1	Antiretroviral drug activity and potential for pre-exposure prophylaxis against COVID-
2	19 and HIV infection
3	
4	Dennis C. Copertino Jr., Bruno C. Casado Lima, Rodrigo R. R. Duarte, Timothy Wilkin, Roy
5	M. Gulick, Miguel de Mulder Rougvie, and Douglas F. Nixon*
6	
7	Division of Infectious Diseases, Weill Cornell Medicine, Cornell University, New York, NY,
8	USA.
9	
10	*Correspondence: Dr. Douglas F. Nixon, Division of Infectious Diseases, Weill Cornell
11	Medicine, Cornell University, Belfer Research Building, Room 530, 413 E. 69th St., New York,
12	NY, 10021, USA. E-mail: dnixon@med.cornell.edu

1

# 2 Abstract

3

COVID-19 is the disease caused by SARS-CoV-2, and has led to over 250,000 deaths by May 4 5 2020. Urgent studies to identify new antiviral drugs, repurpose existing drugs, or identify those 6 drugs that can specifically target the overactive immune response are ongoing around the 7 world. Antiretroviral drugs (ARVs) have been tested in past human coronavirus infections, and 8 also against SARS-CoV-2, but a recent clinical trial of lopinavir and ritonavir failed to show 9 any clinical benefit in COVID-19 disease. However, anecdotal reports suggest either reduced infection or a course of milder COVID-19 disease in people living with HIV (PLWH) on ARVs. 10 We hypothesized ARVs other than lopinavir and ritonavir might be responsible for such effects. 11 12 Here, we used chemoinformatic analyses to predict which ARVs would bind and potentially 13 inhibit the SARS-CoV-2 main protease or RNA-dependent RNA polymerase enzymes, and identified a number of ARVs which bind to SARS-CoV-2 enzymes in silico. Our study identified 14 HIV nucleoside/nucleotide analogue reverse transcriptase inhibitors (abacavir, emtricitabine, 15 16 lamivudine, tenofovir, zidovudine), HIV protease inhibitors (ASC09, atazanavir, indinavir, 17 lopinavir, nelfinavir, ritonavir, saquinavir, tipranavir) and an HIV pharmacokinetic booster (cobicistat), as drug candidates with effective in silico binding to one or both viral enzymes. 18 19 Tenofovir and emtricitabine are FDA-approved as HIV pre-exposure prophylaxis (PrEP) and have an extensive safety profile of use in populations without HIV. Existing or new 20 combinations of antiretroviral drugs could potentially prevent or ameliorate the course of 21 COVID-19, if shown to inhibit SARS-CoV-2 in vitro and/or in clinical trials. Further studies are 22 needed to establish the activity of ARVs for treatment or prevention of SARS-CoV-2 infection. 23

#### 1 Introduction

2

3 The novel coronavirus, now named SARS-CoV-2, was first reported as a new viral infection in humans in late 2019 and over the course of the past few months the virus has 4 5 become a pandemic. Coronavirus disease, COVID-19, has in turn taken the lives of more than 250,000 people by May 2020, with hundreds of thousands of more deaths expected in the 6 7 absence of effective treatments or additional control measures. COVID-19 presents a major 8 worldwide public health emergency. Intense research efforts are underway to find effective 9 antiviral treatments via novel drug design or drug repurposing (Sanders, Monogue, Jodlowski, 10 & Cutrell, 2020) (Duarte et al., 2020) (Elmezayen, Al-Obaidi, Sahin, & Yelekci, 2020)

Some antiretrovirals (ARVs), normally used to treat HIV infection, have also been used 11 in Hepatitis B virus (HBV) infection (Boettiger et al., 2016), and Amyotrophic Lateral Sclerosis 12 13 (ALS) [NCT02437110]. ARVs have been studied both in vitro and in vivo for their activity against Severe Acute Respiratory Syndrome (SARS) (Chu et al., 2004; Yamamoto et al., 14 2004), and Middle East Respiratory Syndrome (MERS) (de Wilde et al., 2014), which led to 15 adoption of two ARVs, lopinavir and ritonavir, as putative antiviral drugs against SARS-CoV-16 17 2. However, a small randomized controlled clinical trial of lopinavir and ritonavir coadministered to hospitalized adults with severe COVID-19 showed no clinical benefit over the 18 19 standard of care (Cao et al., 2020). A recent preprint reported that tenofovir and emtricitabine act as chain terminators in the replication of viral RNA by the SARS-CoV-2 RdRp (Jockusch 20 et al., 2020). In silico studies help us to gauge projected mechanisms and likely molecules 21 which bind to solved targets, in a cost effective, high throughput manner, ideal for primary 22 analyses. In vitro studies provide necessary details for effective inhibitory concentrations of 23 each compound tested, and clinical studies would provide further evidence for effectiveness 24 25 in people.

We used a comprehensive in silico docking analysis of ARVs to catalytically active sites within the Main protease (Mpro) and RNA dependent RNA polymerase (RdRp) of SARS-CoV-2, in order to understand which HIV ARVs might bind and potentially inhibit SARS-CoV-

1 2, and at which specific part of the viral replication cycle. We hypothesized that ARVs would show activity against SARS-CoV-2, and used a small molecule docking algorithm with an 2 empirical scoring function, PLANTSchemplp (Korb, Stutzle, & Exner, 2009) to identify a number 3 of ARVs which we believe should be studied further, either individually or in combination, for 4 5 anti-SARS-CoV-2 activity. Drugs identified to bind the catalytic site of the SARS-CoV-2 Mpro were the HIV protease inhibitors ASC09, atazanavir, indinavir, lopinavir, nelfinavir, ritonavir, 6 7 saquinavir, tipranavir and the HIV pharmacokinetic (PK) booster cobicistat. The drugs shown 8 to bind to the catalytic site of the RdRp include, nucleoside or nucleotide reverse transcriptase 9 inhibitors abacavir, emtricitabine, lamivudine, tenofovir, zidovudine, the protease inhibitors 10 ASC09, atazanavir, indinavir, lopinavir, ritonavir and saquinavir, and a PK booster, cobicistat. In order to better understand the impact of ARVs on COVID-19 infection or disease 11 course, people living with HIV (PLWH) or people at risk of HIV infection on pre-exposure 12 13 prophylaxis (PrEP) could provide critical insights into the course of COVID-19 disease. Anecdotal reports show that PLWH on ARVs can be COVID-19 infected (Blanco et al., 2020), 14 but of the first 543 people admitted to a hospital in Barcelona with the SARS-CoV-2, only five 15 were PLWH. Additional in vitro studies and clinical trials are needed to understand whether 16 17 existing ARV regimens might protect against SARS-CoV-2 infection or ameliorate COVID-19 in PLWH. We attempted to estimate the number of PLWH at risk of death from COVID-19 by 18 August 4th 2020 in the USA and in each state. Therein, we sought to estimate a benchmark 19 for which observational studies could use to measure actual deaths of PLWH by COVID-19, 20 and compare it to the expected number of deaths noted in Supplemental Table 1. 21

As COVID-19 infection is such a rapidly changing problem, prospective placebocontrolled studies in appropriate groups would be preferable to observational studies. However, in silico and in vitro work can often save time and effort in screening effective drugs, in a high throughput manner. Therefore, we sought to use in silico methods to determine whether existing ARVs might bind to key SARS-CoV-2 viral enzymes and inhibit viral replication in doing so. Our results suggest that some commonly used ARVs have binding potential in silico to SARS-CoV-2 including ARVs commonly given to people without HIV as

PrEP or treatment of chronic HBV. Further studies should explore activity of ARVs in vitro, in
animal models, and in clinical trials, to assess whether the ARVs we identified could be used
either in COVID-19 treatment regimens or as pre-exposure prophylaxis for SARS-CoV-2
infection.

5

## 6 Methods

## 7 Protein-Ligand Small Molecule Docking with PLANTS

8 Two of the optimal nonstructural protein targets within the SARS-CoV-2 virus are the Main 9 protease (Mpro) and the RNA dependent RNA polymerase (RdRp). Small molecules which might inhibit these two proteins would bind to key residues typically required for the enzymatic 10 activity of these proteins. For small molecules to effectively inhibit the activity of the Mpro it 11 should disrupt the substrate binding site, marked by residues GIn189, His41 or Cys145 (Zhang 12 13 et al., 2020). For the RdRp, the substrate binding site is marked by conserved residues amongst polymerases 759-SDD-761, within Motif Cs residues, 753-FSMMILSDDAVVCFN-14 15 767 (Gao et al., 2020). In order to identify compounds which are predicted to effectively and specifically bind to these catalytic sites of the Mpro, and RdRp, we performed in silico 16 17 molecular docking analyses using the PLANTS software against a database of FDA-approved 18 small molecules (Korb et al., 2009) (Douguet, 2018). From the database of small molecules, we manually extracted all HIV antiretrovirals for discussion. Docking simulations using the 19 20 main protease (Mpro) (Protein Data Bank [PDB] ID: 6Y2E), and the RNA-dependent RNA 21 polymerase (RdRp) (PDB ID: 6M71) of SARS-Cov-2, using the Protein-Ligand ANT System 22 (PLANTS) (Korb et al., 2009), accessed via https://chemoinfo.ipmc.cnrs.fr/ (Douguet, 2010). The ligand docking sites were specified as the catalytically active sites by Zhang et al. (Zhang 23 24 et al., 2020) and Gao et al. (Gao et al., 2020), using a radius of +10 Å around the assigned 25 residues, while the weight in final score for the protein structures was 1, and all remaining settings were set to the default setting. Protons are added to the structure using the PDB2PQR 26 with an AMBER forcefield option (Dolinsky, Nielsen, McCammon, & Baker, 2004). The 27

1 resulting protein-ligand scores (PLANTS scores) were calculated using the empirical scoring 2 algorithm, CHEMPLP (Douguet, Munier-Lehmann, Labesse, & Pochet, 2005), and reflect the 3 energy change when ligands and proteins come together, with values more negative than (-91.00) suggesting likely protein-ligand interactions. All other settings were set to their default 4 5 parameters. Protein-ligand structures were visualized using PyMol 2.3.5. For the entirety of 6 this paper we will use the terminology ARVs to describe all of the tested drugs, including the 7 PK booster cobicistat, although cobicistat does not have anti-HIV activity perse, and therefore should not truly be considered an HIV ARV. Structures from the FDA approved library did not 8 9 include the metabolically active structures for some of the HIV nucleoside (or nucleotide) 10 analogue reverse transcriptase inhibitors (NRTIs), nor any unapproved drugs such as ASC09. 11 The biologically active structures of these drugs were obtained from PubChem (Kim et al., 12 2019) or other sources noted in **Supplementary Table 2**. The active triphosphorylated NRTIs 13 were then run against each of the viral enzymes individually using the same software and 14 settings stated above. The image portrayed in **Figure 1** was created using Biorender.

15

## 16 Results

#### 17 Molecular docking analyses

We ran analyses to predict the ability of HIV ARV drugs to bind to the catalytic sites of the 18 SARS-CoV-2 main protease (Mpro) and RNA-dependent RNA polymerase (RdRp) enzymes, 19 20 corroborating their repurposing potential, and providing a possible mechanism of action 21 involving direct interactions with viral components (Figure 1). The PLANTSchempton scores 22 calculated here reflect the energy change when the drug comes together with the catalytic site 23 of either the Mpro or RdRp with more negative numbers suggesting a more likely drug-protein 24 interaction (Table 1). HIV ARVs with scores lower than -91 with predicted binding to the Mpro 25 include PIs, ASC09, atazanavir, indinavir, lopinavir, nelfinavir, ritonavir, saquinavir and tipranavir; and the PK booster, cobicistat. HIV ARVs with scores lower than -91 with predicted 26 binding to the RdRp include NRTIs, abacavir, emtricitabine, lamivudine, tenofovir, and 27

zidovudine; PIs, ASC09, atazanavir, indinavir, lopinavir, ritonavir, and saquinavir; and the PK
booster cobicistat. Figure 2 depicts the Mpro and RdRp designated active sites for the
purpose of this analysis. Figure 3 depicts select molecules docking with the Mpro. Figure 4
depicts select molecules docking with the RdRp.

5

# 6 Estimates of COVID-19 and HIV infection

7 There are several important issues that complicate the issue of COVID-19 and HIV. PLWH of color are disproportionally impacted by COVID-19. Half of the PLWH in the USA are over 8 9 50 years of age, and co-morbidities are more common in this group. We estimated the 10 number of PLWH at risk for death from COVID-19 in the USA by August 4th 2020, based upon projected estimates (**Supplemental Table 1**). These estimates are confounded by 11 12 many variables, and since the mortality rates are fluctuating with changing testing 13 algorithms, and data availability, they are limited in utility, yet in order to make sense of data in any observational study, such numbers need to be estimated. An observational study to 14 determine the incidence or disease course from SARS-CoV-2 infection in PLWH who take 15 ARV therapy, compared to those that do not is technically challenging, and problematic 16 17 because ARV therapy is considered the standard of care for HIV, worldwide. A prospective, randomized study of anti-HIV PrEP medications to prevent COVID-19, would be more 18 19 appropriate. In addition, linking COVID-19-related deaths to HIV registries could provide some of these insights as well. 20

21

### 22 Discussion

23

The data presented here, along with anecdotal evidence and recent in vitro studies, present a case for the possible prevention and treatment of COVID-19 with repurposed HIV Antiretroviral drugs (ARVs). ARVs were studied in past human coronavirus infections of SARS-CoV and MERS (Ford et al., 2020), and are being used in SARS-CoV-2 clinical trials 1 as listed in Table 2. Anecdotal initial reports suggest less infection and a milder course of 2 COVID-19 in people living with HIV (PLWH). However, a recent small clinical trial of lopinavir 3 and ritonavir failed to show any clinical benefit in people with severe disease (Cao et al., 2020). Here, we used chemoinformatic analyses to predict which ARVs would bind to the SARS-4 5 CoV-2 Mpro or RdRp enzymes, and identified a number which bound to either or both. Existing 6 or new combinations of ARV regimens for preventing or treating HIV infection could potentially 7 prevent or ameliorate the course of COVID-19. Further studies are needed to establish the value of ARVs in COVID-19. 8

9 Previous published works discuss the similarities between RdRp's from previous 10 coronaviruses, and HIV reverse transcriptase (RT) (Oberg, 2006). More recent publications compared the SARS-CoV viral enzymes with enzymes from SARS-CoV-2. The identity 11 12 between the SARS-CoV and SARS-CoV-2 Mpro is 96% (Chen, Yiu, & Wong, 2020), and 13 identity between SARS-CoV and the SARS-CoV-2 RdRp 96%, with their similarity being 98% (Shannon et al., 2020). The similarity between the enzymes of SARS-CoV and SARS-CoV-2 14 is marked, and provides rationale for testing drugs which were effective in vitro against SARS-15 CoV, like lopinavir and ritonavir (Chu et al., 2004), in the setting of SARS-CoV-2, as 16 17 experimental therapies.

In our study, patterns of HIV inhibitor type seem to correspond closely with each viral 18 enzyme. For example, the ARVs predicted to bind to the Mpro were PIs and cobicistat. ARVs 19 with predicted binding to the RdRp were in the nucleoside and nucleotide reverse transcriptase 20 inhibitor (NRTI), protease inhibitor (PI) families, and the PK booster cobicistat. Cobicistat is a 21 CYP3A inhibitor (L. Xu et al., 2010) used in the treatment of HIV, and is an analog of ritonavir 22 which has proven dual mechanisms as a PI and CYP3A inhibitor, and both share a near exact 23 structure with the exception of 5 atoms (Kumar et al., 1999). Cobicistat's ability to inhibit 24 25 CYP3A gives it the colloquial name of "booster". Mechanistically, cobicistat inhibits CYP3A 26 activity in the liver which is typically used to metabolize certain drugs including most PIs. Thus, for it to have a boosting effect on a drug, the active form of the drug/molecule must be mostly 27 28 or partly broken down by the CYP3A enzyme in the liver. Cobicistat's in silico binding to the 1 Mpro and RdRp follows closely with other PIs, of which it is an analog. Therefore, it may have anti-COVID-19 activity itself, despite no reported protease activity to date. Due to cobicistat's 2 3 effective in silico docking to both the SARS-Cov-2 Mpro and RdRp it should be tested in vitro and if effective, potentially in clinical trials. However, widespread use of protease inhibitors 4 5 would be challenging to implement because of adverse drug-drug interactions and tolerability issues. We (in this study), and others, have shown that PIs bind to the RdRp of SARS-Cov-2 6 7 in silico (Beck, Shin, Choi, Park, & Kang, 2020), but we do not yet understand why this binding 8 occurs. Further in vitro work will be key to elucidating the ability of PIs to bind both the RdRp 9 and Mpro of SARS-CoV-2.

10 Remdesivir, an investigational nucleotide/adenosine analogue currently in clinical studies for COVID-19 has been shown to have potential activity against RdRp of SARS-CoV-11 12 2 (Gordon et al., 2020). Remdesivir is an adenosine analog, and acts on the RdRp by being 13 incorporated into the growing RNA strand and inhibiting further viral transcription. Remdesivir is the first agent to show preliminary clinical efficacy in an NIH randomized, placebo-controlled 14 study (Wang et al., 2020) and the FDA has approved its Emergency Use Authorization for 15 hospitalized patients with severe COVID-19. Tenofovir and emtricitabine can inhibit the RdRp 16 17 of SARS-CoV-2 in vitro (Jockusch et al., 2020), and expected from our in silico analyses. Taken together these studies confirm that both nucleotide analogs and nucleoside analogs 18 19 can be incorporated into growing RNA strands and thus may act as chain terminators of growing viral RNA strands under replication by the RdRp, likely due to the low fidelity of the 20 RdRp to its substrate (Jockusch et al., 2020; McKenna, Kashemirov, Peterson, & Goodman, 21 2010). The remaining NRTIs, if effective against COVID-19, could confer protection through 22 the same mechanism. This suggests at least that PLWH on a backbone regimen of tenofovir 23 and emtricitabine could be partially protected from SARS-CoV-2 infection or COVID-19 24 25 disease, although only additional in vitro studies and clinical trials could confirm this. Our work would also suggest that patients taking abacavir and lamivudine could be protected from 26 COVID-19 by inhibition of the RdRp, but again, in vitro work and clinical trials are needed to 27 28 confirm this. Integrase inhibitors in our study were scored in the -79 to -89 PLANTSchempip

range. These scores are harder to interpret, but at a minimum do warrant further study in vitroand in clinical trials.

Recently the first ever two-drug HIV regimens have been approved for PLWH, including Dovato® (ViiV) dolutegravir and lamivudine, an integrase inhibitor and an NRTI, respectively. The other approved two drug regimen, Juluca® (ViiV), includes dolutegravir and rilpivrine. Observational studies of COVID-19 disease course of different drug combinations are critical for those living with HIV worried about contracting SARS-CoV-2, or acquiring the virus and navigating COVID-19 disease. These questions can only be answered by further in vitro research, observational studies, and clinical trials.

10 It is to be expected that NNRTIs as a class did not bind well to the catalytic site for the SARS-CoV-2 RdRp, which is the nucleoside/nucleotide triphosphate (NTP) binding site, 11 12 where elongation of the RNA strand occurs (Gao et al., 2020). When NRTIs are metabolized 13 and form triphosphate structures, they are capable of being added to the growing RNA/DNA strand and chain termination will follow (Gordon et al., 2020; Jockusch et al., 2020; Wang et 14 al., 2020). However, HIV reverse transcriptase is inhibited by NNRTIs at a site different from 15 16 that which the NRTIs bind to. Instead, NNRTIs inhibit HIV RT in a non-competitive fashion, 17 binding at a site distant from the polymerase active site, usually stopping key protein-protein interactions from occurring, or changing the active site structure (Sluis-Cremer & Tachedjian, 18 19 2008). Past studies have found no potential homologous hydrophobic NNRTI binding site on the previous SARS-CoV RdRp structure (X. Xu et al., 2003), although we identified potential 20 hydrophobic pockets on the surface of the RdRp (data not shown). This will remain an area 21 for further research to address, though COVID-19 research should certainly include second 22 generation NNRTIs with more rotatable bonds and flexibility. 23

Our data suggest select ARVs could also be tested as pre-exposure prophylaxis (PrEP) for COVID-19, if in vitro efficacy was shown. Drugs used for HIV PrEP (tenofovir/ emtricitabine; Truvada® or Descovy®, Gilead) are well tolerated (Charles B. Hare, 2020), and, if effective against SARS-CoV-2 in vitro, studies in trials would be more easily justified given the excellent tolerability, lack of drug-drug interactions, and well characterized safety profile,

when compared to PIs. We know of one randomized, placebo-controlled study of SARS-CoV 2 prophylaxis with tenofovir disoproxil fumarate/emtricitabine vs. hydroxychloroquine vs. both
 (vs placebo) in health care workers in Spain (N=4000) NCT04334928.

At this time the authors of this study would like to caution that this report has not made any conclusion or recommendation on any drug regimen change. However, our studies suggest that further investigations of the role of ARVs in SARS-CoV-2 prevention or amelioration of COVID-19 disease are warranted.

# 1 Acknowledgements

- 2 The authors declare no conflicts of interest.

# 1 References

2 Beck, B. R., Shin, B., Choi, Y., Park, S., & Kang, K. (2020). Predicting commercially available antiviral 3 drugs that may act on the novel coronavirus (SARS-CoV-2) through a drug-target interaction 4 deep learning model. Comput Struct Biotechnol J, 18, 784-790. 5 doi:10.1016/j.csbj.2020.03.025 6 Blanco, J. L., Ambrosioni, J., Garcia, F., Martinez, E., Soriano, A., Mallolas, J., . . . Investigators, C.-i. H. 7 (2020). COVID-19 in patients with HIV: clinical case series. Lancet HIV. doi:10.1016/S2352-8 3018(20)30111-9 9 Boettiger, D. C., Kerr, S., Ditangco, R., Chaiwarith, R., Li, P. C., Merati, T. P., . . . Database, T. A. H. O. 10 (2016). Tenofovir-based antiretroviral therapy in HBV-HIV coinfection: results from the 11 TREAT Asia HIV Observational Database. Antivir Ther, 21(1), 27-35. doi:10.3851/IMP2972 12 Buimovici-Klein, E., Lange, M., Ong, K. R., Grieco, M. H., & Cooper, L. Z. (1988). Virus isolation and 13 immune studies in a cohort of homosexual men. J Med Virol, 25(4), 371-385. Retrieved from 14 https://www.ncbi.nlm.nih.gov/pubmed/2902192 15 Cao, B., Wang, Y., Wen, D., Liu, W., Wang, J., Fan, G., . . . Wang, C. (2020). A Trial of Lopinavir-16 Ritonavir in Adults Hospitalized with Severe Covid-19. N Engl J Med. 17 doi:10.1056/NEJMoa2001282 18 Charles B. Hare, J. C., Peter Ruane, Jean-Michel Molina, Kenneth H. Mayer, Heiko Jessen, Robert M. 19 Grant, Joss J. De Wet, Melanie Thompson, Edwin DeJesus, Ramin Ebrahimi, Robertino Mera 20 Giler, Moupali Das, Diana Brainard, Scott McCallister. (2020). The Phase 3 DISCOVER Study: 21 Daily F/TAF or F/TDF for HIV Preexposure Prophylaxis. CROI. 22 Chen, Y. W., Yiu, C. B., & Wong, K. Y. (2020). Prediction of the SARS-CoV-2 (2019-nCoV) 3C-like 23 protease (3CL (pro)) structure: virtual screening reveals velpatasvir, ledipasvir, and other 24 drug repurposing candidates. F1000Res, 9, 129. doi:10.12688/f1000research.22457.2 25 Chu, C. M., Cheng, V. C., Hung, I. F., Wong, M. M., Chan, K. H., Chan, K. S., . . . Group, H. U. S. S. 26 (2004). Role of lopinavir/ritonavir in the treatment of SARS: initial virological and clinical 27 findings. Thorax, 59(3), 252-256. doi:10.1136/thorax.2003.012658 28 de Wilde, A. H., Jochmans, D., Posthuma, C. C., Zevenhoven-Dobbe, J. C., van Nieuwkoop, S., 29 Bestebroer, T. M., . . . Snijder, E. J. (2014). Screening of an FDA-approved compound library 30 identifies four small-molecule inhibitors of Middle East respiratory syndrome coronavirus 31 replication in cell culture. Antimicrob Agents Chemother, 58(8), 4875-4884. 32 doi:10.1128/AAC.03011-14 33 Dolinsky, T. J., Nielsen, J. E., McCammon, J. A., & Baker, N. A. (2004). PDB2PQR: an automated 34 pipeline for the setup of Poisson-Boltzmann electrostatics calculations. Nucleic Acids Res, 35 32(Web Server issue), W665-667. doi:10.1093/nar/gkh381 36 Douguet, D. (2010). e-LEA3D: a computational-aided drug design web server. Nucleic Acids Res, 37 38(Web Server issue), W615-621. doi:10.1093/nar/gkq322 38 Douguet, D. (2018). Data Sets Representative of the Structures and Experimental Properties of FDA-39 Approved Drugs. ACS Med Chem Lett, 9(3), 204-209. doi:10.1021/acsmedchemlett.7b00462 40 Douguet, D., Munier-Lehmann, H., Labesse, G., & Pochet, S. (2005). LEA3D: a computer-aided ligand 41 design for structure-based drug design. J Med Chem, 48(7), 2457-2468. 42 doi:10.1021/jm0492296 43 Duarte, R. R. R., Copertino Jr, D. C., Iñiguez, L. P., Marston, J. L., Nixon, D. F., & Powell, T. R. (2020). 44 Repurposing FDA-Approved Drugs for COVID-19 Using a Data-Driven Approach. ChemRxiv. 45 doi:10.26434/chemrxiv.12148764.v1 46 Elmezayen, A. D., Al-Obaidi, A., Sahin, A. T., & Yelekci, K. (2020). Drug repurposing for coronavirus 47 (COVID-19): in silico screening of known drugs against coronavirus 3CL hydrolase and 48 protease enzymes. J Biomol Struct Dyn, 1-13. doi:10.1080/07391102.2020.1758791

1 2	Ford, N., Vitoria, M., Rangaraj, A., Norris, S. L., Calmy, A., & Doherty, M. (2020). Systematic review of the efficacy and safety of antiretroviral drugs against SARS, MERS or COVID-19: initial
3	assessment. <i>J Int AIDS Soc, 23</i> (4), e25489. doi:10.1002/jia2.25489
4	Gao, Y., Yan, L., Huang, Y., Liu, F., Zhao, Y., Cao, L., Rao, Z. (2020). Structure of the RNA-
5	dependent RNA polymerase from COVID-19 virus. Science. doi:10.1126/science.abb7498
6	Gordon, C. J., Tchesnokov, E. P., Woolner, E., Perry, J. K., Feng, J. Y., Porter, D. P., & Gotte, M. (2020).
7	Remdesivir is a direct-acting antiviral that inhibits RNA-dependent RNA polymerase from
8	severe acute respiratory syndrome coronavirus 2 with high potency. J Biol Chem.
9	doi:10.1074/jbc.RA120.013679
10	Jockusch, S., Tao, C., Li, X., Anderson, T. K., Chien, M., Kumar, S., Ju, J. (2020). Triphosphates of
11	the Two Components in DESCOVY and TRUVADA are Inhibitors of the SARS-CoV-2
12	Polymerase. <i>BioRxiv, 2020.04.03.022939</i> . doi:10.1101/2020.04.03.022939
13	Kim, S., Chen, J., Cheng, T., Gindulyte, A., He, J., He, S., Bolton, E. E. (2019). PubChem 2019
14	update: improved access to chemical data. Nucleic Acids Res, 47(D1), D1102-D1109.
15	doi:10.1093/nar/gky1033
16	Korb, O., Stutzle, T., & Exner, T. E. (2009). Empirical scoring functions for advanced protein-ligand
17	docking with PLANTS. <i>J Chem Inf Model, 49</i> (1), 84-96. doi:10.1021/ci800298z
18	Kumar, G. N., Dykstra, J., Roberts, E. M., Jayanti, V. K., Hickman, D., Uchic, J., Granneman, G. R.
19	(1999). Potent inhibition of the cytochrome P-450 3A-mediated human liver microsomal
20	metabolism of a novel HIV protease inhibitor by ritonavir: A positive drug-drug interaction.
21	Drug Metab Dispos, 27(8), 902-908. Retrieved from
22	https://www.ncbi.nlm.nih.gov/pubmed/10421617
23	McKenna, C. E., Kashemirov, B. A., Peterson, L. W., & Goodman, M. F. (2010). Modifications to the
24	dNTP triphosphate moiety: from mechanistic probes for DNA polymerases to antiviral and
25	anti-cancer drug design. Biochim Biophys Acta, 1804(5), 1223-1230.
26	doi:10.1016/j.bbapap.2010.01.005
27	Oberg, B. (2006). Rational design of polymerase inhibitors as antiviral drugs. Antiviral Res, 71(2-3),
28	90-95. doi:10.1016/j.antiviral.2006.05.012
29	Sanders, J. M., Monogue, M. L., Jodlowski, T. Z., & Cutrell, J. B. (2020). Pharmacologic Treatments for
30	Coronavirus Disease 2019 (COVID-19): A Review. JAMA. doi:10.1001/jama.2020.6019
31	Shannon, A., Le, N. T., Selisko, B., Eydoux, C., Alvarez, K., Guillemot, J. C., Canard, B. (2020).
32	Remdesivir and SARS-CoV-2: Structural requirements at both nsp12 RdRp and nsp14
33	Exonuclease active-sites. Antiviral Res, 178, 104793. doi:10.1016/j.antiviral.2020.104793
34	Sluis-Cremer, N., & Tachedjian, G. (2008). Mechanisms of inhibition of HIV replication by non-
35	nucleoside reverse transcriptase inhibitors. Virus Res, 134(1-2), 147-156.
36	doi:10.1016/j.virusres.2008.01.002
37	Wang, Y., Zhang, D., Du, G., Du, R., Zhao, J., Jin, Y., Wang, C. (2020). Remdesivir in adults with
38	severe COVID-19: a randomised, double-blind, placebo-controlled, multicentre trial. The
39	Lancet, 0(0). doi:10.1016/s0140-6736(20)31022-9
40	Xu, L., Liu, H., Murray, B. P., Callebaut, C., Lee, M. S., Hong, A., Desai, M. C. (2010). Cobicistat (GS-
41	9350): A Potent and Selective Inhibitor of Human CYP3A as a Novel Pharmacoenhancer. ACS
42	Med Chem Lett, 1(5), 209-213. doi:10.1021/ml1000257
43	Xu, X., Liu, Y., Weiss, S., Arnold, E., Sarafianos, S. G., & Ding, J. (2003). Molecular model of SARS
44	coronavirus polymerase: implications for biochemical functions and drug design. Nucleic
45	Acids Res, 31(24), 7117-7130. doi:10.1093/nar/gkg916
46	Yamamoto, N., Yang, R., Yoshinaka, Y., Amari, S., Nakano, T., Cinatl, J., Yamamoto, N. (2004). HIV
47	protease inhibitor nelfinavir inhibits replication of SARS-associated coronavirus. Biochem
48	<i>Biophys Res Commun, 318</i> (3), 719-725. doi:10.1016/j.bbrc.2004.04.083
49	Zhang, L., Lin, D., Sun, X., Curth, U., Drosten, C., Sauerhering, L., Hilgenfeld, R. (2020). Crystal
50	structure of SARS-CoV-2 main protease provides a basis for design of improved alpha-
51	ketoamide inhibitors. Science. doi:10.1126/science.abb3405

Drug Name	RDRP PLANTS score	Mpro PLANTS score	Drug Class
Abacavir <sub>1</sub>	-107.19	-80.99	NRTI
Cobicistat	-106.57	-108.7	CYP3A Inhibitor
Emtricitabine1	-106.22	-80.84	NRTI
Atazanavir	-99.33	-105.72	PI
Saquinavir	-98.68	-107.57	PI
Ritonavir	-97.79	-111.52	PI
Zidovudine <sub>1</sub>	-96.59	-76.33	NRTI
Lamivudine <sub>1</sub>	-95.23	-78.73	NRTI
Indinavir	-94.83	-103.47	PI
Lopinavir	-94.55	-96.47	PI
ASC093	-92.46	-100.31	PI
<b>Tenofovir</b> 1	-91.16	-75.77	NRTI
Darunavir	-88.76	-87.9	PI
Nelfinavir	-86.22	-94.98	PI
Amprenavir	-85.13	-88.07	PI
Maraviroc	-84.44	-90.46	FI
Tipranavir	-83.84	-99.2	PI
Bictegravir	-81.93	-84.99	INSTI
Raltegravir	-81.08	-85.61	INSTI
Elvitegravir	-80.62	-89.42	INSTI
Dolutegravir	-78.4	-79.49	INSTI
Delavirdine	-75.24	-75.27	NNRTI
Etravirine	-73.13	-73.51	NNRTI
Efavirenz	-71.94	-66.03	NNRTI
Rilpivrine	-70.41	-77.66	NNRTI
Tenofovir <sub>2</sub>	-69.54	-62.65	NRTI
Abacavir <sub>2</sub>	-68.57	-74.97	NRTI
Doravirine	-67.97	-70.19	NNRTI
Emtricitabine <sub>2</sub>	-64.28	-69.82	NRTI
Nevirapine	-64.08	-68.18	NNRTI
Lamivudine <sub>2</sub>	-62.76	-67.67	NRTI
Zidovudine <sub>2</sub>	-49	-64.37	NRTI

<sup>1</sup>Active Form, Triphosphorylated/Diphosphorylated; <sup>2</sup>Inactive form: No additional phosphate groups; <sup>3</sup>ASC09 is also referred to as TMC-310911and is currently not FDA approved; RDRP: RNA-dependent RNA polymerase; Mpro: Main protease; NRTI: Nucleoside/Nucleotide Reverse Transcriptase Inhibitor; NNRTI: Non-Nucleoside Reverse Transcriptase Inhibitor; PI: Protease Inhibitor; FI: Fusion Inhibitor

**Table 1.** Docking scores for HIV drugs tested against SARS-CoV-2 RdRp and Mpro. The HIV ARV inhibitor class is also referenced.

Clinical Trial	Conditions	Interventions
		Leningvin/ritenevin, Undrewschleregwing Culfete, Legenter, Disselses
NC104328012		Lopinavir/ritonavir, Hydroxychloroquine Sulfate, Losartan, Placebos
NC104333953	COVID-19 HIV/AIDS	Other: No Intervention
NCT04321993	COVID-19	Lopinavir/ritonavir, Hydroxychloroquine sulfate, Baricitinib (janus kinase
		inhibitor), Sarilumab (anti-IL-6 receptor)
NCT04343768	COVID-19	Hydroxychloroquine, <b>Lopinavir / Ritonavir</b> , Interferon Beta-1A, Interferon Beta-
		ID Hydrowychloroguina, Blaacha of Hydrowychloroguina, Loninavir and ritanavir
NCT04328285	COVID-19	Placebo of <b>LPV/r</b> Tablets
NCT04350671	COVID-19	Interferon Beta-1A, Lopinavir / Ritonavir, Single Dose of Hydroxychloroquine
NCT04350684	COVID-19	Umifenovir, Interferon-β 1a, <b>Lopinavir / Ritonavir</b> , Single Dose of
		Hydroxycnioroquine, Standards of Care
NCT04330690		Lopinavir/ritonavir, hydroxychioroquine, remdesivir
NC104307693	COVID-19	Lopinavir/ritonavir, Hydroxychloroquine sulfate
NCT04331470	COVID-19	Levamisole Pill + Budesonide+Formoterol inhaler, Lopinavir/Ritonavir + hvdroxvchloroguine
NCT04346147	COVID-19 Pneumonia	Hidroxycloroquine, Lopinavir/ritonavir, Imatinib tablets, Baricitinib Oral Tablet
NCT04345276	COVID-19	Danoprevir+Ritonavir
NCT04291729	COVID-19	Ganovo+ritonavir+/-Interferon nebulization
		Carrimycin, lopinavir/ritonavir tablets or Arbidol or chloroquine phosphate,
NC104286503	COVID-19	basic treatment
NCT04255017	COVID-19	Abidol hydrochloride, Oseltamivir, Lopinavir/ritonavir
NCT04261270	COVID-19 Pneumonia	ASC09F+Oseltamivir, Ritonavir+Oseltamivir, Oseltamivir
		Lopinavir / ritonavir tablets combined with Xiyanping injection,
NC104295551	COVID-19	Lopinavir/ritonavir treatment
		Emtricitabine/tenofovir disoproxil, Hydroxychloroquine, Placebo:
NC104334920	COVID-19	Emtricitabine/tenofovir disoproxil Placebo, Placebo: Hydroxychloroquine
NCT04245049		Remdesivir, Lopinavir/ritonavir, Interferon Beta-1A, HydroxychloroquineOther:
NC104315940	0010-19	Standard of care
NCT04252274	Pneumonia, Pneumocystis	Darunavir and Cobicistat
	Coronavirus	Fixed-duration Hydrocortisone, Shock-dependent hydrocortisone, Ceftriayone
		Moxifloxacin or Levofloxacin Pineracillin-tazohactam Ceftaroline Amoxicillin-
	Community-acquired	clavulanate Macrolide administered for 3-5 days Macrolide administered for un
NCT02735707	Pneumonia Influenza	to 14 days. Five-days oseltamivir. Ten-days oseltamivir. <b>Loninavir/ritonavir</b>
10102100101		Hydroxychloroquine Hydroxychloroquine + Ioninavir/ritonavir Interferon-
		(E<1a Anakinra Eixed-duration higher dose Hydrocortisone Tocilizumah
		Sarilumah
NCT04321174	COVID-19 PrEP	
NCT04261907	COVID-19	ASC09/ritonavir group Joninavir/ritonavir group
NCT04275388	COVID-19 Preumonia	Xivanning injection <b>Loninavir / ritonavir</b> alpha-interferon pebulization
110104270000		Conventional medicines (Oxygen therapy, alfa interferon via aerosol inhalation
		and <b>Ioninavir/ritonavir</b> ) and Traditional Chinese Medicines (TCMs) granules
NCT04251871	COVID-19 Pneumonia	Conventional medicines (Oxygen therapy, alfa interferon via aerosol inhalation
		and lopinavir/ritonavir)
		Chloroquine or Hydroxychloroquine. Lopinavir/Ritonavir Other: Best standard
NCT04351724	COVID-19	of care. Rivaroxaban, Thromboprophylaxis, Candesartan, non-RAS blocking
		antihypertensives. Clazakizumab. placebo for clazakizumab
NCT04276688	COVID-19	Lopinavir/ritonavir, Ribavirin. Interferon Beta-1B
		• •

NCT04278404	COVID-19, multiple other conditions	The POP02 study is collecting bodily fluid samples (i.e., whole blood, effluent samples) of children prescribed the following drugs of interest per standard of care: Oseltamivir, Oxycodone, Risperidone, Sertraline, Zolpidem, Dextroamphetamine /Amphetamine, Azithromycin, Chloroquine, Hydroxychloroquine, <b>Lopinavir/Ritonavir</b> , Ribavirin, (+54 other drugs of interest not listed here)
NCT04359095	COVID-19	Hydroxychloroquine, <b>Lopinavir / Ritonavir Pill</b> , Azithromycin, Standard treatment
NCT04364022	Prevention of COVID-19	Hydroxychloroquine Sulfate 200 MG [Plaquenil], Lopinavir/ritonavir
NCT04372628	COVID-19	Hydroxychloroquine, Lopinavir/ritonavir 400 mg/100 mg, Placebo
NCT04366245	COVID-19	Hyperimmune plasma, Hydroxychloroquine + Azithromycin or <b>Lopinavir/ritonavir</b> + Interferon β-1b + Hydroxychloroquine
NCT04365582	COVID-19	Azithromycin, Hydroxychloroquine, Lopinavir 200Mg/Ritonavir 50Mg Tab
NCT04373044	COVID-19, multiple other conditions (10+)	Baricitinib, Hydroxychloroquine, Lopinavir/Ritonavir, Remdesivir

**Table 2.** Clinical trials with approved and experimental HIV ARVs included in the trial as treatmentfor COVID-19. HIV ARV drugs used in each clinical trial are in bold.



PDB ID: 6M71



**Figure 2.** Image of the Main Protease (Mpro, left) and RNA-dependent RNA Polymerase (RdRp, right) of SARS-CoV-2, catalytic sites are inside white boxes.











Figure 3. PLANTS in silico docking images for tenofovir diphosphate (top left), emtricitabine triphosphate (top right), abacavir triphosphate (bottom left) and lamivudine triphosphate (bottom right) against the RNA-dependent RNA Polymerase (RdRp) of SARS-CoV-2. Polar and Hydrogen interactions are represented by yellow dotted lines.









Figure 4. PLANTS in silico docking images for Cobicistat (top row), Atazanavir (middle row) and Saquinavir (bottom row) against the Main Protease (Mpro) of SARS-CoV-2. Polar and Hydrogen interactions are represented by yellow dotted lines.







