation as evidenced by LC-MS measurement. Water as the potential hydride source was already reported in a reductive Heck reaction and transfer hydrogenations.[50,51] We propose that due to



Scheme 3. Proposed mechanism for the hydrodechlorination.



Scheme 4. Iodination of dechlorinated fidaxomicins **2**-**4**.

steric hindrance of the fully substituted aromatic system, reductive elimination is disfavored allowing formation of a Pd-hydride species which then undergoes reductive elimination to form the product (Scheme 3).

Having dechlorinated compounds **2**-**4** in hands, we further investigated the iodination *via* electrophilic aromatic substitution using periodic acid

and iodine,[52,53] which delivered diiodinated fidaxomicin **5** as well as monoiodinated compounds **6** and **7** (Scheme a). Due to the difficult purification of the dechlorination step, this reaction was performed on a mixture. Interestingly, mono-iodinated compound **8** was only formed when performing the reaction on purified didechlorinated fidaxomicin **2** (Scheme 4b). The biological activities were evaluated by determination of the minimum inhibitory concentration (MIC) against different strains, including *S. aureus*, *M. tuberculosis* as well as several isolates of *C. difficile* (Figure 2 and Table 1). The dechlorinated as well as iodinated fidaxomicin analogs retain their excellent activity against *C. difficile* (Figure 2). Mono-dechlorinated compounds **3** and **4**, as well as chloroiodofidaxomicin **6** and **7** display very similar activity compared to fidaxomicin (**1**) with only a slight decrease in activity. The same trend is evident against *S. aureus* and *M. tuberculosis* (Table 1). Interestingly, chloroiodofidaxomicin **7** displays improved activity against *M. tuberculosis* compared to fidaxomicin (**1**). Against S. aureus, several compounds display equal potency (e.g. **3** and **6**) or improved antiviotics activity, such as **4** and **7**.

Conclusions

In this study, we developed a semisynthetic method towards dechlorinated and iodinated fidaxomicin compounds and investigated their antibiotic activity against the Gram-positive bacterial strains *S. aureus*, *M. tuberculosis* and various isolates of *C. difficile*. These studies revealed that all derivatives maintain their excellent antibacterial properties, even though slightly decreased in comparison to the parent compound **1**. Compared to already known strategies towards fidaxomicin analogs possessing substituents different from Cl at the aromatic moiety, this investigation represents the first semisynthetic approach towards these halogenated compounds and provides a basis for further modifications which are currently under investigation in our labs.

Table 1. Minimum inhibitory concentrations [µg/mL] of compounds **1**-**8** against *M. tuberculosis* and *S. aureus*

Compound	MIC [µg/mL]	
	M. tuberculosis[a]	*S. aureus*
1	0.25	8-16
2	0.25	–
3	0.25	8
4	0.5	4
5	0.25–0.5	16
6	0.25	8
7	0.12	4-8
8	0.25–0.5	16

[a] MIC determined on GFP-expressing *M. tuberculosis* strain (see Experimental Section).[54]

Experimental Section

4‴,6‴-Didechlorofidaxomicin (2), 6‴-Dechlorofidaxomicin (3) and 4‴-Dechlorofidaxomicin (4)

A microwave tube was charged with fidaxomicin (**1**, 100 mg, 94.5 µmol, 1.0 eq.), $Pd_2dba_3{\cdot}CH_3Cl$ (43.3 mg, 41.8 µmol, 44 mol%), CyJohnPhos (49.7 mg, 0.142 mmol, 1.5 eq.), KF (11.0 mg, 189 µmol, 2.0 eq.) and B_2pin_2 (72.0 mg, 0.284 mmol, 3.0 eq.) and the flask was evacuated and flushed with argon several times. The solids were dissolved in dry dioxane (2.0 mL, degassed by freeze-pump-thaw (3x)). Then, H_2O (25 µL, 1.4 mmol, 15 eq.) was added and the mixture was allowed to stir at 100 °C for 24 h. The reaction mixture was filtered over Celite® and the solvent was evaporated. The crude mixture was pre-purified by silica-gel column chromatography (CH_2Cl_2 to MeOH/CH_2Cl_2 1:9) and the product containing fractions were further purified by preparative RP-HPLC (Gemini NX C18, 5 µ, 110 Å, 250 mm × 21.2 mm; solvent A: H_2O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH; 20 mL/min; LC time program (min –% B): 0.0 – 50%, 15.0 – 50%, 45.0 – 55%, 46.0 – 100%] to afford **2** (t_R = 14.2 min, 10.2 mg, 10.3 µmol, 11%), **3** (t_R = 17.3 min, 8.7 mg, 8.5 µmol, 9%) and **4** (t_R = 18.5 min, 4.0 mg, 3.9 µmol, 4%) as colorless solids.

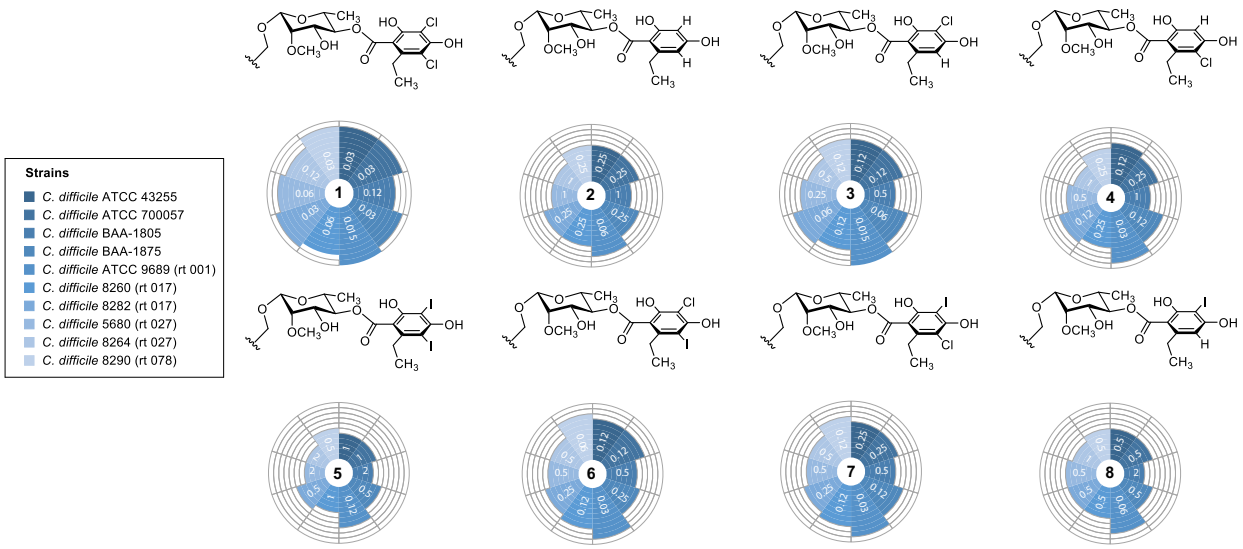


Figure 2. Minimum inhibitory concentrations [µg/mL] of compounds **1**-**8** against *C. difficile* isolates.

2: R_f = 0.44 (MeOH/CH_2Cl_2 1:18); **Specific Rotation** $[\alpha]_D^{25\,°C} = -5.75$ (c = 0.49, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3439, 2977, 2934, 2877, 1700, 1644, 1621, 1589, 1453, 1385, 1371, 1320, 1256, 1197, 1162, 1070, 1029, 1008, 901, 853 cm^{-1}; **^{1}H NMR** (500 MHz, acetone-d_6) δ 7.23 (d, *J* = 11.4 Hz, 1H), 6.68 – 6.58 (m, 1H), 6.31 (d, *J* = 2.5 Hz, 1H), 6.24 (d, *J* = 2.5 Hz, 1H), 5.96 (ddd, *J* = 14.7, 9.6, 4.6 Hz, 1H), 5.83 (s, 1H), 5.63 (t, *J* = 7.0 Hz, 1H), 5.26 – 5.20 (m, 1H), 5.12 (t, *J* = 9.7 Hz, 1H), 5.00 (d, *J* = 10.1 Hz, 1H), 4.78 (d, *J* = 1.2 Hz, 1H), 4.76 – 4.70 (m, 1H), 4.68 (d, *J* = 0.8 Hz, 1H), 4.60 (d, *J* = 11.5 Hz, 1H), 4.41 (d, *J* = 11.4 Hz, 1H), 4.30 – 4.24 (m, 1H), 4.09 – 3.94 (m, 4H), 3.84 – 3.78 (m, 2H), 3.73 (m, 3H), 3.64 – 3.59 (m, 1H), 3.57 (m, 1H), 3.51 (s, 3H), 3.33 – 3.22 (m, 1H), 2.92 – 2.83 (m, 2H), 2.81 – 2.62 (m, 3H), 2.56 (sept, *J* = 7.0 Hz, 1H), 2.52 – 2.40 (m, 2H), 1.98 – 1.90 (m, 1H), 1.81 (d, *J* = 1.3 Hz, 3H), 1.73 (d, *J* = 1.3 Hz, 3H), 1.66 (s, 3H), 1.31 – 1.24 (m, 1H), 1.27 (d, *J* = 6.2 Hz, 3H), 1.23 – 1.12 (m, 15H), 1.09 (s, 3H), 0.83 (t, *J* = 7.4 Hz, 3H) ppm; **^{13}C NMR** (126 MHz, acetone-d_6) δ 176.8, 171.7, 167.8, 165.8, 163.4, 150.6, 145.4, 143.4, 136.9, 136.12, 136.09, 133.8, 128.2, 126.3, 125.4, 124.0, 111.0, 105.3, 101.9, 101.8, 96.8, 93.3, 81.8, 78.2, 76.7, 75.7, 73.8, 72.9, 72.8, 72.4, 70.8, 70.2, 67.7, 63.4, 61.7, 42.0, 37.3, 34.8, 30.2, 28.7, 28.4, 26.5, 20.7, 19.4, 19.2, 18.6, 18.3, 17.5, 16.6, 15.2, 13.8, 11.2 ppm; **HRMS** ESI(+) (MeOH) calculated for $C_{52}H_{76}O_{18}Na$ $[M+Na]^+$: 1011.49239, found: 1011.49031.

3: R_f = 0.15 (MeOH/CH_2Cl_2 1:18); **Specific Rotation** $[\alpha]_D^{25\,°C} = -4.85$ (c = 0.51, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3440, 2977, 2934, 2876, 1700, 1645, 1605, 1417, 1372, 1314, 1258, 1198, 1146, 1071, 1028, 901, 853, 802, 695, 638, 620, 578, 554, 534, 511, 497, 477, 464, 453 cm^{-1}; **^{1}H NMR** (500 MHz, acetone-d_6) δ 7.23 (d, *J* = 11.4 Hz, 1H), 6.63 (dd, *J* = 15.0, 11.5 Hz, 1H), 6.51 (s, 1H), 5.96 (ddd, *J* = 14.7, 9.5, 4.6 Hz, 1H), 5.83 (s, 1H), 5.63 (t, *J* = 8.3 Hz, 1H), 5.22 (dt, *J* = 10.5, 1.3 Hz, 1H), 5.13 (t, *J* = 9.7 Hz, 1H), 5.00 (d, *J* = 10.1 Hz, 1H), 4.78 (s, 1H), 4.76 – 4.70 (m, 1H), 4.69 (s, 1H), 4.60 (d, *J* = 11.54 Hz, 1H), 4.42 (d, *J* = 11.4 Hz, 1H), 4.29 – 4.24 (m, 1H), 4.03 (quint, *J* = 6.3 Hz, 1H), 3.95 (d, *J* = 2.3 Hz, 1H), 3.82 (dd, *J* = 9.8, 3.4 Hz, 1H), 3.76 – 3.71 (m, 2H), 3.65 – 3.60 (m, 1H), 3.58 (d, *J* = 3.3 Hz, 1H), 3.52 (s, 3H), 2.94 – 2.78 (m, 2H), 2.77 – 2.60 (m, 3H), 2.56 (sept, *J* = 6.9 Hz, 1H), 2.51 – 2.38 (m, 2H), 1.99 – 1.91 (m, 1H), 1.81 (s, 3H), 1.73 (s, 3H), 1.66 (s, 3H), 1.30 (d, *J* = 6.1 Hz, 3H), 1.26 – 1.23 (m, 1H), 1.23 – 1.12 (m, 15H), 1.09 (s, 3H), 0.83 (t, *J* = 7.4 Hz, 3H) ppm; **^{13}C NMR** (126 MHz, acetone-d_6) δ 176.8, 171.5, 167.8, 160.8, 159.1, 148.0, 145.4, 143.4, 136.9, 136.12, 136.09, 133.8, 128.2, 126.3, 125.3, 124.0, 110.8, 106.7, 106.2, 101.8, 96.8, 93.3, 81.8, 78.2, 77.3, 75.7, 73.8, 72.86, 72.81, 72.3, 70.7, 70.2, 67.7, 63.4, 61.7, 42.0, 37.3, 34.8, 29.8, 28.7, 28.4, 26.5, 20.7, 19.4, 19.2, 18.6, 18.3, 17.5, 16.5, 15.2, 13.8, 11.2 ppm; **HRMS** ESI(+) (MeOH) calculated for $C_{52}H_{75}ClO_{18}Na$ $[M+Na]^+$: 1045.45341, found: 1045.45320.

4: R_f = 0.15 (MeOH/CH_2Cl_2 1:18); **Specific Rotation** $[\alpha]_D^{25\,°C} = -7.77$ (c = 0.95, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3432, 2977, 2934, 2876, 1698, 1645, 1610, 1580, 1450, 1384, 1370, 1318, 1243, 1199, 1155, 1069, 1028, 901, 848, 799 cm^{-1}; **^{1}H NMR** (500 MHz, acetone-d_6) δ 7.23 (d, *J* = 11.4 Hz, 1H), 6.63 (dd, *J* = 15.0, 11.5 Hz, 1H), 6.48 (s, 1H), 5.96 (ddd, *J* = 14.6, 9.5, 4.6 Hz, 1H), 5.83 (s, 1H), 5.63 (t, *J* = 8.3 Hz, 1H), 5.22 (dt, *J* = 10.5, 1.3 Hz, 1H), 5.09 (t, *J* = 9.7 Hz, 1H), 5.00 (d, *J* = 10.1 Hz, 1H), 4.78 (d, *J* = 1.2 Hz, 1H), 4.76 – 4.70 (m, 1H), 4.67 (s, 1H), 4.60 (d, *J* = 11.5 Hz, 1H), 4.42 (d, *J* = 11.5 Hz, 1H), 4.29 – 4.24 (m, 1H), 4.03 (quint, *J* = 6.2 Hz, 1H), 3.96 (d, *J* = 3.0 Hz, 1H), 3.82 (dd, *J* = 9.8, 3.4 Hz, 1H), 3.76 – 3.71 (m, 2H), 3.65 – 3.60 (m, 1H), 3.58 (d, *J* = 3.3 Hz, 1H), 3.52 (s, 3H), 2.94 – 2.78 (m, 2H), 2.77 – 2.60 (m, 3H), 2.56 (sept, *J* = 6.9 Hz, 1H), 2.51 – 2.38 (m, 2H), 1.99 – 1.91 (m, 1H), 1.81 (s, 3H), 1.73 (s, 3H), 1.66 (s, 3H), 1.28 (d, *J* = 6.2 Hz, 3H), 1.26 – 1.23 (m, 1H), 1.23 – 1.12 (m, 15H), 1.09 (s, 3H), 0.83 (t, *J* = 7.4 Hz, 3H) ppm; **^{13}C NMR** (126 MHz, acetone-d_6) δ 176.8, 170.1, 167.8, 160.9, 158.2, 145.4, 145.2, 143.5, 136.9, 136.10, 136.08, 133.8, 128.1, 126.3, 125.3, 124.0, 114.1, 109.5, 102.8, 101.8, 96.8, 93.3, 81.7, 78.2, 77.0, 75.7, 73.8, 72.9, 72.8, 72.4, 70.8, 70.2, 67.7, 63.4, 61.7, 42.0, 37.3, 34.8, 28.7, 28.3, 26.5, 26.3, 20.7, 19.4, 19.2, 18.6, 18.2, 17.5, 15.2, 14.4, 13.8, 11.2 ppm; **HRMS** ESI(+) (MeOH) calculated for $C_{52}H_{75}ClO_{18}Na$ $[M+Na]^+$: 1045.45341, found: 1045.45374.

4‴,6‴-Diiodofidaxomicin (5), 6‴-Iodofidaxomicin (6) and 4‴-Iodofidaxomicin (7)

A mixture of **2**, **3** and **4** (94.5 µmol, 1.0 eq.) was dissolved in EtOH (1.8 mL) and cooled to 0 °C. Then, I_2 (24.0 mg, 94.5 µmol, 1.0 eq.) was added in one portion and a solution of H_5IO_6 (6.5 mg, 28 µmol, 0.3 eq.) in H_2O (100 µL) was added dropwise. The reaction mixture was stirred at 0 °C for 1 h and was then allowed to slowly warm to room temperature within 2 h. Subsequently, it was diluted with EtOAc (10 mL) and then quenched with 10% aq. $Na_2S_2O_3$ (10 mL). The phases were separated, and the aqueous layer was extracted with EtOAc (3 x 10 mL). The combined organic layers were dried over $MgSO_4$, filtered and the solvent was evaporated under reduced pressure. The crude mixture was purified by preparative RP-HPLC (Gemini NX C18, 5 µ, 110 Å, 250 mm × 21.2 mm; solvent A: H_2O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH; 20 mL/min; LC time program (min –% B): 0.0 – 50%, 15.0 – 50%, 45.0 – 70%, 46.0 – 100%] to afford, after lyophilization, **5** (t_R = 25.0 min, 9.5 mg, 7.6 µmol, 9%), **6** (t_R = 27.5 min, 13.6 mg, 11.8 µmol, 13%), and **7** (t_R = 32.0 min, 6.1 mg, 5.3 µmol, 5%) as colorless solids.

5: R_f = 0.21 (MeOH/CH_2Cl_2 1:18); **Specific Rotation** $[\alpha]_D^{24\,°C} = -1.96$ (c = 0.26, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3432, 2976, 2932, 2875, 1733, 1698, 1644, 1468, 1449, 1384, 1370, 1308, 1244, 1198, 1149, 1069, 1026, 901, 799, 767 cm^{-1}; **^{1}H NMR** (500 MHz, acetone-d_6) δ 7.24 (d, *J* = 11.4 Hz, 1H), 6.68 – 6.58 (m, 1H), 5.96 (ddd, *J* = 14.7, 9.6, 4.6 Hz, 1H), 5.83 (s, 1H), 5.63 (t, *J* = 8.4 Hz, 1H), 5.22 (d, *J* = 10.5 Hz, 1H), 5.10 (t, *J* = 9.7 Hz, 1H), 5.00 (d, *J* = 10.1 Hz, 1H), 4.78 (s, 1H), 4.76 – 4.72 (m, 1H), 4.68 (s, 1H), 4.60 (d, *J* = 11.5 Hz, 1H), 4.43 (d, *J* = 11.5 Hz, 1H), 4.30 – 4.24 (m, 1H), 4.08 – 3.99 (m, 1H), 3.95 (d, *J* = 3.5 Hz, 1H), 3.85 – 3.78 (m, 1H), 3.76 – 3.69 (m, 2H), 3.65 – 3.59 (m, 2H), 3.52 (s, 3H), 3.13 – 3.02 (m, 2H), 2.79 – 2.61 (m, 3H), 2.56 (sept, *J* = 7.1 Hz, 1H), 2.53 – 2.39 (m, 2H), 1.96 – 1.90 (m, 1H), 1.81 (s, 3H), (m, 15H), 1.09 (s, 3H), 0.83 (t, J = 7.4 Hz, 3H) ppm; **^{13}C NMR** (126 MHz, acetone-d_6) δ 176.8, 169.1, 167.8, 160.6, 160.0, 149.4, 145.5, 143.5, 136.9, 136.11, 136.09, 133.8, 128.1, 126.3, 125.2, 123.9, 110.1, 101.7, 96.8, 93.3, 82.4, 81.6, 78.2, 77.6, 75.7, 73.8, 73.2, 72.9, 72.8, 72.4, 70.6, 70.2, 67.7, 63.3, 61.7, 42.0, 37.3, 34.8, 34.3, 28.7, 28.4, 26.5, 20.7, 19.4, 19.2, 18.6, 18.2, 17.5, 15.2, 14.7, 13.8, 11.2 ppm; **HRMS** ESI(+) (MeOH) calculated for $C_{52}H_{74}I_2O_{18}Na$ $[M+Na]^+$: 1263.28567, found: 1263.28567.

6: R_f = 0.23 (MeOH/CH_2Cl_2 1:18); **Specific Rotation** $[\alpha]_D^{26\,°C} = -7.68$ (c = 0.28, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3435, 2977, 2934, 2876, 1732, 1697, 1644, 1453, 1407, 1370, 1308, 1245, 1199, 1164, 1147, 1069, 1026, 901, 799 cm^{-1}; **^{1}H NMR** (500 MHz, acetone-d_6) δ 7.24 (d, *J* = 11.5 Hz, 1H), 6.70 – 6.57 (m, 1H), 5.96 (ddd, *J* = 14.7, 9.6, 4.7 Hz, 1H), 5.83 (s, 1H), 5.68 – 5.58 (m, 1H), 5.22 (d, *J* = 10.5 Hz, 1H), 5.09 (t, *J* = 9.7 Hz, 1H), 5.00 (d, *J* = 10.1 Hz, 1H), 4.78 (s, 1H), 4.73 (q, *J* = 5.3 Hz, 1H), 4.68 (s, 1H), 4.60 (d, *J* = 11.5 Hz, 1H), 4.42 (d, *J* = 11.5 Hz, 1H), 4.29 – 4.23 (m, 1H), 4.08 – 4.00 (m, 1H), 3.96 (d, *J* = 2.9 Hz, 1H), 3.843 – 3.70 (m, 3H), 3.64 – 3.57 (m, 2H), 3.52 (s, 3H), 3.11 – 3.01 (m, 2H), 2.74 – 2.61 (m, 3H), 2.56 (sept, *J* = 7.0 Hz, 1H), 2.52 – 2.39 (m, 2H), 1.97 – 1.87 (m, 1H), 1.81 (s, 3H), 1.73 (s, 3H), 1.66 (d, *J* = 1.1 Hz, 3H), 1.32 (d, J = 6.1 Hz, 3H), 1.30 – 1.23 (m, 1H), 1.22 – 1.11 (m, 15H), 1.09 (s, 3H), 0.83 (t, J = 7.4 Hz, 3H) ppm; **^{13}C NMR** (126 MHz, acetone-d_6) δ 176.8, 169.0, 167.8, 156.3, 156.1, 147.0, 145.5, 143.5, 136.90, 136.86, 136.0, 133.8, 128.1, 126.3, 125.3, 124.0, 111.7, 101.7, 96.8, 93.3, 82.6, 81.6, 78.2, 77.5, 75.7, 73.8, 72.8, 72.7, 72.3, 70.6, 70.2, 67.7, 63.4, 61.7, 42.0, 37.3, 34.8, 33.9, 28.7, 28.4, 26.5, 20.7, 19.4, 19.2, 18.6, 18.2, 17.5, 15.2, 14.8, 13.8, 11.2 ppm; **HRMS** ESI(+) (MeOH) calculated for $C_{52}H_{74}ClIO_{18}Na$ $[M+Na]^+$: 1171.35006, found: 1171.35037.

7: R_f = 0.23 (MeOH/CH_2Cl_2 1:18); **Specific Rotation** $[\alpha]_D^{26\,°C} = -4.28$ (c = 0.55, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3433, 2977, 2934, 2876, 1732, 1697, 1644, 1453, 1385, 1370, 1310, 1242, 1198, 1163, 1148, 1069, 1026, 901, 799, 713 cm^{-1}; **^{1}H NMR** (500 MHz, acetone-d_6) δ 7.24 (d, *J* = 11.5 Hz, 1H), 6.68 – 6.58 (m, 1H), 5.96 (ddd, *J* = 14.6, 9.5, 4.6 Hz, 1H), 5.83 (s, 1H), 5.65 – 5.60 (m, 1H), 5.22 (dt, *J* = 10.4, 1.6 Hz, 1H), 5.12 (t, *J* = 9.7 Hz, 1H), 5.00 (d, *J* = 10.1 Hz, 1H), 4.78 (d, *J* = 1.2 Hz, 1H), 4.76 – 4.71 (m, 1H), 4.69 (s, 1H), 4.60 (d, *J* = 11.5 Hz, 1H), 4.42 (d, *J* = 11.5 Hz, 1H), 4.30 – 4.23 (m, 1H), 4.03 (quint, *J* = 6.4 Hz, 1H), 3.96 (d, *J* = 2.9 Hz, 1H), 3.82 (dd, 1H), 3.77 – 3.68 (m, 2H), 3.67 – 3.54 (m, 2H), 3.52 (s, 3H), 3.11 – 2.96 (m, 2H), 2.79 – 2.61 (m, 3H), 2.56 (sept, *J* = 78.2, 6.7 Hz, 2H), 2.52 – 2.39 (m, 1H), 1.99 – 1.89 (m, 1H), 1.81 (d, *J* = 1.3 Hz, 3H), 1.73 (d, *J* = 1.4 Hz, 3H), 1.65 (d, *J* = 1.4 Hz, 3H), 1.30 (d, *J* = 6.1 Hz, 3H), 1.28 – 1.25 (m, 1H), 1.25 (t, *J* = 7.5 Hz, 3H), 1.20 – 1.11 (m, 12H), 1.09 (s, 3H), 0.83 (t, *J* = 7.4 Hz, 3H) ppm; **^{13}C NMR** (126 MHz, acetone-d_6) δ 176.8, 169.7, 167.8, 160.1, 157.6, 145.5, 144.9, 143.5, 136.9, 136.11, 136.08, 133.8, 128.1, 126.3, 125.2, 124.0, 113.3, 109.1, 101.7, 96.8, 93.3, 81.6, 78.2, 77.8, 75.7, 73.9, 73.8, 72.9, 72.8, 72.3, 70.6, 70.2, 67.7, 63.4, 61.7, 42.0, 37.3, 34.8, 28.7, 28.4, 26.6, 26.5, 20.7, 19.4, 19.2, 18.6, 18.3, 17.5, 15.2, 14.3, 13.8, 11.1 ppm; **HRMS** ESI(+) (MeOH) calculated for $C_{52}H_{74}ClIO_{18}Na$ $[M+Na]^+$: 1171.35006, found: 1171.35027.

4‴-Iodo, 6‴-Dechlorofidaxomicin (8)

Pure dechlorinated compound **2** (8.6 mg, 8.7 µmol, 1.0 eq.) was dissolved in EtOH (180 µL) and cooled to 0 °C. Then, I_2 (2.7 mg, 10.6 µmol, 1.2 eq.) was added in one portion and a solution of H_5IO_6 (1.2 mg, 5.3 µmol, 0.6 eq.) in H_2O (10 µL) was added dropwise. The reaction mixture was allowed to stir at 0 °C for 1 h, was diluted with EtOAc (1 mL) and then quenched with 10% aq. $Na_2S_2O_3$ (1 mL). The phases were separated, and the aqueous layer was extracted with EtOAc (3 x 2 mL). The combined organic layers were dried over $MgSO_4$, filtered and the solvent was evaporated under reduced pressure. The crude mixture was purified by preparative RP-HPLC (Gemini NX C18, 5 µ, 110 Å, 250 mm × 21.2 mm; solvent A: H_2O + 0.1% HCOOH, solvent B: MeCN + 0.1% HCOOH; 20 mL/min; LC time program (min –% B):

0.0 – 50%, 15.0 – 50%, 45.0 – 70%, 46.0 – 100%] to afford, after lyophilization, **8** (t_R = 22.0 min, 1.3 mg, 1.2 µmol, 13%) as a colorless solid.

Specific Rotation $[\alpha]_D^{26\,°C} = -0.52$ (c = 0.58, MeOH); **FT-IR** $\tilde{\nu}$ (film) 3441, 2977, 2933, 2876, 1697, 1643, 1598, 1455, 1407, 1385, 1371, 1311, 1255, 1199, 1163, 1147, 1116, 1069, 1029, 901, 846, 802, 772, 716, 629, 573, 530, 499, 474, 464 cm^{-1}; **^{1}H NMR** (500 MHz, acetone-d_6) δ 7.23 (d, *J* = 11.4 Hz, 1H), 6.69 – 6.56 (m, 1H), 6.51 (s, 1H), 5.96 (s, 1H), 5.88 – 5.78 (m, 1H), 5.63 (t, *J* = 8.3 Hz, 1H), 5.22 (dt, *J* = 10.5, 1.5 Hz, 1H), 5.14 (t, *J* = 9.7 Hz, 1H), 5.00 (d, *J* = 10.1 Hz, 1H), 4.78 (d, *J* = 1.4 Hz, 1H), 4.73 (q, *J* = 5.3 Hz, 1H), 4.69 (d, *J* = 0.9 Hz, 1H), 4.60 (d, *J* = 11.4 Hz, 1H), 4.42 (d, *J* = 11.5 Hz, 1H), 4.29 – 4.25 (m, 1H), 4.07 – 4.00 (m, 1H), 3.96 (s, 1H), 3.85 – 3.80 (m, 1H), 3.77 – 3.71 (m, 2H), 3.63 (dd, *J* = 9.6, 6.2 Hz, 1H), 3.58 (d, *J* = 3.2 Hz, 1H), 3.52 (s, 3H), 2.92 – 2.84 (m, 2H), 2.77 – 2.61 (m, 3H), 2.56 (sept, *J* = 6.8 Hz, 1H), 2.55 – 2.39 (m, 2H), 1.99 – 1.90 (m, 1H), 1.81 (d, *J* = 1.4 Hz, 3H), 1.73 (d, *J* = 1.4 Hz, 3H), 1.66 (s, 3H), 1.28 (d, *J* = 6.2 Hz, 3H), 1.24 (d, 1H), 1.21 (t, *J* = 7.4 Hz, 3H), 1.19 – 1.12 (m, 12H), 1.09 (s, 3H), 0.83 (t, *J* = 7.4 Hz, 3H) ppm; **^{13}C NMR** (126 MHz, acetone-d_6) δ 176.8, 171.5, 167.8, 164.5, 163.4, 150.3, 145.4, 143.4, 136.9, 136.12, 136.09, 133.8, 128.2, 126.3, 125.3, 124.0, 110.3, 105.2, 101.8, 96.8, 93.1, 81.8, 78.2, 77.3, 75.7, 73.8, 72.84, 72.80, 72.75, 72.3, 70.8, 70.1, 67.7, 63.4, 61.7, 42.0, 37.3, 34.8, 30.0, 28.7, 28.4, 26.5, 20.7, 19.4, 19.2, 18.6, 18.3, 17.5, 16.5, 15.2, 13.8, 11.2 ppm; **HRMS** ESI(+) (MeOH) calculated for $C_{52}H_{75}IO_{18}Na$ $[M+Na]^+$: 1137.38903, found: 1137.38932.

General Procedure for the determination of MIC values for *S. aureus*

The strain *Staphylococcus aureus* ATCC 29213 was grown overnight at 37 °C on MH II agar plates. (BD™ BBL™ Mueller Hinton II Agar, BD Diagnostics). MIC values were determined by broth dilution method according to the recommendations of the Clinical and Laboratory Standards Institute (CSLI; U.S.A.). The inoculum size was about $7.5x10^5$ colony forming units/well. The compounds were diluted in H_2O from 1.0 mg/mL stock solutions in 50 % methanol/H_2O in a 2-fold dilution series. The microtiter plates were incubated at 37 °C overnight. Afterwards, the MIC (lowest concentration of the compounds with no bacterial growth observed) was determined by visual inspection.

General procedure for the determination of MIC values for *M. tuberculosis*

MIC determination was essentially conducted as described recently.[54] Briefly, the Green-Fluorescent Protein (GFP) expressing recombinant Mycobacterium tuberculosis H37Rv rpsL[55] transformed with pOLYG-Pr-GFP[56] was grown in Middlebrook 7H9-OADC with 0.05 % Tween 80 until mid-log phase (optical density at 600 nm OD600 = 0.3 – 1.0), diluted to an OD600 of 0.04 and 20 µl of the suspension were added to an equal volume of 12-point two-fold serial dilutions of the compounds in 7H9-OADC-Tween in 384-well plates in triplicates. Compound concentrations were in the range of 62.5 to 0.031 µM. Fluorescence was measured immediately after inoculation (background) and after 10 days of incubation at 37 °C. Dose response curves were fitted with a 4-parameter log-normal model. PMIN [-,-] and PMAX [-, 120] are the minimum and the maximum, respectively, PHill [0,-] indicates the steepness, and EC50 [-,-] the log-back transformed Minimal Effective Concentration 50. The computational and statistical analysis was conducted with R (3.0.1 – 3.1.1; https://www.r-project.org/). Dose response curves were fitted with the ´drc´ package. The inhibitory potency I was calculated with the equation I = 100-[100●(S-P)/(N-P)]. S is the sample's fluorescence while P and N derive from growth inhibition with the control drug (Kanamycin A) and solvent growth control measurements (DMSO 1.25 % vol./vol.), respectively. A fluorescence reduction of 90 % as compared to the no-drug control was reported as Minimal Inhibitory Concentration (MIC90).

General procedure for the determination of MIC values for *C. difficile*

MIC determination was carried out by Micromyx, LLC, 4717 Campus Drive, Kalamazoo, MI, USA 49008.

Approximately 5 mg of each of the test compounds were provided. These were stored at -20°C until testing. On the day of the assay, the test articles were dissolved in 100% DMSO (dimethyl sulfoxide, Sigma; St. Louis, MO, Cat. No. 472301-500ML, Lot No. SHBH5551V) to a stock concentration of 3232 µg/mL. The concentration range tested for these test agents was 16 – 0.015 µg/mL. The comparator agents, metronidazole and clindamycin were supplied by Micromyx, as shown in the table below:

Comparator Drug	Supplier	Catalog No.	Lot No.	Solvent/Diluent	Testing Range (µg/mL)
Metronidazole	Sigma	M3761-100G	095K0693	DMSO/dH2O	64 – 0.06
Clindamycin	Sigma	C5269-100MG	021M1533	dH2O/dH2O	32 – 0.03

Test Organisms

Test organisms consisted of reference strains from the American Type Culture Collection (ATCC; Manassas, VA) and clinical isolates from the Micromyx repository (MMX; Kalamazoo, MI). Organisms were initially received at Micromyx and were streaked for isolation. Colonies were picked by sterile swab from the medium and suspended in the appropriate broth containing cryoprotectant. The suspensions were aliquoted into cryogenic vials and maintained at -80ºC.

Prior to testing, all isolates were streaked onto Brucella Agar supplemented with hemin, Vitamin K and 5% sheep blood (Becton Dickinson [BD]; Sparks, MD, Cat. No. 297716, Lot No. 8256909) and incubated anaerobically at 35-37°C for 44 – 48 hours.

Additionally, Bacteroides fragilis ATCC 25285 and Clostridium difficile ATCC 700057 were tested for purposes of quality control.

Test Medium

The medium employed for anaerobic testing in the broth microdilution MIC assay was Brucella Broth (BD, Cat. No. 211088, Lot No. 7128995), supplemented with hemin (Sigma, Lot No. SLBP5720V), Vitamin K (Sigma, Lot No. MKCG2075) and 5% laked horse blood (LHB, Cleveland Scientific; Bath, OH, Lot No. 474990).

Broth Microdilution Assay

The MIC assay method followed the procedure described by the CLSI[29,30] and employed automated liquid handlers (Multidrop 384, Labsystems, Helsinki, Finland; Biomek 2000 and Biomek FX, Beckman Coulter, Fullerton CA) to conduct serial dilutions and liquid transfers. The wells in columns 2 through 12 in a standard 96-well microdilution plate (Costar) were filled with 150 µL of the appropriate diluent (DMSO for the test agents; dH2O for metronidazole and clindamycin). The drugs (300 µL at 101X the desired top concentration in the test plates) were dispensed into the appropriate well in column 1 of the mother plates. The Biomek 2000 was used to make serial 2-fold dilutions through column 11 in the "mother plate". The wells of column 12 contained no drug and were the organism growth control wells.

The daughter plates for testing of all isolates were loaded with 190 µL per well of supplemented Brucella broth with 5% LHB using the Multidrop 384. The daughter plates were prepared on the Biomek FX instrument which transferred 2 µL of 101X drug solution from each well of a mother plate to the corresponding well of each daughter plate in a single step. The wells of the daughter plates ultimately contained 190 µL of medium, 2 µL of drug solution, and 10 µL of bacterial inoculum prepared in broth.

A standardized inoculum of each organism was prepared per CLSI methods.[57,58] For all bacteria, suspensions were prepared in supplemented Brucella broth supplemented with hemin and Vitamin K to equal the turbidity of a 0.5 McFarland standard. These suspensions were further diluted 1:10 in supplemented Brucella broth with 5% LHB. The inoculum was dispensed into sterile reservoirs (Beckman Coulter) and transferred by hand in the Bactron Anaerobe chamber so that inoculation took place from low to high drug concentration. A 10 µL aliquot of inoculum was delivered into each well. Inoculated daughter plates were stacked and placed in an anaerobic box with GasPak sachets (BD; Lot No. 6309689), covered with a lid on the top plate, and incubated at 35 – 37°C.

The microplates were viewed from the bottom using a plate viewer after 46 hours. For each mother plate, an un-inoculated solubility control plate was observed for evidence of drug precipitation. The MIC was read and recorded as the lowest concentration of drug that inhibited visible growth of the organism.

Supplementary Material

The NMR spectra are available in the Supporting Information.

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Author Contribution Statement

A. D. and K. G. designed the study. A. D. carried out the synthesis and characterization of the derivatives. A. D. and K. G. analyzed and discussed the results. I. S. determined MIC values against *S. aureus*. D. S. and P. S. developed the MIC tests for *M. tuberculosis* and performed the biological evaluation of the derivatives. A. D. and K. G. wrote the manuscript.

A patent application (WO2019135010A1, EP18150671.8A) was filed Jan 8th, 2018 that includes antibiotics presented in this work.

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SUPPORTING INFORMATION

Synthesis of Iodinated Fidaxomicin Derivatives

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EXPERIMENTAL PROCEDURES

General Experimental

Unless otherwise stated, all chemicals were of reagent grade and purchased from *Sigma-Aldrich, Merck, Fluorochem* or *Honeywell*. Fidaxomicin was either obtained by fermentation from *Actinoplanes deccanensis* (ATCC 21983) or purchased from commercial suppliers. Reactions were carried out under protecting gas (N_2 or Ar) and monitored for completion UHPLC-MS (ESI). Solvents for reactions were of p.a. grade. Evaporation of solvents *in vacuo* was carried out on a rotary evaporator at 40 °C bath temperature and appropriate pressure. **Ultra high-performance liquid chromatography coupled to mass spectrometry** (UHPLC-MS): *Ultimate 3000 LC* instrument (*Thermo Fisher Scientific*) coupled to a triple quadrupole *Quantum Ultra EMR* MS (*Thermo Fisher Scientific*) using a reversed-phase column (*Kinetex*® EVO C18; 1.7 µm; 100 Å, 50 x 2.1 mm; *Phenomenex*). The LC was equipped with an *HPG-3400RS* pump, a *WPS-3000TRS* autosampler, a *TCC-3000RS* column oven and a *Vanquish DAD* detector (all *Thermo Fisher Scientific*). The following solvents were applied: H2O + 0.1 % HCOOH (A), MeCN + 0.1 % HCOOH (B). Samples were prepared using HPLC grade solvents (MeCN, MeOH, H_2O) and filtered over a 4 mm syringe filter, PTFE (hydrophilic), pore size: 0.22 µm obtained from BGB Analytik AG. The MS was equipped with an H-ESI II ion source. The source temperature was 250 °C, the capillary temperature 270 °C and capillary voltage 3500 V, and datasets were acquired at resolution 0.7 on Q3 in centroid mode. **High-performance liquid chromatography** (HPLC): *Prominence* modular HPLC instrument (*Shimadzu*) coupled to an *SPD-20A* UV/Vis detector (*Shimadzu*) using a reversed-phase column (*Gemini-NX* C18, 3 µm, 10 Å, 150 mm x 4.6 mm) for analytical HPLC, and a reversed-phase column (*Gemini NX* C18, 5 µm, 110 Å, 250 mm x 21.2 mm; for preparative HPLC. The LC was equipped with a *CBM-20A* system controller, *LC-20A* solvent delivery unit, a *DGU-20A* degassing unit, *FRC-10A* fraction collector (all *Shimadzu*). The following solvents were used: H2O + 0.1 % HCOOH (A), MeCN + 0.1 % HCOOH (B). **Specific optical rotation** $[\alpha]_D^T$: *Jasco P-2000 Polarimeter*; measured at the indicated temperature *T*. All given values for $[\alpha]_D^T$ have the dimension ° mL dm^{-1} g^{-1}. **Infrared spectra** (IR): *SpectrumTwo FT-IR Spectrometer* (*Perkin–Elmer*) equipped with a *Specac Golden Gate™ ATR* (attenuated total reflection) accessory; applied as neat samples or as films; $1/\lambda$ in cm^{-1}. **Nuclear magnetic resonance spectra** (NMR): ^{1}H NMR spectra were recorded in acetone-d_6 on the instruments *AV-500* (500 MHz) or *AV-400* (400 MHz); chemical shift δ in ppm relative to solvent signals (δ = 2.05 ppm for acetone-d_6),[1] coupling constant *J* is given in Hz. ^{13}C NMR spectra were recorded acetone-d_6 on the instruments *Bruker AV-500* (125 MHz) or *AV-400* (100 MHz); chemical shift δ in ppm relative to solvent signals (δ = 29.84 ppm for acetone-d_6).[1] **High-resolution electrospray**

ionization mass spectra (HRMS): *On flow injection*: High-resolution mass spectra were acquired ona *QExactive* instrument (*ThermoFisher Scientific*, Bremen, Germany) equipped with a heated electrospray (ESI) ionization source and connected to a *Dionex Ultimate 3000* UHPLC system (*ThermoFischer Scientifics*, Germering, Germany). The samples were dissolved in MeOH, MeOH/CH_2Cl_2 3:1, MeOH/H_2O 1:1, DMSO/H_2O 1:10 or H_2O at a concentration of ca. 50 µg mL^{-1} thereof 1µL was injected on-flow with a XRS auto-sampler (*CTC*, Zwingen, Switzerland). The mobile phase (120 µL mL^{-1} flow rate) consisting of MeOH + 0.1 % HCOOH or acetonitrile/H_2O 2:8 + 0.1% HCOOH was chosen according to the solubility. Ionsource parameters were set as follow: spray voltage 3.0 kV; capillary temperature 280°C; sheath gas 30 L min^{-1}; aux gas 8; 30 L min^{-1}; s-lens RF level 55.0; and aux gas temperature 250°C. Full scan MS were acquired in the alternating (+)/(–)-ESI mode andoverthe ranges *m/z* 80-1'200, 133-2'000, or 200-3'000at 70'000 resolution (full width half-maximum) and with automatic gain control (AGC) target of 3.00E +06. The maximum allowed ion transfer time (IT) was 30ms. Masses were calibrated below 2 ppm accuracy between *m/z* 130.06619 and 1621.96509 in the positive and between 265.14790 and 1779.96528 in the negative ESI mode usingthe Pierce® ESI calibration solutions (ThermoFisher Scientific, Rockford, USA). Additionally, contaminations of erucamide (*m/z* 338.34174, (+)-ESI) and palmitic acid (*m/z* 255.23295, (–)-ESI) were used as lock masses in (+)-and (–)-ESI, respectively. *LC-MS*: Samples (1 µL injection) were analyzed with a Dionex Ultimate 3000UHPLC system (*ThermoFischer Scientifics*, Germering, Germany) connected to an Acquityeldetector and a *QExactive* high-resolution mass spectrometer (*ThermoFisher Scientific*, Bremen, Germany) equipped with a heated electrospray (ESI) ionization source. Separation was performed with an *AcquityBEH* C18 HPLC column (1.7 µm particle size, 2x100 mm, *Waters*) kept at 30 °C. The mobile phase was consisting of A: H_2O + 0.1% HCOOH and B: CH_3CN + 0.1% HCOOH. A linear gradient was run from 5 to 98% B within 5 min followed by flushing with 98% B for 1 min at 400 µL min^{-1} flow rate. UV spectra were recorded between 200 and 600 nm at 1.2 nm resolution and 20 points s^{-1}. MS ion source parameters were set as follows: spray voltage, 3.5 kV; capillary temperature, 260 °C; sheath gas 45 L min^{-1}; auxgas 15L min^{-1}; sweepgas 2 L min^{-1}; s-lens RF level 450.0, and aux gas temperature 250 °C. Full scan MS were acquired in the (+) ESI mode and over the ranges *m/z* 80-1'200, 133-2'000, or 200-3'000 at 70'000 resolution (full width half-maximum) and with automatic gain control (AGC) target of 3.00E +06. The maximum allowed ion transfer time (IT) was 30 ms. Masses were calibrated below 2 ppm accuracy between *m/z* 130.06619 and 1621.96509 in the positive and between 265.14790 and 1779.96528 in the negative ESI mode using the Pierce® ESI calibration solutions (*ThermoFisher Scientific*, Rockford, USA). Additionally, contaminations of erucamide (*m/z* 338.34174, (+)-ESI) and palmitic acid (*m/z* 255.23295, (-)ESI) were used as lock masses in (+)-and (-)-ESI, respectively.

General procedure for the determination of MIC values of *S. aureus*

Organism	**Strain**
Staphylococcus aureus	ATCC29213

The strain *Staphylococcus aureus* was grown overnight at 37 °C on MH II agar plates. (BD™ BBL™ Mueller Hinton II Agar, BD Diagnostics). MIC values were determined by broth dilution method according to the recommendations of the Clinical and Laboratory Standards Institute (CSLI; U.S.A.). The inoculum size was about $7.5x10^5$ colony forming units/well. The compounds were diluted in H_2O from 1.0 mg/mL stock solutions in 50 % methanol/H_2O in a 2-fold dilution series. The microtiter plates were incubated at 37 °C overnight. Afterwards, the MIC (lowest concentration of the compounds with no bacterial growth observed) was determined by visual inspection.

General procedure for the determination of MIC values of *M. tuberculosis*

MIC determination was essentially conducted as described recently.[2] Briefly, the Green-Fluorescent Protein (GFP) expressing recombinant *Mycobacterium tuberculosis* H37Rv *rpsL*[3] transformed with pOLYG-Pr-GFP[4] was grown in Middlebrook 7H9-OADC with 0.05 % Tween 80 until mid-log phase (optical density at 600 nm OD_{600} = 0.3 – 1.0), diluted to an OD_{600} of 0.04 and 20 µl of the suspension were added to an equal volume of 12-point two-fold serial dilutions of the compounds in 7H9-OADC-Tween in 384-well plates in triplicates. Compound concentrations were in the range of 62.5 to 0.031 µM. Fluorescence was measured immediately after inoculation (background) and after 10 days of incubation at 37 °C. Dose response curves were fitted with a 4-parameter log-normal model. P_{MIN} [-,-] and P_{MAX} [-, 120] are the minimum and the maximum, respectively, P_{Hill} [0,-] indicates the steepness, and EC_{50} [-,-] the log-back transformed Minimal Effective Concentration 50. The computational and statistical analysis was conducted with R (3.0.1 – 3.1.1; https://www.r-project.org/). Dose response curves were fitted with the ´drc´ package. The inhibitory potency I was calculated with the equation I = 100-[100●(S-P)/N-P)]. S is the sample's fluorescence while P and N derive from growth inhibition with the control drug (Kanamycin A) and solvent growth control measurements (DMSO 1.25 % vol./vol.), respectively. A fluorescence reduction of 90 % as compared to the no-drug control was reported as Minimal Inhibitory Concentration (MIC_{90}).

General procedure for the determination of MIC values of *C. difficile*

MIC determination was carried out by Micromyx, LLC, 4717 Campus Drive, Kalamazoo, MI, USA 49008.

Approximately 5 mg of each of the test compounds were provided. These were stored at -20°C until testing. On the day of the assay, the test articles were dissolved in 100% DMSO (dimethyl sulfoxide, Sigma; St. Louis, MO, Cat. No. 472301-500ML, Lot No. SHBH5551V) to a stock concentration of 3232 µg/mL. The concentration range tested for these test agents was 16 – 0.015 µg/mL. The comparator agents, metronidazole and clindamycin were supplied by Micromyx, as shown in the table below:

Comparator Drug	**Supplier**	**Catalog No.**	**Lot No.**	**Solvent/Diluent**	**Testing Range (µg/mL)**
Metronidazole	Sigma	M3761-100G	095K0693	DMSO/$_{d}$H2O	64 – 0.06
Clindamycin	Sigma	C5269-100MG	021M1533	$_{d}$H2O/ $_{d}$H2O	32 – 0.03

Test Organisms

Test organisms consisted of reference strains from the American Type Culture Collection (ATCC; Manassas, VA) and clinical isolates from the Micromyx repository (MMX; Kalamazoo, MI). Organisms were initially received at Micromyx and were streaked for isolation. Colonies were picked by sterile swab from the medium and suspended in the appropriate broth containing cryoprotectant. The suspensions were aliquoted into cryogenic vials and maintained at -80ºC.

Prior to testing, all isolates were streaked onto Brucella Agar supplemented with hemin, Vitamin K and 5% sheep blood (Becton Dickinson [BD]; Sparks, MD, Cat. No. 297716, Lot No. 8256909) and incubated anaerobically at 35-37°C for 44 – 48 hours.

Additionally, *Bacteroides fragilis* ATCC 25285 and *Clostridium difficile* ATCC 700057 were tested for purposes of quality control.

Test Medium

The medium employed for anaerobic testing in the broth microdilution MIC assay was Brucella Broth (BD, Cat. No. 211088, Lot No. 7128995), supplemented with hemin (Sigma, Lot No. SLBP5720V), Vitamin K (Sigma, Lot No. MKCG2075) and 5% laked horse blood (LHB, Cleveland Scientific; Bath, OH, Lot No. 474990).

Broth Microdilution Assay

The MIC assay method followed the procedure described by the CLSI[29,30] and employed automated liquid handlers (Multidrop 384, Labsystems, Helsinki, Finland; Biomek 2000 and Biomek FX, Beckman Coulter, Fullerton CA) to conduct serial dilutions and liquid transfers. The wells in columns 2 through 12 in a standard 96-well microdilution plate (Costar) were filled with 150 µL of the appropriate diluent (DMSO for the test agents; $_{d}$H2O for metronidazole and clindamycin). The drugs (300 µL at 101X the desired top concentration in the test plates) were dispensed into the appropriate well in column 1 of the mother plates. The Biomek 2000 was used to make serial 2-fold dilutions through column 11 in the "mother plate". The wells of column 12 contained no drug and were the organism growth control wells.

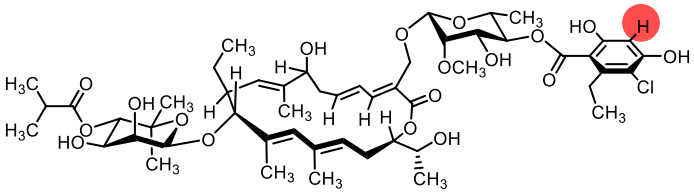
The daughter plates for testing of all isolates were loaded with 190 µL per well of supplemented Brucella broth with 5% LHB using the Multidrop 384. The daughter plates were prepared on the Biomek FX instrument which transferred 2 µL of 101X drug solution from each well of a mother plate to the corresponding well of each daughter plate in a single step. The wells of the daughter plates ultimately contained 190 µL of medium, 2 µL of drug solution, and 10 µL of bacterial inoculum prepared in broth.

A standardized inoculum of each organism was prepared per CLSI methods.[5,6] For all bacteria, suspensions were prepared in supplemented Brucella broth supplemented with hemin and Vitamin K to equal the turbidity of a 0.5 McFarland standard. These suspensions were further diluted 1:10 in supplemented Brucella broth with 5% LHB. The inoculum was dispensed into sterile reservoirs (Beckman Coulter) and transferred by hand in the Bactron Anaerobe chamber so that inoculation took place from low to high drug concentration. A 10 µL aliquot of inoculum was delivered into each well. Inoculated daughter plates were stacked and placed in an anaerobic box with GasPak sachets (BD; Lot No. 6309689), covered with a lid on the top plate, and incubated at 35 – 37°C.

The microplates were viewed from the bottom using a plate viewer after 46 hours. For each mother plate, an un-inoculated solubility control plate was observed for evidence of drug precipitation. The MIC was read and recorded as the lowest concentration of drug that inhibited visible growth of the organism.

Control Experiment Deuterium Incoorporation

A microwave tube was charged with fidaxomicin (**1**, 10 mg, 9.5 µmol, 1.0 eq.), $Pd_2dba_3 \cdot CH_3Cl$ (4.3 mg, 4.2 µmol, 50 mol%), CyJohnPhos (5.0 mg, 14.2 µmol, 1.5 eq.), KF (1.1 mg, 18.9 µmol, 2.0 eq.) and B_2pin_2 (7.2 mg, 28.4 µmol, 3.0 eq.) and the flask was evacuated and flushed with argon several times. The solids were dissolved in dry dioxane (2.0 mL, degassed by freeze-pump-thaw (3x)). Then, D_2O (0.3 µL, 18.9 µmol, 2 eq.) was added and the mixture was allowed to stir at 100 °C for 24 h. The reaction mixture was filtered over Celite® and the solvent was evaporated. The crude was analyzed by UHPLC-MS.



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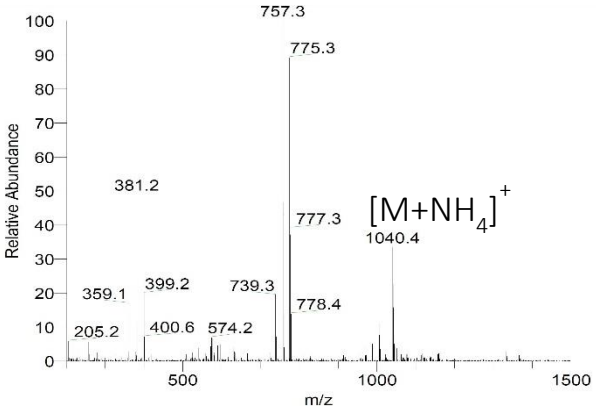

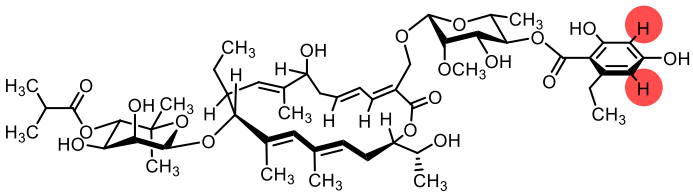

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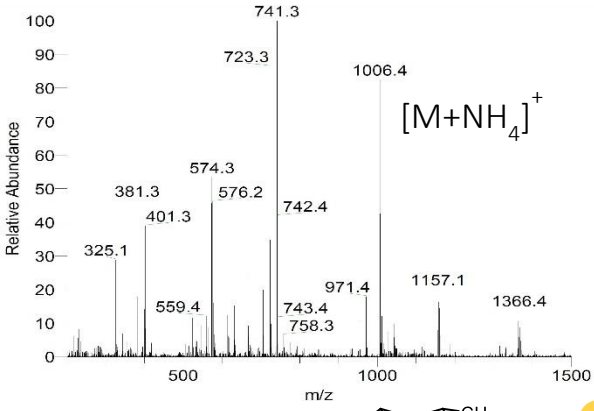

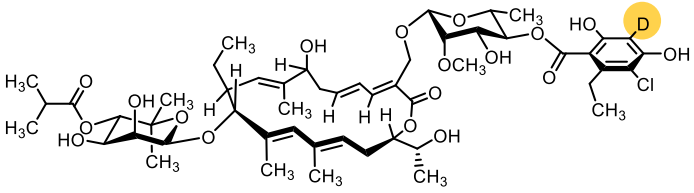

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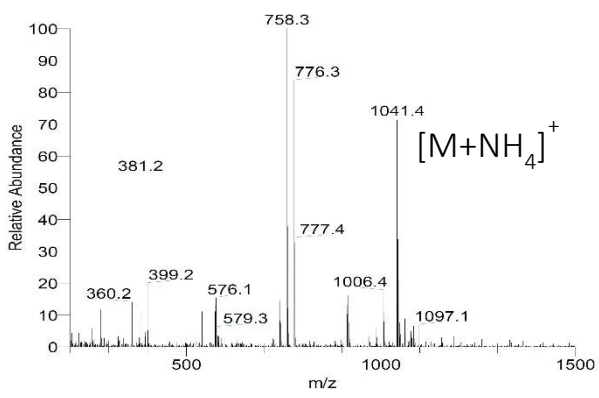

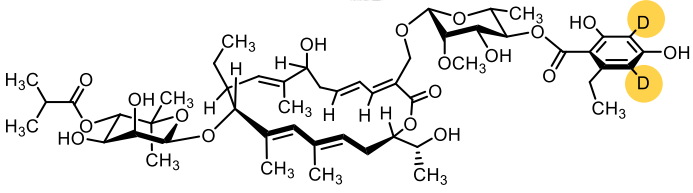

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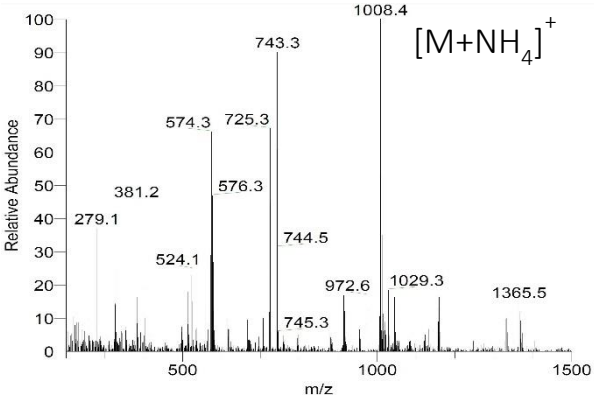
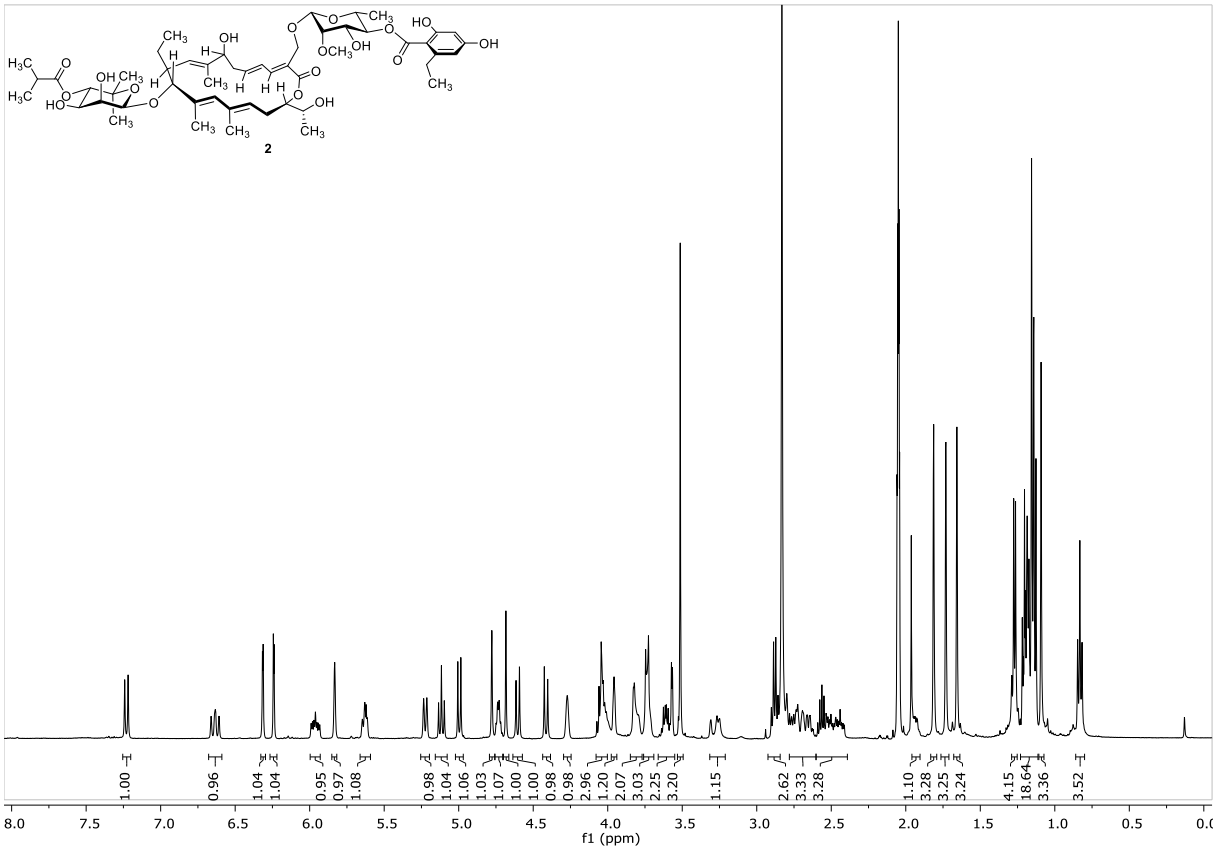
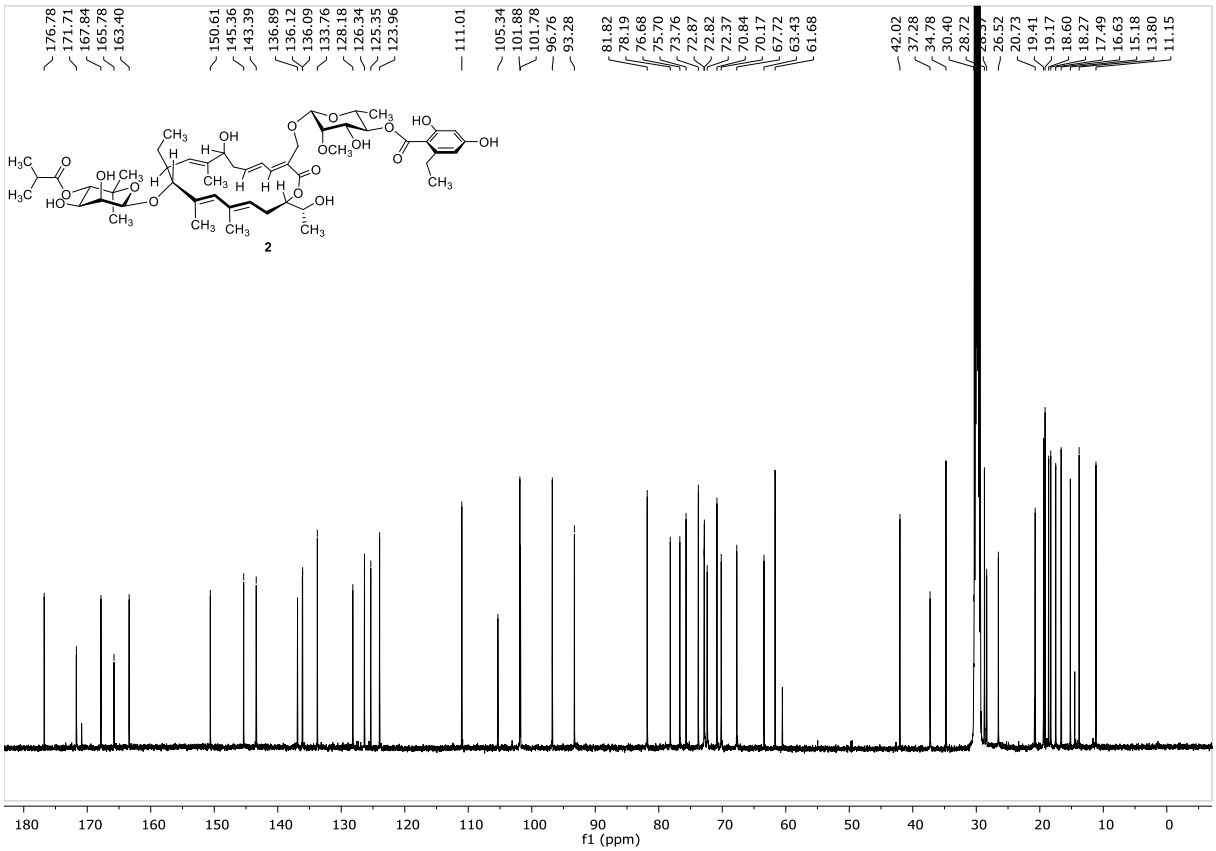


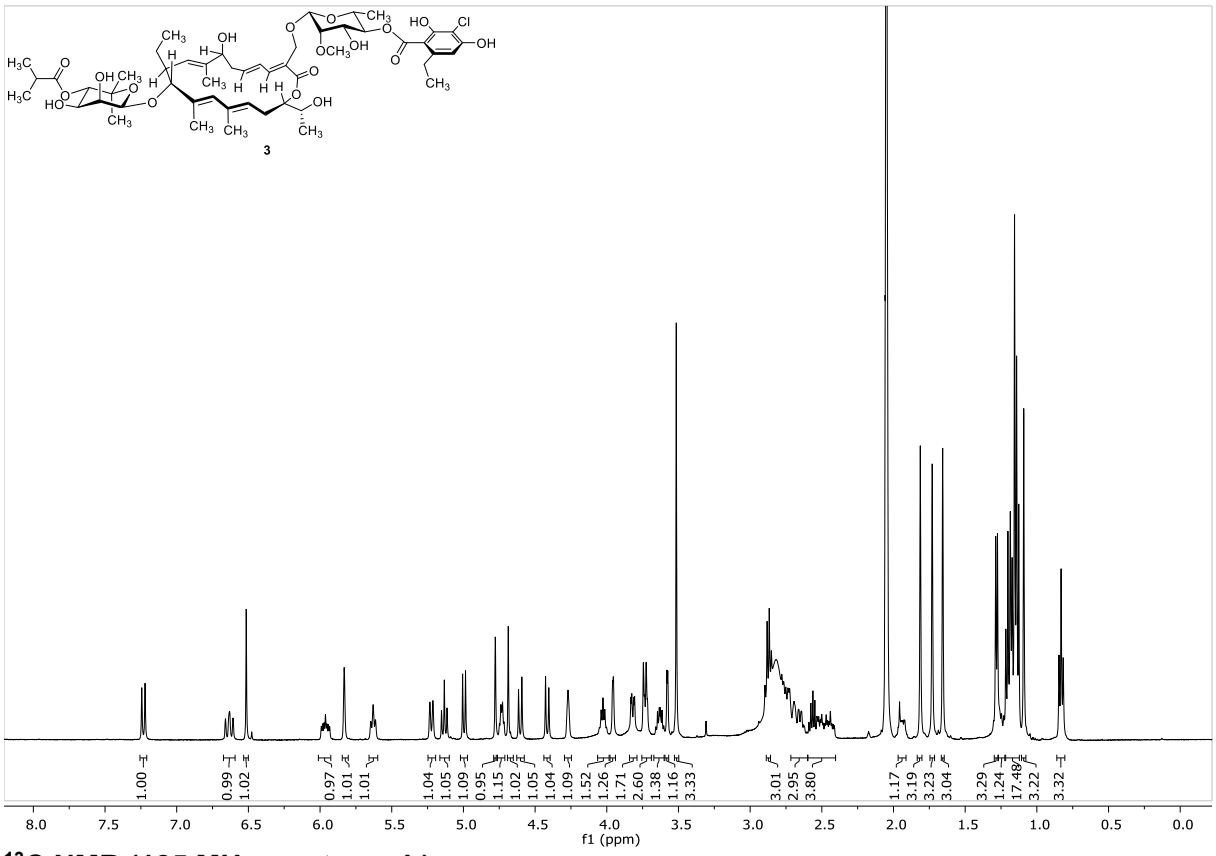
^{1}H NMR (500 MHz, acetone-d_6)



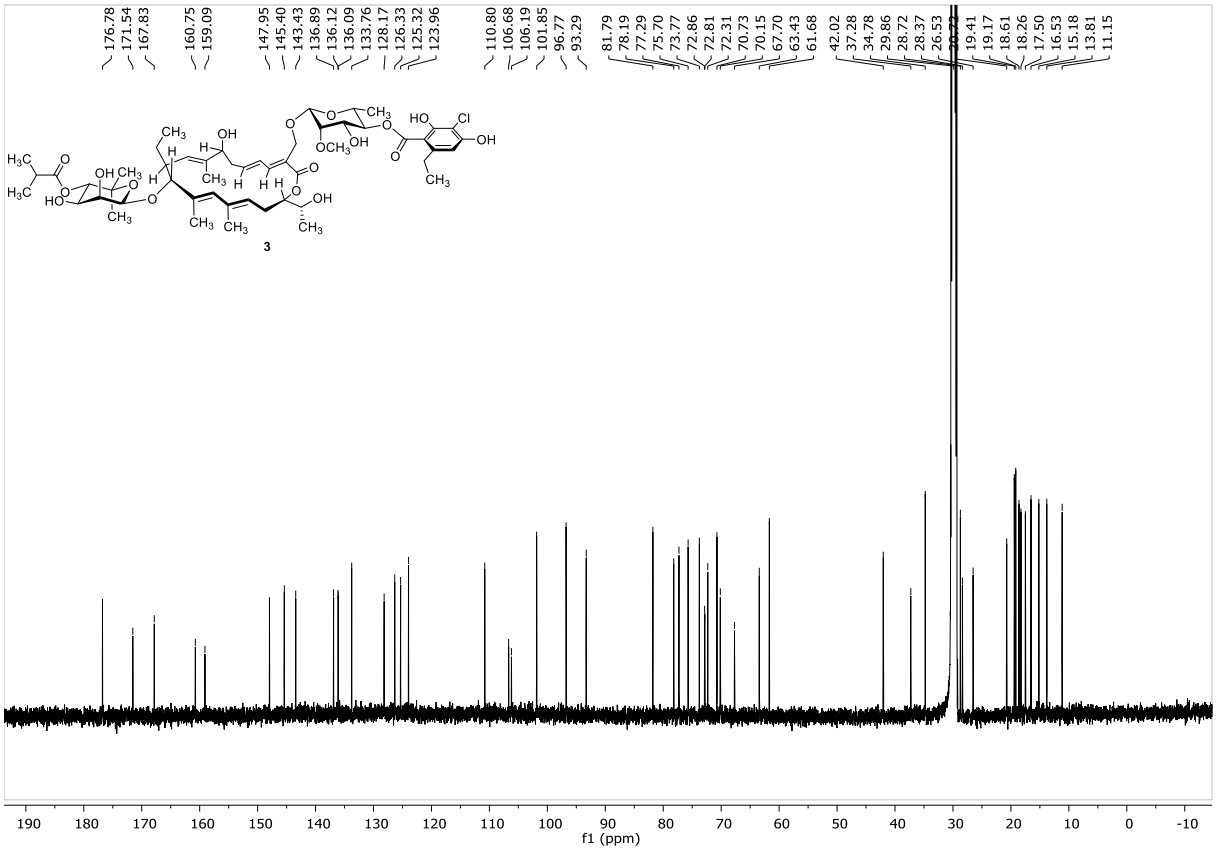
^{13}C NMR (125 MHz, acetone-d_6)



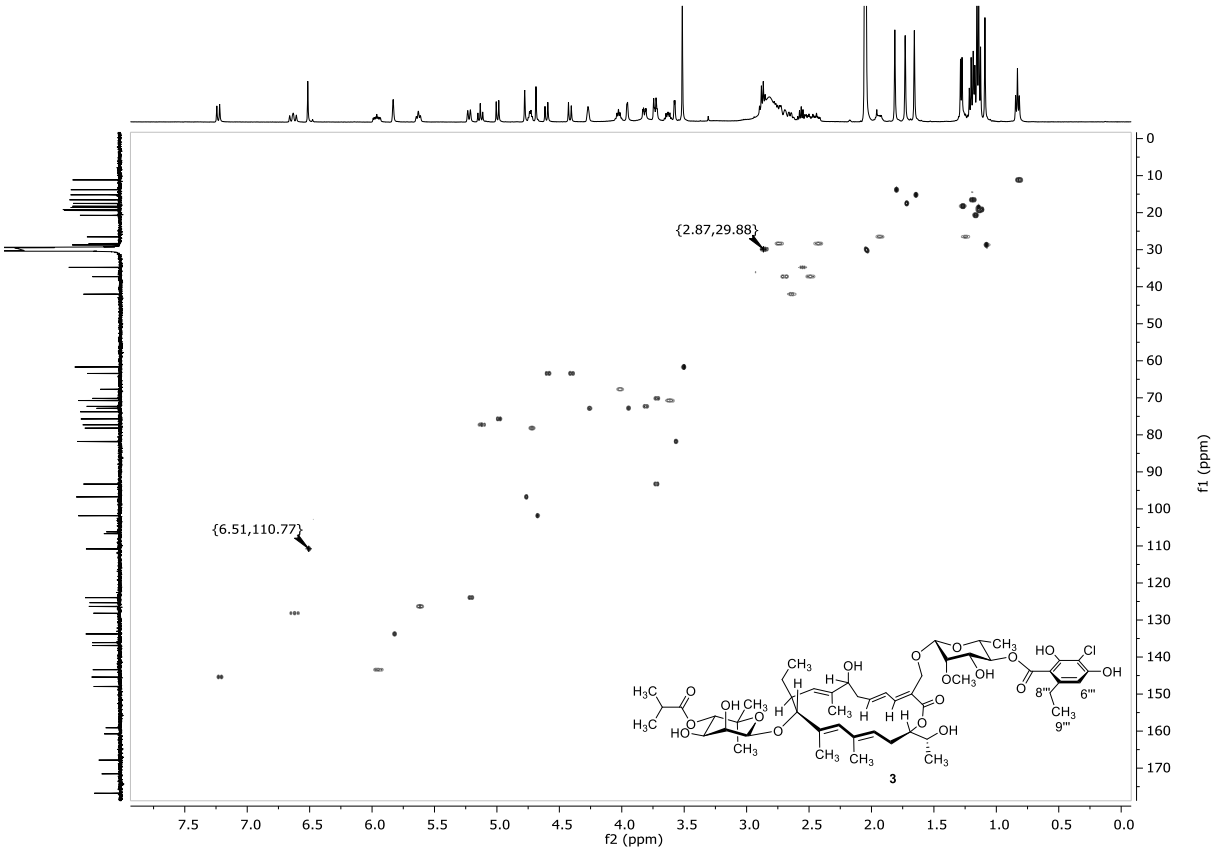
^{1}H NMR (500 MHz, acetone-d_6)



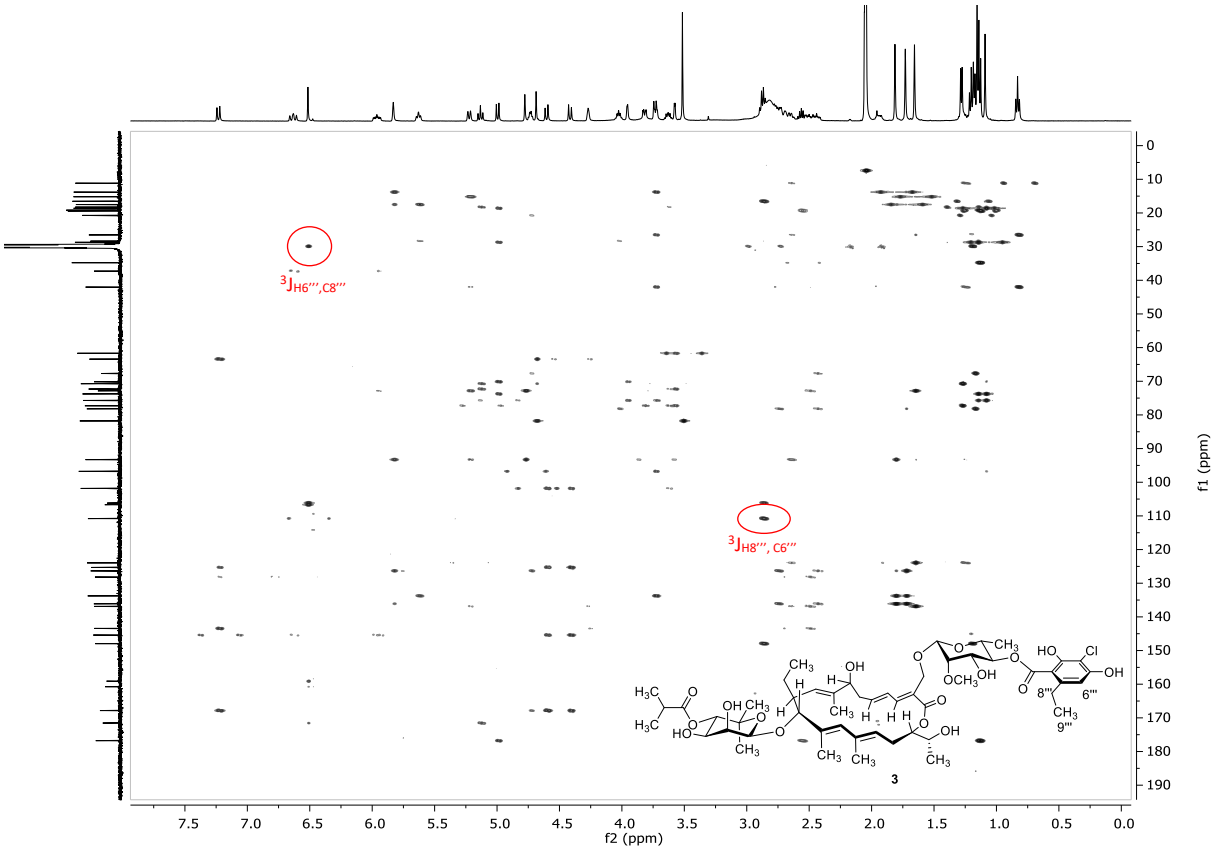
^{13}C NMR (125 MHz, acetone-d_6)



HSQC (500 MHz, acetone-d_6)



HMBC (500 MHz, acetone-d_6)



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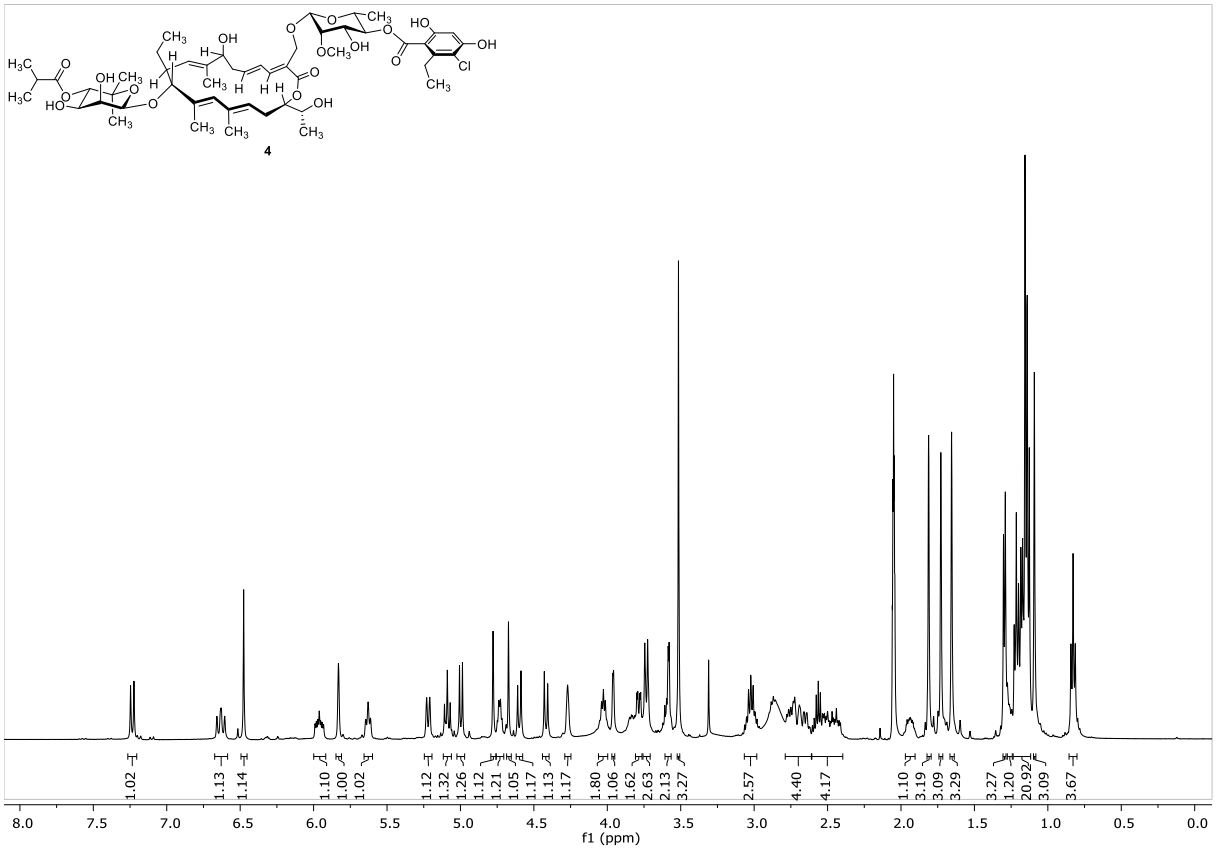

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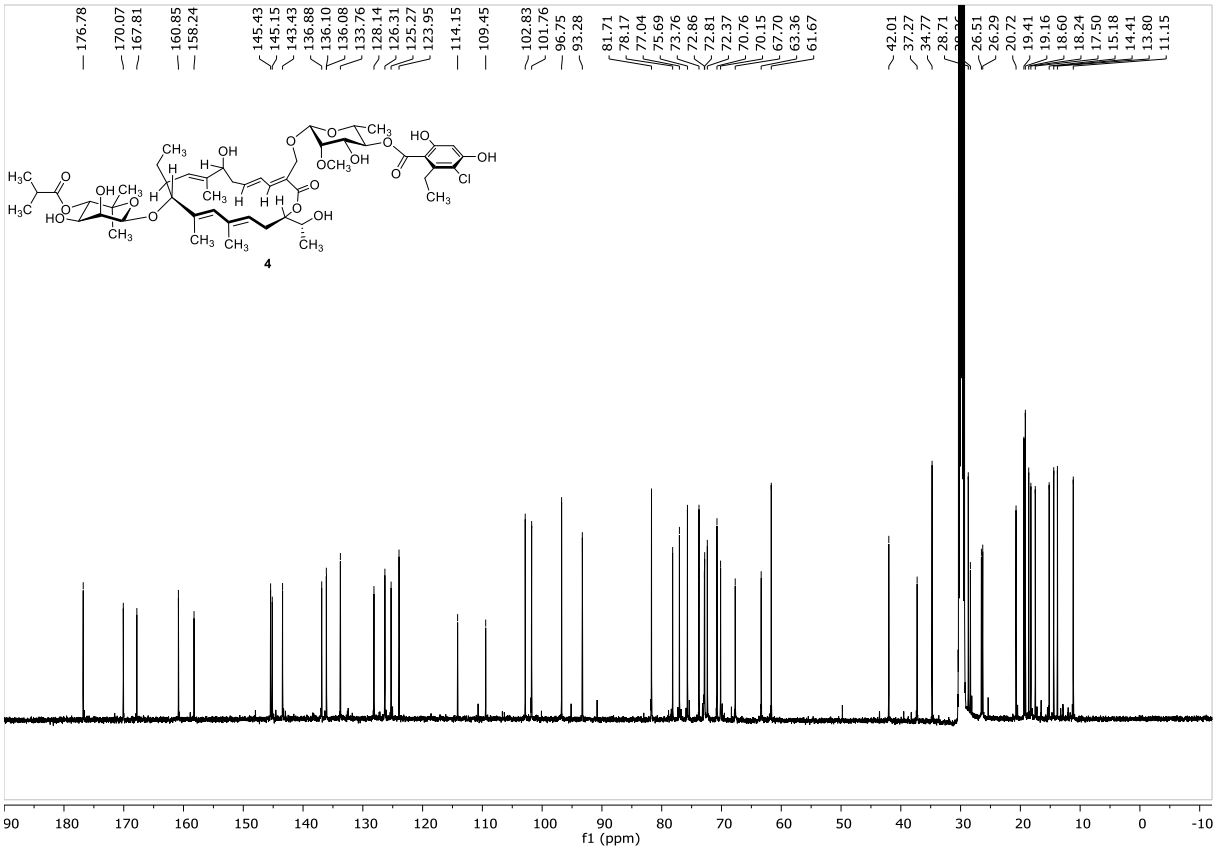
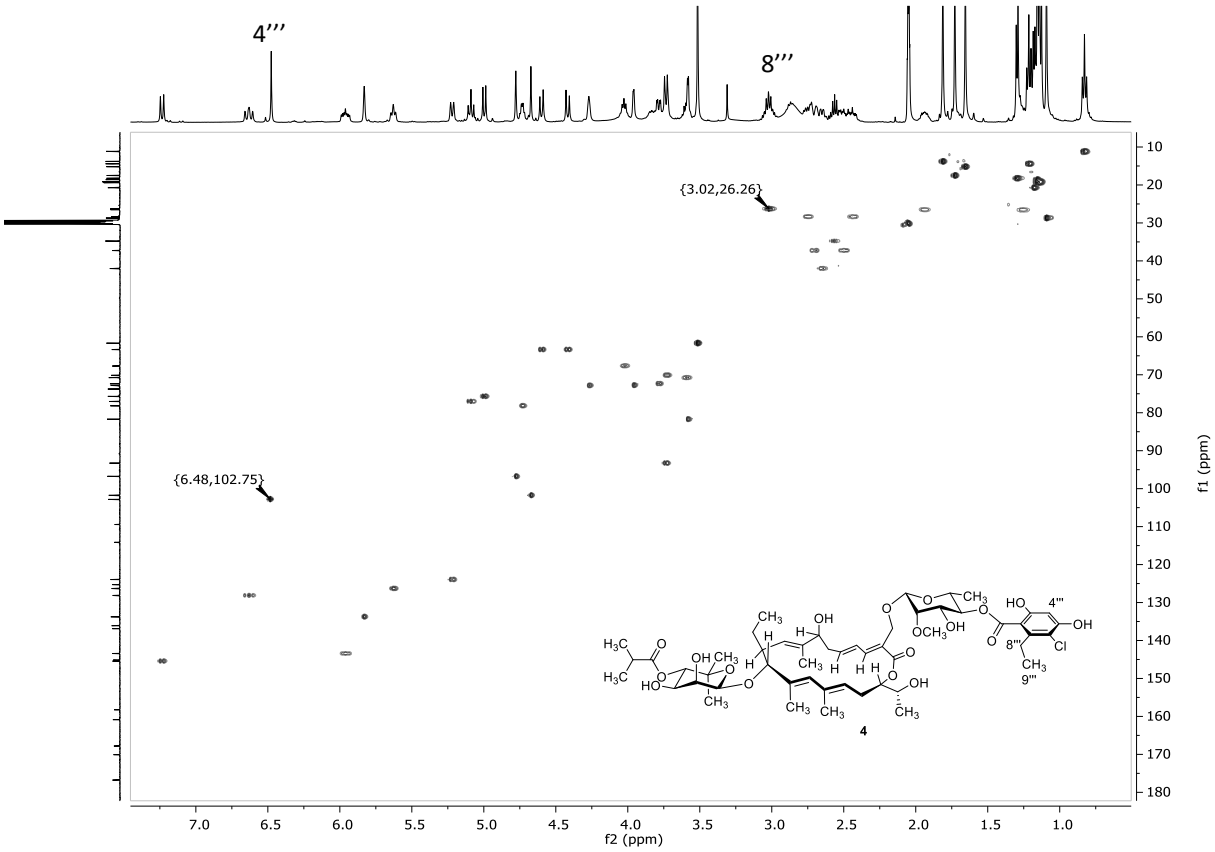


HSQC (500 MHz, acetone-d_6)



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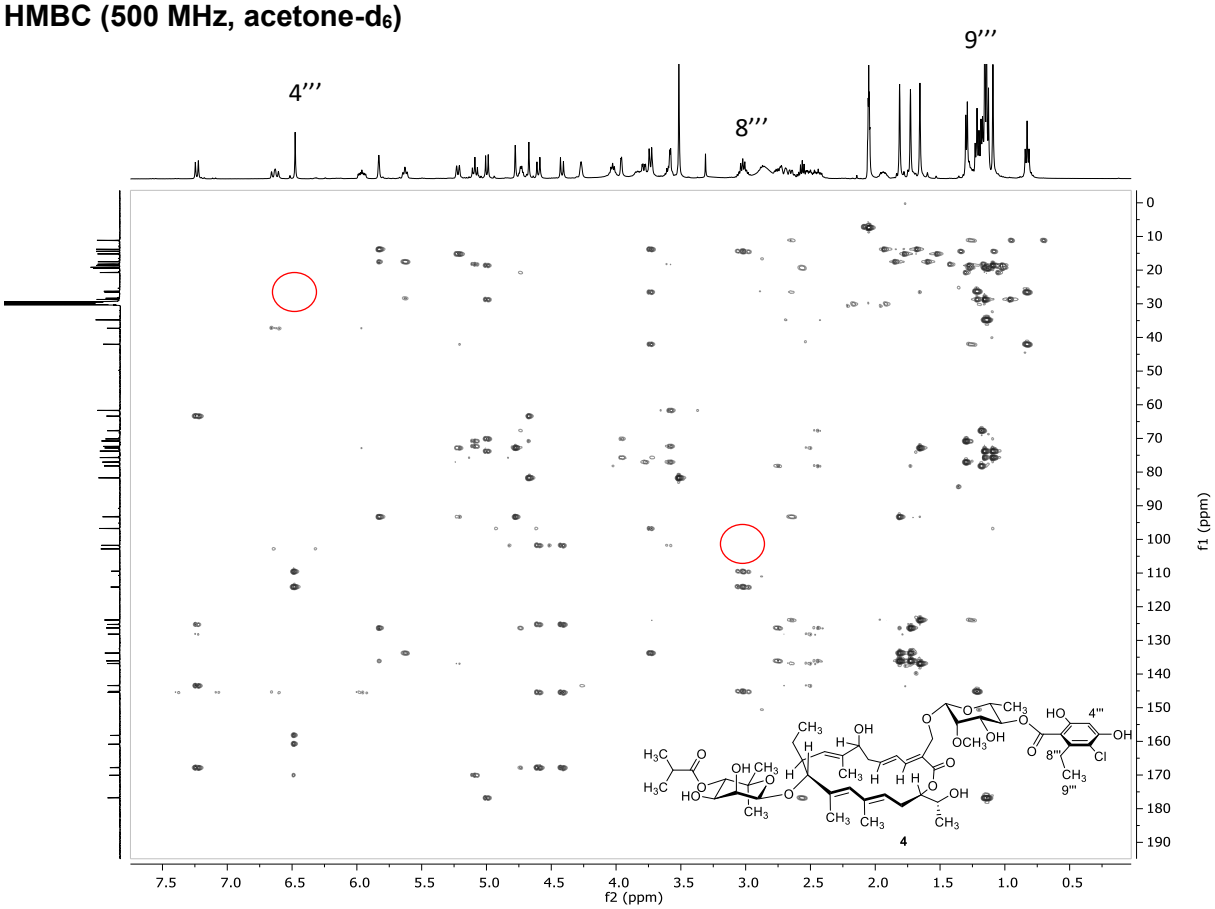

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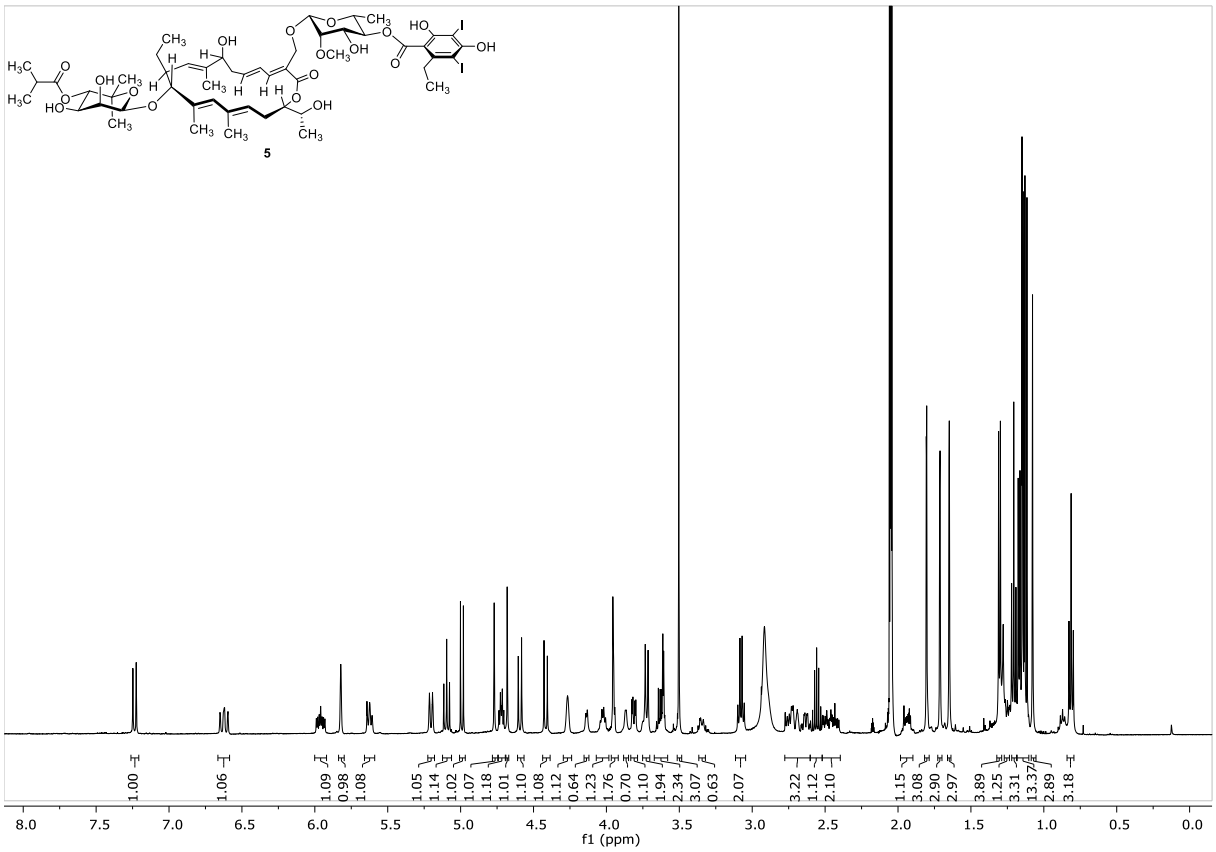

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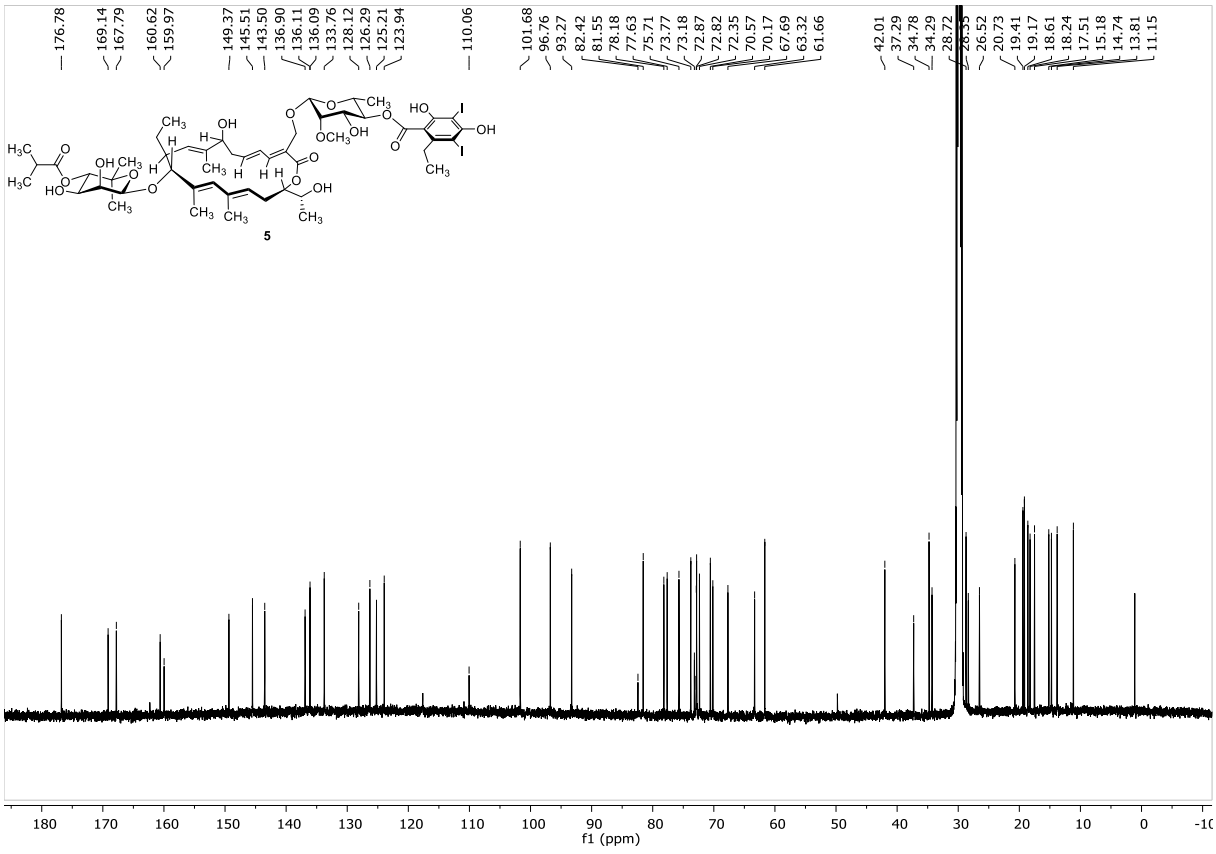
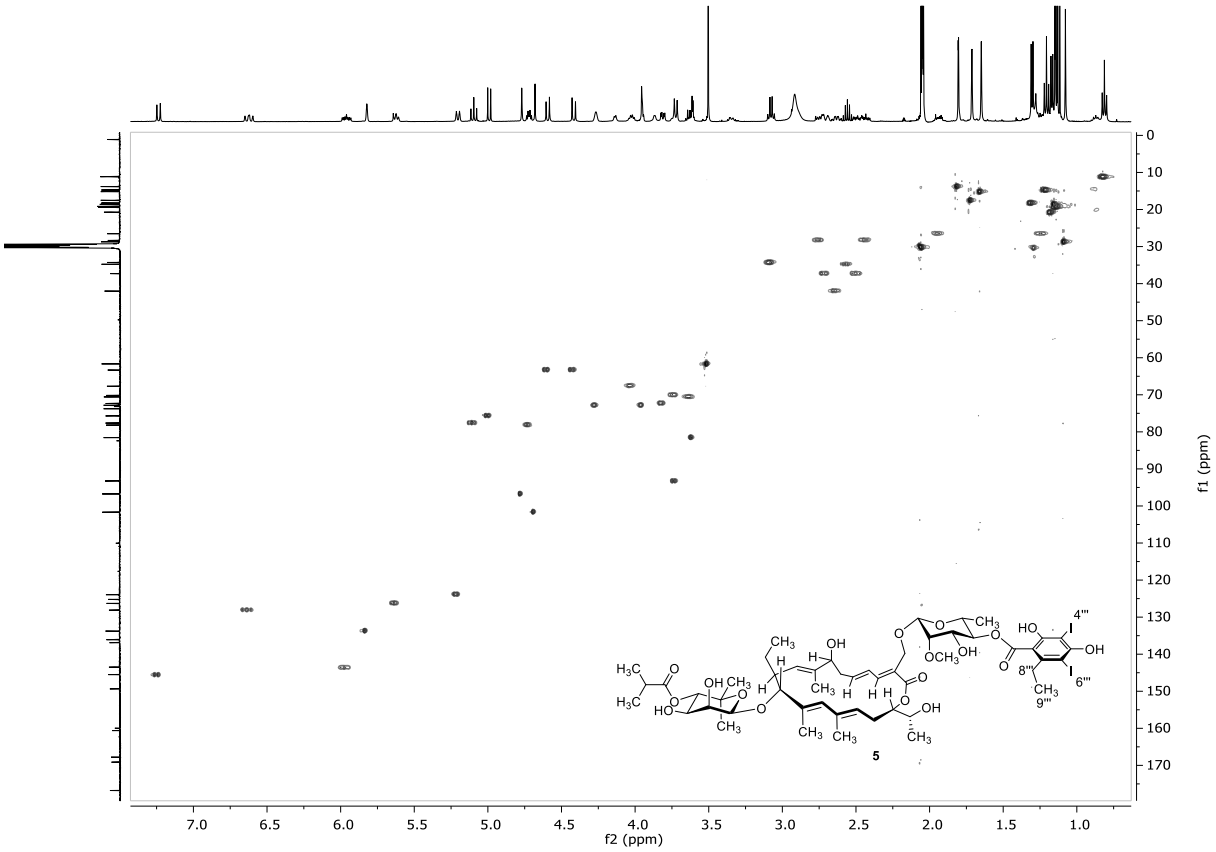
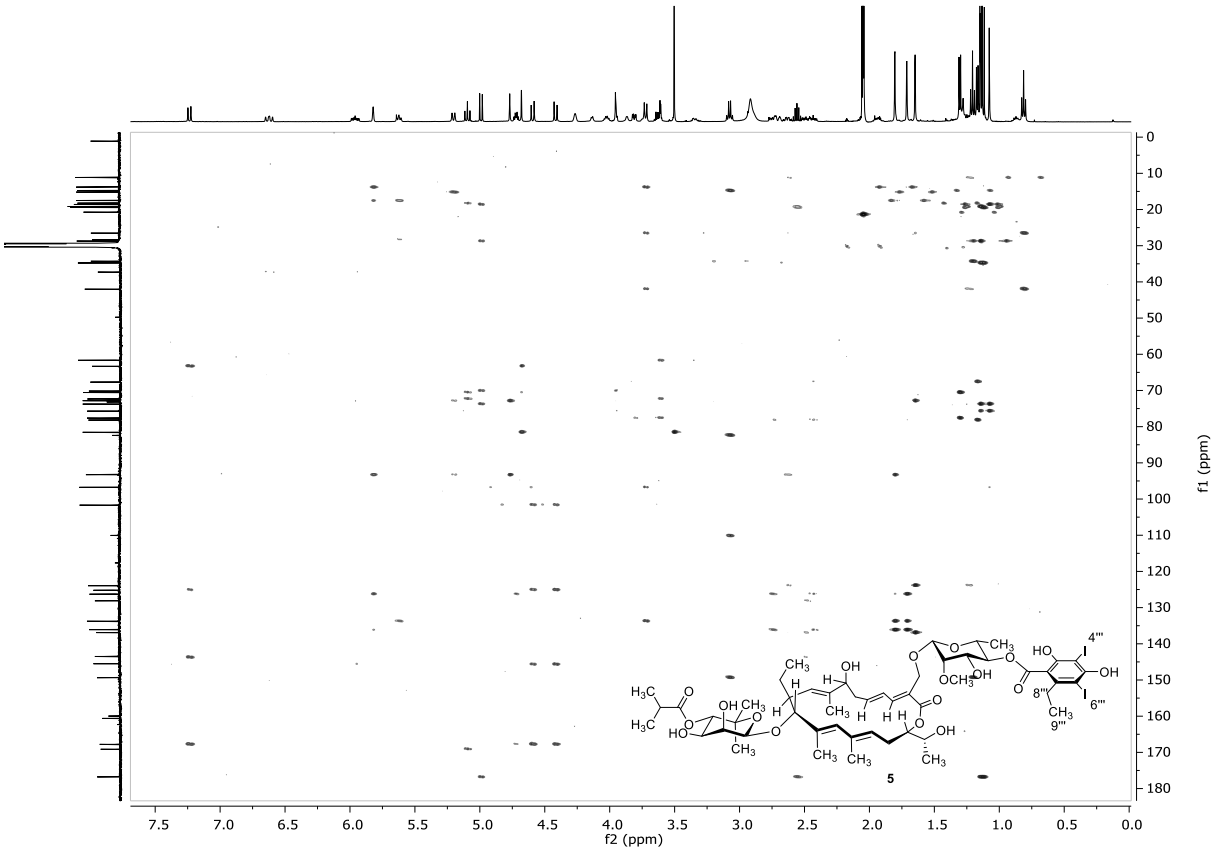


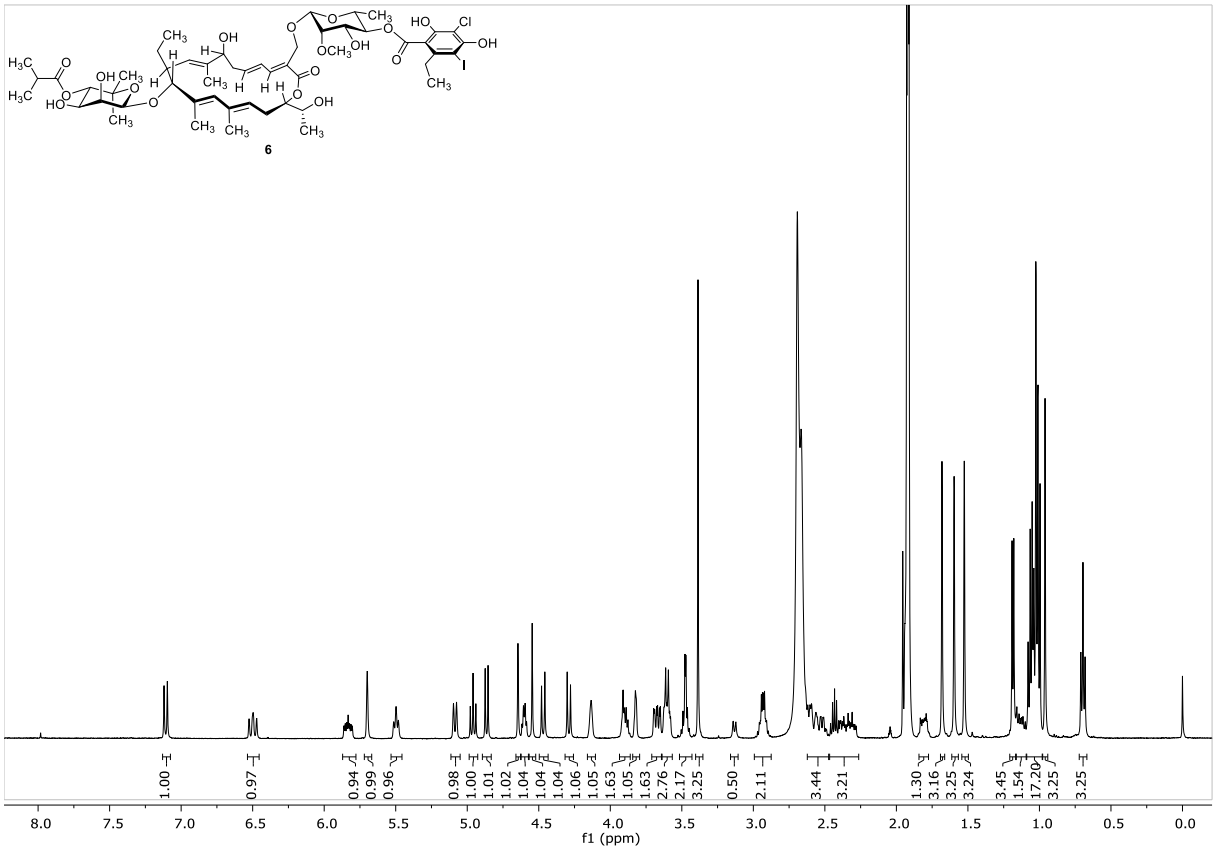
HSQC (500 MHz, acetone-d_6)



HMBC (500 MHz, acetone-d_6)



^{1}H NMR (500 MHz, acetone-d_6)



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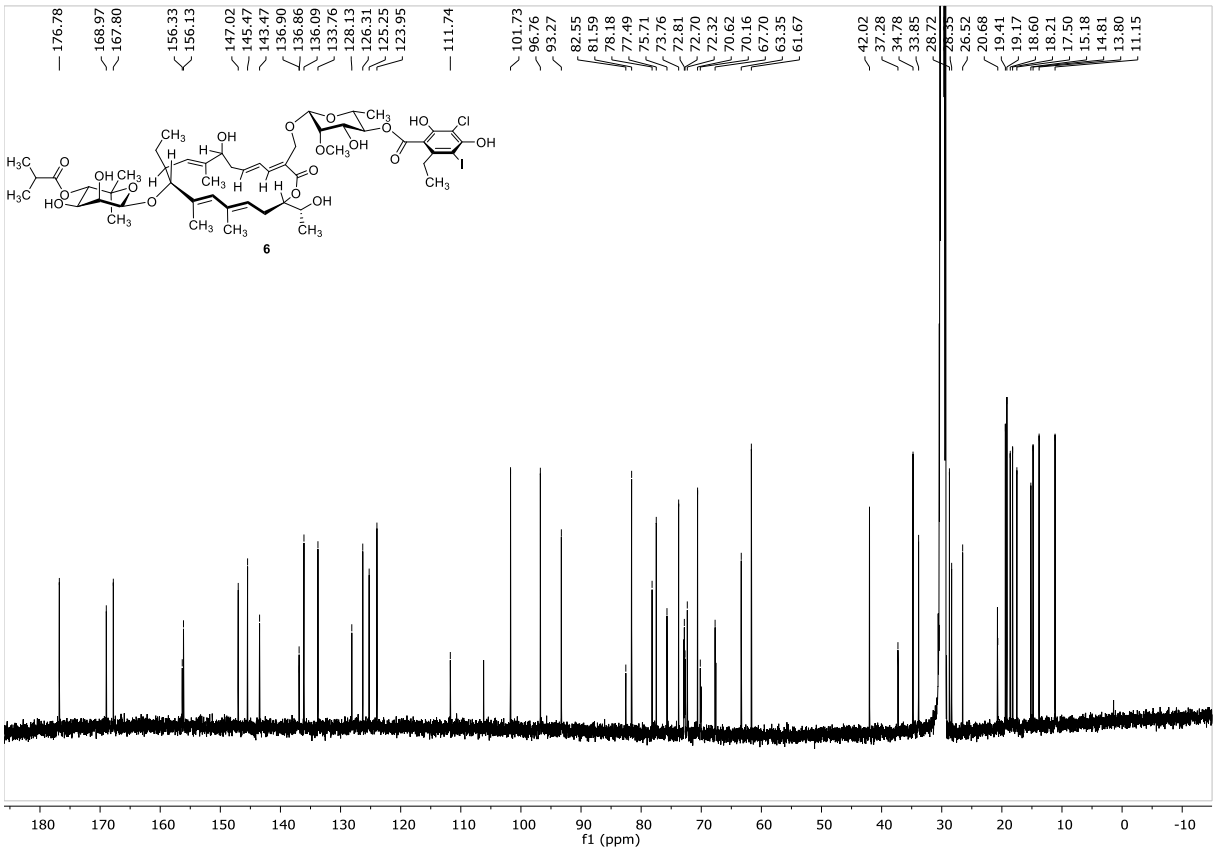
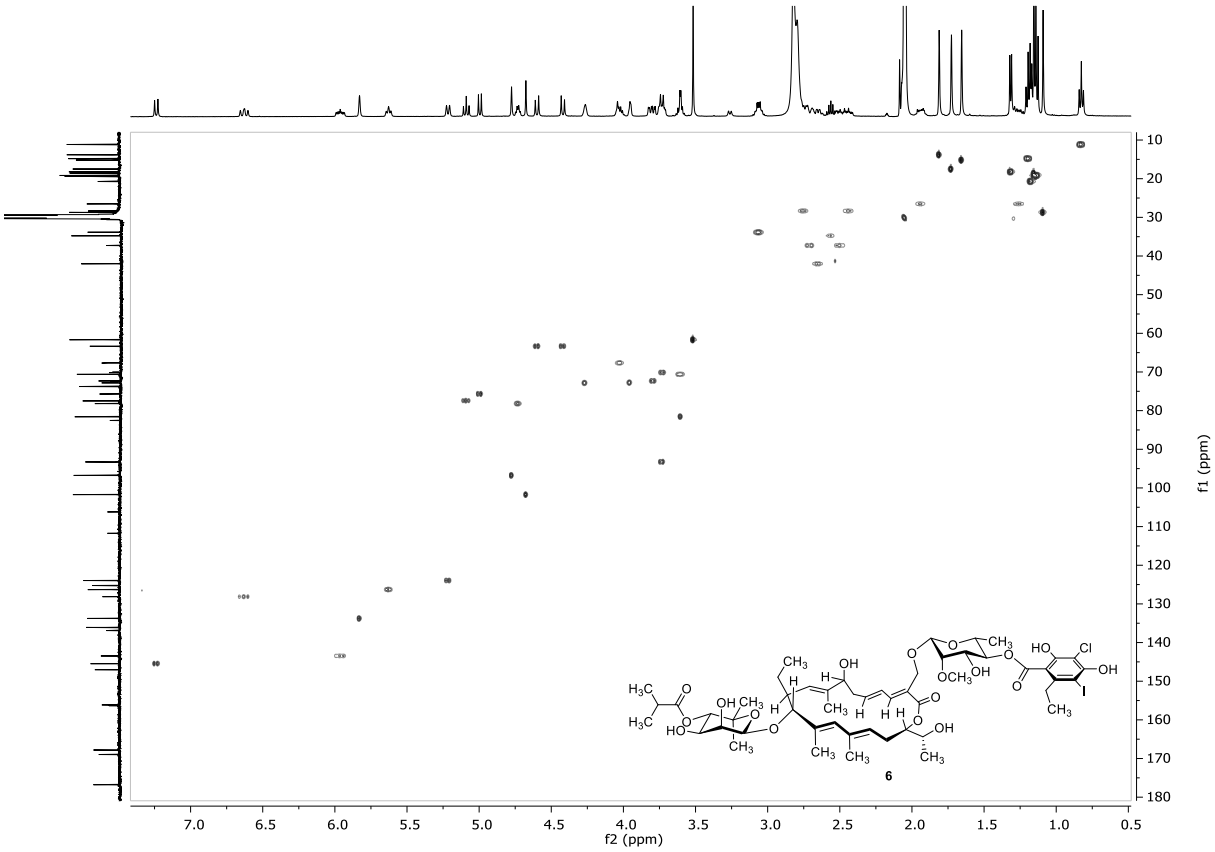
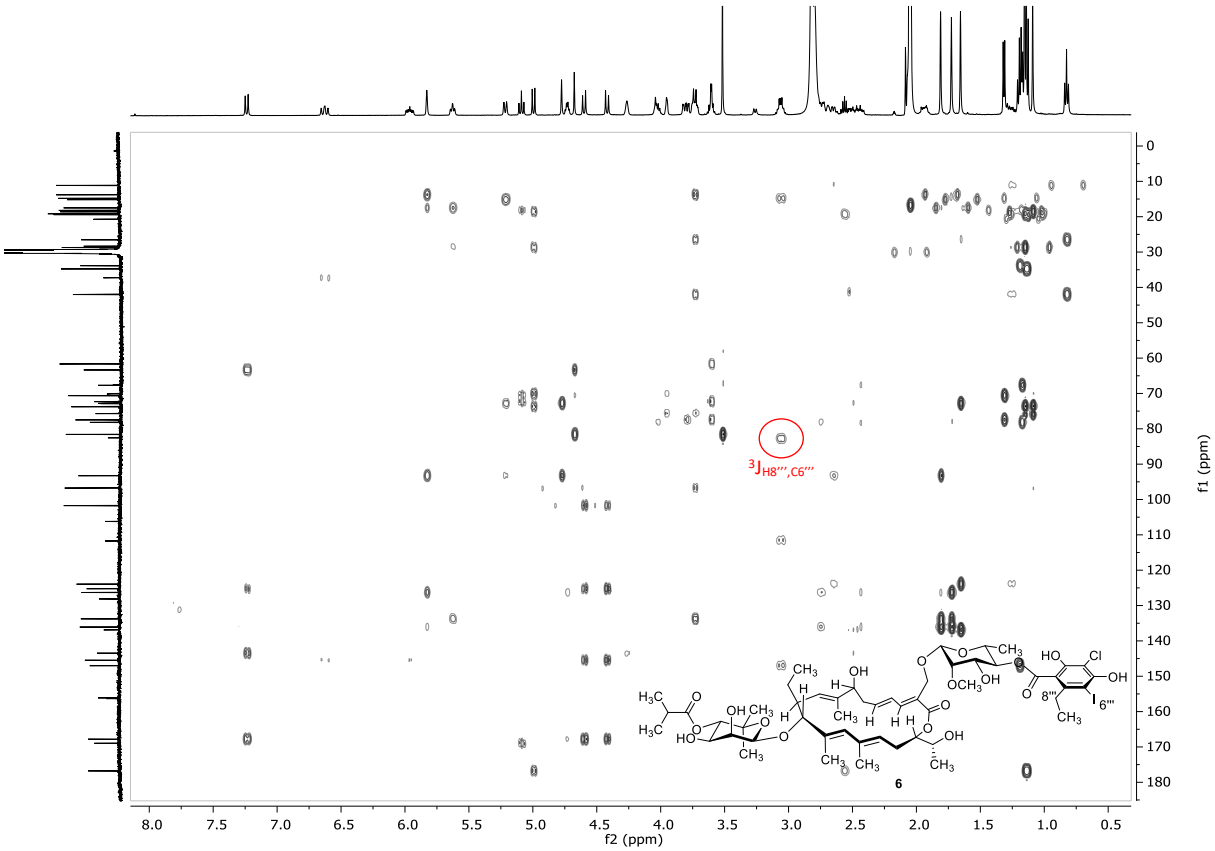


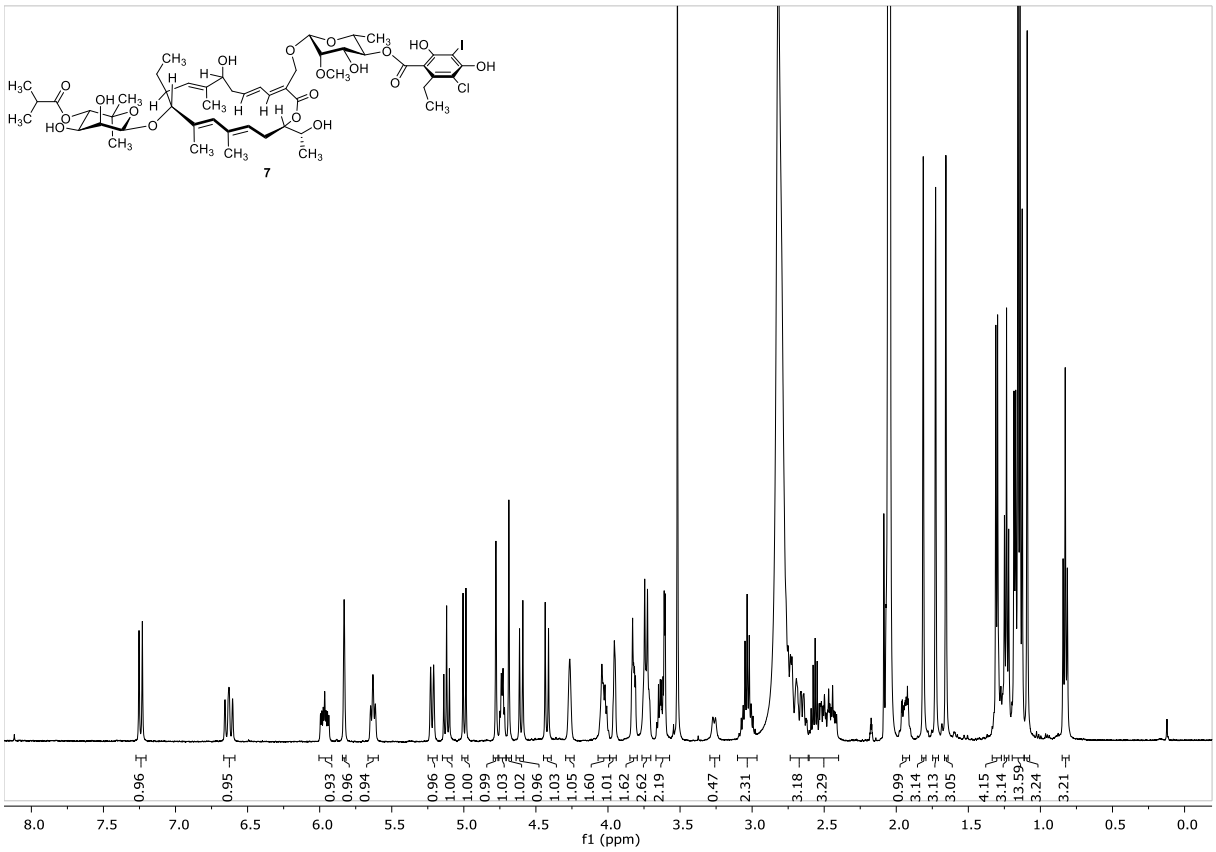
HSQC (500 MHz, acetone-d_6)



HMBC (500 MHz, acetone-d_6)



^{1}H NMR (500 MHz, acetone-d_6)



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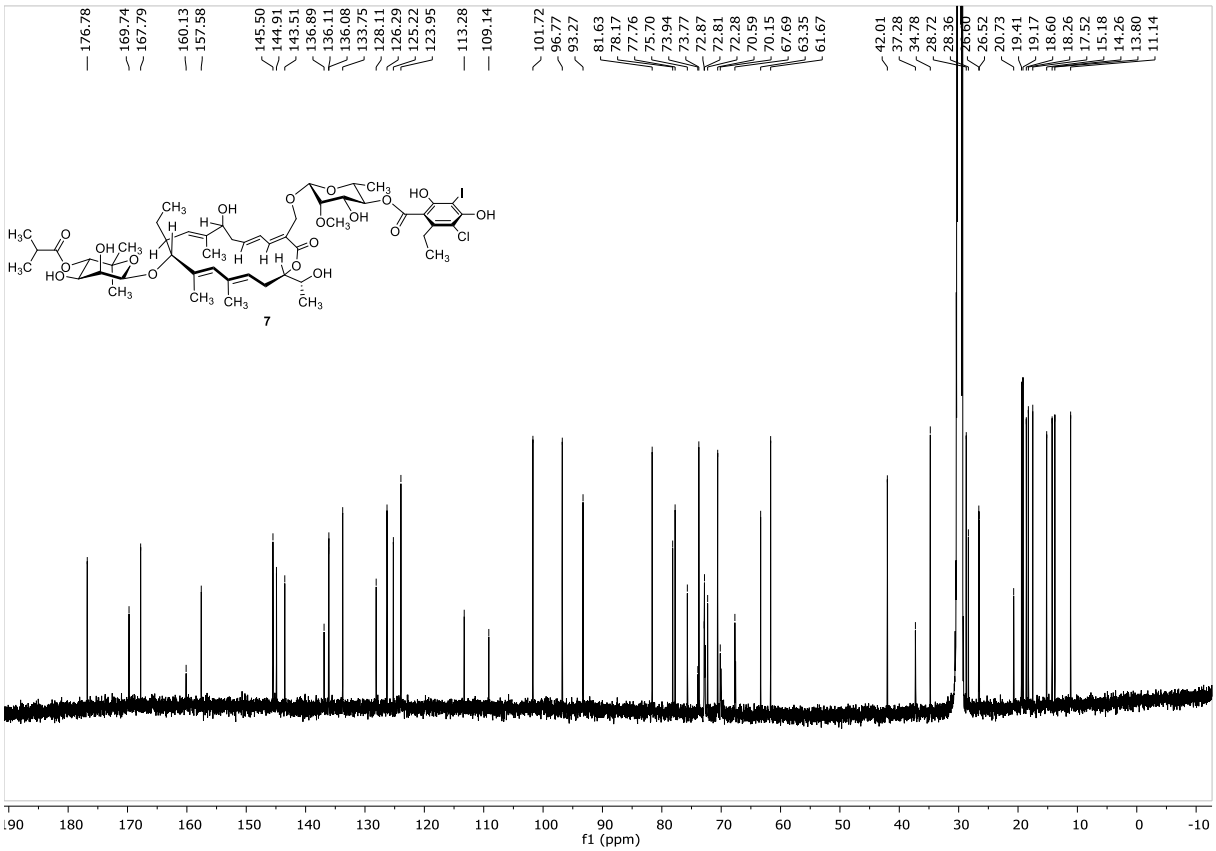

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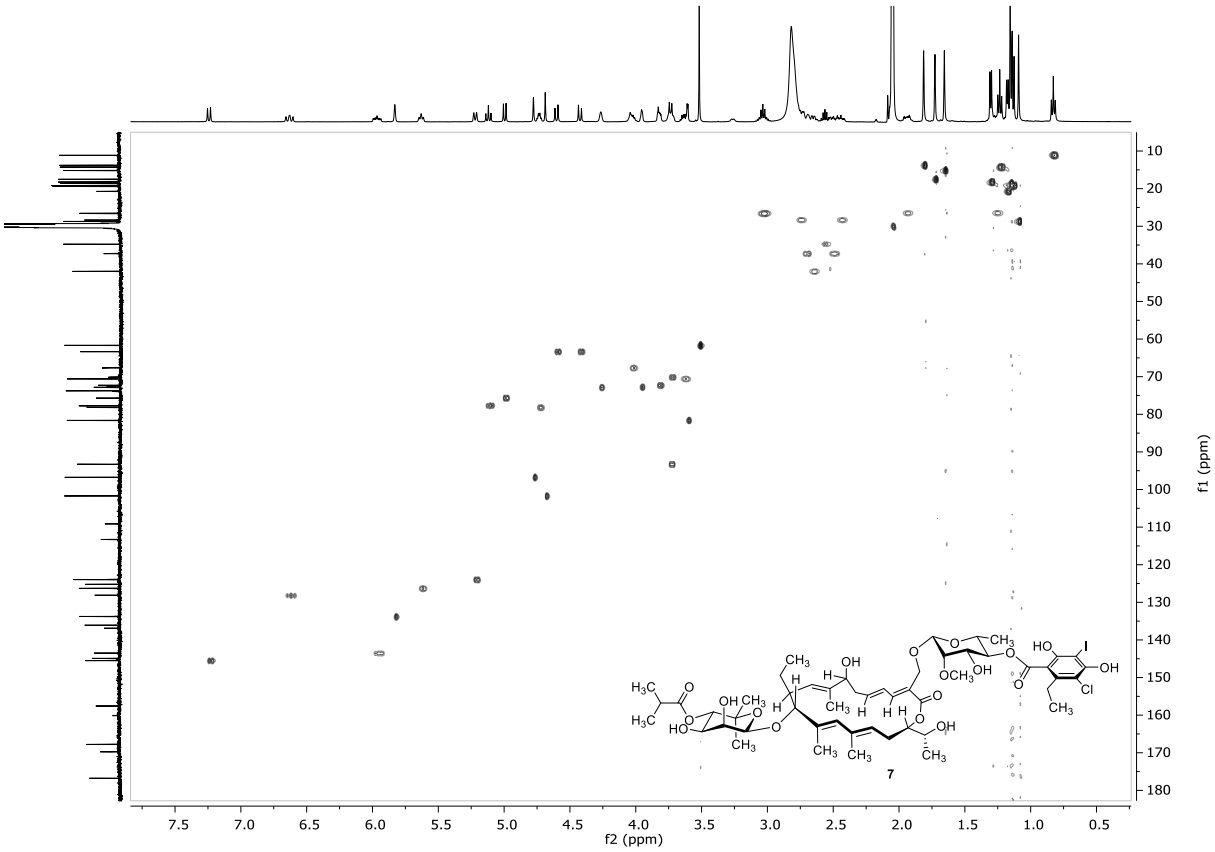

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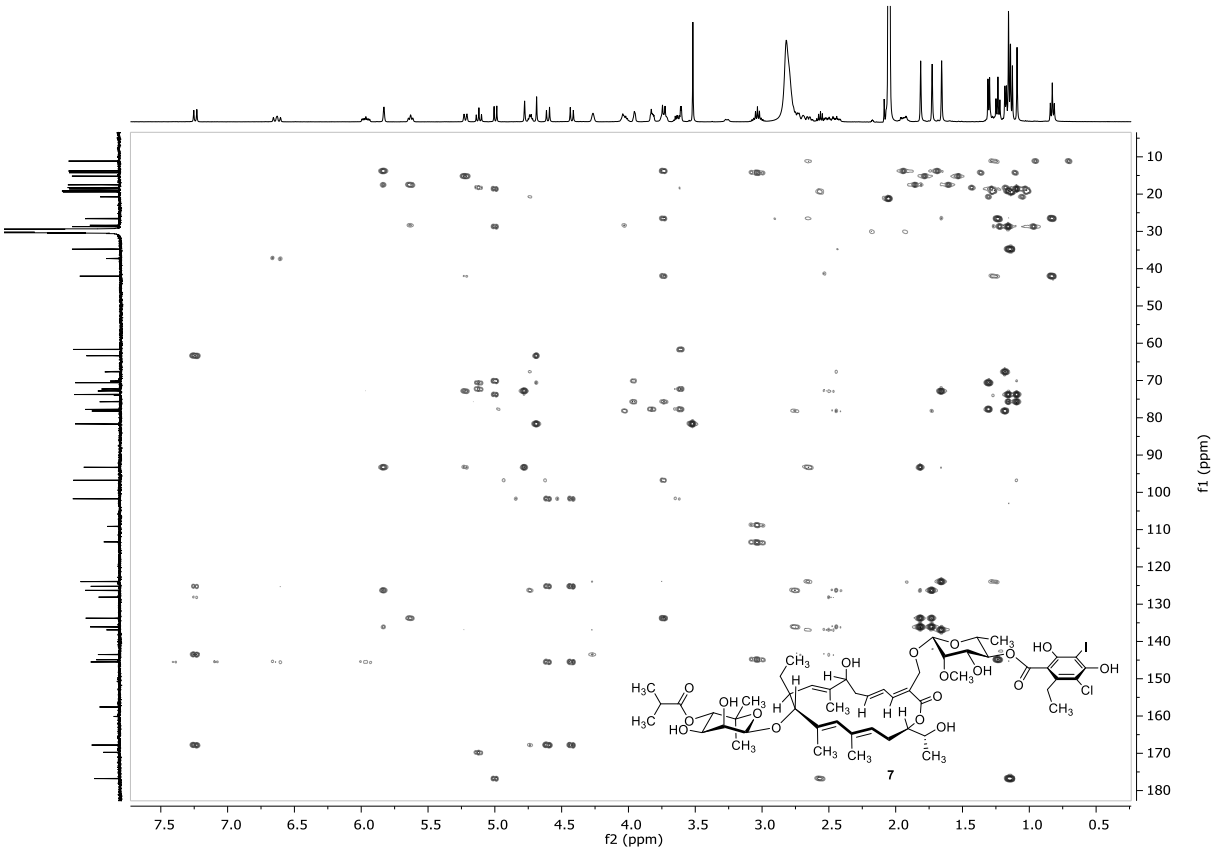

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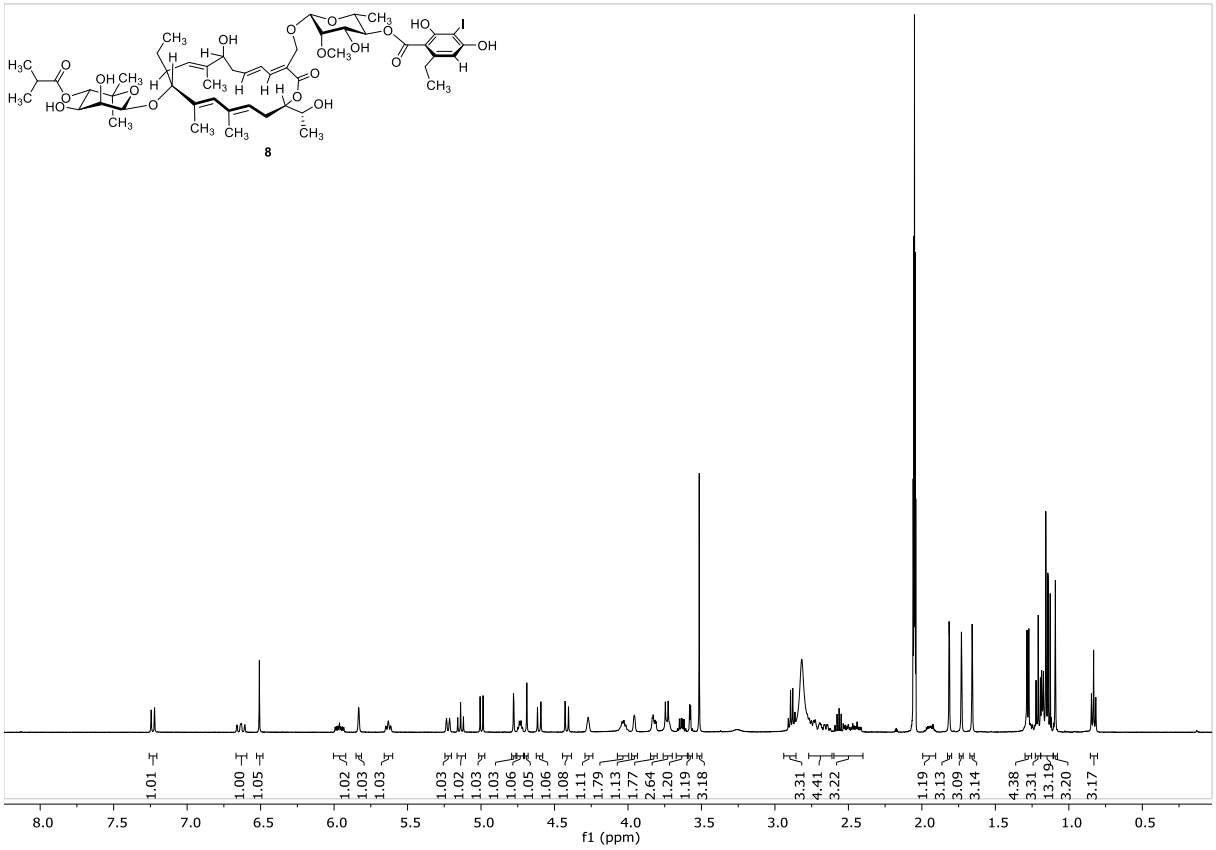

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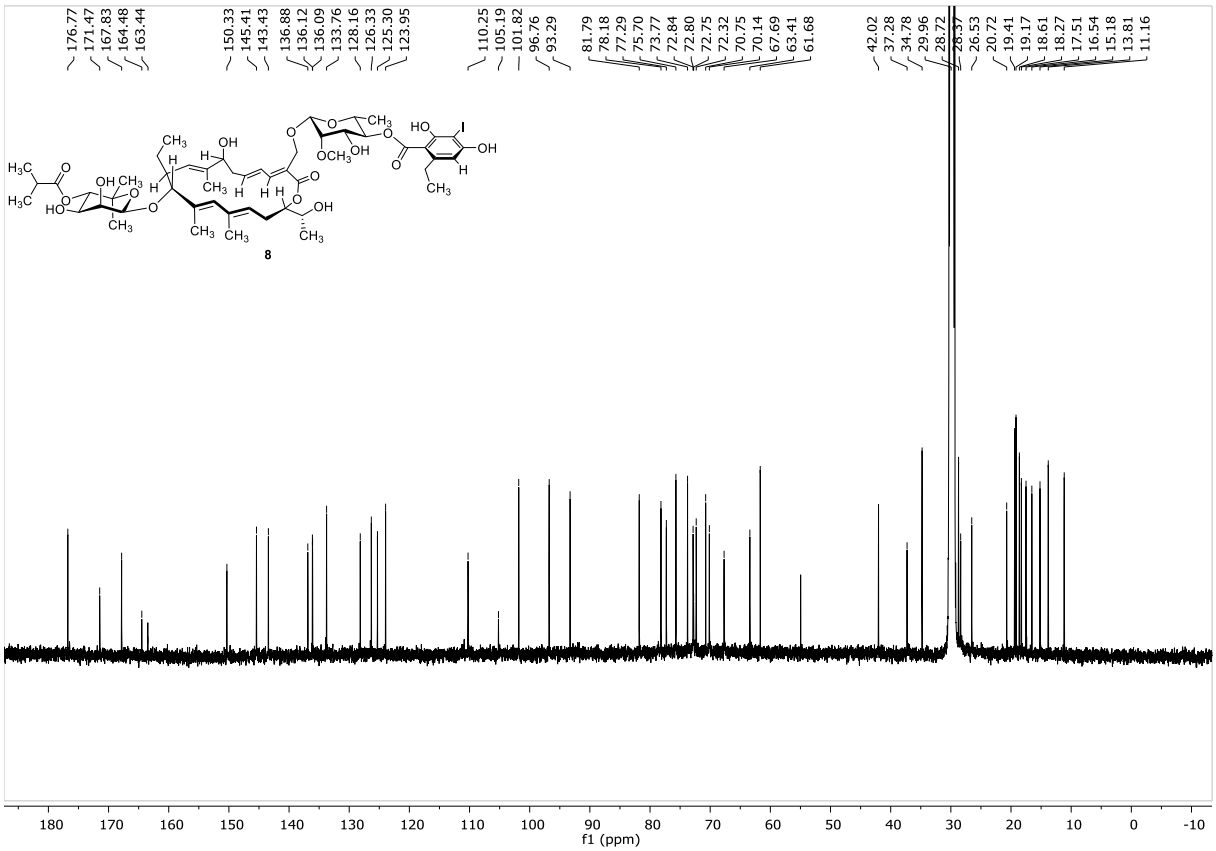

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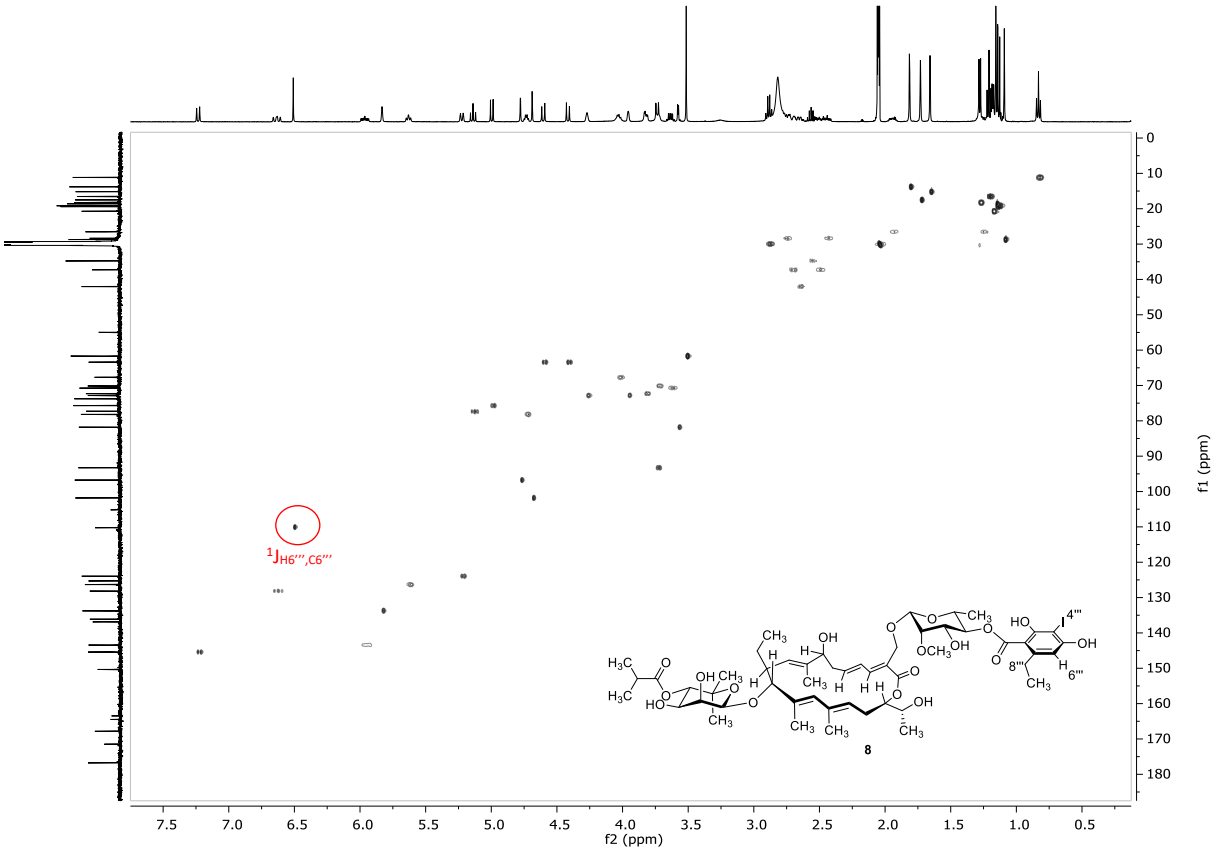

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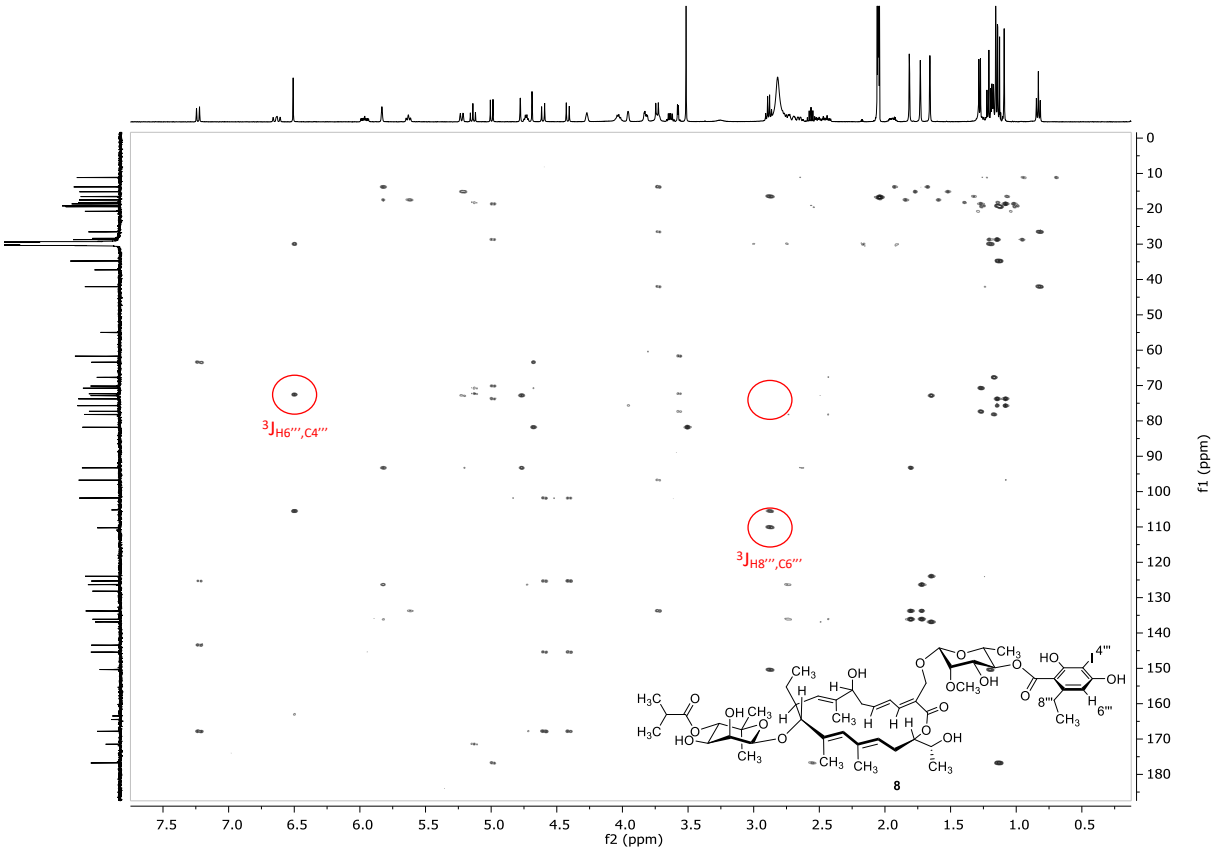

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