

A temperature-sensitive RFID tag for the identification of cold chain failures

F. Vivaldi^{a*}, B. Melai^a, A. Bonini^a, N. Poma^a, P. Salvo^b, A. Kirchhain^a, S. Tintori^a, A. Bigongiari^c,
F. Bertuccelli^c, G. Isola^c, F. Di Francesco^a

^aDepartment of Chemistry and Industrial Chemistry, University of Pisa, via Giuseppe Moruzzi 13, 56124, Pisa, Italy.

^bInstitute of Clinical Physiology (IFC) of National Research Council (CNR), Via Moruzzi 1, 56124, Pisa, Italy.

^cCaen RFID srl, Via Vetraia 11, 55049, Viareggio, Italy.

* Corresponding author, federicomaria.vivaldi@phd.unipi.it

Abstract

Quality and safety of the cold chain undergo strict international regulations that identify storage and shipping temperatures. In fact, the improper handling and transportation of temperature-sensitive products such as food and pharmaceuticals may have harmful effects on human health and a negative economic impact. A passive RFID tag modified with a copper-doped ionic liquid was used to detect the crossing of a temperature threshold (8 °C) during the shipping of medical products. The tag was insensitive to humidity variations and irreversibly changed its status once temperature exceeded the ionic liquid melting point, which can be tuned by changing the concentration of dopant.

Keywords: cold chain; RFID; ionic liquid; temperature sensor; shipment control; perishable goods

1. Introduction

The global market of perishable and temperature-sensitive goods such as pharmaceuticals, medical products and fresh food is constantly growing at a compound annual growth rate (CAGR) of about 7.7% [1]. Other analysts foresee a CAGR of 17.9% from 2019 to 2026 [2]. The transportation management needs sophisticated software and procedures to handle the delivery and provide a high quality service to the customers. For a successful logistic platform, the implementation of an effective

28 cold chain system is crucial to keep temperature-sensitive products in optimal storage and transport
29 conditions. A cold chain can contain a high density of nodes designed to keep goods within a safe
30 temperature range uninterruptedly before the products reach consumers [3]. National and
31 international regulations and directives provide shipping temperature intervals for storage and
32 transportation that depend on the characteristics of the goods [4,5]. For example, most temperature-
33 sensitive medical products must be kept at 2–8 °C [6], fresh fish at 0–4 °C or at -18–15 °C if frozen
34 [7], meat at -25 °C or colder, and fruits like bananas between 12 and 16 °C [8]. However, several
35 factors such as packaging failure, human error, equipment malfunction and non-uniform cooling
36 standards across countries can make cold chain fail. These risks are an important health threat and
37 sources of economic losses. For example, the improper handling and transportation account for an
38 estimated annual loss of more than \$750 billion in the food market [9], whereas Biopharma companies
39 annually spend \$260 billion to ensure the efficacy of their cold chain logistics [10]. Therefore, there
40 is a growing demand of low cost and easy-to-use temperature sensors capable of tracking the cold
41 chain to improve management and identify failures.

42 Different solutions for temperature monitoring and identification of cold chain failures have been
43 described in the scientific literature or are commercially available. Time-temperature indicators
44 (TTIs) are lightweight and inexpensive (few euro cents per piece) colorimetric tags that undergo a
45 mechanical, microbiological, chemical or enzymatic change when temperature exceeds a predefined
46 threshold [11]. There are several off-the-shelf TTIs, e.g. CheckPoint™, 3M™ MonitorMark™,
47 Fresh-Check®, TT Sensor™ and Keep-it®, which can be used for different temperature ranges [12].
48 Since the color change is irreversible, TTIs are disposable; an electronic indicator to improve the
49 color reading [13] is sometimes provided. Although available since 1970s, TTIs have never gained a
50 wide market penetration because of their low accuracy due to the ambiguous quantification of color
51 change [14].

52 Thermistors, resistance temperature detectors (RTDs) and thermocouples are widely used for
53 monitoring temperature in many fields, such as industrial and manufacturing processes, food and
54 beverage products, environment control, consumer electronics, and healthcare [15–18]. These
55 temperature sensors can be coupled or integrated in a datalogger for a continuous monitoring.
56 Thermocouples can be inaccurate if exposed to moisture, voltage noise, or large variations in ambient
57 temperature, whereas RTDs are expensive and rarely used in cold chain logistics [19,20]. Due to good
58 accuracy and small size [21–24], many temperature-tracking systems prefer thermistors, but those
59 allowing high accuracy measurements are expensive, fragile and need filters or DC voltage because
60 of their high resistance [25]. In recent years, other techniques have been investigated to measure
61 temperature in cold chain products, e.g. infrared thermography and fluorescence, but their use is still
62 limited because of high cost, limited applicability and inaccuracy [26–28].

63 Although the aforementioned sensors and technologies allowed temperature measurement, the cold
64 chain management system had poor access to data during shipment before the adoption of wireless
65 data transfer. Technologies such as Bluetooth and Radio frequency identification (RFID) allow the
66 temperature monitoring without a direct access of packagings. RFID is widely adopted in logistics
67 because it is an affordable technology that allows to track the refrigeration status in real-time from
68 the production until the sale of the product [29,30]. Active and semi-passive RFID tags use a power
69 source, typically a battery, whereas passive tags are powered by the external electromagnetic field
70 emitted from the reader. Passive RFID tags are usually preferred for product traceability as they are
71 low cost and long lasting. They consists of a microchip and an antenna [31] and are accessed by a
72 reader operating in the low frequency (125 and 134 KHz), high frequency (13.56 MHz), ultra-high
73 frequency (UHF) (860 – 960 MHz) and microwave (2.4 GHz and 5.8 GHz) bands. Higher frequencies
74 allow the reader to connect with tags in a broader range, from a few centimeters up to several meters
75 [32].

76 Several papers in literature have described RFID tags using commercial or integrated silicon sensors
77 to monitor temperature in cold chain systems [29,33–39]. Bhattacharyya et al. [40] fabricated a
78 temperature-sensitive polymer that actuates a metal plate between two RFID tags. In this paper, we
79 describe the fabrication of a passive RFID tag modified with an ionic liquid, [C₁₂mim][Tf₂N], to sense
80 temperature. Ionic liquids are salts in liquid state at temperatures lower than 100 °C [41] that can be
81 used to fabricate different types of sensors [42], whose melting temperature can be modified by the
82 selection of the cation/anion pair [43]. Our sensor exploits the melting of copper-doped
83 [C₁₂mim][Tf₂N] (Cu-IL) to change the status of a tag and identify failures in the cold chain.

84

85 **2. Materials and Methods**

86 **2.1. Reagents and Materials**

87 1-methylimidazole (purity 98%), lithium bis(trifluoromethanesulfonyl)imide (purity 99%) and
88 bis(trifluoromethanesulfonyl)imide (99% purity) were purchased from Iolitec. 1-Bromododecane
89 (purity 97 %) and dichloromethane (purity 99%), sodium chloride, magnesium nitrate and potassium
90 acetate were purchased from Sigma Aldrich. Copper Oxide (II) was purchased from Carlo Erba.

91 **2.2. Synthesis of the [C₁₂mim][Tf₂N]**

92 The synthesis of [C₁₂mim][Tf₂N] was performed in a two-step reaction. Figure 1 shows the formation
93 of the bromine ionic liquid by the addition of a halogen alkane to a methyl imidazole center. Two
94 aliquots of 1-methylimidazole (10 mL, 0.1218 mol) and deionized water (20 mL) were added to 28
95 mL of 1-bromododecane (0.1218 mol). The mixture was heated and refluxed for 12 h until a unique
96 phase was obtained.

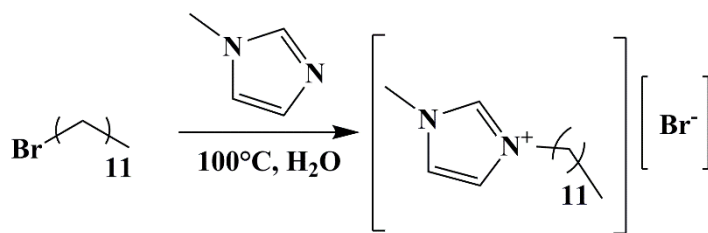


Fig. 1. Synthesis of [C₁₂mim][Br].

100 In the second step, the anion was exchanged with lithium bis(trifluoromethanesulfonyl)imide (Fig.

101 2). An aliquot of 5.152 g Li(Tf₂N) (0.0179 mol) was added to 20 mL of deionized water whereas

102 [C₁₂mim][Br⁻] was supplemented under magnetic stirring at room temperature. After the addition of

103 30 mL of dichloromethane, the solution was stirred for 2 h at 40 °C. The organic phase was extracted

104 removing LiBr after five washing cycles with deionized water. Then, the organic phase was dried

105 under vacuum for 2 h at 60 °C in order to remove the organic solvent and the remaining water.

106 NMR analysis was performed on the synthesized ionic liquid with a Bruker Advance DRX 400:

107 ¹H NMR (401 MHz, C₆D₆), δ (ppm): 8.70 (s, 1H), 7.54 (s, 1H), 7.47 (s, 1H), 4.24 (t, J = 7.1 Hz, 2H),

108 3.96 (s, 3H), 1.95 (s, 2H), 1.39 (d, J = 20.5 Hz, 16H), 0.96 (t, J = 6.5 Hz, 3H).

109

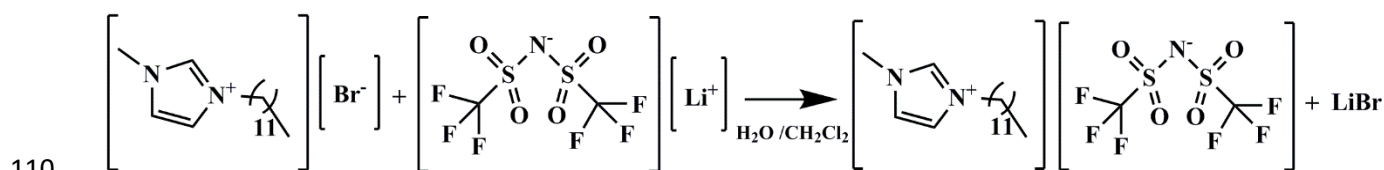


Fig. 2. Anion exchange with lithium bis(trifluoromethanesulfonyl)imide to obtain the hydrophobic ionic liquid [C₁₂mim][Tf₂N].

113 2.3. Preparation of Cu(Tf₂N)₂ and doped ionic liquid

114 A solution of $\text{Cu}(\text{Tf}_2\text{N})_2$ was prepared by mixing 2.303 g of copper oxide (0.0289 mol) with 20.003
115 g of bis(trifluoromethylsulfonil)imide (0.05692 mol) in 20 mL of deionized water. After stirring at
116 room temperature overnight, the solution was filtered to remove any excess of copper oxide. The
117 $\text{Cu}(\text{Tf}_2\text{N})_2$ solution and the $[\text{C}_{12}\text{mim}][\text{Tf}_2\text{N}]$ were mixed (molar ratio 0.5) to obtain Cu-IL. Cu-IL was
118 kept under vacuum at room temperature for about 6 h to remove excess water.

119

120 **2.4. Characterization**

121 In order to calculate the melting and freezing point of the ionic liquids, Differential Scan Calorimetry
122 (DSC) was performed under nitrogen flux (80 mL/min) with a DSC228e Mettler Toledo equipped
123 with a cooling system. The melting and freezing points of Cu-IL were extrapolated from the
124 regression line fitting the onset temperatures at different scan rates (5, 10 and 20 °C/min) calculated
125 by the STARe software (Mettler Toledo). Two identical aluminum pans were used as sample holder
126 and reference for the analysis, respectively.

127 The amount of water contained in each sample was assessed by a thermogravimetric analysis (TGA,
128 Q5000, TA instruments, heating rate 20 °C/min). Flasks at controlled humidity were prepared as
129 previously reported [44]. Briefly, saturated solutions of potassium acetate, magnesium nitrate and
130 sodium chloride were prepared to obtain relative humidity levels of 22.5, 52.9 and 75.3% at room
131 temperature (25 °C).

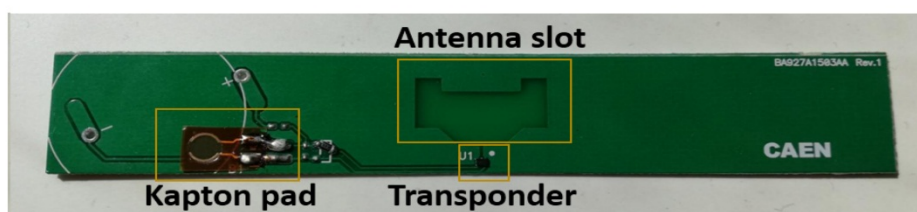
132

133 **2.5. Working principle of the RFID tag**

134 The UHF RFID tag mounts an UCODE G2iM+ transponder (NXP) that can detect the open/closed
135 circuit state between two pads (Figure 3). A drop (10 μL) of Cu-IL is cast between the pads onto the
136 RFID pads kept at low temperature (-40 °C). The instant freezing of the drop avoids any electrical
137 connection between the tracks. Once the transponder is activated from the reader (R1260I SLATE

138 RFID UHF desktop reader, CAEN RFID), the chip injects a current pulse (amplitude 300 nA, duration
 139 200 μ s) through the pads. The integrated circuit detects a closed or an open circuit if the resistance
 140 across the pads is lower than 2 M Ω or higher than 20 M Ω , respectively. The status of the contact is
 141 stored inside transponder's memory and can be retrieved by the reader. If temperature rises above the
 142 Cu-IL melting point, the drop melts and short-circuits the pads, so that the transponder detects a
 143 switch from the open to the closed state.

144



145

146 Figure 3. An illustrative Cu-IL RFID tag.

147 2.6. Electrochemical impedance spectroscopy analysis

148 The Cu-IL impedance over time was recorded using a potentiostat (Palmsens 4, Palmsens) in a two-
 149 electrode configuration and a dedicated software (PSTrace, Palmsens). Table 1 shows the parameters
 150 used for electrochemical impedance spectroscopy (EIS). Temperature was monitored by a thermistor
 151 (NTC2.2K3359I, Betatherm).

152 Table 1. Settings for the electrochemical impedance spectroscopy.

Scan type	Fixed potential
E_{dc}	0.0 V
E_{ac}	0.01 V
t_{run}	1000 s
Frequency type	Fixed frequency
Frequency	10000 Hz

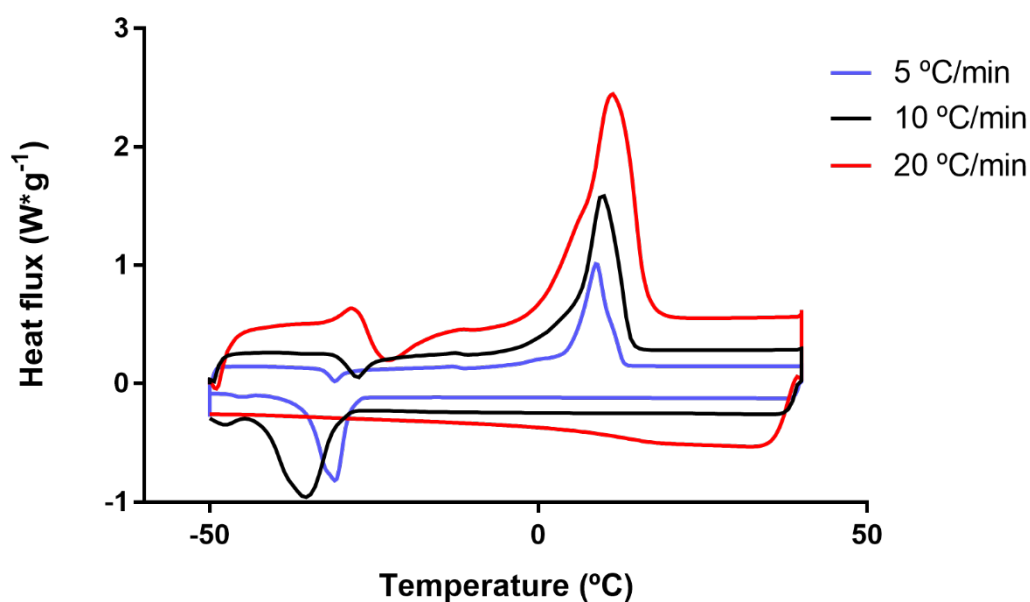
153

154 **3. Results and Discussion**

155 **3.1. DSC and TGA thermograms**

156 Figure 4 shows the DSC thermogram of the Cu-IL. The peaks at about 10 °C are related to the melting
157 of Cu-IL, whereas freezing points are close to -40 °C due to the typical undercooling of these
158 compounds [45]. Several additional smaller peaks can be noticed that are likely related to glass
159 transitions due to ion reorganization within the solid structure [46].

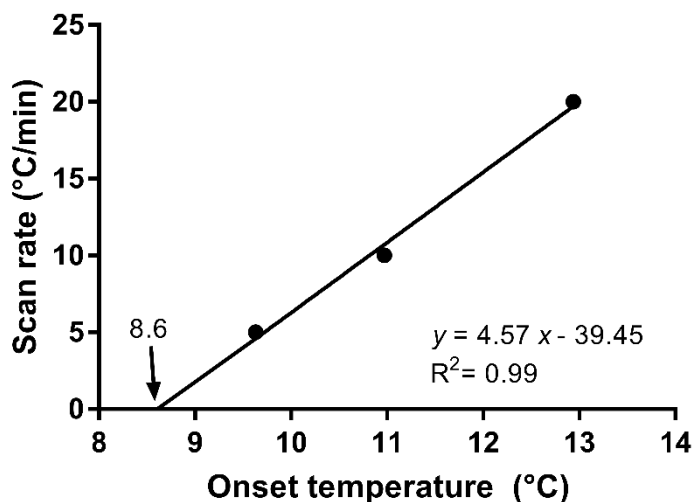
160



161

162 Figure 4. DSC thermograms of Cu-IL at three different temperature scan rates.

163 Figure 4 also shows a dependence of phase transition temperatures from the temperature scan rate, so
164 that the most accurate temperature values are those obtained with extremely slow variations of
165 temperature. Figure 5 shows the extrapolation of the Cu-IL melting and freezing points, i.e. 8.6 and
166 -35.3 °C, respectively, by regression from the onset temperature values estimated at different
167 temperature scan rates. Since the optimal temperature for refrigerated products ranges between 2-8
168 °C [47], the melting point of Cu-IL was set at the upper boundary to verify if the cold chain was
169 preserved.



170

171 Figure 5. Extrapolation of the Cu-IL melting points by regression from the onset temperature values
 172 estimated at different temperature scan rates.

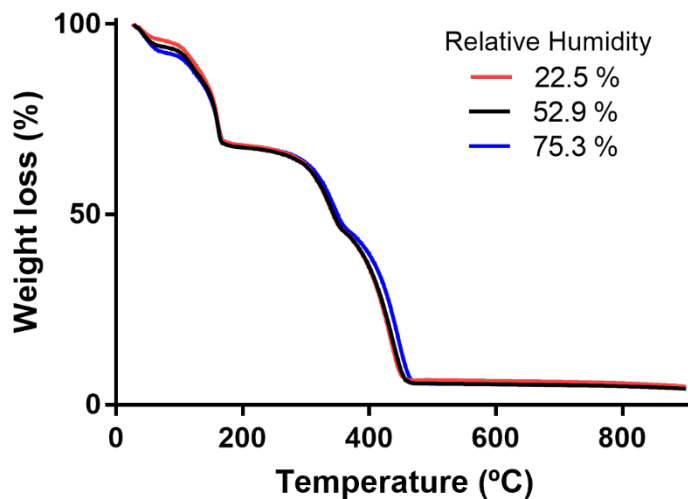
173

174 The supercooling effect of Cu-IL ($\Delta T > 43.9$ °C) prevents re-freezing, and its combination with the
 175 spreading of the drop makes the process irreversible.

176 Since ionic liquids tend to absorb water [48] and this can modify the freezing and melting points, the
 177 behavior of the Cu-IL was tested at three different levels of relative humidity (RH). Figure 6 shows
 178 three similar Cu-IL thermograms at 22.5%, 52.9% and 75.3% RH. The weight loss in the range [25,
 179 100] °C (Table 2) was attributed to the loss of water from the sample, which was followed by the
 180 Cu(Tf₂N)₂ degradation in the range [100, 250] °C. The final part ($T > 250$ °C) can be associated with
 181 the degradation of Cu-IL. The water loss slightly increased at higher RH values, i.e. from 4 to 7.6%,
 182 which can be explained by a predominant hydrophobic behavior from [C₁₂mim][Tf₂N] over the
 183 hydrophilic copper [49].

184

185



186

187

Figure 6. TGA for Cu-IL at three different levels of relative humidity.

188

189 Table 2. Cu-IL weight losses at three relative humidity levels.

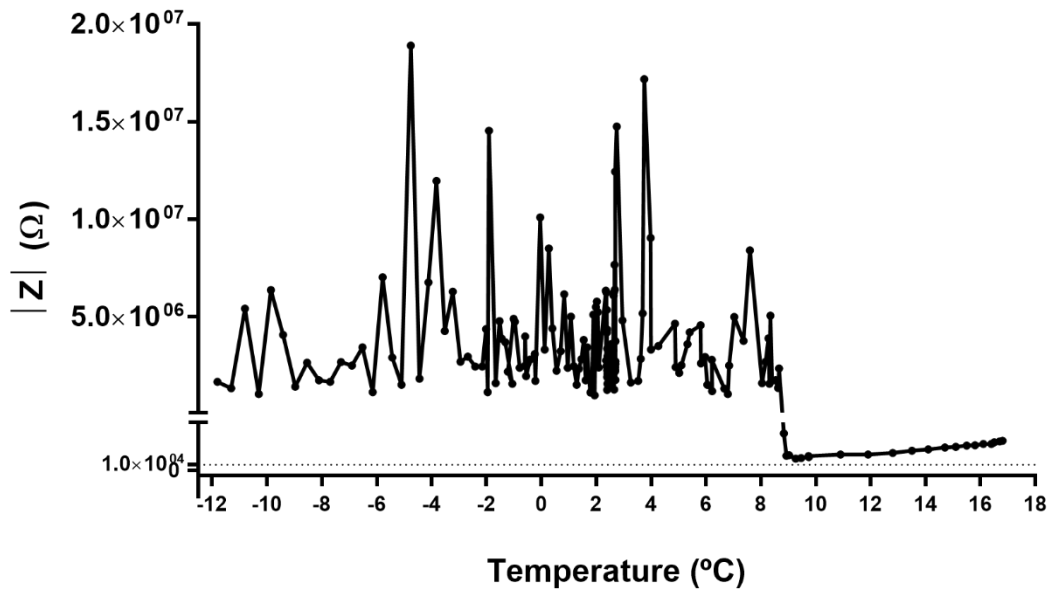
Relative Humidity	Temperature Range	Weight Loss (%)
22.5 %	25 – 100 °C	4
	100 – 250 °C	27.9
	250 – 900°C	68.1
52.9 %	25 – 100 °C	5.9
	100 – 250 °C	26.7
	250 – 900°C	67.5
75.3 %	25 – 100 °C	7.6
	100 – 250 °C	24.6
	250 – 900°C	67.8

190

191

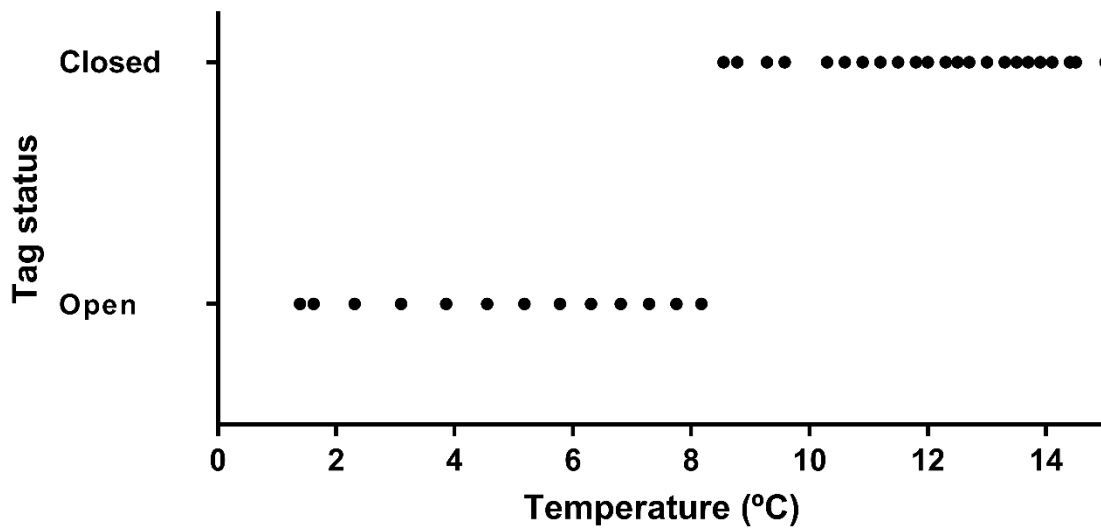
192 **3.2. Electrochemical impedance spectroscopy and temperature measurements**

193 Figure 7 shows the impedance modulus ($|Z|$) over temperature. The noise below 8 °C is intrinsic to
194 the open circuit condition. After melting, the impedance decreased sharply leading to a closed circuit
195 state. The slow $|Z|$ increase after 12 °C could depend on thermal noise. Figure 8 shows the
196 corresponding output of the RFID reader.



197

198 Figure 7. Impedance module of Cu-IL over temperature.



199

200 Figure 8. Tag status versus temperature.

201 **4. Conclusion**

202 A modified RFID tag for the identification of failures in the cold chain during transport of
203 temperature-sensitive products has been proposed. The doping of an ionic liquid with copper ions
204 allowed a fine tuning its melting and freezing points, which are remarkably different due to
205 overcooling. We showed how to exploit the irreversible melting of our Cu-IL upon the crossing of
206 the 8 °C threshold to induce a transition in the RFID tag state. The Cu-IL proved stable at different
207 relative humidity levels, which allows the tag to be used in the cold chain system flawlessly. However,
208 enclosure of the tag in a sealed envelope would at the same time avoid interferences from ambient
209 humidity and prevent contamination from copper. A main advantage of using a modified RFID tag
210 relies in the simultaneous acquisition of information from multiple packages from a same reader
211 without any direct or visual contact. The tags can be inexpensive and the threshold temperature is
212 easily tuned by changing the composition of the CU-IL.

213

214 **Acknowledgements**

215 Regione Toscana and University of Pisa are gratefully acknowledged for supporting this study under
216 the projects eQuality4logistics (POR FESR Toscana 2014-2020) and PRA_2018_26, respectively.

217

218 **References**

- 219 [1] Orbis Research, Global Perishable Goods Transportation Market Growth Opportunities and
220 Forecast 2018-2022, 2018.
- 221 [2] Grand View Research, Cold Chain Market Size, Share & Trends Analysis Report By Type
222 (Transportation, Monitoring Components, Storage), By Packaging, By Equipment, By
223 Application, By Region, And Segment Forecasts, 2019-2025, 2019.

- 224 [3] S. Mercier, S. Villeneuve, M. Mondor, I. Uysal, Time–Temperature Management Along the
225 Food Cold Chain: A Review of Recent Developments, *Compr. Rev. Food Sci. Food Saf.* 16
226 (2017) 647–667. doi:10.1111/1541-4337.12269.
- 227 [4] S.J. James, C. James, The food cold-chain and climate change, *Food Res. Int.* 43 (2010)
228 1944–1956. doi:10.1016/j.foodres.2010.02.001.
- 229 [5] World Health Organization (WHO), Annex 5: WHO good distribution practices for
230 pharmaceutical products, WHO Tech. Rep. Ser. 957 (2010) 235–264.
231 [https://www.who.int/medicines/areas/quality_safety/quality_assurance/GoodDistributionPrac](https://www.who.int/medicines/areas/quality_safety/quality_assurance/GoodDistributionPracticesTRS957Annex5.pdf)
232 [ticesTRS957Annex5.pdf](https://www.who.int/medicines/areas/quality_safety/quality_assurance/GoodDistributionPracticesTRS957Annex5.pdf).
- 233 [6] G.U, *Gazzetta Ufficiale Della Repubblica Italiana*, Italia, 2000.
- 234 [7] EUR-Lex, No Title, (n.d.).
- 235 [8] S. Mercier, S. Villeneuve, M. Mondor, I. Uysal, Time–Temperature Management Along the
236 Food Cold Chain: A Review of Recent Developments, *Compr. Rev. Food Sci. Food Saf.*
237 (2017). doi:10.1111/1541-4337.12269.
- 238 [9] FAO, Food wastage footprint. Impacts on natural resources., 2013. doi:ISBN 978-92-5-
239 107752-8.
- 240 [10] Pharmaceutical Commerce, *The Cold Chain Directory 2015: The 2015 Biopharma Cold*
241 *Chain Landscape*, 2015.
- 242 [11] P.S. Taoukis, Application of Time-Temperature Integrators for Monitoring and Management
243 of Perishable Product Quality in the Cold Chain, in: *Smart Packag. Technol. Fast Mov.*
244 *Consum. Goods*, 2008: pp. 61–74. doi:10.1002/9780470753699.ch4.
- 245 [12] P. Müller, M. Schmid, Intelligent packaging in the food sector: A brief overview, *Foods*.
246 (2019). doi:10.3390/foods8010016.

- 247 [13] V. Kramar, H. Maatta, H. Hinkula, O. Thorsen, G. Cox, Smart-fish system for fresh fish cold
248 chain transportation - Overall approach and selection of sensor materials, in: Conf. Open
249 Innov. Assoc. Fruct, 2018. doi:10.23919/FRICT.2017.8250183.
- 250 [14] A.T.M.M. Rahman, D.H. Kim, H.D. Jang, J.H. Yang, S.J. Lee, Preliminary study on
251 biosensor-type time-temperature integrator for intelligent food packaging, Sensors
252 (Switzerland). (2018). doi:10.3390/s18061949.
- 253 [15] Q. Shan, Y. Liu, G. Prosser, D. Brown, Wireless Intelligent Sensors Network for
254 Refrigerated Vehicle, (2004). doi:10.1109/CASSET.2004.1321941.
- 255 [16] P.H. Chou, C.T. Lee, Z.Y. Peng, J.P. Li, T.K. Lai, C.M. Chang, C.H. Yang, Y.L. Chen, C.C.
256 Nien, L.H. Chen, L.Y. Lai, J.C. Lu, S.C. Hung, A bluetooth-smart insulating container for
257 cold-chain logistics, Proc. - IEEE 6th Int. Conf. Serv. Comput. Appl. SOCA 2013. (2013)
258 298–303. doi:10.1109/SOCA.2013.46.
- 259 [17] A.A. Zaher, Smart Temperature Sensors: History, Current Practices, and Future Trends, in:
260 Process Anal. Des. Intensif. Microfluid. Chem. Eng. IGI Glob., 2019: p. 28.
- 261 [18] S. Wang, J. Tang, F. Younce, Temperature measurement, in: Encycl. Agric. Food, Biol. Eng.,
262 2003: pp. 987 – 993.
- 263 [19] P.S. Taoukis, M.C. Giannakourou, T.N. Tsironi, Monitoring and Control of the Cold Chain,
264 in: Handb. Frozen Food Process. Packag. 2nd Ed. Da-Wen Sun (Ed.), CRC Press. Boca
265 Raton, Florida, 2012.
- 266 [20] A. Tong, Improving the accuracy of temperature measurements, Sens. Rev. (2001).
267 doi:10.1108/02602280110398044.
- 268 [21] B.B. Maskey, J. Sun, K. Shrestha, S. Kim, M. Park, Y. Kim, H. Park, S. Lee, Y. Han, J. Lee,
269 Y. Majima, J. Kim, J. Lee, G. Bahk, G.R. Koirala, G. Cho, A smart food label utilizing roll-

- 270 to-roll gravure printed NFC antenna and thermistor to replace existing “use-by” date system,
271 IEEE Sens. J. (2019). doi:10.1109/jsen.2019.2948752.
- 272 [22] P.J. McColloster, A. Martin-de-Nicolas, Vaccine refrigeration, Hum. Vaccin. Immunother.
273 (2014). doi:10.4161/hv.27660.
- 274 [23] O.C. de Mello Vasconcelos, D. Duarte, J. de Castro Silva, N.F. Oliveros Mesa, B.J. Teruel
275 Mederos, S.T. de Freitas, Modeling ‘Tommy Atkins’ mango cooling time based on fruit
276 physicochemical quality, Sci. Hortic. (Amsterdam). (2019).
277 doi:10.1016/j.scienta.2018.09.068.
- 278 [24] A.A. Van Wyk, Assessment of the performance of thermocouple wires vs. thermistor sensors
279 in a cold room. Report to Perishable Products Export Control Board., 2003.
- 280 [25] T.G. Claggett, R.W. Worrall, B.G. Lipták, P.M.B. Silva Girão, Thermistors, in: Instrum.
281 Eng. Handbook, Vol. One Process Meas. Anal. CRC Press, 2003.
- 282 [26] R. Badia-Melis, U. Mc Carthy, L. Ruiz-Garcia, J. Garcia-Hierro, J.I. Robla Villalba, New
283 trends in cold chain monitoring applications - A review, Food Control. (2018).
284 doi:10.1016/j.foodcont.2017.11.022.
- 285 [27] F. Li, Z. Chen, Brief analysis of application of RFID in pharmaceutical cold-chain
286 temperature monitoring system, in: Proc. 2011 Int. Conf. Transp. Mech. Electr. Eng. TMEE
287 2011, 2011. doi:10.1109/TMEE.2011.6199709.
- 288 [28] K. Likar, M. Jevšnik, Cold chain maintaining in food trade, Food Control. (2006).
289 doi:10.1016/j.foodcont.2004.09.009.
- 290 [29] E. Abad, F. Palacio, M. Nuin, A.G. de Zárata, A. Juarros, J.M. Gómez, S. Marco, RFID
291 smart tag for traceability and cold chain monitoring of foods: Demonstration in an
292 intercontinental fresh fish logistic chain, J. Food Eng. 93 (2009) 394–399.

- 293 doi:10.1016/j.jfoodeng.2009.02.004.
- 294 [30] Y. Bo, L. Danyu, Application of RFID in cold chain temperature monitoring system, in: 2009
295 Second ISECS Int. Colloq. Comput. Commun. Control. Manag. CCCM 2009, 2009.
296 doi:10.1109/CCCM.2009.5270408.
- 297 [31] R. Want, An Introduction to RFID Technology, *IEEE Pervasive Comput.* 5 (2006) 25–33.
298 doi:10.1109/MPRV.2006.2.
- 299 [32] P. Nikitin, K.V.S. Rao, S. Lazar, An overview of near field UHF RFID, in: 2007 IEEE Int.
300 Conf. RFID, *IEEE RFID 2007*, 2007. doi:10.1109/RFID.2007.346165.
- 301 [33] K. Opasjumruskit, T. Thanthipwan, O. Sathusen, P. Sirinamarattana, P. Gadmanee, E.
302 Pootarapan, N. Wongkomet, A. Thanachayanont, M. Thamsirianunt, Self-powered wireless
303 temperature sensors exploit RFID technology, *IEEE Pervasive Comput.* 5 (2006) 54–61.
304 doi:10.1109/MPRV.2006.15.
- 305 [34] L. Boquete, R. Cambrella, J.M. Rodriguez-Ascariz, J.M. Miguel-Jimnez, J.J. Cantos-Frontela,
306 J. Dongil, Portable system for temperature monitoring in all phases of wine production, *ISA*
307 *Trans.* (2010). doi:10.1016/j.isatra.2010.03.001.
- 308 [35] C. Amador, J.P. Emond, M.C. do N. Nunes, Application of RFID technologies in the
309 temperature mapping of the pineapple supply chain, *Sens. Instrum. Food Qual. Saf.* (2009).
310 doi:10.1007/s11694-009-9072-6.
- 311 [36] K. Reiners, A. Hegger, E.F. Hessel, S. Böck, G. Wendl, H.F.A. Van den Weghe, Application
312 of RFID technology using passive HF transponders for the individual identification of
313 weaned piglets at the feed trough, *Comput. Electron. Agric.* (2009).
314 doi:10.1016/j.compag.2009.05.010.
- 315 [37] L. Ruiz-Garcia, L. Lunadei, The role of RFID in agriculture: Applications, limitations and

- 316 challenges, *Comput. Electron. Agric.* (2011). doi:10.1016/j.compag.2011.08.010.
- 317 [38] W. Tingman, Z. Jian, Z. Xiaoshuan, Fish product quality evaluation based on temperature
318 monitoring in cold chain, *African J. Biotechnol.* (2010). doi:10.5897/AJB09.1344.
- 319 [39] R. Jedermann, L. Ruiz-Garcia, W. Lang, Spatial temperature profiling by semi-passive RFID
320 loggers for perishable food transportation, *Comput. Electron. Agric.* 65 (2009) 145–154.
321 doi:10.1016/j.compag.2008.08.006.
- 322 [40] R. Bhattacharyya, C. Di Leo, C. Floerkemeier, S. Sarma, L. Anand, RFID tag antenna based
323 temperature sensing using shape memory polymer actuation, in: *Proc. IEEE Sensors, 2010.*
324 doi:10.1109/ICSENS.2010.5690951.
- 325 [41] T. Welton, Room-Temperature Ionic Liquids. Solvents for Synthesis and Catalysis, *Chem.*
326 *Rev.* 99 (1999) 2071–2084. doi:10.1021/cr980032t.
- 327 [42] T. Schäfer, F. Di Francesco, R. Fuoco, Ionic liquids as selective depositions on quartz crystal
328 microbalances for artificial olfactory systems-a feasibility study, *Microchem. J.* (2007).
329 doi:10.1016/j.microc.2006.06.001.
- 330 [43] F. Di Francesco, C. Ferrari, L. Moni, B. Melai, L. Bernazzani, C. Chiappe, Tuning of the
331 freezing and melting points of [Hmim][NO₃] by the addition of water and nitrate salts, *RSC*
332 *Adv.* 4 (2014) 40407–40413. doi:10.1039/c4ra06290f.
- 333 [44] L. Greenspan, Humidity fixed points of binary saturated aqueous solutions, *J. Res. Natl. Bur.*
334 *Stand.* (1934). 81A (1997). doi:10.2307/2406893.
- 335 [45] M. Galiński, A. Lewandowski, I. Stepniak, Ionic liquids as electrolytes, *Electrochim. Acta.*
336 51 (2006) 5567–5580. doi:10.1016/j.electacta.2006.03.016.
- 337 [46] S.M. Murray, T.K. Zimlich, A. Mirjafari, R.A. O'Brien, J.H. Davis, K.N. West,
338 Thermophysical properties of imidazolium-based lipidic ionic liquids, *J. Chem. Eng. Data.*

- 339 58 (2013) 1516–1522. doi:10.1021/je301004f.
- 340 [47] C. James, B.A. Onarinde, S.J. James, The Use and Performance of Household Refrigerators :
341 A Review, 16 (2017) 160–179. doi:10.1111/1541-4337.12242.
- 342 [48] F. Di Francesco, N. Calisi, M. Creatini, B. Melai, P. Salvo, C. Chiappe, Water sorption by
343 anhydrous ionic liquids, *Green Chem.* (2011). doi:10.1039/c1gc15080d.
- 344 [49] J.G. Huddleston, A.E. Visser, W.M. Reichert, H.D. Willauer, G.A. Broker, R.D. Rogers,
345 Characterization and comparison of hydrophilic and hydrophobic room temperature ionic
346 liquids incorporating the imidazolium cation, *Green Chem.* 3 (2001) 156–164.
347 doi:10.1039/b103275p.
- 348