

Title: Studies on computational molecular interaction between SARS-CoV-2 main protease and natural products

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Abstract

Our objective was to identify the molecule which can inhibit SARS-CoV-2 main protease and can be easily procured or available as a common household food ingredient. For this objective, natural products may provide such molecules and can supplement the current custom chemical synthesis based drug discovery approach. A combination of docking approaches, scoring functions, molecular dynamic simulation, and literature mining have been employed to screen readily available natural products (unique 27256 chemical entities, 598435 unique compounds), which can inhibit the SARS-CoV-2 main protease. Theaflavin digallate, a major constituent of black tea, has been observed to be as three top hits after the virtual screening of 598435 unique conformations. The main protease-theaflavin digallate complex observed to be in the metastable stage and interact with critical active site residues of the main protease during molecular dynamics simulation for 200 ns. *Invitro* evidence on main protease inhibition of 2003 SARS-CoV by theaflavin digallate is available in the scientific literature. As evident by the dynamics of intermolecular interactions, theaflavin digallate, forms approximately three hydrogen bonds with Glu166 of main protease, mostly through hydroxyl groups in the benzene ring of benzo(7) annulen-6-one. Glu166 is the most critical amino acid for main protease dimerization, which in turn, is necessary for catalytic activity. We have employed chloroquine and epigallocatechin gallate (green tea component) as a control set. Based on computational molecular interaction and data available in scientific literature, theaflavin digallate can inhibit the main protease of SARS-CoV-2.

Introduction

The outbreak of SARS-CoV-2 caused an unprecedented event to the world in the recent history of humanity. Most of the affected countries recommended to remain in home isolation, and cities have been shifted to lockdown mode. Natural products offer a wide range of opportunities in this scenario as it provides a diverse chemical scaffold which cannot be represented using conventional chemical libraries. Natural products can have known side effects/pharmacodynamics/pharmacokinetic properties as natural products may have been used historically for several indications. Natural products are chiral rich molecules and hence offer many stereoisomers whose biological exploration might be limited using conventional wet experimental techniques. Furthermore, natural products provide the opportunity to harness the chemical product, which can have resulted due to long evolutionary interaction. The SARS-CoV-2 main protease is necessary for the processing of viral polyproteins. Inhibition of the main protease can block viral replication. Hence, we have used computational approaches and scientific literature mining to identify the molecule, which can have the potential to inhibit SARS-CoV-2 main protease.

Methodology

Structural coordinates of easily available natural products in SDF file formats were downloaded from ZINC (Sterling and Irwin 2015). Chloroquine and epigallocatechin gallate (green tea component) was considered as control drugs. Ligand preparation was performed using the LigPrep module (Schrödinger Release 2018-1: LigPrep, Schrödinger, LLC, New York, NY, n.d.). The force field OPLS 2003e was used for the generation of three-dimensional conformation of natural products. Natural products were desalted and neutralized. Ionization and tautomeric states were prepared at neutral pH using Epik (Greenwood et al. 2010). PDB ID 6ILU7 was used as SARS-CoV-2 main protease. Hydrogen atoms were added and side chains were optimized using prepwizard utility of Schrodinger suite. Ionizable groups at pH 7 were predicted using PROPKA and hydrogen bonds were optimized using ProtAssign utility (Olsson et al. 2011). The structure was minimized with only restrains on heavy atoms using impref utility. The docking grid files are centered around Cys145 and His41.

The protein was docked with a virtual screen workflow combining glide module with high throughput virtual screening (Glide HTVS), single-precision (Glide SP), and extra precision (Glide XP). Glide XP has more extensive conformational sampling such as anchor-and-grow methodology and more stringent scoring function such as high penalty score for receptor-ligand noncomplementary than Glide SP. The compounds were ranked on the basis of glide gscore or docking score. The binding energy was estimated

using Prime MMGBSA for the ligand-receptor complex derived after Glide XP docking. The VSGB2 was used as an implicit solvation model (Li et al. 2011). The complex was also evaluated according to MMGBSA dG_{bind} score where the $dG_{bind} = E_{complex(minimized)} - E_{ligand(minimized)} - E_{receptor(minimized)}$. The molecular dynamic simulation of the top-scoring complex after the Glide XP stage was performed using the Desmond molecular dynamics package and OPLS 2005 force field. The solvation model used for molecular dynamics simulation was TIP3P. 10 Angstrom orthorhombic box has been used as boundary conditions. The charge was neutralized by adding Na⁺ ions and 0.15 M NaCl was added as a salt. Simulations were run for 200 ns in NPT conditions and at 300 K at 1.01 bars.

Results and Discussion

We have used unique natural compounds that are reported to be easily available on the ZINC database. The possible isomeric/ionization/minimized conformation from this set resulted in a total of 598435 distinct ligands. We have used Virtual Screen Workflow default settings, i.e., a combination of HTVS, SP, XP, and prime-MMGBSA. However, we have additionally employed enhanced sampling of ligand conformation and calculation of XP descriptors. Data on the top 10 conformations is presented in table 1. Ligands were sorted according to respective docking scores. 2-D structures of the top 4 compounds observed as top 10 hits are represented in Figure 1. The top 3 hits are from ZINC ID ZINC000195838435, which corresponds to theaflavin digallate (TG) and having 100 percent similarity with ChEMBL IDs, ChEMBL1451483, ChEMBL402609, and ChEMBL434864. Theaflavin digallate has chromane and benzo(7)annulen-6-one as a chemical scaffold.

The exact chemical composition of tea is complex. However, it has mainly two classes of flavonoids/polyphenols viz. catechins and theaflavins. Catechins are the major component of green tea, whereas theaflavins are the major constituent of black tea. The major difference between green tea and black tea is the step of fermentation/oxidation. Green tea does not require fermentation, whereas black tea requires the complete step of fermentation. Theaflavins, as identified in this study, are the result of this oxidation process, and hence black tea contains more theaflavins as compared to green tea.

Using the similarity ensemble approach, TG appears to be in a protease inhibitor target class (Keiser et al. 2007). In the study of screening a natural products library against the 2003 SARS-CoV virus main protease, TG has a half-maximal inhibitory concentration of 7 μ M against the main protease (Chen et al. 2005). Epigallocatechin gallate has a similar structure, the same natural source (tea), and known to have similar antioxidant properties as TG. However, epigallocatechin gallate did not inhibit the main protease

enzymatic activity which reflects the restricted stereo-selectivity of main protease. The other polyphenolic compounds of tea, such as epicatechin, epicatechin gallate, epigallocatechin, theophylline, and caffeine have also shown no inhibitory effect. Similarly, the crude extract of black tea has shown more enzymatic inhibition than the crude extract of green tea (Chen et al. 2005).

As molecular docking can have inherent accuracies, we have included two controls in our studies. The first control was epigallocatechin gallate (ZINC ID ZINC000003978478), the most abundant catechin in green tea, which has a similar structure, source, and activity as of TG. The second control was chloroquine which is known to inhibit SARS-CoV-2. However, chloroquine has a different source, activity and structure (Vincent et al. 2005; Wang et al. 2020). Based on scoring functions, such as docking scores, energy score, XP Gscore, glide gscore, TG observed to be a dominant inhibitor with docking score -15.263 as compared to epigallocatechin gallate for which docking scored varied from -9.401 to -7.541. The docking score of chloroquine was -4.477 to -4.300 and hence it can be inferred that chloroquine may not inhibit SARS-COV-2 by targeting main protease. The molecular interactions of TG with main protease have been presented in figure 2A-C and for control set as figure 2K-2L which reflects TG has dominant interaction with main protease as compared to epigallocatechin gallate/chloroquine.

Whether the addition of milk influences the phenolic component of tea is not extensively delineated (Rashidinejad et al. 2017). In an interventional clinical study on 9 healthy male volunteers, infusion time has shown to increase the concentration of total phenolic concentration, and phenolic concentration was unaffected by the addition of milk (Kyle et al. 2007). In Swiss mice, antioxidative effects have been shown to be diminished by the addition of milk, sugar, and honey and the two-hour dose is recommended for health benefits (Korir et al. 2014)

The molecular interaction of TG with main protease has been evaluated by molecular dynamic simulations. The total number of amino acids in main protease (PDB ID 6ILU7) was 308 and having a charge of -4. Four sodium ions were included to neutralize the system. The molecular dynamics has been performed in NPT at 300K. The total molecular system has 36432 atoms. The protein and ligand were simulated for 200 ns. During the simulation, protein fluctuated around 1.8 Å (Figure 3A left axis) whereas the ligand was fluctuated at around 4 Å (Figure 3A right axis) when superimposed to the docked protein at 0 ns. The protein complex was observed to be achieved a metastable stage throughout the simulation of 200 ns (Figure 3a). A movie of 1000 frames during 200 ns simulation was also available as supplementary data and at YouTube link <https://www.youtube.com/watch?v=yYEwgQymsO4>.

TG approximately makes three hydrogen bonds, either directly or water-mediated with glutamate 166 throughout the simulation of 200 ns. Glu166 of each protomer interacts with “N-finger” of another protomer for dimerizations which is required for the catalytic activity of main protease (Zhang et al. 2020). There are at least 13 residues (Thr 26, His 41, Cys 44, Ser 46, Glu 47, Met 49, Asn 142, Gly143, Cys 145, His 164, Glu 166, Asp 187, Gln 189, Thr 190) in the main protease which interacts with TG for more than 100 ns through hydrogen bonds, hydrophobic, water bridges and ionic interaction (Figure 3B). The catalytic residue Cys 145 predominantly interacts with water bridges, whereas His 41 interacts mainly through hydrophobic interaction (Figure 3B and 3C). The TG makes approximately 15 contacts during the whole simulation (Figure 3 D)

Tea is the second most-consumed beverage after the water. An exact wet experiment or clinical studies providing the evidence on TG efficacy are difficult to conduct because of the huge number of variants available for black tea formulation. However, an observational clinical study and in-vitro experiments with SARS-CoV-2 main protease can provide conclusive evidences.

Acknowledgment

MM is the recipient of senior research associate fellowship under the CSIR-Pool Scientist Scheme. The author acknowledges the computational facility and overall mentorship provided by Prof. Naidu Subbarao, School of computational and integrative sciences, JNU, New Delhi.

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ZINC_id	Energy	docking score	XP GScore	glide gscore	glide energy	glide emodel
ZINC00019583845 (Theaflavin Digallate)	66.45663	-15.263	-15.2929	-15.2929	-80.7874	-114.878
ZINC00019583845 (Theaflavin Digallate)	66.45663	-14.9138	-14.9437	-14.9437	-76.8608	-109.745
ZINC00019583845 (Theaflavin Digallate)	66.45663	-14.4314	-14.4613	-14.4613	-72.5158	-111.75
ZINC33861449	34.39056	-14.4268	-14.5059	-14.5059	-65.6439	-106.16
ZINC00008564535	33.10011	-14.24	-14.24	-14.24	-83.6571	-106.964
ZINC00008564535	30.89951	-14.2216	-14.2216	-14.2216	-72.4829	-105.12
ZINC00008564535	33.10011	-14.2128	-14.2128	-14.2128	-83.4784	-112.139
ZINC67903526	39.81764	-14.1658	-14.1944	-14.1944	-73.0363	-116.279
ZINC00008564535	33.10011	-14.1375	-14.1375	-14.1375	-82.697	-111.94
ZINC00008564535	33.10011	-14.0532	-14.0532	-14.0532	-81.513	-109.506

Table 1: The top 10 ligands after the virtual screening of 54386 unique compounds (909326 unique conformations). The ligands are sorted according to their docking score.