

Characterization of Protease-Activated Receptor (PAR) ligands: Parmodulins are reversible allosteric inhibitors of PAR1-driven calcium mobilization in endothelial cells

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Abstract

Several classes of ligands for Protease-Activated Receptors (PARs) have shown impressive anti-inflammatory and cytoprotective activities, including PAR2 antagonists and the PAR1-targeting parmodylins. In order to support medicinal chemistry studies with hundreds of compounds and to perform detailed mode-of-action studies, it became important to develop a reliable PAR assay that is operational with endothelial cells, which mediate the cytoprotective effects of interest. We report a detailed protocol for an intracellular calcium mobilization assay with adherent endothelial cells in multiwell plates that was used to study a number of different PAR1 and PAR2 ligands, including an alkynylated version of the PAR1 antagonist RWJ-58259 that is suitable for the preparation of tagged or conjugate compounds. Using the cell line EA.hy926, it was necessary to perform media exchanges with automated liquid handling equipment in order to obtain optimal and reproducible antagonist concentration–response curves. The assay is also suitable for study of PAR2 ligands; a peptide antagonist reported by Fairlie was synthesized and found to inhibit PAR2 in a manner consistent with reports using epithelial cells. The assay was used to confirm that vorapaxar acts as an irreversible antagonist of PAR1 in endothelium, and parmodylin 2 (ML161) and the related parmodylin RR-90 were found to inhibit PAR1 reversibly, in a manner consistent with negative allosteric modulation.

1. Introduction

Protease-activated receptors (PARs)¹⁻³ comprise a category of class A G-protein coupled receptors (GPCRs) that are activated by extracellular proteases via cleavage of their N-termini, revealing tethered ligands that serve to self-activate the receptors. This process is illustrated with one of 4 PAR subtypes, PAR1, in Figure 1.^{4,5} PARs mediate a wide range of biological effects in numerous tissues, in particular platelet activation and inflammation in vascular tissues. One consequence of the intramolecular mode of activation of PARs is that competitive antagonism can be very challenging due to the high effective concentration of the tethered ligands. Nonetheless, highly potent antagonists have been discovered and progressed to clinical studies as platelet inhibitors: the PAR1 antagonist vorapaxar⁶ is FDA-approved for treatment of coronary artery disease in certain patients, and the pepducin⁷ PAR1 antagonist PZ-128⁸ and the PAR4 antagonist BMS-986120⁹ were recently in Phase 1 clinical trials.

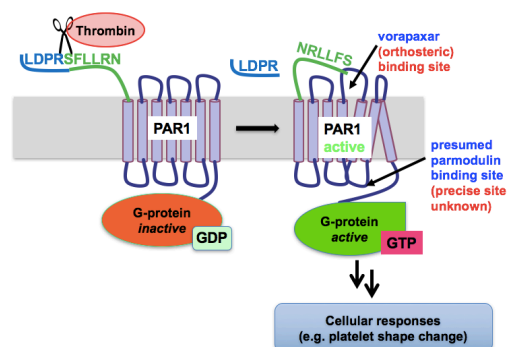
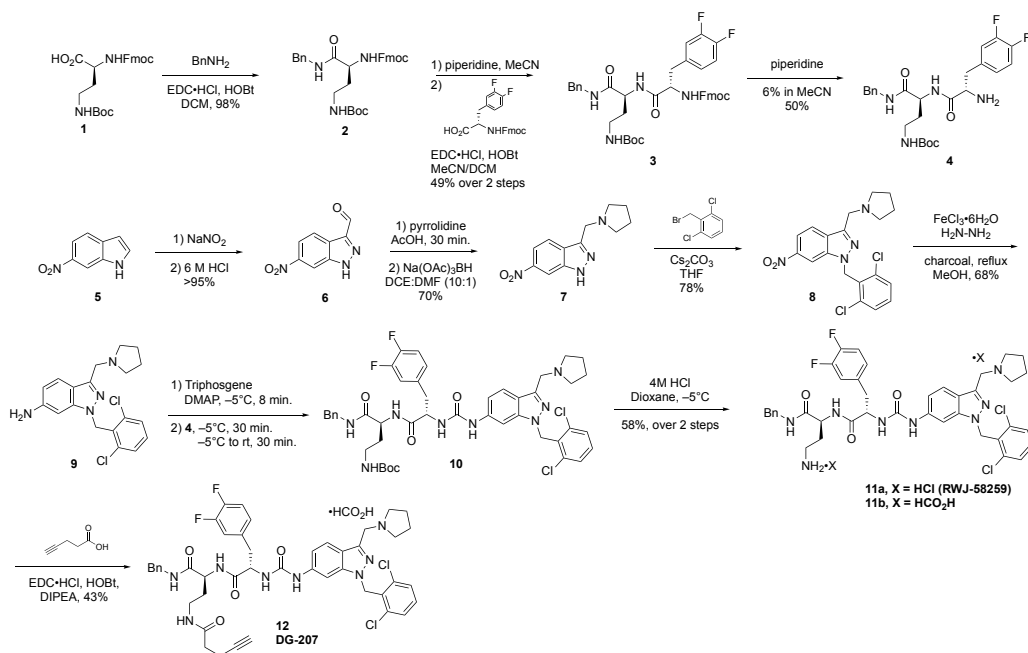


Figure 1. Thrombin-mediated PAR1 signaling

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Scheme 1. Synthesis of PAR1 antagonist RWJ-58259 and alkynylated analog DG-207

As part of our program exploring alternative approaches to the modulation of PAR-mediated signaling,¹⁰ we have been exploring the medicinal chemistry and pharmacology of the parmodulins, a series of small molecules that act as biased ligands at PAR1 via a postulated intracellular, allosteric mechanism (Figure 1).¹¹⁻¹³ Parmodulins were initially of interest to us as antithrombotic agents, due to their biased antagonism of platelet activation: they inhibit Gα_q-driven granule secretion without substantially affecting Gα_{12/13} signaling and platelet shape change. More recently, our lead compound ML161 (also known as parmodulin 2 or PM2) has shown significant promise for its cytoprotective effects in cellular and animal studies, results that are complementary to extensive literature documenting that the PAR-targeting protease Activated Protein C (aPC) can promote cytoprotective effects via PAR1 and/or PAR2,¹⁴⁻²⁵ studies that have been reviewed in recent years.²⁶⁻³¹ In contrast to vorapaxar, ML161 did not block the cytoprotective effects of aPC in endothelial cells exposed to the cytotoxic agents staurosporine and TNF-α,¹³ and more recently it was found to promote on its own a cytoprotective program via PAR1 in the same cells.³² ML161 also significantly decreased cardiac infarct size in a murine model of myocardial ischemia-reperfusion injury.³³ We hypothesize that the biased, cytoprotective signaling of the parmodulins may be due to their presumed allosteric site of action on PAR1 in endothelial cells (Figure 1).

Despite the promising *in vivo* properties of the parmodulins and other PAR ligands, our efforts to study their effects in endothelial cells³⁰ have been hindered by the lack of a reported medium throughput assay protocol. Our present cell lines of interest are Human Umbilical Vein Endothelial Cells (HUVEC) and EA.hy926,³⁴ a hybrid endothelial cell line well-established for *in vitro* measurements of cytoprotective and barrier-protective effects.^{21,22} In order to quantify the signaling bias of the parmodulins and other PAR ligands in these cells, our first objective was to identify a reliable method for the measurement of intracellular calcium mobilization, which can be driven by agonism of PARs through the G-protein Gα_q. Single cell imaging of PAR-driven intracellular calcium mobilization in endothelial cells with different calcium-binding dyes has been reported by several investigators, including Zlokovic³⁵ and Kuckleburg,³⁶ but these assays would not be ideal for rapid assays of large numbers of compounds. Protocols of PAR-

mediated calcium mobilization using plate readers or FLIPR® apparatus have been reported in epithelial cells with PAR2 ligands,³⁷ and with PAR1 ligands in human dermal microvascular endothelial cells (HMEC-1),³⁸ but we are not aware of reports with detailed protocols using adherent endothelial cell lines that were used to generate concentration-responses of PAR1 ligands. Fairlie has, however, reported concentration-responses of PAR2 agonists and antagonists with HUVEC and provided assay protocols.³⁹

2. Results

2.1 Synthesis of tool compounds

We commenced our studies with the syntheses of several tool compounds and functionalized analogs to support future mode-of-action studies. Our interest in building modified and/or labeled versions of various PAR1 antagonists led us to synthesize the tethered ligand peptidomimetic RWJ-58259 (Scheme 1),⁴⁰ versions of which had been under preclinical investigation as antiplatelet agents.⁴¹⁻⁴³ Consideration of published structure-activity relationship (SAR) data with platelets suggested that modification of the primary amine could be tolerated. To this end, we synthesized RWJ-58259 using protocols slightly modified from the original report.⁴⁰ The commercially-available amino acid **1** was coupled sequentially at the acid and amine moieties using standard amide coupling (EDC) and Fmoc-removal conditions (piperidine) to yield dipeptide **4**. According to the protocols reported by Zhang and Maryanoff,⁴⁰ 6-nitroindole **5** underwent nitrosation and rearrangement to yield 3-formylindazole **6**, followed by reductive amination and *N*-alkylation, providing nitroindazole **8**. Several nitro reduction protocols gave high-yielding reactions, including stoichiometric SnCl₂, but these often gave minor byproducts that proved difficult to remove later in the synthesis. The reported protocol using hydrazine and catalytic FeCl₃ gave the desired 7-aminoindazole **9** in high purity after column chromatography. The synthesis of urea **10** via an intermediate 4-nitrophenylcarbamate proved to be low-yielding in our hands, which was also consistent with the original report and a subsequent report from Herranz, who found improved yields using triphosgene and propylene oxide as an acid scavenger.⁴⁴ High-purity urea **10** was obtained by us in acceptable yields via

slow addition of triphosgene to **9**, followed by the addition of dipeptide **4**, as reported by Caddick.⁴⁵ Boc removal using HCl provided RWJ-58259 (**11**), which was profiled with several other PAR1 antagonists (vide infra). Additionally, in order to use RWJ-58259 as a scaffold for the attachment of e.g. fluorophores for future studies, we wished to expand the range of applicable conjugation reactions. We therefore explored the attachment of several different alkyne building blocks for use in copper-catalyzed alkyne/azide cycloadditions (CuAAC).⁴⁶ These efforts were hampered by the poor solubility of several of the intermediates (**2–4**, **10**, **12**) involved in the synthesis of RWJ-58259. However, we successfully coupled **11** with 4-pentynoic acid using EDC/HOBt in DMF (Scheme 1) to yield alkyne DG-207 (**12**).

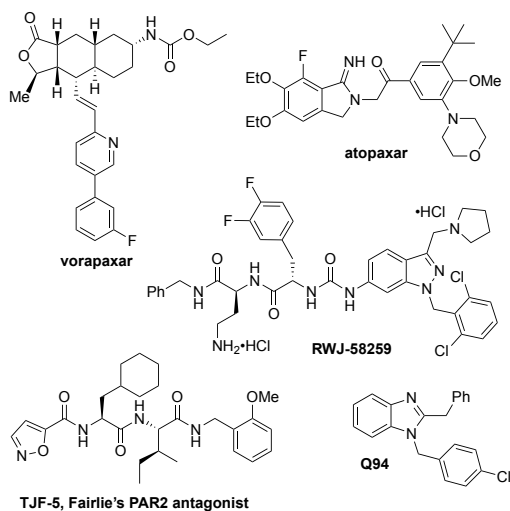
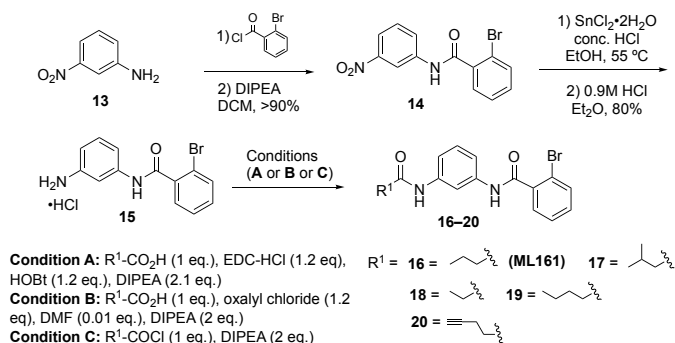


Figure 2. Previously-reported PAR antagonists used in these studies

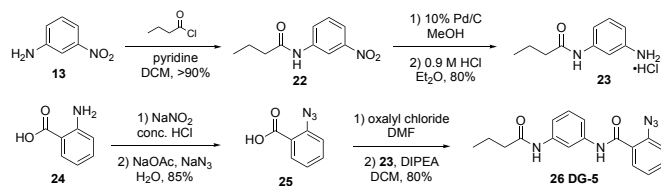
In addition to RWJ-58259 and the commercially available PAR1 antagonists vorapaxar⁶ and atropaxar⁴⁷ (Figure 2), we aimed to profile a selection of our parmodulin analogs (Table 1). “Western” analogs (ML161, CJD-125, RR-90, EMG-21, EMG-23, and RR-10) were prepared as described previously,¹² or via the protocol summarized in Scheme 2. The alkyne RR-10 and the aryl azide DG-5 (Scheme 3) were also prepared to investigate the possibility of generating photoaffinity ligands (via aryl azide to nitrene conversion), with alkyne handles for pulldown experiments.



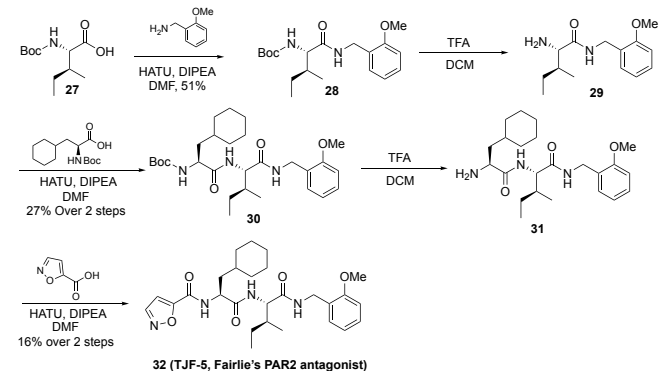
Scheme 2. General synthesis of parmodulins with west side modifications

Our additional interest in the modulation of PAR2 required us to obtain a reliable PAR2 antagonist for confirming that our assay could also be applicable to measuring PAR2 activation in endothelium. For this purpose, we selected Fairlie's peptidic PAR2 antagonist (termed by us TJF-5, Figure 2), which was

previously prepared via solid-phase peptide synthesis.⁴⁸ Since we wished to potentially access larger quantities of this compound, we investigated a solution-phase synthesis of TJF-5, which proceeded uneventfully using Boc-protected building blocks and HATU as coupling agent (Scheme 4).



Scheme 3. Synthesis of azido analog DG-5



Scheme 4. Solution-phase synthesis of TJF-5 (Fairlie's PAR2 antagonist)

2.2 Initial studies of calcium mobilization with adherent endothelial cells

Our assay development studies commenced with the testing of several options for measuring calcium mobilization in adherent endothelial cells. The transformed endothelial cell line EA.hy926,³⁴ widely used in other PAR-related studies, could be grown reliably in tissue culture-treated 96-well plates. Cell seeding of approximately 25,000 cells/well gave confluent monolayers within 48 h. The calcium binding fluorescent dye Fluo-4, dosed as its acetoxymethyl ester (Fluo-4/AM), generated adequate emission signals at 525 nm for us to reliably measure PAR activation using peptide agonists for human PAR1 and PAR2, which correspond to their tethered ligands: TFLLRN-NH₂ (PAR1 selective), SLIGKV-NH₂ (PAR2 selective), and SFLLRN-NH₂ (PAR1/2 selective). Resting cells containing Fluo-4 are depicted in Figure 3A, prior to agonist addition. As with other calcium-binding dyes, the acetoxymethyl ester permeates the cells and is cleaved by esterases, retaining it in the cell.⁴⁹ Anionic transporters are blocked by the concurrent application of probenecid, which slows the efflux of Fluo-4, according to standard protocols. However, we did observe the efflux of Fluo-4 and decreased calcium responses over time. Assays were performed on 16 wells at a time (2 columns of a 96 well plate) using a plate reader (PerkinElmer EnSpire[®]); increased throughput could be easily obtained using an instrument imaging all wells concurrently (e.g. FLIPR[®]).^{50,51} A representative trace of response versus time is shown in Figure 3B. The difference between the maximal emission and the average of the baseline emission at 525 nm was used for all response measurements, which were normalized to the agonist response with no inhibitor present (only vehicle, set to 100%).

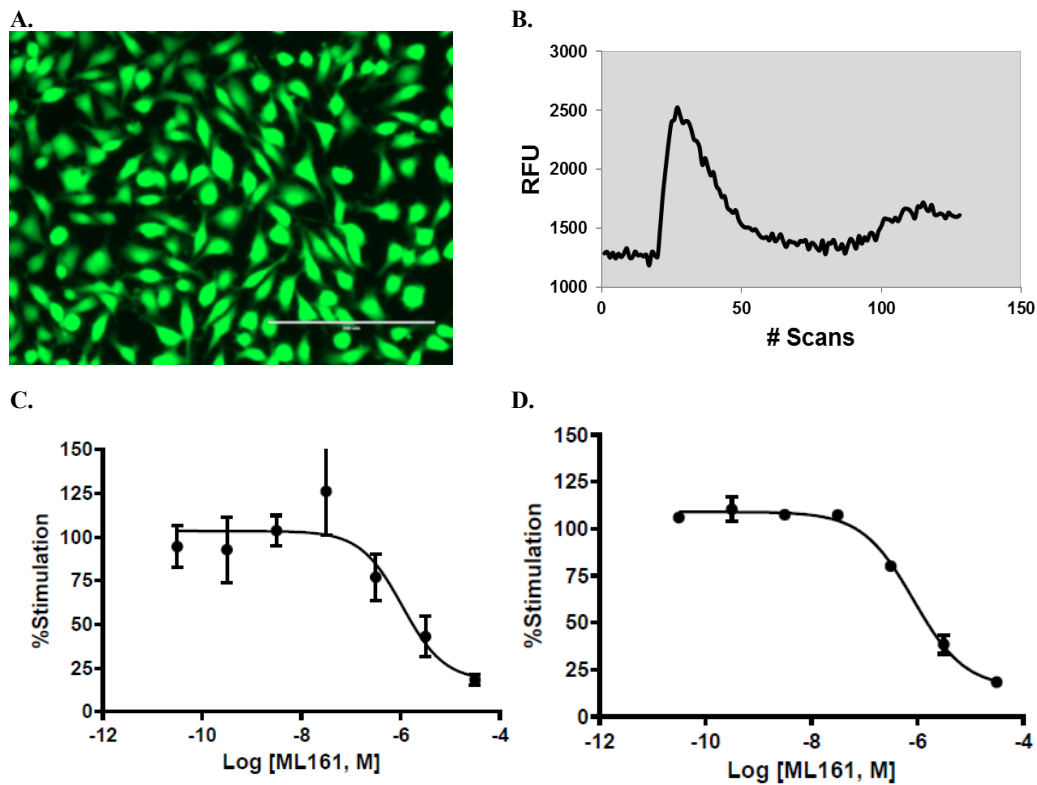
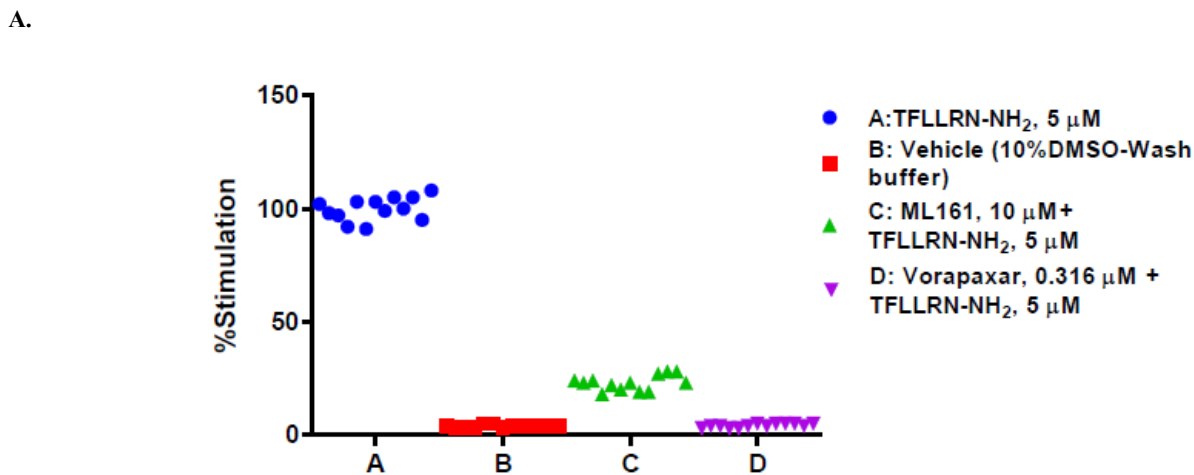


Figure 3. **A:** Adherent Ea.hy926 cells containing Fluo-4 dye. Cells were imaged in a clear-bottom, black wall 96-well plate, using an Evos Fl inverted microscope (20X) with GFP setting. **B:** Representative Fluo-4-Ca²⁺ emission curve (525 nm) in response to TFLLRN-NH₂ (3.16 μM). **C:** Concentration-response curve of a typical (unsuccessful) assay with the PAR1 antagonist ML161 and 3.16 μM TFLLRN-NH₂ before optimization (manual media exchanges), n =3. **D:** Representative concentration-response curve with the PAR1 antagonist ML161 and 5 μM TFLLRN-NH₂ after assay optimization (automated liquid handling).



B.

	Positive Control	Negative Control	Z ² -factor
Agonist Assay (n=12)	TFLLRN-NH ₂ (5 μM)	Vehicle (10% DMSO-HBSS/HEPES)	0.720
Antagonist Assay (n=12)	ML161 (10 μM) + TFLLRN-NH ₂ (5 μM)	TFLLRN-NH ₂ (5 μM)	0.618

Figure 4. **A:** Calcium mobilization responses with PAR1 agonist 5 μM TFLLRN-NH₂ (column A) plus optional PAR1 antagonists 10 μM ML161 (column C) or 0.316 μM vorapaxar (column D). **B:** Table of calculated Z²-factors.

We were pleased to see acceptable signal/noise and measured concentration-response curves with the PAR1 and PAR2 agonists, however we were disappointed to see inconsistent results when measuring antagonist activities. Deviations in normalized emission levels were unacceptably high; a representative plot is given for ML161 in Figure 3C. Adverse effects on the endothelial cells (retraction) were observed when incubated in media containing DMSO required to dissolve the antagonists (1% final concentration). Retraction was minimized when working with 0.1–0.2% DMSO, but high deviations were still observed in many, but not all, assays. We reasoned that the low aqueous solubility of many PAR antagonists, including vorapaxar and ML161, may cause inconsistent results, particularly since turbid suspensions were observed at high concentrations. Several excipients were tested in order to keep compounds in solution at the highest concentrations (10–100 μM), including Tween-20, Tween-80, Triton X-100, BSA, and Pluronic F-127 (0.01 to 0.1%). Pluronic F-127 (up to 0.05%) was effective at solubilizing compounds and did not have observable toxic effects on the cells. Assays proceeded with this excipient in the antagonist solutions, but as previously, inconsistent antagonist concentration-response curves were still obtained. Interestingly, the high deviations themselves were inconsistent, and it became clear that certain wells contained poorly-responding cells. We reasoned that inconsistent pipetting technique, particularly for introduction of the agonists immediately before scanning in the plate reader, could be the source of inconsistencies, but this also did not prove to be the case after extensive experimentation. The addition of higher volumes of agonist led to oscillating calcium levels, perhaps as a result of the temperature differential between the agonist solution (at $\sim 20^\circ\text{C}$) and the cells at 37°C .

We next examined the process of loading the dye (Fluo-4/AM) into the cells. Omitting the probenecid did not have a productive effect. Buffer containing the Fluo-4/AM is added to the cells, then incubated for 0.5–1 h. Longer incubation times with the dye were not helpful and simply led to lower maximum signals, with the Fluo-4 slowly leaking from the cells, which can be problematic for longer experiments. An additional variable was the manner in which the media was changed. Our initial protocol used simple “flicking” of the plates by hand into paper towels to remove media or buffer from the plates, followed by multichannel pipetting of fresh buffer during liquid exchanges. Ensuring that the pipet tips did not touch the bottom of the plate did not improve the quality of the data. Finally, we reasoned that the force of the flicking was inconsistent and could disturb the cells in unpredictable ways, even though wells were confirmed to contain evenly distributed cells that had not obviously detached. Our inconsistent results disappeared after we gained access to an automated liquid handler (Beckman-Coulter BioMek 3000) for media changes. Antagonist concentration-response curves were then consistently obtained with low deviations (e.g. Figure 3D). To determine the suitability of our assay for screening hundreds of compounds, we performed experiments to determine the signal window of the assay by measuring the average response and standard deviation for the PAR1 agonist TFLLRN-NH₂ (5 μM) (designated as the positive control, Figure 4, column A), and comparing to the responses obtained after the same agonist was added to cells treated with the PAR1 antagonists ML161 (10 μM) (column C) or vorapaxar (0.316 μM) (column D). The resulting Z' factors were >0.5 , and the assay is thus considered to be highly suitable for high-throughput screening.⁵²

2.3 Concentration-responses of PAR1 and PAR2 antagonists with endothelial cells

With a reliable calcium mobilization assay in hand, we proceeded to determine concentration-responses with the peptide agonists for PAR1 (Figure 5A) and PAR2 (Figure 5B), which gave EC₅₀s of 4.8 μM and 5.8 μM , respectively. Using 5 μM TFLLRN-NH₂ as agonist, concentration-response curves were also measured with the PAR1 antagonists vorapaxar (Figure 6A, IC₅₀ = 0.032 μM), atopaxar (Figure 6B, IC₅₀ = 0.033 μM), and RWJ-58259 (Figure 6C, IC₅₀ = 0.020 μM). Of special interest to us was the performance of our alkynylated analog of RWJ-58259, DG-207 (12). It is 15-fold less potent in this assay than RWJ-58259 (Figure 6D), but with an IC₅₀ of 0.30 μM it is still anticipated to have sufficient potency for our planned conjugate molecules.

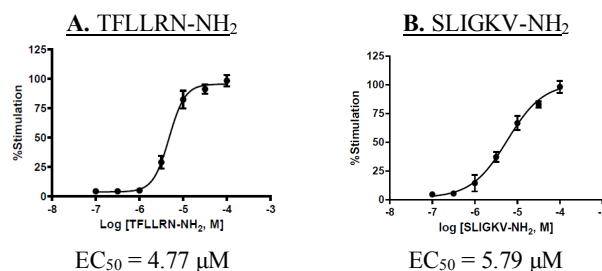


Figure 5. PAR agonist concentration–response with EA.hy926 cells: A) TFLLRN-NH₂ (PAR1)-mediated $i\text{Ca}^{2+}$ mobilization; B) SLIGKV-NH₂ (PAR2)-mediated $i\text{Ca}^{2+}$

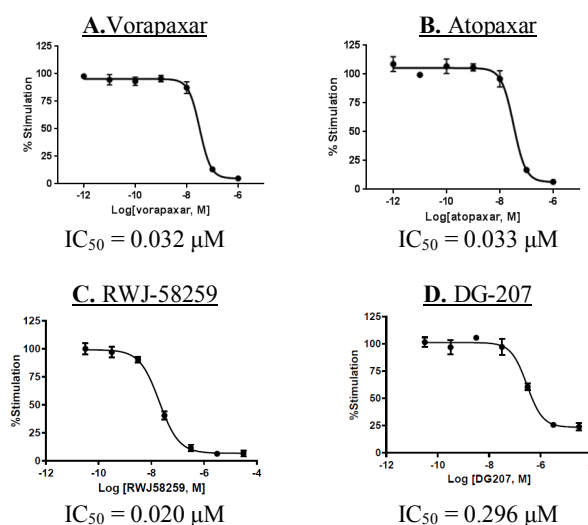


Figure 6. Concentration-response for PAR1 antagonists in the TFLLRN-NH₂-mediated $i\text{Ca}^{2+}$ mobilization assay (5 μM) with EA.hy926 cells: A) vorapaxar, B) atopaxar, C) RWJ-58259 D) DG-207

Concentration-responses were also obtained with several representative parmodulins (Figure 7). With ML161, optimal signal/noise was obtained with 5 μM TFLLRN-NH₂; at this concentration, an IC₅₀ of 0.85 μM was measured (Figure 7A). As previously reported in an alternative P-selectin expression assay with platelets,¹² the analog with one carbon less on the aliphatic chain on the western side (EMG-21) was active but less potent than ML161 (Figure 7B, IC₅₀ = 3.3 μM), and the analog with one additional carbon (EMG-23) was poorly active (Table 1, entry 8). Also in line with our previous results in platelets, the aniline RR-90 was similar in potency to ML161 (Figure 7C, IC₅₀ = 0.97 μM). However, the branched aliphatic amide CJD-125, an analog

of ML161 with improved plasma stability and equipotency in the platelet P-selectin assay,¹² was less potent than ML161 and did not give a good concentration-response with EA.hy926 cells (Table 1, entry 6). Of note is the fact that efficacy (maximal inhibition) of the parmodulins does not appear to be quite as high as with the presumably orthosteric inhibitors of Figure 6, though this comparison is complicated by concentration-limiting solubilities of some of the parmodulins.

Our collaborators in the Flaumenhaft lab have performed experiments with chimeric PAR receptors which support our hypothesis that the parmodulins act at an intracellular location,¹³ which is highly unusual for small molecule GPCR ligands. In order to identify a precise binding site on PAR1 for the parmodulins, we were interested in generating analogs of ML161 suitable for photoaffinity studies. To this end, we prepared analogs possessing aryl azides (for photochemical generation of reactive nitrenes), and alkynes for conjugation or pulldown experiments. Unfortunately, the aryl azide DG-5 (Figure 7D; Table 1, entry 10) had only modest activity. The western alkyne RR-10 (entry 11) also had only modest inhibition of PAR1. Results with all PAR1 ligands are summarized in Table 1.

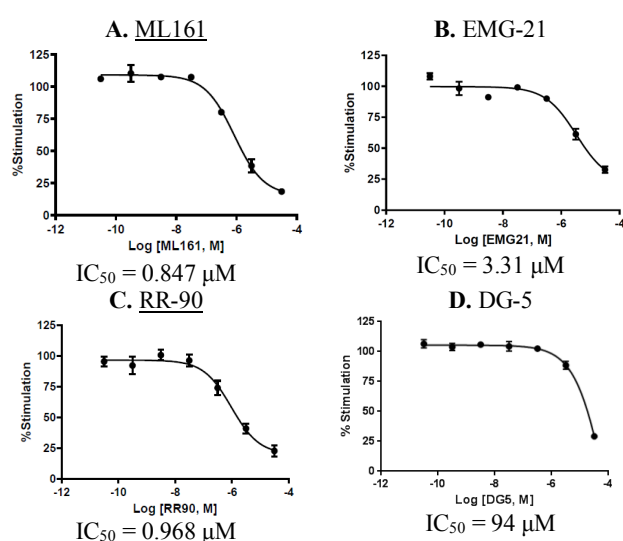


Figure 7. Concentration-response for parmodulins (ML161 analogs) in the TFLLRN-NH₂-mediated (5 μM) *i*Ca²⁺ mobilization assay

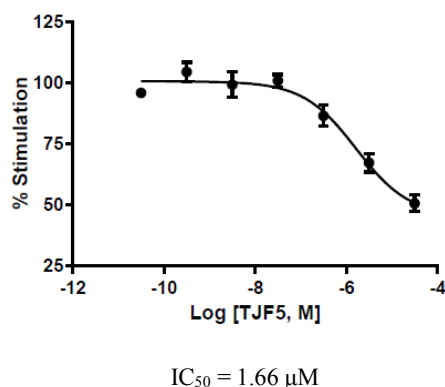


Figure 8. Concentration-response for the PAR2 antagonist TJF5 (Fairlie's PAR2 antagonist) in the SLIGKV-NH₂-mediated (5 μM) *i*Ca²⁺ mobilization assay.

Our assay also proved to be reliable for measuring the inhibition of calcium mobilization with PAR2 antagonists: the peptide inhibitor TJF-5 inhibited the action of the PAR2 activating peptide SLIGKV-NH₂ (5 μM) with an IC₅₀ of 1.7 μM

(Figure 8). Data with other proprietary PAR2 antagonists will be presented in due course.

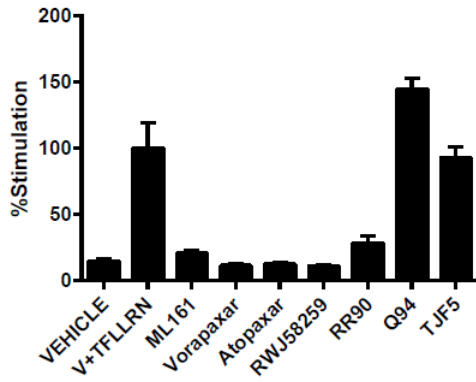
2.4 Selectivity and mode-of-action studies

The activity of the parmodulins and other PAR1 antagonists at PAR2 has not previously been reported, to our knowledge, perhaps because human platelets do not express PAR2. The ability of compounds to inhibit the action of the PAR1 agonist TFLLRN-NH₂ is summarized in Figure 9A. The more potent PAR1 antagonists described previously vorapaxar, atopaxar, and RWJ-58259 were used at concentrations of 0.316 μM, and as expected largely inhibit the activation of the EA.hy926 cells and mobilization of calcium. The parmodulins ML161 and RR-90 also significantly inhibited the PAR1 agonist. Q94 (Figure 2) is a commercially available benzimidazole that was reported to selectively inhibit PAR1 Gαq-mediated calcium mobilization in HMEC-1 endothelial cells.³⁸ It did not act as an antagonist in this assay using EA.hy926 cells, and actually boosted the response to TFLLRN-NH₂. As expected, the PAR2 antagonist TJF-5 also had no measurable effect on TFLLRN-NH₂-induced calcium mobilization. None of the PAR1 ligands had any measurable effect on PAR2-mediated calcium mobilization with SLIGKV-NH₂ (Figure 9B), confirming that they have excellent selectivity for PAR1 over PAR2 in EA.hy926 endothelial cells. These cells did not respond to high concentrations (up to 100 μM) of the PAR4 agonist AYPGFK-NH₂.

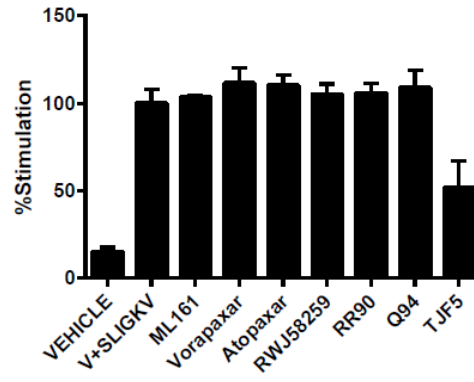
Next, we evaluated if the PAR1 ligands inhibited PAR1 in EA.hy926 cells in an allosteric, non-competitive manner; we have previously shown that ML161 acts as a non-competitive inhibitor of SFLLRN-NH₂ in a platelet P-selectin assay,¹² and it does not significantly inhibit binding of [³H] high-affinity thrombin receptor activating peptide ([³H]TRAP) to platelet membranes.¹³ We measured the concentration-responses of TFLLRN-NH₂ in the presence of increasing concentrations of PAR1 antagonists (Figure 10). Both the presumed orthosteric antagonists (vorapaxar and RWJ-58259) and the parmodulins (ML161 and RR-90) induced a dose-dependent decrease in efficacy of the PAR1 agonist that is consistent with either a lack of reversibility OR action as a negative allosteric modulator. However, it should be noted that the fitted concentration-response curves of TFLLRN-NH₂ in the presence of RWJ-58259 suggest that competitive inhibition might also be possible, whereby the curves are shifted rightward, but the agonist concentrations are not high enough to detect maximum response. The nearly identical ligand RWJ-56110 (with an indole instead of indazole) is a competitive inhibitor of binding of the radiolabeled PAR1 activating peptide [³H]S-(*p*-F-Phe)-homoarginine-L-homoarginine-KY-NH₂.⁴¹ Vorapaxar⁶ is well known to act as a poorly reversible inhibitor of PAR1, with a very low off rate and extended half-life, likely due to its binding site on the top of PAR1 whereby it becomes covered by extracellular loop 2 (ECL-2), as seen in the crystal structure reported by Coughlin and Kobilka.⁵³

To differentiate between irreversible (presumably orthosteric) inhibition and negative allosteric modulation, we performed "wash" studies whereby the endothelial cells were treated with inhibitor, then washed twice with buffer prior to addition of the agonist and measurement of intracellular calcium levels (Figure 11). The activity of vorapaxar was completely retained after washing the cells, confirming that its binding is poorly reversible in endothelial cells. RWJ-58259 and the parmodulins ML161, RR-90, and CJD-125 lost most or all of their activities after washing. Therefore, we conclude that the parmodulins, at least ML161 and RR-90, act as negative allosteric modulators of TFLLRN-NH₂ at PAR1.

A. PAR1 activity



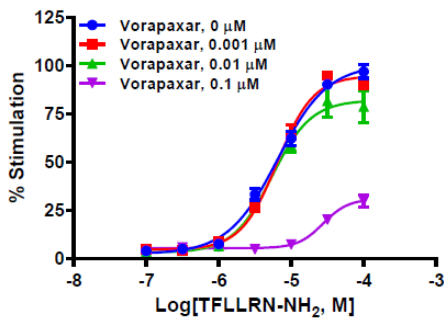
B. PAR2 activity



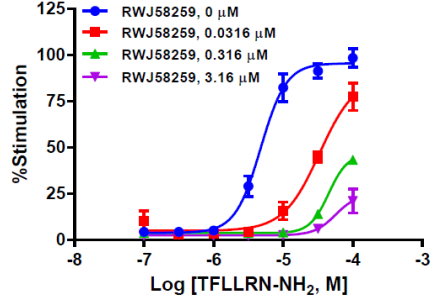
^aML161, RR90, Q94, TJF5 (Fairlie's PAR2 antagonist) were used at 10 μ M; Vorapaxar, Atopaxar, and RWJ-58259 were at 0.316 μ M. TFLLRN-NH₂ and SLIGKV-NH₂ were used at 3.16 μ M; Vehicle (V) = 10% DMSO-HBSS/HEPES

Figure 9. Selectivity data of antagonists in PAR1 (TFLLRN-NH₂)- and PAR2 (SLIGKV-NH₂)-driven *i*Ca²⁺ mobilization^a

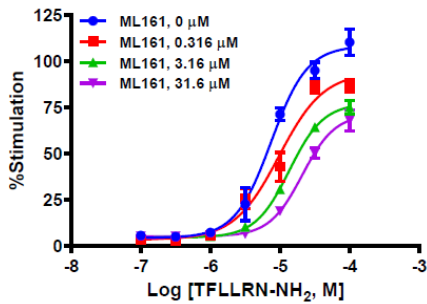
A.



B.



C.



D.

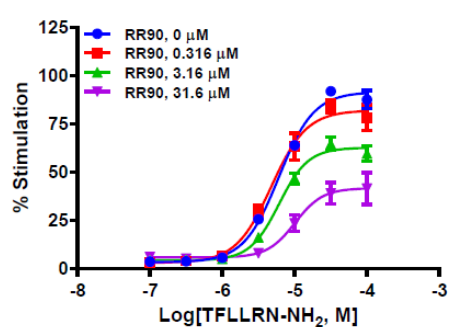


Figure 10. *i*Ca²⁺ concentration-response of the PAR1 agonist TFLLRN-NH₂ in the presence of increasing concentrations of (A) vorapaxar, (B) RWJ 58259, (C) ML161, (D) RR-90

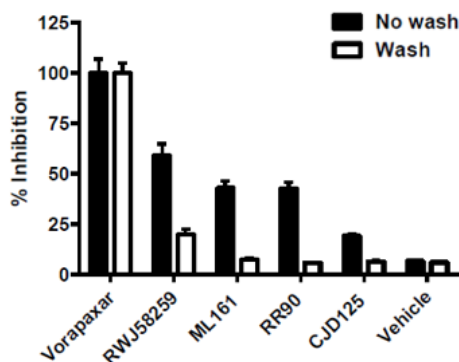


Figure 11. Reversibility studies of the PAR1 antagonists vorapaxar, RWJ-58259, ML161, and RR-90 in the iCa^{2+} assay. Cells containing antagonists were optionally washed with buffer prior to treatment with the PAR1 agonist TFLLRN-NH₂ (5 μ M). Vorapaxar and RWJ-58259 were used at 0.316 μ M; ML161, CJD125, RR-90 were used at 10 μ M.

Table 1. Inhibition data of select antagonists in the PAR1-mediated iCa^{2+} mobilization assay

Entry	ID#	Structure	% Inhibition & IC ₅₀ ^a	Log IC ₅₀ ± Std. Error
1	Vorapaxar		95%; 0.032 μ M	-7.5 ±0.084
2	Atopaxar		94%; 0.033 μ M	-7.5 ±0.10
3	RWJ-58259		94%; 0.020 μ M	-7.7 ±0.065
4	DG-207		76%; 0.29 μ M	-6.5 ±0.074
5	ML161		82%; 0.85 μ M	-6.7 ±0.099
6	CJD-125		74%; 3.16 μ M ^b	N.D.
7	EMG-21		68%; 3.31 μ M	-5.5 ±0.26
8	EMG-23		49%	N.D.
9	RR-90		77%; 0.97 μ M	-6.0 ±0.19
10	DG-5		77%; 94 μ M	-4.0 ±2.9
11	RR-10		79%; 81 μ M	-4.0 ±5.1

^a %Inhibition of PAR1 antagonists in presence of 5 μ M TFLLRN-NH₂. N.D. = not determined

^b Estimated IC₅₀ (incomplete curve at high concentration)

3. Conclusions

A protocol for a medium-throughput assay measuring calcium mobilization in adherent endothelial cells was developed and used to measure concentration-responses of a variety of PAR1 and PAR2 ligands. An alkyne-tethered version of the PAR1 antagonist RWJ-58259 (DG-207) was synthesized. It was a less potent inhibitor of the PAR1 agonist TFLLRN-NH₂ than the parent compound, but still anticipated to have adequate potency for future use in conjugate compounds. In contrast, several alkyne- and azide-containing analogs of the parmodulin ML161 were found to have moderate (micromolar inhibition) or low potency in this assay. A selection of PAR1 and PAR2 ligands were profiled in further detail, and were found to be highly selective for PAR1 or PAR2. The parmodulins ML161 and RR-90 were found to decrease the efficacy of TFLLRN-NH₂ in a dose-dependent manner, but unlike the FDA-approved inhibitor vorapaxar, washing studies showed that their action is reversible in a manner consistent with negative allosteric inhibition of PAR1. Such compounds possess significant potential for the modulation of PARs in a therapeutic manner, in particular for inflammation-related disorders such as sepsis or reperfusion injury.⁵⁴ Our current work is aimed towards measuring the signaling biases of the parmodulins at PAR1 and optimizing their cytoprotective effects.

4. Experimental section

4.1 General information: Synthesis

All reagents and solvents, including anhydrous solvents, were purchased from commercial vendors and used as received. Compound **1** (CAS# 125238-99-5) was obtained from ChemImpex. Compounds **14**, **15**, **16** (ML161), **17**, **21** (RR-90), and **22** were synthesized as previously described.¹² Deionized water was purified by charcoal filtration and used for reaction workups and in reactions with water. NMR spectra were recorded on Varian 300 MHz or 400 MHz spectrometers as indicated. Proton and carbon chemical shifts are reported in parts per million (ppm; δ) relative to tetramethylsilane (¹H δ 0), or CDCl₃ (¹³C δ 77.16), (CD₃)₂CO (¹H δ 2.05, ¹³C δ 29.84), d₆-DMSO (¹H δ 2.50, ¹³C δ 39.5), or CD₃OD (¹H δ 3.31, ¹³C δ 49.00). NMR data are reported as follows: chemical shifts, multiplicity (obs = obscured, app = apparent, br = broad, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, comp = complex overlapping signals); coupling constant(s) in Hz; integration. Unless otherwise indicated, NMR data were collected at 25 °C. Filtration was performed by vacuum using VWR Grade 413 filter paper, unless otherwise noted. Flash chromatography was performed using Biotage SNAP cartridges filled with 40-60 μ m silica gel on Biotage Isolera automated chromatography systems with photodiode array UV detectors. Analytical thin layer chromatography (TLC) was performed on Agela Technologies 0.25 mm glass plates with 0.25 mm silica gel. Visualization was accomplished with UV light (254 nm) and KMnO₄ stain, unless otherwise noted. Chemical names were generated and select chemical properties were calculated using either ChemAxon Marvin suite (<https://www.chemaxon.com>) or ChemDraw Professional 15.1. NMR data were processed using either MestreNova or ACD/NMR Processor Academic Edition (<http://www.acdlabs.com>) using the JOC report format. High-resolution mass spectra (HRMS) were obtained at the University

of Wisconsin-Milwaukee Mass Spectrometry Laboratory with a Shimadzu LCMS-IT-TOF with ESI and APCI ionization.

4.2 LC/MS characterization methods

Tandem liquid chromatography/mass spectrometry (LC-MS) was performed on a Shimadzu LCMS-2020 with autosampler, photodiode array detector, and single-quadrupole MS with ESI and APCI dual ionization using a Peak Scientific nitrogen generator.

Method A

Column: Phenomenex Gemini C₁₈ (100 x 4.6 mm, 3 μ m particle size, 110 Å pore size)

Column temperature: 40 °C

Sample Injection: 1–5 μ L of sample in MeCN or MeOH

Chromatographic monitoring: UV absorbance at 210 or 254 nm

Mobile Phase: Solvent A: H₂O w/ 0.1% formic acid; Solvent B: MeCN w/ 0.1% formic acid

Flow Rate: 1.0 mL/min

Gradient: 0 to 0.5 min: 25% MeCN

0.5 min to 5 min: 25% to 95% MeCN

5 min to 7.3 min: 95% MeCN

7.3 min to 7.5 min: 95% to 25% MeCN

4.3 Preparative HPLC purification methods

Preparative liquid chromatography was performed on a Shimadzu LC-20AP preparative HPLC with autosampler, dual wavelength detector, and fraction collector.

Method A

Column: Phenomenex Gemini C₁₈ Semi Preparative Column (250 x 10 mm, 5 μ m particle size, 110 Å pore size)

Column temperature: 23 °C

Mobile Phase: Solvent A: H₂O w/ 0.1% formic acid; Solvent B: MeCN w/ 0.1% formic acid

Peak collection: measured by UV absorbance at 210 or 254 nm

Sample Injection: 0.1–1.9 mL (2 mL sample loop) of sample in DMSO

Flow Rate: 6.0 mL/min

Gradient: 0 to 1.5 min: 25% MeCN

1.5 min to 8 min: 25% to 95% MeCN

8 min to 10.5 min: 95% MeCN

4.4 Synthesis of RWJ-58259 and alkynylated analog DG-207

4.4.1 (9H-Fluoren-9-yl)methyl tert-butyl (4-(benzylamino)-4-oxobutane-1,3-diyl)(S)-dicarbamate (2). To a round bottom flask with stir bar under nitrogen was added carboxylic acid **1** (500 mg, 1.13 mmol) and anhydrous DCM (25 mL). Benzylamine (0.15 mL, 1.4 mmol), HOBt (229 mg, 1.50 mmol), and EDC•HCl (433 mg, 2.26 mmol) were added and the reaction was stirred under nitrogen for 2 h. A sample aliquot was taken from the reaction, concentrated under reduced pressure, dissolved in a minimal amount of HPLC grade MeCN, and analyzed with LCMS to confirm reaction completion. The reaction was diluted with DCM (75 mL) and washed with saturated NaHCO₃ (30 mL), 1M HCl (30 mL), brine, dried over MgSO₄, filtered, and concentrated under reduced pressure to give **2** (588 mg) as a white solid in 98% yield. mp = 154–156 °C; LC/MS t_R = 6.09 min (Characterization Method A); m/z = 530.05 (M + H⁺); ¹H NMR (300 MHz, CDCl₃) δ = 7.76 (d, J = 7.6 Hz, 2 H), 7.61 – 7.49 (overlapping signals, 3 H), 7.43 – 7.22 (m, 9 H), 5.96 (d, J = 6.7 Hz, 1 H), 5.11 (br. m., 1 H), 4.50 – 4.13

(overlapping m, 6 H), 3.48 – 3.32 (br. s., 1 H), 3.04 – 2.90 (br. m., 1 H), 1.97 – 1.71 (overlapping signals, 2 H), 1.45 – 1.37 (m, 9 H); ¹³C NMR (75 MHz, CDCl₃) δ = 171.2, 157.2, 156.3, 143.9, 141.5, 138.0, 128.8, 127.9, 127.8, 127.6, 127.3, 125.3, 120.2, 80.1, 67.2, 52.2, 47.3, 43.8, 37.0, 34.9, 28.6.

4.4.2 tert-butyl ((S)-3-((S)-2-(((9H-Fluoren-9-yl)methoxy)carbonyl)amino)-3-(3,4-difluorophenyl)propanamido)-4-(benzylamino)-4-oxobutyl)carbamate (3). Part 1: *Fmoc removal*. Dicarbamate **2** (550 mg, 1.03 mmol) was added to a round bottom flask with stir bar and sealed under nitrogen, then MeCN (25 mL), DCM (10 mL), and piperidine (0.220 mL, 2.22 mmol) were added. The reaction was stirred for 2 h. A sample aliquot was taken from the reaction, concentrated under reduced pressure, dissolved in a minimal amount of HPLC grade MeCN, and analyzed with LCMS to confirm reaction completion. The reaction was concentrated under reduced pressure and then re-dissolved in (CHCl₃/EtOAc, 1:1, 75 mL), washed with H₂O (2 x 20 mL) and brine (20 mL). The organic phase was dried over MgSO₄, filtered, and concentrated under reduced pressure to give crude material (496 mg) as a white solid that was carried onto the next reaction without further purification.

Part 2: *Coupling*. To a round bottom flask with stir bar already containing crude material from Part 1 (496 mg crude material, max. yield is 319 mg) was added anhydrous MeCN (30 mL) and anhydrous DCM (15 mL). (S)-2-(((9H-Fluoren-9-yl)methoxy)carbonyl)amino)-3-(3,4-difluorophenyl)propanoic acid (300 mg, 0.709 mmol), HOBt (191 mg, 1.41 mmol), and EDC·HCl (272 mg, 1.41 mmol) were added, and the resulting white suspension was stirred for 2 h. A sample aliquot was taken from the reaction, concentrated under reduced pressure, dissolved in a minimal amount of HPLC grade MeCN, and analyzed with LCMS to confirm reaction completion. The reaction was concentrated under reduced pressure, re-suspended in EtOAc (100 mL), and washed with H₂O (20 mL), 0.5 M HCl (20 mL), saturated NaHCO₃ (20 mL), and brine. The organic phase was dried over MgSO₄, filtered, and concentrated under reduced pressure to give crude material (yellow oil) that was dry loaded using silica onto a 5 g silica gel column and purified with flash chromatography (EtOAc/hexanes, 1–100%) to give **3** (368 mg, 49%) as a white solid over 2 steps. m.p. = 183–185 °C; LC/MS t_R = 6.43 min (Characterization Method A); m/z = 713.15 (M + H⁺), m/z = 757.35 (formic acid adduct); ¹H NMR (300 MHz, DMSO-d₆) δ = 8.43 (dd, J = 5.0, 5.9 Hz, 1 H), 8.22 (d, J = 7.9 Hz, 1 H), 7.86 (d, J = 7.6 Hz, 2 H), 7.68 - 7.53 (m, 2 H), 7.52 - 7.43 (br. s., 1 H), 7.43 - 7.07 (m, 11 H), 6.74 (br. s., 1 H), 4.28 (br. d, J = 5.6 Hz, 4 H), 4.15 (br. s, 3 H), 3.05 - 2.85 (overlapping br. s, 3 H), 2.73 (app. t, J = 12.0 Hz, 1 H), 1.86 - 1.73 (m, 1 H), 1.73 - 1.58 (m, 1H), 1.34 (s, 9 H); ¹³C NMR (75 MHz, DMSO-d₆) δ = 171.8, 171.7, 156.4, 156.1, 150.8 (dd, J = 45.2, 13 Hz), 147.6 (dd, J = 45.8, 12.1 Hz), 144.4, 144.3, 141.35, 141.33, 139.8, 136.6, 128.9, 128.2, 127.7, 127.6, 127.4, 126.7, 125.9, 120.7, 118.9, 118.6, 117.6, 117.4, 78.3, 66.3, 56.4, 51.5, 47.2, 42.7, 37.5, 37.1, 33.1, 28.9

4.4.3 tert-butyl ((S)-3-((S)-2-Amino-3-(3,4-difluorophenyl)propanamido)-4-(benzylamino)-4-oxobutyl)carbamate (4). Carbamate **3** (250 mg, 0.351 mmol) was added to a round bottom flask with stir bar and sealed under nitrogen, then anhydrous MeCN (30 mL) and piperidine (2 mL) were added and the reaction was stirred for 90 min. A sample aliquot was taken from the reaction, concentrated under reduced pressure, dissolved in a minimal amount of HPLC grade MeCN, and analyzed with LCMS to confirm reaction completion. The reaction was concentrated under reduced pressure to give crude

material (light yellow solid) that was triturated with 75/25 hexanes/ether. The material was then dry loaded using silica onto a 10 g silica gel column and purified with flash chromatography (IPA/DCM, 0 to 20%) to give **4** (86 mg) as a white solid in 50% yield. mp = 216–219 °C; LC/MS t_R = 3.15 min (Characterization Method A); m/z = 491.35 (M + H⁺), m/z = 489.30 (M - H⁺), 535.30 (formic acid adduct); ¹H NMR (300 MHz, CDCl₃) δ = 8.06 (d, J = 7.9 Hz, 1 H), 7.72 (br. t, J = 5.0 Hz, 1 H), 7.37 - 7.20 (m, 5 H), 7.14 - 6.95 (m, 2 H), 6.94 - 6.83 (br. m, 1 H), 5.37 (app. t, J = 5.3, 6.2 Hz, 1 H), 4.56 - 4.47 (m, 1 H), 4.47 - 4.35 (m, 2 H), 3.53 - 3.42 (m, 1 H), 3.39 - 3.25 (br. m, 1 H), 3.09 (dd, J = 3.8, 9.7 Hz, 1 H), 3.02 - 2.89 (br. m, 1 H), 2.66 (dd, J = 4.7, 9.1 Hz, 1 H), 2.00 - 1.85 (br. m, 1 H), 1.84 - 1.67 (br. m, 1 H), 1.54 (br. m., 2 H), 1.43 (s, 9 H); ¹³C NMR (75 MHz, CDCl₃) δ = 174.4, 171.2, 156.9, 151.6 (dd, J = 53.5, 12.7 Hz), 148.4 (dd, J = 52.1, 12.4 Hz), 138.1, 134.8 (dd, J = 5.3, 3.9 Hz), 128.8, 127.9 (partially obs.) 127.8, 127.6, 125.4 (dd, J = 6.2, 3.6 Hz), 117.9 (J = 28.5, 17.3 Hz), 79.8, 56.4, 50.4, 43.7, 40.3, 37.0, 34.5, 28.6.

4.4.4 6-Nitro-1H-indazole-3-carbaldehyde (6). To a 500 mL round bottomed flask with stir bar was added NaNO₂ (6.38 g, 92.5 mmol) and H₂O (150 mL). 6-Nitroindole (**5**, 1.5 g, 9.3 mmol) was added to the above solution at 20 °C and the resultant mixture was stirred vigorously until evenly suspended (over 5 min.). To this bright yellow suspension was added 6 M HCl (14 mL) via addition funnel over 30 min. at 20 °C, and the resultant suspension was stirred for 90 min. A sample aliquot was taken from the reaction, filtered, and the precipitate was dissolved in a minimal amount of HPLC grade MeCN. Analysis with LCMS confirmed reaction completion. The product was vacuum filtered and the precipitate was washed with additional H₂O (50 mL). The precipitate was dried to give **6** (1.37 g) as an orange solid in 77% yield. m.p.: changed color from orange to brown between 120 and 160 °C, decomposed after 200 °C; LC/MS t_R = 2.40 min (Characterization Method A); m/z = 190.05 (M - H⁺); ¹H NMR (400 MHz, DMSO-d₆) δ = 10.22 (s, 1 H), 8.57 (d, J = 1.5 Hz, 1 H), 8.29 (dd, J = 0.5, 8.9 Hz, 1 H), 8.13 (dd, J = 2.0, 8.9 Hz, 1 H); ¹³C NMR (100 MHz, DMSO-d₆) δ = 187.7, 146.9, 124.0, 122.4, 118.7, 108.6.

4.4.5 6-Nitro-3-(pyrrolidin-1-ylmethyl)-1H-indazole (7). Aldehyde **6** (1.3 g, 6.80 mmol) was added to a round bottom flask with stir bar and sealed under nitrogen. A pre-mixed solution of anhydrous DCE:DMF:AcOH (300:30:0.3 mL), and pyrrolidine (2.8 mL, 34.01 mmol) was then added. The resulting yellow suspension became a reddish brown solution immediately after the addition of pyrrolidine, and the reaction was stirred for 20 min. Sodium triacetoxyborohydride (8.32 g, 39.24 mmol) was added in 3 portions at 5 min. intervals, and the resultant suspension was stirred at 20°C for 4 h. A sample aliquot was taken from the reaction, diluted with DCM, and washed with half saturated Na₂CO₃. The organic layer was separated, concentrated under reduced pressure, dissolved in a minimal amount of HPLC grade MeCN, and analyzed with LCMS to confirm reaction completion. The reaction was diluted with DCM (100 mL), washed with aqueous NaHCO₃ solution (100 mL), brine, dried over Na₂SO₄, vacuum filtered, and concentrated under reduced pressure to give 2.2 g of dark brown solid. The crude material was dissolved in minimal DCM and loaded onto a 50 g SiO₂ column and purified by flash chromatography (1N NH₃ in MeOH/DCM, 0–18%) to give **7** (1.09 g) as an orange brown shiny solid in 65% yield. mp = 37–40°C; LC/MS t_R = 0.91 min (Characterization Method A); m/z = 247.05 (M + H⁺); m/z = 245.15 (M - H⁺); ¹H NMR (400 MHz, CD₃OD) δ = 8.44 (d, J = 1.5 Hz, 1 H), 8.05 (d, J = 8.8 Hz, 1 H), 7.99 (dd, J = 2.0, 7.0 Hz, 1 H), 4.16 (s, 2 H), 2.78 - 2.71 (m, 4 H), 1.84 (spt, J = 3.3 Hz, 4

H); ¹³C NMR (75 MHz, CD₃OD) δ = 147.0, 142.8, 140.1, 125.2, 121.3, 114.9, 106.8, 53.9, 50.6, 23.1.

4.4.6 1-(2,6-Dichlorobenzyl)-6-nitro-3-(pyrrolidin-1-ylmethyl)-1H-indazole (8). Amine **7** (1.06 g, 4.30 mmol) and 2-(bromomethyl)-1,3-dichlorobenzene (1.03 g, 4.30 mmol) were added to a round bottom flask with stir bar and sealed under nitrogen. Anhydrous THF (50 mL) was added, followed by Cs₂CO₃ (1.4 g, 4.30 mmol) added in 3 portions (1 portion every 5 min.), and the flask was flushed with nitrogen. The resultant orange solution was stirred for 16 h at 20 °C. A sample aliquot was taken from the reaction, diluted with EtOAc, and washed with H₂O. The organic phase was separated, concentrated under reduced pressure, dissolved in CDCl₃ and analyzed with NMR to confirm reaction completion. The reaction was diluted with EtOAc (200 mL), washed with H₂O (200 mL) and brine, dried over Na₂SO₄, vacuum filtered, and concentrated under reduced pressure. The brown crude (1.5 g) was dissolved in minimal DCM and loaded onto a 50 g column and purified with flash chromatography (MeOH/DCM, 0-18%) to give **8** (1.33 g) as an orange brown shiny solid (containing ~10% dibenzylated side-product) in 76% yield. **8** was carried forward to the next step without additional purification. LC/MS t_R = 3.15 min (Characterization Method A); m/z = 404.80 (M + H⁺); for dibenzylated impurity LC/MS t_R = 3.63 min (Characterization Method A); m/z = 579.95 (M + H⁺).

4.4.7 1-(2,6-Dichlorobenzyl)-3-(pyrrolidin-1-ylmethyl)-1H-indazol-6-amine (9). To an oven dried pressure tube with stir bar was added indazole **8** (18 mg, 0.044 mmol) and MeOH (5 mL). FeCl₃·6H₂O (25 mol%, 2.6 mg), hydrazine monohydrate solution (65% active hydrazine, 0.05 mL, 0.7 mmol) and activated charcoal (20 mg) were added, the tube was capped tightly, and the reaction was stirred at 100 °C for 2 h. A sample aliquot was taken from the reaction, concentrated under reduced pressure, dissolved in a minimal amount of HPLC grade MeCN, and analyzed with LCMS to confirm reaction completion. The reaction was filtered, the charcoal and filtered solids were washed with MeOH (10 mL), and the filtrate was concentrated under reduced pressure to give crude material that was dry loaded using silica onto a 5 g silica column and purified with flash chromatography (0 to 20% MeOH/DCM) to give **9** (11.5 mg) as a white solid in 69% yield. LC/MS t_R = 1.44 min (Characterization Method A); m/z = 374.95 (M + H⁺); ¹H NMR (300 MHz, CDCl₃) δ = 7.57 (d, J = 8.5 Hz, 1 H), 7.37 (d, J = 8.2 Hz, 2 H), 7.31 - 7.20 (s, 1 H), 6.63 (d, J = 8.8 Hz, 1 H), 6.58 (s, 1 H), 5.63 (s, 2 H), 4.29 (s, 2 H), 3.96 (br. s., 2 H), 3.13 (br. s., 4 H), 1.84 (br. s., 4 H); ¹³C NMR (75 MHz, CDCl₃) δ = 146.5, 142.5, 137.0, 131.6, 130.3, 128.8, 121.0, 117.7, 113.7, 92.2, 52.3, 48.5, 48.2, 23.9.

4.4.8 (S)-4-Amino-N-benzyl-2-((S)-2-(3-(1-(2,6-dichlorobenzyl)-3-(pyrrolidin-1-ylmethyl)-1H-indazol-6-yl)ureido)-3-(3,4-difluorophenyl)propanamido)butanamide (11a-b, RWJ-58259)

Part 1: Coupling reaction. Aniline **9** (40 mg, 0.11 mmol) was added to an oven-dried round bottom flask with stir bar and sealed under nitrogen, then dry THF (16 mL) was added. The solution was cooled in a NaCl/H₂O bath (-5 °C) for 15 min., and then both DMAP (34.6 mg, 0.282 mmol) and triphosgene (12 mg, 0.04 mmol) were added consecutively. Immediately the reaction became a light orange suspension. The reaction was stirred at -5 °C for 8 min., and then amine **4** (35 mg, 0.071 mmol) was added and the reaction was stirred at -5 °C for 1 h, then for an additional 1 h while warming to room temperature. A sample aliquot was taken from the reaction, concentrated under reduced pressure, dissolved in a minimal amount of HPLC grade MeCN,

and analyzed with LCMS to confirm reaction completion. The reaction was concentrated under reduced pressure to give crude material **10** that could be either carried onto the next reaction without purification, or triturated with DCM/hexanes (1:1) to provide purer material.

Part 2: Boc removal. The round bottom flask already containing either the crude or triturated material **10** was cooled in a NaCl/H₂O bath for 10 min (-5 °C), and then HCl in dioxane (4 M, 3 mL) was slowly added. The reaction immediately became a heterogeneous dark red mixture and was stirred for 40 min. at -5 °C. A sample aliquot was taken from the reaction, concentrated under reduced pressure, dissolved in a minimal amount of HPLC grade MeCN, and analyzed with LCMS to confirm reaction completion. The reaction was concentrated under reduced pressure to give a red-orange solid that was free-based with 0.075 N NH₃ in MeOH and concentrated under reduced pressure. The crude product was dissolved in a minimal amount of DCM, loaded onto a 10 g normal phase silica column, and purified with flash chromatography (0-18% (0.75 N NH₃ in MeOH)/DCM) to give amine **11** (free base) as a light pink oily solid. The product was converted to the bis HCl salt **11a** by adding 1N HCl (1 mL) and lyophilizing overnight to give bis HCl salt as a light pink solid. LCMS Data t_R = (Characterization Method A); m/z = 396.65 [(M/2) + H⁺], 835.30 (M - H⁺, formic acid adduct); ¹H NMR (400 MHz, CD₃OD, free base) δ = 7.98 (s, 1 H), 7.75 (d, J = 8.7 Hz, 1 H), 7.45 - 7.38 (m, 2 H), 7.37 - 7.05 (m, 10 H), 5.56 (br. s., 2 H), 4.64 (s, 2 H), 4.60 - 4.48 (m, 2 H), 4.40 (d, J = 4.5 Hz, 2 H), 3.57 - 3.45 (m, 2 H), 3.29 - 3.12 (m, 3 H), 3.12 - 2.93 (m, 3 H), 2.35 - 2.20 (m, 1 H), 2.15 - 1.85 (m, 5 H); ¹³C NMR (75 MHz, CD₃OD, free base) δ = 173.3, 171.0, 156.2, 141.7, 139.0, 138.2, 136.6, 135.1, 134.3, 131.4, 130.3, 128.3, 128.1, 126.9, 126.7, 125.6, 119.7, 118.3, 118.1, 117.8, 117.0, 116.7, 115.2, 97.4, 55.6, 53.5, 51.0, 48.8, 42.6, 36.8, 36.6, 29.5, 22.6. *Alternatively, the crude product could be dissolved in DMSO and purified in batches with preparative HPLC (method A). Pooled fractions were concentrated under reduced pressure in lukewarm water (to remove MeCN) and lyophilized overnight to give 11b (36.3 mg) as a light pink oil in 58% yield over 2 steps.*

4.4.9 N-((S)-4-(Benzylamino)-3-((S)-2-(3-(1-(2,6-dichlorobenzyl)-3-(pyrrolidin-1-ylmethyl)-1H-indazol-6-yl)ureido)-3-(3,4-difluorophenyl)propanamido)-4-oxobutyl)pent-4-ynamide (12, DG-207). *Note:* If using **11b**, it is necessary to convert this compound to the bis HCl salt (**11a**) prior to performing this EDC coupling reaction as it has been seen that the formate salt will give a mixture of products. To a 5 mL scintillation vial with stir bar was added **11b** (5.5 mg, 6.4 μmol). The vial was purged with N₂ for 5 min and then dry dioxane (0.5 mL) and 4 M HCl in dioxane (0.3 mL, 172 eq.) was added. The reaction was stirred for 5 min, concentrated under reduced pressure, and then lyophilized to give the bis HCl salt (**11a**). The vial containing **11a** was purged with nitrogen and then dissolved in dry DMF (0.5 mL). 4-Pentynoic acid (1 mg, 10 μmol), HOBT (1.2 mg, 8.9 μmol), EDC·HCl (3 mg, 12 μmol), and DIPEA (9.6 μL, 55 μmol) were added consecutively and the reaction was stirred for 5 h under N₂. A sample aliquot was taken from the reaction, concentrated under reduced pressure, dissolved in a minimal amount of HPLC grade MeCN, and analyzed with LCMS to confirm reaction completion. The reaction was diluted with EtOAc (5 mL) and washed with saturated NaHCO₃ (5 mL) and brine (5 mL). The organic layer was dried over MgSO₄, filtered, and concentrated under reduced pressure to give crude material that was dissolved in DMSO and purified with preparative HPLC (method A) to give **12** (2.4 mg) as a colorless oily residue in 43%. ¹H NMR (600 MHz, CD₃OD) δ = 8.54 (br. s, 1 H, formate CHO), 7.94 (d, J = 1.2 Hz, 1 H), 7.67 (d, J = 8.8 Hz,

1 H), 7.44 (s, 1 H), 7.42 (s, 1 H), 7.33 (dd, $J = 1.2, 7.6$ Hz, 1 H), 7.28 - 7.11 (m, 7 H), 7.09 - 7.05 (br. m, 1 H), 6.99 (dd, $J = 1.8, 7.0$ Hz, 1 H), 5.61 (dd, $J = 14.1, 2.3$ Hz, 2 H), 4.55 (dd, $J = 2.9, 5.3$ Hz, 1 H), 4.44 - 4.41 (m, 1 H), 4.39 (s, 2 H), 4.37 - 4.34 (br. m, 2 H), 3.58 (dd, $J = 4.7, 6.5$ Hz, 1 H), 3.51 (dd, $J = 4.7, 6.5$ Hz, 1 H), 3.20 - 3.13 (m, 2 H), 3.12 - 3.04 (br. s, 3 H), 2.99 (dd, $J = 5.9, 8.2$ Hz, 1 H), 2.41 - 2.37 (m, 2 H), 2.34 - 2.29 (m, 2 H), 2.21 - 2.20 (m, 1 H), 2.07 - 1.98 (m, 2 H), 1.93 - 1.86 (m, 4 H). LC/MS $t_R = 3.46$ min (Characterization Method A); $m/z = 871.15$ ($M + H^+$), $m/z = 915.30$ ($M - H^+$, formic acid adduct). HRMS (ESI-TOF) calcd. for $C_{45}H_{47}C_{12}F_2N_8O_4$ ($M + H^+$) 871.3060, found 871.2988.

4.5 General procedures for the syntheses of parmodulins and Fairlie's PAR2 antagonist (**32**, TJF-5)

Method A: Amide coupling using EDC

To a round bottom flask with stir bar under nitrogen were added the appropriate carboxylic acid and anhydrous DCM/DMF (85:15; 0.2-0.6 M). The amine HCl salt to be coupled (1.2 eq.), HOBT (1.2 eq.), EDC·HCl (1.2 eq.), and DIPEA (2.1 eq.) were added and the reaction was stirred under nitrogen. The reaction was diluted with DCM (75 mL), washed with saturated $NaHCO_3$, 1M HCl (30 mL), brine, dried over Na_2SO_4 , filtered, and concentrated under reduced pressure to give crude product.

Method B: Formation and acylation with acid chloride

To an oven-dried round bottom flask with stir bar under nitrogen were added the carboxylic acid, dry DCM, and 3Å molecular sieves. Oxalyl chloride (1.2 eq.) and a catalytic amount of DMF (1-2 mol %) were added and the reaction was stirred while attached to a bubbler (to monitor production of CO_2) at 20 °C for 2-3 h. The amine HCl salt (1 eq.) in DCM and DIPEA (2 eq.) were added and the reaction was stirred under nitrogen for 3-6 h. The reaction was diluted with EtOAc and washed with half-saturated aqueous $NaHCO_3$, 1M HCl (30 mL), brine, dried over Na_2SO_4 , filtered, and concentrated under reduced pressure to give crude material.

Method C: Acylation with acid chloride

To an oven-dried round bottom flask with stir bar and under nitrogen were added the acid chloride, dry DCM, and 3Å molecular sieves. The amine HCl salt (1 eq.) in DCM and DIPEA (2 eq.) were added and the reaction was stirred under nitrogen for 3-6 h. The reaction was diluted with EtOAc and washed with half-saturated aq. $NaHCO_3$, 1M HCl (30 mL), brine, dried over Na_2SO_4 , filtered, and concentrated under reduced pressure to give crude product.

Method D: Amide coupling using HATU

To a round bottom flask with stir bar under nitrogen was added the carboxylic acid and anhydrous DCM. The amine (1.2 eq.), HATU (1.2 eq.), and DIPEA (1.2 eq.) were added and the reaction was stirred under nitrogen. The reaction was diluted with DCM (75 mL) and washed with saturated $NaHCO_3$, 1M HCl (30 mL), brine, dried over Na_2SO_4 , filtered, and concentrated under reduced pressure to give crude material.

Method E: Boc removal

To an 8 mL vial with stir bar and under nitrogen was added the Boc-protected amine, dry DCM, and a large excess of TFA. The reaction was allowed to stir at 20 °C for 1-2 h. The reaction mixture was concentrated under reduced pressure (passed through a base trap) to give crude material that was suspended in EtOAc and washed with half-saturated aq. $NaHCO_3$, and brine. The reaction was dried over Na_2SO_4 , filtered, and concentrated

under reduced pressure to give crude material that was carried onto the next peptide coupling step without further purification.

4.6 Synthesis of parmodulins

4.6.1. 2-Bromo-N-(3-propionamidophenyl)benzamide (18, EMG-21). Analogue **18** (EMG-21) was synthesized by coupling of aniline-HCl salt **15** with propionic acid (16 μ L, 0.20 mmol) according to general method A. The crude material was loaded using Celite onto a 5 g silica gel column and purified by flash chromatography (EtOAc:hexanes, 0-100%) to give **18** as an off-white solid (19 mg) in 32% yield. LC/MS $t_R = 4.47$ min (Characterization Method A); $m/z = 348.70$ ($M + H^+$); 1H NMR (300 MHz, CD_3OD) $\delta = 7.98$ (t, $J = 2.0$ Hz, 1H), 7.69 (dd, $J = 7.9, 1.1$ Hz, 1H), 7.56-7.43 (m, 2H), 7.42-7.35 (m, 3H), 7.29 (t, $J = 8.0$ Hz, 1H), 2.39 (q, $J = 7.6$ Hz, 2H), 1.20 (t, $J = 7.6$ Hz, 3H); HRMS (ESI): m/z [$M + H^+$]⁺ calcd. for $C_{16}H_{15}N_2O_2Br$, 347.0390; found, 347.0357.

4.6.2 2-Bromo-N-(3-pentanamidophenyl)benzamide (19, EMG-23). Analogue **19** was synthesized by peptide coupling of pentanoyl chloride (22 μ L, 0.19 mmol) with aniline·HCl salt **15** according to general method C. The crude material was then dry loaded using Celite onto a 5 g silica gel column and purified by flash chromatography (EtOAc:hexanes, 0-100%) to give **19** as an off white solid (25 mg) in 39% yield. LC/MS $t_R = 5.12$ min (characterization method A); $m/z = 376.75$ ($M + H^+$); 1H NMR (300 MHz, CD_3OD) $\delta = 7.99$ (t, $J = 2.0$ Hz, 1H), 7.69 (dd, $J = 7.9, 1.0$ Hz, 1H), 7.56-7.43 (m, 2H), 7.42-7.34 (m, 3H), 7.29 (t, $J = 8.0$ Hz, 1H), 2.38 (t, $J = 7.3$ Hz, 2H), 1.73-1.68 (m, 2H), 1.41 (h, $J = 7.3$ Hz, 2H), 0.97 (t, $J = 7.3$ Hz, 3H); HRMS (ESI): m/z [$M + H^+$]⁺ calcd. for $C_{18}H_{19}N_2O_2Br$, 375.0703; found, 375.0669.

4.6.3 2-Bromo-N-(3-(pent-4-ynamido)phenyl)benzamide (20, RR-10). Alkyne analogue **20** was synthesized by acylation of pent-4-ynoic acid (50 mg, 0.51 mmol) with aniline·HCl salt **15** according to general method B. The crude material was dissolved in a minimal amount of DCM, loaded onto a 10 g silica column, and purified by flash chromatography (EtOAc:hexanes, 0-100%) to give **20** as a white solid (13 mg) in 7% yield. LC/MS $t_R = 6.44$ min (characterization method A); $m/z = 372.60$ ($M + H^+$); 1H NMR (400 MHz, $CDCl_3$) $\delta = 7.95$ (s, 1 H), 7.71 (s, 1 H), 7.67 - 7.58 (m, 2 H), 7.52 (br. s., 1 H), 7.46 - 7.27 (m, 5 H), 2.66 - 2.52 (m, 4 H), 2.09 - 2.05 (m, 1 H); ^{13}C NMR (75 MHz, $CDCl_3$, 1% CD_3OD) $\delta = 170.5, 167.0, 138.8, 138.4, 133.5, 131.6, 129.6, 129.2, 127.7, 119.6, 116.4, 116.1, 111.7, 83.0, 69.5, 36.0, 14.8$. HRMS (ESI) calcd. for $C_{18}H_{15}BrN_2O_2$ ($M + H^+$), 371.0390, found 371.0364.

4.6.4 N-(3-Aminophenyl)butyramide HCl salt (23). A solution of nitroarene **22** (5.95 g, 28.7 mmol) in MeOH (85 mL) was flushed with nitrogen for 5 min. To a 10 mL scintillation vial under nitrogen was suspended 10% Pd/C (0.95 g, 0.88 mmol) in MeOH/ H_2O (4:1) and the resultant suspension was added to the solution of **22**. A balloon was filled with H_2 gas and inserted into the flask and the reaction was purged with H_2 gas for 2 min. The reaction was left to stir under a full balloon of H_2 for 16 h, after which an aliquot was taken from the reaction, filtered through celite and cotton, and concentrated under reduced pressure. The aliquot was dissolved in a minimal amount of HPLC grade MeCN, and analyzed with LCMS to confirm reaction completion. The reaction was filtered through a funnel packed with Celite. The filter cake was washed with MeOH and the filtrate was concentrated under reduced pressure, to give crude material that was dissolved in DCM and added to 0.9 M HCl in ether (40 mL). The reaction was stirred for 5 min at 20 °C to give a precipitate that was filtered and washed with excess DCM. The

precipitate was dried to give the previously described aniline **23**¹² (5.60 g) as a grey solid in 92% yield.

4.6.5. 2-Azido-*N*-(3-butylamidophenyl)benzamide (**26**, **DG-5**).

Part 1: Diazotization. To a round bottom flask with stir bar was added 2-aminobenzoic acid **24** (1.50 g, 10.9 mmol) and deionized H₂O (100 mL). The reaction was cooled to 0 °C and conc. aq. HCl (5 mL) was added dropwise over 5 min. A solution of NaNO₂ (0.75 g, 11 mmol) in H₂O (1 mL) was added, and the reaction was stirred for 10 min. at 2 °C. A solution of NaOAc (0.63 g, 11 mmol) in H₂O (1 mL) and a solution of NaN₃ (0.71 g, 10.9 mmol) in H₂O (1 mL) were added consecutively and the reaction was stirred for 30 min at 2 °C. A sample aliquot was taken from the reaction, diluted with TBME, extracted and the organic phase was separated, concentrated under reduced pressure, dissolved in a minimal amount of HPLC grade MeCN, and analyzed with LCMS to confirm reaction completion. The reaction was extracted with TBME (100 mL) and the organic layer was washed with brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give 2-azidobenzoic acid **25** (1.5 g) as a yellow solid in 85% yield. TLC: mobile phase: Ethyl acetate/hexanes/formic acid (25/25/1), R_f: 0.4; ¹H NMR (300 MHz, CDCl₃) δ = 8.19 - 8.14 (m, 1 H), 7.67 - 7.59 (m, 1 H), 7.29 (dd, *J* = 2.0, 7.3 Hz, 2 H).

Part 2: Acylation. Crude azide analogue **26** was synthesized by acylation of aniline•HCl salt **23** with 2-azidobenzoic acid **25** according to general method B. The crude material was dissolved in a minimal amount of DCM, loaded onto a 25 g column, and purified with flash chromatography (EtOAc/hexanes, 0–80%) to give **26** (160 mg) as a colorless oil in 80% yield. TLC: mobile phase: ethyl acetate/hexanes (50:50), R_f: 0.6; LC/MS t_R = 5.01 min (characterization method A); *m/z* = 323.80 (M + H⁺) 368.00 (M + formate); ¹H NMR (400 MHz, CDCl₃) δ = 9.36 (br. s., 1 H), 8.26 - 8.19 (m, 1 H), 7.98 (br. s., 1 H), 7.61 - 7.50 (m, 1 H), 7.41 (d, *J* = 7.6 Hz, 2 H), 7.36 - 7.23 (m, 5 H), 2.33 (t, *J* = 7.4 Hz, 2 H), 1.76 (sxt, *J* = 7.4 Hz, 2 H), 0.99 (t, *J* = 7.3 Hz, 3 H); ¹³C NMR (101 MHz, CDCl₃) δ = 162.7, 138.6, 138.4, 136.9, 132.1, 129.6, 125.4, 118.4, 115.6, 111.6, 39.7, 19.0, 13.7; HRMS calcd. for C₁₇H₁₇N₅O₂ (M + H⁺) 324.1455, found 324.1415.

4.7 Synthesis of TJF-5

4.7.1 tert-butyl ((2*S*,3*S*)-1-((2-Methoxybenzyl)amino)-3-methyl-1-oxopentan-2-yl)carbamate (28**).** Analogue **28** was synthesized by peptide coupling of (2*S*,3*S*)-2-(*tert*-butoxycarbonylamino)-3-methyl-pentanoic acid **27** (81 mg, 0.35 mmol) with (2-methoxybenzylamine (38 μL, 0.29 mmol) according to general method D. The crude material was dissolved in a minimal amount of DCM, loaded onto a 10 g silica gel column, and purified by flash chromatography (EtOAc/hexanes, 0–100%) to give **28**⁴⁸ (62 mg) as a white foam in 51% yield. ¹H NMR (300 MHz, CDCl₃) δ = 7.24 (d, *J* = 7.4 Hz, 1 H), 6.93 - 6.83 (m, 2 H), 6.44 (br. s., 1 H), 5.16 (br. s., 1 H), 4.43 (ddd, *J* = 6.3, 8.6 Hz, 2 H), 3.92 (t, *J* = 7.0 Hz, 1 H), 3.84 (s, 3 H), 2.05 - 1.78 (m, 1 H), 1.41 (s, 9 H), 1.14 - 1.01 (m, 1 H), 0.87 (t, *J* = 7.2 Hz, 6 H).

4.7.2 tert-butyl ((*S*)-3-Cyclohexyl-1-(((2*S*,3*S*)-1-((2-methoxybenzyl)amino)-3-methyl-1-oxopentan-2-yl)amino)-1-oxopropan-2-yl)carbamate (30**).** **Part 1. Boc removal.** Intermediate **28** was deprotected to give **29** according to general method E and was advanced to the amide coupling step.

Part 2. Amide coupling. Analogue **30** was synthesized from **29** (29.0 mg, 0.12 mmol) and (2*S*)-2-(*tert*-butoxycarbonylamino)-3-cyclohexyl-propanoic acid hydrate (40 mg, 0.14 mmol) according to general method D. The crude material was dissolved in a

minimal amount of DCM, loaded onto a 10 g silica gel column, and purified by flash chromatography (MeOH/DCM, 0–12%) to give **30**⁴⁸ (51 mg) as a white solid in 37% yield over 2 steps. ¹H NMR (300 MHz, CDCl₃) δ = 7.23 (d, *J* = 7.6 Hz, 2 H), 6.89 (q, *J* = 7.6 Hz, 2 H), 6.69 (d, *J* = 8.8 Hz, 1 H), 6.56 - 6.46 (m, 1 H), 4.90 (br. d, *J* = 7.3 Hz, 1 H), 4.42 (ddd, *J* = 9.7, 16.4 Hz, 2 H), 4.25 (t, *J* = 8.8 Hz, 1 H), 3.84 (s, 3 H), 3.07 (s, 1 H), 2.95 (s, 1 H), 1.75 - 1.55 (m, 8 H), 1.48 - 1.35 (m, 15 H), 1.23 - 1.01 (m, 6 H).

4.7.3 *N*-((*S*)-3-Cyclohexyl-1-(((2*S*,3*S*)-1-((2-methoxybenzyl)amino)-3-methyl-1-oxopentan-2-yl)amino)-1-oxopropan-2-yl)isoxazole-5-carboxamide (32**).** **Part 1. Boc removal.** Intermediate **30** was deprotected to give free amine **31** according to general method E and advanced to the amide coupling step.

Part 2. Amide coupling. Crude **32** (**TJF-5**) was synthesized from amine intermediate **31** (21.5 mg, 0.05 mmol) and isoxazole-5-carboxylic acid (7.3 mg, 0.06 mmol) according to general method D. The crude material was dissolved in a minimal amount of DCM, loaded onto a 10 g silica gel column, and purified by flash chromatography (MeOH/DCM, 0–5%) to give **32**⁴⁸ (9.1 mg) as a white solid in 16% yield over 2 steps. LC/MS t_R = 6.46 min (Characterization Method A); *m/z* = 499.00 (M + H⁺), 521.00 (M + Na⁺), 498.25 (M - H⁺), 543.25 (M + formic acid adduct); ¹H NMR (300 MHz, CDCl₃) δ = 8.32 (d, *J* = 1.5 Hz, 1 H), 7.30 (s, 1 H), 7.33 - 7.21 (m, 1 H), 6.96 - 6.82 (m, 4 H), 6.46 (app. t, *J* = 5.6 Hz, 1 H), 4.74 (dd, *J* = 8.5 Hz, 1 H), 4.44 (ddd, *J* = 8.8, 20.8 Hz, 2 H), 4.27 (t, *J* = 7.6 Hz, 1 H), 3.86 (s, 3 H), 1.89 - 1.73 (m, 3 H), 1.72 - 1.58 (m, 7 H), 1.52 - 1.38 (m, 1 H), 1.37 - 1.24 (m, 2 H), 1.22 - 1.02 (m, 4 H), 0.98 - 0.88 (m, 2 H), 0.88 - 0.79 (m, 6 H).

4.8 General information: Assays

Water for solutions was deionized and filtered through charcoal (Milli-Q by Millipore) to a resistance of 18 MΩ. TFLLRN-NH₂ (PAR1 agonist) was obtained from AnaSpec (cat# AS-62937) or was synthesized (as the TFA salt) by Trudy Holyst, Blood Research Institute Protein Chemistry Core. SFLLRN-NH₂ (TFA salt) (PAR1/2 agonist) was obtained from Bachem (cat# H-2936.0005) or was synthesized (as the TFA salt) by Trudy Holyst, Blood Research Institute Protein Chemistry Core. SLIGKV-NH₂ (TFA salt) (PAR2 agonist) was obtained from Bachem (cat# H-5042.0025) or AnaSpec (cat# AS-60217-1). AYPGFK-NH₂ (TFA salt) (PAR4 agonist) was obtained from AnaSpec (cat# AS-60218-1). Atopaxar (E5555 HBr) was obtained from Axon Medchem (cat# 2030). Vorapaxar (SCH 530348) was obtained from Axon Medchem (cat# 1755). Q94 (HCl) was obtained from Axon Medchem (cat# 2055).

Adherent EA.hy926 cells (ATCC CRL-2922) were used for all assays performed. The cells were cultured using the suggested protocol from the manufacturer (ATCC), except that 150 cm² tissue culture flasks were used, and DMEM complete media was prepared as described below. Cells were frozen in 40% fetal bovine serum (FBS), 50% complete media, and 10% DMSO in liquid nitrogen, and used between passages 2 and 11. All manipulations prior to the assays were performed in a sterile laminar flow cell culture hood.

Cell counting was performed with an automated cell counter (Countess, Invitrogen) using Invitrogen cell counting chamber slides and trypan blue stain. All assays were run in 96-well plates (Corning Costar #3603, polystyrene black wall, clear bottom). Unless otherwise noted, media exchanges with 96-well plates were performed using an automated liquid handler (Beckmann-

Coulter, Biomek 3000) with 220 μ L pipette tips. Cells were imaged with an Evos Fl inverted microscope. All assays were run on a multimode plate reader (Perkin Elmer EnSpire). Data was exported to Microsoft Excel for nominal processing then analyzed with GraphPad Prism (versions 5 or 6).

4.9 Preparation of stock solutions

Complete cell media (for EA.hy926 cells). Dulbecco's Modified Eagle Medium (DMEM) with 4.5 g/L glucose (500 mL) was supplemented with 200 mM L-glutamine (10 mL, 4 mM final conc.), 100 mM sodium pyruvate (2.5 mL, 1 mM final conc.), 7.5% (w/v) sodium bicarbonate (5.0 mL, 0.75 g/L final conc.), 100X penicillin and streptomycin (Corning #30-002-Cl, 2.0 mL) and heat inactivated FBS (50 mL). The prepared media was stored in a refrigerator (5 °C) until needed.

Probenecid solution (250 mM, aqueous). Probenecid (Alfa Aesar, catalog # B20010-88, 36 mg) was added to a 1 mL microcentrifuge tube and dissolved in 0.6 M NaOH (500 μ L). NaOH (0.6 M) was prepared by dissolving NaOH (480 mg) in H₂O (20 mL). This solution was made the day of the experiment and was sufficient for up to 50 mL of buffer (for 2 96-well plates).

HBSS-HEPES wash buffer. HEPES (1.19 g) was added to a 500 mL bottle of Ca/Mg/phenol red-free HBSS to make a 10 mM solution in HEPES. MgCl₂ (1 M solution, 500 μ L) and CaCl₂ (1 M solution, 500 μ L) were added and the solution was stored in the refrigerator. Before use, the desired amount (46 mL total per 96 well plate) was warmed to room temperature and added to a 50 mL centrifuge tube, then supplemented with 250 mM probenecid solution (1:100, i.e. 46 mL of buffer needs 460 μ L of probenecid solution) to give a 2.5 mM probenecid solution. The pH was adjusted to 7.4 if necessary by adding aq. HCl.

10% Pluronic F-127 in DMSO. Pluronic F-127 (Sigma Aldrich, 20 mg) was added to a 1 mL microcentrifuge tube and dissolved with DMSO (200 μ L), to make a 10% w/v solution. This solution was stored at room temperature.

Fluo-4/AM loading buffer. Fluo-4/AM (Invitrogen via Fisher, catalogue # F23917, 50 μ g) was dissolved in DMSO (24 μ L), then 6 μ L of the solution (light-sensitive) was transferred to a 200 μ L microcentrifuge tube and mixed with 6 μ L of 10% pluronic F-127 in DMSO (sufficient for 1 96-well plate). The remainder of the Fluo-4/AM solution was stored in the freezer at -30 °C. The solution was then transferred to a foil-covered 14 mL centrifuge tube already containing HBSS-HEPES wash buffer (6 mL) and vortexed for 5 seconds. 50 μ L of this prepared dye solution (2 μ M) can be added to each well of a 96-well plate containing adherent cells.

4.10 Preparation of agonist/antagonist solutions

Prior to the assay, stock solutions of PAR1 and PAR2 agonists were prepared by dissolving the agonists in 100% H₂O (sterile, deionized) to give stock solutions (10 mM, 5 mM, or 1 mM depending on agonist). These were aliquoted into sterile microcentrifuge tubes and stored in a -30 °C freezer, and thawed before use. Stock solutions of antagonists were prepared by dissolving with 100% DMSO, generally in glass vials, to give stock solutions (31.6 mM, 10 mM, or 1 mM depending on antagonist used) that were stored in the freezer at -30 °C, and thawed in a desiccator before use.

Agonist solutions: Prepared as 10 mM stock solutions in 100% water (sterile, deionized). Diluted solutions were then prepared with water to give half log concentrations.

Antagonist solutions: Vehicle (diluent) was prepared by mixing 1:9 DMSO:HBSS-HEPES wash buffer. An aliquot from the antagonist stock solution (in 100% DMSO) was then diluted 10x with vehicle to give a 10% DMSO solution, then subsequent dilutions were made with vehicle to give a consistent concentration of DMSO (1%).

4.11 Protocol for calcium mobilization assay with adherent endothelial cells

Part 1. Cell Preparations (Aseptic Conditions)

Unless otherwise noted, cells were plated at 25,000 cells per well in 100 μ L, which corresponds to 250,000 cells/mL. Complete cell media (25 mL per plate) was transferred to a 50 mL centrifuge tube and warmed to 37 °C in a water bath. A vial of frozen EA.hy926 cells (ATCC CRL-2922) was taken from liquid N₂ storage and warmed to 37 °C in a water bath for 5 min. Complete cell media (1 mL) was added dropwise to the vial and the contents were mixed and transferred to a 14 mL centrifuge tube. The cells were centrifuged at 130 g for 3 min and the supernatant was removed via aspiration with a Pasteur pipet. The cells were re-suspended in complete cell media (2 mL) and the suspension was mixed thoroughly with a pipette to gently break up the cell pellet to give an evenly distributed suspension in the tube. A small fraction of this suspension (25 μ L) was mixed with 25 μ L of trypan blue solution and a cell count was taken. Based on the cell count, an appropriate volume of complete cell media was added to the 2 mL of cell suspension to assure that each well of the 96-well plate would have 25,000 cells. The suspension was gently mixed, and the contents of the tube were slowly poured into a sterile disposable pipetting reservoir. This suspension containing the cells was then added (column by column, 100 μ L per well) to a 96-well plate (Corning Costar #3603, black wall, clear bottom) with a multichannel pipette. The plate was gently rocked back and forth to get even coverage of cells and the plate was incubated at 37 °C (5% CO₂) for 40–48 h.

Part 2. Loading of Fluo-4 dye

The 96-well plate was removed from the incubator after 40–48 h and then checked with an EVOS Fl microscope and optionally imaged (20X magnification) to confirm that cells were >80% confluent and viable. Note: All media exchanges were performed with the Biomek 3000, unless otherwise noted. The complete cell media was removed and HBSS-HEPES wash buffer (100 μ L) was added to each well. The plate was left to sit at room temperature for 5 min. to allow the probenecid to be absorbed, then the media was removed. Fluo-4/AM solution (50 μ L per well) was added (in the dark), and the plate was incubated at 37 °C (5% CO₂) for 40–50 min. The dye solution was removed from each well and the wells were washed with HBSS-HEPES wash buffer (50 μ L/well) twice. HBSS-HEPES wash buffer (200 μ L/well) was added to each well and the plate was imaged with the EVOS Fl microscope (20X magnification) to verify that the dye has been absorbed into the cells.

Part 3. Running the assay

Basic plate reader info and settings:

Instrument: Perkin Elmer EnSpire plate reader

Temperature: 37°C

Excitation wavelength: 485 nm

Emission wavelength: 525 nm

Measurement height: 7.8 mm (optimized)

Number of flashes: 100

Number of repeats: 250, every 5 sec. Experiment was halted after the emission signal starts to decrease from the maximum.

Vehicle or antagonist (2 μ L) were added (single or multichannel pipette) to each well, and the plate was incubated in the plate reader for 15 min at 37 °C. Response to agonists was measured 2 columns at a time, since this plate reader scanned wells individually, but plate readers that measure wells concurrently could optionally support assays with all agonists added concurrently, for higher throughput. Upon starting the assay, the basal level of the 2 columns was measured (20 scans) prior to the addition of the agonist. The agonist (2 μ L) was then added (over 10 seconds with a multichannel pipette or over 90–120 seconds with a single channel pipette), and the wells were analyzed with the plate reader using the settings above.

Part 4. Data analysis

Concentration-response curves were obtained by measuring the maximal increase in emission intensity (Δ Relative Fluorescence Units, RFU) from basal levels (average of 20 scans prior to agonist addition), then dividing by the average basal levels to give Δ RFU/basal. From this data, % stimulation was obtained by normalizing Δ RFU/basal for each column of the plate with a well from the same column where vehicle had been added instead of antagonist solution. The Δ RFU/basal value from the vehicle well was considered to be 100% stimulation for each column. This normalization was necessary due to increasing background signal over time, presumably due to dye efflux. Data (% stimulation versus log concentration) was then plotted for 3 or more repetitions and analyzed using GraphPad Prism (versions 5 or 6). All concentration-response curves were fitted using 4-variable non-linear regression, showing the standard error of the mean (SEM) for each concentration.

4.12 Reversibility Assay Protocol

Part 1. Preparation of Plate for Assay

Cell preparations (under aseptic conditions), loading of probenecid and Fluo-4 dye, the washing/removal of dye, and the replacement with HBSS-HEPES buffer solution (200 μ L) were performed as detailed above.

Part 2. Wash Procedure

Vehicle or antagonist (2 μ L) was added to the wells of interest (already containing 200 μ L of HBSS-HEPES wash buffer) and the plate was incubated in the plate reader for 20–30 min. The wash buffer was carefully removed from the wells with a multichannel pipette, replaced with fresh HBSS-HEPES wash buffer (200 μ L) and incubated in the plate reader for 10 min. at 37 °C. Wash buffer was carefully removed from the wells again with a multichannel pipette and replaced with fresh HBSS-HEPES wash buffer (200 μ L). The plate was incubated again in the plate reader for 10 min., then the assay was performed according to the regular protocol above. Note that the washing steps can alternatively be performed with the automated liquid handler.

Author Contributions

D.M.G.: Synthesized parmodulins; validated and completed the synthesis of RWJ-58259 and derivatives; developed the calcium mobilization assay; assayed compounds; processed and analyzed data; trained and mentored junior contributors; wrote sections of the S.I. and manuscript; **M.W.M.:** Validated and completed the synthesis of RWJ-58259 and derivatives; developed the calcium mobilization assay; assayed compounds; processed and analyzed data; wrote sections of the S.I. and manuscript; **R.R.:** Synthesized parmodulins, trained and mentored junior contributors. **K.K.:** Assisted with the synthesis

of RWJ-58259. **T.J.F.:** Synthesized Fairlie's PAR2 antagonist. **E.G.:** Synthesized parmodulins. **C.D.:** Conceived the project; developed the calcium mobilization assay; processed and analyzed data; trained and mentored junior contributors; wrote the manuscript; obtained funding.

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References

- (1) Coughlin, S. R. (2000) Thrombin signalling and protease-activated receptors. *Nature* 407, 258–264.
- (2) Adams, M. N., Ramachandran, R., Yau, M.-K., Suen, J. Y., Fairlie, D. P., Hollenberg, M. D., and Hooper, J. D. (2011) Structure, function and pathophysiology of protease activated receptors. *Pharmacology and Therapeutics* 130, 248–282.
- (3) Hamilton, J. R., and Trejo, J. (2017) Challenges and Opportunities in Protease-Activated Receptor Drug Development. *Annu. Rev. Pharmacol. Toxicol.* 57, 349–373.
- (4) Vu, T.-K. H., Hung, D. T., Wheaton, V. I., and Coughlin, S. R. (1991) Molecular cloning of a functional thrombin receptor reveals a novel proteolytic mechanism of receptor activation. *Cell* 64, 1057–1068.
- (5) Hein, L., Ishii, K., Coughlin, S. R., and Kobilka, B. K. (1994) Intracellular targeting and trafficking of thrombin receptors. A novel mechanism for resensitization of a G protein-coupled receptor. *Journal of Biological Chemistry*.
- (6) Chackalamannil, S., Wang, Y., Greenlee, W. J., Hu, Z., Xia, Y., Ahn, H.-S., Boykow, G., Hsieh, Y., Palamanda, J., Agans-Fantuzzi, J., Kurowski, S., Graziano, M., and Chintala, M. (2008) Discovery of a Novel, Orally Active Himbacine-Based Thrombin Receptor Antagonist (SCH 530348) with Potent Antiplatelet Activity. *J. Med. Chem.* 51, 3061–3064.
- (7) O'Callaghan, K., Kuliopulos, A., and Covic, L. (2012) Turning Receptors On and Off with Intracellular Pepducins: New Insights into G-protein-coupled Receptor Drug Development. *Journal of Biological Chemistry* 287, 12787–12796.
- (8) Gurbel, P. A., Bliden, K. P., Turner, S. E., Tantry, U. S., Gesheff, M. G., Barr, T. P., Covic, L., and Kuliopulos, A. (2016) Cell-Penetrating Pepducin Therapy Targeting PAR1 in Subjects With Coronary Artery Disease. *Arteriosclerosis, Thrombosis, and Vascular Biology* 36, 189–197.
- (9) Wong, P. C., Seiffert, D., Bird, J. E., Watson, C. A., Bostwick, J. S., Giancarli, M., Allegretto, N., Hua, J., Harden, D., Guay, J., Callejo, M., Miller, M. M., Lawrence, R. M., Banville, J., Guy, J., Maxwell, B. D., Priestley, E. S., Marinier, A., Wexler, R. R., Bouvier, M., Gordon, D. A., Schumacher, W. A., and Yang, J. (2017) Blockade of protease-activated receptor-4

- (PAR4) provides robust antithrombotic activity with low bleeding. *Sci Transl Med* 9, eaaf5294.
- (10) Ramachandran, R., Noorbakhsh, F., DeFea, K., and Hollenberg, M. D. (2012) Targeting proteinase-activated receptors: therapeutic potential and challenges. *Nat Rev Drug Discov* 11, 69–86.
- (11) Dowal, L., Sim, D. S., Dilks, J. R., Blair, P., Beaudry, S., Denker, B. M., Koukos, G., Kuliopulos, A., and Flaumenhaft, R. (2011) Identification of an antithrombotic allosteric modulator that acts through helix 8 of PAR1. *Proceedings of the National Academy of Sciences* 108, 2951–2956.
- (12) Dockendorff, C., Aisiku, O., Verplank, L., Dilks, J. R., Smith, D. A., Gunnink, S. F., Dowal, L., Negri, J., Palmer, M., Macpherson, L., Schreiber, S. L., and Flaumenhaft, R. (2012) Discovery of 1,3-Diaminobenzenes as Selective Inhibitors of Platelet Activation at the PAR1 Receptor. *ACS Med Chem Lett* 3, 232–237.
- (13) Aisiku, O., Peters, C. G., De Ceunynck, K., Ghosh, C. C., Dilks, J. R., Fustolo-Gunnink, S. F., Huang, M., Dockendorff, C., Parikh, S. M., and Flaumenhaft, R. (2015) Parmodulins inhibit thrombus formation without inducing endothelial injury caused by vorapaxar. *Blood* 125, 1976–1985.
- (14) Riewald, M., Petrovan, R. J., Donner, A., Mueller, B. M., and Ruf, W. (2002) Activation of endothelial cell protease activated receptor 1 by the protein C pathway. *Science* 296, 1880–1882.
- (15) Cheng, T., Liu, D., Griffin, J. H., Fernandez, J. A., Castellino, F., Rosen, E. D., Fukudome, K., and Zlokovic, B. V. (2003) Activated protein C blocks p53-mediated apoptosis in ischemic human brain endothelium and is neuroprotective. *Nat Med* 9, 338–342.
- (16) Feistritz, C., Lenta, R., and Riewald, M. (2005) Protease-activated receptors-1 and -2 can mediate endothelial barrier protection: role in factor Xa signaling. *J. Thromb. Haemost.* 3, 2798–2805.
- (17) Bae, J.-S., Yang, L., Manithody, C., and Rezaie, A. R. (2007) The ligand occupancy of endothelial protein C receptor switches the protease-activated receptor 1-dependent signaling specificity of thrombin from a permeability-enhancing to a barrier-protective response in endothelial cells. *Blood* 110, 3909–3916.
- (18) Kerschen, E. J., Fernandez, J. A., Cooley, B. C., Yang, X. V., Sood, R., Mosnier, L. O., Castellino, F. J., Mackman, N., Griffin, J. H., and Weiler, H. (2007) Endotoxemia and sepsis mortality reduction by non-anticoagulant activated protein C. *Journal of Experimental Medicine* 204, 2439–2448.
- (19) Soh, U. J., and Trejo, J. (2011) Activated protein C promotes protease-activated receptor-1 cytoprotective signaling through β -arrestin and dishevelled-2 scaffolds. *Proc. Natl. Acad. Sci. U.S.A.* 108, E1372–E1380.
- (20) Madhusudhan, T., Wang, H., Straub, B. K., Grone, E., Zhou, Q., Shahzad, K., Muller-Krebs, S., Schwenger, V., Gerlitz, B., Grinnell, B. W., Griffin, J. H., Reiser, J., Grone, H. J., Esmon, C. T., Nawroth, P. P., and Isermann, B. (2012) Cytoprotective signaling by activated protein C requires protease-activated receptor-3 in podocytes. *Blood* 119, 874–883.
- (21) Schuepbach, R. A., Madon, J., Ender, M., Galli, P., and Riewald, M. (2012) Protease-activated receptor-1 cleaved at R46 mediates cytoprotective effects. *J. Thromb. Haemost.* 10, 1675–1684.
- (22) Mosnier, L. O., Sinha, R. K., Burnier, L., Bouwens, E. A., and Griffin, J. H. (2012) Biased agonism of protease-activated receptor 1 by activated protein C caused by noncanonical cleavage at Arg46. *Blood* 120, 5237–5246.
- (23) Burnier, L., and Mosnier, L. O. (2013) Novel mechanisms for activated protein C cytoprotective activities involving noncanonical activation of protease-activated receptor 3. *Blood* 122, 807–816.
- (24) Liang, H. P. H., Kerschen, E. J., Hernandez, I., Basu, S., Zogg, M., Botros, F., Jia, S., Hessner, M. J., Griffin, J. H., Ruf, W., and Weiler, H. (2015) EPCR-dependent PAR2 activation by the blood coagulation initiation complex regulates LPS-triggered interferon responses in mice. *Blood* 125, 2845–2854.
- (25) Áinle, F. N., O'Donnell, J. S., Johnson, J. A., Brown, L., Gleeson, E. M., Smith, O. P., and Preston, R. J. S. (2011) Activated Protein C N-Linked Glycans Modulate Cytoprotective Signaling Function on Endothelial Cells. *J. Biol. Chem.* 286, 1323–1330.
- (26) Rothmeier, A. S., and Ruf, W. (2012) Protease-activated receptor 2 signaling in inflammation. *Semin Immunopathol* 34, 133–149.
- (27) Bouwens, E., Stavenuiter, F., and Mosnier, L. O. (2013) Mechanisms of anticoagulant and cytoprotective actions of the protein C pathway. *Journal of Thrombosis and Haemostasis* 11, 242–253.
- (28) Hollenberg, M. D., Mihara, K., Polley, D., Suen, J. Y., Han, A., Fairlie, D. P., and Ramachandran, R. (2014) Biased signalling and proteinase-activated receptors (PARs): targeting inflammatory disease. *British Journal of Pharmacology* 171, 1180–1194.
- (29) Bock, F., Shahzad, K., Vergnolle, N., and Isermann, B. (2014) Activated protein C based therapeutic strategies in chronic diseases. *Thromb. Haemost.* 111, 610–617.
- (30) Rezaie, A. R. (2014) Protease-activated receptor signalling by coagulation proteases in endothelial cells. *Thromb. Haemost.* 112, 876–882.
- (31) Griffin, J. H., Zlokovic, B. V., and Mosnier, L. O. (2015) Activated protein C: biased for translation. *Blood* 125, 2898–2907.
- (32) De Ceunynck, K., Peters, C. G., Jain, A., Higgins, S. J., Aisiku, O., Fitch-Tewfik, J. L., Chaudhry, S. A., Dockendorff, C., Parikh, S. M., Ingber, D. E., and Flaumenhaft, R. (2018) PAR1 agonists stimulate APC-like endothelial cytoprotection and confer resistance to thromboinflammatory injury. *Proc. Nat. Acad. Sci. USA in press*
<https://doi.org/10.1073/pnas.1718600115>
- (33) Nazir, S., Gadi, I., Al-Dabet, M. M., Elwakiel, A., Kohli, S., Ghosh, S., Manoharan, J., Ranjan, S., Bock, F., Braun-Dullaeus, R. C., Esmon, C. T., Huber, T. B., Camerer, E., Dockendorff, C., Griffin, J. H., Isermann, B., and Shahzad, K. (2017) Cytoprotective activated protein C averts Nlrp3 inflammasome induced ischemia reperfusion injury via mTORC1 inhibition. *Blood* 129, 2017–05–782102.
- (34) Ahn, K., Pan, S., Beningo, K., and Hupe, D. (1995) A permanent human cell line (EA.hy926) preserves the characteristics of endothelin converting enzyme from primary human umbilical vein endothelial cells. *Life Sci* 56, 2331–2341.
- (35) Dömötör, E., Benzakour, O., Griffin, J. H., Yule, D., Fukudome, K., and Zlokovic, B. V. (2003) Activated protein C alters cytosolic calcium flux in human brain endothelium via binding to endothelial protein C receptor and activation of protease activated receptor-1. *Blood* 101, 4797–4801.
- (36) Kuckleburg, C. J., and Newman, P. J. (2013) Neutrophil Proteinase 3 Acts on Protease-Activated Receptor-2 to Enhance Vascular Endothelial Cell Barrier Function. *Arteriosclerosis, Thrombosis, and Vascular Biology* 33, 275–284.
- (37) Barry, G. D., Suen, J. Y., Le, G. T., Cotterell, A., Reid, R. C., and Fairlie, D. P. (2010) Novel Agonists and Antagonists for Human Protease Activated Receptor 2. *J. Med. Chem.* 53, 7428–7440.
- (38) Asteriti, S., Daniele, S., Porchia, F., Dell'Anno, M. T., Fazzini, A., Pugliesi, I., Trincavelli, M. L., Taliani, S., Martini,

- C., Mazzone, M. R., and Gilchrist, A. (2012) Modulation of PAR1 signalling by benzimidazole compounds. *British Journal of Pharmacology* 167, 80–94.
- (39) Suen, J. Y., Barry, G. D., Lohman, R. J., Halili, M. A., Cotterell, A. J., Le, G. T., and Fairlie, D. P. (2012) Modulating human proteinase activated receptor 2 with a novel antagonist (GB88) and agonist (GB110). *British Journal of Pharmacology* 165, 1413–1423.
- (40) Zhang, H.-C., Derian, C. K., Andrade-Gordon, P., Hoekstra, W. J., McComsey, D. F., White, K. B., Poulter, B. L., Addo, M. F., Cheung, W.-M., Damiano, B. P., Oksenberg, D., Reynolds, E. E., Pandey, A., Scarborough, R. M., and Maryanoff, B. E. (2001) Discovery and Optimization of a Novel Series of Thrombin Receptor (PAR-1) Antagonists: Potent, Selective Peptide Mimetics Based on Indole and Indazole Templates. *J. Med. Chem.* 44, 1021–1024.
- (41) Andrade-Gordon, P., Maryanoff, B. E., Derian, C. K., Zhang, H.-C., Addo, M. F., Darrow, A. L., Eckardt, A. J., Hoekstra, W. J., McComsey, D. F., and Oksenberg, D. (1999) Design, synthesis, and biological characterization of a peptide-mimetic antagonist for a tethered-ligand receptor. *Proc. Natl. Acad. Sci. U.S.A.* 96, 12257–12262.
- (42) Zhang, H.-C., White, K. B., McComsey, D. F., Addo, M. F., Andrade-Gordon, P., Derian, C. K., Oksenberg, D., and Maryanoff, B. E. (2003) High-affinity thrombin receptor (PAR-1) ligands: a new generation of indole-based peptide mimetic antagonists with a basic amine at the C-terminus. *Bioorg. Med. Chem. Lett.* 13, 2199–2203.
- (43) Derian, C. K., Damiano, B. P., Addo, M. F., Darrow, A. L., D'Andrea, M. R., Nedelman, M., Zhang, H.-C., Maryanoff, B. E., and Andrade-Gordon, P. (2003) Blockade of the thrombin receptor protease-activated receptor-1 with a small-molecule antagonist prevents thrombus formation and vascular occlusion in nonhuman primates. *J. Pharmacol. Exp. Ther.* 304, 855–861.
- (44) Valdivielso, Á. M., García-López, M. T., and Herranz, R. (2008) Improved synthesis of the PAR-1 thrombin receptor antagonist RWJ-58259. *Arkivoc* 2008, 287.
- (45) Robinson, E., Knight, E., Smoktunowicz, N., Chambers, R. C., Inglis, G. G., Chudasama, V., and Caddick, S. (2016) Identification of an active metabolite of PAR-1 antagonist RWJ-58259 and synthesis of analogues to enhance its metabolic stability. *Org. Biomol. Chem.* 14, 3198–3201.
- (46) Meldal, M., and Tornøe, C. W. (2008) Cu-Catalyzed Azide–Alkyne Cycloaddition. *Chem. Rev.* 108, 2952–3015.
- (47) Suzuki, S., Kotake, M., Miyamoto, M., Kawahara, T., Kajiwara, A., Hishinuma, I., Okano, K., Miyazawa, S., Clark, R., Ozaki, F., Sato, N., Shinoda, M., Kamada, A., Tsukada, I., Matsuura, F., Naoe, Y., Terauchi, T., Oohashi, Y., Ito, O., Tanaka, H., Musya, T., Kogushi, M., Kawada, T., Matsuoka, T., Kobayashi, H., Chiba, K.-I., Kimura, A., Ono, N., Eisai Co. Ltd. (2002) 2-iminopyrrolidine derivatives.
- (48) Yau, M.-K., Liu, L., Lim, J., Lohman, R.-J., Cotterell, A. J., Suen, J. Y., Vesey, D. A., Reid, R. C., and Fairlie, D. P. (2016) *Bioorganic & Medicinal Chemistry Letters*. *Bioorg. Med. Chem. Lett.* 26, 986–991.
- (49) Tsien, R. Y. (1981) A non-disruptive technique for loading calcium buffers and indicators into cells. *Nature* 290, 527–528.
- (50) Gopalakrishnan, S. M., Mammen, B., Schmidt, M., Otterstaeetter, B., Amberg, W., Wernet, W., Kofron, J. L., Burns, D. J., and Warrior, U. (2005) An offline-addition format for identifying GPCR modulators by screening 384-well mixed compounds in the FLIPR. *J. Biomol. Screen* 10, 46–55.
- (51) Tovey, S. C., Sun, Y., and Taylor, C. W. (2006) Rapid functional assays of intracellular Ca²⁺ channels. *Nat. Protoc.* 1, 259–263.
- (52) Zhang, J., Chung, T., and Oldenburg, K. (1999) A Simple Statistical Parameter for Use in Evaluation and Validation of High Throughput Screening Assays. *J. Biomol. Screen* 4, 67–73.
- (53) Zhang, C., Srinivasan, Y., Arlow, D. H., Fung, J. J., Palmer, D., Zheng, Y., Green, H. F., Pandey, A., Dror, R. O., Shaw, D. E., Weis, W. I., Coughlin, S. R., and Kobilka, B. K. (2012) High-resolution crystal structure of human protease-activated receptor 1. *Nature* 492, 387–392.
- (54) Flaumenhaft, R., and De Ceunynck, K. (2017) Targeting PAR1: Now What? *Trends in Pharmacological Sciences* 38, 701–716.

Supplementary Material

NMR spectra and LC-MS traces are in the accompanying file.