

1 Protoenzymes: The Case of Hyperbranched Polymer- 2 Scaffolded ZnS Nanocrystals

3 Irena Mamajanov,^{1,*} Melina Caudan,¹ Tony Z. Jia^{1,2}

4 ¹ Earth Life Science Institute, Tokyo Institute of Technology, Meguro, Tokyo 152-8550, Japan.

5 ² Blue Marble Institute for Science, 1001 4th Ave, Suite 3201, Seattle, WA 98154, USA.

6 * Correspondence: irena.mamajanov@elsi.jp

7 **Abstract:** Enzymes could be described as small-molecule, metal, or cluster catalysts augmented by
8 biopolymeric scaffolds. It is conceivable that early in chemical evolution, ancestral enzymes opted
9 for simpler, easier to assemble scaffolds. Herein, we describe such possible protoenzymes:
10 hyperbranched polymer-scaffolded metal-sulfide nanocrystals. Hyperbranched polyethyleneimine
11 (HyPEI) and glycerol citrate polymer-supported ZnS nanocrystals (NCs) are formed in a simple,
12 abiotically plausible process. Transmission electron microscopy (TEM) analyses of HyPEI-
13 supported NCs reveal spherical particles with an average size of 10nm that undergo only a modest
14 aggregation over a 14-day incubation. The polymer-supported ZnS NCs are shown to possess a high
15 photocatalytic activity in an eosin B photodegradation assay, making them an attractive model for
16 the study of the origin of life under the “Zn world” theory dominated by a photocatalytic proto-
17 metabolic redox reaction network. The catalyst, however, could be easily adapted to apply broadly
18 to different protoenzymatic systems.

19 **Keywords:** protoenzyme; hyperbranched polymers; photocatalytic nanoparticles; polymer-
20 supported nanoparticles; metal-sulfide clusters

21
22

23 1. Introduction

24 Metabolism is one of the defining characteristics of life. The primary purpose of metabolism is
25 the efficient utilization of external sources of energy to fuel cellular processes, such as growth and
26 replication [1]. Enzymes, highly selective and efficient biocatalysts, govern metabolism. Enzymes
27 offer a selective advantage to some reactions by enhancing their rate up-to 10²³-fold [2,3], unmatched
28 by other types of catalysts, effectively shaping metabolic transformations. This level of control and
29 order sets metabolism apart from any other set of chemical reactions. The crucial role of enzymes in
30 life has prompted some to argue that catalytic enzyme precursors, or protoenzymes, played a critical
31 role in the emergence of life [4,5].

32 Nearly all enzymes are based upon globular proteins, save for some RNA strands that have
33 catalytic activity (ribozymes) [1]. While the discovery of the ribozyme [6,7] has become the basis of
34 the RNA world origins of life theory [8,9], the question of how long, functional biopolymers were
35 spontaneously formed remains unsolved. Previous studies have thus considered potentially more
36 prebiotically reasonable transition metal complexes [10], small molecules [11], and mineral surfaces
37 [12, 13] as primitive protoenzymes. While metal cations and small molecules catalyze chemical
38 reactions by lowering the activation energy or altering reaction pathways, macromolecular enzymes
39 offer an additional advantage of selective binding, orienting, and enclosing reactants within a
40 modulated microenvironment [14]. In the prebiotic world, in the absence of long coded proteins or
41 RNA, mineral surface catalysis could have performed some catalytic functions. Encapsulation of
42 organic molecules between mica sheets has been proposed to trigger protoenzymatic processes [15]
43 and clay minerals have been shown to facilitate the polymerization of activated nucleotides,
44 presumably by orienting the reactants [16]. The orientation of glycine molecules on oxide mineral
45 surfaces has also been shown to affect its reactivity towards polymerization [17].

46 In the synthetic community, the application of enzymatic systems to modern synthetic processes
47 has become a desirable goal [18]. The high cost of enzyme isolation and enzyme incompatibility with
48 synthesis conditions outside of physiological has prompted studies of non-biopolymer-based
49 enzyme mimics. Approaches to the design of artificial enzymes range from stripping the cofactors of
50 the biopolymeric scaffold, synthesis of supramolecular active sites, surveying enzyme-mimetic
51 properties of various nanoparticles, and investigation of non-biological polymeric scaffolds [19]. In
52 the context of prebiotic chemistry, it is conceivable that the first enzymes may not have been purely
53 or entirely protein- or RNA-based. Herein, we describe an effort to combine the principles of artificial
54 enzyme construction discovered by synthetic chemists to gain new insights into the question of the
55 provenance of enzymes.

56 Catalytic nanoparticles are an attractive avenue to explore in the context of protoenzymes. The
57 catalytic properties of nanoparticles, in particular, arise from their large surface area compared to the
58 nanoparticle's total number of atoms [20]. Many nanoparticle catalysts have been shown to catalyze
59 enzymatic reactions to earn the term "nanozyme" [21]. Nanomaterials are usually thought of as
60 synthetic or anthropogenic; however, they can form through natural abiotic processes and could have
61 been present on prebiotic Earth. All mineral formation processes undergo a sometimes persistent
62 nanophase stage during formation. For example, volcanic ash clouds contain polydisperse particles
63 that range from 100 to 200 nm in size and are primarily composed of silicate and iron compounds
64 [22]. In geological processes, nanoparticles can be generated through mineral weathering or by
65 mechanical grinding associated with earthquake-generating faults in Earth's crust [23]. Could
66 chemical evolution harness the catalytic capacity of natural nanomaterials? To answer this question,
67 we investigated the potential role of metal-sulfide nanocrystals (NCs) as possible precursors to metal
68 sulfide clusters found in some modern enzymes. Metal sulfide deposits near deep-sea hydrothermal
69 vents have become the basis of the iron-sulfur world theory [24]. The model suggests that these
70 minerals would catalyze complex sequences of reactions, driven by the energy from the vents,
71 eventually leading to life. In the subaerial "Zinc world" scenario [25], life emergence would have been
72 driven by light energy harnessed by photoactive semiconductive zinc sulfide (ZnS) minerals. Under
73 "Zinc world" assumptions, the geochemical formation of long-lived ZnS nanoparticles would extend
74 the catalytic capacity of the material due to the significantly increased specific surface area of the
75 catalyst and the exciton confinement effects [26]. NCs, however, tend to spontaneously aggregate;
76 therefore, it is a common practice to include polymeric supports [27] or other capping agents [28,29]
77 in NC formulations to decrease particle overgrowth and aggregation. As a plausible prebiotic
78 polymeric support candidate, we introduce hyperbranched polymers. Hyperbranched polymers [30]
79 and dendrimers [31,32] have been long considered for biomimetic catalysts in synthetic applications.
80 Our previous studies have demonstrated the plausibly prebiotic synthesis of hyperbranched
81 polyesters from citric acid and glycerol [33-35] and the ability of hyperbranched polyesters to catalyze
82 the Kemp elimination reaction [36]. Here, we report on the prebiotically plausible formation of
83 photocatalytic hyperbranched polymer-scaffolded ZnS NCs.

84 2. Materials and Methods

85 All chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA) and used without
86 further purification.

87 *Synthesis of hyperbranched polyethyleneimine (HyPEI)-supported ZnS nanocrystals (NCs):*

88 To produce 10mg of supported ZnS NCs, a solution of 0.4g HyPEI (124.9ml, pH=6) was titrated
89 with 2.56ml of 40mM ZnCl₂ solution and magnetically stirred for 1 hour. 2.56mL of a 40mmol solution
90 of Na₂S was then added dropwise and was allowed to stir for 1 hr magnetically.

91 *Synthesis of glycerol citrate polyesters:*

92 A polyesterification was conducted, starting with 10mL of an aqueous solution containing
93 330mM citric acid and 660mM glycerol. Some formulations contained also 165mM CoCl₂ or ZnCl₂.
94 The samples were incubated, uncovered, at 85°C for 72 h. The resulting gel-like product was
95 redissolved and dialyzed through a 500-1000Da cutoff Float-A-Lyzer membrane (Repligen, Waltham,
96 MA, USA) and lyophilized.

97 *Measurement of the divalent cation loading capacity of the polymers:* 100mg of glycerol citrate
98 polyester (formulation without added cations), COOH-terminated oligo- 2,2-
99 bis(hydroxymethyl)propionic acid dendrimer, trimethylol propane core, generation 1, C₄₅H₆₂O₃₀
100 (Sigma-Aldrich, 806099), OH-terminated oligo- 2,2-bis(hydroxymethyl)propionic acid dendrimer,
101 trimethylol propane core, generation 1, C₂₁H₃₈O₁₂(Sigma-Aldrich, 805920) polyester dendrimers, and
102 HyPEI was equilibrated overnight in 1mL of a 165mM solution of CoCl₂. The step was omitted for
103 the glycerol citrate polyesters prepared in the presence of CoCl₂. The solutions were then dialyzed
104 through a 500-1000Da cutoff Float-A-Lyzer membrane and lyophilized. The content of Co²⁺ per unit
105 weight of the resulting polymer was determined spectroscopically utilizing a JASCO (Hachioji,
106 Tokyo, Japan) V-670 UV-vis spectrometer.

107 *Synthesis of glycerol citrate polyester-supported ZnS NCs:* The loading capacity towards the
108 absorption of Zn²⁺ was estimated to be the same or similar, as in the case of Co²⁺ ions. An excess of
109 the theoretical amount of the Zn²⁺-bearing polyester was used. 0.8g of Zn²⁺ -bearing polyester was
110 dissolved in 27.4mL of water. The pH of the solution was adjusted to pH=6 with NaOH. 2.56mL of a
111 40mmol solution of Na₂S was then added dropwise and was allowed to stir for 1 hr magnetically.

112 *Mass spectrometry:* MALDI-MS spectra were collected on an ultrafleXtreme Bruker Daltonics
113 MALDI-TOF-MS (Bruker Corporation, Billerica, MA, USA) in positive ion mode. External mass
114 calibration was conducted using standard peptide mixtures. The sample preparation matrix, 90:10
115 mixture of 2,5-dihydroxybenzoic acid and 2-hydroxy-5-methoxy benzoic acid (SDHB), was dissolved
116 in deionized water. Subsequently, the freeze-dried samples and the matrix were mixed at a 1:10 [v/v]
117 ratio in advance, and then the mixture was applied to the plate before analysis.

118 *Transmission Electron Microscopy (TEM):* Transmission Electron Microscopy and Field-emission
119 (FE)-TEM imaging were performed at the Tokyo Institute of Technology Materials Analysis Division
120 Technical Department in Meguro-ku, Tokyo, with the assistance of Akira Genseki and Ryohei
121 Kikuchi. Samples were prepared by placing a droplet of ZnS nanoparticle solution (See above for
122 synthesis method) on a Collodion Film COL-C15 copper grid (Okenshoji Co., Ltd., Chuo-ku, Tokyo,
123 Japan) and wicking away the liquid to deposit a thin layer onto the grid. The grid was allowed to dry
124 at room temperature for 2 days before imaging. After confirmation using a Hitachi (Chiyoda-ku,
125 Tokyo, Japan) H7650 Zero A TEM (100 kV), high-resolution imaging was performed at room
126 temperature on a JEOL (Akishima, Tokyo, Japan) JEM-2010F FE-TEM at 200 kV with the following
127 specifications: a ZrO/W(100) Schottky cathode, ultra-resolution pole piece, Gatan (Sarasota, Florida,
128 USA) Digiscan System Model 688, Gatan MSC 794 CCD Camera, and an EDAX (Minato-ku, Tokyo,
129 Japan) Genesis energy dispersive X-ray spectrometer.

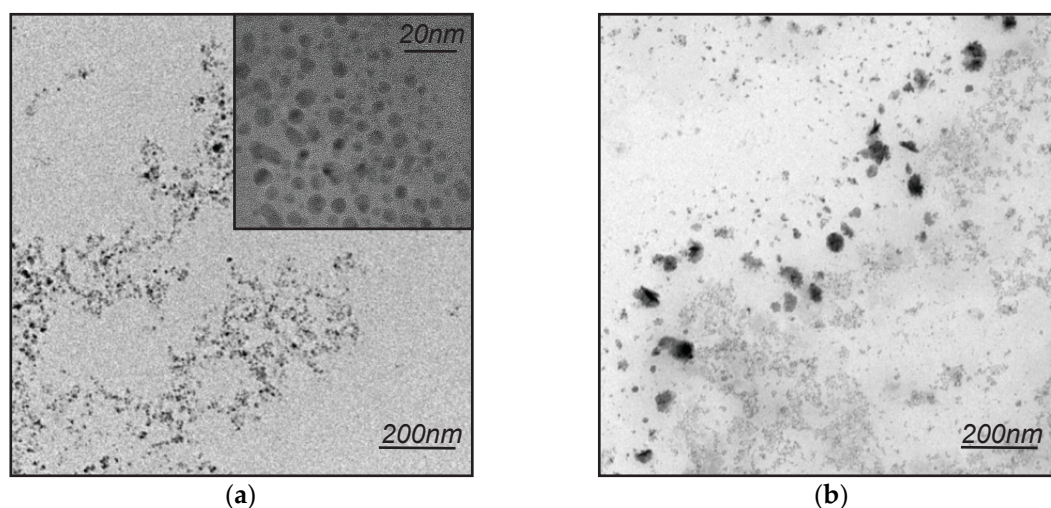
130 *Photocatalytic activity measurement:* A jacketed pyrex flask (capacity ca. 40mL) was used as the
131 photoreactor vessel. For a typical experiment, a solution adjusted to pH=6 by NaOH containing eosin
132 B (5.0 x10⁻⁵M, 30mL) and polymer-supported ZnS NC catalysts (10mg of ZnS, corresponding amount
133 of polymer) was magnetically stirred in the dark for 30 min to reach the adsorption equilibrium of
134 eosin B with the catalyst and then exposed to light from a 100W high-pressure mercury lamp (Handy
135 100, Mizuka Planning, Amagasaki, Hyogo, Japan). The temperature of the solution was maintained
136 constantly by circulating ice water through the flask's jacket. An aliquot was collected every 15 min;
137 UV/Vis absorption spectra of each aliquot were recorded on a JASCO V-670 UV-vis spectrometer.

138 3. Results

139 The procedure for the preparation of hyperbranched polyethyleneimine (HyPEI) was adapted
140 from a protocol described by Hassan and Ali [37]. The protocol devised by Hassan and Ali utilized
141 high molecular weight HyPEI (M_w=25,000 – 50,000); however, we used HyPEI of a low molecular
142 weight HyPEI (M_w=800Da) comparable to that of hyperbranched polyesters synthesized under
143 prebiotically plausible conditions [33,34] In our experiments, when Na₂S solution was titrated into a
144 solution of ZnCl₂ at neutral pH, the clear solution quickly turned cloudy with a white precipitate,
145 presumably, crystalline zinc sulfide (ZnS), settling within an hour. However, when Na₂S solution was
146 titrated into a ZnCl₂ solution in the presence of hyperbranched polyethyleneimine (at neutral pH, the
147 resulting solution remained unclouded for at least 14 days suggesting that the ZnS product remained

148 soluble. Figure 1A shows a low-magnification TEM (transmission emission microscope) micrograph
 149 of a freshly prepared ZnS/HyPEI sample, which indicates that the sample contains many
 150 nanoparticles in a clustered pattern, possibly owing to the hyperbranched polymer matrix. The high-
 151 magnification FE (field-emission)-TEM micrograph in Figure 1A (inset) reveals spherical particles
 152 with an average size under 10nm. The low-magnification TEM micrograph in Figure 1B shows the
 153 presence of aggregated particles (~50nm in diameter) in a solution of ZnS/HyPEI that was kept at
 154 room temperature for 14 days, confirming our previous observation that the nanoparticular ZnS
 155 remained stable in solution for this time period.

156



157 **Figure 1.** TEM images of HyPEI-supported ZnS nanocrystals. **(a)** Low magnification image of a freshly
 158 prepared sample; inset: high magnification FE-TEM micrograph of a freshly prepared sample. **(b)**
 159 Low magnification image of a sample aged for 14 days.

160

161 HyPEI contains abundant primary, secondary, and tertiary amine groups, and has a strong
 162 affinity towards the complexation of transition metals [38]. HyPEI, consequently, has been used in
 163 water treatment and as a support for nanoparticles [37,39], although it is not very prebiotically
 164 plausible. We, therefore, next considered hyperbranched polyesters that could plausibly form in
 165 geochemical settings [33-35]. We assessed the hyperbranched polyesters formed upon the reaction
 166 between citric acid and glycerol, as well as commercially available oligo-2,2-
 167 bis(hydroxymethyl)propionic acid dendrimers terminated with either carboxylic acid or alcohol
 168 functional groups. Neither commercial polyester dendrimers nor the glycerol citrate polymer has
 169 exhibited high affinity towards transition metal binding (Table 1). Interestingly, the citric acid
 170 glycerol polymers formed in the presence of CoCl_2 retained a significant amount of Co^{2+} through
 171 dialysis (see the Materials and Methods section). We have previously shown that the presence of
 172 divalent cations during citric acid and glycerol polyesterification alters the structure and the size of
 173 the resulting polymer [33]. The cation-containing formulations resulted in shorter polymers that
 174 incorporated more citric acid moieties compared to neat solution formulations. Stoichiometrically, the
 175 increased number of citric acid moieties, in turn, increases the number of unreacted carboxylic groups
 176 changing the metal affinity. The response of the polyesterification system to the presence of divalent
 177 cations by adjusting the polymer product structure to accommodate cation absorption is an example
 178 of rudimentary molecular imprinting, a primitive mechanism of molecular memory encoding
 179 suggested to be prevalent in prebiotic world prior to the onset of nucleic acid replication [40]. Thus,
 180 to incorporate a prebiotically plausible polymer that has molecular imprinting ability, we synthesized
 181 a polyester by heating at 85°C a dry mixture of citric acid, glycerol, and ZnCl_2 (1:2:0.5, mole ratios).
 182 The mass spectral analysis of the polymer (Figure S1) reveals a heterogeneous mixture of polymeric
 183 products up to ~1400Da. The resulting polymer solution was titrated with Na_2S to form polymer-
 184 scaffolded ZnS NCs.

185
186**Table 1.** Loading Capacity of Co²⁺ of hyperbranched polyesters and HyPEI.

Polymer	Co ²⁺ Loading Capacity ($\mu\text{mol Co}^{2+}/\text{g polymer}$)
Hyperbranched Polyethyleneimine M _w =800	736
COOH-terminated oligo- 2,2- bis(hydroxymethyl)propionic acid dendrimer, trimethylol propane core, generation 1, C ₄₅ H ₆₂ O ₃₀ (bis- MPA-COOH)	4.5
OH-terminated oligo- 2,2- bis(hydroxymethyl)propionic acid dendrimer, trimethylol propane core, generation 1, C ₂₁ H ₃₈ O ₁₂ (bis- MPA-OH)	1.6
Citric acid – glycerol polymer, prepared in neat	non detectable
Citric acid – glycerol, prepared in the presence of CoCl ₂	542

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

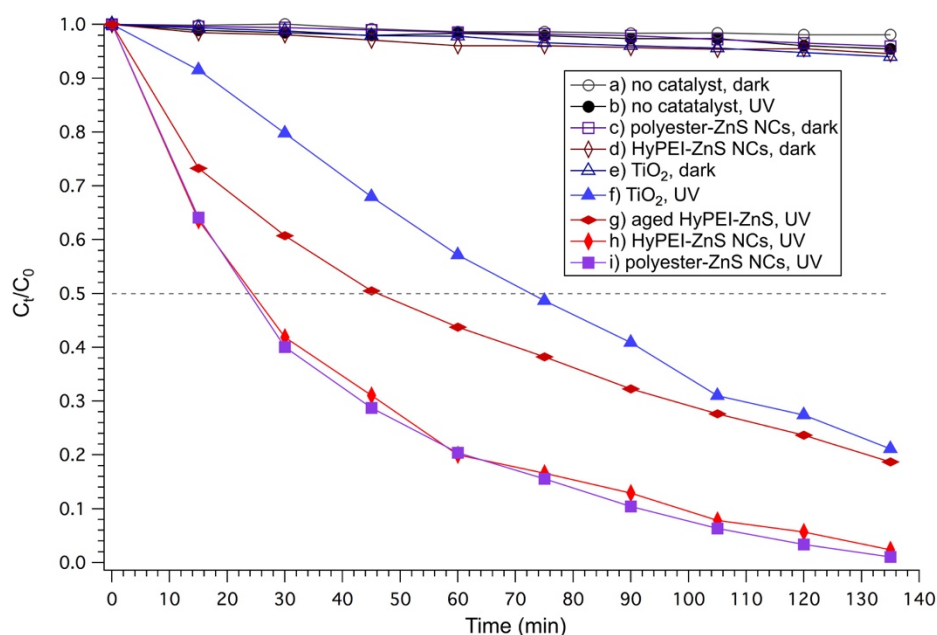
216

217

218

To demonstrate the potential applicability of the polymer-supported ZnS NCs to the prebiotic chemistry in "Zinc world" settings, we have followed the analytical procedure described by Hu et al. [41]. We investigated the photocatalytic activity of the polymer-supported ZnS NCs relative to those of bulk ZnS and the commercial photocatalyst TiO₂ (21nm), with the photocatalytic degradation of eosin B as a test reaction. We have chosen the characteristic absorption of eosin B at 520 nm as the monitored parameter for the photocatalytic degradation process. 30 ml of 5.0×10⁻⁵M solution of eosin B was subjected to incubation with periodic sampling under the following conditions (Figures 2, S2): a) without a catalyst, in the dark b) without a catalyst, under UV irradiation, c) with polyester-supported ZnS NCs (10mg of NCs), in the dark, d) with HyPEI supported ZnS NCs, in the dark, e) with 21nm TiO₂ nanoparticles, in the dark f) with 21nm TiO₂ nanoparticles, under UV, g) with HyPEI-supported ZnS NCs, aged at room temperature for 14 days, under UV h) with freshly prepared HyPEI-supported ZnS NCs, under UV, i) with polyester-supported ZnS NCs, under UV, j) with unsupported ZnS₂, k) with monomeric citric acid, glycerol, ZnCl₂, and Na₂S, under UV. When appropriate, 10 mg of TiO₂ nanoparticles or ZnS NCs was used. Under the experimental conditions from (a) to (e), the photocatalytic effect on the solution degradation without catalysts but under exposure to UV light is almost the same as that with catalyst but no exposure to UV light. For example, a slight decrease in the concentration of eosin B was detected in the absence of any catalyst (Figure 2, curve b). Exposure to UV light for 135 min resulted in only 5-6% degradation of the dye with and without the catalyst. The curves measured under the experimental conditions (f) to (i), those containing a catalyst and with exposure to UV light, are indicative of a significantly higher catalytic activity. For example, the fresh HyPEI-supported ZnS NC (Figure 2, curve h) sample was completely decolorized after 135 min of exposure to UV light with a half-life of ~24 mins. The aged HyPEI-supported ZnS NC (Figure 2, curve g), and commercial TiO₂ particle (Figure 2, curve f) samples exhibit a lower photocatalytic activity with half-lives of ~45 and 72 min, respectively. This difference in the photocatalytic activity between fresh HyPEI-ZnS NC sample and aged HyPEI ZnS NC and TiO₂ samples can be explained by the smaller diameter of the ZnS NC (~10nm of the fresh HyPEI-ZnS NCs vs. 20nm of TiO₂ particles vs. ~50nm of the aged HyPEI-ZnS NCs), hence resulting in a larger catalytic surface area. The polyester-supported ZnS NCs activity (Figure 2, curve i) closely resembles that of the fresh HyPEI-supported ZnS NCs (Figure 2, curve h). Unsupported ZnS undergoes quick aggregation and precipitation under the irradiation (Figures S2, S3). In this sample (Figure S2, curve

219 j), the solution was rapidly decolorized. However, the dye was adsorbed and preserved on the surface
 220 of the precipitated ZnS particles suggesting that decolorization derived from the solid-liquid phase
 221 partitioning rather than photodegradation (Figure S3). The photocatalytic activity of polyester-
 222 supported ZnS NCs was further compared to that of a solution containing *unreacted* citric acid and
 223 glycerol as well as ZnCl₂ and Na₂S (Figure S2, curve k). Interestingly, ZnS did precipitate out of the
 224 solution containing citric acid and glycerol as it did in a neat solution. Moreover, citric acid, glycerol,
 225 ZnCl₂, and Na₂S solution did not exhibit a detectable photocatalytic activity. It is conceivable that the
 226 process of chelation of zinc cations by the unreacted citric acid in the sample (i) interferes with the
 227 formation of ZnS particulates.



228

229 **Figure 2.** Time-lapse measurement of photodegradation of eosin B ($5.0 \times 10^{-5} \text{M}$, 30mL) under different
 230 conditions: a) without a catalyst, in the dark b) without a catalyst, under UV irradiation, c) with
 231 polyester-supported ZnS NCs (10mg of NCs), in the dark, d) with HyPEI-supported ZnS NCs, in the
 232 dark, e) with 21nm TiO₂ nanoparticles, in the dark f) with 21nm TiO₂ nanoparticles, under UV, g) with
 233 HyPEI-supported ZnS NCs, aged at room temperature for 14 days, under UV h) with freshly prepared
 234 HyPEI-supported ZnS NCs, under UV, i) with polyester-supported ZnS NCs, under UV. When
 235 appropriate, 10 mg of TiO₂ nanoparticles or ZnS NCs was used. The normalized concentration (C_t/C_0)
 236 was derived from the UV absorbance values at 520nm.

237 4. Discussion

238 Our results show that HyPEI- or glycerol citrate polyester-supported ZnS NCs make excellent
 239 photocatalysts. Their straightforward formation makes them plausible agents at the early stages of
 240 chemical evolution. While we so far have only explored the catalyst and the processes relevant to the
 241 "Zinc World" scenario, the demonstrated synthesis of the catalytic complexes could be applicable in
 242 a variety of geological settings. Therefore, by considering different particles or cofactors, as well as
 243 different plausible polymeric scaffolds, one could extend the repertoire of similar catalysts to other
 244 chemical evolution models. With minimal modifications, the model could be adjusted to study the
 245 possible chemical evolution of FeS or MoS cluster bearing proteins, e.g., ferredoxins, hydrogenases,
 246 and nitrogenases [42]. Utilizing NCs rather than bulk surfaces as a model for possible cluster
 247 precursors permits the investigation of size-dependent properties. The concept of hyperbranched
 248 scaffolds could also be applicable broadly to the models of chemical evolution. In these models,
 249 cofactors other than metal sulfides, i.e., small molecules and cations could be used. Furthermore, in
 250 lieu of hyperbranched polyimides and polyester, other polymers could be utilized, i.e.,
 251 hyperbranched polyamidoamines [43] and polypeptides [44]. In addition to forming efficient

252 prebiotically plausible catalysts, the structure of cofactors scaffolded by globular polymers,
 253 superficially similar to contemporary enzymes, is intriguing in the context of chemical evolution as
 254 well. When considering enzyme-like prebiotic catalysts, the abundance of particular small-molecule
 255 cofactors, inorganic clusters, and cations are conceivable; however, functional high proteins and RNA
 256 molecules are unlikely to have been present at the early stages of chemical evolution. Several studies
 257 have tackled the prebiotic formation of peptide [45,46] and phosphodiester bonds [16,47]. These
 258 studies, however, so far, did not address the mechanisms controlling the primary structure of a
 259 biopolymer that is responsible for folding and function. The intrinsically globular hyperbranched
 260 polymers are a reasonable candidate for primitive protoenzymatic scaffolds.

261 In summary, we have presented a straightforward, plausibly prebiotic process of a spontaneous
 262 formation of stabilized polymer-supported nanoparticles. The structure of these complexes, a
 263 catalytic agent scaffolded by globular polymers, is superficially reminiscent of the enzymatic
 264 structure and therefore is a compelling model for the study of the chemical evolution of enzymes. An
 265 enzyme could be described as a catalytic agent augmented by a biopolymer scaffold. The
 266 hyperbranched polymer scaffold could have been the early primitive augmenting scaffold to be
 267 replaced with more sophisticated ones throughout chemical evolution. This primitive scaffold
 268 furthermore provides a means to study the aspects of small particle catalysis in prebiotic chemistry.
 269 Although herein, we have only explored the photocatalytic ZnS NC complex relevant to the "Zinc
 270 world" hypothesis of the origin of life, the model, however, can be easily extended to other
 271 protoenzymatic systems.
 272

273 **Supplementary Materials:** The following are available online at www.mdpi.com/xxx/s1, Figure S1: MALDI
 274 mass spectrum of the ZnCl₂-bearing glycerol citrate polyester, Figure S2: Time-lapse measurement of
 275 photodegradation of eosin B (5.0×10⁻⁵M, 30mL) under additional conditions, Figure S3: Visual progression of the
 276 eosin B degradation assay catalyzed by unsupported ZnS.

277 **Author Contributions:** Conceptualization, I.M.; methodology, I.M. and T.Z.J.; formal analysis, I.M., M.C. and
 278 T.Z.J.; investigation, I.M. and M.C.; resources, I.M. and T.Z.J.; writing—original draft preparation, I.M.; writing—
 279 review and editing, I.M., M.C. and T.Z.J.; visualization, I.M.; funding acquisition, I.M. and T.Z.J. All authors have
 280 read and agreed to the published version of the manuscript.

281 **Funding:** This research was supported by the World Premier International Research Center Initiative (WPI),
 282 Ministry of Education, Culture, Sports, Science and Technology, Japan, as well as Grant-in-Aid for Scientific
 283 Research from the Japan Society for the Promotion of Science Kakenhi Kiban C Grant 17K01943 (to I.M.) and
 284 Wakate Grant 18K14354 (to T.Z.J.), as well as a Tokyo Institute of Technology Seed Grant "Tane" 1798 (to T.Z.J.).

285 **Acknowledgments:** We would like to thank Ryohei Kikuchi and Akira Genseki at the Tokyo Institute of
 286 Technology Ookayama Materials Analysis Division, Technical Department for assistance in TEM images.

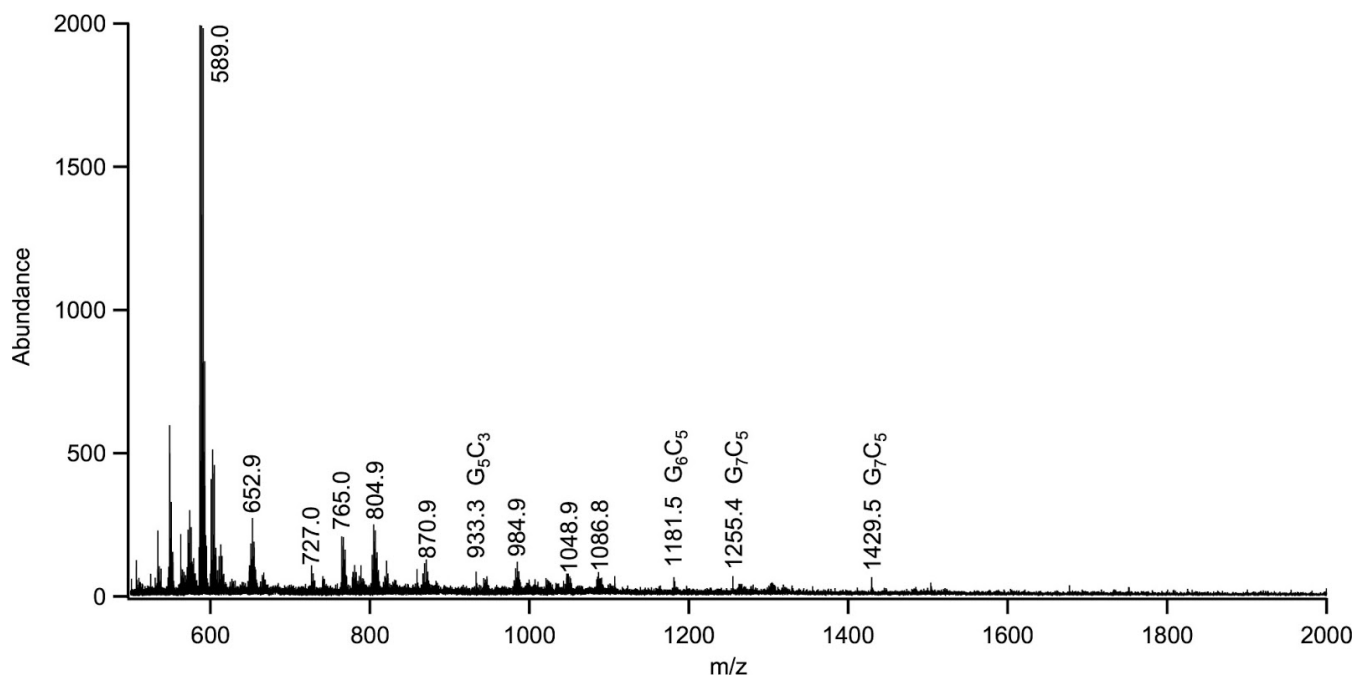
287 **Conflicts of Interest:** The authors declare no conflict of interest.

288 References

- 289 1. Campbell, M. K.; Farrell, S. O. *Biochemistry*; Cengage Learning, 2011.
- 290 2. Radzicka, A.; Wolfenden, R. A Proficient Enzyme. *Science* **1995**, *267* (5194), 90–93.
- 291 3. Richard, J. P.; Amyes, T. L.; Reyes, A. C. Orotidine 5'-Monophosphate Decarboxylase: Probing the Limits
 292 of the Possible for Enzyme Catalysis. *Acc. Chem. Res.* **2018**, *51* (4), 960–969.
- 293 4. Szöke, A.; Scott, W. G.; Hajdu, J. Catalysis, Evolution and Life. *FEBS Lett.* **2003**, *553* (1), 18–20.
- 294 5. Canepa, C. The Role of Catalysis on the Formation of an Active Proto-Enzyme in the Prebiotic Aqueous
 295 Environment. *Nat. Sci.* **2013**, *05* (05), 549–555.
- 296 6. Kruger, K.; Grabowski, P. J.; Zaug, A. J.; Sands, J.; Gottschling, D. E.; Cech, T. R. Self-Splicing RNA:
 297 Autoexcision and Autocyclization of the Ribosomal RNA Intervening Sequence of Tetrahymena. *Cell* **1982**,
 298 *31* (1), 147–157.
- 299 7. Guerrier-Takada, C.; Gardiner, K.; Marsh, T.; Pace, N.; Altman, S. The RNA Moiety of Ribonuclease P Is
 300 the Catalytic Subunit of the Enzyme. *Cell* **1983**, *35* (3, Part 2), 849–857.

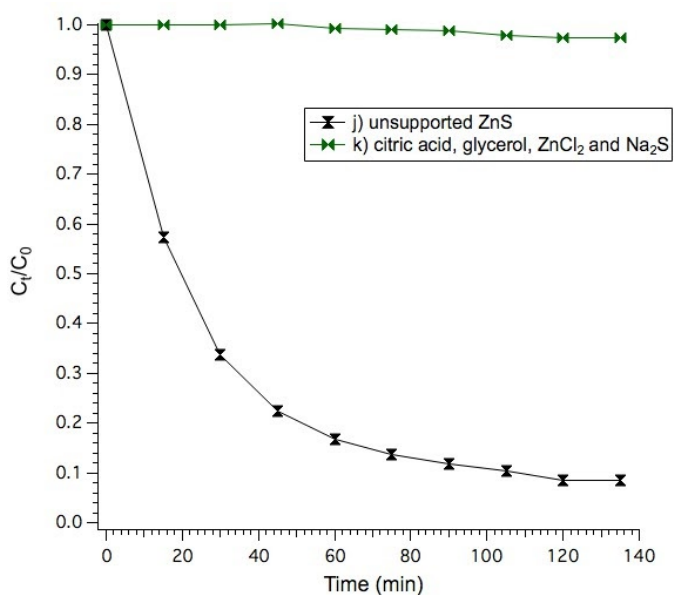
- 301 8. Cech, T. R. The RNA Worlds in Context. *Cold Spring Harb. Perspect. Biol.* **2012**, *4*(7), a006742.
- 302
- 303 9. Orgel, L. E. Some Consequences of the RNA World Hypothesis. *Orig. Life Evol. Biosph.* **2003**, *33* (2), 211–218.
- 304 10. Morowitz, H. J.; Srinivasan, V.; Smith, E. Ligand Field Theory and the Origin of Life as an Emergent Feature
- 305 of the Periodic Table of Elements. *Biol. Bull.* **2010**, *219* (1), 1–6.
- 306 11. Shapiro, R. Small Molecule Interactions Were Central to the Origin of Life. *Q. Rev. Biol.* **2006**, *81* (2), 105–
- 307 125.
- 308 12. Cleaves, H. J.; Scott, A. M.; Hill, F. C.; Leszczynski, J.; Sahai, N.; Hazen, R. Mineral–Organic Interfacial
- 309 Processes: Potential Roles in the Origins of Life. *Chem. Soc. Rev.* **2012**, *41* (16), 5502–5525.
- 310 13. Russell, M. J.; Barge, L. M.; Bhartia, R.; Bocanegra, D.; Bracher, P. J.; Branscomb, E.; Kidd, R.; McGlynn, S.;
- 311 Meier, D. H.; Nitschke, W.; Shibuya T.; Vance S., White L.; Kanik I. Drive to Life on Wet and Icy Worlds.
- 312 *Astrobiology* **2014**, *14* (4), 308–343
- 313 14. Bruice, T. C.; Benkovic, S. J. Chemical Basis for Enzyme Catalysis. *Biochemistry* **2000**, *39* (21), 6267–6274.
- 314 15. Hansma, H. G. Possible Origin of Life between Mica Sheets: Does Life Imitate Mica? *J. Biomol. Struct.*
- 315 *Dyn.* **2013**, *31* (8), 888–895.
- 316 16. Ferris, J. P.; Hill, A. R.; Liu, R.; Orgel, L. E. Synthesis of Long Prebiotic Oligomers on Mineral Surfaces.
- 317 *Nature* **1996**, *381* (6577), 59–61.
- 318 17. Kitadai, N.; Oonishi, H.; Umamoto, K.; Usui, T.; Fukushi, K.; Nakashima, S. Glycine Polymerization on
- 319 Oxide Minerals. *Orig. Life Evol. Biosphere* **2017**, *47* (2), 123–143.
- 320 18. Breslow, R. Biomimetic Chemistry and Artificial Enzymes: Catalysis by Design. *Acc. Chem. Res.* **1995**, *28* (3),
- 321 146–153.
- 322 19. Kuah, E.; Toh, S.; Yee, J.; Ma, Q.; Gao, Z. Enzyme Mimics: Advances and Applications. *Chem. – Eur. J.* **2016**,
- 323 *22* (25), 8404–8430.
- 324 20. Moshfegh, A. Z. Nanoparticle Catalysts. *J. Phys. Appl. Phys.* **2009**, *42* (23), 233001.
- 325 21. Liang, M.; Yan, X. Nanozymes: From New Concepts, Mechanisms, and Standards to Applications. *Acc.*
- 326 *Chem. Res.* **2019**, *52* (8), 2190–2200.
- 327 22. Griffin, S.; Masood, M. I.; Nasim, M. J.; Sarfraz, M.; Ebokaiwe, A. P.; Schäfer, K.-H.; Keck, C. M.; Jacob,
- 328 C. Natural Nanoparticles: A Particular Matter Inspired by Nature. *Antioxidants* **2017**, *7* (1), 3.
- 329 23. Hochella, M. F.; Lower, S. K.; Maurice, P. A.; Penn, R. L.; Sahai, N.; Sparks, D. L.; Twining, B. S.
- 330 Nanominerals, Mineral Nanoparticles, and Earth Systems. *Science* **2008**, *319* (5870), 1631–1635.
- 331 24. Wächtershäuser, G. Groundworks for an Evolutionary Biochemistry: The Iron-Sulphur World. *Prog.*
- 332 *Biophys. Mol. Biol.* **1992**, *58* (2), 85–201.
- 333 25. Mulkidjanian, A. Y. On the Origin of Life in the Zinc World: 1. Photosynthesizing, Porous Edifices Built of
- 334 Hydrothermally Precipitated Zinc Sulfide as Cradles of Life on Earth. *Biol. Direct* **2009**, *4*, 26.
- 335 26. Stroyuk, A. L.; Raevskaya, A. E.; Korzhak, A. V.; Kuchmii, S. Y. Zinc Sulfide Nanoparticles: Spectral
- 336 Properties and Photocatalytic Activity in Metals Reduction Reactions. *J. Nanoparticle Res.* **2007**, *9* (6), 1027–
- 337 1039.
- 338 27. Sarkar, S.; Guibal, E.; Quignard, F.; SenGupta, A. K. Polymer-Supported Metals and Metal Oxide
- 339 Nanoparticles: Synthesis, Characterization, and Applications. *J. Nanoparticle Res.* **2012**, *14* (2), 715.
- 340 28. Donegá, C. de M. Synthesis and Properties of Colloidal Heteronanocrystals. *Chem. Soc. Rev.* **2011**, *40* (3),
- 341 1512–1546.
- 342 29. Campisi, S.; Schiavoni, M.; Chan-Thaw, C. E.; Villa, A. Untangling the Role of the Capping Agent in
- 343 Nanocatalysis: Recent Advances and Perspectives. *Catalysts* **2016**, *6* (12), 185.
- 344 30. Kirkorian, K.; Ellis, A.; Twyman, L. J. Catalytic Hyperbranched Polymers as Enzyme Mimics; Exploiting
- 345 the Principles of Encapsulation and Supramolecular Chemistry. *Chem. Soc. Rev.* **2012**, *41* (18), 6138–6159.
- 346 31. Liu, L.; Breslow, R. Dendrimeric Pyridoxamine Enzyme Mimics. *J. Am. Chem. Soc.* **2003**, *125* (40), 12110–
- 347 12111.
- 348 32. Astruc, D.; Chardac, F. Dendritic Catalysts and Dendrimers in Catalysis. *Chem. Rev.* **2001**, *101* (9), 2991–
- 349 3024.
- 350 33. Mamajanov, I.; Callahan, M. P.; Dworkin, J. P.; Cody, G. D. Prebiotic Alternatives to Proteins: Structure and
- 351 Function of Hyperbranched Polyesters. *Orig. Life Evol. Biospheres* **2015**, *45*(1-2), 123-137
- 352 34. Mamajanov, I. Wet-Dry Cycling Delays the Gelation of Hyperbranched Polyesters: Implications to the
- 353 Origin of Life. *Life* **2019**, *9* (3).

- 354 35. Mamajanov I. Selective Synthesis of Hyperbranched Polyesters under Wet-Dry Cycling Conditions *ALife*
355 *Conference Proceedings*, **2018**, 580-581.
- 356 36. Mamajanov, I.; Cody, G. D. Protoenzymes: The Case of Hyperbranched Polyesters. *Philos. Trans. R. Soc.*
357 *Math. Phys. Eng. Sci.* **2017**, *375*(2109), 20160537.
- 358 37. Hassan, M. L.; Ali, A. F. Synthesis of Nanostructured Cadmium and Zinc Sulfides in Aqueous Solutions of
359 Hyperbranched Polyethyleneimine. *Cryst. Growth* **2008**, *310*, 5252–5258.
- 360 38. Jia, J.; Wu, A.; Luan, S. Spectrometry Recognition of Polyethyleneimine towards Heavy Metal Ions. *Colloids*
361 *Surf. Colloids Surf. Physicochem. Eng. Asp.* **2014**, *449*, 1–7.
- 362 39. Sang, L.-J.; Wu, Y.-Y.; Wang, H.-F. Polyethyleneimine/Manganese-Doped ZnS Nanocomposites: A
363 Multifunctional Probe for Two-Color Imaging and Three-Dimensional Sensing. *ChemPlusChem* **2013**, *78* (5),
364 423–429.
- 365 40. Drexler, K. E. Molecular Imprinting: The Missing Piece in the Puzzle of Abiogenesis? arXiv:1807.07065 [q-
366 bio.PE] **2018**.
- 367 41. Hu, J.-S.; Ren, L.-L.; Guo, Y.-G.; Liang, H.-P.; Cao, A.-M.; Wan, L.-J.; Bai, C.-L. Mass Production and High
368 Photocatalytic Activity of ZnS Nanoporous Nanoparticles. *Angew. Chem.* **2005**, *117*, 1295–1299.
- 369 42. Stiefel, E. I.; George, G. N. 7 Ferredoxins , Hydrogenases , and Nitrogenases : Metal-Sulfide Proteins.
370 *Bioorganic Chemistry* (Eds.: I. Bertini, H. B. Gray, S. J. Lippard, J. S. Valentine), University Science Books,
371 Mill Valley, **1994**, pp. 365-453.
- 372 43. Pérignon, N.; Marty, J.-D.; Mingotaud, A.-F.; Dumont, M.; Rico-Lattes, I.; Mingotaud, C. Hyperbranched
373 Polymers Analogous to PAMAM Dendrimers for the Formation and Stabilization of Gold Nanoparticles.
374 *Macromolecules* **2007**, *40*, 3034–3041.
- 375 44. Peng, Q.; Zhu, J.; Yu, Y.; Hoffman, L.; Yang, X. Hyperbranched Lysine–Arginine Copolymer for Gene
376 Delivery. *J. Biomater. Sci. Polym. Ed.* **2015**, *26*, 1–35.
- 377 45. Forsythe, J. G.; Yu, S.-S.; Mamajanov, I.; Grover, M. A.; Krishnamurthy, R.; Fernández, F. M.; Hud, N. V.
378 Ester-Mediated Amide Bond Formation Driven by Wet–Dry Cycles: A Possible Path to Polypeptides on the
379 Prebiotic Earth. *Angew. Chem. Int. Ed.* **2015**, *54*, 9871–9875
- 380 46. Rodriguez-Garcia, M.; Surman, A. J.; Cooper, G. J. T.; Suárez-Marina, I.; Hosni, Z.; Lee, M. P.; Cronin, L.
381 Formation of Oligopeptides in High Yield under Simple Programmable Conditions. *Nat. Commun.* **2015**, *6*,
382 8385.
- 383 47. Costanzo, G.; Pino, S.; Ciciriello, F.; Mauro, E. D. Generation of Long RNA Chains in Water. *J. Biol. Chem.*
384 **2009**, *284* (48), 33206–33216.
- 385
386

387 **Supplementary Materials**

388

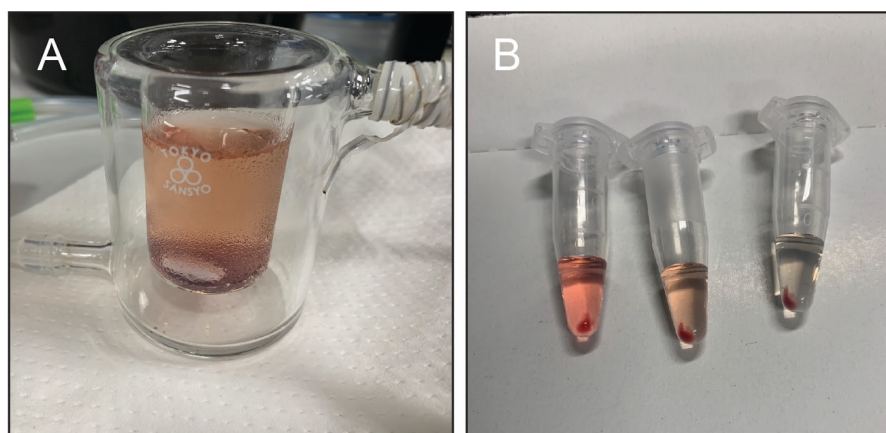
389 **Figure S1.** MALDI mass spectrum of the ZnCl₂-bearing glycerol citrate polyester. The mass spec is
 390 indicative of a heterogeneous mixture of polymeric species. The assigned peaks indicate the masses consistent
 391 with molecular formulae of x glycerol (G) units and y citrate units (C). The assignments were possible in the
 392 case of sodiated signals; zinc complexes were difficult to assign unequivocally due to the complexity of the
 393 zinc isotopic pattern.



394

395 **Figure S2.** Time-lapse measurement of photodegradation of eosin B (5.0×10^{-5} M, 30mL) under additional
 396 conditions: j) with unsupported ZnS particles, under UV, k) with *unreacted* citric acid, glycerol, ZnCl₂, Na₂S,

397 under UV.
398



399

400 **Figure S3.** Visual progression of the eosin B degradation assay catalyzed by unsupported ZnS. A)
401 Photograph of the reaction vessel showing colored precipitate at the end of the reaction. B) Photograph of the
402 centrifuged aliquots taken over the course of the measurement (left to right indicates increasing time) featuring
403 the colored precipitate.