Whole Genome Sequences Analysis and Homology Modelling of a 3C Like Peptidase and a Non-Structural Protein 3 of the SARS-CoV-2 Shows Protein Ligand Interaction with an Aza-Peptide and a Noncovalent Lead Inhibitor with Possible Antiviral Properties

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Abstract

The family of viruses belonging to Coronaviridae mainly consist of virulent pathogens that have a zoonotic property, Severe Respiratory Syndrome (SARS-CoV) and Middle East Respiratory Syndrome (MERS-CoV) of this family have emerged before and now the SARS-CoV-2 has emerged in China. Characterization of spike glycoproteins, polyproteins and other viral proteins from viruses are important for vaccine development. Homology modelling of these proteins with known templates offers the opportunity to discover ligand binding sites and explore the possible antiviral properties of these protein ligand complexes. Any information emerging from these protein models can be used for vaccine development. In this study we did a complete bioinformatic analysis, sequence alignment, comparison of multiple sequences and homology modelling of the SARS-CoV-2 whole genome sequences, the spike protein and the polyproteins for homology with known proteins, we also analysed receptor binding sites in these models for possible binding with ligands that exhibit antiviral properties. Our results showed that the tertiary structure of the polyprotein isolate SARS-CoV-2_HKU-SZ-001_2020 had 98.94 percent identity with SARS-Coronavirus NSP12 bound to NSP7 and NSP8 co-factors. Our results indicate that a part of the viral genome (residues 3268 -3573 in Frame 2 with 306 amino acids) of the SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3) when modelled with template 2a5i of the PDB database had 96 percent identity with a 3C like peptidase of SARS-CoV which has ability to bind with Aza-Peptide Epoxide (APE) which is known for irreversible inhibition of SARS-CoV main peptidase. The part of the genome (residues 1568-1882 in Frame 2 with 315 amino acids) when modelled with template 3e9s of the PDB database had 82 percent identity with a papain-like protease/deubiquitinase which when complexed with ligand GRL0617 acts as inhibitor which can block SARS-CoV replication. It is possible that these viral inhibiters can be used for vaccine development for the SARS-CoV-2.

Introduction

More than a decade has passed since the emergence human Coronavirus that caused Severe Respiratory Syndrome (SARS-CoV) and it is about 7 years since the emergence of another

type of Coronavirus - Middle East Respiratory Syndrome (MERS-CoV) and now the SARS-CoV-2 has emerged in China. This repeated onslaught of these viruses goes to show that it can assume pandemic proportions at any time and at any place.

The family of viruses belonging to Coronaviridae mainly consist of virulent pathogens that have a zoonotic property and this large family of corona viruses, have been known to be circulating in animals including camels, cats and bats. It has been seen in the past that Severe Acute Respiratory Syndrome associated coronavirus (SARS-CoV) and Middle East Respiratory Syndrome-associated coronavirus (MERS-CoV) belonging to this family of viruses can be transmitted from animals to humans and can cause respiratory diseases. Human to human transmission on this virus has been a concern and due to this search for antiviral compounds and vaccine development for this family of virus becomes the need of the hour.

The SARS was first seen in 2002 in Guangdong province of China, and later spread globally and has caused close to about 8096 cases (WHO 2004, de Vit et al., 2016). In 2012, a novel betacoronavirus, designated Middle East respiratory syndrome coronavirus or MERS-CoV associated with severe respiratory disease in humans, emerged in the Arabian Peninsula (de Wit et al., 2013).

The World Health Organization (WHO), China Country Office was informed of cases of pneumonia of unknown aetiology in Wuhan City, Hubei Province, on 31 December 2019 (WHO 2020). A novel coronavirus currently termed SARS-COV-2 was officially announced as the causative agent by Chinese authorities on 7 January 2020. As on 20 Feb 2020 China's National Health Commission reported that there are 74,280 confirmed cases in China (Fig.1). The World Health Organization reported 924 confirmed cases in 25 countries outside China (WHO Situation Report 29 2020). This novel corona virus has been designated as SARS-CoV-2.

Coronaviruses are RNA viruses and have large genomes structures and due to this they can have high error in replication as compared to host genomes. It is also known that various CoVs can do effective recombination of their genomes after infecting host cells (Luo et al 2018). This recombination can be a factor for their evolution to novel types which may have new animals as their intermediate hosts. These factors give the CoVs high adaptive ability and the capability to jump across species and have a relatively large host range.

Characterization of Spike glycoproteins from viruses are important for vaccine development. Any information coming from the protein model can be used for vaccine development. *In Silico* Epitope, polyprotein and spike protein-based peptide vaccine designing for infectious viruses is a way that can hasten the process of vaccine development. Spike (S) protein, polyprotein and other viral proteins of the SARS-CoV-2 as a target for the development of vaccines and therapeutics for the prevention and treatment of infection is an important approach. In the case of SARS-CoV, these proteins can mediate binding of the virus with its receptor and promotes the fusion between the viral and host cell membranes and virus entry into the host cell, hence peptides, antibodies, organic compounds and short interfering RNAs that interact with the spike protein can have a potential role in vaccine development (Du et al 2009).

There are multiple domain functions that are active in the replication of the coronavirus and these domains are present in a protein designated as Non-structural protein 3 (nsp3) which is the largest protein in the coronavirus genome (Chen et al 2015). 3C like protease (3CLpro) and Papain like Protease (PLpro) are two important class of proteases that are involved in the process of translation of the polypeptide from the genomic RNA to protein components that are required structurally or non-structurally for replication and packaging of new generation viruses (Liu et al 2020)

We hypothesised that there can be some proteins in the large chuck of proteins in the SARS-CoV-2 that could have homology with the Non-structural protein 3 (nsp3) SARS CoV and these proteins can possibly have binding sites with ligands that can bind with known ligand with antiviral properties.

Here in this study we did a complete bioinformatic analysis, sequence alignment, comparison of multiple sequences of the SARS-CoV-2 whole genome sequences, the Spike protein and the polyproteins for homology with known spike proteins and also analysed receptor binding sites for possible vaccine development.

Materials and Methods

Six complete viral genome sequences, seven polyproteins (RdRp region) and seven glycoproteins available on NCBI portal on 4 Feb 2020 were taken for analysis. The sequence details and GenBank accession numbers are listed in Supplementary Table 1. Amongst the seven polyproteins, five are of Wuhan pneumonia virus isolate SARS-COV-2 and two sequences are of Wuhan pneumonia virus isolate SI200040-SP. The seven Glycoproteins are of the same isolate, Wuhan pneumonia virus isolate SARS-COV-2.

The available polyproteins (RdRp region) and glycoprotein sequences were retrieved from Genbank, NCBI (Benson et al., 2000). These sequences were translated to amino acid sequences using sorted six frame translation with Bioedit (Hall et al., 2011). Multiple sequence alignment of the translated protein sequences was performed and phylogenetic tree was constructed using Mega-X (Kumar et al., 2018). The alignment shows that amongst the seven polyproteins, five sequences were identical being from the same isolate and two other sequences of the other isolate are identical. Similar analysis of the seven glycoproteins was done, all the seven glycoprotein sequences were found to be identical. Therefore, further analysis was carried out for three sequences.

1. MN938385.1 SARS-CoV-2 virus isolate SARS-COV-2 _HKU-SZ-001_2020 ORF1ab polyprotein, RdRp region, (orf1ab) gene, partial cds: 0 to 284: Frame 3 95 aa

2. MN970003.1 SARS-CoV-2 virus isolate SI200040-SP orf1ab polyprotein, RdRP region, (orf1ab) gene, partial cds: 2 to 289: Frame 2 96 aa

3. MN938387.1 SARS-CoV-2 virus isolate SARS-COV-2 _HKU-SZ-001_2020 surface glycoprotein (S) gene, partial cds: 1 to 105: Frame 1 35 aa

Expasy proteomics server (Gasteiger et al., 2003) was used to study the protein sequence and structural details. These peptides were studied for their physio-chemical properties using the tool Protparam (Gasteiger et al., 2005). The secondary structure analysis was done using Chou and Fasman algorithm with CFSSP (Kumar, 2013). To generate the 3D structure from the fasta sequence, homology modelling was performed and the templates were identified. The model was built using the template with highest identity. Swiss-model (Schwede et al., 2003) was used to build and validate the 3D model, structural assessment was also performed to validate the model built.

Complete genome sequence of the SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3) which has 29903 bp ss-RNA linear was translated sorted 6 frame with minimum ORF of 20 with any start codon and the resultant protein sequence was used for homology modelling, homology models where done with large chunks of proteins 21503 to 25381 in Frame 2 with 1293 amino acids, 13450 to 21552 in Frame 1 with 2701 amino acids and 254 to 13480 in Frame 2 with 4409 amino acids.

SWISS-MODEL server was used for homology modelling (Waterhouse et al 2018) where computation was on ProMod3 engine which is based on Open Structure (Biasini et al 2013). Structural information is extracted from the template, sequence alignment is used to define insertions and deletions.

Protein ligand interaction profile with hydrogen bonding, hydrophobic interactions, salt bridges and π-Stacking was done with PLIP server (Salentin et al., 2015)

Results and Discussion

The physico- chemical properties and primary structure parameters of the 7 polyproteins RdRp region of the SARS-CoV-2 virus isolate is given in Table 1. RdRP forms an important part of the viral genome where in the RNA viruses its function is to catalyze the synthesis of the RNA strand complementary to a given RNA template.

The isolates SI200040-SP orf1ab polyprotein and the isolate SI200121-SP orf1ab polyprotein had 2 reading frames as compared to the rest of the isolates which had 3 reading frames. The presence of multiple reading frames suggests the possibility of overlapping genes as seen in many virus and prokaryotes and mitochondrial genomes. This could affect how the proteins are made. The number of amino acid residues in all the polyproteins were the same expect one isolate SI200040-SP which had one amino acid more than the other polyproteins. The extinction coefficients of the two isolates SI200040-SP orf1ab polyprotein and the isolate SI200121-SP orf1ab polyprotein was much higher compared to the rest of the polyproteins. The extinction coefficient is important when studying protein-protein and protein-ligand interactions. The instability index of these two isolates was also high when compared to the others indicating the that these two isolates are instable. Regulation of gene expression by polyprotein processing is known in viruses and this is seen in many viruses that are human pathogens (Yost et al 2013).

The isolates here like many other viruses may be using replication strategy which could involve the translation of a large polyprotein with subsequent cleavage by viral proteases. The two isolates SI200040-SP orf1ab polyprotein and the isolate SI200121-SP orf1ab polyprotein also showed shorter half-lives as compared to the other isolates indicating that they are susceptible to enzymatic degradation.

155 The tertiary structure analysis of the isolate SARS-CoV-2 _HKU-SZ-001_2020 ORF1ab
156 polyprotein is given in Table 2. It is seen that the polyprotein has a 98.94 percent identity with
157 PDB structure 6nur.1.A which is a hetero-1-2-1-mer. The polyprotein is an RNA directed RNA
158 polymerase. The protein is identical to the SARS-Coronavirus NSP12 bound to NSP7 and
159 NSP8 co-factors (Kirchdoerfer and Ward 2019). In SARS it is basically a nonstructural protein
160 with NSP12 being the RNA dependent RNA polymerase and the co factors NSP 7 and NSP 8
161 having the function of forming hexadecameric complexes and also act as processivity clamp
162 for RNA polymerase and primase (Fehr et al., 2016). This structure as in SARS CoV here in
163 SARS-CoV-2 may be involved in the machinery of core RNA synthesis and can be a template
164 for exploring antiviral properties.

165 The phylogenetic tree of the seven polyproteins is shown in Fig.2. It is seen that two
166 polyproteins were distinctly different from the rest. The Phylogenetic tree of the seven
167 glycoproteins of the SARS-CoV-2 virus isolate is shown in Fig.3, it is seen that the
168 glycoproteins are similar in all the isolates. Multiple alignment of the Polyproteins of the
169 SARS-CoV-2 is shown in Supplementary Fig.1.

170 This structure as in SARS CoV here in SARS-CoV-2 may be involved in the machinery of core
171 RNA synthesis and can be a template for exploring antiviral properties. Based on its functions
172 in the SARS CoV and its identity to the SARS-CoV-2, it is possible that it has the same
173 functions in SARS-CoV-2 an RNA polymerase which does de novo initiation and primer
174 extension with possible exonuclease activities, the activity itself being primer dependent useful
175 for understanding the mechanism of SARS-CoV-2 replication and can be used as an antiviral
176 target (Te Velthuis et al 2012; Te Velthuis et al 2010; Subissi et al 2014; Subissi et al 2014).

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The polyprotein also has an identity of 19.74 percent with an ABC-type uncharacterized transport system periplasmic component-like protein, this protein is known to be a substrate binding protein and possible binding can be explored here (Bae et al 2019).

The homology model developed from the residues 254 to 13480 in Frame 2 with 4409 amino acids from the Complete genome sequence of the SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3) which has 29903 bp with linear ss-RNA linear showed interesting template alignments, in all the model aligned with 50 templates from the PDB database with most of them being replicase polyprotein 1ab which is a SARS-CoV papain-like protease (Daczkowski 2017). The maximum similarity of 97.3 percent was with template structure of a Nsp9 protein from SARS-coronavirus indicating that this novel coronavirus has high degree of similarity with the SARS-coronavirus and this can be used for gaining insights into vaccine development. Nsp 9 is an RNA binding protein and has an oligosaccharide/oligonucleotide fold-like fold, this protein can have an important function in the replication machinery of the virus and can be important when designing antiviral for this virus (Egloff et al 2004).

Two models were developed, one from residues 3268 -3573 in Frame 2 with 306 amino acids and the other from the part of the genome residues 1568-1882 in Frame 2 with 315 amino acids of the SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3). The models had similarity with the 3C like proteinase and a papain-like protease/deubiquitinase protein which are known antiviral drug targets. Ligand binding with these proteins and their action is on viral replication and inactivation can be useful in stopping the viral replication (Baez-Santos et al 2015).

The homology models of the 4409 amino acid residues of the whole genome of the SARS-CoV-2 virus isolate Wuhan-Hu-1 with the ligand association with templates 2a5i and 3e9s are shown in Fig. 4 and Fig. 5 respectively.

The statistics of structural comparison with PDB templates is given in Table 5, it is seen that the proteins from the SARS-CoV-2 are significantly close to the proteins of SARS CoV and the amino acid alignment in the biding region is the same in both the viruses.

The alignment of the 305 residues from 3268-3573 aa of the Novel Coronavirus COVI-19 with the template 2a5i is shown in Fig.6 and the alignment of the 315 residues from 1568-1882 aa of the Novel Coronavirus COVI-19 with the template 3e9s is shown in Fig.7.

The important templates that aligned with this 4409 amino acid residues of the whole genome of the SARS-CoV-2 virus isolate Wuhan-Hu-1were 2a5i of the PDB database which is a crystal structure of SARS coronavirus main peptidase inhibited by an Aza-Peptide epoxide in the space group C2 (Lee et al 2005) and 3e9s of the PDB database which is new class of papain-like protease/deubiquitinase which when combined with ligand GRL0617 acts as inhibitors blocking SARS virus replication (Ratia et al 2008). The model with template 2a5i of the PDB database shows that Aza-Peptide Epoxide (APE; kinact/Ki=1900(±400) M^{-1} s^{-1}) which is a known anti SARS agent can be used to develop a molecular target with irreversible inhibitor properties. The protein ligand interaction analysis of the Novel Coronavirus C3 like peptidase and aza-peptide epoxide is shown in Fig.8. The substrate binding properties and structural and chemical complementarity of this Aza-Peptide Epoxide can be explored as an anti - Coronavirus SARS-COV-2 agent. The APE which is ethyl (2S)-4-[(3-amino-3-oxo-propyl)-[[(2S)-2-[[(2S)-4-methyl-2-phenylmethoxycarbonylamino-pentanoyl]amino]-3-phenyl-propanoyl]amino]amino]-2-hydroxy-4-oxo-butanoate structure is shown in Fig.9.

The model with template 3e9s of the PDB database shows that the Coronavirus viral protein can have a ligand which is a papain-like protease (PLpro) that is known to be a potent inhibitor of viral replication in SARS (Ratia et al 2008).

The two parts of the Main protein from the whole genome of the SARS-CoV-2 aligned with two SAR proteins and the ligand binding sites were similar, the alignment positions, number of amino acids and ligand and the interacting residues is given in Table 3

The complete genome of MN908947.3 SARS-CoV-2 virus isolate Wuhan-Hu-1 encodes a 4409aa long protein along with the other glycoproteins and polyproteins. The homology modelling of this protein showed sequence and structural alignment with two SARS proteases with structural accession numbers 3e9s.1 and 2a5i.1 at positions 1568-1882 and 3268-3573 respectively. Reports suggests inhibition of virus replication by TTT ligand and an aza-peptide epoxide inhibiting the main peptidase. The structural similarity of these templates are 83% and 96% respectively. The multiple sequence alignment shows complete conservation of the sequence suggesting a high degree of homology. The protein ligand interaction analysis of the Novel Coronavirus non structural protein and papain-like protease is shown in Fig. 10.

The Comparison of Hydrophobic interaction, hydrogen bonding, salt bridges of the constructed model of the Novel Coronavirus protein from region 3268-3573 aa to ligand AZP with Hydrophobic interaction, hydrogen bonding, salt bridges of the template 2a5i is given in Suppl. Table 2, when comparing both it is seen that the binding properties are the same expect for the presence of water bridge in the template 2a5i.

The Comparison of Hydrophobic interaction, hydrogen bonding, π-Stacking of the constructed model of the Novel Coronavirus protein from region 1568-1882 aa to ligand Small molecule Noncovalent Lead Inhibitor with the Hydrophobic interaction, hydrogen bonding, π-Stacking

of the template 3e9s is given in Suppl. Table 3, when comparing both it is seen that the binding properties are the same except or an additional π-Stacking at Tyr in the template 2a5i. This shows that there is high possibility of binding of these antiviral compounds with the regions of Novel Coronavirus protein that is in homology with the SARS protein.

Comparison of the hydrophobic interaction of the biding of the ligand AZP between the SARS-CoV-2 protein and the template 2a5i of SARS CoV is shown in Fig.11 and the comparison of the hydrophobic interaction of the biding of the ligand AZP between the SARS-CoV-2 protein and the template 3e9s of SARS CoV is shown in Fig.12. It is seen that the interaction is the same in both proteins with the same amino acids participating in the interaction indicating that there is a possibility that these ligands with antiviral properties can bind to the new virus.

The similarity in the amin acids involved in the Hydrophobic interactions which are short range interactions and have an important role in the affinities of the ligands and receptors shows that the proteins of the SARS-CoV-2 may bind with the same affinity as seen in the SARS CoV and this also shows a similar action of the ligand as seen in SARS CoV, indicating that these ligands can be used as antivirals in the SARS-CoV-2.

The targeting of this part of the genome of the SARS-CoV-2 with the antiviral compounds which have shown to bind in the similar region of the SARS virus can have implication in the development of an effective antiviral compound against the SARS-CoV-2 . The SARS-CoV-2 shows homology with the SARS coronaviral proteases, papain-like protease (PLpro) and 3C-like protease (3CLpro), these proteins have the function of processing the viral polyprotein and also they perform the function of stripping ubiquitin and the ubiquitin-like interferon (IFN)-stimulated gene 15 (ISG15) from the hosts to facilitate coronavirus replication and help in evading immune response of the host, these inhibitors can also have a role in disrupting signalling cascades in infected cells and protecting the uninfected cells.

The chemical GRL0617 is 5-Amino-2-methyl-N-[(1R)-1-(1-naphthalenyl)ethyl]benzamide and is known to inhibit the papainlike protease that is present in SARS CoV . This protease is a potential target for antiviral compounds (Chaudhuri et al., 2011). We found the SARS-CoV-2 has homology with this and the binding sites for this in the structural protein of the SARS-CoV-2 is the same (Table 4). This compound inhibits the enzyme that is required for the cleavage of the viral protein from the virus in SARS CoV, it also cleaves ubiquitin and has a structural homology with the Deubiquitinases (DUBs) of the Ubiquitin-Specific Proteases Compound GRL0617 binds in the S4 and S3 enzyme subsite that gets the C terminal tail of the Ubiquitin (King and Finley 2014; Schauer et al., 2019). Our results show that Aza-Peptide Epoxide an irreversible protease inhibitor and GRL0617 a viral replication inhibitor can be used to develop inhibitors of the Novel Coronavirus SARS-COV-2.

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Figure Captions

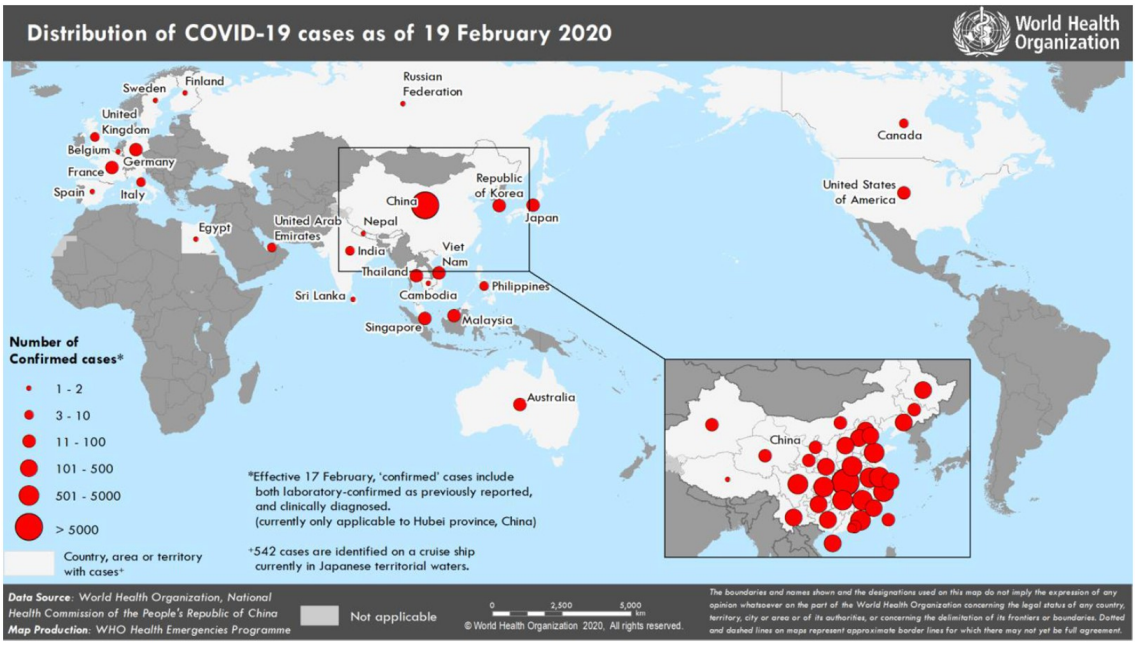


Fig.1 Countries, territories or areas with reported confirmed cases of SARS-COV-2 , 3 February 2020 Source WHO (https://www.who.int/docs/default-ource/coronaviruse/situation-reports/20200219-sitrep-30-covid-19.pdf?sfvrsn=6e50645_2)

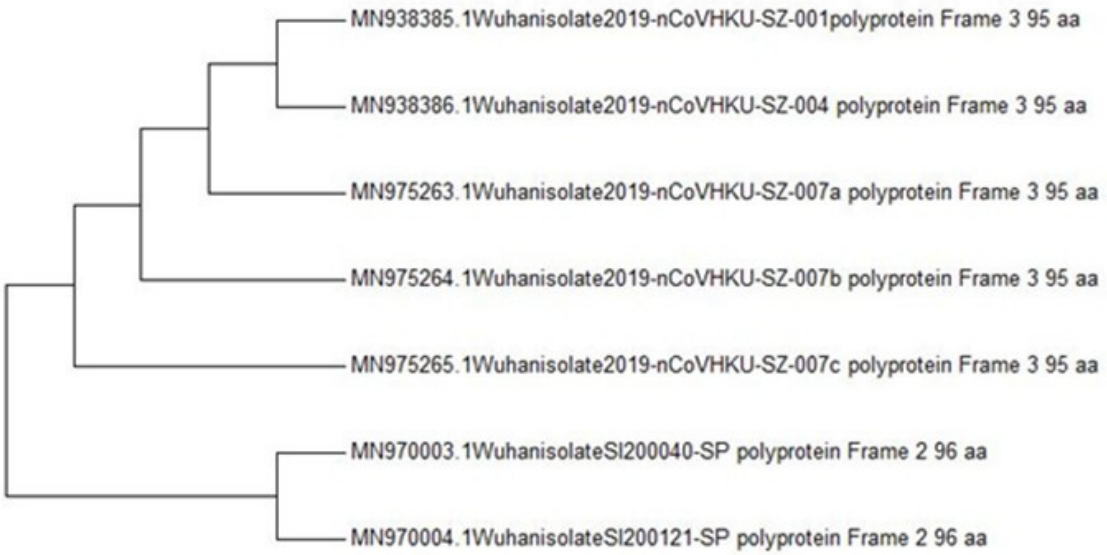


Fig.2 Phylogenetic tree of the seven polyproteins of Severe acute respiratory syndrome coronavirus 2 isolate virus isolates

Fig.3 Phylogenetic tree of the seven glycoproteins of Severe acute respiratory syndrome coronavirus 2 isolate virus isolates

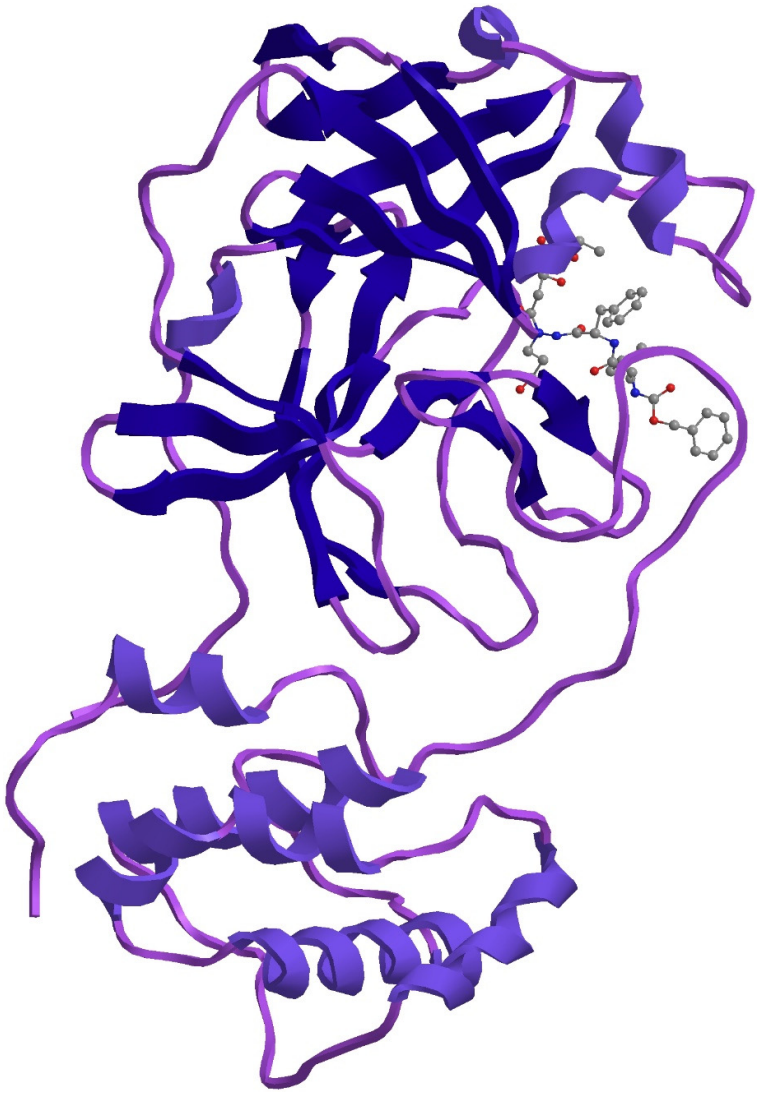


Fig. 4 Homology model with ligand binding of protein from amino acids 3268 -3573 in Frame 2 with 306 amino acids of the Complete genome sequence of the SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3) which has 29903 bp linear ss-RNA with 2a5i of the PDB database as template.

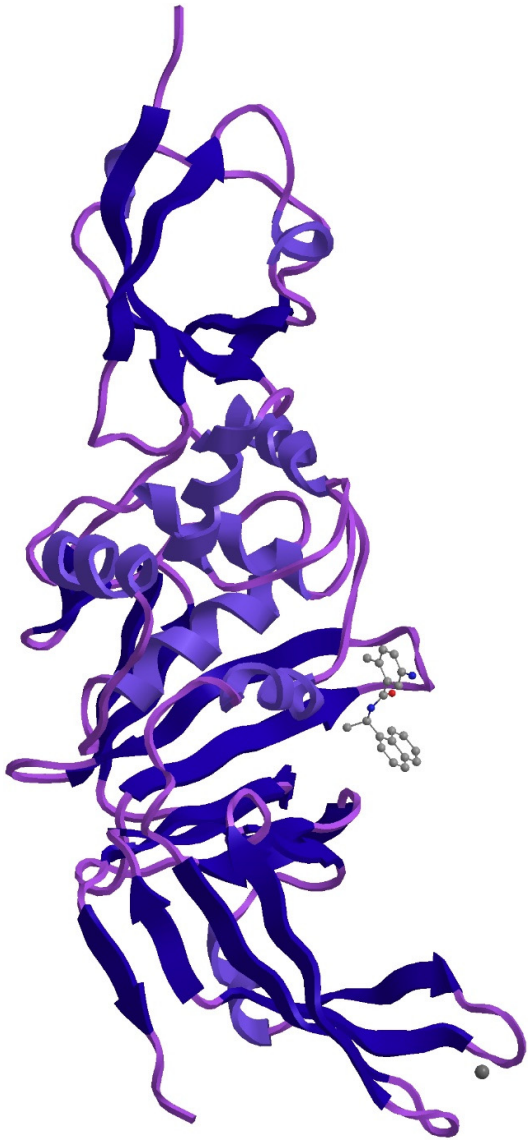
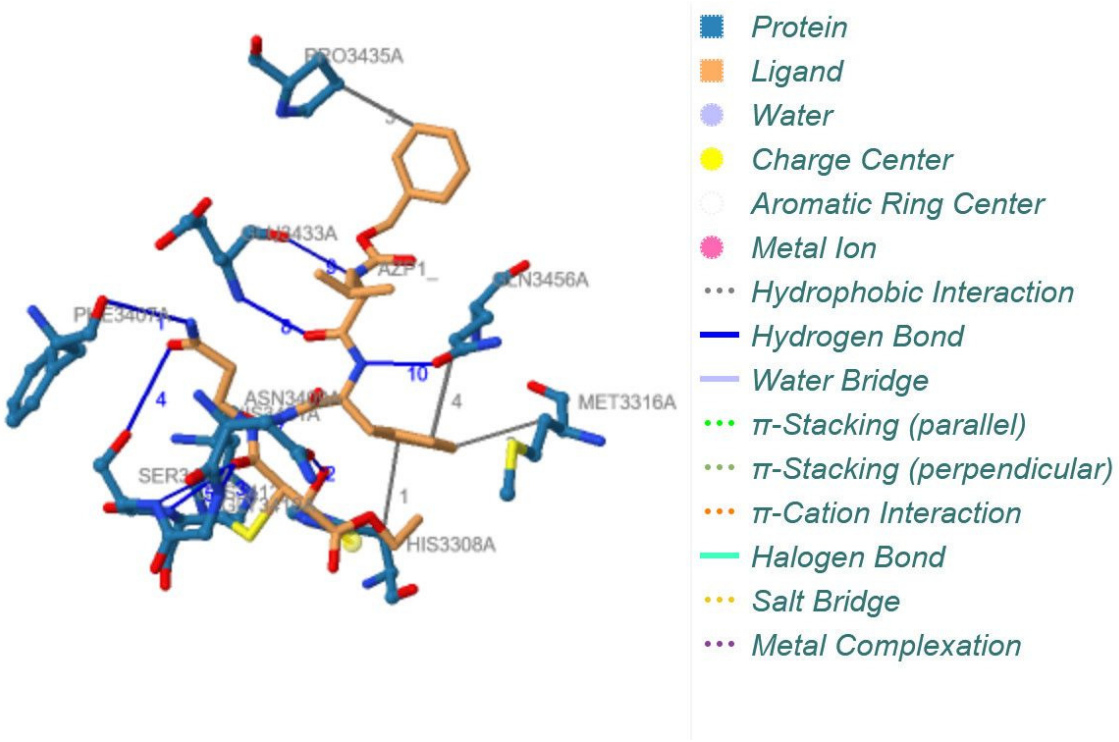


Fig. 5 Homology model with ligand binding of protein from residues 1568-1882 in Frame 2 with 315 amino acids of the Complete genome sequence of the SARS-CoV-2 virus isolate Wuhan-Hu-1 (Genbank Accession Number MN908947.3) which has 29903 bp linear ss-RNA with 3e9s of the PDB database as template.

Fig. 6 Alignment of the 305 residues from 3268-3573 aa of the Novel Coronavirus COVI-19 with the template 2a5i

Fig.7 the alignment of the 315 residues from 1568-1882 aa of the Novel Coronavirus COVI-19 with the template 3e9s



429

430 **Fig.8** Protein Ligand interaction between the C3 like peptidase with aza-peptide epoxide of the

431 model with the template 2a5i

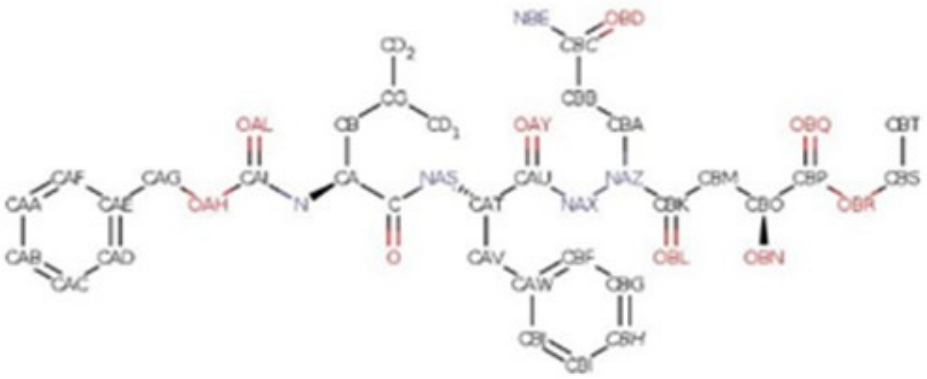


Fig. 9 Structure of Aza-Peptide Epoxide (APE) ethyl (2S)-4-[(3-amino-3-oxo-propyl)-[[(2S)-2-[[(2S)-4-methyl-2-phenylmethoxycarbonylamino-pentanoyl]amino]-3-phenyl-propanoyl]amino]amino]-2-hydroxy-4-oxo-butanoate with possible anti Coronavirus activity – (Source https://www.rcsb.org/ligand/AZP)

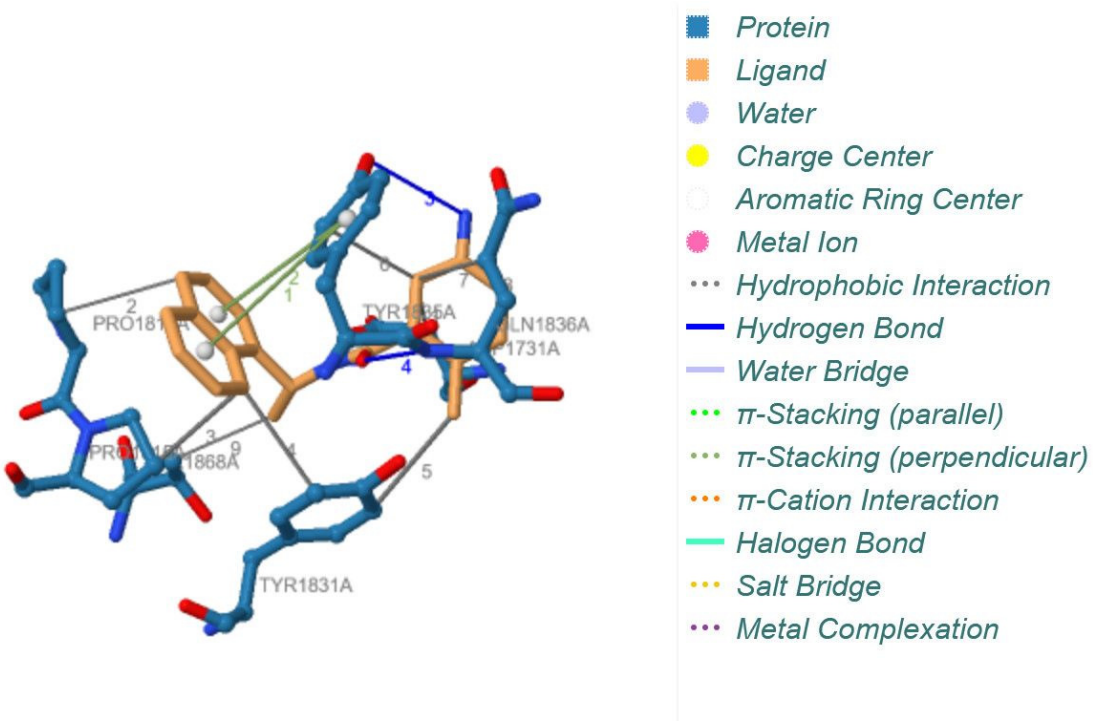


Fig.10 Protein Ligand interaction between the Novel Coronavirus non structural protein and papain-like protease of the model with the template 3e9s

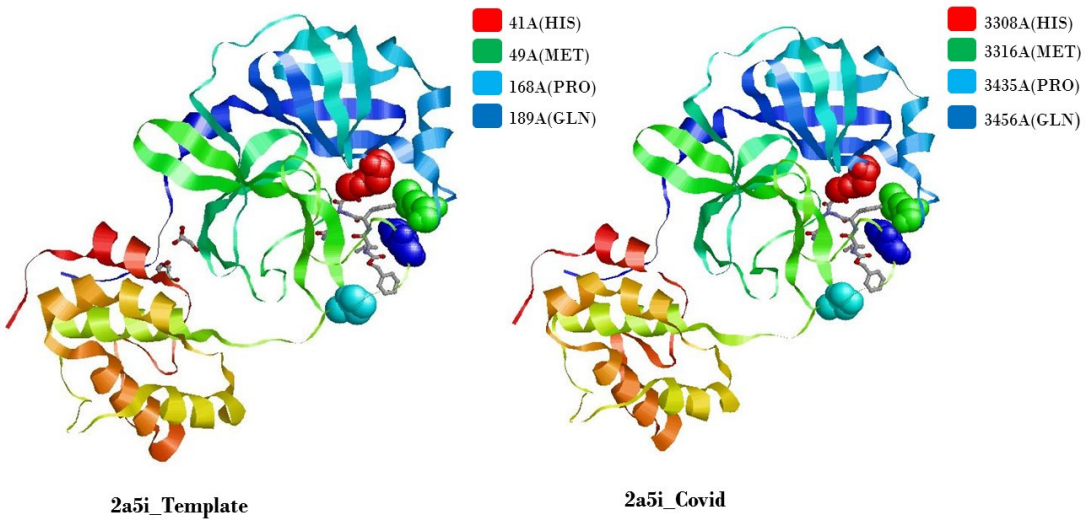


Fig.11 Comparison of the hydrophobic interaction of the biding of the ligand AZP between the SARS-CoV-2 protein and the template 2a5i of SARS CoV

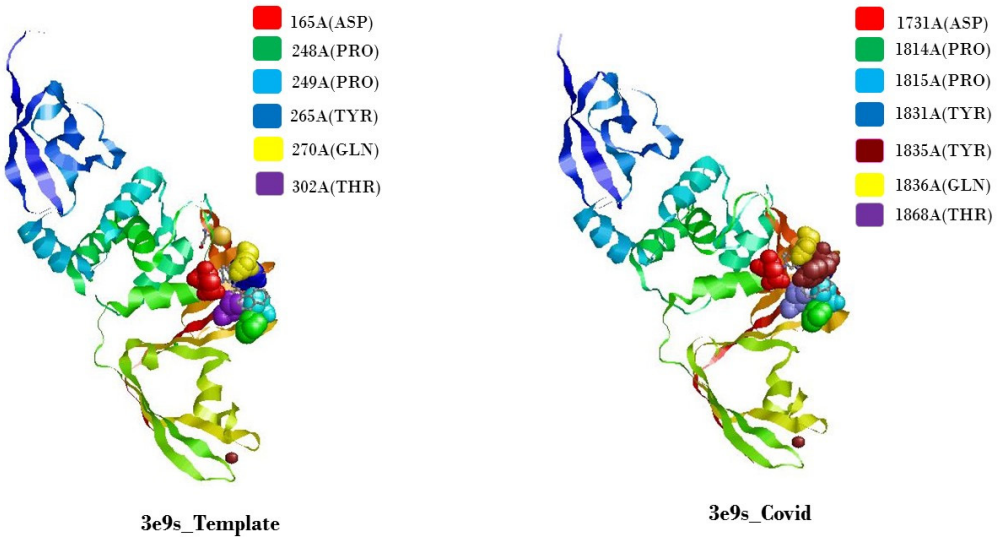
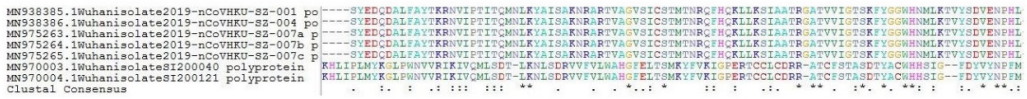


Fig.12 comparison of the hydrophobic interaction of the biding of the ligand AZP between the SARS-CoV-2 protein and the template 3e9s of SARS CoV



Supplementary Fig.1 Multiple alignment of the Polyproteins of Severe acute respiratory syndrome coronavirus 2 isolate virus isolates

Table 1 Physico-chemical properties of polyproteins of SARS-CoV-2 virus isolates

Accession Number	MN938385.1	MN938386.1	MN975263.1	MN975264.1	MN975265.1	MN970003.1	MN970004.1
Reading Frame	3	3	3	3	3	2	2
Number of amino acids	95	95	95	95	95	96	96
Molecular weight	10640.22	10640.22	10640.22	10640.22	10640.22	11239.26	11239.26
Theoretical pI	9.87	9.87	9.87	9.87	9.87	8.9	8.9
Formula	$C_{472}H_{752}N_{134}O_{138}S_4$	$C_{472}H_{752}N_{134}O_{138}S_4$	$C_{472}H_{752}N_{134}O_{138}S_4$	$C_{472}H_{752}N_{134}O_{138}S_4$	$C_{472}H_{752}N_{134}O_{138}S_4$	$C_{516}H_{786}N_{132}O_{132}S_9$	$C_{516}H_{786}N_{132}O_{132}S_9$
Total number of atoms	1500	1500	1500	1500	1500	1575	1575
Extinction coefficients	12950	12950	12950	12950	12950	24200	24200
Instability index	20.51	20.51	20.51	20.51	20.51	29.66	29.66
Aliphatic index	80.11	80.11	80.11	80.11	80.11	89.27	89.27
Grand average of hydropathicity (GRAVY)	-0.264	-0.264	-0.264	-0.264	-0.264	0.161	0.161
Estimated half-life	1.9 hours (mammal	1.9 hours (mammali	1.9 hours (mammali	1.9 hours (mammali	1.9 hours (mammali	1.3 hours (mammali	1.3 hours (mammal

	ian reticulocytes, in vitro).	an reticulocytes, in vitro).	an reticulocytes, in vitro).	an reticulocytes, in vitro).	an reticulocytes, in vitro).	an reticulocytes, in vitro).	ian reticulocytes, in vitro).
	>20 hours (yeast, in vivo).	>20 hours (yeast, in vivo).	>20 hours (yeast, in vivo).	>20 hours (yeast, in vivo).	>20 hours (yeast, in vivo).	3 min (yeast, in vivo).	3 min (yeast, in vivo).
	>10 hours (Escherichia coli, in vivo).	>10 hours (Escherichia coli, in vivo).					

Table 2 Tertiary Structure of SARS-CoV-2 virus isolate SARS-COV-2 _HKU-SZ-001_2020 ORF1ab polyprotein alignment with templates

PDB Template	Gene	Identity
6nur.1.A	NSP12	98.947
1khv.1.A	RNA-directed RNA polymerase	8.97
1khv.2.A	RNA-directed RNA polymerase	8.97
5z6v.1.A	ABC-type uncharacterized transport system periplasmic component-like protein	19.74

6k1y.1.A	ABC-type uncharacterized transport system periplasmic component-like protein	19.74
2ckw.1.A	RNA-directed RNA polymerase	10.53
2uuw.1.A	RNA-directed RNA polymerase	10.67
2wk4.1.A	Protease-polymerase p70	10.67
2wk4.1.B	Protease-polymerase p70	10.67
2yan.1.A	Glutaredoxin-3	12.50
2yan.2.A	Glutaredoxin-3	12.50

Table 3 Main Protein with a sequence length – 4409aa of SARS-CoV-2 Virus showing structural alignment with two other proteins of SARS-CoV

Template ID	Template Title	Alignment Positions	Number of aa	Ligands	Interacting Residues
3e9s.1	A new class of papain-like protease/deubiquitinase inhibitors blocks SARS virus replication	1568-1882	315	TTT	Chain A: L.1729, G.1730, D.1731, P.1814, P.1815, Y.1831, Y.1835, Q.1836, Y.1840, T.1868
2a5i.1	Crystal structures of SARS coronavirus main peptidase inhibited by an aza-peptide epoxide in the space group C2	3268-3573	306	AZP	Chain A: T.3292, T.3293, H.3308, M.3316, Y.3321, F.3407, L.3408, N.3409, G.3410, S.3411, C.3412, H.3430, H.3431, M.3432, E.3433, P.3435, H.3439, D.3454, R.3455, Q.3456, T.3457, A.3458, Q.3459

Table 4 Statistics of structural comparison with PDB templates

Structure	Template	Similarity	p-Value	No. of equivalent positions	RMSD	Raw Score
3e9s_covid	3e9s	Significantly Similar	0.00e+00	314	0.10	935.61
2a5i_covid	2a5i	Significantly Similar	0.00e+00	306	0.08	911.72

Supplementary Table 1 List of Severe Acute Respiratory Syndrome coronavirus 2 isolate sequences taken for bioinformatic analysis

Genbank Accession Number	Title	Description
MN988713.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV/USA-IL1/2020	Complete genome
MN938384.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-002a_2020	Complete genome
MN975262.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-005b_2020	Complete genome
MN985325.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV/USA-WA1/2020	Complete genome
NC_045512.2	Wuhan seafood market pneumonia virus isolate Wuhan-Hu-1	Complete genome
MN908947.3	Severe acute respiratory syndrome coronavirus 2 isolate Wuhan-Hu-1	Complete genome
MN938385.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-001_2020 ORF1ab polyprotein, RdRp region, (orf1ab) gene, partial cds	Polyprotein, RdRp region
MN938386.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-004_2020 ORF1ab polyprotein, RdRp region, (orf1ab) gene, partial cds	Polyprotein, RdRp region
MN975263.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-007a_2020 ORF1ab polyprotein, RdRp region, (orf1ab) gene, partial cds	Polyprotein, RdRp region
MN975264.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-007b_2020 ORF1ab polyprotein, RdRp region, (orf1ab) gene, partial cds	Polyprotein, RdRp region
MN975265.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-007c_2020 ORF1ab polyprotein, RdRp region, (orf1ab) gene, partial cds	Polyprotein, RdRp region
MN970003.1	Severe acute respiratory syndrome coronavirus 2 isolate SI200040-SP orf1ab polyprotein, RdRP region, (orf1ab) gene, partial cds	Polyprotein, RdRp region

MN970004.1	Severe acute respiratory syndrome coronavirus 2 isolate SI200121-SP orf1ab polyprotein, RdRP region, (orf1ab) gene, partial cds	Polyprotein, RdRp region
MN938387.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-001_2020 surface glycoprotein (S) gene, partial cds	Glycoprotein
MN938388.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-002b_2020 surface glycoprotein (S) gene, partial cds	Glycoprotein
MN938389.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-004_2020 surface glycoprotein (S) gene, partial cds	Glycoprotein
MN938390.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-005_2020 surface glycoprotein (S) gene, partial cds	Glycoprotein
MN975266.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-007a_2020 surface glycoprotein (S) gene, partial cds	Glycoprotein
MN975267.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-007b_2020 surface glycoprotein (S) gene, partial cds	Glycoprotein
MN975268.1	Severe acute respiratory syndrome coronavirus 2 isolate 2019-nCoV_HKU-SZ-007c_2020 surface glycoprotein (S) gene, partial cds	Glycoprotein

Supplementary Table 2 Comparison of binding properties of Novel Coronavirus protein from region 3268-3573 (2a5i _SARS-CoV-2) and 2a5i template to ligand AZP

Hydrophobic Interactions

Index	Residue		AA		Distance		Ligand Atom		Protein Atom	
	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i
1	3308A	41A	HIS	HIS	3.64	3.70	2393	2461	307	308
2	3316A	49A	MET	MET	3.81	3.86	2395	2463	368	368
3	3435A	168A	PRO	PRO	3.42	3.73	2376	2443	1303	1347
4	3456A	189A	GLN	GLN	3.84	3.93	2396	2464	1462	1507

Hydrogen Bonds

Index	Residue		AA		Distance H-A		Distance D-A		Donor Angle		Protein donor?		Sidechain		Donor Atom		Acceptor Atom	
	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i
1	3407A	140A	PHE	PHE	2.61	2.46	3.47	3.33	146.60	147.28	✖	✖	✖	✖	2404 [Nam]	2472 [Nam]	1081 [O2]	1112 [O2]
2	3409A	142A	ASN	ASN	2.51	2.52	2.87	2.87	102.18	100.93	✖	✖	✔	✔	2410 [O3]	2478 [O3]	1103 [O2]	1134 [O2]

3	3410A	143A	GLY	GLY	1.94	1.83	2.78	2.73	142.36	150.57	✓	✓	✗	✗	1105 [Nam]	1136 [Nam]	2407 [O2]	2475 [O2]
4	3411A	144A	SER	SER	3.37	3.44	3.76	3.80	106.27	104.79	✓	✓	✓	✓	1114 [O3]	1145 [O3]	2405 [O2]	2473 [O2]
5	3411A	144A	SER	SER	2.60	2.61	3.24	3.20	122.58	118.23	✓	✓	✗	✗	1109 [Nam]	1140 [Nam]	2407 [O2]	2475 [O2]
6	3412A	145A	CYS	CYS	2.50	2.57	3.39	3.37	150.10	137.96	✓	✓	✗	✗	1115 [Nam]	1146 [Nam]	2407 [O2]	2475 [O2]
7	3431A	164A	HIS	HIS	1.70	1.73	2.63	2.67	156.72	157.31	✗	✗	✗	✗	2399 [Nam]	2467 [Nam]	1266 [O2]	1307 [O2]
8	3433A	166A	GLU	GLU	2.08	2.01	3.04	2.97	165.52	163.22	✓	✓	✗	✗	1281 [Nam]	1325 [Nam]	2387 [O2]	2455 [O2]
9	3433A	166A	GLU	GLU	2.06	2.13	2.88	2.92	140.23	135.67	✗	✗	✗	✗	2370 [Nam]	2438 [Nam]	1284 [O2]	1328 [O2]
10	3456A	189A	GLN	GLN	1.90	1.82	2.84	2.77	158.82	161.93	✗	✗	✓	✓	2388 [Nam]	2456 [Nam]	1464 [O2]	1509 [O2]

Salt Bridges

Index	Residue		AA		Distance		Protein positive?		Ligand Group		Ligand Atoms	
	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i
1	3308A	41A	HIS	HIS	5.10	5.08	✔	✔	Carboxylate	Carboxylate	2412, 2413	2481, 2480

Water Bridges

Index	Residue		AA		Dist. A-W		Dist. D-W		Donor Angle		Water Angle		Protein donor?		Donor Atom		Acceptor Atom		Water Atom	
	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i	2a5i _SARS-CoV-2	2a5i
1	189A	-	GLN	-	4.07	-	3.93	-	115.60	-	91.13	-	✔	-	1510 [Nam]	-	2456 [Nam]	-	2543	-

Supplementary Table 3 Comparison of binding properties of Novel Coronavirus protein from region 1568-1882 (3e9s_SARS-CoV-2) and 3e9s template to ligand Small molecule Noncovalent Lead Inhibitor

Hydrophobic Interactions

Index	Residue		AA		Distance		Ligand Atom		Protein Atom	
	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s
1	1731A	165A	ASP	ASP	3.82	3.83	2502	2504	1308	1320
2	1814A	248A	PRO	PRO	3.79	3.80	2501	2503	1955	1952
3	1815A	249A	PRO	PRO	3.52	3.75	2503	2505	1963	1960
4	1831A	265A	TYR	TYR	3.50	3.57	2503	2505	2090	2089
5	1831A	265A	TYR	TYR	3.63	3.67	2504	2506	2091	2090
6	1835A	-	TYR	-	3.62	-	2502	-	2121	-
7	1836A	270A	GLN	GLN	3.60	3.59	2502	2504	2130	2129
8	1836A	270A	GLN	GLN	3.58	3.62	2507	2509	2130	2129
9	1868A	302A	THR	THR	3.37	3.54	2514	2516	2381	2385

Hydrogen Bonds

Index	Residue	AA	Distance H-A	Distance D-A	Donor Angle	Protein donor?	Sidechain	Donor Atom	Acceptor Atom

	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s
1	1731A	165A	ASP	ASP	2.12	2.19	2.98	3.03	151.51	149.71	✔	✔	✔	✔	1311 [O3]	1323 [O3]	2512 [Nam]	2514 [Nam]
2	1731A	165A	ASP	ASP	2.00	2.05	2.98	3.03	173.11	175.80	✖	✖	✔	✔	2512 [Nam]	2514 [Nam]	1311 [O3]	1323 [O3]
3	1835A	269A	TYR	TYR	2.78	2.81	3.61	3.64	143.08	142.33	✖	✖	✔	✔	2511 [Npl]	2513 [Npl]	2124 [O3]	2123 [O3]
4	1836A	270A	GLN	GLN	1.87	1.77	2.83	2.75	164.21	174.65	✔	✔	✖	✖	2125 [Nam]	2124 [Nam]	2509 [O2]	2511 [O2]

π-Stacking

Index	Residue		AA		Distance		Angle		Offset		Type		Ligand Atoms	
	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s	3e9s_SARS-CoV-2	3e9s
1	1835A	269A	TYR	TYR	5.30	5.31	84.17	85.46	1.70	1.70	T	T	2497, 2500, 2503, 2506, 2508, 2516	2499, 2502, 2505, 2508, 2510, 2518

2	1835A	269A	TYR	TYR	5.09	5.10	84.10	85.40	0.83	0.85	T	T	2497, 2498, 2500, 2501, 2515, 2517	2499, 2500, 2502, 2503, 2517, 2519
3	-	269A	-	TYR	-	5.18	-	73.97	-	1.96	-	T	-	2498, 2501, 2504, 2507, 2509, 2512