

# Facile Synthesis of Formate-Functionalized Graphene Quantum Dots

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Graphene quantum dots have proven useful for a variety of applications due to their impressive bandgap tunability, which can be achieved through structural modification methods including edge functionalization and doping. Here, we investigate the functionalization of graphene quantum dots with formate esters using a method of noncatalyzed electrophilic substitution. We also examine the optical and structural properties of these formate-functionalized graphene quantum dots using UV-Vis Spectrophotometry, IR Spectroscopy, and NMR Spectroscopy, with the functionalization causing a redshift in absorption consistent with a reduction in bandgap. The findings in this paper have the potential to be used in applications ranging from single photon emission in quantum computing to tunable fluorescence in bioimaging.

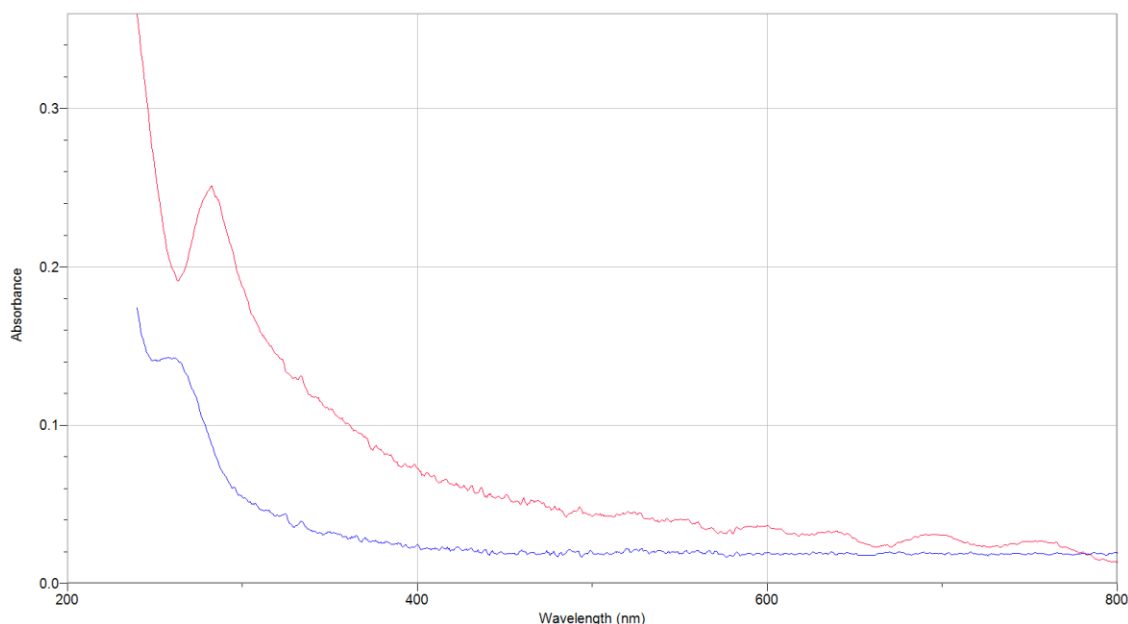
## INTRODUCTION

Graphene, an allotrope of carbon, has garnered much attention since its experimental discovery in 2004 due to its unique properties.<sup>1</sup> Being one atom thick and containing a hexagonal,  $sp^2$ -hybridized lattice-type structure, graphene possesses a low mass coupled with impressive structural strength.<sup>2</sup> Additionally, graphene's electronic band structure allows for high electron mobility, making it an excellent conductor of heat and electricity.<sup>3</sup> Finally, graphene - even in quantum dot form - poses low health risks,<sup>4</sup> and is easily obtainable from graphite, a common material composed of stacked sheets of graphene. This combination of ideal properties permits many applications of graphene in electronics, bioimaging, biosensing, and more.

In recent years, a new application of graphene has received attention - graphene quantum dots (GQDs), or 0-dimensional particles of graphene.<sup>5</sup> GQDs inherit many of graphene's desirable physical properties, such as high electron mobility and low toxicity. Additionally, due to their size (on the order of  $10^{-8}$  m), GQDs are also subject to quantum confinement, and exhibit tunable photoluminescence as a result.<sup>6</sup> Due to these ideal properties, GQDs have found a home in a variety of applications, including solar cells,<sup>7</sup> lithium-ion batteries,<sup>8</sup> bioimaging,<sup>9</sup> and single-photon emission.<sup>10</sup>

As with other types of quantum dots, such as those composed of amorphous carbon, there exists the potential to enhance the photoluminescence (PL) properties of GQDs through modification of their chemical structures. One common method of doing so is the introduction of a dopant into the structure of a GQD - for instance, Ain *et al.* have examined the effects of doping GQDs with boron, nitrogen, chlorine, sodium, and potassium, and have found significant differences in PL properties following the introduction of the dopants.<sup>11</sup> Another avenue for altering PL characteristics, which forms the focus of this paper, involves functionalizing the edge structures of graphene quantum dots. Multiple approaches to edge functionalization have been examined in the past: Kumar *et al.*, among others, have examined amino-functionalized graphene oxide QDs,<sup>12</sup> and Li *et al.* have performed a theoretical study on GQDs functionalized with a variety of chemical groups, many containing C=O double bonds.<sup>13</sup> However, there still exist a variety of potential candidates for edge functionalization that have yet to be explored. As such, the focus of this paper is to create and experimentally verify a protocol for the functionalization of GQDs using formate, which, to our best knowledge, has yet to be experimentally achieved. We also aim to examine the effect of formate functionalization on the optical absorbance properties of GQDs.

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**Figure 1:** Image of the UV-Vis spectrum for GQDs (blue curve) and functionalized GQDs (red curve), showing a redshift in absorption due to a reduction in bandgap as a result of functionalization. Different concentrations of functionalized/non-functionalized quantum dots were used.

## EXPERIMENTAL METHODS

### I. Graphene Quantum Dot Fabrication

To fabricate non-functionalized GQDs, an electrochemical process derived from the work of Ahirwar *et al.* was used.<sup>14</sup> To start, 500 mL of a solution containing 0.10 M citric acid monohydrate (Showa Chemical) and 0.30 M sodium hydroxide (Choneye Pure Chemicals), dissolved in Milli-Q water, was prepared. This 1:3 molar ratio of citric acid to NaOH was chosen for its ability to produce GQDs with low amounts of oxygen-containing functional groups, in addition to high amounts of C-H bonds on the GQDs' surface.<sup>14</sup> Two graphite rods were then heated at  $\sim 1000^{\circ}\text{C}$  for  $\sim 10$  minutes to introduce surface impurities before being rinsed in Milli-Q water, dried, and placed vertically in the solution 25 mm apart.

Using a DC power supply, a voltage was passed through the rods for 15 minutes, alternating between +0.5 and +1.2 V every  $\sim 5$  seconds with current between 0.005 and 0.010 A. Following this, a constant voltage of +10.0 V (with current equal to 0.200 A) was applied to the rods for 30 minutes. The solution gradually

turned from clear to yellow, indicating the formation of GQDs. 0.15 M calcium chloride was then added to the solution and heated to  $70^{\circ}\text{C}$  for 15 minutes to precipitate calcium citrate. The precipitate was filtered out of solution, and the solution was placed in dialysis tubing (#684216, Carolina) for 7 days.

After 7 days, a yellow-colored solid was obtained after the removal of water via rotary evaporation. UV-Vis spectrophotometry (VSP-UV, Vernier), IR spectroscopy (UATR Two, PerkinElmer), and  $^1\text{H}$  NMR spectroscopy were performed to characterize the sample. Prior to  $^1\text{H}$  NMR spectroscopy (Avance, Bruker), the sample was dispersed in ethanol to isolate GQDs from any remaining calcium citrate from the synthesis process, and then reduced to dryness and rehydrated in  $\text{D}_2\text{O}$ .

### II. Graphene Quantum Dot Functionalization with Formate

To functionalize the GQD sample with formate, an excess of formic acid (determined using 1:1 molar ratio based on total GQD mass

and molar mass of carbon) was added to the sample in a flask.

The resulting solution was refluxed and stirred for 48 hours, at 95°C, inside of a fume hood. For UV-Vis spectrophotometry (VSP-UV, Vernier) and IR spectroscopy (UATR Two, PerkinElmer), this functionalized GQD sample was placed in dialysis tubing (#684216, Carolina) for 7 days to filter any excess formic acid. For  $^1\text{H}$  NMR spectroscopy (Avance, Bruker), this functionalized GQD sample was dispersed in ethanol to isolate GQDs from any remaining calcium citrate from the synthesis process, and then reduced to dryness and rehydrated in  $\text{D}_2\text{O}$ .

## RESULTS AND DATA

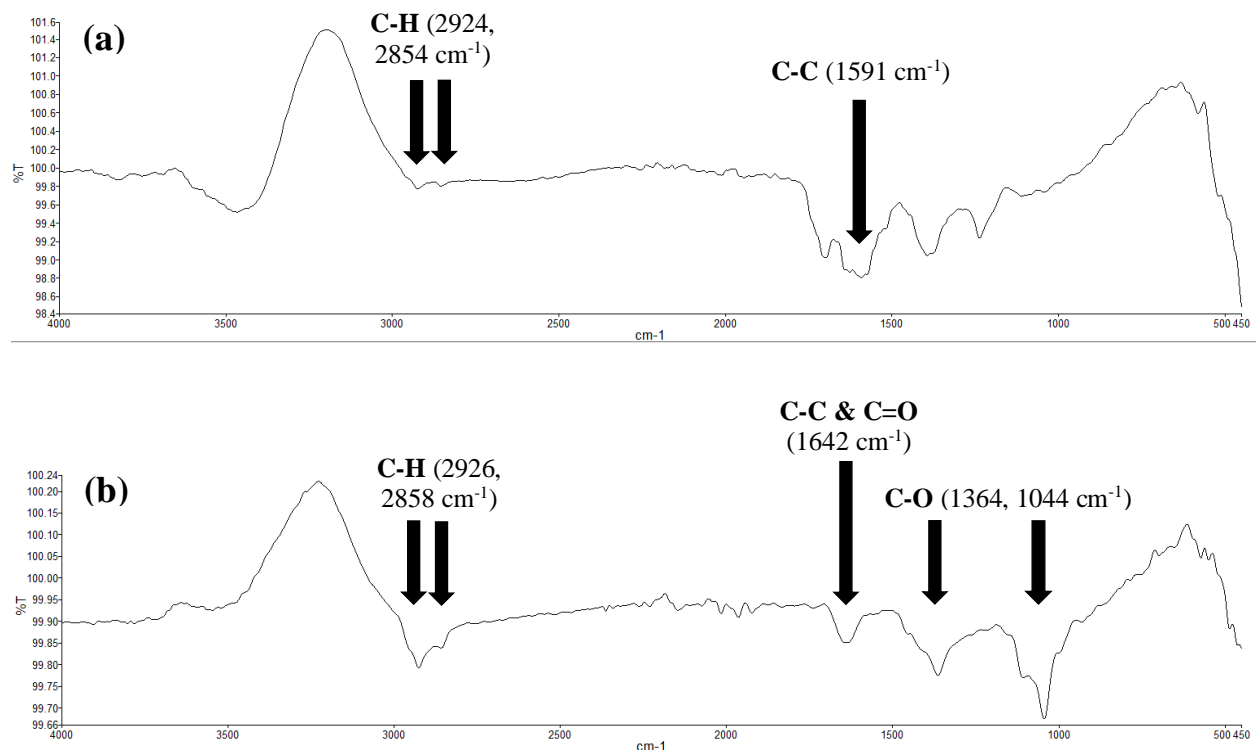
### I. UV-Vis Spectrophotometry

Results of the formate-functionalized GQDs demonstrate marked differences compared to non-functionalized GQDs. UV-Vis spectrophotometry demonstrates a prominent shift in absorbance, from a peak at 255 nm for

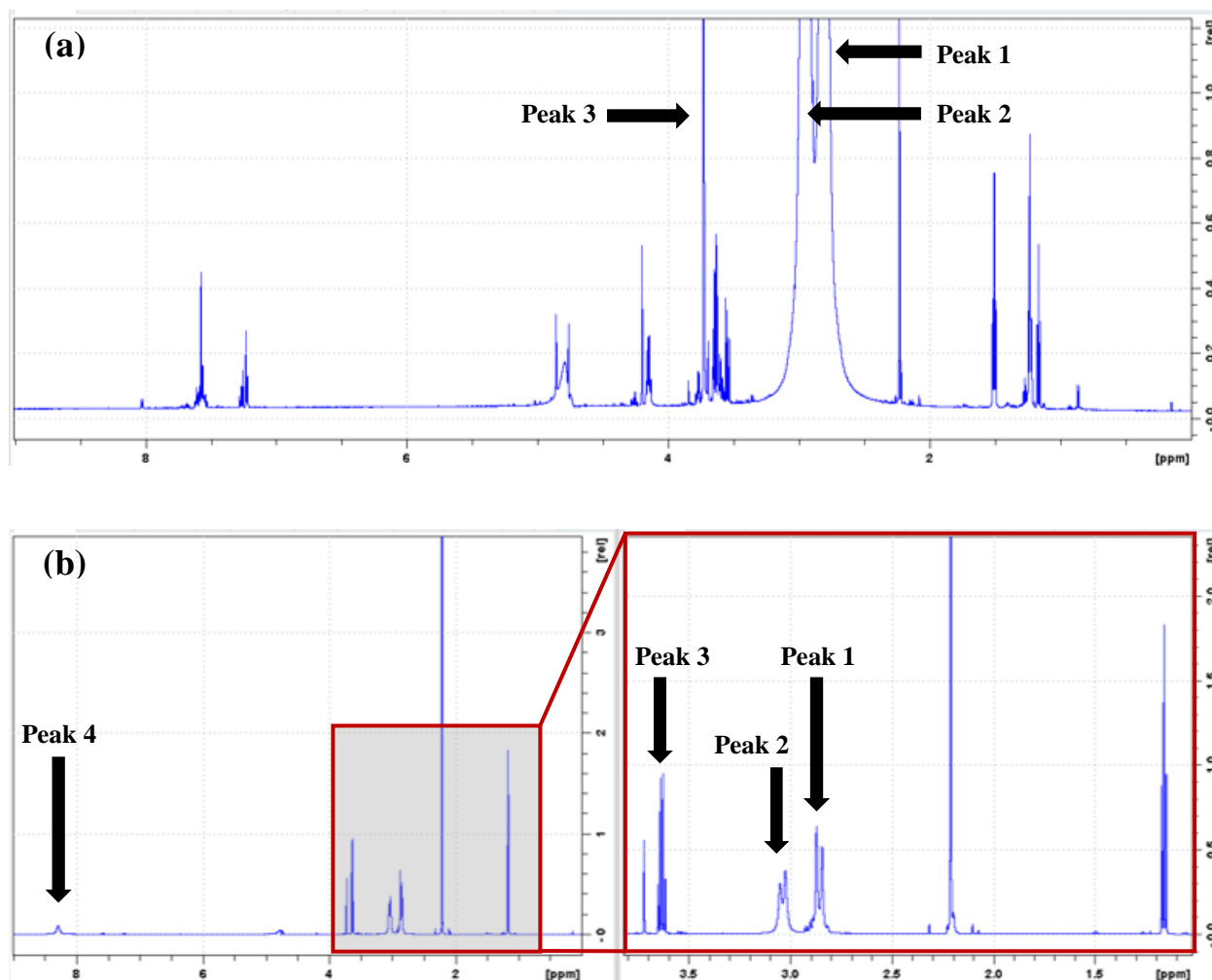
non-functionalized GQDs to a peak at 282 nm for functionalized GQDs (**Figure 1**). This redshift in absorbance demonstrates a decreased bandgap width in the formate-functionalized GQD as compared to the non-functionalized GQD. This is consistent with the theoretical results of Li *et al.* for other functional groups – namely, a longer wavelength of light implies a lower energy of emitted light and therefore a smaller bandgap.<sup>13</sup>

### II. IR-Spectroscopy Results

The IR spectrum of the non-functionalized GQDs shows a well-defined peak at 1591  $\text{cm}^{-1}$  corresponding to hybridized carbon-carbon bonds in the structure of graphene.<sup>15</sup> Additionally, peaks at 2924 and 2854  $\text{cm}^{-1}$  correspond to C-H bonds found in the edge structure of graphene, which appear in prior literature at 2922 and 2850  $\text{cm}^{-1}$ .<sup>15</sup> Additional peaks at  $\sim 1700$ ,  $\sim 1400$ , and  $\sim 1250$   $\text{cm}^{-1}$  are thought to be due to residual impurities, especially citric acid, from the synthesis procedure (**Figure 2a**).<sup>16</sup>



**Figure 2:** Image of the IR-spectroscopy graph for the (a) non-functionalized and (b) formate-functionalized GQD samples, with the peaks of interest highlighted and labeled. Different concentrations of functionalized/non-functionalized quantum dots were used.



**Figure 3:** Images of the <sup>1</sup>H NMR spectra for (a) GQDs and (b) functionalized GQDs. Peaks of interest in each graph are labelled, and the shaded portion of spectrum (b) was enlarged for clarity. Note: Samples were of different concentrations (approximately 5 mg/mL).

The IR spectrum of the formate-functionalized GQD sample shows a pair of small peaks at 2926 and 2858  $\text{cm}^{-1}$ , two broad peaks located at 1642 and 1364  $\text{cm}^{-1}$ , and a sharp peak at 1044  $\text{cm}^{-1}$  (**Figure 2b**).

The broad peak at 1642  $\text{cm}^{-1}$  is attributed to the hybridized carbon-carbon bonds inherent in graphene's structure,<sup>15</sup> as well as the C=O bonds inherent in formate.<sup>17</sup> Likewise, the two peaks at 2926 and 2858  $\text{cm}^{-1}$  correspond to C-H bonds in the GQDs.<sup>15</sup>

Finally, the peaks located at 1364 and 1044  $\text{cm}^{-1}$ , after comparison with the IR spectra for pure formic acid and graphene oxide,<sup>15,17</sup> are associated with C-O bonds.

### III. NMR Spectroscopy

Additionally, <sup>1</sup>H NMR spectroscopy was performed to characterize the GQDs' structure (**Figure 3**).

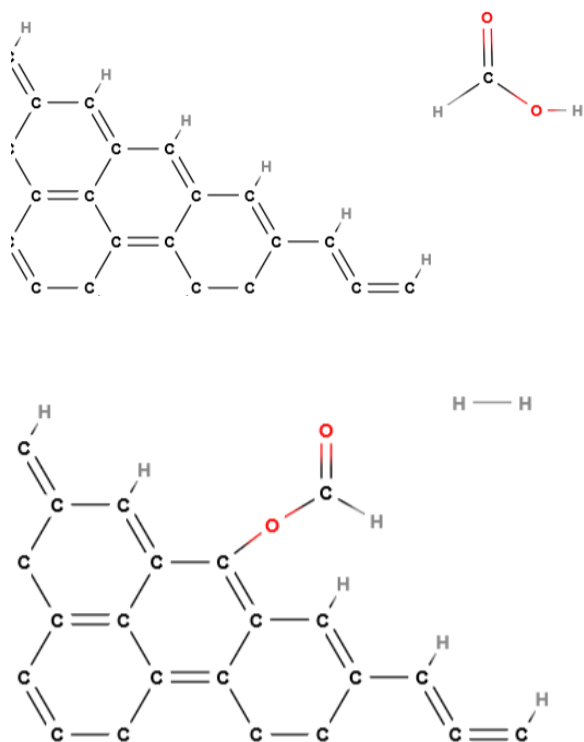
Comparison with NMR spectra for formic acid suggests that peak 4 in the functionalized GQD spectrum (**Figure 3b**), which does not appear in the non-functionalized GQD spectrum, corresponds to the formate functional group.<sup>18</sup>

NMR spectra for GQDs suggests that peaks 1, 2, and 3 in both **Figures 3a** and **3b** correspond to the structure of a non-functionalized GQD, independent of any functional groups.<sup>19</sup>

The additional peaks in both samples correspond to residual ethanol and H<sub>2</sub>O from the purification process.

### FURTHER ANALYSIS

The edge-functionalization protocol detailed here is derived from the process of electrophilic substitution, in which one electrophile replaces another in the edge structure of a GQD. Therefore, formate, an electrophile, will tend to replace hydrogen atoms in the edge structure of a GQD through electrophilic substitution, thus functionalizing the GQD (**Figure 4**).



**Figure 4:** Diagram of proposed mechanism, before (top) and after (bottom) GQD functionalization with formate.

As a part of this functionalization mechanism, the edge structures of non-functionalized GQDs are assumed to contain abundant C-H bonds. Typically, top-down synthesis methods yield significant oxygen-containing functional groups on the surface of GQDs, such as -OH and -COOH.<sup>20</sup> However, for

GQDs synthesized using a 1:3 molar ratio of citric acid to NaOH, the XPS data of Ahirwar *et al.* show an abundance of C-H bonds (by At. %) in the GQDs' surface structures. Furthermore, the XPS data also show that these C-H bonds are significantly greater in amount than C-O and C=O bonds, which represent oxygen-containing functional groups.<sup>14</sup> Thus, **Figure 4** depicts edges entirely containing C-H bonds for illustrational purposes only; our GQDs could contain low amounts of other functional groups.

Evidence of the functionalized GQD structure, shown in **Figure 4**, can be found in the IR spectra. Specifically, the peak at 1364 cm<sup>-1</sup> in **Figure 2b** is attributed to the C-O bond in the formate functional group. This peak appears, with a ~150 cm<sup>-1</sup> shift, in the IR spectrum for pure formic acid.<sup>17</sup> The observed shift can be attributed to the change from a C-OH bond to a C-O-C bond when the formate ion attaches to the GQD. Meanwhile, the peak at 1044 cm<sup>-1</sup> in **Figure 2b** aligns closely with the 1084 cm<sup>-1</sup> C-O peak observed in the IR spectrum for graphene oxide,<sup>15</sup> a compound that has oxygen bonded to graphene in a similar way as in our proposed structure.

Additionally, the C-O peaks at 1364 cm<sup>-1</sup> and 1044 cm<sup>-1</sup> seen in **Figure 2b** are not observed in **Figure 2a**, indicating that C-O bonds were formed during functionalization. Therefore, the C-O peaks observed in the spectrum for functionalized GQDs reinforce our predicted structure.

Further evidence of this proposed structure can be found in peak 4 of the <sup>1</sup>H NMR spectrum for functionalized GQDs (**Figure 3b**), which shows a slight shift to ~8.3 ppm from the peak at 8.44 ppm associated with formic acid.<sup>18</sup> This shift was attributed to the change from a C-OH bond in formic acid to a C-O-C bond linking the formate ester to the GQD. Therefore, the NMR spectra indicate the presence of formate functional groups in the edge structures of functionalized GQDs, as predicted.

### CONCLUSION

Through a facile experimental protocol, we have successfully executed edge

functionalization of GQDs with formate. Furthermore, we have used IR Spectroscopy and NMR Spectroscopy to analyze the structural properties of our functionalized GQDs and have found that they support our theoretical structure. Finally, we have analyzed the absorbance properties of these GQDs and confirmed a redshift in absorption, characteristic of a reduction in bandgap.

These findings hold the potential for several real-world applications, including bioimaging and single photon emission in quantum computing. These applications require the precise bandgap tunability and control over optical properties observed in this study. Furthermore, owing to the low toxicity of GQDs and the relatively low toxicity of formate, we predict acceptably low toxicity levels for use in such applications, especially compared to GQDs functionalized with other groups such as halogens.

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## AUTHOR CONTRIBUTIONS

JAB: Conceptualization, Methodology & Data Curation, Investigation, Validation, Formal Analysis, Visualization, and Writing - Original Draft & Editing.

JCC: Writing - Review & Editing, Funding Acquisition, Supervision & Resources, and Project Administration.

## FINANCIAL DISCLOSURE

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## COMPETING INTERESTS

The authors have declared that no competing interests exist.

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- [1] Geim, A.K.; Novoselov, K.S. *Nat. Mater.* **2007**, *6* (Mar 01), 183–191.
  - [2] Papageorgiou, D.; Kinloch, I.A.; Young, R.J. *Prog. Mater. Sci.* **2017**, *90* (Oct), 75–127.
  - [3] Castro Neto, A.H.; Guinea, F.; Peres, N.M.R.; Novoselov, K.S.; Geim, A.K. *Rev. Mod. Phys.* **2009**, *81* (1), 109.
  - [4] Wang, S.; Cole, I.S.; Li, Q. *RSC Adv.* **2016**, *6* (Sep 08), 89867–89878.
  - [5] Chen, W.; Lv, G.; Hu, W.; Li, D.; Chen, S.; Dai, Z. *Nanotechnol. Rev.* **2018**, *7* (2), 157–185.
  - [6] Zhu, S.; Song, Y.; Wang, J.; Wan, H.; Zhang, Y.; Ning, Y.; Yang, B. *Nano Today* **2017**, *13* (Apr), 10–14.
  - [7] Diao, S.; Zhang, X.; Shao, Z.; Ding, K.; Jie, J.; Zhang, X. *Nano Energy* **2017**, *31* (Jan), 359–366.
  - [8] Guo, J.; Zhu, H.; Sun, Y.; Tang, L.; Zhang, X. *J. Mater. Chem. A* **2016**, *4* (13), 4783–4789.
  - [9] Zhang, R.; Ding, Z. *Journal of Analysis and Testing* **2018**, *2* (1), 45–60.
  - [10] Zhao, S.; Lavie, J.; Rondin, L.; Orcin-Chaix, L.; Diederichs, C.; Roussignol, P.; Chassagneux, Y.; Voisin, C.; Müllen, K.; Narita, A.; Campidelli, S.; Lauret, J.S. *Nat. Commun.* **2018**, *9* (1).
  - [11] Ain, N.; Eriksson, M.O.; Schmidt, S.; Asghar, M.; Lin, P.C.; Holtz, P.O.; Syväjärvi, M.; Yazdi, G.R. *Nanomaterials* **2016**, *6* (11), 198.
  - [12] Kumar, G.S.; Roy, R.; Sen, D.; Ghorai, U.K.; Thapa, R.; Mazumder, N.; Saha, S.; Chattopadhyay, K.K. *Nanoscale* **2014**, *6* (Dec 05), 3384–3391.
  - [13] Li, Y.; Shu, H.; Niu, X.; Wang, J. *J. Phys. Chem. C* **2015**, *119* (44), 24950–24957.
  - [14] Ahirwar, S.; Mallick, S.; Bahadur, D. *ACS Omega* **2017**, *2* (11), 8343–8353.

- [15] Sudesh; Kumar, N.; Das, S.; Bernhard, C.; Varma, G.D. *Supercond. Sci. Technol.* **2013**, 26 (9). (Figure 1)
- [16] William E. Wallace, "Infrared Spectra" in **NIST Chemistry WebBook, NIST Standard Reference Database Number 69**, Eds. P.J. Linstrom and W.G. Mallard, National Institute of Standards and Technology, Gaithersburg MD, 20899, <https://doi.org/10.18434/T4D303>, (retrieved July 17, 2019).
- [17] Bergantini, A.; Pilling, S.; Rothard, H.; Boduch, P.; Andrade, D.P.P. *Mon. Not. R. Astron. Soc.* **2014**, 437 (3). (Figure 1)
- [18] Wishart, D.S.; Knox, C.; Guo, A.C.; Eisner, R.; Young, N.; Gautam, B.; Hau, D.D.; Psychogios, N.; Dong, E.; Bouatra, S.; Mandal, R.; Sinelnikov, I.; Xia, J.; Jia, L.; Cruz, J.A.; Lim, E.; Sobsey, C.A.; Shrivastava, S.; Huang, P.; Liu, P.; Fang, L.; Peng, J.; Fradette, R.; Cheng, D.; Tzur, D.; Clements, M.; Lewis, A.; De Souza, A.; Zuniga, A.; Dawe, M.; Xiong, Y.; Clive, D.; Greiner, R.; Nazyrova, A.; Shaykhutdinov, R.; Li, L.; Vogel, H.J.; Forsythe, I. *Nucleic Acids Res.* 2009, 37 (Database Issue).
- [19] Song, L.; Shi, J.; Lu, J.; Lu, C. *Chem. Sci.* **2015**, 6 (8), 4846-4850. (Figure 4b)
- [20] Li, L.; Wu, G.; Yang, G.; Peng, J.; Zhao, J.; Zhu, J.-J. *Nanoscale* **2013**, 5 (Jan 10), 4015-4039.