

Whole Genome Sequence Analysis and Homology Modelling of a 3C Like Peptidase and a Non-Structural Protein 3 of the SARS-CoV-2 Shows Protein Ligand Interaction with an Aza-Peptide and a Noncovalent Lead Inhibitor with Possible Antiviral Properties

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Abstract

The family of viruses belonging to Coronaviridae mainly consist of virulent pathogens that have a zoonotic property, Severe Acute Respiratory Syndrome (SARS-CoV) and Middle East Respiratory Syndrome (MERS-CoV) of this family have emerged before and now the SARS-CoV-2 has emerged in China. Characterization of spike glycoproteins, polyproteins and other viral proteins from viruses are important for vaccine development. Homology modelling of these proteins with known templates offers the opportunity to discover ligand binding sites and explore the possible antiviral properties of these protein ligand complexes. Any information emerging from these protein models can be used for vaccine development. In this study we did a complete bioinformatic analysis, sequence alignment, comparison of multiple sequences and homology modelling of the SARS-CoV-2 whole genome sequences, the spike protein and the polyproteins for homology with known proteins, we also analysed receptor binding sites in these models for possible binding with ligands that exhibit antiviral properties. Our results showed that the tertiary structure of the polyprotein isolate SARS-CoV-2_HKU-SZ-001_2020 had 98.94 percent identity with SARS-Coronavirus NSP12 bound to NSP7 and NSP8 co-factors. Our results indicate that a part of the viral genome (residues 3268 -3573 in Frame 2 with 306 amino acids) of the SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3) when modelled with template 2a5i of the PDB database had 96 percent identity with a 3C like peptidase of SARS-CoV which has ability to bind with Aza-Peptide Epoxide (APE) which is known for irreversible inhibition of SARS-CoV main peptidase. This region was conserved in 98 genomes of SARS-CoV-2. The part of the genome (residues 1568-1882 in Frame 2 with 315 amino acids) when modelled with template 3e9s of the PDB database had 82 percent identity with a papain-like protease/deubiquitinase which when complexed with ligand GRL0617 acts as inhibitor which can block SARS-CoV replication. This region was conserved in 91 genomes of SARS-CoV-2. It is possible that these viral inhibitors can be used for vaccine development for the SARS-CoV-2.

Introduction

More than a decade has passed since the emergence human Coronavirus that caused Severe Respiratory Syndrome (SARS-CoV) and it is about 7 years since the emergence of another

34 type of Coronavirus - Middle East Respiratory Syndrome (MERS-CoV) and now the SARS-
35 CoV-2 has emerged in China. This repeated onslaught of these viruses goes to show that it can
36 assume pandemic proportions at any time and at any place.

37 The family of viruses belonging to Coronaviridae mainly consist of virulent pathogens that
38 have a zoonotic property and this large family of corona viruses, have been known to be
39 circulating in animals including camels, cats and bats. It has been seen in the past that Severe
40 Acute Respiratory Syndrome associated coronavirus (SARS-CoV) and Middle East
41 Respiratory Syndrome-associated coronavirus (MERS-CoV) belonging to this family of
42 viruses can be transmitted from animals to humans and can cause respiratory diseases. Human
43 to human transmission on this virus has been a concern and due to this search for antiviral
44 compounds and vaccine development for this family of virus becomes the need of the hour.

45 The SARS was first seen in 2002 in Guangdong province of China, and later spread globally
46 and has caused close to about 8096 cases (WHO 2004, de Wit et al., 2016). In 2012, a novel
47 betacoronavirus, designated Middle East respiratory syndrome coronavirus or MERS-CoV
48 associated with severe respiratory disease in humans, emerged in the Arabian Peninsula (de
49 Wit et al., 2013).

50 The World Health Organization (WHO), China Country Office was informed of cases of
51 pneumonia of unknown aetiology in Wuhan City, Hubei Province, on 31 December 2019
52 (WHO 2020). A novel coronavirus currently termed SARS-CoV-2 was officially announced
53 as the causative agent by Chinese authorities on 7 January 2020. As on 20 Feb 2020 China's
54 National Health Commission reported that there are 74,280 confirmed cases in China (Fig.1).
55 The World Health Organization reported 924 confirmed cases in 25 countries outside China
56 (WHO Situation Report 29 2020). This novel corona virus has been designated as SARS-CoV-
57 2.

58

59 Coronaviruses are RNA viruses and have large genomes structures and due to this they can
60 have high error in replication as compared to host genomes. It is also known that various CoVs
61 can do effective recombination of their genomes after infecting host cells (Luo et al 2018). This
62 recombination can be a factor for their evolution to novel types which may have new animals
63 as their intermediate hosts. These factors give the CoVs high adaptive ability and the capability
64 to jump across species and have a relatively large host range.

65 Characterization of Spike glycoproteins from viruses are important for vaccine development.
66 Any information coming from the protein model can be used for vaccine development. *In Silico*
67 Epitope, polypeptide and spike protein-based peptide vaccine designing for infectious viruses
68 is a way that can hasten the process of vaccine development. Spike (S) protein, polypeptide and
69 other viral proteins of the SARS-CoV-2 as a target for the development of vaccines and
70 therapeutics for the prevention and treatment of infection is an important approach. In the case
71 of SARS-CoV, these proteins can mediate binding of the virus with its receptor and promotes
72 the fusion between the viral and host cell membranes and virus entry into the host cell, hence
73 peptides, antibodies, organic compounds and short interfering RNAs that interact with the spike
74 protein can have a potential role in vaccine development (Du et al 2009).

75 There are multiple domain functions that are active in the replication of the coronavirus and
76 these domains are present in a protein designated as Non-structural protein 3 (nsp3) which is
77 the largest protein in the coronavirus genome (Chen et al 2015). 3C like protease (3CLpro) and
78 Papain like Protease (PLpro) are two important class of proteases that are involved in the
79 process of translation of the polypeptide from the genomic RNA to protein components that
80 are required structurally or non-structurally for replication and packaging of new generation
81 viruses (Liu et al 2020)

82 We hypothesised that there can be some proteins in the large chunk of proteins in the SARS-
83 CoV-2 that could have homology with the Non-structural protein 3 (nsp3) SARS CoV and
84 these proteins can possibly have binding sites with ligands that can bind with known ligand
85 with antiviral properties.

86 Here in this study we did a complete bioinformatic analysis, sequence alignment, comparison
87 of multiple sequences of the SARS-CoV-2 whole genome sequences, the Spike protein and
88 the polyproteins for homology with known spike proteins and also analysed receptor binding
89 sites for possible vaccine development.

90 **Materials and Methods**

91 Six complete viral genome sequences, seven polyproteins (RdRp region) and seven
92 glycoproteins available on NCBI portal on 4 Feb 2020 were taken for analysis. The sequence
93 details and GenBank accession numbers are listed in Supplementary Table 1. Amongst the
94 seven polyproteins, five are of Wuhan pneumonia virus isolate SARS-COV-2 and two
95 sequences are of Wuhan pneumonia virus isolate SI200040-SP. The seven Glycoproteins are
96 of the same isolate, Wuhan pneumonia virus isolate SARS-COV-2.

97 The available polyproteins (RdRp region) and glycoprotein sequences were retrieved from
98 Genbank, NCBI (Benson et al., 2000). These sequences were translated to amino acid
99 sequences using sorted six frame translation with Bioedit (Hall et al., 2011). Multiple sequence
100 alignment of the translated protein sequences was performed and phylogenetic tree was
101 constructed using Mega-X (Kumar et al., 2018). The alignment shows that amongst the seven
102 polyproteins, five sequences were identical being from the same isolate and two other
103 sequences of the other isolate are identical. Similar analysis of the seven glycoproteins was
104 done, all the seven glycoprotein sequences were found to be identical. Therefore, further
105 analysis was carried out for three sequences.

1. MN938385.1 SARS-CoV-2 virus isolate SARS-COV-2 _HKU-SZ-001_2020 ORF1ab polyprotein, RdRp region, (orf1ab) gene, partial cds: 0 to 284: Frame 3 95 aa

2. MN970003.1 SARS-CoV-2 virus isolate SI200040-SP orf1ab polyprotein, RdRP region, (orf1ab) gene, partial cds: 2 to 289: Frame 2 96 aa

3. MN938387.1 SARS-CoV-2 virus isolate SARS-COV-2 _HKU-SZ-001_2020 surface glycoprotein (S) gene, partial cds: 1 to 105: Frame 1 35 aa

Expasy proteomics server (Gasteiger et al., 2003) was used to study the protein sequence and structural details. These peptides were studied for their physio-chemical properties using the tool Protparam (Gasteiger et al., 2005). The secondary structure analysis was done using Chou and Fasman algorithm with CFSSP (Kumar, 2013). To generate the 3D structure from the fasta sequence, homology modelling was performed and the templates were identified. The model was built using the template with highest identity. Swiss-model (Schwede et al., 2003) was used to build and validate the 3D model, structural assessment was also performed to validate the model built.

Complete genome sequence of the SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3) which has 29903 bp ss-RNA linear was translated sorted 6 frame with minimum ORF of 20 with any start codon and the resultant protein sequence was used for homology modelling, homology models were done with large chunks of proteins 21503 to 25381 in Frame 2 with 1293 amino acids, 13450 to 21552 in Frame 1 with 2701 amino acids and 254 to 13480 in Frame 2 with 4409 amino acids.

SWISS-MODEL server was used for homology modelling (Waterhouse et al 2018) where computation was on ProMod3 engine which is based on Open Structure (Biasini et al 2013). Structural information is extracted from the template, sequence alignment is used to define insertions and deletions.

Protein ligand interaction profile with hydrogen bonding, hydrophobic interactions, salt bridges and π -Stacking was done with PLIP server (Salentin et al., 2015)

Results and Discussion

The physico- chemical properties and primary structure parameters of the 7 polyproteins RdRp region of the SARS-CoV-2 virus isolate is given in Table 1. RdRP forms an important part of the viral genome where in the RNA viruses its function is to catalyze the synthesis of the RNA strand complementary to a given RNA template.

The isolates SI200040-SP orflab polyprotein and the isolate SI200121-SP orflab polyprotein had 2 reading frames as compared to the rest of the isolates which had 3 reading frames. The presence of multiple reading frames suggests the possibility of overlapping genes as seen in many virus and prokaryotes and mitochondrial genomes. This could affect how the proteins are made. The number of amino acid residues in all the polyproteins were the same except one isolate SI200040-SP which had one amino acid more than the other polyproteins. The extinction coefficients of the two isolates SI200040-SP orflab polyprotein and the isolate SI200121-SP orflab polyprotein was much higher compared to the rest of the polyproteins. The extinction coefficient is important when studying protein-protein and protein-ligand interactions. The instability index of these two isolates was also high when compared to the others indicating that these two isolates are instable. Regulation of gene expression by polyprotein processing is known in viruses and this is seen in many viruses that are human pathogens (Yost et al 2013).

The isolates here like many other viruses may be using replication strategy which could involve the translation of a large polyprotein with subsequent cleavage by viral proteases. The two isolates SI200040-SP orflab polyprotein and the isolate SI200121-SP orflab polyprotein also showed shorter half-lives as compared to the other isolates indicating that they are susceptible to enzymatic degradation.

The tertiary structure analysis of the isolate SARS-CoV-2 _HKU-SZ-001_2020 ORF1ab polyprotein is given in Table 2. It is seen that the polyprotein has a 98.94 percent identity with PDB structure 6nur.1.A which is a hetero-1-2-1-mer. The polyprotein is an RNA directed RNA polymerase. The protein is identical to the SARS-Coronavirus NSP12 bound to NSP7 and NSP8 co-factors (Kirchdoerfer and Ward 2019). In SARS it is basically a nonstructural protein with NSP12 being the RNA dependent RNA polymerase and the co factors NSP 7 and NSP 8 having the function of forming hexadecameric complexes and also act as processivity clamp for RNA polymerase and primase (Fehr et al., 2016). This structure as in SARS CoV here in SARS-CoV-2 may be involved in the machinery of core RNA synthesis and can be a template for exploring antiviral properties.

The phylogenetic tree of the seven polyproteins is shown in Fig.2. It is seen that two polyproteins were distinctly different from the rest. The Phylogenetic tree of the seven glycoproteins of the SARS-CoV-2 virus isolate is shown in Fig.3, it is seen that the glycoproteins are similar in all the isolates. Multiple alignment of the Polyproteins of the SARS-CoV-2 is shown in Supplementary Fig.1.

This structure as in SARS CoV here in SARS-CoV-2 may be involved in the machinery of core RNA synthesis and can be a template for exploring antiviral properties. Based on its functions in the SARS CoV and its identity to the SARS-CoV-2, it is possible that it has the same functions in SARS-CoV-2 an RNA polymerase which does de novo initiation and primer extension with possible exonuclease activities, the activity itself being primer dependent useful for understanding the mechanism of SARS-CoV-2 replication and can be used as an antiviral target (Te Velthuis et al 2012; Te Velthuis et al 2010; Subissi et al 2014; Subissi et al 2014).

The polyprotein also has an identity of 19.74 percent with an ABC-type uncharacterized transport system periplasmic component-like protein, this protein is known to be a substrate binding protein and possible binding can be explored here (Bae et al 2019).

The homology model developed from the residues 254 to 13480 in Frame 2 with 4409 amino acids from the Complete genome sequence of the SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3) which has 29903 bp with linear ss-RNA linear showed interesting template alignments, in all the model aligned with 50 templates from the PDB database with most of them being replicase polyprotein 1ab which is a SARS-CoV papain-like protease (Daczkowski 2017). The maximum similarity of 97.3 percent was with template structure of a Nsp9 protein from SARS-coronavirus indicating that this novel coronavirus has high degree of similarity with the SARS-coronavirus and this can be used for gaining insights into vaccine development. Nsp 9 is an RNA binding protein and has an oligosaccharide/oligonucleotide fold-like fold, this protein can have an important function in the replication machinery of the virus and can be important when designing antiviral for this virus (Egloff et al 2004).

Two models were developed, one from residues 3268 -3573 in Frame 2 with 306 amino acids and the other from the part of the genome residues 1568-1882 in Frame 2 with 315 amino acids of the SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3). The models had similarity with the 3C like proteinase and a papain-like protease/deubiquitinase protein which are known antiviral drug targets. Ligand binding with these proteins and their action is on viral replication and inactivation can be useful in stopping the viral replication (Baez-Santos et al 2015).

201 The homology models of the 4409 amino acid residues of the whole genome of the SARS-
202 CoV-2 virus isolate Wuhan-Hu-1 with the ligand association with templates 2a5i and 3e9s are
203 shown in Fig. 4 and Fig. 5 respectively.

204 The statistics of structural comparison with PDB templates is given in Table 5, it is seen that
205 the proteins from the SARS-CoV-2 are significantly close to the proteins of SARS CoV and
206 the amino acid alignment in the binding region is the same in both the viruses.

207 The alignment of the 305 residues from 3268-3573 aa of the Novel Coronavirus COVI-19 with
208 the template 2a5i is shown in Fig.6 and the alignment of the 315 residues from 1568-1882 aa
209 of the Novel Coronavirus COVI-19 with the template 3e9s is shown in Fig.7.

210 A PSI-BLAST of a length of 306 amino acid residues 3268 -3573 in Frame 2 from the SARS-
211 CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3) was conducted
212 to ascertain the conservation of these amino acids in 101 genome sequences of SARS-CoV-2
213 and it was found that there was a complete match with 98 of these genomes and 99.67 percent
214 identity with 3 genomes in the orf1a polyprotein of the virus. The fact that the region is
215 conserved in all these SARS-CoV-2 sequences further emphasizes this ligand interaction of
216 Aza-Peptide epoxide with the protein can be used as an antiviral in SARS-CoV-2. Similarly
217 A PSI-BLAST of a length of 315 amino acid residues 3268 -3573 in Frame 2 with 315 amino
218 acid residues 1568-1882 in Frame 2 from SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank
219 Accession Number MN908947.3) was conducted to ascertain the conservation of these amino
220 acids in 101 genome sequences of SARS-CoV-2 and it was found that there was a complete
221 match with 91 of these genomes and 99.68 percent identity with 10 genomes in the orf1a
222 polyprotein of the virus. The fact that the region is conserved in all these SARS-CoV-2
223 sequences further emphasizes this ligand interaction of ligand GRL0617 with the protein can
224 be used as an antiviral in SARS-CoV-2.

The important templates that aligned with this 4409 amino acid residues of the whole genome of the SARS-CoV-2 virus isolate Wuhan-Hu-1 were 2a5i of the PDB database which is a crystal structure of SARS coronavirus main peptidase inhibited by an Aza-Peptide epoxide in the space group C2 (Lee et al 2005) and 3e9s of the PDB database which is new class of papain-like protease/deubiquitinase which when combined with ligand GRL0617 acts as inhibitors blocking SARS virus replication (Ratia et al 2008). The model with template 2a5i of the PDB database shows that Aza-Peptide Epoxide (APE; $k_{inact}/K_i=1900(\pm 400) \text{ M}^{-1} \text{ s}^{-1}$) which is a known anti SARS agent can be used to develop a molecular target with irreversible inhibitor properties. The protein ligand interaction analysis of the Novel Coronavirus C3 like peptidase and aza-peptide epoxide is shown in Fig.8. The substrate binding properties and structural and chemical complementarity of this Aza-Peptide Epoxide can be explored as an anti - Coronavirus SARS-COV-2 agent. The APE which is ethyl (2S)-4-[(3-amino-3-oxo-propyl)-[[[(2S)-2-[[[(2S)-4-methyl-2-phenylmethoxycarbonylamino-pentanoyl]amino]-3-phenyl-propanoyl]amino]amino]-2-hydroxy-4-oxo-butanoate structure is shown in Fig.9.

The model with template 3e9s of the PDB database shows that the Coronavirus viral protein can have a ligand which is a papain-like protease (PLpro) that is known to be a potent inhibitor of viral replication in SARS (Ratia et al 2008).

The two parts of the Main protein from the whole genome of the SARS-CoV-2 aligned with two SAR proteins and the ligand binding sites were similar, the alignment positions, number of amino acids and ligand and the interacting residues is given in Table 3

The complete genome of MN908947.3 SARS-CoV-2 virus isolate Wuhan-Hu-1 encodes a 4409aa long protein along with the other glycoproteins and polyproteins. The homology modelling of this protein showed sequence and structural alignment with two SARS proteases

with structural accession numbers 3e9s.1 and 2a5i.1 at positions 1568-1882 and 3268-3573 respectively. Reports suggests inhibition of virus replication by TTT ligand and an aza-peptide epoxide inhibiting the main peptidase. The structural similarity of these templates are 83% and 96% respectively. The multiple sequence alignment shows complete conservation of the sequence suggesting a high degree of homology. The protein ligand interaction analysis of the Novel Coronavirus non structural protein and papain-like protease is shown in Fig. 10.

The Comparison of Hydrophobic interaction, hydrogen bonding, salt bridges of the constructed model of the Novel Coronavirus protein from region 3268-3573 aa to ligand AZP with Hydrophobic interaction, hydrogen bonding, salt bridges of the template 2a5i is given in Suppl. Table 2, when comparing both it is seen that the binding properties are the same except for the presence of water bridge in the template 2a5i.

The Comparison of Hydrophobic interaction, hydrogen bonding, π -Stacking of the constructed model of the Novel Coronavirus protein from region 1568-1882 aa to ligand Small molecule Noncovalent Lead Inhibitor with the Hydrophobic interaction, hydrogen bonding, π -Stacking of the template 3e9s is given in Suppl. Table 3, when comparing both it is seen that the binding properties are the same except or an additional π -Stacking at Tyr in the template 2a5i. This shows that there is high possibility of binding of these antiviral compounds with the regions of Novel Coronavirus protein that is in homology with the SARS protein.

Comparison of the hydrophobic interaction of the biding of the ligand AZP between the SARS-CoV-2 protein and the template 2a5i of SARS CoV is shown in Fig.11 and the comparison of the hydrophobic interaction of the biding of the ligand AZP between the SARS-CoV-2 protein and the template 3e9s of SARS CoV is shown in Fig.12. It is seen that the interaction is the

272 same in both proteins with the same amino acids participating in the interaction indicating that
273 there is a possibility that these ligands with antiviral properties can bind to the new virus.

274 The similarity in the amino acids involved in the Hydrophobic interactions which are short range
275 interactions and have an important role in the affinities of the ligands and receptors shows that
276 the proteins of the SARS-CoV-2 may bind with the same affinity as seen in the SARS CoV
277 and this also shows a similar action of the ligand as seen in SARS CoV, indicating that these
278 ligands can be used as antivirals in the SARS-CoV-2.

279 The targeting of this part of the genome of the SARS-CoV-2 with the antiviral compounds
280 which have shown to bind in the similar region of the SARS virus can have implication in the
281 development of an effective antiviral compound against the SARS-CoV-2. The SARS-CoV-2
282 shows homology with the SARS coronaviral proteases, papain-like protease (PLpro) and 3C-
283 like protease (3CLpro), these proteins have the function of processing the viral polyprotein and
284 also they perform the function of stripping ubiquitin and the ubiquitin-like interferon (IFN)-
285 stimulated gene 15 (ISG15) from the hosts to facilitate coronavirus replication and help in
286 evading immune response of the host, these inhibitors can also have a role in disrupting
287 signalling cascades in infected cells and protecting the uninfected cells.

288 The chemical GRL0617 is 5-Amino-2-methyl-N-[(1R)-1-(1-naphthalenyl)ethyl]benzamide
289 and is known to inhibit the papainlike protease that is present in SARS CoV. This protease is
290 a potential target for antiviral compounds (Chaudhuri et al., 2011). We found the SARS-CoV-
291 2 has homology with this and the binding sites for this in the structural protein of the SARS-
292 CoV-2 is the same (Table 4). This compound inhibits the enzyme that is required for the
293 cleavage of the viral protein from the virus in SARS CoV, it also cleaves ubiquitin and has a
294 structural homology with the Deubiquitinases (DUBs) of the Ubiquitin-Specific Proteases
295 Compound GRL0617 binds in the S4 and S3 enzyme subsite that gets the C terminal tail of the

Ubiquitin (King and Finley 2014; Schauer et al., 2019). Our results show that Aza-Peptide Epoxide an irreversible protease inhibitor and GRL0617 a viral replication inhibitor can be used to develop inhibitors of the Novel Coronavirus SARS-COV-2.

References

Bae, J.E., Kim, I.J., Kim, K.J. and Nam, K.H., 2018. Crystal structure of a substrate-binding protein from *Rhodothermus marinus* reveals a single α/β -domain. Biochemical and Biophysical Research Communications, 497(1), pp.368-373.

Baez-Santos, Y.M., John, S.E.S. and Mesecar, A.D., 2015. The SARS-coronavirus papain-like protease: structure, function and inhibition by designed antiviral compounds. Antiviral Research, 115, pp.21-38.

Benson, D. A., Karsch-Mizrachi, I., Lipman, D. J., Ostell, J., Rapp, B. A., & Wheeler, D. L. (2000). GenBank. Nucleic Acids Research, 28(1), 15-18.

Biasini, M., Schmidt, T., Bienert, S., Mariani, V., Studer, G., Haas, J., Johnner, N., Schenk, A.D., Philippsen, A. and Schwede, T., 2013. OpenStructure: an integrated software framework for computational structural biology. Acta Crystallographica Section D: Biological Crystallography, 69(5), pp.701-709.

Chaudhuri, R., Tang, S., Zhao, G., Lu, H., Case, D.A. and Johnson, M.E., 2011. Comparison of SARS and NL63 papain-like protease binding sites and binding site dynamics: inhibitor design implications. Journal of molecular biology, 414(2), pp.272-288.

Chen, Y., Savinov, S.N., Mielech, A.M., Cao, T., Baker, S.C. and Mesecar, A.D., 2015. X-ray structural and functional studies of the three tandemly linked domains of non-structural

318 protein 3 (nsp3) from murine hepatitis virus reveal conserved functions. *Journal of*
319 *Biological Chemistry*, 290(42), pp.25293-25306.

320 Daczkowski, C.M., Dzimianski, J.V., Clasman, J.R., Goodwin, O., Mesecar, A.D. and Pegan,
321 S.D., 2017. Structural insights into the interaction of coronavirus papain-like proteases
322 and interferon-stimulated gene product 15 from different species. *Journal of Molecular*
323 *Biology*, 429(11), pp.1661-1683.

324 de Wit, E., Rasmussen, A.L., Falzarano, D., Bushmaker, T., Feldmann, F., Brining, D.L.,
325 Fischer, E.R., Martellaro, C., Okumura, A., Chang, J. and Scott, D., 2013. Middle East
326 respiratory syndrome coronavirus (MERS-CoV) causes transient lower respiratory tract
327 infection in rhesus macaques. *Proceedings of the National Academy of Sciences*,
328 110(41), pp.16598-16603.

329 de Wit, E., van Doremalen N., D. Falzarano, V. J. Munster, SARS and MERS: recent insights
330 into emerging coronaviruses. *Nat Rev Microbiol* 14, 523-534 (2016).

331 Du, L., He, Y., Zhou, Y., Liu, S., Zheng, B.J. and Jiang, S., 2009. The spike protein of SARS-
332 CoV—a target for vaccine and therapeutic development. *Nature Reviews*
333 *Microbiology*, 7(3), pp.226-236.

334 Egloff, M.P., Ferron, F., Campanacci, V., Longhi, S., Rancurel, C., Dutartre, H., Snijder, E.J.,
335 Gorbalenya, A.E., Cambillau, C. and Canard, B., 2004. The severe acute respiratory
336 syndrome-coronavirus replicative protein nsp9 is a single-stranded RNA-binding
337 subunit unique in the RNA virus world. *Proceedings of the National Academy of*
338 *Sciences*, 101(11), pp.3792-3796.

339 Fehr, A.R. and Perlman, S., 2015. Coronaviruses: an overview of their replication and
340 pathogenesis. In *Coronaviruses* (pp. 1-23). Humana Press, New York, NY.

341 Gasteiger, E., Gattiker, A., Hoogland, C., Ivanyi, I., Appel, R. D., & Bairoch, A. (2003).
 342 ExPASy: the proteomics server for in-depth protein knowledge and analysis. *Nucleic*
 343 *Acids Research*, 31(13), 3784-3788.

344 Gasteiger, E., Hoogland, C., Gattiker, A., Wilkins, M. R., Appel, R. D., & Bairoch, A. (2005).
 345 Protein identification and analysis tools on the ExPASy server. In *The proteomics*
 346 *protocols handbook* (pp. 571-607). Humana press.

347 Hall, T., Biosciences, I., & Carlsbad, C. (2011). BioEdit: an important software for molecular
 348 biology. *GERF Bull Biosci*, 2(1), 60-61.

349 King, R.W. and Finley, D., 2014. Sculpting the proteome with small molecules. *Nature*
 350 *chemical biology*, 10(11), p.870.

351 Kirchdoerfer, R.N. and Ward, A.B., 2019. Structure of the SARS-CoV nsp12 polymerase
 352 bound to nsp7 and nsp8 co-factors. *Nature Communications*, 10(1), pp.1-9.

353 Kumar, S., Stecher, G., Li, M., Knyaz, C., & Tamura, K. (2018). MEGA X: molecular
 354 evolutionary genetics analysis across computing platforms. *Molecular Biology and*
 355 *Evolution*, 35(6), 1547-1549.

356 Kumar, T. A. (2013). CFSSP: Chou and Fasman secondary structure prediction server. *Wide*
 357 *Spectrum*, 1(9), 15-19.

358 Lee, T.W., Cherney, M.M., Huitema, C., Liu, J., James, K.E., Powers, J.C., Eltis, L.D. and
 359 James, M.N., 2005. Crystal structures of the main peptidase from the SARS coronavirus
 360 inhibited by a substrate-like aza-peptide epoxide. *Journal of Molecular Biology*, 353(5),
 361 pp.1137-1151.

362 Liu, W., Morse, J.S., Lalonde, T. and Xu, S., 2020. Learning from the Past: Possible Urgent
 363 Prevention and Treatment Options for Severe Acute Respiratory Infections Caused by
 364 2019-nCoV. *ChemBioChem*.

365 Luo, C.M., Wang, N., Yang, X.L., Liu, H.Z., Zhang, W., Li, B., Hu, B., Peng, C., Geng, Q.B.,
 366 Zhu, G.J. and Li, F., 2018. Discovery of novel bat coronaviruses in south China that
 367 use the same receptor as Middle East respiratory syndrome coronavirus. *Journal of*
 368 *Virology*, 92(13), pp.e00116-18.

369 Ratia, K., Pegan, S., Takayama, J., Sleeman K., Coughlin, M., Baliji, S., Chaudhuri, R., Fu,
 370 W., Prabhakar, B.S., Johnson, M.E. and Baker, S.C., 2008. A noncovalent class of
 371 papain-like protease/deubiquitinase inhibitors blocks SARS virus replication.
 372 *Proceedings of the National Academy of Sciences*, 105(42), pp.16119-16124.

373 Salentin, S., Schreiber, S., Haupt, V.J., Adasme, M.F. and Schroeder, M., 2015. PLIP: fully
 374 automated protein–ligand interaction profiler. *Nucleic acids research*, 43(W1),
 375 pp.W443-W447

376 Schauer, N.J., Magin, R.S., Liu, X., Doherty, L.M. and Buhrlage, S.J., 2019. Advances in
 377 Discovering Deubiquitinating Enzyme (DUB) Inhibitors. *Journal of medicinal*
 378 *chemistry*.

379 Schwede, T., Kopp, J., Guex, N., & Peitsch, M. C. (2003). SWISS-MODEL: an automated
 380 protein homology-modeling server. *Nucleic Acids Research*, 31(13), 3381-3385.

381 Subissi, L., Imbert, I., Ferron, F., Collet, A., Coutard, B., Decroly, E. and Canard, B., 2014.
 382 SARS-CoV ORF1b-encoded nonstructural proteins 12–16: replicative enzymes as
 383 antiviral targets. *Antiviral research*, 101, pp.122-130.

384 Subissi, L., Posthuma, C.C., Collet, A., Zevenhoven-Dobbe, J.C., Gorbalenya, A.E., Decroly,
385 E., Snijder, E.J., Canard, B. and Imbert, I., 2014. One severe acute respiratory syndrome
386 coronavirus protein complex integrates processive RNA polymerase and exonuclease
387 activities. *Proceedings of the National Academy of Sciences*, 111(37), pp.E3900-
388 E3909.

389 Te Velthuis, A.J., Arnold, J.J., Cameron, C.E., van den Worm, S.H. and Snijder, E.J., 2010.
390 The RNA polymerase activity of SARS-coronavirus nsp12 is primer dependent.
391 *Nucleic acids research*, 38(1), pp.203-214.

392 Te Velthuis, A.J., van den Worm, S.H. and Snijder, E.J., 2012. The SARS-coronavirus nsp7+
393 nsp8 complex is a unique multimeric RNA polymerase capable of both de novo
394 initiation and primer extension. *Nucleic acids research*, 40(4), pp.1737-1747.

395 Waterhouse, A., Bertoni, M., Bienert, S., Studer, G., Tauriello, G., Gumienny, R., Heer, F.T.,
396 de Beer, T.A.P., Rempfer, C., Bordoli, L. and Lepore, R., 2018. SWISS-MODEL:
397 homology modelling of protein structures and complexes. *Nucleic Acids Research*,
398 46(W1), pp.W296-W303.

399 World Health Organization (WHO) 2004. [Accessed 11 Feb 2020]
400 https://www.who.int/csr/don/2004_05_18a/en/

401 World Health Organization (WHO). Coronavirus. Geneva: WHO; 2020 [Accessed 4 Feb
402 2020]. Available from: <https://www.who.int/health-topics/coronavirus>

403 Yost, S.A. and Marcotrigiano, J., 2013. Viral precursor polyproteins: keys of regulation from
404 replication to maturation. *Current Opinion in Virology*, 3(2), pp.137-142.

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406 **Figure Captions**

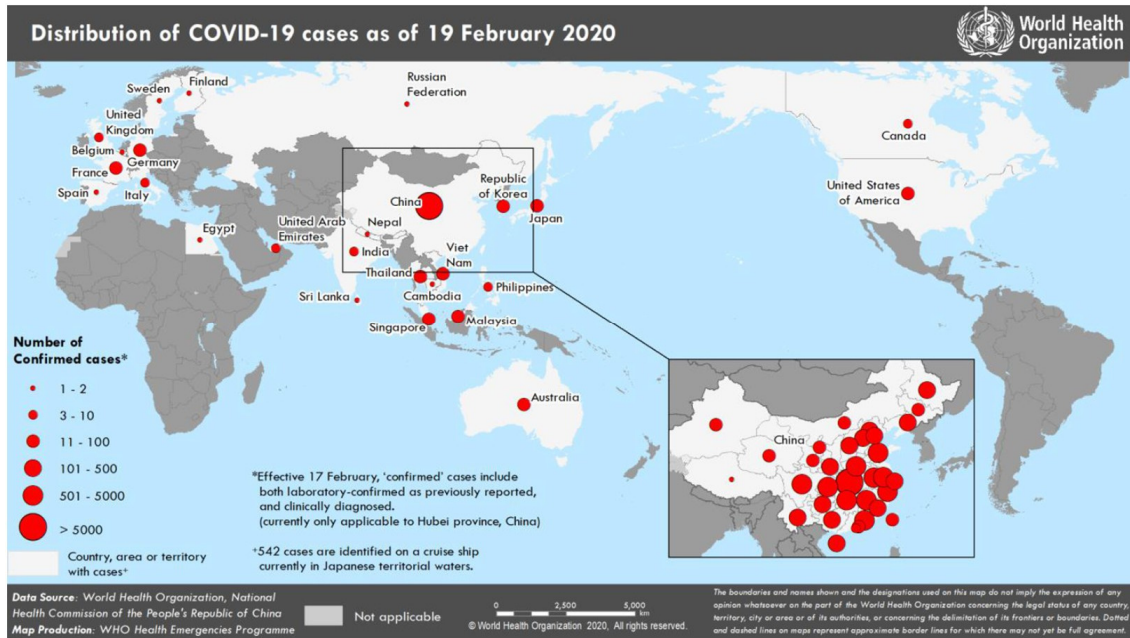
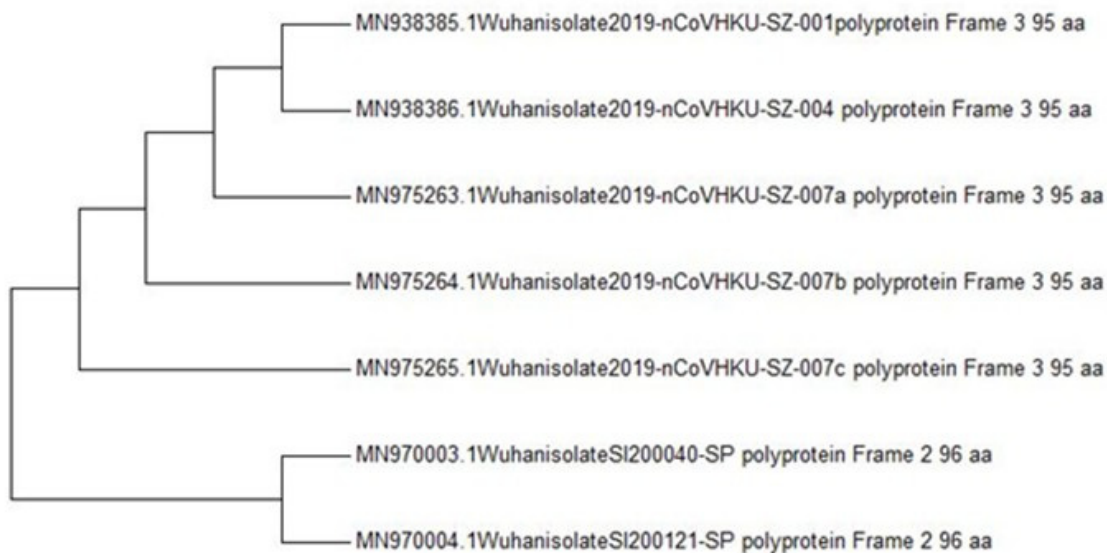
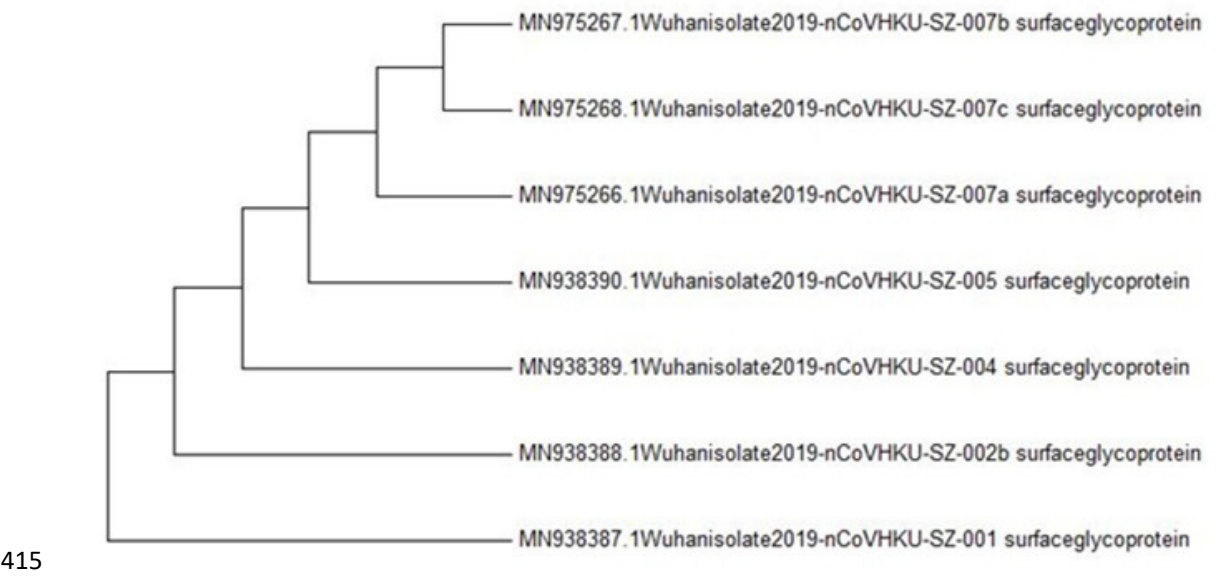


Fig.1 Countries, territories or areas with reported confirmed cases of SARS-COV-2 , 3
February 2020 Source WHO (https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200219-sitrep-30-covid-19.pdf?sfvrsn=6e50645_2)



413 **Fig.2** Phylogenetic tree of the seven polyproteins of Severe acute respiratory syndrome
414 coronavirus 2 isolate virus isolates



416 **Fig.3** Phylogenetic tree of the seven glycoproteins of Severe acute respiratory syndrome
417 coronavirus 2 isolate virus isolates

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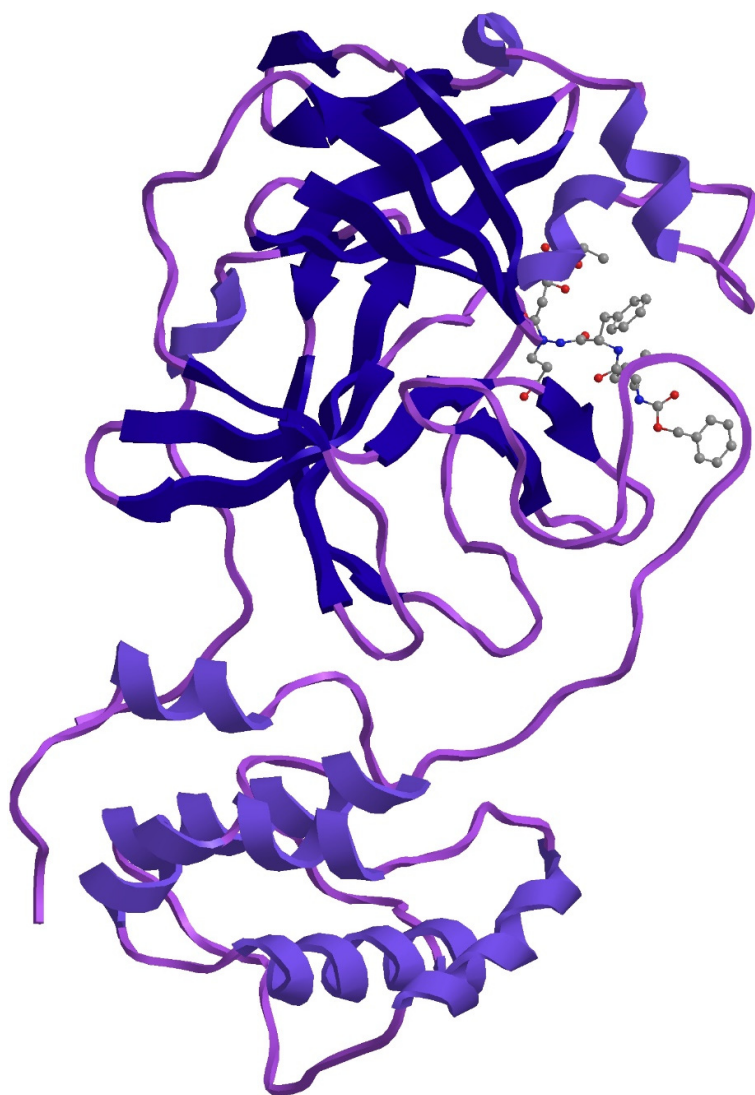
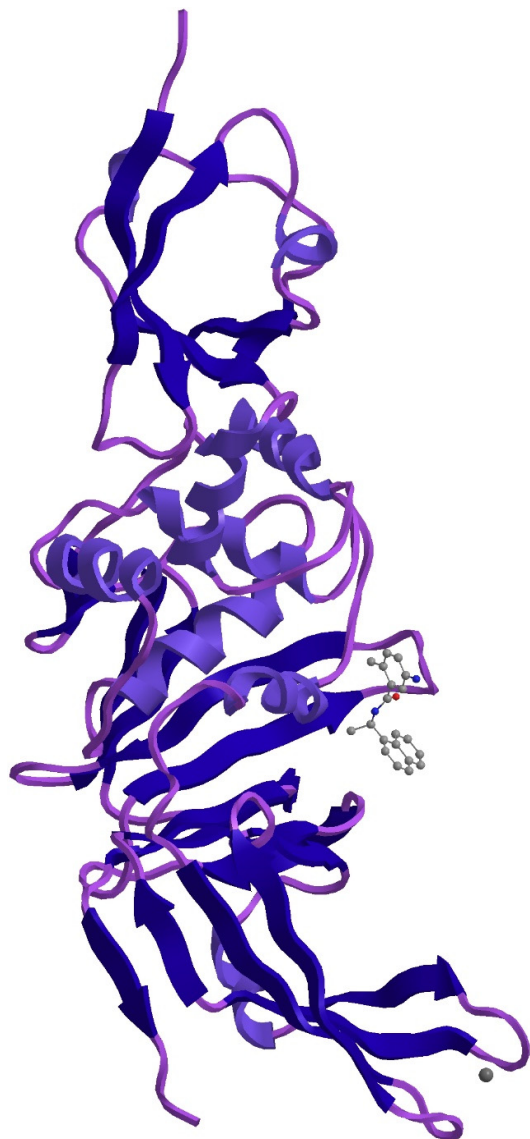


Fig. 4 Homology model with ligand binding of protein from amino acids 3268 -3573 in Frame 2 with 306 amino acids of the Complete genome sequence of the SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3) which has 29903 bp linear ss-RNA with 2a5i of the PDB database as template.

429



430

431 **Fig. 5** Homology model with ligand binding of protein from residues 1568-1882 in Frame 2
432 with 315 amino acids of the Complete genome sequence of the SARS-CoV-2 virus
433 isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3) which has 29903 bp
434 linear ss-RNA with 3e9s of the PDB database as template.

435

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437

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Seqres  SGFRKMAPPSGKVEGCMVQVTCGTTTNGLNLDLDDTVYCPRHVICTAEDMLNPNYEDLLIRKSNHSFLVQAGNVQLRVIGH 80
2a5i.1. (AB) SGFRKMAPPSGKVEGCMVQVTCGTTTNGLNLDLDDTVYCPRHVICTAEDMLNPNYEDLLIRKSNHSFLVQAGNVQLRVIGH 80

Seqres  SMQNCILRLKVDTSNPKTFKPKYKRVRIQPGQTFSVLACVNGSPSGVYQCAMRPNHTIKGSFLNGSCGSVGFNIDYDCVSFC 160
2a5i.1. (AB) SMQNCILRLKVDTSNPKTFKPKYKRVRIQPGQTFSVLACVNGSPSGVYQCAMRPNHTIKGSFLNGSCGSVGFNIDYDCVSFC 160

Seqres  YMHMMELPTGVHAGTDLEGRKFGYGFVDRQTAQAAGTDTTITLNVLAWLAAVINGDRWFLNRFTTTLNDFNLVAMKYNVE 240
2a5i.1. (AB) YMHMMELPTGVHAGTDLEGRKFGYGFVDRQTAQAAGTDTTITLNVLAWLAAVINGDRWFLNRFTTTLNDFNLVAMKYNVE 240

Seqres  FLTQDHDVDILGPLSAQTGIAVLDMCAALKELLQNGMNGRTILGSTILEDEFTPFDDVVRQCSGVTFQ 306
2a5i.1. (AB) FLTQDHDVDILGPLSAQTGIAVLDMCAALKELLQNGMNGRTILGSTILEDEFTPFDDVVRQCSGVTFQ 306

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438

439 **Fig. 6** Alignment of the 305 residues from 3268-3573 aa of the Novel Coronavirus COVI-19
 440 with the template 2a5i

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Seqres  ASMEVKTIKVFETVDNTNLHTQLVDMSMTYGGQFGFTYLDGADVTIKIKPHVNHGKTFEVLPSDDTLRSEAFEYVHTLDESFLGR 65
3e9s.1.A ASMEVKTIKVFETVDNTNLHTQLVDMSMTYGGQFGFTYLDGADVTIKIKPHVNHGKTFEVLPSDDTLRSEAFEYVHTLDESFLGR 65

Seqres  YMSALNHTKKWKFPQVGGTTSIKWADNNCYLSSVLLALQOLEVKFNAPALQEAAYRARAGDAANFCALILAYSNKTVGEIGDVE 170
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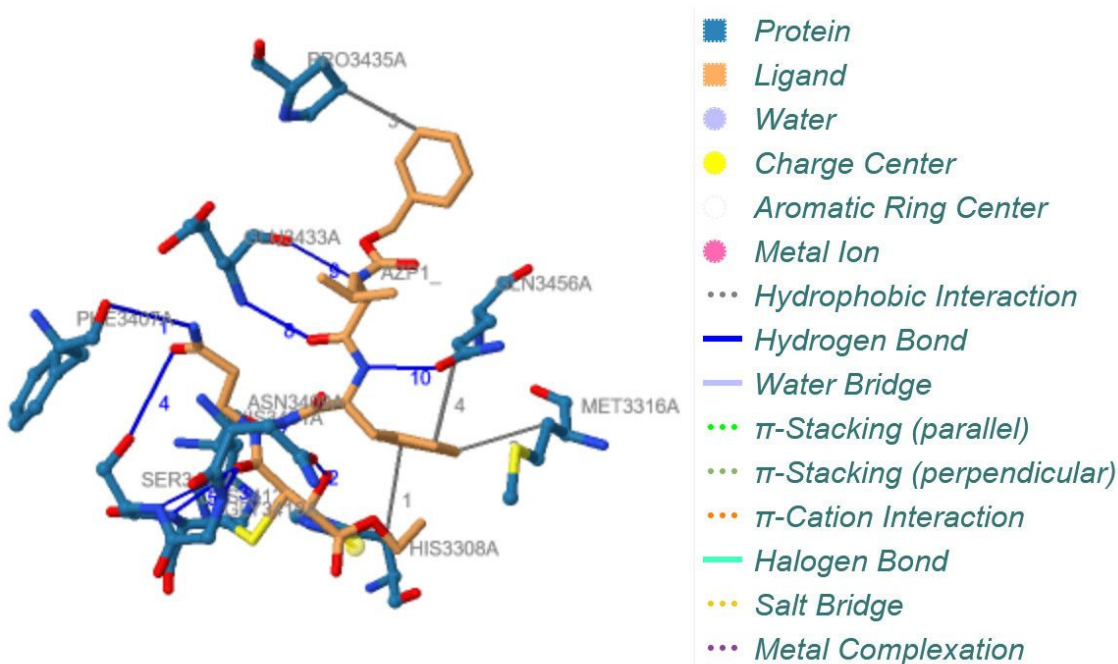
Seqres  TMTHLLQHANLESARKVLNVYCKKCGQKTTTLTGVEAVMYMGTLSYDNLKTGVSI PCVCGRDATQYLVQQESSFVMM SAPPAEYK 255
3e9s.1.A TMTHLLQHANLESARKVLNVYCKKCGQKTTTLTGVEAVMYMGTLSYDNLKTGVSI PCVCGRDATQYLVQQESSFVMM SAPPAEYK 255

Seqres  LQQGTFLCANEYTSNYQCGHYTHITAKETLYRIDGAHLTKMSEYKGVTDVFEYKETSMTTIT 318
3e9s.1.A LQQGTFLCANEYTSNYQCGHYTHITAKETLYRIDGAHLTKMSEYKGVTDVFEYKETSMTTIT 317

```

441

442 **Fig.7** the alignment of the 315 residues from 1568-1882 aa of the Novel Coronavirus COVI-
 443 19 with the template 3e9s



444

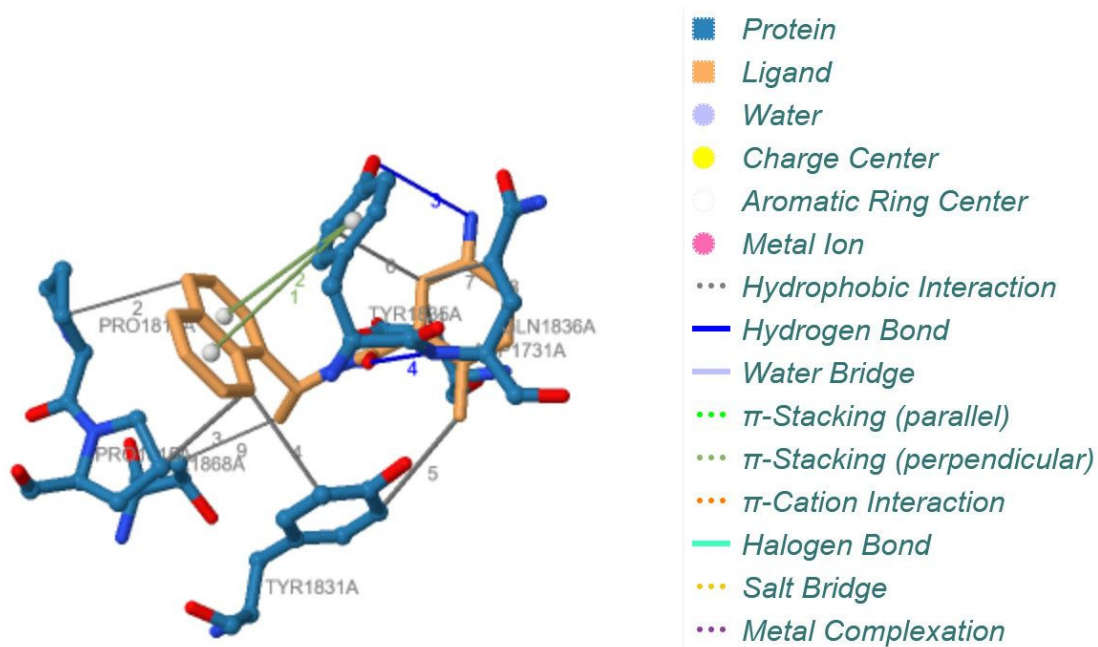


Fig.10 Protein Ligand interaction between the Novel Coronavirus non structural protein and papain-like protease of the model with the template 3e9s

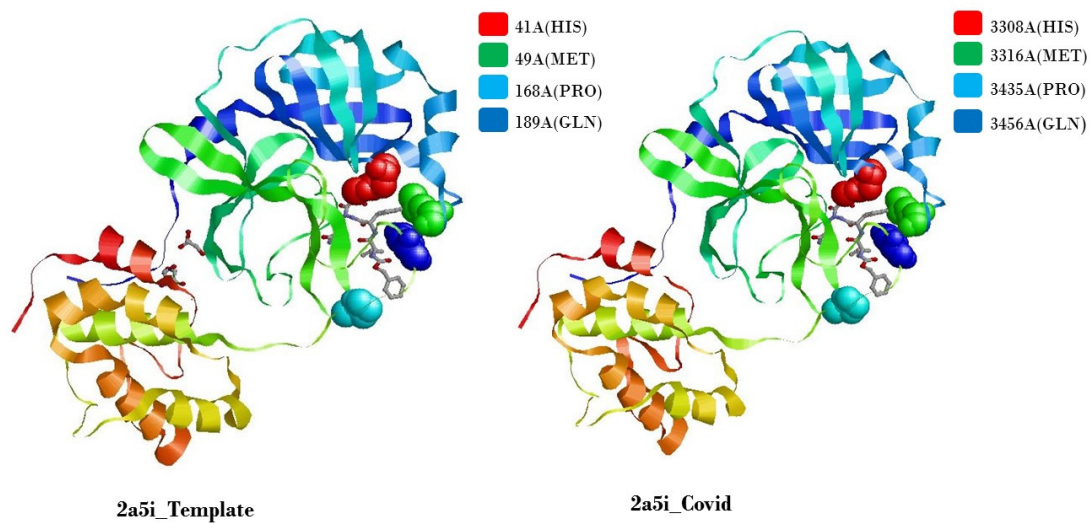
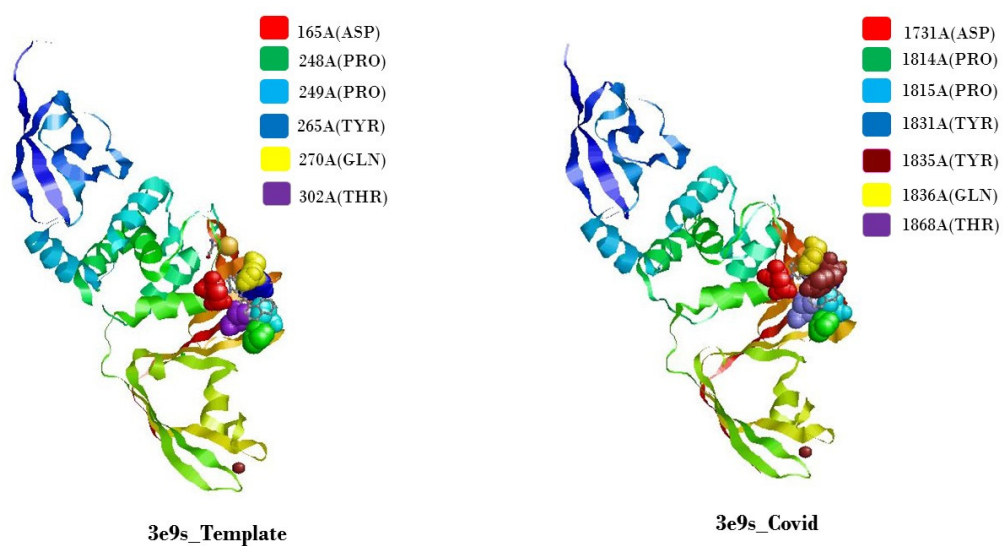


Fig.11 Comparison of the hydrophobic interaction of the binding of the ligand AZP between the SARS-CoV-2 protein and the template 2a5i of SARS CoV

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481

482 **Fig.12** comparison of the hydrophobic interaction of the binding of the ligand AZP between the
483 SARS-CoV-2 protein and the template 3e9s of SARS CoV

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