Understanding the Role of AgNO₃ Concentration and Seed Morphology to Achieve Tunable Shape Control in Gold Nanostars

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ABSTRACT: Gold nanostars are one of the most fascinating anisotropic nanoparticles. Nanostar morphology can be controlled by changing various synthetic parameters; however, the detailed growth mechanisms are not fully understood. Herein, we investigate this process in six-branched nanostars, focusing first on the properties of the single crystalline seed, which evolves to include penta-twinned defects as the gateway to anisotropic growth into 6-branched nanostars. In particular, we report on a high-yield seed-mediated protocol for the synthesis of these particles with high monodispersity in the presence of Triton-X, ascorbic acid, and AgNO₃. Detailed spectroscopic and microscopic analyses have allowed the identification of several key intermediates in the growth process, revealing that it proceeds via penta-twinned intermediate seeds. Importantly, we report the first experimental evidence tracking the location of silver with sub-nanometer resolution and prove its role as stabilizing agent in these highly branched nanostructures. Our results indicate that metallic silver on the spikes stabilizes the nanostar morphology, and that the remaining silver, present when AgNO₃ is added at high concentration,

deposits on the core and between the base of neighboring spikes. Importantly, we also demonstrate the possibility to achieve monodispersity, reproducibility, and tunability in colloidal gold nanostars that are substantially higher than previously reported, which could be leveraged to carry out holistic computational-experimental studies to understand, predict, and tailor their plasmonic response.

INTRODUCTION

Among the various types of gold nanoparticle systems providing features facilitating field localization, gold nanostars have been widely recognized owing to their tunable plasmon resonances, which have the potential to be extended from the visible to the infrared.¹⁻⁴ Generally speaking, there is a strong correlation between optical properties and morphology in plasmonic nanoparticles, with sharp edges and tips being further able to create localized electric field enhancement in proximity to the nanoparticle's surface. Due to the structure- and size-dependence of the localized surface plasmon resonance (LSPR) bands, morphological control during the synthesis is very important.⁵⁻⁶ For instance, we have observed how small differences in the shape and number of branches of a nanostar lead to drastic shifts in the LSPR band position and width, with highly branched nanostars being characterized by broad and blue shifted resonances.⁷ For this reason, an increasing number of synthetic procedures have been developed to control size and shape of plasmonic nanoparticles, among which both seeded and non-seed mediated methods have shown to afford morphological tunability.⁸⁻¹⁰ Importantly, it is crucial to obtain colloidal nanoparticles with high yield and reproducibility and large batch monodispersity. However, morphological control in gold nanostars is still not satisfactory.^{3, 11-12} To improve monodispersity and reproducibility in gold nanostars, it is necessary to develop a fundamental understanding of the role of the chemical variables and their impact on the growth mechanism, and to investigate

the key factors affecting growth. One of the issues in achieving this goal is the difficulty to trap reaction intermediates due to the generally fast reaction kinetics for these particles, and to understand, as a consequence, how any ill-defined morphology can affect the final products.¹³ One of the goals of this work was therefore to develop a fundamental understanding of the growth mechanism of these nanoparticles, and to identify a simple synthetic methodology that could yield highly monodispersed gold nanostars with consistent reproducibility.

The use of Ag⁺ is common in seed-mediated syntheses of gold nanostars to tune spike morphology up to a certain length by gradually increasing AgNO₃ concentration in the growth solution.¹⁴ This phenomenon is similar to what observed in gold nanorods.¹⁵⁻¹⁷ However, while several mechanisms have been proposed to explain in detail the role of AgNO₃, they are still highly debated despite the decade-long scientific efforts.¹⁶ For example, a face-specific cetyltrimethylammonium-Br-Ag+ capping complex was hypothesized to be formed to block specific gold facets thus leading to the formation of nanorods.¹⁸ In another proposed mechanism, underpotential deposition (UPD) of monolayers or sub-monolayers of silver on the gold nanorod surface was invoked as the factor leading to anisotropic growth.¹⁹ In this mechanism, silver deposits on the surface of the nanorods and selectively blocks selected facets, such as the {110}, rather than others, for instance {100} or {111}.²⁰ Unfortunately, while the presence of trace amounts of silver has been reported on the surface of the gold nanorods,²⁰⁻²² a mechanistic investigation on the role of AgNO₃ at different concentrations in the growth of gold nanorods or nanostars has not been carried out yet. In this respect, structural and elemental characterization could provide essential insight into the growth mechanism, ultimately allowing to improve the monodispersity and reproducibility of gold nanostars.

In this manuscript, we describe a novel type of gold nanostars, possessing only an average of 6branches (see Figure S1 for the definition of *average*), and elucidate the role of the synthetic parameters in the surfactant-based seed-mediated protocol employed to synthesize them. We also propose a growth mechanism that focuses on the properties of the seeds and how they affect the final nanoparticle morphology. These nanoparticles show exceptionally high monodispersity compared to nanostar systems reported before,²³⁻²⁶ and can be synthesized with high reproducibility. The reduced number of spikes, whose length and shape can be rationally controlled, limits side-by-side spike cross-talk, thus reducing LSPR peak broadening, and enables establishing fundamental relationships between morphology and plasmonic properties, which has been not possible so far due to the irreproducibility of the traditional synthetic protocols for gold nanostars.²⁷ For these reasons, these nanoparticles represent the first example of branched nanostructure that can be synthesized by design to possess pre-determined physical and optical properties. Based on this potential, it is important to understand the role of the reagents and their interplay during nanostar growth. To address this need, we performed a systematic study to determine how the concentrations of the surfactant (Triton X), the reducing agent (ascorbic acid), and AgNO₃, and their interaction with the evolving seeds affect the morphology, and thus the LSPR bands, of the resulting gold nanostars. By investigating the mechanism of gold nanostar formation in this surfactant-mediated synthesis, we propose a kinetically-controlled process as the basis of the growth.

RESULTS AND DISCUSSION

The two main recognized parameters affecting the growth of anisotropic gold nanostructures are the properties of the seeds and the concentration of AgNO₃. In the specific synthesis explored in this work, investigating their interplay with the added surfactant (Triton X) and the reducing agent

(ascorbic acid) also provides useful insight. To us however, the most important goal was to understand how seed evolution and silver concentration and position affect growth. Therefore, we first carried out the synthesis varying the concentrations of the four main reagents (i.e., Triton X, ascorbic acid, AgNO₃, and seeds) and characterized the obtained nanoarticles to extrapolate trends to identify the optimal synthetic conditions to achieve 6-branched nanostars, the ideal starting point for a more in-depth analysis. In this seed-mediated synthesis, seed and growth solutions were prepared separately but with equal concentration of surfactant, starting from concentration values similar to what reported in the initial manuscript of Pallavicini *et al.* but departing from them to achieve monodispersity and eliminate byproducts.²⁸ By varying the concentration of the four variables independently, we have investigated and determined how to obtain highly monodispersed 6-branched gold nanostars. While we are aware that multiple parameter spaces could likely provide ideal conditions to achieve similar results, we deemed it beyond the scope of this work to search for additional concentration values, considering more important to instead focus on understanding the growth mechanism in this specific set of conditions. In **Figure 1**, we have investigated the role of Triton X and ascorbic acid by modifying their concentrations while keeping the concentration of AgNO₃ and seeds constant at 100 μ M and 0.14 nM, respectively. We have examined four different concentrations of Triton X (0.01 M, 0.04 M, 0.15 M, and 0.3 M), and three different concentrations of ascorbic acid (0.8 mM, 1.6 mM, and 3.9 mM). In Figure 1, the concentration of Triton X varies along a column while the concentration of ascorbic acid is kept constant. For example, in **column 1** of **Figure 1**, the concentration of Triton X was increased from 0.01 M to 0.3 M (0.01 M (1a), 0.04 M (1d), 0.15 M (1g), and 0.3 M (1j)), while the concentration of ascorbic acid was kept constant (0.8 mM). On the other hand, the concentration of Triton X was kept constant throughout a row while the concentration of ascorbic acid was

varied. For example, in **row 1** of **Figure 1**, the concentration of ascorbic acid was increased from 0.8 mM to 3.9 mM (0.8 mM (1a), 1.6 mM (1b), and 3.9 mM (1c)) while the concentration of Triton X was kept constant (0.01 M).

TEM micrographs in Figure 1 reveal that the 6-branched morphology can be obtained only at ideal concentrations of both surfactant (Triton X) and reducing agent (ascorbic acid). In a column of Figure 1, when the concentration of Triton X was increased at a constant ascorbic acid concentration, the morphology changed from polyhedral nanoparticles to 6-branched stars. For example, in column 1 (Figure 1a, d, g, and j), the morphology of the nanoparticles changed from polyhedral to 6-branched stars at very high Triton X (0.3 M) and very low ascorbic acid (0.8 mM) concentration. However, the change in morphology from polyderal to 6-branched stars was also observed at moderately high Triton X (0.15 M) and moderate ascorbic acid (1.6 mM) concentration in column 2 (Figure 1h), and at moderately low Triton X (0.04 M) and high ascorbic acid (3.9 mM) concentration in column 3 (Figure 1f). Interestingly, at a very low concentration of Triton X (0.01M) and very high concentration of ascorbic acid (3.9 mM) (Figure 1c) multibranched stars were formed, where multiple (n >> 6) branches were grown from the central core. Complex hyperbranched nanoparticles, with multiple side branches on each of the six main branches (Figure 1k, and 1i), were formed when both Triton X and ascorbic acid concentration were higher than the ideal concentrations for 6-branched stars formation. Multibranched hollow gold nanoparticles were formed at a very high concentration of Triton X and ascorbic acid (Figure 11); we are still investigating their mechanism of formation. We observed similar morphology changes from polyhedral to 6-branched stars to complex hyperbranched nanoparticles in a row when the ascorbic acid concentration was increased at constant Triton X concentration (Figure 1g-i).



Figure 1. (a-1) TEM images of the gold nanoparticles where the concentration of the two chemical variables (Triton X and ascorbic acid) were progressively changed. In a column, the concentration of Triton X (TX) was varied from 0.01 to 0.3 M while it was kept constant in a row. For example, the concentration of Triton X in column 1 were 0.01 M (a), 0.04 M (d), 0.15 M (g), and 0.3 M (j). On the other hand, the concentration of ascorbic acid (AA) was kept constant in a column while it was increased from 0.8 mM to 3.9 mM in a row. For example, the concentration of ascorbic acid in row 1 were 0.8 mM (a), 1.6 mM (b), and 3.9 mM (c). The concentrations of AgNO₃ (100 μ M) and seeds (0.14 nM) were kept constant.

At ideal ascorbic acid (and AgNO₃) concentrations for 6-branched nanostar synthesis, the final morphology of the particles depends strongly on the concentration of Triton X. We observed that multibranched stars were formed in the absence of Triton X (Figure S2), while hyperbranched nanostars were formed at high Triton X concentration (Figure 1k). In agreement with previous reports, it appears that at very low concentration of Triton X, gold ions are not tightly bound to the surface by the surfactant Triton X and can be easily reduced by ascorbic acid.²⁸ By increasing the surfactant concentration above the critical micelle concentration (CMC) of Triton X (0.3 mM), the number of Au-encapsulating Triton X micelles increases,²⁹ thus decreasing the amount of free Au ions. The reduction in available free Au ions leads to more controllable gold reduction and branch generation during the growth process. A similar effect has been observed in other surfactantmediated gold nanostars syntheses, in which morphological changes from polyhedral to branched stars were also observed at increasing surfactant concentration.²⁶ For gold nanorods, it has been reported that a high concentration of surfactant (cetyltrimethylammonium bromide, CTAB) is necessary to achieve high aspect ratios, as high amounts of CTAB limit the number of free Au ions and reduce secondary nucleation events thus leading to the formation of longer nanorods.³⁰ However, because it is still unclear how micelle encapsulation for 6-branched nanostars occurs, we can only hypothesize that at extremely high concentrations of Triton X (>0.15 M) unzipping of the surfactant at the surface of the spikes may be taking place (mediated by the excess surfactant in solution) leading to the generation of secondary nucleation events on the spikes,³¹ and thus resulting in the formation of hyperbranched nanostars (Figure 1k).

Ascorbic acid can tune the morphology of the resulting nanoparticles as well, with higher amounts leading to the desired 6-branched nanostars. We believe this to be due to the increase in negative charge on the growing seeds due to excess ascorbic acid in the growth solution, resulting in the migration of gold atoms toward low surface energy facets, such as {111}, from high surface energy facets like {110}, or to an increase in crystal growth kinetics at higher ascorbic acid concentration.³²⁻³³ Clearly, additional experiments will be necessary to provide further evidence on the nature of the observed nanostar reshaping.

Having established the ideal parameter space for Triton X and ascorbic acid, we then focused our investigation on the determination of seed quality (size and crystallinity) (**Figure 2**). To avoid Ostwald ripening of the seeds on the grid during TEM analysis, we employed *n*-pentanethiol as a capping agent to quench the reaction and stabilize the seeds, as previously done to analyze the morphology of growth intermediates. HRTEM micrographs of the seeds disclose that most of the particles have single crystalline morphology, with an interplanar spacing of 0.23 nm, typical of (111) plane in FCC gold, and diameters of around 3 nm (**Figure 2a**). We have observed that at moderate ascorbic acid concentrations, ideal to produce 6-branched nanostars, the *in situ* evolution of single crystalline seeds into five-fold twinned seeds occurs (**Figure 2b**). On the other hand, at very high ascorbic acid concentration the intermediate seed possesses multiple twin defects (**Figure 2d**), but these conditions do not lead to 6-branched nanostars, rather to multibranched particles. Therefore, although multiple nucleation centers at low surface energy {111} facets are necessary for spike growth, their identity and number is what determines the final nanostar morphology.

We also followed spike growth over time by TEM. It has been proposed that twinned seeds may be fundamental to ensure spike growth on nanostars;^{14, 34} however, the direct correlation between defect nature and nanostar morphology has never been shown. We have therefore carried out a detailed analysis of the crystallographic properties of the seeds to correlate them to the resulting nanostar product. The morphology was investigated 10 s after addition of Triton X, ascorbic acid, and AgNO₃ for 6-branched stars and multibranched stars, leading to the products in Figure 2b and 2d. The morphology of the seed turned from single crystalline to twinned crystal, with all gold planes belonging to the {111} family (Figure 2c, and 2e). To be specific, penta-twinned intermediate seeds were formed when the concentration of Triton X, ascorbic acid, and AgNO₃ were ideal for the formation of 6-branched stars (Figure 2b-c), whereas multiply twinned intermediate seeds having multiple $\{111\}$ facets were formed when the concentration of Triton X, ascorbic acid, and AgNO₃ were kept at values observed to produce multibranched stars (Figure **2d-e**). The formation of multiple {111} facets in a multiply twinned intermediate supports our hypothesis that at high ascorbic acid concentration multiple low energy facets {111} are formed, whereas the reduction rate of gold (III) at 0.15 M Triton-X, 1.6 mM ascorbic acid, and 100 µM AgNO₃ leads only to the formation of penta-twinned defects. Interestingly, STEM micrographs of 6-branched nanostars show an anisotropic growth over the {111} facets on either side of the twin boundaries for penta-twinned intermediate seeds (Figure 2f-i), which arises due to low twinning energy and angle strain.³⁵ The corresponding fast Fourier transform image further confirms that the two {111} crystal facets are oriented with a common {111} twin plane (Figure 2h). This is further confirmed by the STEM micrographs of the tips (Figure 2j-p) which provide a clear view of the crystallographic structure of the penta-twinned seeds from which the spikes were grown, as illustrated in Figure 2q. In comparing Figure 2j-p and 2q, one can observe that while in principle these particles should possess either five or seven spikes, the distribution reported in Figure 2q identified quite comparable numbers of 5-, 6-, and 7-branched nanostars, with higher frequency of 6-branched nanostars. Further TEM tomography will allow us to better evaluate these numbers and clearly tease out the correlation between seed defects and spike numbers.



Figure 2. a) TEM micrographs of the seeds illustrating that their average diameter was 3 nm. HRTEM micrograph (inset) shows the single crystalline morphology of the seed, with interplanar spacing characteristic of {111} planes in FCC gold. (b-c) TEM and HRTEM micrographs of pentatwinned intermediate seeds of 6-branched stars. (d-e) TEM and HRTEM images of multiply twinned intermediate seeds of multibranched stars. (f-i) STEM images of the spike showing the presence of only a twin boundary where the FFT (inset) and lattice fringes represent {111} planes reversely oriented with respect to a common twin plane. j) TEM image of the 6-branched stars. k) STEM micrograph of the tip showing five twinning planes, which indicates that the spike is forming on a twinning axis of a pentagonal unit. l-p) STEM micrographs of the tips shows four

twinning planes, indicating that the spike is forming on a twinning axis of a tetrahedral unit. Red arrows show the twin planes. q) Representation of spike growth from decahedral seeds. Top image: Schematic side view identifying the spike growth directions on seeds such as that reported in Figure 71. A maximum number of five spikes can grow along the equatorial directions identified by the red arrows, which are determined by four neighboring facets (T1, T2, T3, and T4). Bottom image: Schematic top view representing the structure reported in Figure S12, in which the spike growth is expected to occur from penta-twinned defects, in which five neighboring facets are present (T1, T2, T3, T4, and T5).

We have observed that polyhedral nanoparticles were formed in the absence of AgNO₃, thus indicating that AgNO₃ is necessary for the formation of the desired 6-branched nanostars (Figure S3). AgNO₃ concentration affects the stability of the particle as well, as we observed spherical impurities at or below 30 µM AgNO₃, likely due to nanostar reshaping. However, we have not seen the transformation from polyhedral nanoparticles to 6-branched stars by increasing the concentration of AgNO₃ at non-ideal concentrations of Triton-X and ascorbic acid (Figure S4). We have investigated the effect of AgNO₃ when the concentrations of the other reagents are optimized to obtain highly monodisperse and reproducible 6-branched stars (0.15 M Triton X, 1.6 mM ascorbic acid, and 0.14 nM seeds) (Figure 1h). The concentration of AgNO₃ was increased in small increments of 10 µM to determine the possibility of finely-tuning the morphology through AgNO₃ (Figure 3 and Figure S5). The spike length increased rapidly by roughly 8 nm by increasing AgNO₃ concentration in 10 μ M increments from 30 to 60 μ M AgNO₃. Then it slowly increased by around 2-3 nm by increasing the amount of AgNO₃ from 60 to 100 μ M in 10 μ M steps (Figure 3k and Figure S6). This evolution was followed by monitoring the red-shift of the longitudinal LSPR band in the UV-Vis spectrum (Figure 3j and Figure S5). The red-shift was also

accompanied by a visible change in the solution color from blue to brown as the concentration of AgNO₃ increased. At AgNO₃ concentration higher than 100 μ M, the spike length did not further increase (Figure 3j-k), rather a blue shift in the LSPR band, reported to indicate a shortening or thickening of the spike, was instead observed.^{6, 14} The blue shift of the LSPR at 110 μ M AgNO₃ concentration could also however be due to the deposition of atomic Ag on the core (*vide infra*), as the ascorbic acid in excess can reduce remaining silver ions resulting in the nanostar core diameter to increase from 25 to 35 nm. Vo-Dinh and coworkers reported that silver overgrowth on gold nanostars is possible and can blue shift the plasmon resonance of the longitudinal mode.³⁶



Figure 3. (a-i) TEM micrographs of nanostars formed under different AgNO₃ concentrations- 30 μ M (a), 40 μ M (b), 50 μ M (c), 60 μ M (d), 70 μ M (e), 80 μ M (f), 90 μ M (g), 100 μ M (h), and 110 μ M (i). The concentration of other two chemical variables (ascorbic acid and Triton-X) were 1.6 mM and 0.15 M, respectively. Scale bars are 20 nm (inset). j) UV–vis spectra (normalized) for each of the colloidal dispersions which shows a gradual red shift with increasing AgNO₃

concentration as the spike length is increased, and blue shift after 100 μ M AgNO₃ as silver is deposited on the core. k) Evolution of the average spike length as a function of AgNO₃. concentration.

To study the fate of silver and its role on the evolution of spike morphology, we first monitored the branch sharpness, which is the ratio between the spike widths at core and tip, observing that it increased from 1.2 to 1.9 (Figure S7) with increasing AgNO₃ concentration from 30 µM to 100 μ M, possibly due to the migration of gold atoms from the tip toward the core. This result further motivated us to study the evolution of the spike morphology. Spike growth was investigated in detail by arresting the reaction at different time points and examining the intermediates via TEM to correlate morphology evolution to LSPR position, when the concentration of Triton X, ascorbic acid, and AgNO₃ were 0.15 M, 1.6 mM, and 30 µM respectively (Figure 3 and Figure S8). TEM micrographs (Figure 3a-l and Figure S8a-l) at different time intervals reveal that the branches grew gradually and reached maximum length (100 nm) after 5 minutes. After that, they shrank, and the process was completed after 12 hours, with an overall shrinking in spike length by 30 nm, from 100 nm (5 min) to 70 nm (12 hours). The time-dependent evolution of this reaction was monitored by UV-Vis spectrophotometry (Figure 3m), which elucidated that the LSPR band gradually red shifted as the spike length increased between 30 seconds and 5 minutes. The LSPR band reached its maximum redshift to 1071 nm after 5 min, then blue shifted from 1071 nm to 734 nm after 12 hours. The blue shift was associated to the migration of gold atoms from high energy sites on the tip toward lower energy ones at the core, resulting in a decrease in spike length from 100 nm to 70 nm. The LSPR shift was also accompanied by a visual color change in the solution from blue to green to brown to blue, as observed before for the growth of multibranched gold

nanostars and gold nanorods when the spike length was reduced in the late stages of the reaction.⁴, 17, 22, 37

The nanostars morphology at 30 µM AgNO₃ concentration for a given growth time was further investigated by STEM (**Figure 4n-p**) to study spike morphology in detail. STEM micrographs revealed that the gold nanostars grown for 5 minutes contained spherical penta-twinned tips where twin boundaries having {111} facets bridging the sides with the tips can be observed (**Figure 4n**). However, after 6 hours the tip was observed to be less spherical (**Figure 4o**) and to become oblate after 12 hours, with no clearly distinguishable facets detectable, as the penta-twinned morphology of the tips disappeared after 6 hours. (**Figure 4o-p**). We believe that the driving force for these morphological changes is the surface energy minimization that is achieved by removing reactive edge atoms located at the twin boundaries of highly faceted penta-twinned spikes.^{15, 38}



Figure 4. a-1) TEM micrographs of gold nanostars when the concentrations of the growth solution are- Triton-X-0.15 M, ascorbic acid- 1.6 mM, and AgNO₃- 30 μ M for which growth was arrested at the indicated reaction times (30 sec (a), 1 min (b), 1 min 30 sec (c), 2 min (d), 5 min (e), 10 min (f), 30 min (g), 60 min (h), 120 min (i), 240 min (j), 480 min (k), and 720 min (l)). m) Corresponding UV–vis spectra (normalized) taken from each aliquot sample which indicates that an initial red shift of the longitudinal plasmon peak occurred, which reversed after 5 minutes, and was followed by a permanent blue shift. Scale bars are 20 nm. (n-p) STEM micrographs of the

spike at 5 min (n), 240 min (o), and 720 min (p) showing the morphology from twinned spherical tip to oblate tip.

To gain further insight into the evolution of the spikes, we have investigated in detail the changes in overall nanostar morphology at the 5-minute (Figure S9) and 12-hour (Figure 2b-d, 2f, and 2h) marks, by varying the concentration of AgNO₃, starting from the observation that maximum spike length is reached after 5 minutes and the minimum after 12 hours, for reactions with 30 µM AgNO₃. We have used five different additional concentrations of AgNO₃ (40 µM, 50 µM, 60 µM, 80μ M, and 100μ M), and observed that the spike length is maximized after 5 minutes (100 nm) for all AgNO₃ concentrations, with additional substantial shape reconstruction occurring at the 12hour time point. For nanostars synthesized with 30 μ M AgNO₃ the shape reconstruction was substantial, with spike length reduction from 100 nm to 70 nm and loss of the sphere at the tip (Figure 4n-p). However, the extent of deformation decreased with increasing the concentration of AgNO₃ from 30 µM, becoming the lowest for nanostars synthesized at 100 µM AgNO₃ (Figure **S9a-f**). For instance, while a 341 nm blue shift was observed for 30 μ M AgNO₃ stars, only a 6 nm blue shift was observed for 100 µM AgNO₃ stars (Figure 5a-f). We also observed visually that the color did not change from brown to blue when the concentration of AgNO₃ was kept at 100 µM AgNO₃. These results led us to postulate that the 5-minute morphology might be the kinetically trapped version of the thermodynamically-stable 12-hour morphology, and that deposited Ag atoms might reduce significantly the atom diffusion typically observed for highly energetic gold facets on gold nanoparticles. These observations also established the added important role for AgNO₃ (i.e. to stabilize the nanostar shape and size) beyond the well-known shape-inducing role. We attribute the shape reconstruction observed at longer reaction times for low AgNO₃ concentration to the fact that the highly energetic gold atoms at the tips can easily diffuse along

the spike migrating to more energetically favorable positions on the nanostar, such as the base of the spike. However, when the concentration of AgNO₃ was 100 μ M, the Ag atoms appeared to stabilize the highly energetic gold atoms, thus inhibiting their diffusion toward the core. A similar result was reported by Tong *et al.*, who observed a blue shift of the longitudinal plasmon peak of gold nanorods when they kept from 2 hrs. to 13 weeks.¹⁵ The possible reason behind the stability gained at above 30 μ M AgNO₃ concentration is that the presence of submonolayer silver atoms act as a protective agent for the underlying the gold atoms in form of Au-Ag_(UPD)-Cl on the surface of the nanoparticles.³⁹⁻⁴⁰



Figure 5. (a-f) UV-Vis spectra of 6-branched gold nanostars formed after 5 min and 720 min reaction time under different AgNO₃ concentration-30 μ M (a), 40 μ M (b), 50 μ M (c), 60 μ M (d), 80 μ M (e), and 100 μ M (f). UV-Vis spectrum shows a 341 nm, 212 nm, 99 nm, 42 nm, 12 nm, and 6 nm blue shift for 30 μ M, 40 μ M, 50 μ M, 50 μ M, 60 μ M, 80 μ M, and 100 μ M AgNO₃ respectively.

We have investigated further the role of Ag by using scanning transmission electron microscopyefficiency energy dispersive X-ray spectroscopy (STEM-EDS), to determine whether or not Ag exists as adsorbed species on the Au surface or becomes fully alloyed to Au on the nanostar spike. We also wanted to ascertain whether or not the deposition of increasing amounts of silver at high concentrations of AgNO₃ is responsible for the observed morphology stabilization and LSPR blue shifts. Figure 6 shows the STEM image of 6-branched gold nanostars having three different AgNO₃ concentrations (30, 100, and 110 μ M) along with corresponding STEM-EDS map showing the gold signal (red scale) and the silver signal (blue scale), which reveal that silver was alloyed with gold in the nanostars. Moreover, we have observed that the relative silver signal increased with increasing AgNO₃ concentration, going from 3.86% (2.21% at core, and 5.51% at spike) to 14.66% (13.87% at core, and 15.45% at spike) when the concentration of AgNO₃ was increased from 30 μ M to 100 μ M, and even further increased to 16.45% (18.94% at core, and 13.97% at spike) when the concentration of AgNO₃ was 110 µM. Interestingly, the line-scanned EDS elemental profiles of the spike and the core of 30 µM AgNO₃ showed that the amount of Ag was uniform throughout the spike and the core and (Figure 6a'-b'), While it was higher at the side wall of the spike for 100 µM AgNO₃, which supports our hypothesis that a submonolayer of silver stabilizes the surface Au atoms (Figure 6c'-d'). Moreover, silver deposition increased at core when the concentration of AgNO₃ was 110 μ M (Figure 6e'). This growth mechanism also supports the observed patterns in gold nanorod growth, where silver deposits on the side wall of the rod rather than the tips.²⁰ We further carried out an area scanned analysis of the EDS map to obtain more information about the amount of Ag present on the gold nanostars (Table S1). Area scan results of the spike at 30 µM AgNO₃ show 3.28% (A1), 8.84% (A2), and 4.12% (A3) of Ag present at the tip, side wall, and middle portion of the spike, respectively, which reveals that the

amount of Ag was almost uniform on the spike. On the other hand, the amount of Ag was significantly lower at the core (A4, 2.46 % Ag). However, we have seen an increase in the amount of Ag at the side wall of the spike of the stars at 100 μ M and 110 μ M AgNO₃ (36.87% for 100 μ M (A7) and 33.70% for 110 μ M (A15)) compared to the middle portion of the spike (23.61% for 100 μ M (A8) and 13.78% for 110 μ M (A16)). Interestingly, we have seen that the amount of Ag increased at core when the concentration increased from 100 μ M (8.34 % Ag (A10)) to 110 μ M (25.05 % Ag (A13)) which supports our hypothesis that the observed LSPR blue shift is also due to Ag deposition at the core (**Figure 3**).



Figure 6. (a) TEM micrograph and (b-i) HAADF-STEM micrograph and elemental maps of the spike (b-e) and the core (f-i) of 30 μ M AgNO₃ after 12 hours. (j) TEM micrograph and (k-r) HAADF-STEM micrograph and elemental maps of the spike (k-n) and the core (o-r) of 100 μ M AgNO₃ after 12 hours. (s-x) HAADF-STEM micrograph and elemental maps of the core of 100 μ M AgNO₃ after 12 hours. (a'-e') Line scan elemental profiles of 6-branched nanostars at 30 μ M (spike a', and core b'), 100 μ M (spike c', and core d'), and 110 μ M (core e') which reveal that Au

and Ag are miscible in all samples. At increasing AgNO₃ concentrations, metallic Ag first saturates deposition sites along the side wall of the tips, and then proceeds to deposit at the core.

The concentration of seeds to be added to the growth solution is also a very important parameter to monitor, as the seed is the primary nucleation center from which the spikes are formed. Moreover, it is reported that multibranched hollow gold nanostars can be formed in the absence of seeds.¹⁰ We have determined the concentration of seeds following a reported method,⁴¹ and investigated their effect by increasing their concentration from 0.02 nM to 0.14 nM, in 0.04 nM increments (Figure 7a-d and Figure S10). We determined that 0.14 nM is the smallest amount of seeds necessary to achieve 6-branched stars with high monodispersity (Figure 7d), which is associated to both spike number and spike length reduction (Figure 7a-d). On the contrary, multibranched stars were formed at 0.02 nM seed concentration (Figure 7a). Interestingly, the spike number never increased above six by increasing the concentration of seeds above 0.1 nM (Figure 7c). The high sample monodispersity achieved for 0.14 nM seeds was evidenced in the UV-Vis spectra reported in **Figure 7e** in the form of narrower LSPR bands (green curve) compared to what observed at lower seed concentrations. During the growth process, the availability of free gold atoms is very high when the concentration of seeds in the growth solution is low. These gold atoms can easily associate to the seeds and generate nucleation centers in high numbers, thus leading to the formation of multibranched stars. However, the availability of free gold atoms saturates at or above 0.1 nM seed concentration, thus leading to nanostars with fewer spikes and shorter spike lengths, as larger amounts of seeds at equal Au concentration create more primary growth centers. A similar observation was reported by Barbosa et al. who noted that the branching of PVP-capped gold nanostars increased by decreasing seed concentration in the growth solution.⁴²

One of the most interesting aspects of this synthesis is the possibility to leverage the interplay between Triton X, ascorbic acid, and seeds to modify the number of branches in the nanostars. For instance, in **Figure 1** we have seen that the number of branches can be increased by either decreasing Triton X (Figure 1c, and 1f) or by increasing ascorbic acid (Figure 1g, and 1h) at constant seed concentration. In Figure 7 (7a-d) the number of branches was increased by decreasing the seed amount when the other variables were kept constant. To examine how these three variables are connected to each other, we ran two additional control experiments starting from the multibranched stars reported in Figure 7a and 7b. These nanoparticles were synthesized in conditions ideal to specifically obtain multibranched stars (0.15 M Triton X, 1.6 mM ascorbic acid, 100 µM of AgNO₃, and either 0.02 nM or 0.06 nM seeds). In our first control, we increased the concentration of Triton X from 0.15 M to 0.3 M, while the other concentrations were kept constant, and observed that in both cases the morphology changed from multibranched to 6branched stars (Figure 7f-g). In the second control experiment, we decreased the concentration of ascorbic acid form 1.6 mM to 0.8 mM keeping the other variables constant. In these conditions we did observe a decrease in the number of branches with decreasing ascorbic acid concentration (Figure 7h-i), but this was not sufficient to produce 6-branched nanostars at 0.02 nM seed concentration (Figure 7h). These results show that the basic process of forming the 6-branched stars can be tweaked by independently modifying the concentration of Triton X, ascorbic acid, and seeds, which provides a useful knob to rationally tuning morphology.



Figure 7. (a, d) TEM micrographs of gold nanostars synthesized by adding different amounts of seeds (0.02 nM (a), 0.06 nM (b), 0.1 nM (c), and 0.14 nM (d)) to the growth solution containing 0.15 M Triton X, 1.6 mM ascorbic acid, and 100 μ M of AgNO₃. (e) UV-Vis spectra (normalized) of the nanostars at different concentration of seeds showing a blue-shifted narrower LSPR band with increasing seed concentration, which indicates that lower branching and higher monodispersity of the stars can be achieved at 0.14 nM seeds concentration. (f, g) TEM micrographs of gold nanostars synthesized by adding different amounts of seeds (0.02 nM (f), and 0.06 nM (g)) to the growth solution containing 0.3 M Triton X, 1.6 M ascorbic acid, and 100 μ M of AgNO₃. (h, i) TEM micrographs of gold nanostars synthesized by adding different amounts of seeds (0.02 nM (f), and 0.06 nM (i)) to the growth solution containing 0.3 M Triton X, 1.6 M ascorbic acid, and 100 μ M of AgNO₃. (h, i) TEM micrographs of gold nanostars synthesized by adding different amount of seeds (0.02 nM (h), and 0.06 nM (i)) to the growth solution containing 0.15 M Triton X, 0.8 mM ascorbic acid, and 100 μ M of AgNO₃. A decrease in spike number from figures a and b is evident. At low ascorbic acid concentration (h and i) 6-branched nanostars cannot be obtained at low seed concentration, as opposed to the other conditions. Scale bars are 20 nm in figures f-i.

CONCLUSION

In this study, we have reported a detailed systematic study of the seed-mediated growth mechanism of 6-branched gold nanostars. The interplay of various synthetic parameters (Triton X, ascorbic acid, AgNO₃, and seed concentrations) is shown to influence the growth and final morphology of stars. After extrapolating the fundamental growth parameters a identifying the ideal parameter space for Triton X, ascorbic acid, AgNO₃, and seeds to yield to the expected 6-branched products, we explored in detail the role of the nature of the seeds and the concentration of AgNO₃. Analysis of the kinetic data and microscopic images reveals that during this synthesis the single crystalline seeds transform into two different types of intermediate seeds - multiply-twinned intermediate seeds for multibranched stars and penta-twinned intermediate seeds for 6-branched stars. Moreover, the evolution of the spikes of 6-branched stars shows that the shape and size of the spikes are highly dependent on AgNO₃ concentration, proceeding via a common intermediate having maximum spike length (100 nm), with final spike length determined by the amount of AgNO₃ in solution. We have demonstrated the important role of silver in the stabilization of the evolving crystal, confirmed by the observation that at low AgNO₃ concentrations kineticallytrapped nanostars, at 5-minute time points, evolve substantially before reaching thermodynamic equilibrium at 12 hours. Most importantly, the presence of metallic silver both at the side walls of the spikes (at low AgNO₃ concentration) at also at the core (at high AgNO₃ amounts) reveals in detail the importance of this reagent in tuning nanostar morphology: Increasing amounts of deposited silver appear to stabilize the five-fold twinned morphology, which would instead be lost at low Ag concentrations due to the substantial strain present in highly-curved twinned regions. These nanostars display high monodispersity, batch-to-batch reproducibility, and plasmon tunability between the visible and the short wave infrared, which could prove extremely useful in

several quantitative applications or fundamental studies for which the *rational* design of multibranched nanoparticles is necessary. Looking ahead, it is possible to envision how this synthesis could lend itself as a model for the implementation of machine learning tools in materials design.

EXPERIMENTAL SECTION

Materials. Gold (III) chloride trihydrate (HAuCl₄.3H₂O), silver nitrate (AgNO₃; 99.995%), L(+)-ascorbic acid, sodium borohydride (NaBH₄), and TritonX-100 were purchased from Sigma-Aldrich. All these chemicals were used without further purification. Ultrapure MilliQ water (18.2 M Ω ·cm) was used in all syntheses. All glassware was aqua regia cleaned before each synthesis.

Instrumentation. Absorption spectra were collected on a Thermo Scientific Evolution 300 UV-Visible spectrophotometer using a quartz cuvette with 1 cm path. Nanoparticle morphology was determined using a Topcon 002B TEM. HRTEM analysis was performed on a JEOL 2010 F highresolution transmission electron microscope. The particle sizes (spike length, spike width) were analyzed using ImageJ. Particle morphology was analyzed using Gatan DigitalMicrograph (TM) 3.11.1 for GMS 1.6.1. The values of average *d* spacing were obtained from Fourier transform analysis of high-magnification images.

STEM were obtained using a FEI Titan Themis transmission electron microscope (TEM) operated at 200kV. Energy dispersive X-ray spectroscopy maps (EDX maps) were obtained in scanning mode of TEM (STEM). The point resolution in this aberration-corrected mode is 0.08nm. 1nm resolution EDX maps with an average beam current of 100pA are routine with this microscope.

Synthesis of 6-branched Gold Nanostars. The synthesis of 6 branched gold nanostars was first proposed by Pallavicini *et al.*²⁸ However, their nanoparticles did not display sufficient purity and

monodispersity. We therefore modified and varied the synthetic parameters to achieve high monodispersity and tunable morphology. Briefly, the seed solution was prepared by addition of a freshly prepared ice-cold solution of NaBH₄ (0.6 ml, 0.01 M) into a solution mixture of HAuCl₄ (10 ml, 0.25 mM) and Triton X, whose concentration was ranging from 0.01 to 0.3 M. The solution turned immediately from pale yellow to orange after addition of NaBH₄. The mixture was stirred for 2 minutes and aged for 10 minutes at 4°C before use.

The growth solution was prepared by adding 0.4 ml of 25 mM HAuCl₄ solution to a 20 ml Triton-X solution where the concentration of Triton-X was the same for both the seed and growth solutions. This step was followed by addition of ascorbic acid (ranging from 0.8 to 3.9 mM), AgNO₃ (ranging from 30 to 110 μ M), and Au seeds (ranging from 0.02 nM to 0.14 nM) to the growth solution. The solution was stirred for 12 hours and then centrifuged at 4,000 g for 10 min and dispersed with 5 ml of Ultrapure MilliQ water (18.2 MQ·cm).

Arrested Growth Studies. We have observed that *n*-pentanethiol works best to trap reaction intermediates compared to the more commonly used mPEG-SH (MW 5000), as we observed surface modifications and nanoparticle restructuring using the latter. Briefly, an aliquot (1 ml) of growth solution at the desired time was added to the solution of 1 ml 8.4 mM *n*-pentane in ethanol. Then, the solution was mixed well and centrifuged at 8000 g for 10 minutes. UV-Vis and TEM analysis of the particles were performed immediately after re-dispersion of the particles in 500 μ L of MilliQ water.

ASSOCIATED CONTENT

Additional TEM micrographs and relevant statistical analyses of 6-branched nanostars and their reaction intermediates.

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REFERENCES

1. Indrasekara, A. S. D. S.; Meyers, S.; Shubeita, S.; Feldman, L. C.; Gustafsson, T.; Fabris, L., *Nanoscale* **2014**, *6* (15), 8891-8899.

2. Wang, Y.; Serrano, A. B.; Sentosun, K.; Bals, S.; Liz-Marzán, L. M., *Small* **2015**, *11* (34), 4314-4320.

3. Kedia, A.; Kumar, P. S., Journal of Materials Chemistry C 2013, 1 (30), 4540-4549.

4. Khoury, C. G.; Vo-Dinh, T., *The journal of physical chemistry. C, Nanomaterials and interfaces* **2008**, 2008 (112), 18849-18859.

5. Trigari, S.; Rindi, A.; Margheri, G.; Sottini, S.; Dellepiane, G.; Giorgetti, E., *Journal of Materials Chemistry* **2011**, *21* (18), 6531-6540.

6. Hsiangkuo, Y.; Christopher, G. K.; Hanjun, H.; Christy, M. W.; Gerald, A. G.; Tuan, V.-D., *Nanotechnology* **2012**, *23* (7), 075102.

7. Atta, S.; Tsoulos, T. V.; Fabris, L., *The Journal of Physical Chemistry C* **2016**, *120* (37), 20749-20758.

8. Kawamura, G.; Yang, Y.; Fukuda, K.; Nogami, M., *Materials Chemistry and Physics* 2009, 115 (1), 229-234.

9. Ndokoye, P.; Li, X.; Zhao, Q.; Li, T.; Tade, M. O.; Liu, S., Journal of Colloid and Interface Science 2016, 462, 341-350.

10. Blanch, A. J.; Döblinger, M.; Rodríguez-Fernández, J., Small 2015, 11 (35), 4550-4559.

11. Ramsey, J. D.; Zhou, L.; Kyle Almlie, C.; Lange, J. D.; Burrows, S. M., New Journal of Chemistry 2015, 39 (12), 9098-9108.

12. Guerrero-Martínez, A.; Barbosa, S.; Pastoriza-Santos, I.; Liz-Marzán, L. M., Current Opinion in Colloid & Interface Science **2011**, *16* (2), 118-127.

13. Sajitha, M.; Vindhyasarumi, A.; Gopi, A.; Yoosaf, K., RSC Advances 2015, 5 (119), 98318-98324.

14. Yuan, H.; Ma, W.; Chen, C.; Zhao, J.; Liu, J.; Zhu, H.; Gao, X., *Chemistry of Materials* **2007**, *19* (7), 1592-1600.

15. Tong, W.; Katz-Boon, H.; Walsh, M. J.; Weyland, M.; Etheridge, J.; Funston, A. M., *Chemical Communications* **2018**, *54* (24), 3022-3025.

16. Tong, W.; Walsh, M. J.; Mulvaney, P.; Etheridge, J.; Funston, A. M., *The Journal of Physical Chemistry C* 2017, *121* (6), 3549-3559.

17. Nikoobakht, B.; El-Sayed, M. A., Chemistry of Materials 2003, 15 (10), 1957-1962.

18. Hubert, F.; Testard, F.; Spalla, O., Langmuir 2008, 24 (17), 9219-9222.

- 19. Liu, M.; Guyot-Sionnest, P., The Journal of Physical Chemistry B 2005, 109 (47), 22192-22200.
- 20. Orendorff, C. J.; Murphy, C. J., *The Journal of Physical Chemistry B* **2006**, *110* (9), 3990-3994.
- 21. Jackson, S. R.; McBride, J. R.; Rosenthal, S. J.; Wright, D. W., Journal of the American Chemical Society 2014, 136 (14), 5261-5263.
- 22. Sau, T. K.; Murphy, C. J., Langmuir 2004, 20 (15), 6414-6420.
- 23. Xie, J.; Lee, J. Y.; Wang, D. I. C., Chemistry of Materials 2007, 19 (11), 2823-2830.
- 24. Bakr, O. M.; Wunsch, B. H.; Stellacci, F., Chemistry of Materials 2006, 18 (14), 3297-3301.
- 25. Jeong, G. H.; Lee, Y. W.; Kim, M.; Han, S. W., *Journal of Colloid and Interface Science* **2009**, *329* (1), 97-102.

26. Pandian Senthil, K.; Isabel, P.-S.; Benito, R.-G.; Abajo, F. J. G. d.; Luis, M. L.-M., *Nanotechnology* **2008**, *19* (1), 015606.

- 27. Ted V., T.; Supriya, A.; Maureen J., L.; Philip E., B.; George, T.; Laura, F. ChemRxiv. 2018.
- 28. Pallavicini, P.; Dona, A.; Casu, A.; Chirico, G.; Collini, M.; Dacarro, G.; Falqui, A.; Milanese,
- C.; Sironi, L.; Taglietti, A., Chemical Communications 2013, 49 (56), 6265-6267.
- 29. Mandal, A. B.; Nair, B. U.; Ramaswamy, D., Langmuir 1988, 4 (3), 736-739.
- 30. Takenaka, Y.; Kawabata, Y.; Kitahata, H.; Yoshida, M.; Matsuzawa, Y.; Ohzono, T., *Journal of Colloid and Interface Science* **2013**, 407, 265-272.

31. Zhou, H.; Jia, H.; Zhang, A.; Zhang, L.; Jia, C.; Zheng, L., *Journal of Molecular Liquids* **2015**, 208, 27-33.

- 32. Waqqar, A.; Kooij, E. S.; Arend van, S.; Bene, P., Nanotechnology 2010, 21 (12), 125605.
- 33. Novo, C.; Mulvaney, P., Nano Letters 2007, 7 (2), 520-524.
- 34. Kuo, C.-H.; Huang, M. H., Langmuir 2005, 21 (5), 2012-2016.
- 35. Sau, T. K.; Rogach, A. L.; Döblinger, M.; Feldmann, J., Small 2011, 7 (15), 2188-2194.
- 36. Fales, A. M.; Yuan, H.; Vo-Dinh, T., *The Journal of Physical Chemistry C* **2014**, *118* (7), 3708-3715.
- 37. Keul, H. A.; Möller, M.; Bockstaller, M. R., Langmuir 2007, 23 (20), 10307-10315.
- 38. Alpay, D.; Peng, L.; Marks, L. D., *The Journal of Physical Chemistry C* **2015**, *119* (36), 21018-21023.

39. Langille, M. R.; Personick, M. L.; Zhang, J.; Mirkin, C. A., *Journal of the American Chemical Society* **2012**, *134* (35), 14542-14554.

40. Personick, M. L.; Langille, M. R.; Zhang, J.; Mirkin, C. A., *Nano Letters* **2011**, *11* (8), 3394-3398.

41. Liu, X.; Atwater, M.; Wang, J.; Huo, Q., *Colloids and Surfaces B: Biointerfaces* **2007**, *58* (1), 3-7.

42. Barbosa, S.; Agrawal, A.; Rodríguez-Lorenzo, L.; Pastoriza-Santos, I.; Alvarez-Puebla, R. A.; Kornowski, A.; Weller, H.; Liz-Marzán, L. M., *Langmuir* **2010**, *26* (18), 14943-14950.