Concise, gram-scale synthesis of furo[2,3-*b*]pyridines, with functional handles for chemoselective cross-coupling

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Abstract: A concise 4-step synthesis of furo[2,3-b]pyridines, with handles in the 3 and 5 positions for palladium mediated cross-coupling reactions, is described. The synthetic route has been optimized, with only one step requiring purification by column chromatography. The route is amenable to scaling-up and was successfully executed on a multi-gram scale. Furopyridines are of growing interest in medicinal chemistry, and this route should enable easy access to the core for structure-activity relationship (SAR) studies.

1. Introduction

Protein kinases play an essential role in signal transduction and regulation of cellular activities including metabolism, cell cycle progression, cell differentiation and apoptosis.[1] Deregulation of kinase function has been implicated in cancers, immunological, neurological, metabolic, and infectious diseases.[2] Chromosomal mapping of the kinome links 244 kinases to a disease loci or cancer amplicon, emphasizing the potential of therapeutics in this area.[3] Kinases are an extremely important group of drug targets, second only to G-protein coupled receptors.[4, 5] To date 54 kinase inhibitor have been approved by the FDA and more than 200 are in clinical trials worldwide.[6-8]

The majority of kinase inhibitors bind to the ATP-binding site, a deep cleft between the N- and C- lobes of the protein's catalytic domain.[4] Most inhibitors bind the active conformation of kinase (type 1), although a significant portion bind to the inactive state (type 2). Other inhibitor types include allosteric inhibitors (type 3), substrate-directed inhibitors (type 4) and covalent inhibitors (type 5).[9]

There are many heterocyclic hinge binding pharmacophores found in kinase inhibitors, some of which are considered privileged fragments.[10, 11] One versatile example is the 7-azaindole core (**Figure 1 - 1**) which is employed as the hinge binder in vemurafenib (**2**), an FDA approved serine/threonine-protein kinase B-Raf (B-Raf) inhibitor used in the treatment of melanoma.[12] Azaindoles are excellent hinge binders, making two hydrogen bonds with the kinase hinge region. Unfortunately, inhibitors with multiple hinge binding interactions may suffer from poor selectivity across the kinome, leading to off-target toxicity.[10, 11, 13]

One strategy to change a kinase inhibitor's selectivity profile is to use isosteric replacements, which can elicit changes in binding affinity between closely related kinases.[14] There is strong evidence to support that modification of

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the hydrogen bond interactions between a kinase inhibitor and the enzyme's hinge region can improve selectivity without sacrificing potency.[11] An isostere of azaindole, the furo[2,3-*b*]pyridine core (**3**), has been gaining traction as a hinge-binding template useful for the synthesis of kinase inhibitors. The furopyridine core contains an electron-deficient pyridine ring and an electron-rich furan ring. Furopyridines are relatively uncommon in natural compounds, with the most prominent examples being furochinoline alkaloids (**4**) isolated from *Rutaceae* plant family and furomegistine I and II (**5** & **6**) alkaloids isolated from the bark of *Sarcomelicope megistophylla* (**Figure 1**).[15, 16]

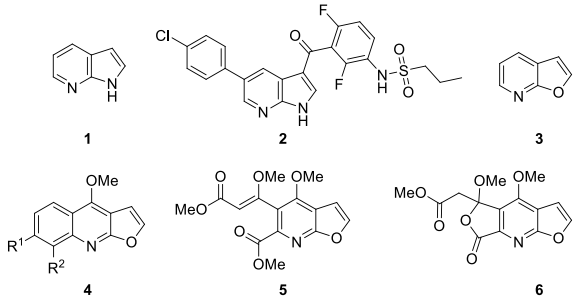


Figure 1: **1** - The azaindole core; **2** - The B-Raf kinase inhibitor vemurafenib; **3** – The furo[2,3-*b*]pyridine core; **4** – The general structure of the furoquinoline alkaloids isolated from *Rutaceae* sp.; **5**–**6**: Alkaloids isolated from *Sarcomelicope megistophylla* - Furomegistine I and II.

Although the furopyridine core is only infrequently found in nature, it is present in several synthetic drug molecules. One of the earliest reports of the furopyridine core being utilized is in the HIV protease inhibitor L-754,394 (**7**).[17] A recent publication describes a set of substituted furopyridine derivatives based on **8** with activity against multidrug-resistant *Mycobacterium tuberculosis* (**Figure 2**).[18] In the kinase field furopyridines have been reported as inhibitors of B-Raf, lymphocyte-specific protein tyrosine kinase (Lck), epidermal growth factor receptor (EGFR) (**9**), insulin-like growth factor 1 receptor (IGF-1R), and AKT (**10**).[19-23]

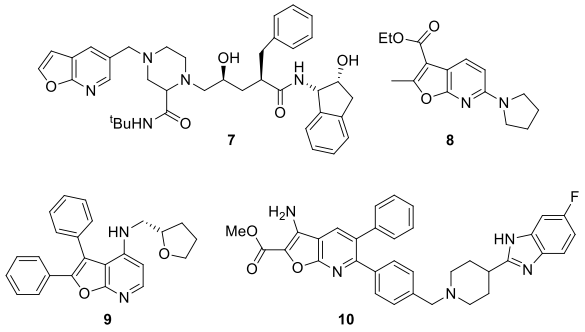
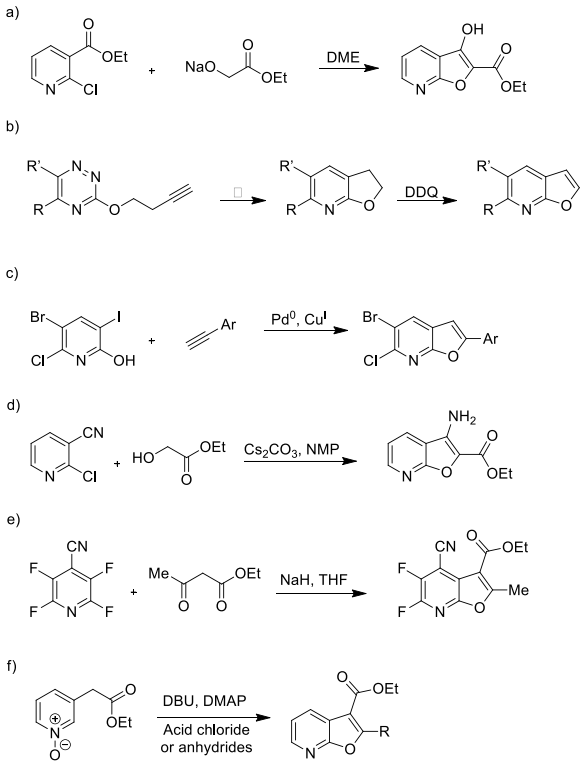


Figure 2: **7** - The HIV protease inhibitor L-754,394; **8** – A furo[2,3-*b*]pyridine which has activity against multidrug resistant *Mycobacterium tuberculosis*; **9** – A furo[2,3-*b*]pyridine based EGFR inhibitor; **10** – A furo[2,3-*b*]pyridine based AKT inhibitor.

Despite their interesting chemical and biological activities, there are only a handful of robust synthetic routes to the furopyridine core. Approaches that have been employed in the synthesis of furopyridines are summarized in **Scheme 1**. Nucleophilic aromatic substitution on 2-halopyridines, followed by subsequent ring closure, has been utilized in several routes to substituted furo[2,3-*b*]pyridines (**Scheme 1** – a), d) and e)).[24-27] An intramolecular Diels-Alder reaction between a triazine and alkyne afforded a dihydrofuro[2,3-*b*]pyridine which was oxidized to the furo[2,3-*b*]pyridine

with DDQ (**Scheme 1** – b)).[28] There are several examples of palladium catalyzed one-pot syntheses of furopyridines in which Sonogashira couplings were followed by Wacker-type heteroannulations (**Scheme 1** – c)).[29] The most recent methodology for synthesis of furopyridines was *via* pyridine *N*-oxides which yielded 2,3-substituted furo[2,3-*b*]pyridines (**Scheme 1** – f)).[30] A comprehensive review on the synthesis of furopyridines has been published by Sirakanyan *et al*.[31]

Our interest in the furo[2,3-*b*]pyridine core was its use as an isosteric hinge binding replacement of a promiscuous azaindole scaffold for the synthesis of a series of kinase inhibitors. We required rapid access on a multigram scale to the furopyridine core that allowed subsequent functionalization at the 3 and 5 positions. Of the synthetic strategies described we decided to utilize the route described by Morita and Shiotani (**Scheme 1** – a).[24] Using this methodology and starting with a trisubstituted pyridine such as **12** should give furopyridine product **13** (Scheme 2). We envisioned that a sequence of saponification of the ester in the 2-position followed by decarboxylation would provide 5-chlorofuro[2,3-*b*]pyridin-3-ol **14**. Conversion of the 3-hydroxy functionality into a triflate would provide us with a di-substituted furopyridine core compatible with versatile palladium mediated coupling reactions at the 3- and 5-positions.

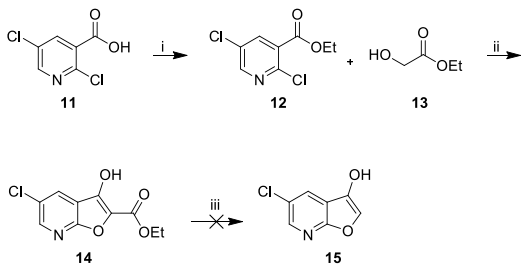


Scheme 1: Reported synthetic routes to the furo[2,3-*b*]pyridine core.

2. Results

a. Furo[2,3-b]pyridine core synthesis

Our initial route is described in **Scheme 2**. 2,5-dichloronicotinic acid, **11**, was converted to the ethyl ester **12** under acidic conditions. Ethyl 2-hydroxyacetate **13** was then deprotonated to the nucleophilic alkoxide, which undergoes an S_NAr reaction to displace the 2-chloro group of **12** with intramolecular cyclization of the putative intermediate to afford the furo[2,3-*b*]pyridine **14**.



Scheme 2: Initial synthetic route to the furo[2,3-*b*]pyridine core **15**. i) EtOH, H_2SO_4, 80 ºC, 16 h, 95%. ii) 1.1 eq. **13**, 3.5 eq NaH, THF, 0–70 ºC, 3 h, 88%. iii) 3 eq. KOH, EtOH, 100 ºC, 20 min then aq. HCl, 100 ºC, 20 min, 0%.

With the furo[2,3-*b*]pyridine **14** constructed, the hydrolysis and decarboxylation steps were attempted. The conditions described by Morita, aqueous potassium hydroxide in refluxing ethanol, failed to yield any saponified or decarboxylated product with only starting material recovered. Changing the base from potassium to sodium or lithium hydroxide also failed to afford **15** (**Table 1 - 1**). Increasing the equivalents of base, switching solvent from ethanol to THF, in combination with longer reaction times for both the hydrolysis and acidification steps, yielded the product **15** in moderate yield (46%) (**Table 1 - 2**). The reaction was attempted under acidic conditions, and also with potassium trimethylsilanolate (**Table 1 - 3/4**). Although the product was isolated from several of these reaction conditions conversion was not quantitative.

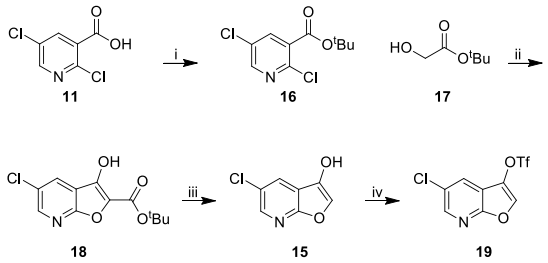
Table 1: Conditions and yields for the one-pot hydrolysis-decarboxylation reaction.

Entry	Conditions	Yield
1	i) 3 eq. KOH*, EtOH,100 °C, 20 min, ii) 3 M HCl, 100 °C, 20 min	0%
2	i) 10 eq. LiOH, THF, H_2O, 60 °C, 16 h ii) 3 M HCl, 100 °C, 1 h	46%
3	6 M HCl, dioxane (1:2), 100 °C, 6 h	57%
4	i) $KOSiMe_3$, THF, 60 °C, 16 h, ii) 4 M HCl, 1 h, 100 °C	63%

*Substituting with NaOH or LiOH afforded the same result.

To further optimise the hydrolysis-decarboxylation reaction, we decided to switch to an acid liable *tert*-butyl ester, which we envisioned cleaving efficiently with TFA. The carboxylic acid of 2,5-dichloronicotinic acid was converted to the *tert*-butyl ester using an acid catalyzed dehydration of concentrated sulfuric acid on magnesium sulfate in the presence of *tert*-butanol, which mediates the formation of isobutylene *in situ*.[32] The conversion of acid **11** to ester **16** proceeded smoothly in excellent yield (92%). A small excess of *tert*-butyl 2-hydroxyacetate **17** was deprotonated with

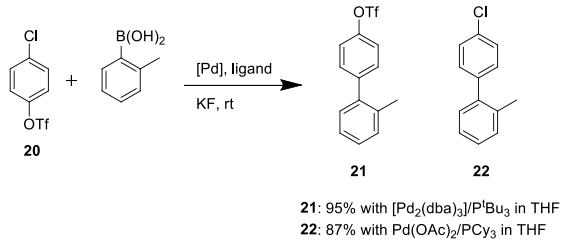
3 equivalents of fresh sodium hydride and utilized in the tandem S_NAr-cyclisation reaction to afford the furo[2,3-*b*]pyridine **18** in excellent yield (86%). Gratifyingly, the TFA mediated *tert*-butyl ester cleavage and decarboxylation afforded the furo[2,3-*b*]pyridine **15** in excellent yield (89%). Notably, the three steps from **11** to **15** were conducted on gram scale without the need for column chromatography at any stage. Conversion of the alcohol **15** to triflate **19** proceeded smoothly. Triflate **19** was synthesised in 71% yield, with an overall yield of 50% from **11,** with only the final step requiring purification *via* column chromatography.



Scheme 3: Revised route to the furo[2,3-*b*]pyridine **19**. i) H_2SO_4, Mg_2SO_4, ^tBuOH, CH_2Cl_2, rt, 16 h, 92%. ii) 1.1 eq. **17**, 3.5 eq.NaH, THF, 0–50 ºC, 3 h, 86%. iii) TFA, CH_2Cl_2, rt, 16 h. 89%. iv) Tf_2O, DIPEA, CH_2Cl_2, -10–25 ºC, 3 h, 71%.

b. Chemoselectivity testing

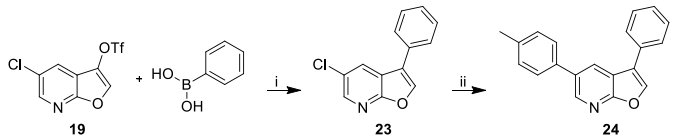
Generally, aryl triflates are considered to have greater reactivity than aryl chlorides in palladium catalyzed C-C bond formation. There are only a few reported systems with selectivity for aryl chlorides over triflates. The first was published by Fu and co-workers in 2000.[33] Fu used $Pd_2(dba)_3$ with the bulky tri-*tert*-butylphosphine ligand P^tBu_3 in THF to obtain selectivity for an aryl chloride in the presence of an aryl triflate (**Scheme 4**). Complementarily, switching to $Pd(OAc)_2$ with the smaller tricyclohexylphosphine (PCy_3) ligand reversed selectivity favoring reactivity at the triflate. The selectivity was rationalized by the ligation state of palladium. P^tBu_3 forms a mono-ligated palladium species which favors C-Cl insertion, but the smaller PCy_3 forms a bis-ligated species which favors C-OTf insertion.



Scheme 4: The chemoselective Suzuki-Miyaura cross-coupling conditions developed by Fu and coworkers.[33] Chemoselectivity is achieved with the appropriate palladium and ligand combination.

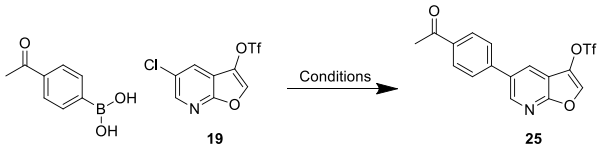
Proutiere and Schoenebeck later found that solvent polarity plays an important role in the selectivity.[34] They demonstrated that, with the $Pd_2(dba)_3/P^tBu_3$ system, changing from THF to DMF switched the selectivity from the chloride to the triflate. The nonpolar toluene retained selectivity for the chloride. Most recently, Neufeldt and co-workers used *N*-heterocycle carbene ligands to achieve chemoselectivity in the Pd-catalyzed Suzuki−Miyaura cross-coupling of chloroaryl triflates.[35]

Controlling chemoselectivity in our furopyridine derivative would be a valuable asset in our medicinal chemistry efforts, so we set out to investigate conditions that would allow this. Our primary objective was to first substitute the triflate at the 3-position followed by the chloro in the 5-position. Effective conditions to couple aryl triflates had already been utilized in our group using palladium tetrakis ($Pd(PPh_3)_4$) and cesium carbonate. The triflate of **19** selectively reacted in a Suzuki reaction using $Pd(PPh_3)_4$ to afford the product **23** in 92% yield. The chlorine was subsequently substituted using a system of $Pd_2(dba)_3$/XPhos to efficiently afford **24** in 84% yield.



Scheme 5: Coupling reaction of **19**. i) $Pd(PPh_3)_4$, Cs_2CO_3, dioxane/water (3:1), 100 ºC, 16 h, 92%. ii) 4-methylphenylboronic acid, $Pd_2(dba)_3$, XPhos, Cs_2CO_3, dioxane/water (3:1), 100 ºC, 16 h, 84%.

Our initial attempts to achieve selectivity for the chlorine over the triflate used the reported system of $Pd_2(dba)_3$/P^{t}Bu$_3$ in THF, either at rt or 70 ºC. 4-acetylphenylboronic acid was used as a coupling partner. We envisioned that the acetyl group could cause a moderate change in polarity and greatly simplify separation of products during chromatography, however after 16 hours only a low yield of unreacted starting material was recovered. We next employed toluene and xylene as solvents to run the reaction at higher temperatures. Frustratingly use of either solvent yielded none of the desired product, although <10% of the triflate substituted product was isolated. Additional aliquots of palladium and ligand with longer reaction times did not improve the result. The NHC carbene pre-catalyst PEPPSI™ SIPr which was reported by Neufeldt as selective and high yielding in coupling of aryl chlorides did not prove effective either, with only starting material recovered.



Scheme 6: Attempted chemoselective Suzuki coupling of the chloride of **19**. Reaction conditions are listed in **Table 2**.

Table 2: Conditions utilized towards the chemoselective coupling of furo[2,3-*b*]pyridine **19**.

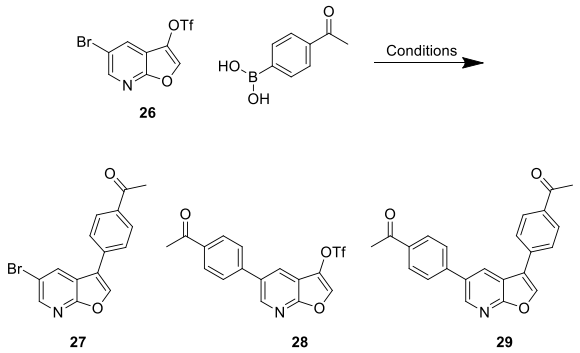
Conditions	Outcome
$Pd_2(dba)_3$, $PtBu_3$, KF, THF, 70 ºC, 16 h	SM recovered[†]
$Pd_2(dba)_3$, $PtBu_3$, KF, PhMe, 110 ºC, 16 h	SM recovered[†]
$Pd_2(dba)_3$, $PtBu_3$, KF, Xylene, 170 ºC, 16 h	SM recovered[†]

PEPPSI-SiPr, KF, PhMe, 110 ºC, 16 h	SM recovered†

† Amounts of starting material recovered varied from 0–48%.

As we could not find condition that led to reaction at the chlorine instead of the triflate, we opted to make bromo analogue of compound **19**. Aryl bromides are often considered to have similar reactivity to triflates, but we wanted to investigate if selectivity for the bromide could be achieved over the triflate. Following the same route as shown in **Scheme 3**, the bromo-triflate **26** was synthesized in 80% overall yield.

Initial attempts to couple the bromide utilized Fu's conditions of $Pd_2(dba)_3$ and P^tBu_3 in THF at room temperature (**Scheme 7** and **Table 3**). The reaction failed to afford product, and only limited starting material was isolated after purification. Increasing the reaction temperature to 70 ºC did afford the product **28**, but in poor yield (16%) along with small amounts of **27** and **29**. Multiple conditions were subsequently explored, affording various mixtures of mono- and di-substituted products. Several conditions reported to selectively couple the triflate over the bromide were also attempted[33-35] (**Table 3**), but all failed to give clean reactions.



Scheme 7: Couplings of the furo[2,3-*b*]pyridine **26** afforded mixtures mono-substituted **27** and **28**, and di-substituted product **29**. The conditions and isolated yields are shown in **Table 3**.

3. Discussion

Our initial synthetic route to the furopyridine core **12** was successful, although the hydrolysis-decarboxylation to **14** was not as efficient as anticipated. Under basic conditions, the stability of the ethyl ester **13** could be due to the ability of the deprotonated starting material to chelate with the positively charged metal counter ion, forming a pseudo 6-membered ring (**Figure 3** – **30a/b**). This intermediate is analogous to the acetylacetonate anion (**31a/b**), which is commonly used as bidentate ligand in metal complexes.[36] The chelation could hinder nucleophilic attack of hydroxide into the ester carbonyl group and suppress the saponification reaction.

Table 3: Conditions attempted to selectively couple the triflate or bromide of **26**. Yields were calculated after isolation. $^{\phi}$Predicted selectivity is based on the results in the relevant literature. *These conditions resulted in the precipitation of palladium metal in the microwave vial, causing intense localised heating and shattering of the microwave vial. This happened on two attempts after which safety concerns led us to stop using the microwave reactor for this reaction. †The amount of starting material recovered varied from 0–71%.

Entry	System	Conditions	Sel$^{\phi}$	26:27:28:29
1	$Pd_2(dba)_3$, P^tBu_3	THF, KF, rt, 16 h	Br	SM recovered†
2	$Pd_2(dba)_3$, P^tBu_3	THF, KF, 70 ºC, 16 h	Br	37%:<5%:16%:<5 %
3	$Pd_2(dba)_3$, P^tBu_3	PhMe, KF, 110 ºC, 16 h	Br	61%:<5%:14%:<5 %
4	$Pd_2(dba)_3$, P^tBu_3	Dioxane, KF, 50 ºC, 16 h	Br	63%:<5%:17%:9%
5	$Pd_2(dba)_3$, P^tBu_3	Dioxane, KF, 100 ºC, 16 h	Br	12%:9%:29%:15%
6	$Pd_2(dba)_3$, P^tBu_3	Xylene, KF, 170 ºC, 16 h	Br	37%:<5%:36%:6%
7	$Pd_2(dba)_3$, P^tBu_3	Xylene, KF, 200 ºC, µW, 30 min	Br	*
8	PEPPSI-SiPr	THF, KF, rt, 16 h	Br	SM recovered†
9	PEPPSI-SiPr	PhMe, KF, 110 ºC, 16 h	Br	SM recovered†
10	$Pd(OAc)_2$/ PCy_3	MeCN, KF, rt, 16 h	OTf	SM recovered†
11	$Pd(OAc)_2$/ PCy_3	MeCN, KF, 70 ºC, 16 h	OTf	51%:44%:<5%:<5 %
12	$Pd(OAc)_2$ /PCy_3	Dioxane/H_2O, KF, 100 ºC, 16 h	OTf	12%:19%:<5%:13 %
13	$Pd(PPh_3)_4$	Dioxane/H_2O, KF, rt, 16 h	OTf	17%:36%:<5%:<5 %
14	$Pd(PPh_3)_4$	Dioxane/ H_2O KF, 50 ºC, 16 h	OTf	13%:42%:11%:<5 %
15	$Pd(PPh_3)_4$	MeCN, KF, 70 ºC, 16 h	OTf	15%:19%:<5%:<5 %

Switching to acidic hydrolysis conditions did afford product, but longer reaction times led to decomposition of the product and reduction in yield. Potassium trimethylsilanolate proved effective but capricious, with the yield on repeated reactions proving variable. Switching to the acid labile *tert*-butyl ester proved to be an effective strategy to facilitate the hydrolysis-decarboxylation step, proceeding cleanly in high yield and on multigram scale with TFA. The conversion of the aryl-alcohol to the triflate proceeded smoothly to afford the furo[2,3-*b*]pyridine with handles in the 3- and 5-positions. The triflate and chloro of **19** could be subjected to Pd-catalyzed coupling reaction sequentially and chemoselectively, which accomplished our primary objective.

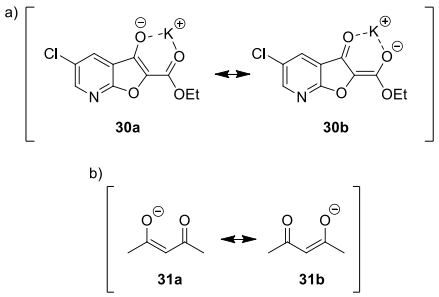


Figure 3: a) The deprotonated species **30** can coordinate a metal counter-ion forming a pseudo 6-membered ring stabilized by resonance forms. This may hinder the nucleophilic attack of hydroxide anions, preventing the saponification of the ester. b) The acetylacetonate ($acac^-$) anion **31** which is often used as a bidentate ligand in inorganic metal complexes.

Unfortunately, all attempts to switch the chemoselectivity by coupling the chloride of **19** before the triflate proved futile. The chlorine group is positioned at the least reactive carbon of the pyridine ring, hindering the oxidative insertion of the active palladium species. This observation is in line the with predictive model of regioselectivity developed by Handy *et al* based on the ^{1}H NMR chemical shift values.[37] Review of published NMR data and predicted NMR spectrums indicate the 5- position has the smallest ^{1}H chemical shift and therefore is often the least reactive.

Switching intermediates to the more reactive bromine did afford some success, but the yields of bromo-substituted product were not high enough for this to be an effective method to support analog synthesis. This approach also led to problems in the chemoselective coupling of the triflate of **26** over the bromide. Having a 5-chloro instead of a 5-bromo increased our ability to achieve selectivity for the triflate in the 3-position. Conditions which couple the triflate in **19** are indiscriminate when used on the bromo-triflate analogue **26**, resulting in non-selective addition to the bromine and triflate. Reducing the temperature from 100 ºC to rt or 50 ºC, did enhance selectivity of the triflate, but the yields were greatly reduced, leading us back to chloro triflate 19 as our key intermediate.

Literature conditions reported to chemoselectively couple aryl halides and triflates performed poorly, and is likely reflective of these challenging furopyridine substrates, which push the current methodologies for Pd-catalyzed reactions to their limits. Initial attempts to couple the bromine afforded only starting material. Changing to higher boiling non-polar solvents, with accompanying increases in temperature did produce low yields of product. The most successful condition employed xylene at 170 ºC, but only afforded the isolated product in 36% yield. Longer reaction times may have improved the yield but running reactions at 170 ºC for over 24 hours did not meet our goal of a practical synthesis.

In conclusion, we successfully developed a 4-step high yielding and scalable synthesis of chloro-triflate furopyridine **19,** requiring only one chromatographic purification. The route is, to the best of our knowledge, the most efficient way to access a furo[2,3-*b*]pyridine core containing synthetic handles for further derivatization. This intermediate has allowed us to undertake a large kinase medicinal chemistry program, the results of which will be published soon.

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References and notes

1 Zhang, J.; Yang, P. L.; Gray, N. S. *Nat. Rev. Cancer,* **2009**, *9*, 28.

2 Ferguson, F. M.; Gray, N. S. *Nat. Rev. Drug Discov.,* **2018**, *17*, 353.

3 Manning, G.; Whyte, D. B.; Martinez, R.; Hunter, T.; Sudarsanam, S. *Science* **2002**, *298*, 1912.

4 Cohen, P. *Nat. Rev. Drug Discov.,* **2002**, *1*, 309.

5 Sriram, K.; Insel, P. A. *Molecular Pharmacology,* **2018**, *93*, 251.

6 Roskoski, R. *Pharmacological Research,* **2019**, *144*, 19.

7 Compiled by Roskoski, R. J.; available online from http://www.brimr.org/PKI/PKIs.htm

8 Carles, F.; Bourg, S.; Meyer, C.; Bonnet, P. *Molecules,* **2018**, *23*.

9 Bhullar, K. S.; Lagarón, N. O.; McGowan, E. M.; Parmar, I.; Jha, A.; Hubbard, B. P.; Rupasinghe, H. P. V. *Mol. Cancer,* **2018**, *17*, 48.

10 Ghose, A. K.; Herbertz, T.; Pippin, D. A.; Salvino, J. M.; Mallamo, J. P. *J. Med. Chem.,* **2008**, *51*, 5149.

11 Xing, L.; Klug-Mcleod, J.; Rai, B.; Lunney, E. A. *Bioorg. Med. Chem.,* **2015**, *23*, 6520.

12 Bollag, G.; Tsai, J.; Zhang, J.; Zhang, C.; Ibrahim, P.; Nolop, K.; Hirth, P. *Nat. Rev. Drug Discov.,* **2012**, *11*, 873.

13 Davis, M. I.; Hunt, J. P.; Herrgard, S.; Ciceri, P.; Wodicka, L. M.; Pallares, G.; Hocker, M.; Treiber, D. K.; Zarrinkar, P. P. *Nat. Biotech.,* **2011**, *29*, 1046.

14 Meanwell, N. A. *J. Med. Chem.,* **2011**, *54*, 2529.

15 Aldona, A.-S.; Kazimierz, G.; Tomasz, B. *Current Issues in Pharmacy and Medical Sciences* **2016**, *29*, 33.

16 Fokialakis, N.; Magiatis, P.; Aligiannis, N.; Mitaku, S.; Tillequin, F.; Sévenet, T. *Phytochemistry* **2001**, *57*, 593.

17 Vacca, J. P.; Dorsey, B. D.; Schleif, W. A.; Levin, R. B.; McDaniel, S. L.; Darke, P. L.; Zugay, J.; Quintero, J. C.; Blahy, O. M.; Roth, E. *P. Nat. A Sci.,* **1994**, *91*, 4096.

18 Fumagalli, F.; de Melo, S. M. G.; Ribeiro, C. M.; Solcia, M. C.; Pavan, F. R.; da Silva Emery, F. *Bioorg. Med. Chem. Lett.,* **2019**, *29*, 974.

19 Buckmelter, A. J.; Ren, L.; Laird, E. R.; Rast, B.; Miknis, G.; Wenglowsky, S.; Schlachter, S.; Welch, M.; Tarlton, E.; Grina, J.; Lyssikatos, J.; Brandhuber, B. J.; Morales, T.; Randolph, N.; Vigers, G.; Martinson, M.; Callejo, M. *Bioorg. Med. Chem. Lett.,* **2011**, *21*, 1248.

20 Ren, L.; Wenglowsky, S.; Miknis, G.; Rast, B.; Buckmelter, A. J.; Ely, R. J.; Schlachter, S.; Laird, E. R.; Randolph, N.; Callejo, M.; Martinson, M.; Galbraith, S.; Brandhuber, B. J.; Vigers, G.; Morales, T.; Voegtli, W. C.; Lyssikatos, J. *Bioorg. Med. Chem. Lett.,* **2011**, *21*, 1243.

21 Wu, Z.; Robinson, R. G.; Fu, S.; Barnett, S. F.; Defeo-Jones, D.; Jones, R. E.; Kral, A. M.; Huber, H. E.; Kohl, N. E.; Hartman, G. D.; Bilodeau, M. T. *Bioorg. Med. Chem. Lett.,* **2008**, *18*, 2211.

22 Martin, M. W.; Newcomb, J.; Nunes, J. J.; Bemis, J. E.; McGowan, D. C.; White, R. D.; Buchanan, J. L.; DiMauro, E. F.; Boucher, C.; Faust, T.; Hsieh, F.; Huang, X.; Lee, J. H.; Schneider, S.; Turci, S. M.; Zhu, X. *Bioorg. Med. Chem. Lett.,* **2007**, *17*, 2299.

23 Hempel, C.; Najjar, A.; Totzke, F.; Schächtele, C.; Sippl, W.; Ritter, C.; Hilgeroth, A. *Med. Chem. Comm.,* **2016**, *7*, 2159.

24 Morita, H.; Shiotani, S. *J. Het. Chem.,* **1986**, *23*, 1465.

25 Cailly, T.; Lemaître, S.; Fabis, F.; Rault, S. *Synthesis* **2007**, *2007*, 3247.

26 Cartwright, M. W.; Parks, E. L.; Pattison, G.; Slater, R.; Sandford, G.; Wilson, I.; Yufit, D. S.; Howard, J. A. K.; Christopher, J. A.; Miller, D. D. *Tetrahedron* **2010**, *66*, 3222.

27 Jasselin-Hinschberger, A.; Comoy, C.; Chartoire, A.; Fort, Y. *Eur. J. Org.Chem.,* **2015**, *2015*, 2321.

28 Taylor, E. C.; Macor, J. E. *Tetrahedron Lett.,* **1986**, *27*, 431.

29 Eastman, K. J.; Parcella, K.; Yeung, K.-S.; Grant-Young, K. A.; Zhu, J.; Wang, T.; Zhang, Z.; Yin, Z.; Beno, B. R.; Sheriff, S.; Kish, K.; Tredup, J.; Jardel, A. G.; Halan, V.; Ghosh, K.; Parker, D.; Mosure, K.; Fang, H.; Wang, Y.-K.; Lemm, J.; Zhuo, X.; Hanumegowda, U.; Rigat, K.; Donoso, M.; Tuttle, M.; Zvyaga, T.; Haarhoff, Z.; Meanwell, N. A.; Soars, M. G.; Roberts, S. B.; Kadow, J. F. *Med. Chem. Comm.* **2017**, *8*, 796.

30 Fumagalli, F.; da Silva Emery, F. *J. Org. Chem.,* **2016**, *81*, 10339.

31 Sirakanyan, S. N.; Hovakimyan, A. A.; Noravyan, A. S. *Russ. Chem. Rev.,* **2015**, *84*, 441.

32 Wright, S. W.; Hageman, D. L.; Wright, A. S.; McClure, L. D. *Tetrahedron Lett.* **1997**, *38*, 7345.

33 Littke, A. F.; Dai, C.; Fu, G. C. *J. Am. Chem. Soc.,* **2000**, *122*, 4020.

34 Proutiere, F.; Schoenebeck, F. *Angew. Chem. Int. Ed.,* **2011**, *50*, 8192.

35 Reeves, E. K.; Humke, J. N.; Neufeldt, S. R. *J. Org.Chem.,* **2019**, *84*, 11799.

36 Manbeck, K. A.; Boaz, N. C.; Bair, N. C.; Sanders, A. M. S.; Marsh, A. L. *J. Chem. Ed.,* **2011**, *88*, 1444.

37 Handy, S. T.; Zhang, Y. *Chem. Comm.,* **2006**, 299.

Supplementary data

Supplementary data associated with this article contains experimental procedures and characterization data for all new compounds.

Supporting Information for "Concise, gram-scale synthesis of furo[2,3-*b*]pyridines, with functional handles for chemoselective cross-coupling"

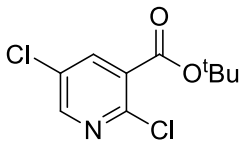
Section S1: Experimental procedures of compound synthesis

Section S2: NMR spectra

Section S1:

General experimental

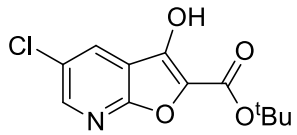
All reagents and solvents, unless specifically stated, were used as obtained from commercial sources without further purification. Air and moisture sensitive reactions were performed under an inert atmosphere using nitrogen in a previously oven-dried or flame-dried reaction flask, and addition of reagents was done using a syringe. All microwave (μW) reactions were carried out in a Biotage Initiator EXP US 400W microwave synthesizer. Thin layer chromatography (TLC) analyses were performed using 200 μm pre-coated sorbtech fluorescent TLC plates and spots were visualized using UV light. High resolution mass spectrometry samples were analyzed with a ThermoFisher Q Exactive HF-X (ThermoFisher, Bremen, Germany) mass spectrometer coupled with a Waters Acquity H-class liquid chromatograph system. Column chromatography was undertaken with a Biotage Isolera One instrument. Nuclear magnetic resonance (NMR) spectrometry was run on a Varian Inova 400 MHz or Bruker Avance III 700 MHz spectrometer equipped with a TCI H-C/N-D 5 mm cryoprobe and data was processed using the MestReNova processor. Chemical shifts are reported in ppm with residual solvent peaks referenced as internal standard.



***Tert*-butyl 2,5-dichloronicotinate 16**

Sulfuric acid (1.67 mL, 31.2 mmol) was added to a stirred suspension of magnesium sulfate (15.0 g, 125 mmol) in dichloromethane (180 mL). The suspension was stirred vigorously for 15 minutes then 2,5-dichloronicotinic acid (6.00 g, 31.2 mmol) and *tert*-butanol (15 mL, 156 mmol) was added. The suspension was stirred for 16 h. A solution of sodium bicarbonate (5% w/v, 180 mL) was added and stirred until the Mg_2SO_4 was dissolved. The organic layer was separated, washed with brine (180 mL), then dried (Na_2SO_4) and concentrated *in vacuo* to afford the *tert*-butyl ester (7.1 g, 92%) as a clear yellow oil.

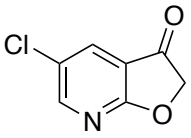
^{1}H NMR ($CDCl_3$, 400 MHz) δ: 8.42 (1H, d, $^4J_{H\text{-}H}$ = 2.6 Hz), 8.02 (1H, d, $^4J_{H\text{-}H}$ = 2.6 Hz, Ar*H*), 1.60 (9H, s, 3 × CCH_3); ^{13}C NMR ($CDCl_3$, 100 MHz) δ: 162.6, 149.8, 147.3, 139.2, 130.6, 129.3, 84.0, 27.9; HRMS calculated for $C_{10}H_{12}Cl_2NO_2$ $[M+H]^+$: m/z 248.0245, found m/z 248.0234.



***Tert*-butyl 5-chloro-3-hydroxyfuro[*2,3-b*]pyridine-2-carboxylate 18**

Tert-butyl 2,5-dichloronicotinate (5.00 g, 20.1 mmol) and *tert*-butyl 2-hydroxyacetate (2.8 g, 21.1 mmol) were dissolved in THF (120 mL) and the solution was cooled to 0 ºC. Sodium hydride (2.4 g, 60.5 mmol) was added portion wise over 30 minutes. Once the evolution of gas stopped the solution was allowed warm to rt. After 1 h the temperature of the reaction was raised to 50 ºC and stirred for 3 h. The reaction was cooled to 0 ºC and the excess sodium hydride was quenched with 1 M aq. HCl and the pH adjusted to 6. The solution was diluted with water (80 mL) and extracted with CH_2Cl_2 (3 × 150 mL). The organic layers were combined, dried (Na_2SO_4) and concentrated *in vacuo* to afford the furopyridine (5.4 g, 86%) as a yellow-orange solid.

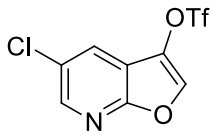
^{1}H NMR ($CDCl_3$, 400 MHz) δ: 8.43 (1H, d, $^4J_{H\text{-}H}$ = 2.4 Hz, ArH), 8.06 (1H, d, $^4J_{H\text{-}H}$ = 2.4 Hz, ArH), 4.82 (1H, s, OH), 1.65 (9H, s, 3 × CCH_3); ^{13}C NMR ($CDCl_3$, 100 MHz) δ 156.8, 155.8, 147.6, 133.3, 129.3, 127.7, 127.5, 113.6, 84.4, 28.3; HRMS calculated for $C_{12}H_{13}ClNO_4$ $[M+H]^+$: m/z 270.0533, found m/z 270.0520; MP Range: 105–107 ºC.



5-chlorofuro[*2,3-b*]pyridin-3-ol 15

Tert-butyl 5-chloro-3-hydroxyfuro[*2,3-b*]pyridine-2-carboxylate (5.00 g, 18.5 mmol) was dissolved in CH_2Cl_2 (45 mL) and trifluoroacetic acid (45 mL, 0.6 mol) was added. The solution was stirred at rt for 16 h. The volatiles were removed *in vacuo* and the residue dissolved in EtOAc (100 mL) before the addition of sat. aqueous $NaHCO_3$ (100 mL). The layers were separated and the organic washed with brine (100 mL), dried (Na_2SO_4) and concentrated *in vacuo* to afford the decarboxylated furopyridine (2.8 g, 89%) as a brown-orange solid.

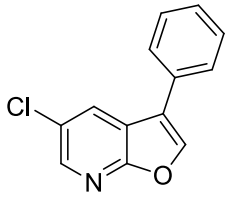
^{1}H NMR ($CDCl_3$, 400 MHz) δ: 8.54 (1H, d, *J* = 2.6 Hz, ArH), 8.00 (1H, d, *J* = 2.6 Hz, ArH), 4.82 (2H, s, CH_2); 13C NMR ($CDCl_3$, 101 MHz) δ 195.8, 175.8, 155.8, 133.4, 126.7, 114.3, 75.6; HRMS calculated for $C_7H_5ClNO_2$ $[M+H]^+$: m/z 170.0008, found m/z 170.0000; MP Range: 115–118 ºC.



5-Chlorofuro[*2,3-b*]pyridin-3-yl trifluoromethanesulfonate 19

A solution of 5-chlorofuro[*2,3-b*]pyridin-3-ol (2.00 g, 11.8 mmol) in CH_2Cl_2 (80 mL) was cooled to -10 ºC. DIPEA (2.69 mL, 14.2 mmol) was added dropwise resulting in a red solution. Triflic anhydride (2.38 mL, 16.5 mmol) was added dropwise and the resulting solution was allowed warm to rt over 2 h. The reaction was quenched with water (80 mL) and the layers separated. The aqueous layer was extracted with CH_2Cl_2 (2 × 80 mL), and the combined organics were dried (Na_2SO_4), filtered and concentrated *in vacuo.* The crude residue was purified using column chromatography (0-5% EtOAc/hexanes) to afford the triflate (2.50 g, 71%) as a pale-yellow oil.

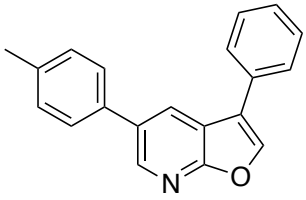
^{1}H NMR ($CDCl_3$, 400 MHz) δ: 8.41 (1H, d, J = 2.4 Hz, ArH), 8.00 (1H, d, J = 2.4 Hz, ArH), 7.95 (1H, s, ArH); ^{13}C NMR ($CDCl_3$, 101 MHz) δ 157.0, 145.2, 136.9, 131.7, 128.8, 127.4, 120.3 (q, $^1J_{C-F}$ = 321.8 Hz), 113.7; HRMS calculated for $C_8H_4ClF_3NO_4S$ $[M+H]^+$: m/z 301.9501 found m/z 301.9489.



5-Chloro-3-phenylfuro[*2,3-b*]pyridine 23

5-Chlorofuro[*2,3-b*]pyridin-3-yl trifluoromethanesulfonate (100 mg, 0.33 mmol) and phenylboronic acid (42 mg, 0.35 mmol) were dissolved in a 10:1 mixture of dioxane and water (2.0 mL). Palladium tetrakis (19 mg, 16 μmol) and cesium carbonate (324 mg, 0.99 mmol) were added. The flask was flushed with nitrogen gas and stirred for 100 ºC for 16 h. Once cooled the volatiles were removed *in vacuo*, and the crude material purified by column chromatography (0–5% EtOAc/hexane) to afford the 3-aryl furopyridine (70 mg, 92%) as a white solid.

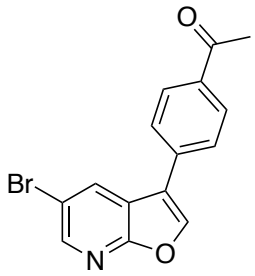
^{1}H NMR ($CDCl_3$, 400 MHz) δ: 8.33 (1H, d, J = 2.3 Hz, ArH), 8.14 (1H, d, J = 2.4 Hz, ArH), 7.91 (1H, s, ArH), 7.61 – 7.55 (2H, m, 2 × ArH), 7.53 – 7.45 (2H, m, 2 × ArH), 7.46 – 7.37 (1H, m, ArH); 13C NMR ($CDCl_3$, 101 MHz) δ 160.6, 143.1, 142.1, 130.5, 129.3, 129.0, 128.2, 127.5, 127.1, 121.4, 119.7; HRMS calculated for $C_{13}H_9NOCl$ $[M+H]^+$: m/z 230.0372 found m/z 230.0356; MP Range: 96–98 ºC.



3-Phenyl-5-(*p*-methylphenyl)furo[*2,3-b*]pyridine 24

5-Chloro-3-phenylfuro[*2,3-b*]pyridine (100 mg, 0.44 mmol) and 4-methylphenylboronic acid (65 mg, 0.48 mmol) were were dissolved in a 10:1 mixture of dioxane and water (2.0 mL). $Pd_2(dba)_3$ (20 mg, 22 μmol), XPhos (21 mg, 44 μmol) and cesium carbonate (426 mg, 1.3 mmol) were added. The flask was flushed with nitrogen gas and stirred at 120 ºC for 16 h. Once cooled the volatiles were removed *in vacuo*, and the crude material purified by column chromatography (5% EtOAc/hexane) to afford the target molecule (104 mg, 84%) as a white solid.

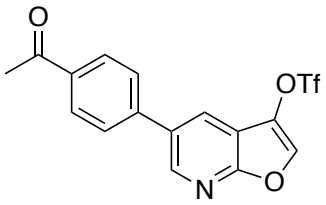
^{1}H NMR ($CDCl_3$, 400 MHz) δ 8.59 (1H,d, *J* = 2.2 Hz, ArH), 8.30 (1H, d, *J* = 2.2 Hz, ArH), 7.91 (1H, s, ArH), 7.67–7.63 (2H, m, ArH), 7.51 (4H, t, *J* = 7.7 Hz, ArH), 7.43 (1H, dd, *J* = 4.8, 3.6 Hz, ArH), 7.32–7.28 (2H, m, 1H), 2.43 (3H, s, CH_3); ^{13}C NMR ($CDCl_3$, 101 MHz) δ 161.9, 143.5, 141.1, 137.6, 135.4, 133.4, 131.2, 129.7 (2 ×C), 129.1 (2 ×C), 127.9, 127.8, 127.4 (2 ×C), 127.1 (2 ×C), 121.6, 118.5, 21.1; HRMS calculated for $C_{20}H_{16}NO$ $[M+H]^+$: m/z 286.1231 found m/z 286.1213; MP Range: 126-129 ºC



1-(4-(5-Bromofuro[*2,3-b*]pyridin-3-yl)phenyl)ethan-1-one 27

5-Bromofuro[*2,3-b*]pyridin-3-yl trifluoromethanesulfonate (75 mg, 0.22 mmol) and 4-acetylphenylboronic acid (36 mg, 0.22 mmol) were loaded into a microwave vial before the addition of degassed acetonitrile (2.0 mL). $Pd(OAc)_2$ (13 mg, 11 µmol), PCy_3 (6 mg, 22 µmol) and KF (38 mg, 0.65 mmol) were added. The flask was flushed with nitrogen gas and stirred at 70 ºC for 16 h. Once cooled the volatiles were removed *in vacuo*, and the crude material purified by column chromatography (0–10% EtOAc/hexane) to afford the aryl-substituted furopyridine (25 mg, 36%) as a white solid. The disubstituted product, 1,1'-(furo[*2,3-b*]pyridine-3,5-diylbis(4,1-phenylene))bis(ethan-1-one), was also isolated from this reaction (5 mg, 6%).

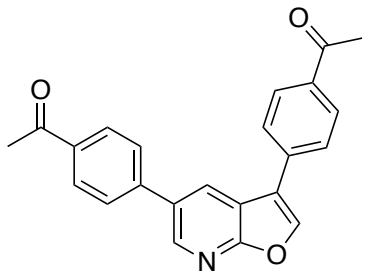
1H NMR (400 MHz, Chloroform-d) δ 8.46 (1H, d, J = 2.2 Hz, ArH), 8.31 (1H, d, J = 2.2 Hz, ArH), 8.10 (1H, d, J = 8.4 Hz, ArH), 8.00 (1H, s, ArH), 7.70 (1H, d, J = 8.4 Hz, ArH); 13C NMR (101 MHz, Chloroform-d) δ 197.2, 161.0, 145.6, 142.9, 136.6, 135.2, 131.9, 129.3, 127.0, 120.4, 119.9, 115.7, 26.6; HRMS calculated for $C_{15}H_{11}NO_2Br$ $[M+H]^+$: m/z 315.9973 found m/z 315.9957; MP Range: 132–134 ºC.



5-(4-Acetylphenyl)furo[*2,3-b*]pyridin-3-yl trifluoromethanesulfonate 28

5-Bromofuro[*2,3-b*]pyridin-3-yl trifluoromethanesulfonate (75 mg, 0.22 mmol) and 4-acetylphenylboronic acid (36 mg, 0.22 mmol) were loaded into a microwave vial before the addition of degassed *p*-xylene (2.0 mL) . $Pd_2(dba)_3$ (13 mg, 11 µmol), P^tBu_3 (6 mg, 22 µmol) and KF (38 mg, 0.65 mmol) were added. The flask was flushed with nitrogen gas and stirred at 170 ºC for 16 h. Once cooled the volatiles were removed *in vacuo*, and the crude purified by column chromatography (0–10% EtOAc/hexane) to afford the aryl-substituted furopyridine (25 mg, 37%) as a white solid. The disubstituted product, 1,1'-(furo[*2,3-b*]pyridine-3,5-diylbis(4,1-phenylene))bis(ethan-1-one), was also isolated from this reaction (5 mg, 6%).

1H NMR (400 MHz, Chloroform-d) δ 8.70 (1H, d, J = 2.2 Hz, ArH), 8.19 (1H, d, J = 2.2 Hz, ArH), 8.15–8.09 (2H, m, 2 × ArH), 7.98 (1H, s, ArH), 7.78–7.69 (2H, m, 2 × ArH), 2.68 (3H, s, CH_3); 13C NMR (101 MHz, Chloroform-d) δ 197.4, 158.7, 145.6, 141.7, 136.7, 136.2, 133.5, 132.6, 129.3, 127.8, 126.5, 118.7 (q, $^1J_{C-F}$ = 321.6 Hz), 112.96, 26.71; HRMS calculated for $C_{16}H_{11}F_3NO_5S$ $[M+H]^+$: m/z 386.03100 found m/z 386.0288; MP Range: 113-115 ºC.

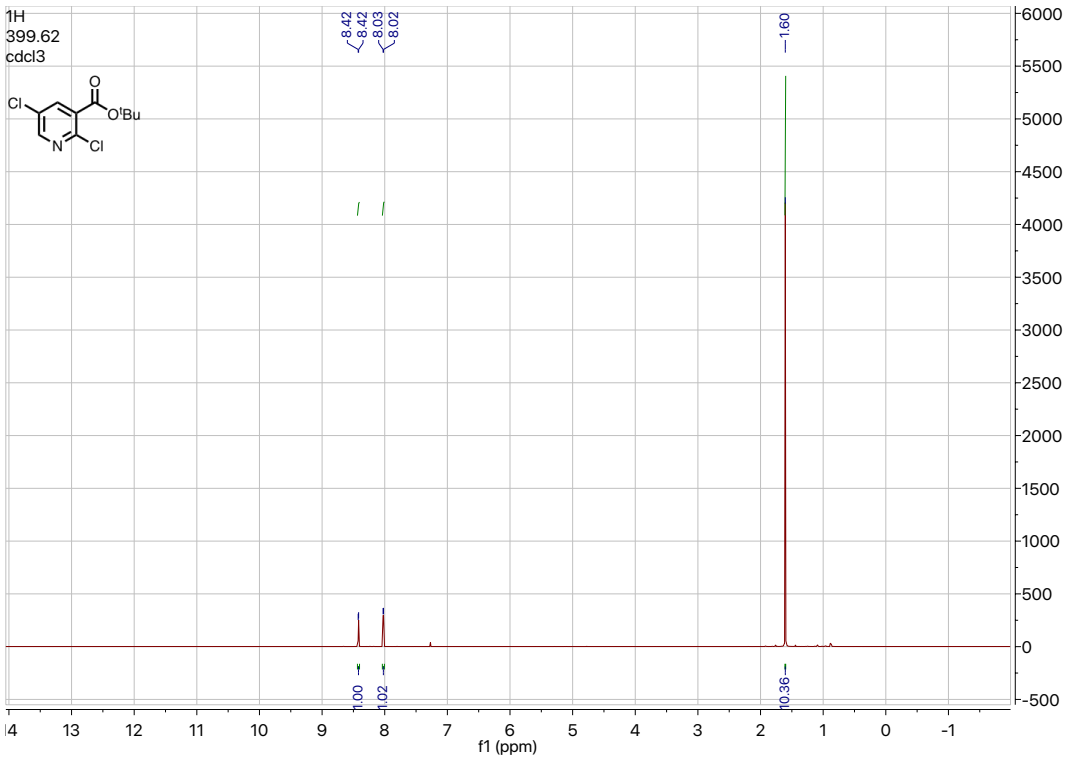


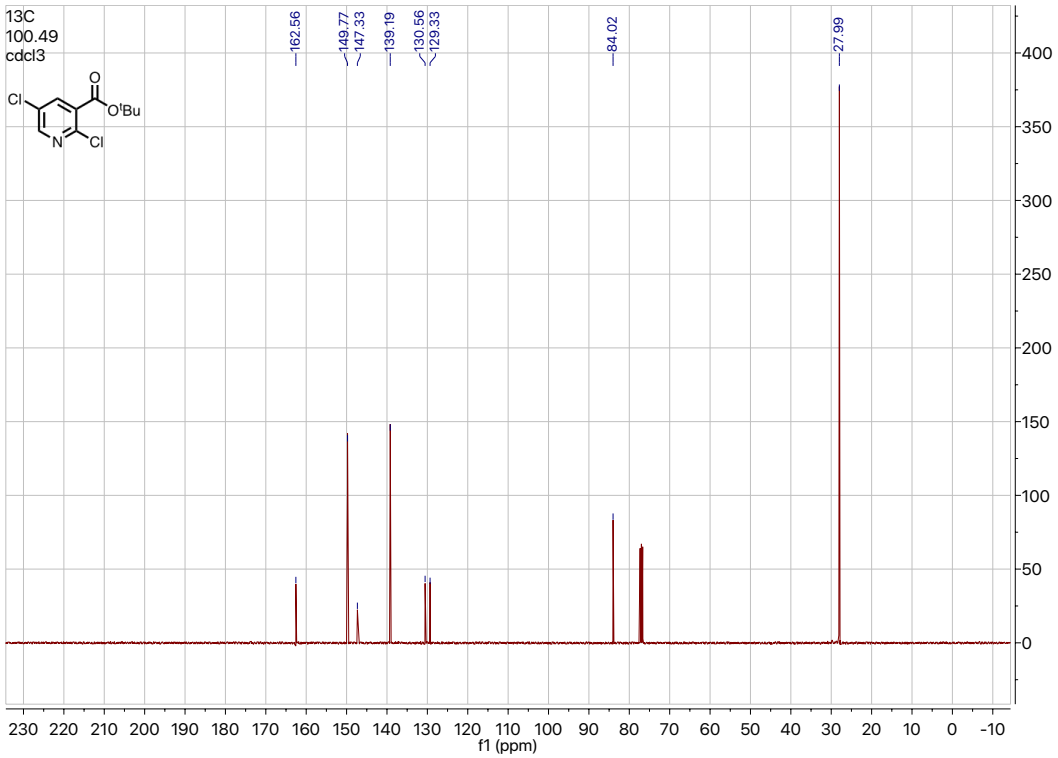
1,1'-(Furo[*2,3-b*]pyridine-3,5-diylbis(4,1-phenylene))bis(ethan-1-one) 29

1H NMR (400 MHz, Chloroform-d) δ 8.67 (1H, d, J = 2.2 Hz, ArH), 8.38 (1H, d, J = 2.2 Hz, ArH), 8.19–8.08 (4H, m, 4 ArH), 8.06 (1H, s, ArH), 7.81–7.69 (4H, m, 4 × ArH), 2.68 (6H, app d, J = 1.0 Hz, 2 × CH_3); 13C NMR (101 MHz, Chloroform-d) δ 197.5, 197.3, 162.4, 144.0, 142.7, 142.5, 136.5, 136.4, 135.8, 132.6, 129.3, 129.2, 128.2, 127.7, 127.1, 120.9, 118.3, 26.7, 26.6; HRMS calculated for $C_{23}H_{18}NO_3$ $[M+H]^+$: m/z 356.1286 found m/z 356.1269; MP Range: 124-125 ºC.

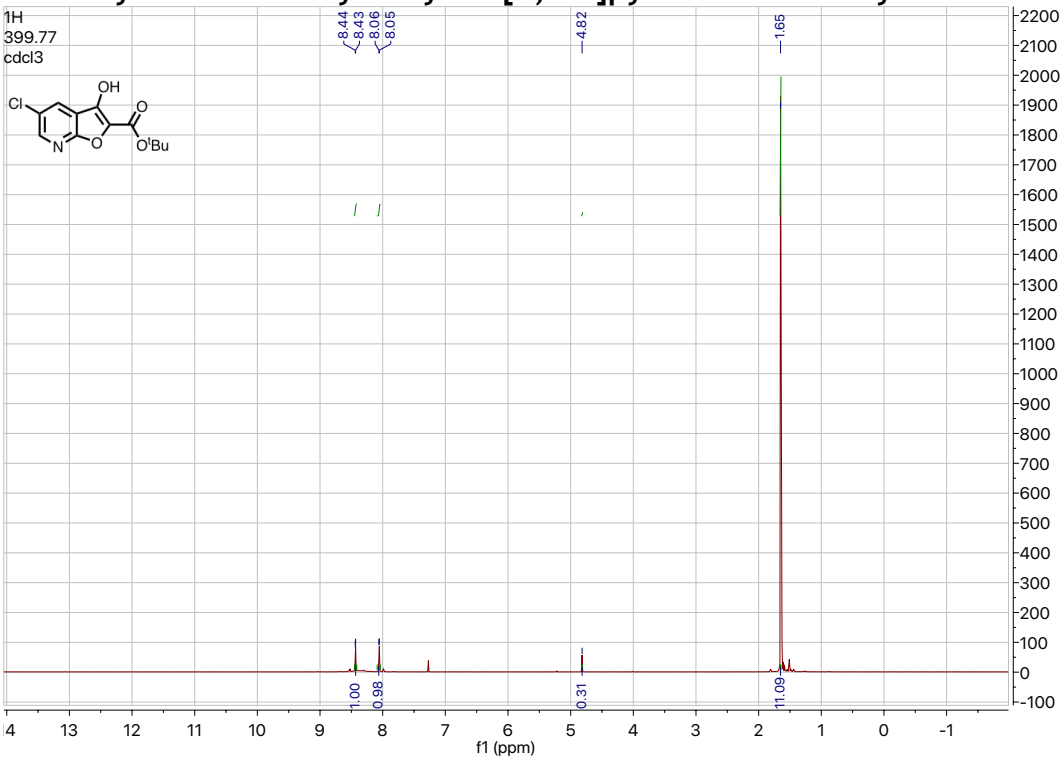
Section S2:

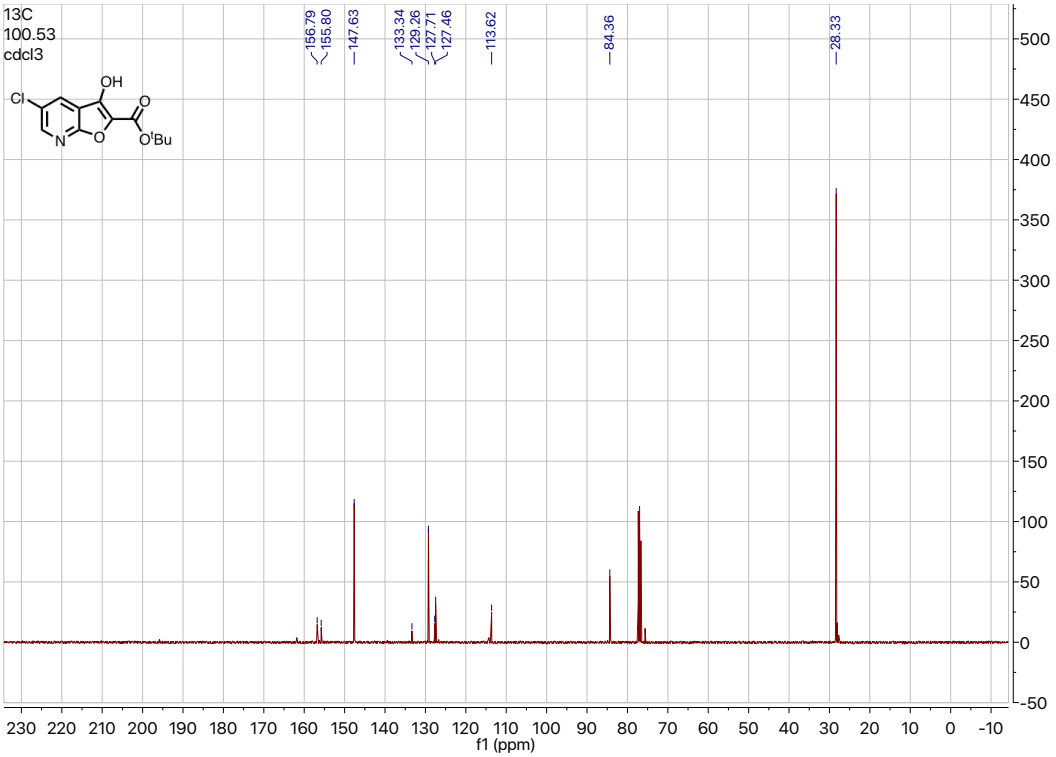
Tert-butyl 2,5-dichloronicotinate 16



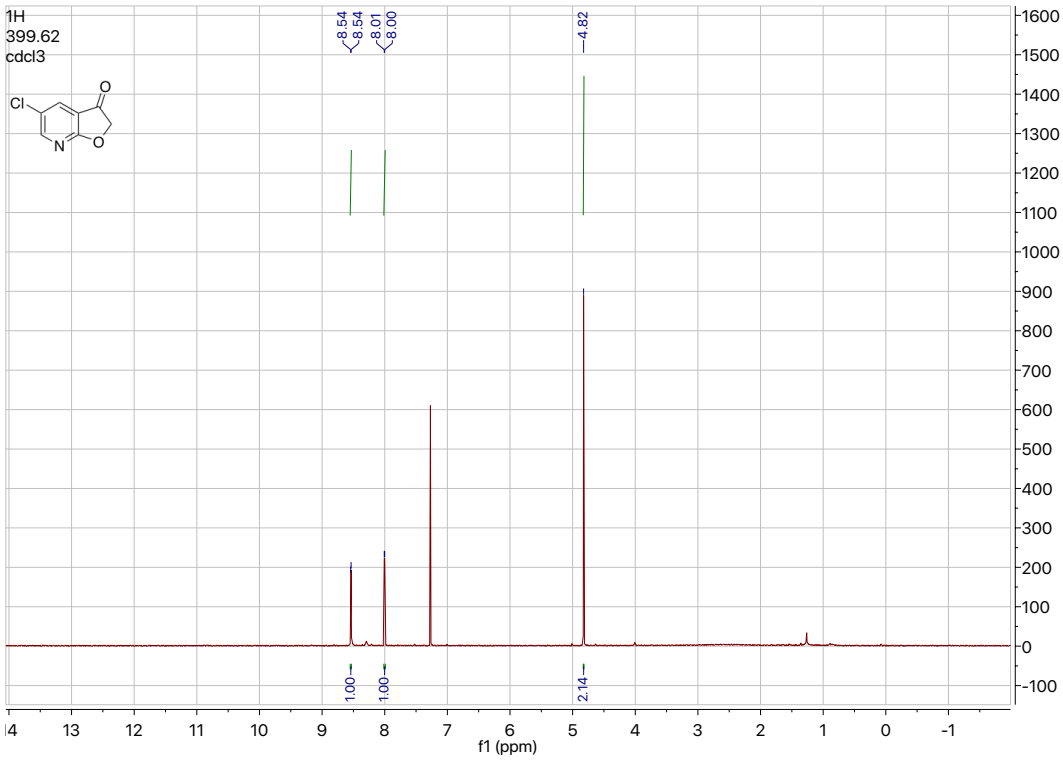


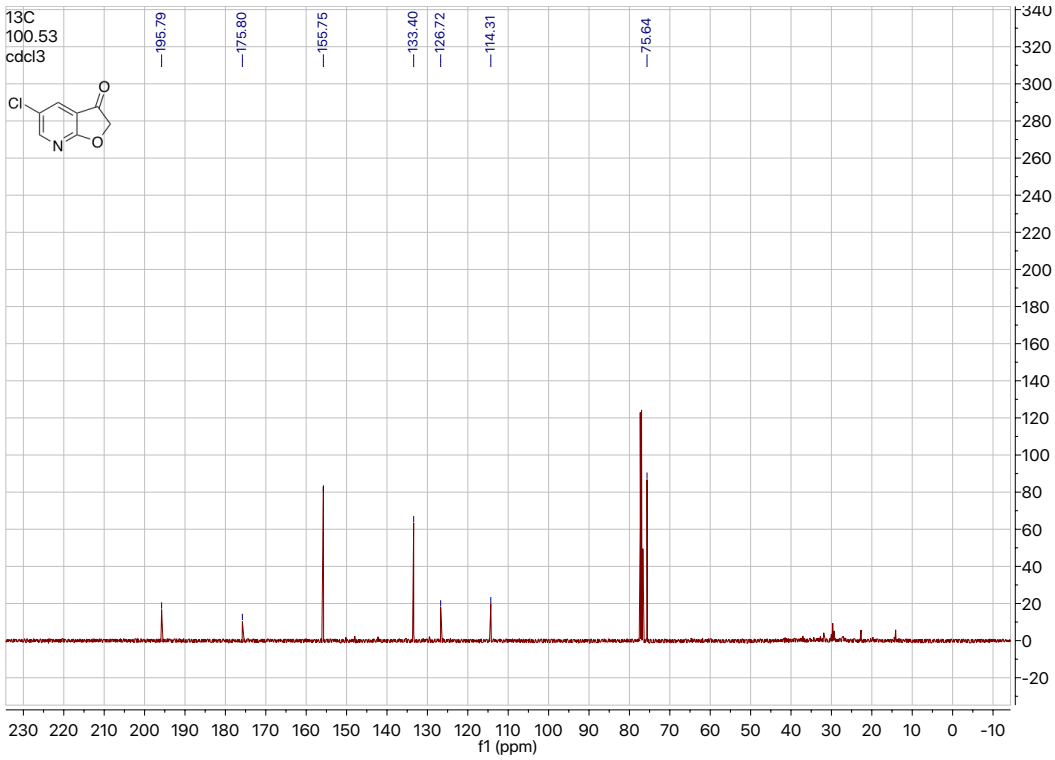
Tert-butyl 5-chloro-3-hydroxyfuro[*2,3-b*]pyridine-2-carboxylate 18



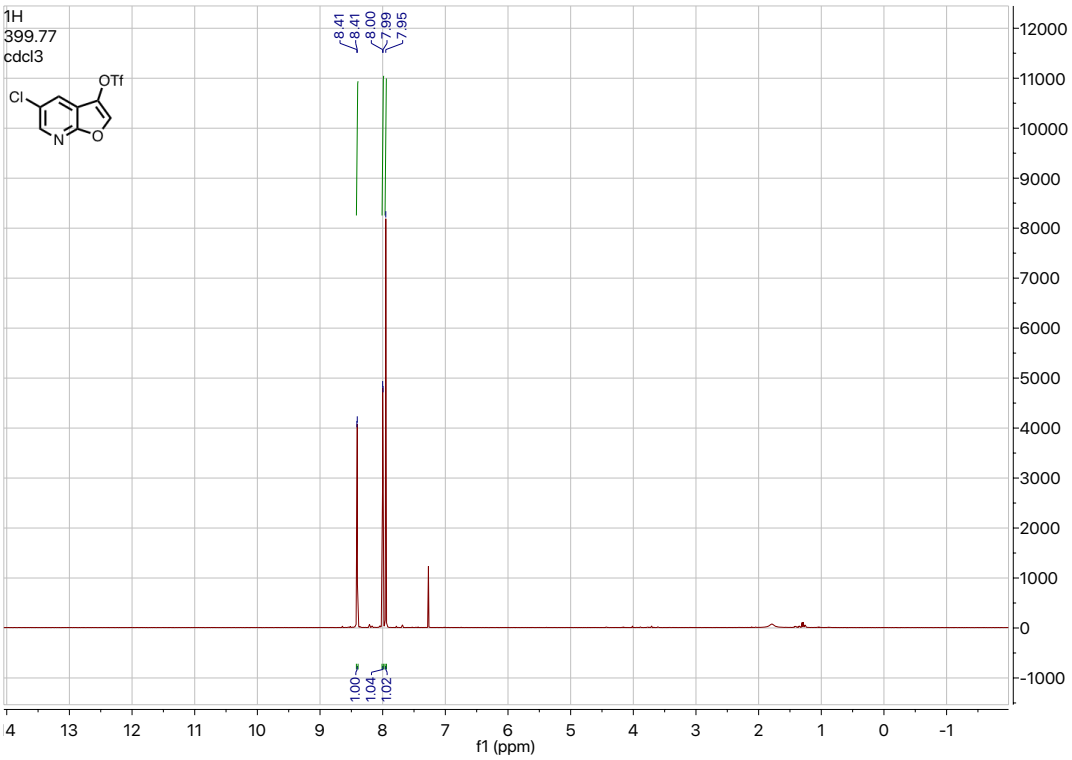


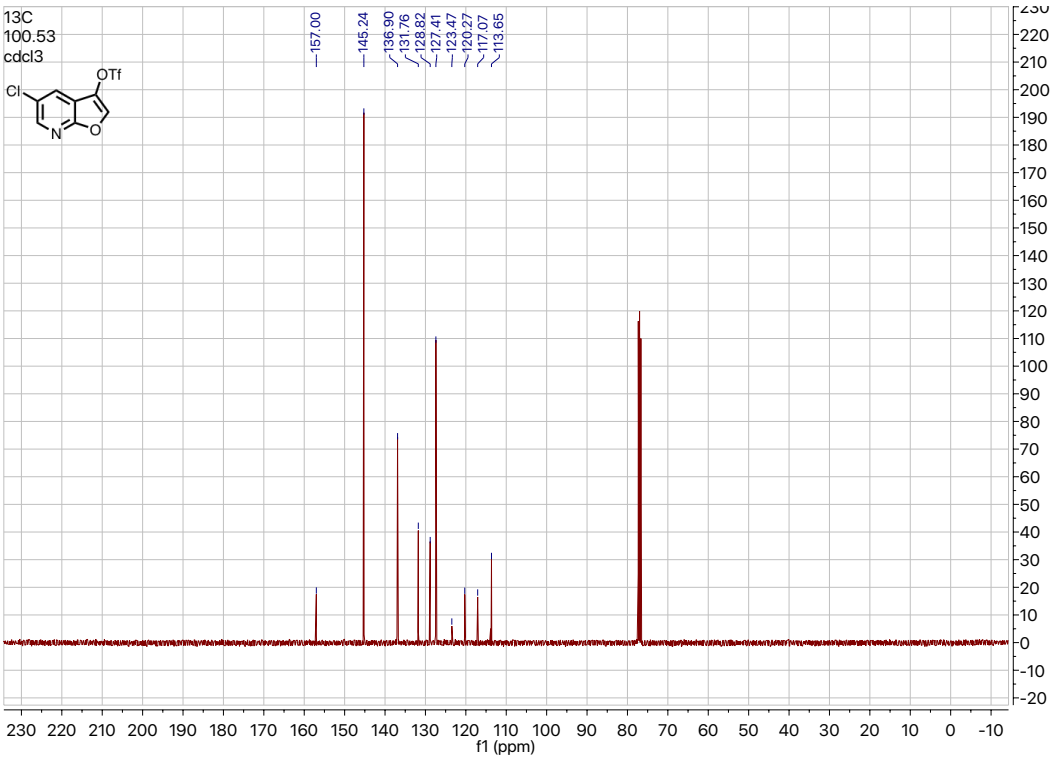
5-Chlorofuro[*2*,*3*-*b*]pyridin-3-ol 15

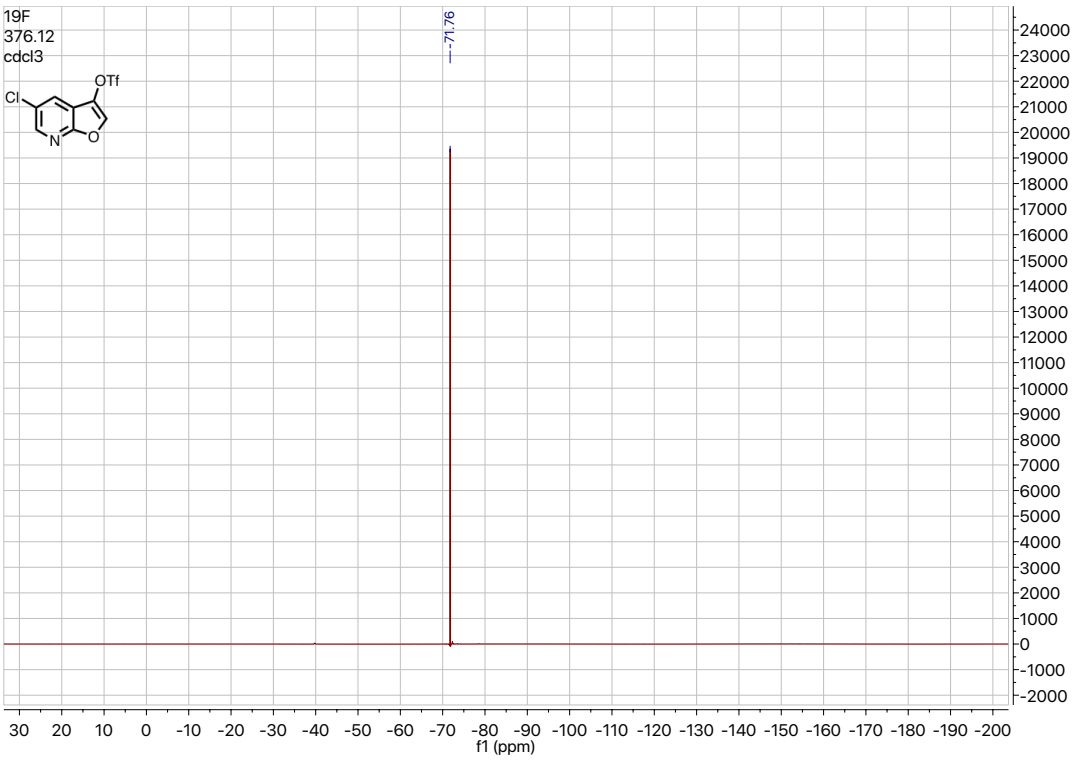




5-Chlorofuro[*2*,*3*-*b*]pyridin-3-yl trifluoromethanesulfonate 19



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OTf
Cl
N
O
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23000
22000
21000
20000
19000
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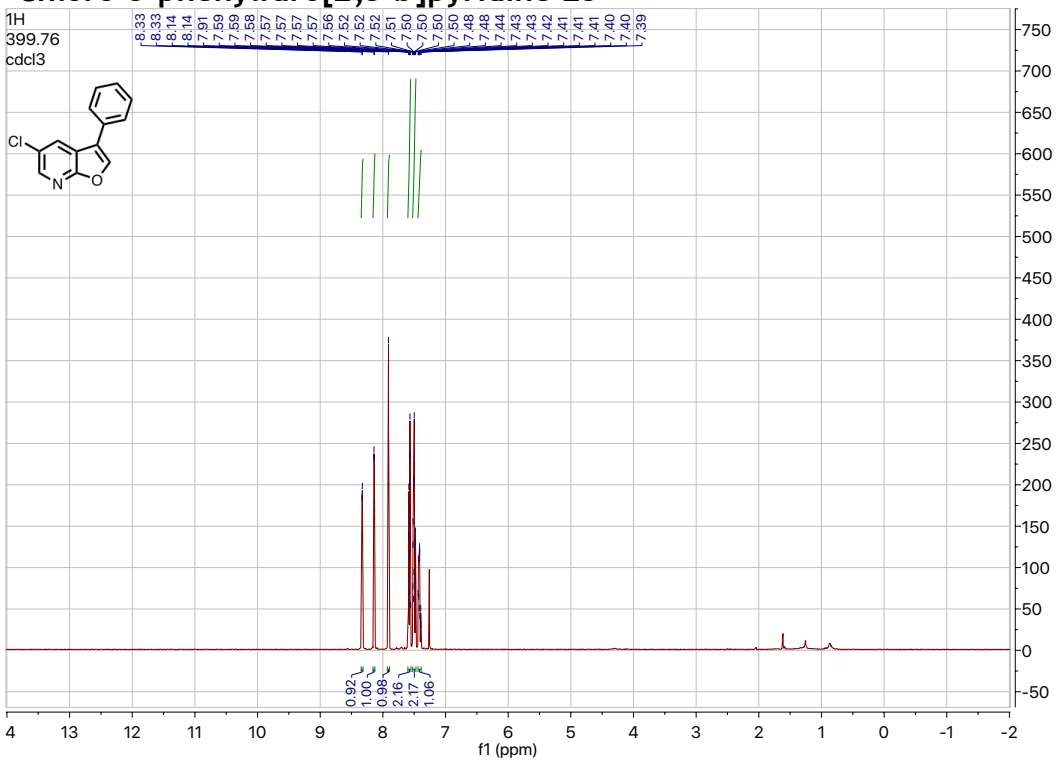
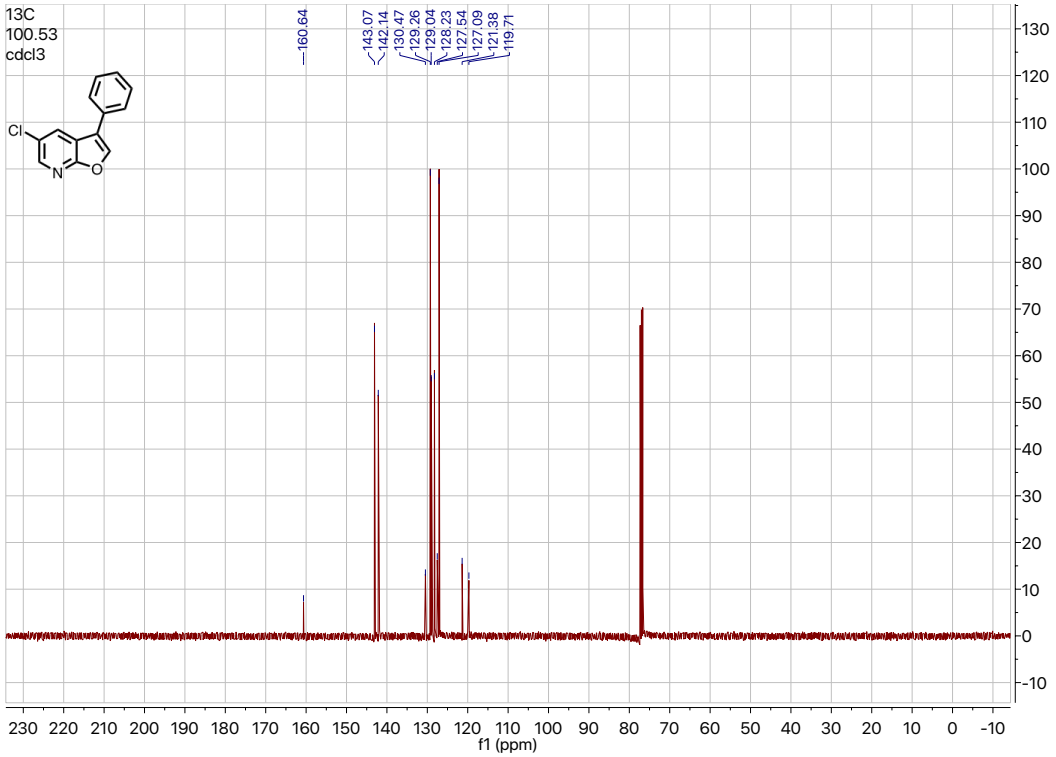
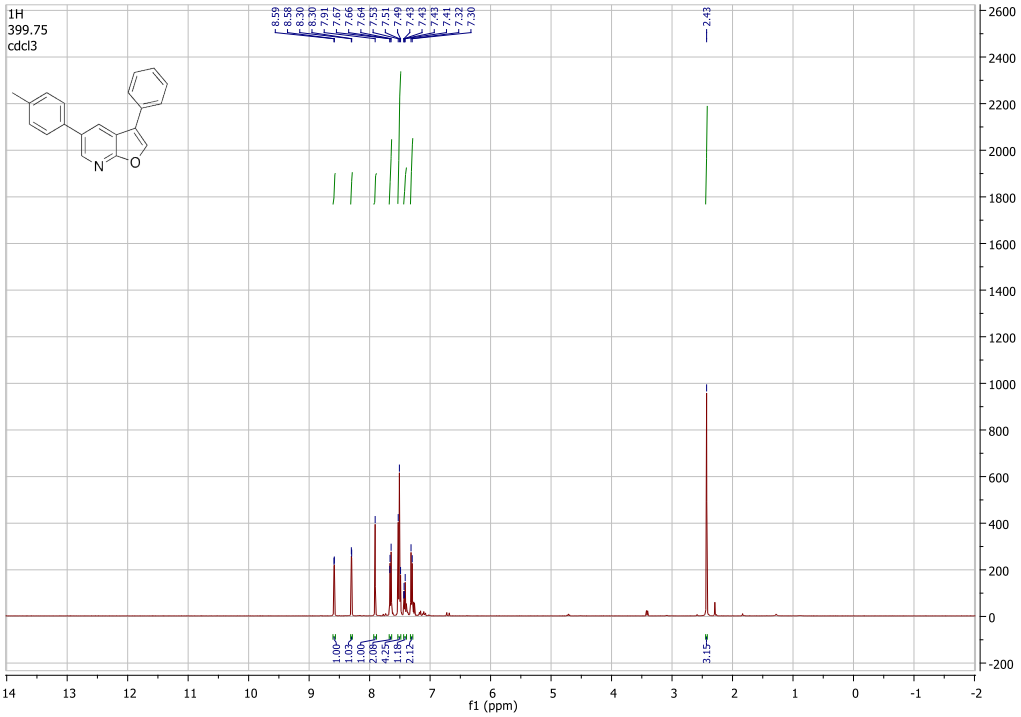
17000
16000
15000
14000
13000
12000
11000
10000
9000
8000
7000
6000
5000
4000
3000
2000
1000
0
-1000
-2000
30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200
f1 (ppm)

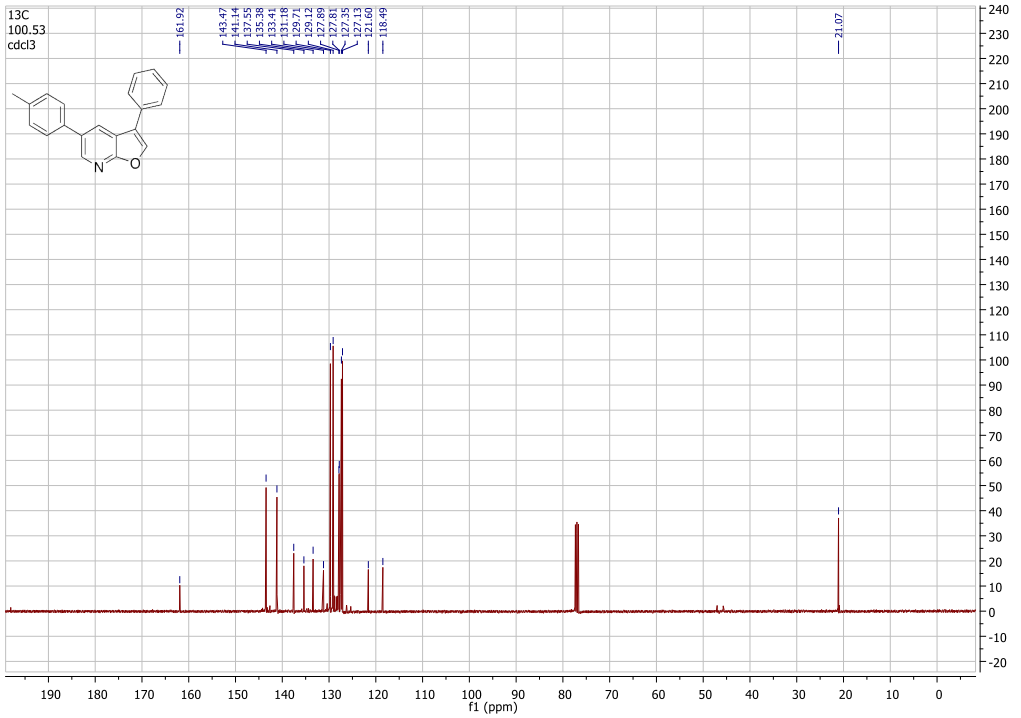
5-Chloro-3-phenylfuro[2,3-*b*]pyridine 23



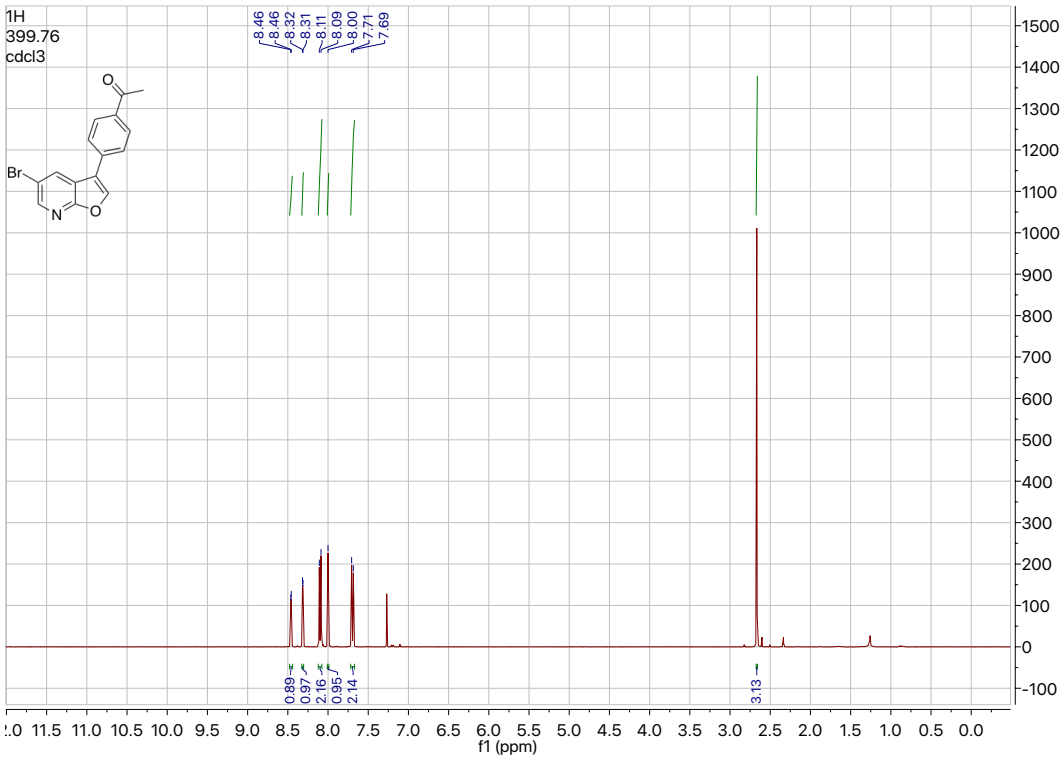


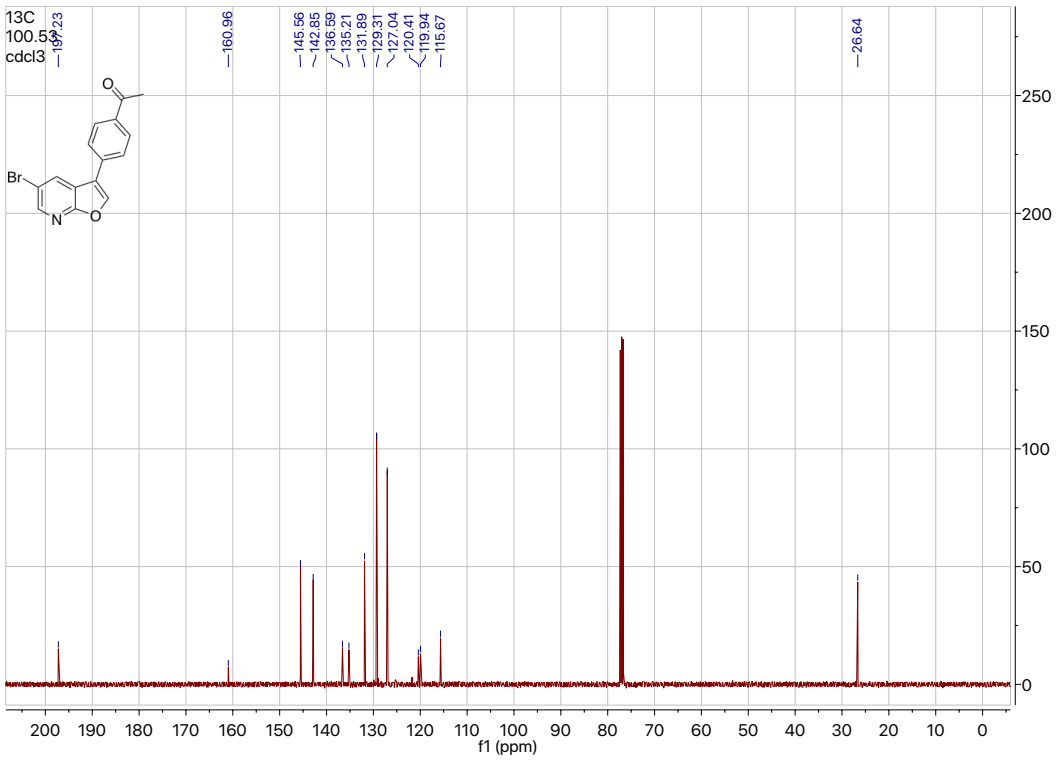
3-Phenyl-5-(*p*-methylphenyl)furo[*2*,*3*-*b*]pyridine 24



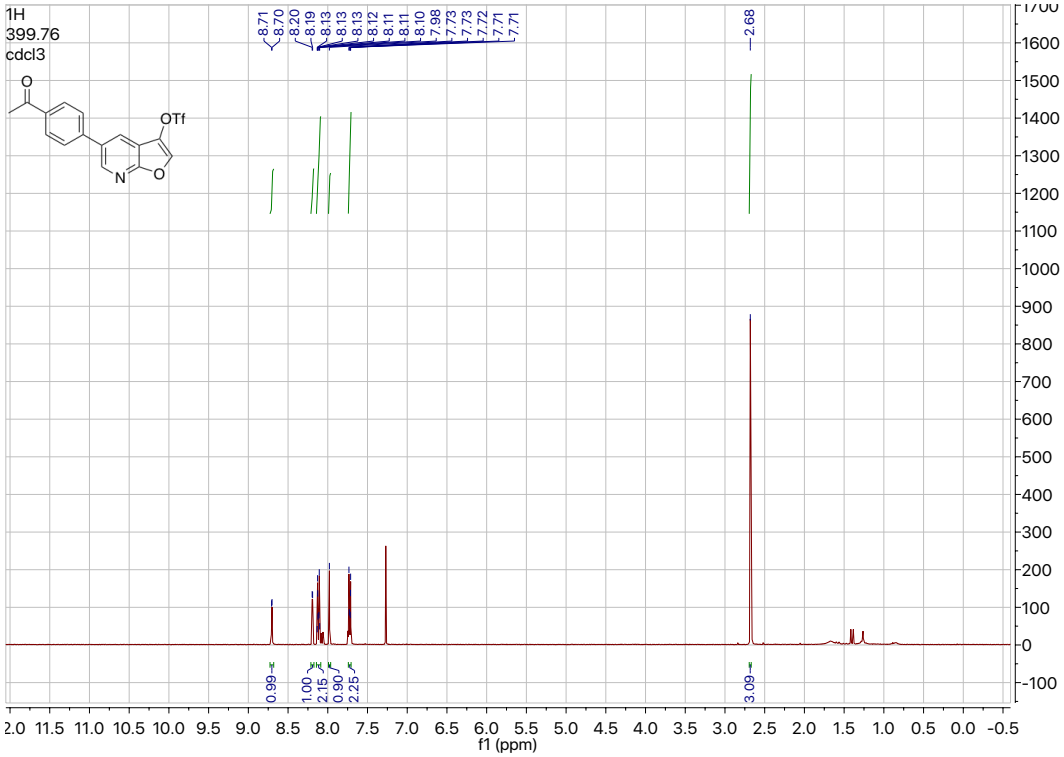


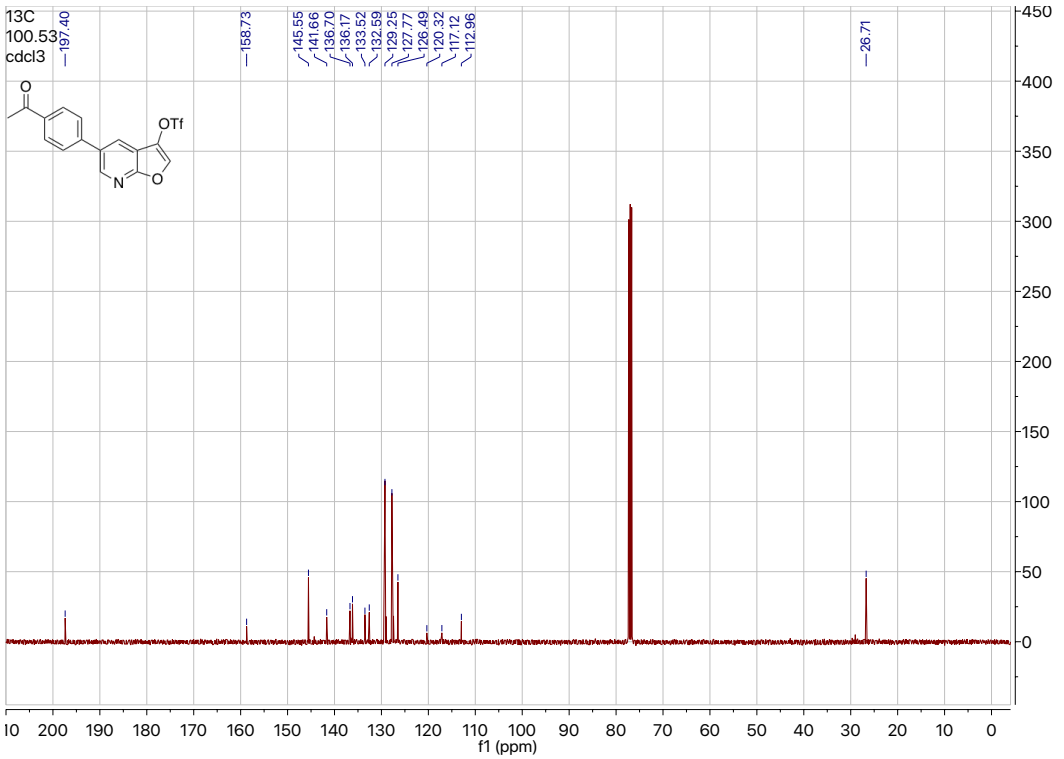
1-(4-(5-Bromofuro[*2*,*3*-*b*]pyridin-3-yl)phenyl)ethan-1-one 27

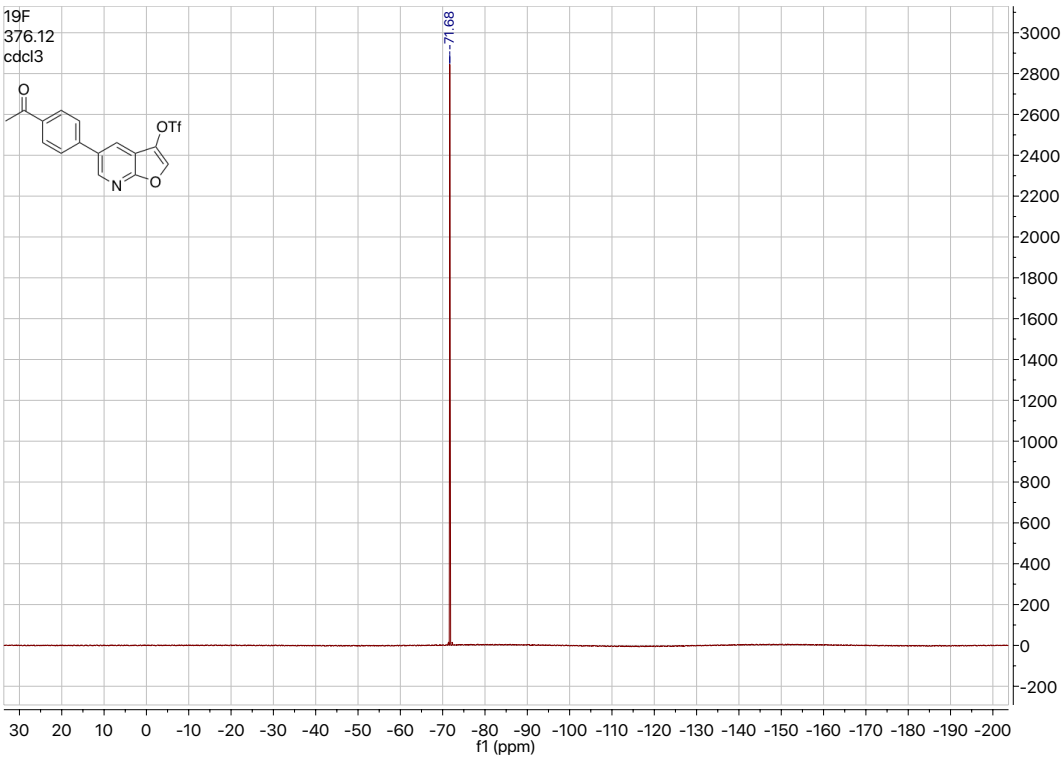




5-(4-Acetylphenyl)furo[*2,3-b*]pyridin-3-yl trifluoromethanesulfonate 28



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OTf
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2800
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2400
2200
2000
1800
1600
1400
1200
1000
800
600
400
200
0
-200
30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200
f1 (ppm)

1,1'-(Furo[*2*,*3*-*b*]pyridine-3,5-diylbis(4,1-phenylene))bis(ethan-1-one) 29